## A peer-reviewed version of this preprint was published in PeerJ on 10 January 2019.

<u>View the peer-reviewed version</u> (peerj.com/articles/6252), which is the preferred citable publication unless you specifically need to cite this preprint.

Andrew NR, Miller C, Hall G, Hemmings Z, Oliver I. 2019. Aridity and land use negatively influence a dominant species' upper critical thermal limits. PeerJ 6:e6252 <u>https://doi.org/10.7717/peerj.6252</u>

# Ant thermal tolerances under climate, land cover and land use change

Nigel R Andrew Corresp., 1, 2 , Cara Miller <sup>3</sup> , Graham Hall <sup>2</sup> , Zac Hemmings <sup>1, 2</sup> , Ian Oliver <sup>2, 4</sup>

<sup>1</sup> UNE Centre of Excellence for Behavioural and Physiological Ecology, Natural History Museum, University of New England, Armidale, NSW, Australia

<sup>2</sup> School of Environmental and Rural Science, University of New England, Armidale, NSW, Australia

<sup>3</sup> School of Science and Technology, University of New England, Armidale, NSW, Australia

<sup>4</sup> Office of Environment and Heritage, Armidale, NSW, Australia

Corresponding Author: Nigel R Andrew Email address: nigel.andrew@une.edu.au

Thermal stress is a key issue for species dominant within ecosystems especially those that carry out key ecosystem service roles. When assessing the impacts of climate change it is critical to assess its biotic impacts relative to other anthropogenic changes to landscapes including the reduction of native vegetation cover, landscape fragmentation and changes in land use intensity. Here we integrate the observed phenotypic plasticity of the dominant and ubigitous meat ant Iridomyrmex purpureus in critical thermal limits across altitudinal, land cover and land use gradients to: (i) predict the adaptive capacity of a key terrestrial ecosystem service provider to changes in climate, land cover and land use, and (ii) assess the ability of multiple use landscapes to confer maximum resilience to terrestrial biodiversity in the face of a changing climate. The research was carried out along a 270km aridity gradient spanning 840m in altitude in northern New South Wales, Australia. When we assessed critical thermal maximum temperatures (CT<sub>max</sub>) of meat ants in relation to the environmental variables, and within the model we had critical thermal minimums of meat ants (CT<sub>min</sub>) as a random slope and as a fixed effect we detected a negative aridity effect on CT<sub>max</sub>, a negative effect of land use intensity, and no overall correlation between CT<sub>max</sub> and CT<sub>min</sub>. We also found a negative relationship with warming tolerance of *I. purpureus* and landscape aridity. In conclusion, we expect to see a reduction in the physiological resilience of *I. purpureus* as land use intensity increases and as the climate becomes more arid. Meat ants are key ecosystem engineers and as they are put under more stress, wider ecological implications may occur if populations decline or disappear.

### 1 Ant thermal tolerances under climate, land cover and land use change

2 N. R. Andrew<sup>1, 2</sup>, C. Miller<sup>3</sup>, G. Hall<sup>2</sup>, Z. Hemmings<sup>1, 2</sup>, and I. Oliver<sup>2, 4</sup>

<sup>1</sup> UNE Centre of Excellence for Behavioural and Physiological Ecology, Natural History Museum,
University of New England, Armidale, NSW, 2351.

- <sup>5</sup> <sup>2</sup> School of Environmental and Rural Science, University of New England, Armidale, NSW, 2351.
- <sup>6</sup> <sup>3</sup> School of Science and Technology, University of New England, Armidale, NSW, 2351
- <sup>7</sup> <sup>4</sup> Office of Environment and Heritage, PO Box U221, Armidale, NSW, 2351
- 8

### 9 Abstract

Thermal stress is a key issue for species dominant within ecosystems especially those that 10 carry out key ecosystem service roles. When assessing the impacts of climate change it is 11 12 critical to assess its biotic impacts relative to other anthropogenic changes to landscapes including the reduction of native vegetation cover, landscape fragmentation and changes in 13 14 land use intensity. Here we integrate the observed phenotypic plasticity of the dominant and ubigitous meat ant Iridomyrmex purpureus in critical thermal limits across altitudinal, land 15 cover and land use gradients to: (i) predict the adaptive capacity of a key terrestrial ecosystem 16 service provider to changes in climate, land cover and land use, and (ii) assess the ability of 17 multiple use landscapes to confer maximum resilience to terrestrial biodiversity in the face of a 18 19 changing climate. The research was carried out along a 270km aridity gradient spanning 840m in altitude in northern New South Wales, Australia. When we assessed critical thermal 20 21 maximum temperatures (CT<sub>max</sub>) of meat ants in relation to the environmental variables, and within the model we had critical thermal minimums of meat ants (CT<sub>min</sub>) as a random slope and 22 as a fixed effect we detected a negative aridity effect on CT<sub>max</sub>, a negative effect of land use 23 intensity, and no overall correlation between CT<sub>max</sub> and CT<sub>min</sub>. We also found a negative 24 relationship with warming tolerance of I. purpureus and landscape aridity. In conclusion, we 25 expect to see a reduction in the physiological resilience of *I. purpureus* as land use intensity 26 27 increases and as the climate becomes more arid. Meat ants are key ecosystem engineers and as they are put under more stress, wider ecological implications may occur if populations declineor disappear.

