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Novel epitopes identified from efflux pumps of *Mycobacterium tuberculosis* could induce cytotoxic T lymphocyte response

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Overcoming drug-resistance is one of the major challenges to control tuberculosis (TB). The up-regulation of efflux pumps is one common mechanism that leads to drug-resistance. Therefore, immunotherapy targeting these efflux pump antigens could be promising strategy to be combined with current chemotherapy. Considering that CD8+ cytotoxic T lymphocytes (CTLs) induced by antigenic peptides (epitopes) could elicit HLA-restricted anti-TB immune response, efflux pumps from classical ABC family (*Mycobacterium tuberculosis*, Mtb) were chosen as target antigens to identify CTL epitopes. HLA-A2 restricted candidate peptides from Rv2937, Rv2686c and Rv2687c of *Mycobacterium tuberculosis* were predicted, synthesized and tested. Five peptides could induce IFN- γ release and cytotoxic activity in PBMCs from HLA-A2⁺ PPD⁺ donors. Results from HLA-A2/K^b transgenic mice immunization assay suggested that four peptides Rv2937-p168, Rv2937-p266, Rv2686c-p151, and Rv2686c-p181 could induce significant CTL response *in vivo*. These results suggested that these novel epitopes could be used as immunotherapy candidates to TB drug-resistance.

1 **Novel epitopes identified from efflux pumps of *Mycobacterium tuberculosis* could induce**
2 **cytotoxic T lymphocyte response**

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26

27 INTRODUCTION

28 Tuberculosis is a serious infection disease in developing countries, which is caused by
29 *Mycobacterium tuberculosis* (Dye & Williams 2010). To date, the available strategies against
30 tuberculosis mainly rely on chemotherapeutic regimens. However, with the emergence of
31 multidrug-resistant tuberculosis (MDR-TB) and extensively drug resistant tuberculosis
32 (XDR-TB), it becomes more difficult to fight against tuberculosis. In addition to acquired
33 drug resistance (Ramaswamy & Musser 1998), intrinsic drug resistance has been proved in
34 *Mycobacterium tuberculosis* (Mtb) (De Rossi et al. 2006), in which active drug efflux pump
35 does make contributions to drug resistance (Escribano et al. 2007; Spies et al. 2008).

36 As an intracellular pathogen living in macrophages, more and more evidence suggested the
37 important role of cellular immunity in controlling the dissemination of Mtb. It has been
38 proved that, in addition to CD4⁺ T cells, MHC class I restricted CD8⁺ T cells also play very
39 important role in immune responses against Mtb. And large amounts of previous results have
40 proved the role of CD8⁺ T cell-mediated immune responses in Mtb challenged mouse models,
41 nonhuman primates as well as patients (D'Souza et al. 2000; Flynn & Chan 2001; Flynn et al.
42 1992; Lazarevic & Flynn 2002; Sousa et al. 2000).

43 Although a lot of researchers are trying to develop new vaccines, BCG is the only approved
44 vaccine against tuberculosis, which efficacy in adults has been questioned (Black et al. 2002).
45 Considering all these reasons mentioned above, we believed that it would be very worthy to
46 identify HLA-A2 restricted cytotoxic T lymphocyte epitopes derived from drug efflux pump
47 antigens of Mtb. It would help us to develop effective subunit vaccines against TB, especially
48 XDR and MDR strains.

49 Classic ABC family is the most well-known efflux pump responsible for intrinsic drug
50 resistance. It was shown that the over-expression of Rv2686c-Rv2687c-Rv2688c in *M.*

51 *smegmatis* increased the minimum inhibitory concentrations of ciprofloxacin (Pasca et al.
52 2004). And Rv2937 (drrB) behaved as a functional efflux pump referring to rifampin,
53 tetracycline and erythromycin together with drrA (Choudhuri et al. 2002). Because that DrrA
54 and Rv2688c have homology with some proteins in human, Rv2937, Rv2686c, and Rv2687c
55 were chosen as target antigens to identify HLA-A2 restricted cytotoxic T lymphocyte
56 epitopes.

57 Eight potential peptides derived from Rv2937, Rv2686c, and Rv2687c were predicted by
58 using the online tools SYFPEITHI, BIMAS, and NetCTL. These peptides were synthesized
59 and their ability to induce immune response was tested both in PBMCs of HLA-A2⁺ donors
60 (*in vitro*) and HLA-A2/K^b transgenic mice (*in vivo*).

