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# Impact of copper toxicity on stone-head cabbage (*Brassica oleracea* var. *capitata*) in hydroponics

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Arable soils are frequently subjected to contamination with copper as the consequence of imbalanced fertilization with manure and organic fertilizers and/or extensive use of copper-containing fungicides. In the present study, the exposure of Stone-head cabbage (*Brassica oleracea* Var. *capitata*) to elevated  $\text{Cu}^{2+}$  levels resulted in leaf chlorosis and lesser biomass yield at  $\geq 2 \mu\text{M}$ . Root nitrate content was not statistically affected by  $\text{Cu}^{2+}$  levels, although it was substantially decreased at  $\geq 5 \mu\text{M}$   $\text{Cu}^{2+}$  in the shoot. The decrease in nitrate contents can be related to lower nitrate uptake rates because of growth inhibition by Cu-toxicity. Shoot sulfate content increased strongly at  $\geq 2 \mu\text{M}$   $\text{Cu}^{2+}$  indicating an increase in demand for sulfur under Cu stress. Furthermore, at  $\geq 2 \mu\text{M}$  concentration, concentration of water-soluble non-protein thiol increased markedly in the roots and to a smaller level in the shoot. When exposed to elevated concentrations of  $\text{Cu}^{2+}$  the improved sulfate and water-soluble non-protein thiols need further studies for the evaluation of their direct relation with the synthesis of metal-chelating compounds (i.e. phytochelatins).

# IMPACT OF COPPER TOXICITY ON STONE-HEAD CABBAGE (*BRASSICA OLERACEA* VAR. *CAPITATA*) IN HYDROPONICS

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## INTRODUCTION

Transition metals such as copper (Cu), zinc (Zn) and molybdenum (Mo) are essential for the growth and development of plants, but they rapidly get toxic at higher levels (Kopsell and Kopsell, 2007). Cu contamination in agricultural soils as a consequence of mining metals, dispersal of sewage sludge, arbitrary and improper application of agrochemicals, addition of organic fertilizers and frequent use of irrigation with low quality water is a well-known problem (Dach and Starmans, 2005; Yruela, 2009). Cu being redox active metal can exist in both  $\text{Cu}^{2+}$  and  $\text{Cu}^+$  forms in living organisms. At the protein level, Cu serves as a co-factor for various enzymes such as Cu/Zn-superoxide dismutase (Cu/ZnSOD), cytochrome c oxidase, ascorbate oxidase, amino oxidase, laccase, plastocyanin (PC), and polyphenol oxidase (Yruela, 2005; 2009; Pilon et al., 2006). However, redox cycling between  $\text{Cu}^{2+}$  and  $\text{Cu}^+$  could induce oxidative stress by producing highly toxic hydroxyl radicals (Yruela, 2005, 2009).

Plants exposed to elevated levels of Cu show unspecific toxicity symptoms. Elevated Cu levels in soils primarily result in stunted root growth and leaf chlorosis (Kopsell and Kopsell, 2007; Shahbaz et al., 2010a). Copper toxicity-induced reduction in chlorophyll contents hinder the development of chloroplast, thalokoid membrane and photosystem II (PSII), which are considered as the most sensitive Cu toxicity sites (Caspi et al., 1999; Pätsikkä et al., 1998, 2002; Burkhead et al., 2009; Yruela, 2005, 2009; Shahbaz et al., 2010). At cellular level, toxicity may lead to binding of sulfhydryl groups in proteins, insufficiency or excess of other essential ions, oxidative damage and reduced cell transport (De Vos et al., 1993; Yruela, 2009). Furthermore, Cu-toxicity can change the mineral composition of plants i.e. for instance, Fe contents may decrease in the shoot (Pätsikkä et al., 1998; 2002; Kopsell and Kopsell, 2007; Shahbaz et al., 2010b), Ca and Mg may decrease in the root and Zn contents may increase in both root and shoot upon Cu exposure at elevated levels (Shahbaz et al., 2010b).

Root growth is more severely affected by elevated Cu than shoot growth and the major proportion of Cu uptake retains in the root. Increased Cu contents in the plant tissues induce the synthesis of metal-binding compounds (*viz.* phytochelatins), which are most likely glutathione-derived compounds. (Inouhe, 2005; Ernst et al., 2008). Induction of phytochelatins presume that more sulfur is needed for synthesis of these compounds which results into higher absorption and incorporation of sulfate. Nonetheless, the role of phytochelatins in detoxification of Cu still is not very clear (Ernst et al., 2008; Yruela, 2005; 2009; Shahbaz et al., 2010a).