### 30 Introduction

31 Assessing the performance and physiological responses of ectotherms is critical to understanding biotic responses to climate change (Andrew & Terblanche 2013), particularly the 32 effects of exposure to thermal stress and temperature extremes (Vasseur et al. 2014) on key 33 ecosystem service providers. Biochemical and physiological reactions are mediated by 34 temperature and thermal stress can negatively influence development, growth, metabolism, 35 36 movement and reproduction, leading to changes in community and ecosystem level processes 37 (Dell et al. 2011; Grigaltchik et al. 2012). Thermal performance curves identify how the 38 performance or fitness of an ectotherm is influenced by body temperature (Sinclair et al. 2016), 39 a key response to thermal stress identified by these curves are critical thermal limits: the 40 functional endpoint that identify upper and lower limits of temperatures that insects can tolerate from which they are unable to escape (Lighton & Turner 2004). 41

42 Thermal stress is a key issue for all taxa including those that provide key ecosystem services 43 and are dominant within ecosystems (Andrew 2013; Andrew et al. 2016; Andrew et al. 2013a; Mooney et al. 2009). Through many terrestrial ecosystems worldwide, ants provide key 44 ecosystem services and mediate key ecosystem processes (Del Toro et al. 2012; Hölldobler & 45 Wilson 1990). The resilience of these ecosystem service providers to thermal stress may change 46 among populations as they are exposed to different environmental conditions, particularly in a 47 more variable climate (Greenslade 1976). This phenotypic (or physiological) plasticity is a core 48 driver of adaptive responses to climatic variation (Kingsolver & Huey 1998). 49

It is especially critical to take into account the impacts of climate change relative to other anthropogenic changes to landscapes including the reduction of native vegetation cover, landscape fragmentation and changes in land use intensity (Oliver & Morecroft 2014; Sala et al. 2000). Assessing the synergistic effects of changes in land cover, land use and climate are critical to enable decision makers to make better determinations in regards to the management

55 and conservation of biodiversity, ecosystem and environmental services both now and into the future (Mawdsley et al. 2009). Previously we assessed how adapting landscapes may improve 56 insect biodiveristy conservation via a study of the additive and synergistic effects of climate 57 58 with land cover and land use change (Oliver et al. 2016). From the main-effects models 59 developed, it was found that a greater amount of woody plant canopy cover increases ant richness (species and genus) and diversity; whereas a higher amount of land cultivation, 60 61 grazing, exotic plant groundcover and bare ground reduced species richness. At sites with warmer and drier climates (i.e. a higher aridity index), native plant canopy cover had greatest 62 benefit, and exotic plant cover had the most negative effects, on ant species richness (Oliver et 63 64 al. 2016). From this, we predict that the effects of landscape change on diversity may also affect 65 the thermal physiology of insect populations.

Changes in a dominant and widespread ectotherm's critical thermal limits across climatic, 66 67 land use and land cover gradients may occur (Angilletta et al. 2007): leading to changes in community structure and the provision of ecological services (Traill et al. 2010). Exposure to 68 69 different microclimates may influence ectotherm physiology in more unpredictable ways than 70 just exposure to warmer temperatures individually. Microclimates that ants are exposed to (e.g. 71 Andrew et al. 2013a; Hemmings & Andrew 2017) may change substantially across surfaces 72 within different habitat spaces: such as those with substantive bare ground, a high grazing 73 intensity, exotic plant species cover, and woody ground cover.

74 Here we focus on meat ants (Iridomyrmex purpureus (Smith, F., 1858)) as they are a dominant and ubiquitous part of the landscape (Andersen 2000; Greaves 1971; Greenslade 75 1976). Iridomyrmex purpureus can have a substantive impact on the availability of resources 76 77 and the use of these resources by other species in different landscapes (Gibb 2005). 78 *Iridomyrmex purpureus* are also excellent at resource exploitation and interference competiton to enable them to dominate and control resources quickly (Gibb & Hochuli 2004). They can also 79 maximise their foraging times by displaying opportunistic thermal responses and adjusting 80 81 foraging behaviour to deal with high trail temperatures (Andrew et al. 2013a).

82 Warming tolerance defines how much warming an ectotherm can tolerate before lethal levels are attained (Deutsch et al. 2008): it is calculated by taking the difference between the 83 upper critical thermal limit and the habitat ambient temperature. These values can change 84 85 substantially based on the method from which habitat temperatures are derived. For example, Andrew et al. (2013a) found I. purpureus' warming tolerance at a site in temperate Australia 86 (Armidale, New South Wales (NSW)) to be relatively high (25.8°C) when habitat temperatures 87 were based on closest weather station annual averages, but warming tolerance reduced when 88 closest weather station summer average temperatures and then microclimate summer average 89 temperatures were used (19.52°C and 19.12°C respectively). Warming tolerance decreased 90 91 substantially (to 7.81°C) when microclimate temperatures based on summer temperatures 92 between 10am and 4pm (when ants are most surface active) were used.

In this study we integrate observed ant phenotypic plasticity in critical thermal limits across altitudinal, land cover and land use gradients to: (i) predict the adaptive capacity of terrestrial invertebrate biodiversity to changes in climate, land cover and land use, and (ii) assess the ability of multiple use landscapes to confer maximum resilience to terrestrial biodiversity in the face of a changing climate.

98 The following questions were addressed:

What were the critical thermal limits (CT<sub>max</sub> and CT<sub>min</sub>) for *I. purpureus* across sites
 representing the main climatic, vegetation, and land-use regimes?