61

62 **MATERIALS AND METHODS**

63 **Prediction and synthesis of candidate peptides**

64 By using epitope prediction tools, BIMAS (http://bimas.dcrf.nih.gov/molbio/hla_bind/)
65 (Parker et al. 1994), SYFPEITHI
66 (<http://www.syfpeithi.de/Scripts/MHCServer.dll/EpitopePrediction.htm>) (Rammensee et al.
67 1999), and NetCTL (<http://www.cbs.dtu.dk/services/NetCTL/>) (Larsen et al. 2007), potential
68 HLA-A2-restricted T cell epitopes derived from Rv2937, Rv2686c, and Rv2687c were
69 predicted. Peptides with relative high scores were synthesized by using standard solid phase
70 Fmoc strategy. The peptides were purified by reverse phase-high performance liquid
71 chromatography (RP-HPLC), and then their molecular weights were confirmed by
72 electrospray ionization-mass spectrometry (ESI-MS). The HBV core antigen-derived T helper
73 epitope (sequence128–140: TPPAYRPPNAPIL) was used to enhance the immune activity in
74 the mice vaccination experiment (Milich et al. 1988; Vissers et al. 1999).

75

76 **Blood samples, animals and cell lines**

77 Whole blood was prepared from HLA-A2⁺ (PPD⁺ or PPD⁻) healthy donors. The
78 HLA-A2.1/K^b transgenic mice were previously gifted by professor Xue-tao Cao (Second
79 Military Medical University) (Vitiello et al. 1991). All mice at 8 to 12 weeks in the
80 experiments were housed in a specific pathogen-free environment in our laboratory. The

81 sample collection from healthy donors and animal experiments were approved by the Ethics
82 Committee of Zhengzhou University (No. 20120312). The human transporter associated with
83 antigen processing (TAP) -deficient T2 cell line was kindly provided by professor Yu-zhang
84 Wu (Third Military Medical University, China), and the cells were cultured in RPMI 1640
85 medium supplemented with 10% fetal bovine serum (FBS) and maintained at 37°C in an
86 incubator with a humidified atmosphere containing 5% CO₂.

87

88 **Generation of CTLs from HLA-A2 healthy donors**

89 The procedures of generation of CTLs *in vitro* were performed in accordance with the
90 protocols described by our laboratory (Liu et al. 2012; Shi et al. 2013). Briefly, PBMCs were
91 isolated from six HLA-A2⁺ PPD⁺ and an HLA-A2⁺ PPD⁻ healthy donors with centrifugation
92 at a Ficoll-Paque density gradient and then cultured in IMDM medium supplemented with
93 10% FBS under the condition of 37°C, 5% CO₂ (Han et al. 2006). After 24h, PBMCs at the
94 concentration of 1×10⁶/ml were stimulated with the candidate peptides (10μg/ml) in the
95 presence of 3μg/ml β2-m for 4h. The next day, human recombinant IL-2 (50 U/ml) and IL-7
96 (50 U/ml) were added. Once a week, these cells were re-stimulated same as the procedures
97 above. Seven days after the third round of stimulation, the cytotoxic assay and ELISPOT
98 assay were performed.

99

100 **Generation of CTLs from HLA-A2.1/K^b transgenic mice**

101 CTLs from HLA-A2.1/K^b transgenic mice were generated as previously described (Liu et
102 al. 2012; Shi et al. 2013). Briefly, HLA-A2.1/K^b transgenic mice were grouped randomly, they
103 were injected subcutaneously at the base of the tail with 100μg each peptide emulsified in
104 incomplete Freund's adjuvant (IFA) in the presence of 140μg of the T helper epitope every
105 five days (Eguchi et al. 2006; Tourdou et al. 2000). On day 11, splenocytes of each mouse
106 were separated and then re-stimulated with the corresponding peptide (10μg/ml) *in vitro* for
107 another five days. Then, the LDH cytotoxicity and ELISPOT assays were employed.

