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Intragenomic polymorphisms among high-copy loci: A genus-wide study of nuclear ribosomal DNA in *Asclepias* (Apocynaceae)

Despite knowledge that concerted evolution of high-copy loci is often imperfect, few studies investigate the extent of intragenomic polymorphisms and comparisons across a large number of species are rarely made. We present a bioinformatic pipeline for characterizing polymorphisms within an individual among copies of a high-copy locus. Results are presented for nuclear ribosomal DNA (nrDNA) across the milkweed genus, *Asclepias*. The 18S-26S portion of the nrDNA cistron of *Asclepias syriaca* served as a reference for assembly of the region from 124 samples representing 90 species of *Asclepias*. Reads were mapped back to each individual's consensus and at each position reads differing from the consensus were tallied using a custom perl script. Low frequency polymorphisms existed in all individuals (mean = 5.8%). Most nrDNA positions (91%) were polymorphic in at least one individual, with polymorphic sites being less frequent in subunit regions and loops. Highly polymorphic sites existed in each individual, with highest abundance in the “noncoding” ITS regions. Phylogenetic signal was present in the distribution of intragenomic polymorphisms across the genus. Intragenomic polymorphisms in nrDNA are common in *Asclepias*, being found at higher frequency than any other study to date. The high and variable frequency of polymorphisms across species highlights concerns that phylogenetic applications of nrDNA may be error-prone. The new analytical approach provided here is applicable to other taxa and other high-copy regions characterized by low coverage genome sequencing (genome skimming).

1 Intragenomic polymorphisms among high-copy loci: A genus-wide study of nuclear
2 ribosomal DNA in *Asclepias* (Apocynaceae)

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13 **INTRODUCTION**

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With the advent of DNA sequencing technology to infer phylogenetic relationships, investigators began searching for genetic loci that were both phylogenetically informative and readily sequenced in most organisms. The use of nuclear ribosomal DNA (nrDNA) soon became a popular choice for phylogenetic inference (Hamby & Zimmer, 1988; Hillis & Dixon, 1991; Baldwin, 1992; Baldwin et al., 1995; Álvarez & Wendel, 2003). Nuclear ribosomal DNA offered several advantages over other loci: the combination of highly conserved and variable regions allowed phylogenetic inference across a broad range of evolutionary time scales, conserved regions allowed the use of “universal” PCR primers applicable to a wide range of taxa, the high copy number of nrDNA repeats allowed reliable amplification from lower quality DNA extractions, and the process of concerted evolution ensured that these copies were similar within individuals (Baldwin et al., 1995). The use of nrDNA, particularly the variable internal transcribed spacer (ITS) regions, became widespread, to the extent that many studies were based exclusively on ITS data (Álvarez & Wendel, 2003).

However, nrDNA loci have been shown to harbor limitations in their phylogenetic utility. Nuclear ribosomal DNA copies are assembled as tandem repeats at one or more loci in the genome, with each locus being known as an array. The number of repeats present within an array is labile, as is the number and location of arrays (Álvarez & Wendel, 2003). The process of nrDNA copy homogenization is thought to occur much more frequently within than among arrays (Schlötterer & Tautz, 1994). Thus, differing nrDNA alleles may become fixed in different arrays within a genome, creating paralogy that, if unrecognized, may confound phylogenetic inference (Álvarez & Wendel, 2003). Moreover, these events can create pseudogenes which, freed from selective pressures, may evolve through processes quite different from the functional loci and provide misleading evidence for between-individual genetic divergences if compared to functional copies. These events may occur at a greater rate than inter-array homogenization via concerted evolution (Gernandt & Liston, 1999).

Due to the technical difficulty of systematically sequencing individual nrDNA loci because of their high copy number, studies characterizing the abundance and patterns of intragenomic polymorphisms have been rare. Recently, studies utilizing whole-genome shotgun sequencing have begun to reveal levels of intragenomic polymorphism in *Drosophila* (Stage & Eickbush, 2007), nematodes (Bik et al., 2013), and fungi (Ganley & Kobayashi, 2007). However, these studies included a small number of species (12, 6, and 5, respectively) and did not attempt to place patterns of polymorphism in a phylogenetic context. Other studies have used intragenomic polymorphisms to identify populations of *Arabidopsis* (Simon et al., 2012) and infer intraspecific phylogenies of *Saccharomyces* (West et al., 2014). Studies of intragenomic polymorphism patterns across many species have not been performed in plants (but see Straub et al., 2012).