101 What were the key environmental drivers (climatic, vegetation, and land-use regimes)102 influencing thermal limits?

103 What is the relationship between *I. purpureus'* warming tolerance and aridity along the 104 environmental gradient?

### 105 Methods

106 Site selection

107 The study was carried out along a 270km aridity gradient spanning 840m in altitude in northern NSW, Australia (Table 1). The area has some of the most fertile soils in Australia, with 108 much of the farming practices dominated by livestock grazing on modified pastures and native 109 110 vegetation, and dryland and irrigated cropping (BRS 2009). Native remnant vegetation is dominated by semi-arid woodlands at lower altitudes through to grassy woodlands and dry 111 sclerophyll forest at higher altitudes (Keith 2004). Eleven sites were chosen to represent a 112 range of climatic, land-use and native woody vegetation cover along this gradient (Table 1) 113 covering the total number of sites (87) that were used to assess ant species diversity (Oliver et 114 al. 2016). Sites were chosen to maximise the range in climate (Aridity: based on rainfall and 115 evaporation collected from modelled climate data from ANUCLIM 6.1 (Xu & Hutchinson 2011) 116 117 over three time periods: 3 months, 12 months and 36 months), land cover (total native woody cover (Canopy) and bare ground), land use (intensity of use: Land Use Intensity - LUI, and exotic 118 groundcover) and soil pH and clay content. Land use intensity is a semi-quantitative index 119 based on cultivation and grazing severity and age: so more intensively managed sites have 120 higher values (ranging between 0 and 12). More information on how these variables were 121 calculated and justified can be found in Oliver et al. (2016). 122

123 We used the ant dataset collected from Oliver et al. (2016) to identify a common and widespread species to assess for critical thermal limits. Iridomyrmex purpureus was chosen for 124 physiological tolerance comparisons as it was the most abundant species at each site. A 125 126 minimum of 15 individual ants were collected from each site between April and May 2014, and then held at 25°C for two hours to avoid effects of time of day of capture differences along the 127 gradient. Previous work on *I. purpureus* found no effect of time of day of capture/ nest 128 129 temperature on thermal tolerances (determined via thermolimit respirometry) from a single 130 site (Andrew et al. 2016).

131 Critical thermal maximum and minimum assessments (CT<sub>max</sub> and CT<sub>min</sub>)

132 CT<sub>max</sub> measurements were carried out in a Grant R4 waterbath with a GP200 heater using
133 distilled water. Ten individuals from each site were each put into a single 50ml vial for testing,
134 and a reading for each ant was taken. Waterbath temperature was initialised for 10 minutes at

135 25°C and then ramped at 0.25°C/minute until CT<sub>max</sub> was reached. CT<sub>max</sub> was identified when an individual ant could not perform coordinated motor functions in the vial to right itself after 136 being turned onto its side (Andrew et al. 2013a). CT<sub>max</sub> could go up to 55°C (equivalent to 120 137 minutes/individual – 30°C temperature change at 0.25°C/min). Ramping at 0.25°C is considered 138 139 the most 'standard' temperature ramping rate, at which the body temperature of ants is in equilibrium with their surroundings (Andrew et al. 2013a; Chown et al. 2009; Lighton & Turner 140 141 2004; Nguyen et al. 2014; Terblanche et al. 2007). CT<sub>min</sub> was carried out in a similar fashion to 142 CT<sub>max</sub> using 1:1 distilled water/glycol mix. Waterbath temperature was initialised for 10 minutes at 5°C and then decreased at 0.25°C/minute until CT<sub>min</sub> was reached. CT<sub>min</sub> was identified when 143 an individual ant could not perform coordinated motor functions in a 50ml vial to right itself 144 after being turned onto its side (Andrew et al. 2013a). CT<sub>min</sub> could go down to -15°C (equivalent 145 to 80 minutes/species –  $20^{\circ}$ C temperature change at  $0.25^{\circ}$ C/min). To measure temperatures 146 that ants were exposed to within each vial, a Type-T thermocouple was placed within another 147 50ml vial that was plunged with the ants and connected to a temperature datalogger (Testo 148 175 T3) with data logged as waterbath temperatures were ramped: the Testo temperature was 149 used to identify ant  $CT_{max/min}$ . 150

151 Model fitting

We used R (R Core Team 2017) and the R package *lme4* (Bates et al. 2015) to perform a 152 153 linear mixed effects analysis of the relationship between CT<sub>max</sub> as a response variable against 154 the environmental variables of aridity, LUI (converted to a proportion), soil clay content, exotic 155 plant ground cover, and total native woody cover (Canopy) designated as fixed effects. We 156 explored singular interaction effects of Aridity:LUI, Canopy:LUI, and Clay:LUI in some models as 157 well as the impact of dropping main effect variables. With this framework we considered 158 random intercept models by site only, and by both site and CT<sub>min</sub> (individually). We also 159 considered a random intercept, random slope model with CT<sub>min</sub> within Site as the random effect. We repeated this model selection process with CT<sub>min</sub> as the response variable and CT<sub>max</sub> 160 as the predictor variable where appropriate. All variables were centred and scaled. Models 161 162 were initially fit with REML and then refitted with ML for comparison in Likelihood ratio tests.

Minimum AIC values and p-values of less than 0.05 were used to aid model selection. Visual inspection of residual plots of the preferred models were used to assess obvious deviations from homoscedasticity or normality. Visualization of random effects were undertaken using R package *sjPlot* (Lüdecke 2017). Standard errors and confidence intervals for predicted values of preferred models were undertaken using parametric bootstrapping (n = 1000) within R package *bootpredictIme4* (Duursma 2017) and visualized within R package *visreg* (Breheny & Burchett 2017).