108

109 **ELISPOT assay**

110 ELISPOT assay was performed according to the instruction of the commercial kit (Dakewe,
111 China). Peptide-pulsed T2 cells (stimulator cells, 1×10⁵), along with the induced CTLs
112 (effector cells, 1×10⁵), were seeded into an anti-human (or anti-mouse) IFN-γ antibody coated
113 96-well plate (Ding et al. 2009). After incubation for 16 h at 37°C, cells were removed and
114 plates were processed. Spots were counted with a computer-assisted spot analyzer (Dakewe,

115 China).

116

117 **Cytotoxicity assay**

118 Cytotoxic activity was tested by the non-radioactive cytotoxicity assay kit (Promega, US)
119 at gradient E:T ratio according to the manufacturer's instruction. T2 cells were loaded with
120 10µg/ml peptide for 1h at 37°C as target cells. The effector cells were co-cultured with target
121 cells (1×10^4 /well) at various effector/target ratios for 5h at 37°C under 5% CO₂. The
122 percentage of specific lysis of the target cells was determined according to the following
123 formula. Percentage of specific lysis = [(experimental release – effector spontaneous release –
124 target spontaneous release) / (target maximum release – target spontaneous release)] × 100.

125

126 **Statistical analysis**

127 All data were presented as means ± S.D. Comparisons between experimental groups and
128 relevant controls were analyzed by Student's t test. $P < 0.05$ was considered as a statistically
129 significant difference.

130

131 **RESULTS**

132 **Peptides selected as potential CTL epitopes**

133 By using the on-line prediction tools, eight potential HLA-A2 restricted T cell epitopes
134 were selected from the three candidate efflux pump antigens, Rv2937, Rv2686c, and Rv2687c.
135 The peptides were synthesized and their molecular weights were confirmed by ESI-MS.
136 (Table 1).

137

138 **IFN-γ release ELISPOT assay *in vitro***

139 CTLs were induced from the PBMCs of six HLA-A2⁺ PPD⁺ and an HLA-A2⁺ PPD⁻ healthy
140 donors. IFN-γ release ELISPOT assay was employed to test the capacity of the eight peptides
141 to induce CTL response. As shown in Fig. 1, among the six HLA-A2⁺ PPD⁺ donors,
142 Rv2937-p168, Rv2937-p266, Rv2686c-p151, Rv2686c-p181, and Rv2686c-p184 could
143 induce more frequent and potent CTL response, while all these peptides could only induce
144 very weak response in the HLA-A2⁺ PPD⁻ donor (Fig. S1).

145

146 ***In vitro* cytotoxic activity of peptide-specific CTLs**

147 Based on the results of the ELISPOT assay, Rv2937-p168, Rv2937-p266, Rv2686c-p151,
148 Rv2686c-p181, and Rv2686c-p184 were selected to investigate whether the specific T cells

149 they induced could lyse target cells. In the cytotoxicity assay, peptide-pulsed T2 cells were
150 considered as target cells, the effector cells were obtained from CTLs induced from PBMCs
151 of HLA-A2⁺ PPD⁺ donor. As shown in Fig. 2, the specific lysis percentages of the CTLs
152 induced by Rv2937-p168, Rv2937-p266, Rv2686c-p151, Rv2686c-p181, and Rv2686c-p184
153 from PBMCs of HLA-A2⁺ PPD⁺ healthy donors were increased gradiently from E/T ratio
154 12.5:1 to 50:1. However, after incubating with anti-HLA-A2 monoclonal antibody, the
155 specific lysis rates of the CTLs derived from HLA-A2⁺ PPD⁺ were greatly reduced. These
156 results indicated that these peptides could induce HLA-A2-restricted CTL response in PPD⁺
157 healthy donor.

158

159 **Cytotoxic T lymphocyte response in HLA-A2.1/K^b transgenic mice**

160 HLA-A2.1/K^b transgenic mice immunization model is a widely used putative model to
161 study the *in vivo* CTL activity of HLA-A2-restricted epitopes. Since Rv2937-p168,
162 Rv2937-p266, Rv2686c-p151, Rv2686c-p181, and Rv2686c-p184 could induce good immune
163 response *in vitro*, we then investigated whether these peptides could stimulate CTL response
164 in HLA-A2.1/K^b transgenic mice. After immunization of the candidate peptides with Th
165 epitope and IFA adjuvant, the splenocytes were isolated and stimulated with the
166 corresponding peptide *in vitro* for another five days. IFN- γ release ELISPOT assay and LDH
167 cytotoxicity assay were performed. As shown in Fig. 3, all of the five peptides showed more
168 potent activity to induce IFN- γ release than the negative control group.