The present study used the hydroponics system which allows very close control over water soluble Cu and other mineral concentrations as compared to soil-grown system. Cabbage is a very important vegetable in all over the world. In Pakistan, cabbage is cultivated on almost 4.4 thousand hectares with 69 thousand tonnes annual production (Annonymus, 2005). The present study was aimed to investigate the response of growth, pigment contents and sulfur metabolism of Stone-head cabbage grown in hydroponics to the copper exposure.

## **MATERIAL AND METHODS**

Stone-head cabbage (*Brassica Oleracea*) Var. Capitata F<sub>1</sub> seeds were germinated on sand in a green house. The seedlings collected at ten days after germination were grown on a 25% Hoagland nutrient solution having a pH 5.9-6.0, 1.25 mM Ca(NO<sub>3</sub>)<sub>2</sub>·4H<sub>2</sub>O, 1.25 mM KNO<sub>3</sub>, 0.25 mM KH<sub>2</sub>PO<sub>4</sub>, 0 / 0.5 mM MgSO<sub>4</sub>·7H<sub>2</sub>O, 11.6 μM H<sub>3</sub>BO<sub>3</sub>, 2.4 μM MnCl<sub>2</sub>·4H<sub>2</sub>O, 0.24 μM ZnSO<sub>4</sub>·7H<sub>2</sub>O, 0.08 μM CuSO<sub>4</sub>·5H<sub>2</sub>O, 0.13 μM Na<sub>2</sub>MoO<sub>4</sub>·2H<sub>2</sub>O and 22.5 μM Fe<sup>3+</sup>-EDTA supplemental concentrations of 0, 2, 5 and 10 μM CuCl<sub>2</sub> (3 plants set<sup>-1</sup> and 12 sets container<sup>-1</sup>) in a greenhouse for 10 days. The photoperiod was 14 h. 30 and 25°C (±5°C) temperatures were set for day and night respectively, whereas the relative humidity was maintained at 60-70%.

Whole shoot was homogenized (in 100% acetone @ 10 mL per g FW) followed by centrifugation at 800 g for 20 minutes. Lichtenthaler (1987) was followed for the determination of chlorophyll *a*, *b* and total carotenoid contents.

### **Nitrate and sulfate contents**

Frozen root and shoot material was homogenized in de-mineralized water (10 mL per g fresh weight) and one layer of Miracloth filter was used to filter the homogenate. The supernatant was incubated in a water bath at 100°C for 10 min. The remainder was centrifuged for 15 min (0°C) at 30,000 g. The anions were separated by HPLC and Maas et al. (1986) was followed for their refractometric determination.

### **Water-soluble non-protein thiols**

Extraction medium containing 80 mM sulfosalicylic acid, 1 mM EDTA, and 0.15% (w/v) ascorbic acid with an Ultra Turrax at 0°C (10 mL per g fresh weight) was used for the homogenization of fresh plant matter. The resultant homogenous material was passed through one layer of Miracloth which was then centrifuged at 30,000 g for 15 min (0°C). De Kok et al. (1988) was followed for the determination of total water-soluble non-protein thiol content colorimetrically at 413 nm after reaction with 5, 5'-dithiobis [2-nitrobenzoic acid].

## RESULTS

### Plant biomass in response to Cu exposure

Exposure of Stone-head cabbage to higher concentrations of  $\text{Cu}^{2+}$  ( $\geq 2 \mu\text{M}$ ) in nutrient solution caused the chlorosis of both the shoot and young emerging leaves, that ultimately reduced both root and shoot biomass production (Fig. 1). A 10 day exposure to increasing  $\text{Cu}^{2+}$  concentrations in nutrient media led to a significant reduction of both root and shoot biomass production at  $\geq 2 \mu\text{M}$   $\text{Cu}^{2+}$ . Shoot to root ratio improved at  $\geq 5 \mu\text{M}$   $\text{Cu}^{2+}$ , demonstrating that when exposed to copper, root growth was more affected than that of shoot growth (Fig. 1). Root dry matter content increased at  $10 \mu\text{M}$   $\text{Cu}^{2+}$ , whereas that of shoot increased at  $\geq 5 \mu\text{M}$   $\text{Cu}^{2+}$  (Fig. 2).