#### 170 Warming Tolerance

171 Warming tolerance was calculated using the equation of Deutsch et al. (2008) and Diamond et al. (2012): CT<sub>max</sub> - T<sub>hab</sub>. The T<sub>hab</sub> calculation may include different calculations (e.g. annual 172 average; summer average; microclimate summer average; and microclimate summer 10am-173 4pm summer average) which are ecologically relevant and to identify the most appropriate to 174 assess ectotherm stress (e.g. Andrew et al. 2013a). For Than here, we did not have access to 175 microclimate data, so we modelled site location data using ANUCLIM V6.1 (Xu & Hutchinson 176 2011) from the closest weather stations based on 3 month summer average 2009, 12 month 177 average for 2009 and 36 month (2007-2009) average day temperatures. These weather data 178 were used, as the data were generated for all sites at the time of sampling ant species richness 179 in Oliver et al. (2016). 180

#### 181 **Results**

Critical thermal maxima of individual ants ranged between 41.5°C and 56.1°C, and CT<sub>min</sub> between 0.3°C and 7.1°C in this study. There was no consistent relationship between CT<sub>min</sub> and CT<sub>max</sub> across the 11 sites sampled (Figure 1), suggesting no causal relationship between the two end points.

186 The preferred model proposed for explaining meat ants CT<sub>max</sub> across the landscape is:

187  $CT_{max} \sim LUI + Canopy + Exotic + Clay + Aridity + (CT_{min}|SITE_ID)$ 

188 The fixed effects for this model are shown in Table 2. The overall random effects for the

189 model above are (in terms of variance): Site: 1.0448; CT<sub>min</sub>|Site: 0.1937; and Residuals: 1.4886.

190 As shown in Figure 2, Sites are an important source of variation (much more so than CT<sub>min</sub>

191 although the inclusion of this was still significant as per the model selection process). However,

there is still additional (unaccounted for) variation in the residuals. For the variables of LUI and

193 Clay, there were significant relationships with  $CT_{max}$  (Figure 3). As LUI increases,  $CT_{max}$ 

194 decreases; whereas clay content was positively correlated with CT<sub>max</sub>.

For explaining meat ants CT<sub>min</sub> across the landscape a similar model is proposed as that for
 CT<sub>max</sub>:

197  $CT_{min} \sim LUI + Canopy + Exotic + Clay + Aridity + (CT_{max}|SITE_ID)$ 

The fixed effects for this model are shown in Table 3. The overall random effects for the model above are (in terms of variance): Site: 1.3011;  $CT_{max}$  [Site: 0.3484; Residuals: 1.3769. As with  $CT_{max}$ , the sites also exhibit a high amount of variation (Figure 2). The prediction intervals for  $CT_{min}$  also show similar results as those for  $CT_{max}$ , (Figure 3c and d) however the relationships are weaker for both LUI and Clay content.

We found a negative relationship between the warming tolerances of *l. purpureus* and
landscape aridity (Figure 4). This relationship was consistent among all measures of mean
temperatures (no significant difference in Test for Common Slope across Groups: Test Statistic =
1.488, p = 0.482). There was a significant difference in the slope elevation of warming tolerance
between the three month and thirty six month mean temperature calculations (Figure 4; d.f. =
2, WALD = 95.299, p < 0.0001).</li>

### 209 Discussion

The phenotypic plasticity in critical thermal limits and physiological responses of insects to a changing climate is crucial for understanding how individuals and populations will respond to changes in their local environment (Andrew et al. 2013b; Andrew & Terblanche 2013). These responses are becoming a key area of research interest (Andrew et al. 2013a). The assessment

of common species responses to a changing climate needs to be thoroughly assessed, as
changes in these taxa's population structure can have large implications for the ecosystems in
which they provide key services (Andrew 2013; Gaston 2011; Inger et al. 2014). In addition, as
landscapes become more fragmented and anthropogenic, common and dominant species
responses to changes may also be limited. Here, critical thermal maxima and minima were
determined for ants that encompassed an extensive distribution along an environmental
gradient.

There was no strong pattern in  $CT_{max}$  and  $CT_{min}$  associated with the environmental variables tested. The results of the  $CT_{max}$  measurements indicates there is a high variation of  $CT_{max}$  across sites, this may be due to the ants being field fresh and so their previous exposure to a variety of stresses may influence their thermal capabilities. However, this is also important, as it indicates that no one individual stress dominates the thermal abilities of *I. purpureus* workers on site.

Critical thermal maxima of individual ants ranged between 41.5°C and 56.1°C, and CT<sub>min</sub> 226 between 0.3°C and 7.1°C in this study. This is a very wide range of readings for CT<sub>max</sub>, and could 227 be due to age, nutritional status, stress or prior heat exposure that the ants may have been 228 229 exposed to (Nyamukondiwa & Terblanche 2009; Sørensen et al. 2001). Upper thermal limits are 230 thought to be less plastic compared to lower limits, however it is known that environmental 231 exposure does influence these limits (Hoffmann et al. 2013), and this is seen with the relationships with both LUI and Clay in this study. Here we measured  $CT_{max}$  and  $CT_{min}$  by 232 observing an individual ant's ability to right itself while temperatures were increasing at 0.25°C 233 min<sup>-1</sup>. The calculation of critical thermal limits using ant righting behaviour may be more 234 235 variable than using physiological critical limits such as upper lethal temperatures where ants are exposed to static temperatures for two hours (Andrew et al. 2013a) or thermolimit 236 237 respirometry where CT<sub>max</sub> is derived from metabolic measurements using flow-through CO<sub>2</sub>based respirometry and optical detection, when temperatures are ramped at a consistent rate 238 (Andrew et al. 2016; Lighton & Turner 2004). As an alternative measure of CT<sub>max</sub> thermolimit 239 respirometry (Lighton & Turner 2004) may be more robust, as the method explicitly measures 240 the ceasing of metabolism (release of carbon dioxide) of the ant; but it is also a different 241

242 measure of CT<sub>max</sub>, as there is no ability for the ants to recover from heat exposure in
243 thermolimit respirometry.