The current study utilizes high throughput technology to survey a large sample of species and individuals in the angiosperm genus *Asclepias* (Apocynaceae) in order to characterize levels of intragenomic nrDNA polymorphism and place these within a phylogenetic context. The methods presented here are expanded from those we have previously developed as part of the Milkweed Genome Project (Straub et al., 2011, 2012), and generalized for use with a large number of taxa and any high-copy locus, such as those that may be obtained from a genome-skimming or Hyb-Seq study (Straub et al., 2012; Weitemier et al., 2014).

METHODS

Sampling and sequencing

One hundred twenty-five individuals representing 90 *Asclepias* species and subspecies were sampled (Table 1) and sequencing libraries were produced as described in Straub et al. (2012). Two individuals of putatively hybrid origin were included: *A. albicans* $\times$ *subulata* and *A. speciosa* $\times$ *syriaca*. These individuals were collected from wild populations and identified as hybrids through expression of intermediate morphological characteristics (M. Fishbein, unpubl.

data; see also Fishbein et al., 2011). Samples were multiplexed in approximately equimolar ratios, with up to 21 individuals per lane, and sequenced with 80 bp single-end reads on an Illumina GAIIX instrument (Illumina, San Diego California, USA). *Asclepias subverticillata* was multiplexed in a lane with 32 samples and sequenced with 101 bp paired-end reads on an Illumina HiSeq 2000 instrument, with reads analyzed as though they were single-end. One individual of *A. syriaca* was sequenced at higher coverage: this individual was sequenced in a single lane on an Illumina GAIIX with 40 bp single-end reads (Straub et al., 2011). To allow more efficient assembly downstream, read pools were filtered to remove plastid reads (using the custom script `sort_fastq_v1.pl` modified to retain Ns; Knaus, 2010).

An *A. syriaca* haploid genome size estimate of 420 Mbp and a nrDNA copy number estimate of 960 were used for estimates of sequencing depth and for comparisons with other organisms. These estimates are modified from Straub et al. (2011), where an incorrect estimate of the average *A. syriaca* 2C value led to a haploid genome size estimate of 820 Mbp and a nrDNA copy number estimate of 1,845. The current values are based on 2C estimates from Bai et al. (2012) and Bainard et al. (2012).

Polymorphism quantification

The method for determining polymorphisms present among nrDNA copies within an individual while retaining information about position homology across a group of distantly related individuals included four general steps: 1) A sequence was selected to serve as a reference for the whole group. 2) A consensus sequence was obtained for each individual taxon and aligned against the group reference, allowing tailored read-mapping for each individual while associating positions along the individual consensus with their homologous positions in the group reference. 3) Reads for each individual were mapped onto that individual's consensus sequence. 4) At each position the reads differing from the individual consensus were tallied.

1) Group reference

The nrDNA cistron of the high-coverage *A. syriaca* individual was previously assembled (Straub et al., 2011; GenBank JF312046). The nontranscribed spacer and external transcribed spacers from each end were removed due to the presence of internal repeats, and the more conserved 18S and 26S subunits used as the boundaries of the alignment. The resulting reference sequence contained 5839 bp.

2) Individual consensus sequences

Read pools were examined prior to consensus assembly and reads that were exact duplicates were reduced to a single representative, retaining the highest average quality score (using the custom script fastq_collapse.py, available at <https://github.com/listonlab>). Sequences for each individual were constructed via reference-guided assembly, with *A. syriaca* as the reference, using Alignreads ver. 2.25 (Straub et al., 2011). Alignreads is a pipeline that includes the short-read assembler YASRA (Ratan, 2009), utilities from the MUMmer ver. 3.0 suite (Delcher et al., 2002; Kurtz et al., 2004), and custom scripts. Parameters were selected to ensure high identity of reads mapping to the nrDNA reference (95%), but allow the reconstructed sequence to differ from the reference. In addition to assembling the individual consensus sequence, Alignreads outputs a file associating each position in the group reference with those in the individual consensus.

