

1 **No evidence for differential biomass and mineral content in adult plants grown**  
2 **from dimorphic seeds of *Suaeda aralocaspica***

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## Abstract

Producing two or more types of seeds by a single plant is known as seed heteromorphism. Comparison of seed traits or growth between plants grown from heteromorphic seeds has received considerable attention. However, information is scarce regarding the comparison of mineral content of adult plants from heteromorphic seeds. Here we present biomass and mineral profiles (Nitrogen, Phosphorus, Potassium, Sodium and Chloride) of adult plants grown from dimorphic seeds (non-dormant brown seeds and black seeds with non-deep physiological dormancy) of annual desert halophyte *Suaeda aralocaspica* at different levels of nutrient and salinity. The results showed that, although nutrient and salinity could affect dry weight and mineral content, seed-dimorphic plants of *S. aralocaspica* did not show significant difference under the same experimental conditions. This study is one of the few to compare the physiological responses between seed-heteromorphic plants and suggests that mineral status is exactly consistent with growth performance for these plants.

**Keywords:** dimorphic seeds, halophyte, mineral content, salinity, seed heteromorphism

## Introduction

Plants may produce different types of seeds on a single individual, called seed heteromorphism. Heteromorphic seeds usually differ in their ecological and biological characteristics, such as seed color and shape (Brändel 2007; Song et al. 2008; Ei-Keblawy et al. 2014), germination (Khan et al. 2001; Yao et al. 2010; Aguado et al. 2011), dispersability (Talavera et al. 2010) and soil seed bank (Mandák and Pyšek 2001; Cao et al. 2012). For example, the differences between peripheral and central achenes of annual heterocarpic species *Leontodon longirrostris* result in differences in dispersal and germination (Ruiz de Clavijo 2001).

Carry-over of differential growth and reproduction characteristics has also been reported in many seed-heteromorphic plants, including seedling establishment (Venable and Levin 1985), plant size (Cheplick and Quinn 1982), salt tolerance (Redondo-Gómez et al. 2008; Jiang et al. 2012), phenotypic plasticity (Zhang 1995), competitive ability (Imbert et al. 1997) and reproductive allocation (Sadeh et al. 2009; Lu et al. 2013). Plants grown from heteromorphic seeds generally show different growth characteristics under competitive conditions (Ellison 1987; Imbert et al. 1997). However, under certain conditions, such differences may disappear in the later life-cycle stage. For instance, *Emex spinosa* plants grown from subterranean seeds have larger leaf area and stems than plants grown from aerial achenes in mix culture, but these differences do not exist in monoculture when harvested (Weiss 1980).

Growth indices (biomass, plant height and so on) are generally used as the measure to compare seed-heteromorphic plants (Cheplick and Quinn 1982; Ruiz de

60 Clavijo and Jiménez 1998; Ruiz de Clavijo 2001; Mandák and Pyšek 2005). Few  
61 studies compare the physiological responses (antioxidative enzyme activities,  
62 chlorophyll content, chlorophyll fluorescence, gas exchange and ion accumulation)  
63 between seedlings or adult plants grown from heteromorphic seeds (Redondo-Gómez  
64 et al. 2008; Xu et al. 2011; Jiang et al. 2012). These studies show plants grown from  
65 different types of seeds maintain differences in physiological responses. As far as we  
66 know, there is no record of no physiological differences between plants grown from  
67 heteromorphic seeds.

68 *Suaeda aralocaspica* (Bunge) Freitag & Schütze (Amaranthaceae), is an annual  
69 halophyte restricted to saline-alkaline soils of Gobi desert in central Asia. *Suaeda*  
70 *aralocaspica* germinates from late March to April, flowering occurs from late August  
71 to September and fruits mature and disperse in October (Wang et al. 2012). This  
72 species produces two types of seeds on a single plant. Brown seeds are oblate,  
73 non-dormant and can germinate rapidly at relative high salinity; black seeds are  
74 biconvex, have non-deep Type 2 physiological dormancy and germinate slowly even  
75 at low salinity (Wang et al. 2008).