### Pigment content in response to Cu exposure

The total chlorophyll (Chl. *a+b*) and carotenoids contents of Stone-head cabbage were significantly decreased upon exposure at  $\geq 2 \mu\text{M}$   $\text{Cu}^{2+}$  (Fig. 3). There were significant decreases in chlorophyll *a/b* and chlorophyll/carotenoid ratios when exposed to increased  $\text{Cu}^{2+}$  concentrations ( $10 \mu\text{M}$   $\text{Cu}^{2+}$ ).  $10 \mu\text{M}$   $\text{Cu}^{2+}$  exposure resulted in the start of rapid development of shoot chlorosis and significantly faster reduction in chlorophyll *a* contents than that of chlorophyll *b* and carotenoids, ultimately leading to a significant reduction in chlorophyll *a/b* and chlorophyll/carotenoid ratios (Fig. 3).

### Sulfate and water-soluble non-protein thiol contents in response to Cu exposure

Elevated  $\text{Cu}^{2+}$  levels showed a significant effect on concentration of the nitrate, sulfate and water-soluble non-protein thiol in stone-head cabbage. The nitrate contents of the roots showed a non significant response to the Cu exposure at different levels, however in shoots it were significantly decreased at  $\geq 5 \mu\text{M}$   $\text{Cu}^{2+}$  (Fig. 4). Sulfate contents in the roots were not affected; however  $\text{Cu}^{2+}$  treatments of  $\geq 2 \mu\text{M}$  substantially increased the sulfate contents of the shoot (Fig. 4). There was slight decrease in nitrate to sulfate ratio in root and a strong decrease in the shoot when exposed to elevated levels of Cu (Fig. 4). Furthermore, the exposure to  $\geq 2 \mu\text{M}$   $\text{Cu}^{2+}$  resulted in a solid raise in water-soluble non-protein thiol contents in the roots and to a smaller degree in the shoots at  $10 \mu\text{M}$   $\text{Cu}^{2+}$  (Fig. 5).

## DISCUSSION

Cu exposure at elevated levels ( $< 2 \mu\text{M}$   $\text{Cu}^{2+}$ ) to Stone-head cabbage significantly decreased the production of root and shoot biomass and raised the ratio of the shoot to the root.

Copper contamination in the root environment generally results in retarded production of root and shoot biomass and a reduced photosynthetic activity. Moreover, it causes chlorosis, necrosis and bleaching of pigments (Yruela, 2005, 2009; Sheldon and Menzies, 2005; Shahbaz et al. 2010a, b). In cabbage, the reduced production of biomass when exposed to elevated Cu levels coincided with decreased pigment contents (chl a, b, carotenoids; Fig. 3) which may have resulted in reduced activity of photosynthesis and the dark respiration rate (Shahbaz et al., 2010a). It has been shown that Cu-toxicity damages chloroplasts either by inducing iron deficiency or by replacing Mg in the chlorophyll by Cu (Pätsikkä et al., 2002; Küpper et al., 2003).

It is shown that the production of root biomass was more influenced than that of the shoot biomass production. The relatively higher reduction in the root biomass upon exposure to metal contamination could be due the fact that roots come in direct contact with toxic metals (Cd, Cu). Toxic metal-induced hindered root growth also reduces the uptake of essential nutrients (Sheldon and Menzies, 2005).

Plants have evolved a tightly-controlled mechanism for the absorption, allocation and assimilation of sulfate under normal conditions. (Hawkesford and De Kok, 2006). Enhanced introduction of cabbage to Cu concentrations considerably affects the contents and allocation of sulfur compounds in the root and shoot of cabbage (Shahbaz et al., 2010a). The raised shoot sulfate contents might be attributed to Cu-toxicity induced upregulation of the sulfate suppliers in roots (Shahbaz et al., 2010a). To maintain rapid growth rates under stress conditions, *Brassica* species increase their demand for sulfur supply (Koralewska et al., 2008; 2009). Disclosure of Stone-head cabbage to elevated levels of Cu did not affect the nitrate contents in roots, however at  $\geq 5 \mu\text{M Cu}^{2+}$  there was a significant decrease in the shoot nitrate contents. The reduced production of plant biomass at  $\geq 5 \mu\text{M Cu}^{2+}$  could be attributed to reduced supply of nitrate in the shoot. The decrease in nitrate: sulfate ratio in the shoot of Stone-head cabbage under elevated Cu levels may be attributed to enhanced sulfate contents, however the link between uptake rates of nitrate and sulfate is not evident yet (Stulen and De Kok, 2012). Since both nitrate and sulfate are involved in amino acid and protein synthesis, their uptake rates are related with growth rates (Stulen and De Kok, 2012).