When the fitted models were used to assess critical thermal limits, it is clear that site specific differences strongly influenced the results found. However, land use and soil clay content also played a significant role in influencing ant physiological end-points. This suggests that ant populations that were exposed to higher levels of habitat modification (via land use intensity) showed lower climatic resilience relative to less disturbed habitats. However, there is still additional unaccounted for variation in the residuals which suggests that there may be other variables (unmeasured) that may have an effect on the meat ants' climatic resilience.

251 For ants, much of the research on local effects of habitat disturbance has been carried out 252 on changes in communities (Andersen & Majer 2004; Andrew et al. 2000; Bromham et al. 253 1999). Previous work carried out along the gradient (Oliver et al. 2016) used for this study 254 found clear evidence for landscape adaptation to maintain and restore species richness of ant 255 communities at the site level. For ant communities, higher woody native cover and shrub cover, and lower exotic plant groundcover have a positive effect on ant species richness. Interestingly, 256 257 Land Use Intensity had no significant impact on the species richness within any of the ant genera assessed across the gradient. 258

However, clay did have a positive influence on *Iridomyrmex* spp. diversity in Oliver et al. 259 260 (2016). Clay and clay-like substrate is an important component for ant nest development 261 (Monaenkova et al. 2015), and is critical for other insect taxa, such as termites in giving their feeding galleries structural support to assist with load bearing (Oberst et al. 2016). As I. 262 *purpureus* nests are known to be located in the same location for over 70 + years (Greenslade 263 1975), substantive structural elements are required to keep it maintained during this time. The 264 amount of clay in a I. purpureus nest is representative of the surrounding non-nest soil 265 (Ettershank 1968). Iridomyrmex purpureus nests are also not found on quartz sand soils, even 266 when climatic factors are suitable, indicating that soil type can be a limiting distributional factor 267 268 (Greaves 1971). As clay plays a role in the distribution of the species, it also clearly pays a role in the physiological breadth of individuals. 269

For the  $CT_{max}$  model fitting, there was one model with a lower AIC (in which the interaction between LUI\*Canopy) was included. This was a more complex model, and so was not deemed the most appropriate to best explain the  $CT_{max}$  relationship. In addition, the current model is simpler to interpret and also in line with the results for  $CT_{min}$ .

We calculated warming tolerance using three different measures of habitat temperature, all 274 generated based on location data using ANUCLIM. These all indicated, as expected, that an 275 increase in aridity reduces ant tolerance to warming. When the warming tolerance was 276 277 previously calculated for *I. purpureus* at a higher altitude (Armidale, NSW: 980 masl), similar calculations were made: a warming tolerance of 19.5°C was calculated on weather station 278 summer average, and 25.8°C based on weather station annual average (Andrew et al. 2013a). 279 280 As Armidale is a more temperate site than those tested here, it would be at the lower scale of 281 the aridity index. Across the aridity index there is a 10°C difference in warming tolerance for *I*. 282 *purpureus*. With a prediction of global increases in air temperature of 2°C and 6°C over the 21<sup>st</sup> 283 Century, and in the region assessed there is an 80% probability of a 3°C warming and a 30% 284 probability of a 4°C warming with a likelihood of reduced annual rainfall of 3-5% (CSIRO-ABM 285 2012), aridity of the region assessed will only continue to increase.

#### 286 Conclusion

From this study we have found that habitat type (e.g. soils) and land-use intensity are more limiting factors on meat ant  $CT_{max}$  and  $CT_{min}$  than climatic factors (here we tested aridity). These populations are key ecosystem engineers and as they are put under more stress, wider ecological implications may occur if population abundances decline, as we expect to see a reduction in the physiological resilience of *I. purpureus* as land use intensity increases.

292

### 293 **References**

294

295	Andersen AN. 2000. The Ants of Northern Australia: A guide to the monsoonal fauna.
296	Collingwood: CSIRO.
297	Andersen AN, and Majer JD. 2004. Ants show the way Down Under: invertebrates as
298	bioindicators in land management. Frontiers in Ecology and the Environment 2:291-298.
299	10.1890/1540-9295(2004)002[0292:ASTWDU]2.0.CO;2
300	Andrew N, Rodgerson L, and York A. 2000. Frequent fuel-reduction burning: the role of logs and
301	associated leaf litter in the conservation of ant biodiversity. Austral Ecology 25:99-107.
302	10.1046/j.1442-9993.2000.01015.x
303	Andrew NR. 2013. Population dynamics of insects: impacts of a changing climate. In: Rohde K,
304	ed. The Balance of Nature and Human Impact: Cambridge University Press, 311-323.
305	Andrew NR, Ghaedi B, and Groenewald B. 2016. The role of nest surface temperatures and the
306	brain in influencing ant metabolic rates. Journal of Thermal Biology 60:132-139.
307	10.1016/j.jtherbio.2016.07.010
308	Andrew NR, Hart RA, Jung M-P, Hemmings Z, and Terblanche JS. 2013a. Can temperate insects
309	take the heat? A case study of the physiological and behavioural responses in a common
310	ant, Iridomyrmex purpureus (Formicidae), with potential climate change. Journal of
311	Insect Physiology 59:870-880. 10.1016/j.jinsphys.2013.06.003
312	Andrew NR, Hill SJ, Binns M, Bahar MH, Ridley EV, Jung M-P, Fyfe C, Yates M, and Khusro M.
313	2013b. Assessing insect responses to climate change: What are we testing for? Where
314	should we be heading? PeerJ 1:e11. 10.7717/peerj.11
315	Andrew NR, and Terblanche JS. 2013. The response of insects to climate change. In: Salinger J,
316	ed. Living in a Warmer World: How a changing climate will affect our lives. Auckland:
317	David Bateman Ltd 38-50.
318	Angilletta MJ, Jr., Wilson RS, Niehaus AC, Sears MW, Navas CA, and Ribeiro PL. 2007. Urban
319	physiology: City ants possess high heat tolerance. PLoS ONE 2:e258.
320	10.1371/journal.pone.0000258