76 In this study, we hypothesized that mineral content of adult plants grown from  
77 heteromorphic seeds was consistent with growth performance. In order to test our  
78 hypothesis, we designed a single-plant pot experiment to compare the biomass and  
79 mineral profiles (Chloride (Cl), Sodium (Na), Nitrogen (N), Phosphorus (P) and  
80 Potassium (K)) of adult plants grown from dimorphic seeds of desert halophyte *S.*  
81 *aralocaspica* under different levels of nutrients and salinity.

82

## 83 **Materials and methods**

### 84 *Seed collection*

85 In early October 2008, freshly matured fruits of *S. aralocaspica* were collected from a  
86 natural population of approximately 600 individuals (44°14'N, 87°44'E; 445 m a.s.l.)  
87 in saline desert in the northern part of Xinjiang Province, China (for a detailed  
88 description of this site please refer to Wang et al. (2012)). In the laboratory, dry fruits  
89 were manually rubbed to detach the seeds enclosed by the membranous pericarps.  
90 Then the brown and black seeds were separately stored at 4 °C until used.

### 91 *Experimental design*

92 The experiment was carried out in a screen house covered with sun shade net (ca.  
93 25 % porosity) at Fukang Desert Ecosystem Observation and Experimental Station of  
94 the Chinese Academy of Sciences (44°17'26"N, 87°55'58"E; 460 m a.s.l.).

95 A randomized block design with six replicates was used. Each block contained  
96 18 pots representing a combination of two seed types (brown and black), three  
97 fertilization treatments (low, moderate and high) and three salinity levels (0 as control,  
98 low and high). For the fertilization treatment, granular lawn fertilizer (Osmocote 301,  
99 Scotts, Marysville, OH, USA; 15N : 11P : 13K : 2Mg) was used as the basic fertilizer  
100 and a commercial fertilizer (Peters1, Scotts; 20N : 20P : 20K) as the supplemental  
101 nutrient. In the fertilization treatment, each pot received (1) 1.2 g Osmocote 301 and  
102 100 mL Peters1 nutrient solution (0.046 g L<sup>-1</sup>) once a week (low fertilization), (2) 6 g  
103 Osmocote 301 and 100 mL Peters1 nutrient solution (0.23 g L<sup>-1</sup>) once a week

(moderate fertilization) or (3) 12 g Osmocote 301 and 100 mL Peters1 nutrient solution ( $0.46 \text{ g L}^{-1}$ ) once a week (high fertilization). Addition of the Peters1 nutrient solution began 3 weeks after sowing. For the salinity treatment, a mixed salt with a  $20\text{NaCl} : 20\text{Na}_2\text{SO}_4 : 1\text{NaHCO}_3$  (mass ratio) was used. Each pot received (1) 100 mL tapwater ( $0 \text{ g L}^{-1}$ ) (control) once a week, (2) 100 mL salt solution ( $1 \text{ g L}^{-1}$ ) once a week or (3) 100 mL salt solution ( $7 \text{ g L}^{-1}$ ) once a week. To avoid salt shock,  $7 \text{ g L}^{-1}$  salinity was applied gradually by adding  $1 \text{ g L}^{-1}$  salinity per week. Adding the salt solution started 4 weeks after planting.

About twenty seeds of brown or black seeds were sown into plastic pots (17 cm deep and 16 cm in diameter) filled with  $2000 \text{ cm}^3$  of a vermiculite quartz-sand mix (4:1 v/v) in late April 2009. Germinated seedlings were cultured for nearly 3 weeks and then thinned to one for each container. To reduce variation of initial seedling size, only seedlings of the similar height were used for each seed type. Pots were watered every 2 days, and the same amount of water/ nutrient solution/ salt solution was used for each pot. Five months after sowing, plants for each treatment were harvested.

### ***Measurements and chemical analyses***

Shoot samples were oven-dried to a constant weight at  $75^\circ\text{C}$  for 2 days, then weighed by an electronic analytical balance (accuracy:  $0.01 \text{ g}$ ). Oven-dried shoot samples were ground and stored in plastic bags.