3) Read mapping

Prior to mapping reads from an individual onto its consensus, reads with an average quality (Phred) score below 20 were removed, and bases in the remaining reads with a score below 20 were converted to Ns using FASTX-Toolkit ver. 0.0.13 (Gordon, 2008). Read mapping was performed with the program BWA ver. 0.5.7 (Li & Durbin, 2009), and output files processed with the SAMtools ver. 0.1.13 utilities (Li et al., 2009).

4) Polymorphism counting

A perl script was developed to tally the number of reads differing from the consensus at each position (polymorphic_read_counter_bwaPileup.pl ver. 3.03b, available at <https://github.com/listonlab>). For example, a base covered by 10 reads might have 7 reads with a G in that position and 3 with a C. In this case the consensus would have called a G at that position, with 30% of the reads differing. See Supplemental File 1 for exact parameters and a pipeline of commands used.

Positions with 2% or more of reads differing from the consensus base were considered polymorphic. This cutoff is the same used by Straub et al. (2011) and comparable to that used by Nguyen et al. (2011) under a similar quality-filtering scheme. The control *PhiX* lane of the higher coverage *A. syriaca* individual was examined by Straub et al. (2011, Additional file 1 from that study) and found to have an error rate much less than 2%, indicating that the cutoff used here may be somewhat conservative. In addition to counting positions that were polymorphic, positions were recorded as “highly polymorphic” if 10% or more of the reads differed from the consensus.

In order to keep homologous bases aligned across individuals, only those positions that were present in the *A. syriaca* (group) reference were kept in the analysis (i.e., insertions relative to the reference were discarded). Note that deletions (relative to the reference) fixed within an individual are also not considered because zero reads called a base at that position.

RNA structure determination

The secondary structure of each subunit and spacer region was predicted for the *A. syriaca* reference from the minimum free energy structure found by the program RNAfold for the 18S, ITS1, and ITS2 regions and RNAcofold for the 5.8S+26S regions (Lorenz et al., 2011). Program default model parameters were used (37° C, unpaired bases can participate in up to one dangling end). Positions along the cistron were then categorized as either paired (stems) or unpaired (loops).

138 ***Polymorphisms within the cistron***

139 The effects of cistron position (subunit or spacer) and secondary structure position (stem
140 or loop) on the likelihood of a position being polymorphic or highly polymorphic were assessed
141 in two ways. Differences in the likelihood that at least one individual was polymorphic at a
142 position (e.g. positions coded as either polymorphic or invariant) were assessed via a two factor
143 multiple logistic regression, as implemented in R ver. 3.1.0 using the MASS ver. 7.3.33 package
144 (Venables & Ripley, 2002; R Core Team, 2014). Differences in the abundance of polymorphic
145 individuals at a position were assessed using square-root transformed data with a two-way
146 ANOVA and type III sum of squares for unbalanced design, as implemented using the car ver.
147 2.0.20 package (Fox & Weisberg, 2011). The ANOVA analysis is not reported for the highly
148 polymorphic individuals because the data diverge substantially from assumptions of a normal
149 distribution.

150 ***Phylogenetic context***

151 A maximum likelihood estimate of phylogenetic relationships within *Asclepias* was
152 produced by Fishbein et al. (2011, Figure 2, TreeBase #27576). This tree was pruned to match the
153 sampling in this study, and counts of polymorphic positions were recorded for each taxon. Taxa
154 sampled in this study, but not present in Fishbein et al. (2011), were omitted from further
155 analyses. Counts were averaged for taxa with multiple individuals in this study, but sampled only
156 once in Fishbein et al. (2011). Ancestral states for the number of polymorphic positions in the
157 rDNA cistron were reconstructed using squared-change parsimony in the Mesquite phylogenetic
158 suite ver. 2.75+ build 573 (Maddison & Maddison, 2011; Maddison, Maddison & Midford,
159 2011).