Roots accumulated a slightly raised level of water-soluble non-protein thiols when compared with that of the shoot. Only a small proportion of the rise in thiol content might be

attributed to a Cu-induced synthesis of phytochelatins in Chinese cabbage (Shahbaz et al., 2010a). Previous reports suggest that the formation of sulfur-rich metal-chelating compounds (i.e. water-soluble non-protein thiols) when disclosed to prospective toxic metals may perhaps require an enhanced demand for sulfur, viz. the absorption rate and incorporation of sulfate (Sirko and Gotor, 2007; Ernst et al., 2008). The possible significance of sulfur nutrition upon elevated copper exposure needs be further investigated.

## CONCLUSIONS

This investigation has shown that elevated copper intensities in the root surroundings proved toxic for Stone-head cabbage. Copper exposure at  $\geq 2 \mu\text{M Cu}^{2+}$  negatively affected the plant biomass production and pigment contents. Furthermore, elevated Cu content considerably affected the concentration of sulfate and water-soluble non-protein thiol of Stone-head cabbage which might be due to the induction of phytochelatins to detoxify excess copper.

## REFERENCES

- Burkhead, J., K.A.G. Reynolds, S.E. Abdel-Ghany, C. M. Cohu and M. Pilon. 2009. Copper homeostasis. *New Phytol.* 182(4):799-816.
- Dach, J. and D. Starmans. 2005. Heavy metals balance in Polish and Dutch agronomy: Actual state and provisions for the future. *Agric. Ecosys. Environ.* 107: 309-316.
- Kok, D., F. Buwalda and W. Bosma. 1988. Determination of cysteine and its accumulation in spinach leaf tissue upon exposure to excess sulfur. *J. Pl. Physiol.* 133(4): 502-505.
- De Vos, C.H.R., T. M. Ten Bookum, R. Vooijs, H. Schat and L.D. De Kok. 1993. Effect of copper on fatty acid composition and peroxidation of lipids in roots of copper tolerant and sensitive *Silenecucubalus*. *Pl. Physiol. Biochem.* 31(2): 151-158.
- Ernst, W.H.O., G.J. Krauss, G. A. C. Verkleij and D. Wesenberg. 2008. Interaction of heavy metals with the sulphur metabolism in angiosperms from an ecological point of view. *Pl. Cell Environ.* 31(1): 123-143.
- Hawkesford, M.J. and L.J. De Kok. 2006. Managing sulfur metabolism in plants. *Pl. Cell Environ.* 29(3): 382-395.



- Inouhe, M. 2005. Phytochelatins. *Braz. J. Pl. Physiol.* 17(1): 65-78.
- Kopsell, D. E. and D.A. Kopsell. 2007. Copper. In: Barker, A.V., Pilbeam, D.J. (Eds.), *Handbook of Plant Nutrition*. Taylor and Francis Group, Boca Raton, p.p. 293-328.
- Küpper, H., E. Šetlík, N. Šetliková, M. Ferimazova and F.C. Küpper. 2003. Copper-induced inhibition of photosynthesis: limiting steps of *in vivo* copper chlorophyll formation in *Scenedesmusquadricauda*. *Funct. Pl. Bio.* 30(12): 1187-1196.
- Lichtenthaler, H.K. 1987. Chlorophylls and carotenoids: pigments of the photosynthetic biomembranes. *Methods Enzymol.* 148: 350-382.
- PätsikkäE, M., F. Kairavuo, E.M. Šerešen and E. Tyystjärvi. 2002. Excess copper predisposes photosystem II to photoinhibition *in vivo* by outcompeting iron and causing decrease in leaf chlorophyll. *Pl. Physiol.* 129(3): 1359-1367.
- Pätsikkä, E., E.M. Aroan and E. Tyystjärvi. 1998. Increase in the quantum yield of photoinhibition contributes to copper toxicity *in vivo*. *Pl. Physiol.* 117(2): 619-627.
- Pilon, M., S.E. Abdel-Ghany, C.M. Cohu, K.A. Gogolin and H. Ye. 2006. Copper cofactor delivery in plant cells. *Curr. Opin. Pl. Bio.* 9: 256-263.
- Shahbaz, M., M.H. Tseng, C.E.E. Stuiver, A. Koralewska, F.S. Posthumus, J.H. Venema, S. Parmar, M.J. Hawkesfordans and L.J. De Kok. 2010a. Copper exposure interferes with the regulation of the uptake, distribution and metabolism of sulfate in Chinese cabbage. *J. Pl. Physiol.* 167(6): 438-446.
- Shahbaz, M., M.H. Tseng, C.E.E. Stuiver, F.S. Posthumus, S. Parmar, A. Koralewska, M.J. Hawkesford and L.J. De Kok. 2010b. Impact of copper exposure on physiological functioning of Chinese cabbage (*Brassica pekinensis*). In: *More Sustainability in Agriculture: New Fertilizers and Fertilizer Management* (Sequi, P., Ferri, D., Rea, E.,