Na and Cl were extracted by water extract method. Na content was determined by flame photometry and Cl content by Titrimetric method. For the total contents of N, P and K,  $0.5 \text{ g}$  samples was transferred into  $100 \text{ mL}$  digestion flask and  $5 \text{ mL}$

concentrated H<sub>2</sub>SO<sub>4</sub> was added to flask. Then 4 ml H<sub>2</sub>O<sub>2</sub> were added twice. The flasks were heated for ten minutes at 360 °C and 2 ml of H<sub>2</sub>O<sub>2</sub> was added. This step was repeated until the color of the digested samples turned to colorless. N content was determined by distillating method, P content by vanadium molybdate yellow colorimetric method, and K by flame photometry.

### ***Statistical analysis***

Three-way ANOVA was used to determine the effects of plant type, salinity and nutrients and their interactions on dry matter yields and mineral concentrations. Tukey's test were performed for multiple comparisons to determine significant ( $P < 0.05$ ) differences between individual treatments. All data were analyzed using SPSS Version 16.0 (SPSS Inc., Chicago, Illinois, U.S.A).

## **Results**

### ***Effects of different treatments on biomass***

There was no significant effect of plant type on biomass ( $P = 0.634$ ). In addition, there were no significant interactive effects between all three factors. In contrast, there were significant effects of salinity or nutrient levels on biomass (Table 1). Salinity significantly increased the biomass at moderate and high nutrient levels, but not at low nutrient level. Nutrient supplement significantly increased plant biomass. Plants had the highest biomass at high salinity and high nutrient level (Fig. 1).

### ***Effects of different treatments on Cl and Na concentrations***

The main effects of plant type on Cl concentration and Na concentration were not significant (Table 1). However, the main effects of salinity on Cl and Na concentrations were significant, i.e. salinity significantly increased Cl concentration at any nutrient levels. In contrast, nutrient supplement significantly decreased Cl concentration. Plants had the highest Cl concentration at high salinity and low nutrient levels, and had higher Na concentration at higher salinity under moderate or high nutrient conditions (Fig. 2). There were significant plant type x salinity, and plant type x nutrient interaction effect on Na concentration, and significant salinity x nutrient interaction effect on Cl concentration.

#### ***Effects of different treatments on N, P and K concentrations***

The main effects of plant type on N concentration ( $P = 0.393$ ), P concentration ( $P = 0.536$ ) and K concentration ( $P = 0.115$ ) were not significant. However, the main effects of salinity on N and P concentrations, and the main effects of nutrients on N, P and K concentrations were significant (Table 1). There were significant interaction effect of plant type x salinity on P, plant type x nutrient on K, and salinity x nutrient on N, P and K.

Overall, there were no significant differences in N, P and K concentrations between plants from brown seeds and black seeds under the conditions of this experiment. Salinity significantly decreased the N and P concentrations at moderate and high nutrient levels, but not at low nutrient level (Fig. 3).

## Discussion

Although differences between plants grown from heteromorphic seeds have been studied extensively (for reviews see Mandák 1997; Imbert 2002), our results are the first in which no significant mineral differences between adult plants grown from heteromorphic seeds have been documented. Dimorphic seeds of *S. aralocaspica* have different mass, dormancy and germination characteristics (Wang et al. 2008). However, the differences disappear for adult seed-dimorphic plants. This is reflected by not only biomass but also mineral content. So the results of this study support our hypothesis that mineral status is exactly consistent with growth performance of plants grown from heteromorphic seeds.

It is often hypothesized that plants grown from heteromorphic seeds have different competitive ability and stress tolerance. Some seed-heteromorphic species can maintain carry-over differences through the whole life cycle, even in the absence of stress (Weiss 1980; Venable and Levin 1985; Mandák and Pyšek 2005). In contrast, seed-heteromorphic *Crepis sancta* has similar reproductive biomass in the absence of competition (Imbert et al. 1997). There is even no difference in production between mixtures and monocultures of the ray plants of *Dimorphotheca sinuata* (Beneke et al. 1992). It seems that there is no clear fixed pattern for seed-heteromorphic plants.