169 The cytotoxic activity of these splenocytes was also measured by an LDH cytotoxicity
170 assay. Peptide-loaded T2 cells were served as target cells and the effector/target ratios were
171 20:1, 40:1, and 80:1. As shown in Fig. 4, at the E:T ratio of 80:1, the CTLs induced by
172 Rv2937-p168, Rv2937-p266, Rv2686c-p151, and Rv2686c-p181 could significantly kill the
173 target cells. To our surprise, although Rv2686c-p184 could induce the most potent IFN- γ
174 release activity in PBMCs of HLA-A2⁺ PPD⁺ donors, it could not induce CTLs with killing
175 effects in HLA-A2.1/K^b transgenic mice.

176

177 **DISCUSSION**

178 Tuberculosis is considered as a major public health concern worldwide because of the high
179 mortality and morbidity associated with it. And as reported, the emergence of XDR-TB and
180 MDR-TB forced patients having fewer options for treatment and risking higher mortalities,
181 especially in HIV/TB co-infected ones (Principi & Esposito 2015). Development of novel
182 therapeutic vaccines provides a promising strategy to combine with chemotherapy.

6

183 Accumulating data showed that subunit vaccine containing peptide or protein antigens could
184 exhibit protective activity and very good safety (Ivanyi 2014).

185 HLA-A2 is one of the most common supertypes of human leukocyte antigen particularly in
186 Asian with an estimated frequency of nearly 50% (Mehra et al. 2001), and cell-mediated
187 immunity to Mtb is believed important to control the latent Mtb infection, therefore,
188 identification of HLA-A2 restricted cytotoxic T lymphocyte epitopes derived from efflux
189 pump antigens could be helpful to develop vaccines against drug-resistant TB caused by
190 intrinsic drug efflux. Recently, most researchers who hammer at identification of CTL
191 epitopes of Mtb antigens focused their work on secretory protein, such as ESAT-6(Lalvani et
192 al. 1998) , 19-kDa lipoprotein(Mohaghehpour et al. 1998), Ag85B(Geluk et al. 2000),
193 16-kDa antigen(Caccamo et al. 2002) and MPT51(Suzuki et al. 2004), and so on. We also
194 identified such kind of epitopes from antigen CFP21 and RD region (Chen et al. 2012; Lv et
195 al. 2010). Furthermore, we firstly reported that efflux pumps could also be considered as
196 target antigens for TB immunotherapy and found that Rv1410c could serve as a candidate to
197 the vaccine design against drug-resistant Mtb (Zhu et al. 2011). Then we screened classical
198 efflux pump family members in the genome of TB to find more promising target antigens. As
199 shown in the present study, we found that epitopes, Rv2937-p168, Rv2937-p266,
200 Rv2937-p168, Rv2686c-p181, and Rv2686c-p184 identified from ABC family members,
201 elicited good capacity to induce CTL response in HLA-A2⁺ PPD⁺ healthy donors and/or
202 HLA-A2.1/K^b transgenic mice. Work is still need to be done to identify more promising
203 antigens related to drug-resistant Mtb. We are now also working on extrinsic in TB. Hopefully,
204 we can combine all these epitopes derived from secretory and drug-resistant antigens to
205 develop multi-valent subunit vaccines.

206 As we all known, early diagnosis of TB is fundamental for tuberculosis control. In the last
207 decade, ELISPOT assay in the TB diagnosis is considered to have high specificity and
208 sensitivity (Lalvani & Pareek 2010; Milotic et al. 2011). Most of the five candidate peptides
209 showed good activity among the six HLA-A2⁺ PPD⁺ healthy donors, but not in HLA-A2⁺
210 PPD⁻. Although we do not know whether these PPD⁺ donors have drug-resistant Mtb infection,
211 the results indicated that these epitopes might also be used as TB diagnosis.

212 In the present study, we used so called 'reversal immunology' strategy to predict antigen
213 epitopes by using on-line tools instead of time-consuming overlapping peptides method,
214 which was very efficient. However, using just one computational algorithm to predict CTL
215 epitopes may lead to large amounts of false positive and false negatives. So we used epitope
216 prediction tool NetCTL combined with the widely used BIMAS and SYFPEITHI databases.