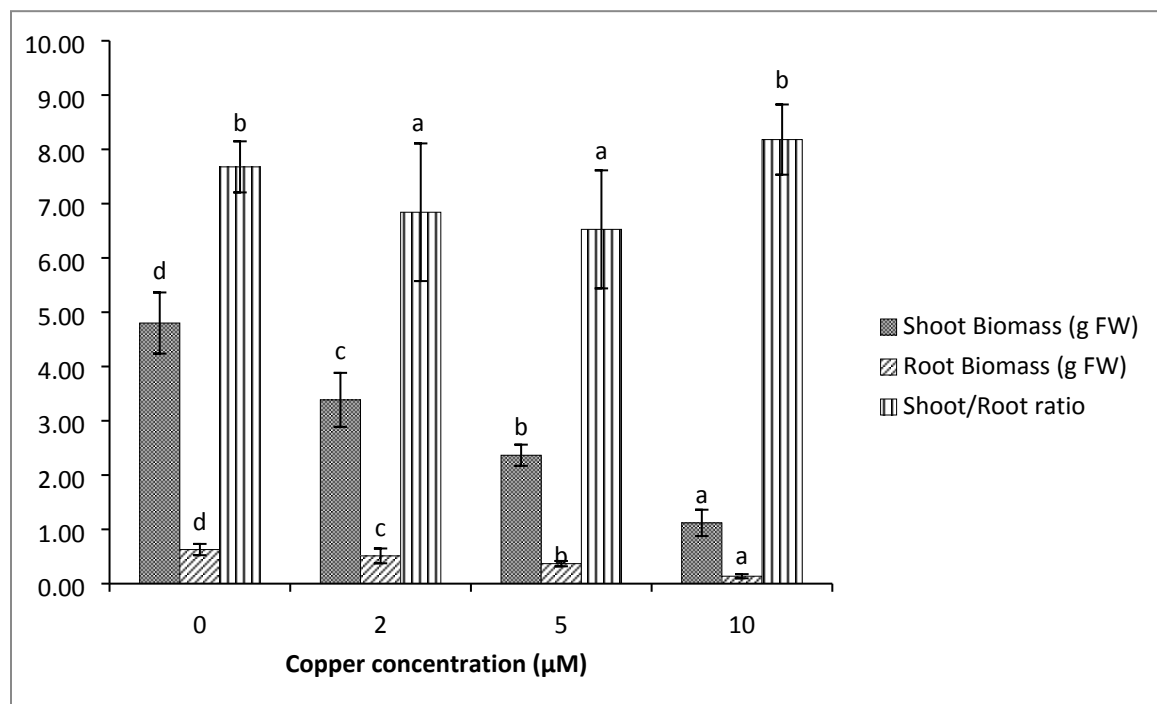
Montemurro, A.V. and Fornado, F. eds.), 18th International Symposium of CIEC, Fertilitas Agrorum, pp. 318-324.

Sheldon, A. and N.W. Menzies. 2005. The effect of copper toxicity on growth and morphology of Rhodes grass (*Chlorisgayana*) in solution culture. Pl. Soil 278(1-2): 341-349.