Growth performance of seed-heteromorphic plants can be related to the physiological measures. Seedlings grown from brown and black seeds of *Suaeda splendens* grow well at 400 mmol L<sup>-1</sup> NaCl, but seedlings grown from brown seeds does not perform well at low salinity. Growth difference of seed-dimorphic plants can

be reflected by chlorophyll fluorescence and gas exchange (Redondo-Gómez et al. 2008). Similarly, for *Suaeda salsa*, Jiang et al. (2012) found that growth status of adult plants derived from dimorphic seeds was related to chlorophyll content and photosynthetic rate. Seedlings grown from yellow seeds of *Atriplex centralasiatica*, which have high salt tolerance, produce greater amounts of NO than do those from brown seeds (Xu et al. 2011). It's clear that physiological differences can reflect growth difference of seed-heteromorphic plants. Furthermore, our results show no difference in growth performance between seed-heteromorphic plants can also be reflected by no difference in mineral content.

Although plants grown from the two seed morphs of *S. aralocaspica* turned out to have similar responses to salinity and nutrient availability under the treatments in this experiment, mineral content are more sensitive to environmental factors than biomass. This is showed by the significant interaction effects between plant type and salinity or plant type and nutrient on different mineral concentrations. In other words, when the availability of salinity and nutrient is beyond the range applied in this study, the possible differences between plants grown from dimorphic seeds will be displayed by mineral concentration firstly.

In conclusion, in this study we documented direct evidence that adult plants grown from dimorphic seeds of *S. aralocaspica* had similar performance in biomass and mineral content under the same environmental conditions. Further researches are necessary to clarify whether the absence of differences in heteromorphic plants is related to other environmental constraints.

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218 **Competing Interests**

219 The authors declare there are no competing interests.

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## References

- Aguado M, Martínez-Sánchez JJ, Reig-Armiñana J, García-Breijo FJ, Franco JA, Vicente MJ (2011) Morphology, anatomy and germination response of heteromorphic achenes of *Anthemis chrysantha* J. Gay (Asteraceae), a critically endangered species. *Seed Sci Res* 21:283-294
- Beneke K, Van Rooyen MW, Theron GK (1992) Fruit polymorphism in ephemeral species of Namaqualand: VI. Intermorphic competition among plants cultivated from dimorphic diaspores. *S Afr J Bot* 58:469-477
- Brändel M (2007) Ecology of achene dimorphism in *Leontodon saxatilis*. *Ann Bot* 100:1189-1197
- Cao DC, Baskin CC, Baskin JM, Yang F, Huang ZY (2012) Comparison of germination and seed bank dynamics of dimorphic seeds of the cold desert halophyte *Suaeda corniculata* subsp. *mongolica*. *Ann Bot* 110:1545-1558
- Cheplick GP, Quinn JA (1982) *Amphicarpum purshii* and the "pessimistic strategy" in amphicarpic annuals with subterranean fruit. *Oecologia* 52:327-332
- El-Keblawy AA, Bhatt A, Gairola S (2014) Perianth colour affects germination behaviour in wind-pollinated *Salsola rubescens* in Arabian deserts. *Botany* 92:69-75
- Ellison AM (1987) Effect of seed dimorphism on the density-dependent dynamics of experimental populations of *Atriplex triangularis* (Chenopodiaceae). *Am J Bot* 74:1280-1288