- Bates D, Mächler M, Bolker B, and Walker S. 2015. Fitting linear mixed-effects models using
  Ime4. 2015 67:48. 10.18637/jss.v067.i01
  Breheny P, and Burchett W. 2017. visreg: Visualization of Regression Models. R package
  version 2.4-1. <u>https://CRAN.R-project.org/package=visreg</u>.
  Bromham L, Cardillo M, Bennett AF, and Elgar MA. 1999. Effects of stock grazing on the ground
  invertebrate fauna of woodland remnants. *Australian Journal of Ecology* 24:199-207.
- BRS. 2009. Land use summary border rivers/Gwydir NRM Region—NSW. Catchment scale land
- 328 use mapping for Australia. Update May 2009 dataset. Bureau of Rural Sciences.
- 329 <u>http://www.daff.gov.au/abares/aclump/pages/land-use/catchment-scale-land-use-</u>
- 330 <u>reports.aspx</u>. Accessed July 2015.
- 331 Chown SL, Jumbam KR, Sørensen JG, and Terblanche JS. 2009. Phenotypic variance, plasticity
- and heritability estimates of critical thermal limits depend on methodological context.
- 333 Functional Ecology 23:133-140.
- 334 CSIRO-ABM. 2012. State of the Climate 2012. Canberra. Available at
- 335 http://www.csiro.au/Outcomes/Climate/Understanding/State-of-the-Climate-
- 336 <u>2012.aspx</u>: CSIRO and the Australian Bureau of Meteorology.
- 337 Del Toro I, Ribbons RR, and Pelini SL. 2012. The little things that run the world revisited: a
- review of ant-mediated ecosystem services and disservices (Hymenoptera: Formicidae).
- 339 *Myrmecological News* 17:133-146.
- 340 Dell AI, Pawar S, and Savage VM. 2011. Systematic variation in the temperature dependence of
- physiological and ecological traits. *Proceedings of the National Academy of Sciences*108:10591-10596. 10.1073/pnas.1015178108
- 343 Deutsch CA, Tewksbury JJ, Huey RB, Sheldon KS, Ghalambor CK, Haak DC, and Martin PR. 2008.
- Impacts of climate warming on terrestrial ectotherms across latitude. *Proceedings of the National Academy of Sciences* 105:6668-6672. 10.1073/pnas.0709472105
- Diamond SE, Sorger DM, Hulcr J, Pelini SL, Toro ID, Hirsch C, Oberg E, and Dunn RR. 2012. Who
- 347 likes it hot? A global analysis of the climatic, ecological, and evolutionary determinants
- of warming tolerance in ants. *Global Change Biology* 18:448-456. 10.1111/j.1365-
- 349 2486.2011.02542.x

- 350 Duursma R. 2017. bootpredictlme4 GitHub repository, <u>https://github.com/remkoduursma</u>.
- 351 Ettershank G. 1968. The three dimensional gallery structure of the nest of the meat ant
- 352 Iridomyrmex Purpureus (SM.) (Hymenoptera : Formicidae). *Australian Journal of*
- 353 Zoology 16:715-723. <u>https://doi.org/10.1071/Z09680715</u>
- 354 Gaston KJ. 2011. Common Ecology. *BioScience* 61:354-362.
- Gibb H. 2005. The effect of a dominant ant, *Iridomyrmex purpureus*, on resource use by ant
- assemblages depends on microhabitat and resource type. *Austral Ecology* 30:856-867.
  10.1111/j.1442-9993.2005.01528.x
- 358 Gibb H, and Hochuli DF. 2004. Removal experiment reveals limited effects of a behaviorally
- dominant species on ant assemblages. *Ecology* 85:648-657. 10.1890/03-0007
- 360 Greaves T. 1971. The distribution of the three forms of the meat ant *Iridomyrmex purpureus*
- 361 (Hymenoptera: Formicidae) in Australia. *Australian Journal of Entomology* 10:15-21.
- 362 10.1111/j.1440-6055.1971.tb00004.x
- 363 Greenslade P. 1975. Dispersion and history of a population of the meat ant *Iridomyrmex*
- 364 *purpureus* (Hymenoptera: Formicidae). *Australian Journal of Zoology* 23:495-510.
- 365 <u>https://doi.org/10.1071/Z09750495</u>
- 366 Greenslade PJM. 1976. The meat ant *Iridomyrmex purpureus* (Hymenoptera: Formicidae) as a
- dominant member of ant communities. *Australian Journal of Entomology* 15:237-240.
- 368 10.1111/j.1440-6055.1976.tb01700.x
- 369 Grigaltchik VS, Ward AJW, and Seebacher F. 2012. Thermal acclimation of interactions:
- 370 differential responses to temperature change alter predator–prey relationship.
- 371 *Proceedings of the Royal Society B: Biological Sciences* 279:4058-4064.
- 372 10.1098/rspb.2012.1277
- 373 Hemmings Z, and Andrew NR. 2017. Effects of microclimate and species identity on body
- temperature, and thermal tolerance, of ants (Hymenoptera: Formicidae). *Austral Entomology* 56:104-114. 10.1111/aen.12215
- 376 Hoffmann AA, Chown SL, and Clusella-Trullas S. 2013. Upper thermal limits in terrestrial
- ectotherms: How constrained are they? *Functional Ecology* 27:934-949.
- Hölldobler B, and Wilson EO. 1990. *The Ants*. Cambridge, MA: Belknap Press.