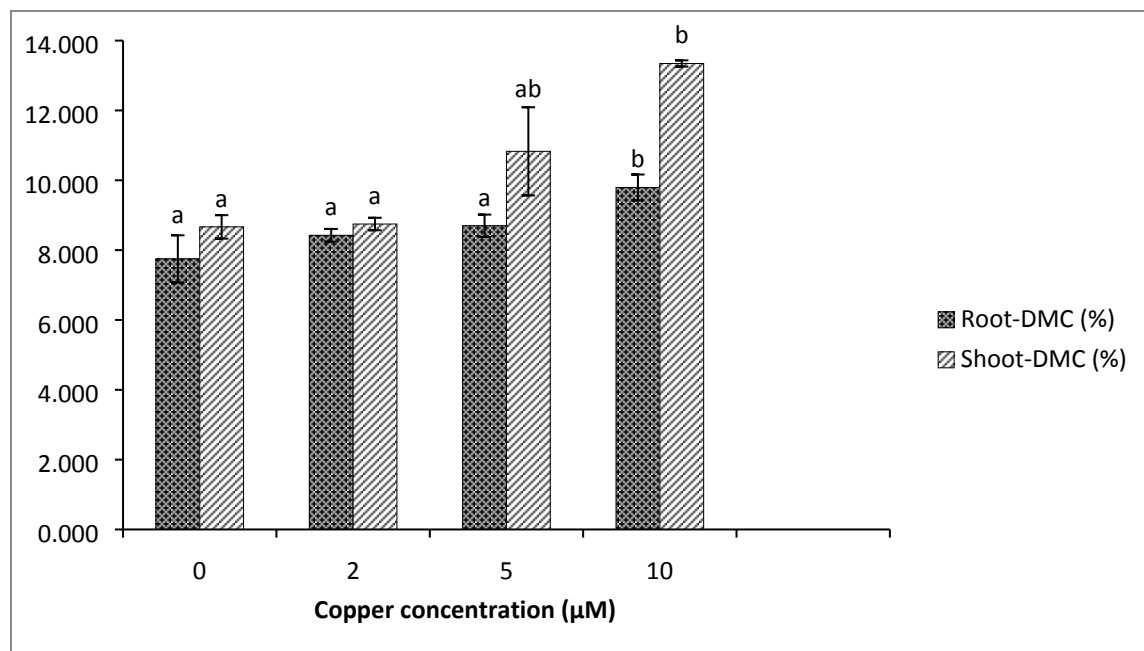
Sirko, A. and C. Gotor. 2007. Molecular links between metals in the environment and plant sulfur metabolism. In: Hawkesford M.J. and De Kok, L.J. editors. Sulfur in plants - an ecological perspective. Springer p. 169-195.

Yruela, I. 2005. Copper in plants. Braz. J. Pl. Physiol. 17(1): 145-156.

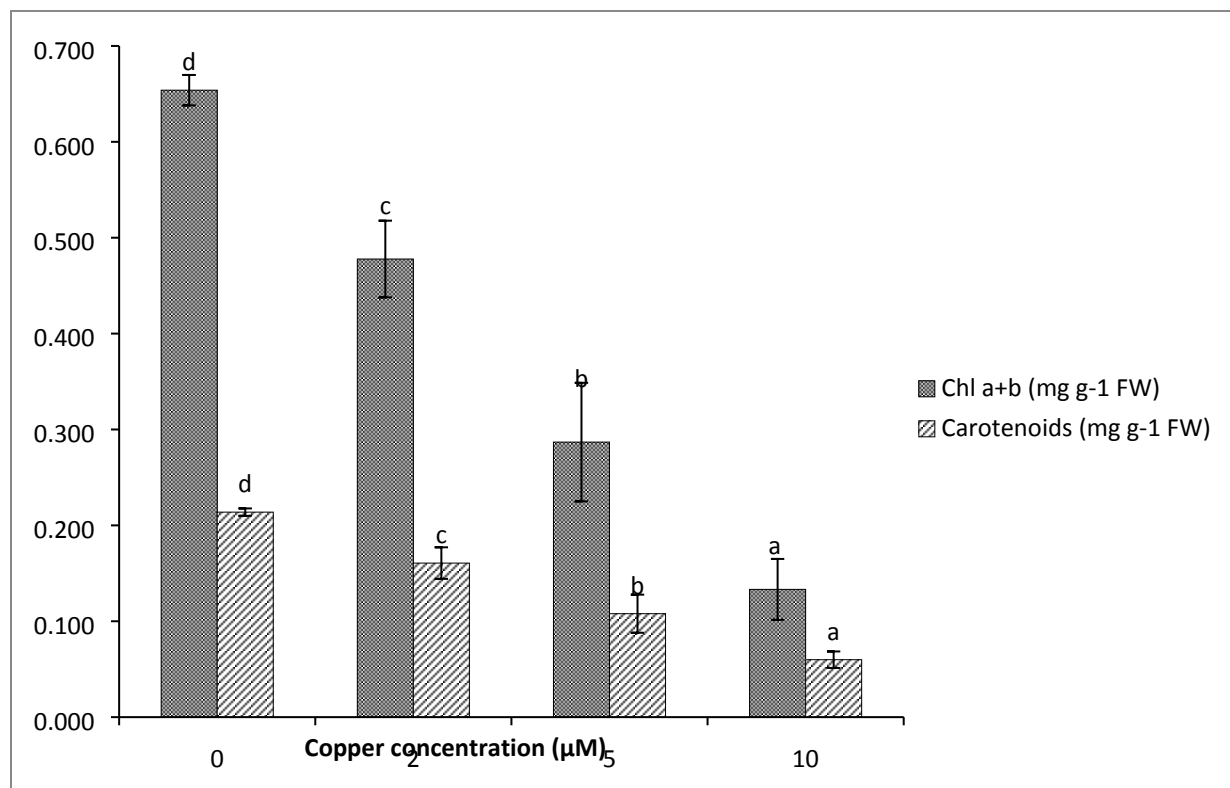
Yruela, I. 2009. Copper in plants: acquisition, transport and interactions. Funct. Pl. Biol. 36(5):409-430.