- 243 Imbert E (2002) Ecological consequences and ontogeny of seed heteromorphism.  
244 Perspect Plant Ecol Evol Syst 5:13-36
- 245 Imbert E, Escarré J, Lepart J (1997) Seed heteromorphism in *Crepis sancta*  
246 (Asteraceae): Performance of two morphs in different environments. Oikos  
247 79:325-332
- 248 Jiang L, Wang L, Yin CH, Tian CY (2012) Differential salt tolerance and similar  
249 responses to nitrogen availability in plants grown from dimorphic seeds of  
250 *Suaeda salsa*. Flora 207:565-571
- 251 Khan MA, Gul B, Weber DJ (2001) Germination of dimorphic seeds of *Suaeda*  
252 *moquinii* under high salinity stress. Aust J Bot 49:185-192
- 253 Lu JJ, Ma WB, Tan DY, Baskin JM, Baskin CC (2013) Effects of environmental  
254 stress and nutlet morph on proportion and within-flower number-combination of  
255 morphs produced by the fruit-dimorphic species *Lappula duplicicarpa*  
256 (Boraginaceae). Plant Ecol 214:351-362
- 257 Mandák B (1997) Seed heteromorphism and the life cycle of plants: a literature  
258 review. Preslia 69:129-159
- 259 Mandák B, Pyšek P (2001) Fruit dispersal and seed banks in *Atriplex sagittata*: the  
260 role of heterocarpy. J Ecol 89:159-165
- 261 Mandák B, Pyšek P (2005) How does seed heteromorphism influence the life history  
262 stages of *Atriplex sagittata* (Chenopodiaceae)? Flora 200:516-526
- 263 Redondo-Gómez S, Mateos-Naranjo E, Cambrollé J, Luque T, Figueroa ME, Davy AJ  
264 (2008) Carry-over of differential salt tolerance in plants grown from dimorphic

265 seeds of *Suaeda splendens*. Ann Bot 102:103-112  
 266 Ruiz de Clavijo E, Jiménez MJ (1998) The influence of achene type and plant density  
 267 on growth and biomass allocation in the heterocarpic annual *Catananche lutea*  
 268 (Asteraceae). Int J Plant Sci 159:637-647  
 269 Ruiz de Clavijo E (2001) The role of dimorphic achenes in the biology of the annual  
 270 weed *Leontodon longirrostris*. Weed Res 41:275-286  
 271 Sadeh A, Guterman H, Gersani M, Ovadia O (2009) Plastic bet-hedging in an  
 272 amphicarpic annual: an integrated strategy under variable conditions. Evol Ecol  
 273 23:373-388  
 274 Song J, Fan H, Zhao YY, Jia YH, Du XH, Wang BS (2008) Effect of salinity on  
 275 germination, seedling emergence, seedling growth and ion accumulation of a  
 276 euhalophyte *Suaeda salsa* in an intertidal zone and on saline inland. Aquat Bot  
 277 88:331-337  
 278 Talavera M, Ortiz PL, Arista M, Berjano R, Imbert E (2010) Disentangling sources of  
 279 maternal effects in the heterocarpic species *Rumex bucephalophorus*. Perspect  
 280 Plant Ecol Evol Syst 12:295-304  
 281 Venable DL, Levin DA (1985) Ecology of achene dimorphism in *Heterotheca*  
 282 *latifolia*: II. Demographic variation within populations. J Ecol 73:743-755  
 283 Wang L, Baskin JM, Baskin CC, Cornelissen JHC, Dong M, Huang ZY (2012) Seed  
 284 dimorphism, nutrients and salinity differentially affect seed traits of the desert  
 285 halophyte *Suaeda aralocaspica* via multiple maternal effects. BMC Plant Biol  
 286 12:170

- 287 Wang L, Huang ZY, Baskin CC, Baskin JM, Dong M (2008) Germination of  
288 dimorphic seeds of the desert annual halophyte *Suaeda aralocaspica*  
289 (Chenopodiaceae), a C<sub>4</sub> plant without Kranz anatomy. Ann Bot 102:757-769
- 290 Weiss PW (1980) Germination, reproduction and interference in the amphicarpic  
291 annual *Emex spinosa* (L.) Campd. Oecologia 45:244-251
- 292 Xu J, Yin HX, Yang LL, Xie ZZ, Liu XJ (2011) Differential salt tolerance in  
293 seedlings derived from dimorphic seeds of *Atriplex centralasiatica*: from  
294 physiology to molecular analysis. Planta 233:859-871
- 295 Yao SX, Lan HY, Zhang FC (2010) Variation of seed heteromorphism in  
296 *Chenopodium album* and the effect of salinity stress on the descendants. Ann Bot  
297 105:1015-1025
- 298 Zhang JH (1995) Differences in phenotypic plasticity between plants from dimorphic  
299 seeds of *Cakile edentula*. Oecologia 102:353-360