217 NetCTL integrates prediction of binding affinity, transporter of antigenic peptide efficiency
218 and proteasomal cleavage (Larsen et al. 2007), SYFPEITHI is a motif-matrix-based
219 prediction method for MHC binding prediction (Rammensee et al. 1999) and BIMAS is based
220 on peptide/MHC complex half-life(Parker et al. 1994). Our results suggested that this strategy
221 could be very efficient and successful.

222

223 CONCLUSIONS

224 In conclusion, we have identified five HLA-A2 restricted cytotoxic T lymphocyte epitopes
225 derived from drug efflux pump antigens of *Mycobacterium tuberculosis*. The epitopes
226 Rv2937-p168, Rv2937-p266, Rv2686c-p151, Rv2686c-p181, and Rv2686c-p184 showed
227 good capacity to induce CTL response in HLA-A2⁺ PPD⁺ donors. Except Rv2686c-p184,
228 other epitopes could also elicit CTL response when immunized in HLA-A2.1/K^b transgenic
229 mice. These epitopes could serve as candidates for TB diagnosis and immunotherapy.

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231

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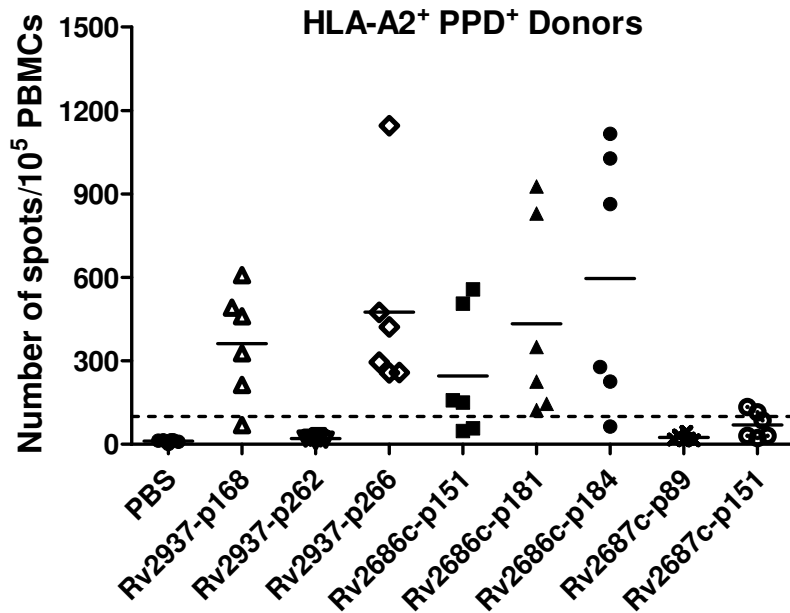
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Figure 1. IFN- γ release ELISPOT assay by CTLs induced from PBMCs of HLA-A2⁺ PPD⁺ donors. Each peptide was tested by using the samples from the same group of six donors. PBMCs from healthy donors were separated and stimulated once a week with synthetic peptides and IL-2 for three rounds.