379	Inger R, Gregory R, Duffy JP, Stott I, Voříšek P, and Gaston KJ. 2014. Common European birds
380	are declining rapidly while less abundant species' numbers are rising. Ecology
381	<i>Letters</i> :n/a-n/a. 10.1111/ele.12387
382	Keith DA. 2004. Ocean shores to desert dunes: the native vegetation of New South Wales and
383	the ACT Sydney: Department of Environment and Conservation.
384	Kingsolver JG, and Huey RB. 1998. Evolutionary analyses of morphological and physiological
385	plasticity in thermally variable environments. American Zoologist 38:545-560.
386	Lighton JR, and Turner RJ. 2004. Thermolimit respirometry: an objective assessment of critical
387	thermal maxima in two sympatric desert harvester ants, Pogonomyrmex rugosus and P.
388	californicus. Journal of Experimental Biology 207:1903-1913. 10.1242/jeb.00970
389	Lüdecke D. 2017. sjPlot: Data Visualization for Statistics in Social Science. R package version
390	2.3.2, <u>https://CRAN.R-project.org/package=sjPlot</u> .
391	Mawdsley JR, O'Malley R, and Ojima DS. 2009. A review of climate-change adaptation strategies
392	for wildlife management and biodiversity conservation. Conservation Biology 23:1080-
393	1089. 10.1111/j.1523-1739.2009.01264.x
394	Monaenkova D, Gravish N, Rodriguez G, Kutner R, Goodisman MAD, and Goldman DI. 2015.
395	Behavioral and mechanical determinants of collective subsurface nest excavation. The
396	Journal of Experimental Biology 218:1295-1305. 10.1242/jeb.113795
397	Mooney H, Larigauderie A, Cesario M, Elmquist T, Hoegh-Guldberg O, Lavorel S, Mace GM,
398	Palmer M, Scholes R, and Yahara T. 2009. Biodiversity, climate change, and ecosystem
399	services. Current Opinion in Environmental Sustainability 1:46-54.
400	http://dx.doi.org/10.1016/j.cosust.2009.07.006
401	Nguyen C, Bahar MH, Baker G, and Andrew NR. 2014. Thermal tolerance limits of diamondback
402	moth in ramping and plunging assays. PLoS ONE 9:e87535.
403	10.1371/journal.pone.0087535
404	Nyamukondiwa C, and Terblanche JS. 2009. Thermal tolerance in adult Mediterranean and
405	Natal fruit flies (Ceratitis capitata and Ceratitis rosa): Effects of age, gender and feeding
406	status. Journal of Thermal Biology 34:406-414.

407	Oberst S, Lai JCS, and Evans TA. 2016. Termites utilise clay to build structural supports and so
408	increase foraging resources. Scientific Reports 6:20990. 10.1038/srep20990
409	Oliver I, Dorrough J, Doherty H, and Andrew NR. 2016. Additive and synergistic effects of land
410	cover, land use and climate on insect biodiversity. Landscape Ecology 31:2415-2431.
411	10.1007/s10980-016-0411-9
412	Oliver TH, and Morecroft MD. 2014. Interactions between climate change and land use change
413	on biodiversity: attribution problems, risks, and opportunities. Wiley Interdisciplinary
414	Reviews: Climate Change 5:317-335. 10.1002/wcc.271
415	R Core Team. 2017. R: A language and environment for statistical computing. R Foundation for
416	Statistical Computing, Vienna, Austria.
417	Sala OE, Stuart Chapin F, III, Armesto JJ, Berlow E, Bloomfield J, Dirzo R, Huber-Sanwald E,
418	Huenneke LF, Jackson RB, Kinzig A, Leemans R, Lodge DM, Mooney HA, Oesterheld Mn,
419	Poff NL, Sykes MT, Walker BH, Walker M, and Wall DH. 2000. Global biodiversity
420	scenarios for the year 2100. Science 287:1770-1774. 10.1126/science.287.5459.1770
421	Sinclair BJ, Marshall KE, Sewell MA, Levesque DL, Willett CS, Slotsbo S, Dong Y, Harley CDG,
422	Marshall DJ, Helmuth BS, and Huey RB. 2016. Can we predict ectotherm responses to
423	climate change using thermal performance curves and body temperatures? Ecology
424	Letters 19:1372-1385. 10.1111/ele.12686
425	Sørensen JG, Dahlgaard J, and Loeschcke V. 2001. Genetic variation in thermal tolerance among
426	natural populations of Drosophila buzzatii: down regulation of Hsp70 expression and
427	variation in heat stress resistance traits. <i>Functional Ecology</i> 15:289-296.
428	doi:10.1046/j.1365-2435.2001.00525.x
429	Terblanche JS, Deere JA, Clusella-Trullas S, Janion C, and Chown SL. 2007. Critical thermal limits
430	depend on methodological context. Proceedings of the Royal Society B: Biological
431	Sciences 274:2935-2943. 10.1098/rspb.2007.0985
432	Traill LW, Lim MLM, Sodhi NS, and Bradshaw CJA. 2010. Mechanisms driving change: altered
433	species interactions and ecosystem function through global warming. Journal of Animal
434	Ecology 79:937-947. 10.1111/j.1365-2656.2010.01695.x

435 Vasseur DA, DeLong JP, Gilbert B, Greig HS, Harley CDG, McCann KS, Savage V, Tunney TD, and

- 436 O'Connor MI. 2014. Increased temperature variation poses a greater risk to species than
- 437 climate warming. *Proceedings of the Royal Society B: Biological Sciences* 281.