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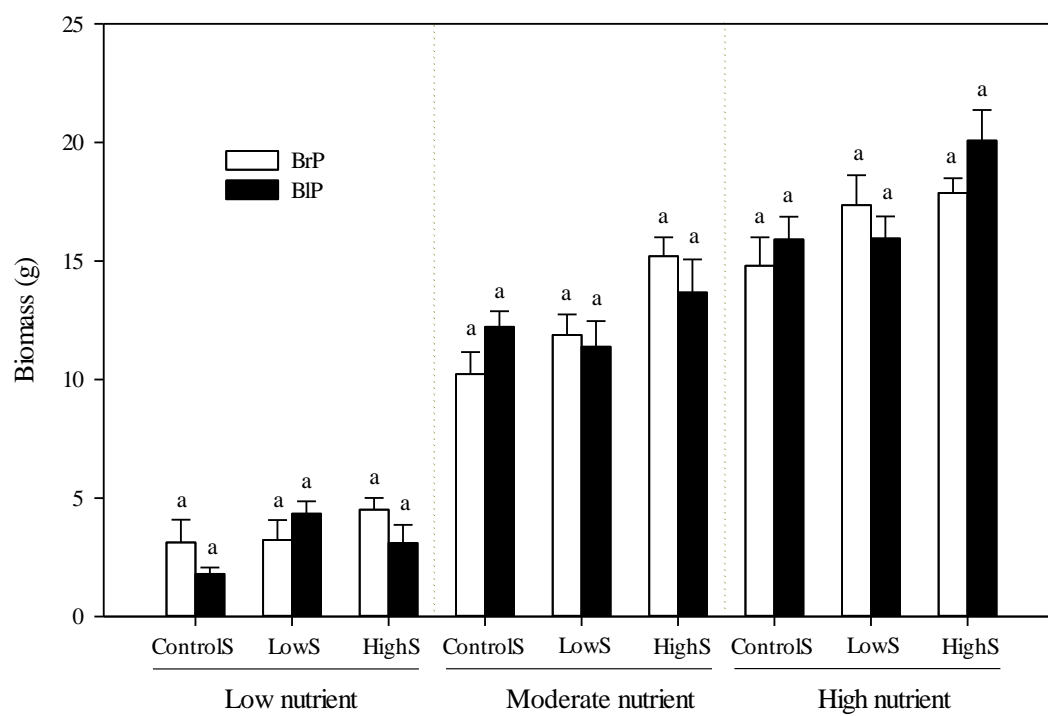
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## Figure legends