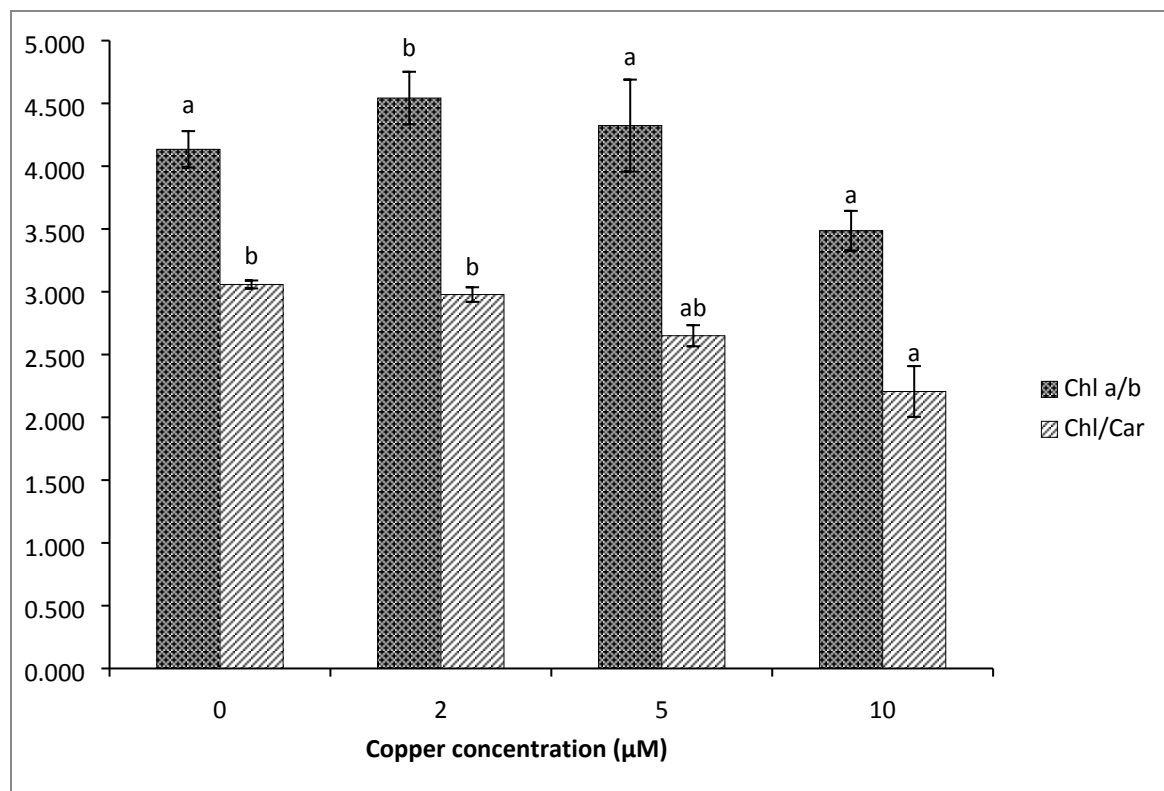
**Figure 1:** Impact of elevated levels of  $\text{Cu}^{2+}$  on biomass production of Stone-head cabbage (*Brassica Oleracea Var. Capitata*). 10-day-old seedlings of Stone-head cabbage (*Brassica Oleracea Var. Capitata*) were grown on a 25% Hoagland solution containing 0, 2, 5 and 10  $\mu\text{M}$   $\text{CuCl}_2$  in the root environment. The initial shoot and root fresh weight was  $0.059 \pm 0.004$  and  $0.009 \pm 0.003$  g, respectively. Data on biomass production (g FW) and shoot/root ratio represent the mean of 2 independent experiments with 9 measurements having 3 plants in each treatment ( $\pm$  SD). Means with different letters differ significantly at  $p \leq 0.01$  (Student's t-test).