438 10.1098/rspb.2013.2612

- 439 Xu T, and Hutchinson MN. 2011. ANUCLIM version 6, users guide. . Canberra: Australian
- 440 National University.

441

### Table 1(on next page)

Characteristics of the 11 sites used in this study (from a total of 87). See Oliver *et al.* ( 2016 ) for details on the full complement of sites.

1 Table 1. Characteristics of the 11 sites used in this study (from a total of 87). See Oliver *et al.* (2016) for details on the full complement of sites.

					Total				
			Land		native	Exotic			
			Use	Soil	woody	Ground			
Site name	Aridity	Altitude	Intensity	Clay	cover	Cover	рН	Lat	Long
Smokey Mountain (38)	0.401	891	2	5	23	50	4.585	-29.966	151.271
Furrocabad Station									
(44	0.346	1047	3	28.8	30	69	5.25	-29.83	151.608
Furrocabad Station									
(45)	0.366	1008	8	28.8	97	68	5.64	-29.823	151.598
Delunga 52	0.617	338	0	37.5	76	9	6.16	-29.835	150.554
62	0.716	203	4	27.5	16	43	5.675	-29.379	149.797
63	0.715	204	5	30	66	2	5.855	-29.378	149.796
87C	0.732	163	6	57.5	1	83	6.425	-29.693	149.23
Towarra (96)	0.537	643	1	28.8	70	1	5.56	-30.125	150.76
Myall Creek (117)	0.583	457	1	65	68	18	6.465	-29.823	150.74
West Oaks (126C)	0.491	730	4	15	97	2	5.89	-29.359	151.429
West Oaks (127C)	0.508	683	5	53.8	1	10	5.695	-29.36	151.412

2

### Table 2(on next page)

Estimated fixed effects for the selected  $CT_{max}$  model. Standard errors and 95% confidence intervals are also presented. All variables have been centred.

- 1 Table 2. Estimated fixed effects for the selected  $CT_{max}$  model. Standard errors and 95% confidence
- 2 intervals are also presented. All variables have been centred.

	Estimate	Std. Error	2.5 %	97.5 %
(Intercept)	46.04	0.30	45.52	46.54
Land Use	-0.28	0.15	-0.48	-0.06
Intensity (LUI)				
Total native	0.01	0.01	-0.014	0.02
woody cover				
(Canopy)				
Exotic	0.01	0.01	-0.01	0.03
groundcover				
Soil Clay Content	0.04	0.02	0.01	0.06
Aridity Index	2.82	2.54	-6.37	0.18

3

### Table 3(on next page)

Estimated fixed effects for the selected  $CT_{min}$  model. Standard errors and 95% confidence intervals are also presented. All variables have been centred.

- 1 Table 3. Estimated fixed effects for the selected  $CT_{min}$  model. Standard errors and 95% confidence
- 2 intervals are also presented. All variables have been centred.

	Estimate	Std. Error	2.5%	97.5%
(Intercept)	3.7	0.37	2.85	4.28
Land Use Intensity	-0.091	0.18	-0.34	0.15
Total native woody	0.00	0.013	-0.03	0.02
cover (Canopy)				
Exotic Groundcover	0.004	0.016	-0.02	0.023
Soil Clay Content	0.017	0.025	-0.01	0.06
Aridity Index	0.29	3.3	-4.24	5.01

3

### Figure 1(on next page)

Sites used in this study in northern New South Wales, Australia (see insert) with relative values for  $CT_{max}$  (A), Land Use Intensity (B), Aridity (C), and Clay (D) shown.

Maps generated using Map data © OpenStreetMap contributors. The size of the circle is indicative of the mean value of the given variable (i.e. the larger the circle, the higher the value, and the contrary). Image produced using the Leaflet package (version 1.1.0.9000, http://rstudio.github.io/leaflet/) within R statistical software (version 3.4.3). The R package OpenStreetMap is licensed under a GNU General Public License (GPL-2) (https://cran.r-project.org/web/packages/OpenStreetMap/index.html) and was used to extract map tiles from OpenStreetMap which is licensed on terms of the Open Database License, "ODbL" 1.0. (http://wiki.osmfoundation.org/wiki/Licence).



### Figure 2(on next page)

Random effect estimates of model coefficients using Best Linear Unbiased Prediction (BLUP) and 95% confidence intervals of the intercepts and  $CT_{max}$  (centred – Ctmaxc) and  $CT_{min}$  (centred – Ctminc) across sites.



PeerJ Preprints | https://doi.org/10.7287/peerj.preprints.26854v1 | CC BY 4.0 Open Access | rec: 13 Apr 2018, publ: 13 Apr 2018 Best Linear Unbiased Prediction (BLUP)

Sites

### Figure 3(on next page)

95% confidence intervals on the selected  $CT_{max}$  and  $CT_{min}$  models for the standardised factors of Land Use Intensity (LUI) and Clay.

### NOT PEER-REVIEWED





LUI (standardised)

PeerJ Preprints | https://doi.org/10.7287/peerj.preprints.26854v1 | CC BY 4.0 Open Access | rec: 13 Apr 2018, publ: 13 Apr 2018

### Figure 4(on next page)

Three measures of warming tolerance (3 months, 12 months and 36 months); based on location modelled ANUCLIM data for three different sampling periods) relative to aridity index for each site.



Peer/ Preprints | https://doi.org/10.7287/peeri.pr