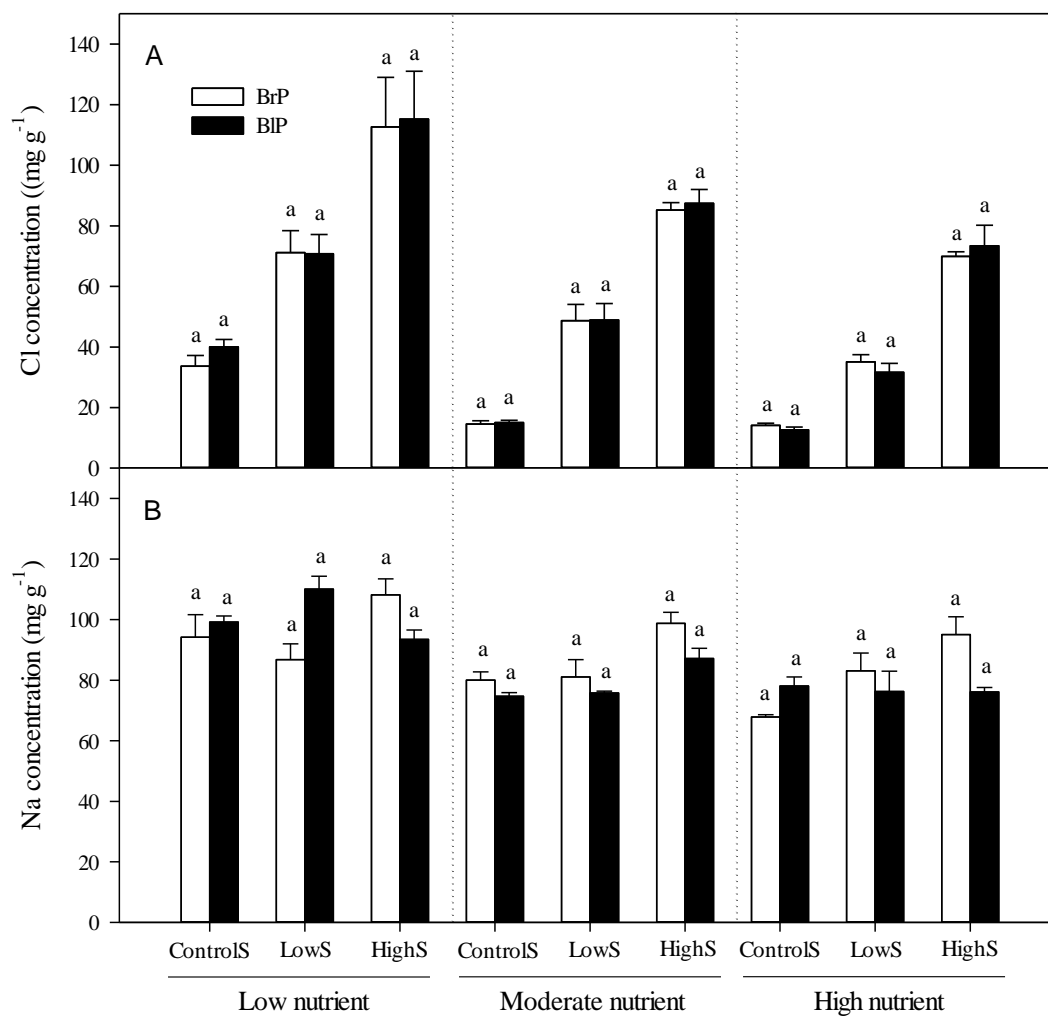
**Figure 1.** Effects of plant type, salinity and nutrient levels on biomass. BrP: plant grown from brown seed; BIP: plant grown from black seed; ControlS: Control salinity; LowS: Low salinity; HighS: High salinity. Bars with the same letters indicate no significant differences in dry weight of seed-dimorphic plants at a given salinity under a particular nutrient treatment at  $P < 0.05$  level.

**Figure 2.** Effects of plant type, salinity and nutrient levels on Cl and Na concentration. BrP: plant grown from brown seed; BIP: plant grown from black seed; ControlS: Control salinity; LowS: Low salinity; HighS: High salinity. Bars with the same letters indicate no significant differences in Cl or Na concentration of seed-dimorphic plants at a given salinity under a particular nutrient treatment at  $P < 0.05$  level.