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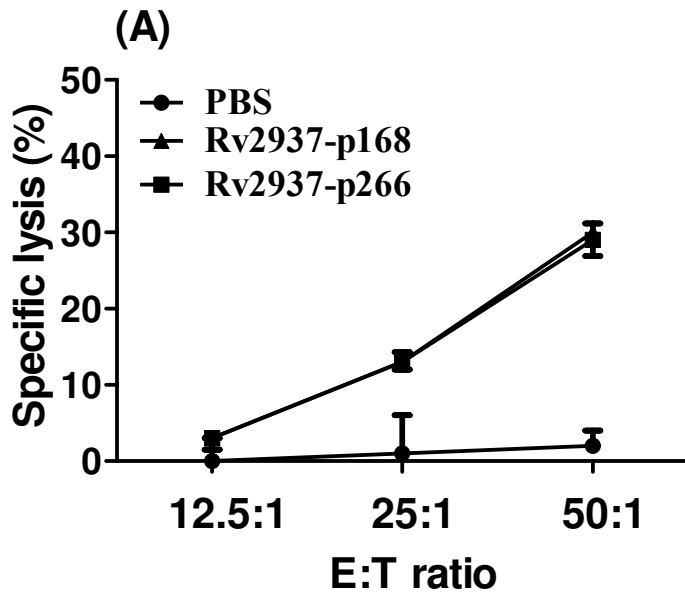
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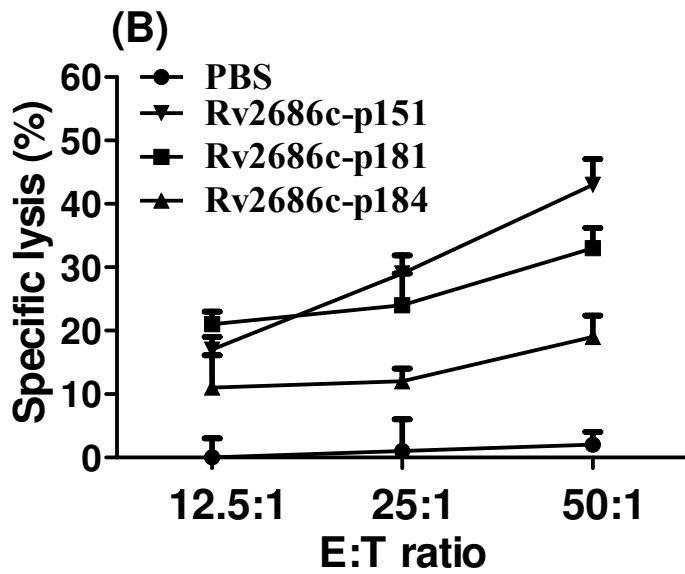
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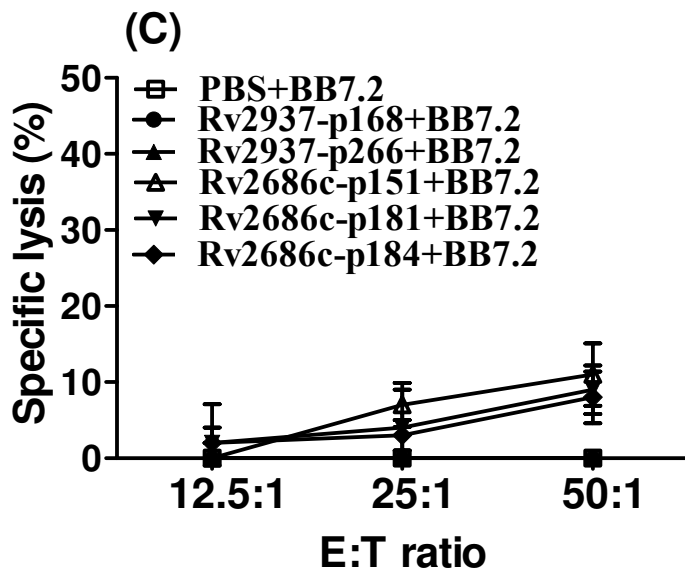
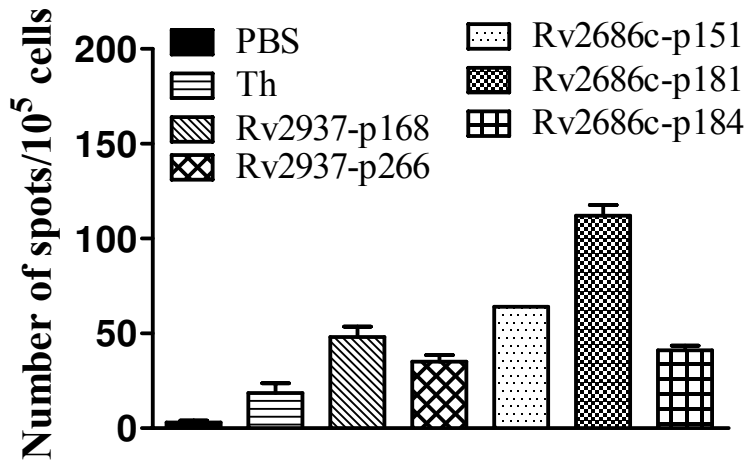


Figure 2. Specific lysis of T2 cells loaded with synthetic peptides by the CTLs generated from PBMCs of HLA-A2⁺ PPD⁺ donors. (A-B) The effector cells were obtained from CTLs induced from PBMCs of HLA-A2⁺ PPD⁺ donors. (C) The HLA-A2 molecules on the T2 cells surface were blocked by anti-HLA-A2 monoclonal antibody.