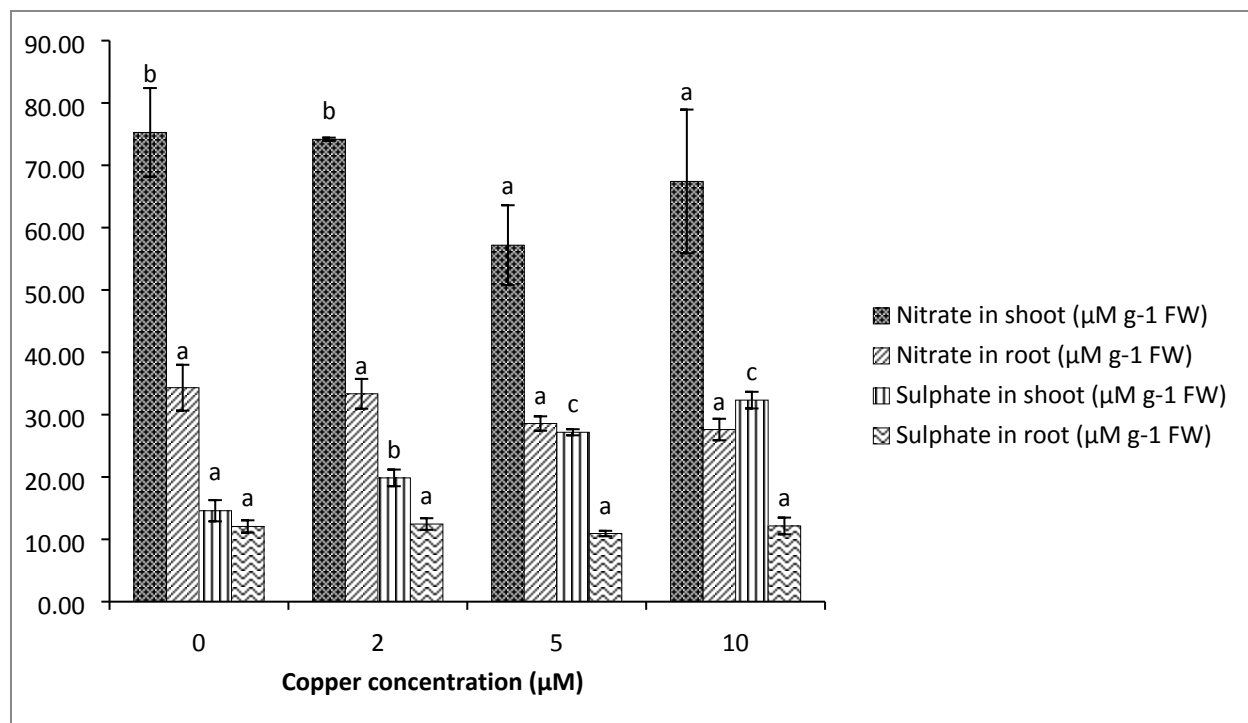
**Figure 2:** Impact of elevated levels of  $\text{Cu}^{2+}$  on dry matter content of Stone-head cabbage (*Brassica Oleracea Var. Capitata*). For experimental details see legends Fig. 1. Data on dry matter content (DMC %) represent the mean of 1 experiment with 9 measurements having 3 plants in each treatment ( $\pm$  SD). Means with different letters differ significantly at  $p \leq 0.01$  (Student's t-test).



**Figure 3:** Impact of elevated levels of  $\text{Cu}^{2+}$  on pigment content of Stone-head cabbage (*Brassica Oleracea* Var. *Capitata*). For experimental details see legends Fig. 1. Data on chlorophyll content (chl $a+b$ ; mg g<sup>-1</sup> FW) and carotenoids content (mg g<sup>-1</sup> FW) represent the mean of 3 measurements with 3 plants in each treatment ( $\pm$  SD). Means with different letters differ significantly at  $p \leq 0.01$  (Student's t-test).



**Figure 4:** Impact of elevated levels of  $\text{Cu}^{2+}$  on pigment content of Stone-head cabbage (*Brassica Oleracea Var. Capitata*). For experimental details see legends Fig. 1. Data on chlorophyll *a* to chlorophyll *b* ratio (Chl*a*/*b*) and chlorophyll to carotenoid ratio (Chl/Car) represent the mean of 3 measurements with 3 plants in each treatment ( $\pm$  SD). Means with different letters differ significantly at  $p \leq 0.01$  (Student's *t*-test).



**Figure 5:** Impact of elevated levels of  $\text{Cu}^{2+}$  on nitrate and sulfate content of Stone-head cabbage (*Brassica Oleracea* Var. *Capitata*). For experimental details see legends Fig. 1. Data on nitrate and sulfate contents ( $\mu\text{mol g}^{-1}$  FW) represent the mean of 3 measurements with 3 plants in each treatment ( $\pm$  SD). Means with different letters differ significantly at  $p \leq 0.01$  (Student's t-test).