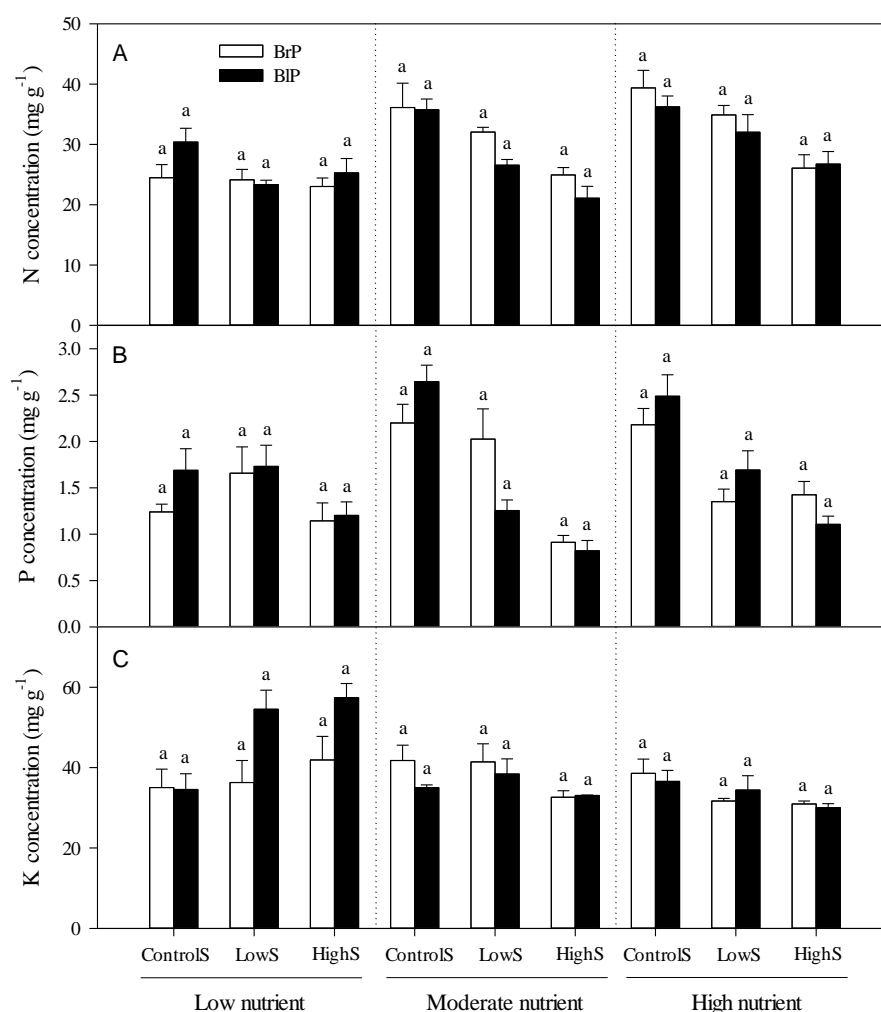
**Figure 3.** Effects of plant type, salinity and nutrient levels on N, P and K concentration. BrP: plant grown from brown seed; BIP: plant grown from black seed; ControlS: Control salinity; LowS: Low salinity; HighS: High salinity. Bars with the same letters indicate no significant differences in N, P or K concentration of seed-dimorphic plants at a given salinity under a particular nutrient treatment at  $P < 0.05$  level.



**Fig 1**



**Fig 2**



**Fig 3**

**Table 1** Three-way ANOVA of the effects of plant type, salinity, nutrient levels and their interactions on biomass and mineral content (Cl, Na, N, P and K) of *Suaeda aralocaspica*

Source	Biomass		Cl		Na		N		P		K	
	<i>F</i> -value	<i>P</i> -value	<i>F</i> -value	<i>P</i> -value	<i>F</i> -value	<i>P</i> -value	<i>F</i> -value	<i>P</i> -value	<i>F</i> -value	<i>P</i> -value	<i>F</i> -value	<i>P</i> -value
Plant type (PT)	0.04	0.947	0.05	0.825	1.74	0.193	0.741	0.393	0.387	0.536	2.565	0.115
Salt (S)	13.248	<0.001	461.22	<0.001	9.911	<0.001	28.763	<0.001	40.287	<0.001	0.828	0.442
Nutrient (N)	336.666	<0.001	111.618	<0.001	33.885	<0.001	18.992	<0.001	3.209	0.048	11.567	<0.001
PT*S	0.415	0.661	0.293	0.747	9.294	<0.001	1.335	0.272	3.833	0.028	3.035	0.056
PT*N	0.609	0.546	0.883	0.419	3.264	0.046	2.974	0.06	1.275	0.288	6.824	0.002
S*N	1.538	0.198	9.496	<0.001	1.51	0.212	3.474	0.013	7.805	<0.001	6.529	<0.001
PT*S*N	2.347	0.06	0.427	0.788	3.271	0.018	0.612	0.656	2.049	0.1	0.959	0.438