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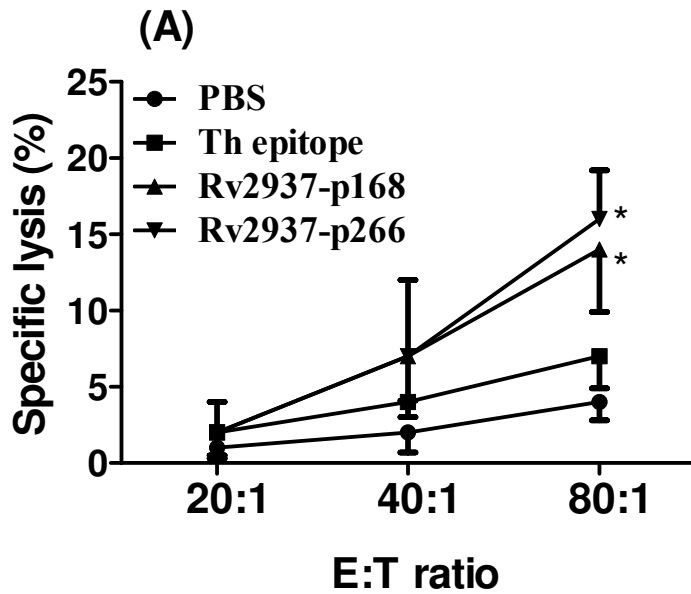
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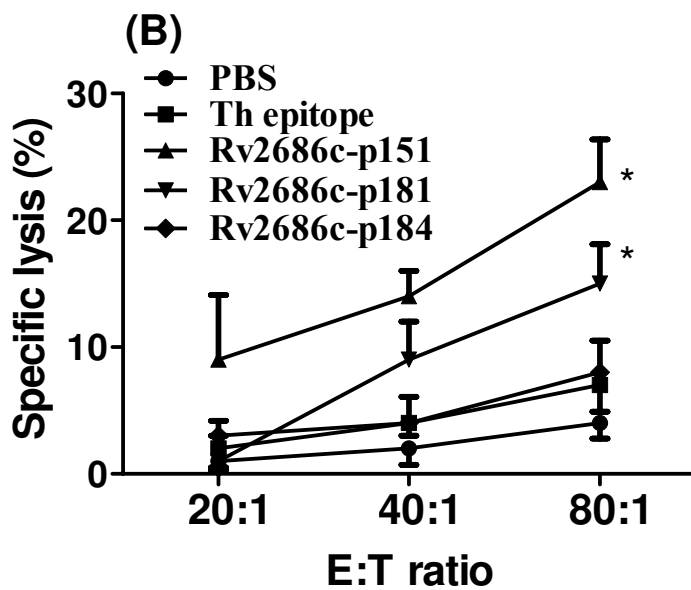
Figure 3. IFN- γ release ELISPOT assay of the splenocytes from the immunized HLA-A2.1/K^b transgenic mice. HLA-A2.1/K^b transgenic mice were immunized with 100 μ g each peptide emulsified in incomplete Freund's adjuvant (IFA) in the presence of 140 μ g of the T helper epitope every five days for three times.



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417 Figure 4. Specific lysis of T2 cells loaded with synthetic peptides by the CTLs generated from
 418 the immunized HLA-A2.1/K^b transgenic mice ($n=5$). The data from each peptide immunized
 419 group were compared with the Th epitope alone negative control.

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424 Table 1. HLA-A2 CTL epitopes predicted from efflux pumps of *Mycobacterium tuberculosis*

Antigen	Position	Sequence	SYFPEITHI	NetCTL
Rv2937	168	YIVGFCLLV	24	1.2629
	262	VMAPLTWL	27	1.2098
	266	TLTWLFAFV	22	1.1149
Rv2686c	151	GLVAGLSAV	28	0.9644
	181	ALGMLIAGL	30	0.9898
	184	MLIAGLPCL	29	1.3358
Rv2687c	89	YLAAKLTVL	29	1.3019
	151	FLAAVIPLA	22	1.2362

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