160 Phylogenetic signal in the distribution of the number polymorphic positions was tested in
161 two ways. In the first method, the total length of the tree (parsimony steps) in terms of changes in
162 number of polymorphic positions was compared to a distribution of tree lengths created by

10,000 permutations of polymorphic positions across tips. In the second method, a likelihood ratio test (LRT) was performed between models where character evolution followed a Brownian motion model across the tree. In the first model, the parameter lambda (describing how well the phylogeny correctly predicts the covariance among taxa for a trait) was found that maximized the model's likelihood (Pagel, 1999). This was compared to the likelihood found when lambda was held at zero, representing phylogenetic independence among species for that trait. Parsimony permutations were performed in Mesquite (Maddison & Maddison, 2011; Maddison, Maddison & Midford, 2011), and likelihood ratio tests were performed in R with the phytools ver. 0.4.05 and ape ver. 3.1.2 packages (Paradis, Claude & Strimmer, 2004; Revell, 2012; R Core Team, 2014).

To determine if any clades held intragenomic polymorphism frequencies that were significantly high or low relative to the rest of the phylogeny, polymorphism rates were simulated along the tree and the true polymorphism counts compared to the distribution of simulated counts (Garland et al., 1993). Two tips in the tree separated by zero branch length (*A. asperula* ssp. *asperula* and ssp. *capricornu*) were collapsed, and the average of the polymorphism counts for the four sampled individuals of the species used for the new tip. A model of trait evolution under Brownian motion, using the lambda parameter estimated from the LRT above, was found from 10,000 random starting points using the fitContinuous function from the geiger ver. 2.0.3 R package (Harmon et al., 2008). This model was used to simulate polymorphism counts across the phylogeny 10,000 times, with lower and upper bounds of 0 and infinity, respectively, using the fastBM function in phytools ver. 0.4.31 (Revell, 2012). For each node, the ancestral state was estimated for the true data and the simulated data using squared change parsimony as implemented in Mesquite (Maddison & Maddison, 2011; Maddison, Maddison & Midford, 2011).

RESULTS

188 Average coverage of the nrDNA region was ~97x for all individuals (median = 88x;
189 Sequence Read Archive ###-###). Despite high overall coverage of the nrDNA region, not all
190 positions of the cistron were assembled for all individuals; therefore results for polymorphic
191 positions are presented both as counts and as percentages of sequenced bases. The *A. syriaca*
192 reference had total genome coverage of ~0.8x (Sequence Read Archive SRX040889).

193 ***Intragenomic polymorphism***

194 All individuals were polymorphic at several positions homologous with the *A. syriaca*
195 reference (Table 1). Number of polymorphic positions ranged from 23 (*A. verticillata*, 0.41% of
196 sequenced bases) to 882 (*A. solanoana*, 15.13%), with a mean of 333 (5.77%, Fig. 1). A very
197 high percentage (91%) of positions in the *A. syriaca* reference were polymorphic in at least one
198 individual (Fig. 2A).

199 Positions were significantly more likely to be polymorphic if they were in a spacer region
200 (ITS1, ITS2; Fig. 2A) or stem (Fig. 3A). This is true both when considering the number of
201 individuals polymorphic at a position (Table 2), and whether any sample was polymorphic at that
202 position (Table 3A).

203 The number of highly polymorphic positions ranged from 2 (*A. arenaria*, 0.03%) to 111
204 (*A. boliviensis*, 1.91%) with a mean of 28 (0.50%). Positions highly polymorphic in more than 10
205 individuals were found in the 18S, ITS1, ITS2, and 26S regions (Fig. 2B). The most polymorphic
206 position was 4172 (using the *A. syriaca* reference), in the 26S region, which was highly
207 polymorphic in 29 individuals. Highly polymorphic positions were dramatically less frequent in
208 the subunit regions (18S=19%, 5.8S=26%, 26S=31%) than in the spacer regions (ITS1=61%,
209 ITS2=57%; Table 3B, Fig. 2B). Positions in secondary structure stems were moderately more
210 likely to be highly polymorphic in at least one individual than loop positions (Table 3B, Fig. 3B).

211 ***Phylogenetic Signal***

The number of polymorphic base pair positions exhibited strong phylogenetic signal across *Asclepias* under both the permutation test ($P < 0.0012$) and the likelihood ratio test (estimated $\lambda = 0.51$, $P = 0.0067$; Table 4; Fig. 4). This signal remained even after the ITS regions were removed from the dataset (permutation test $P < 0.0200$, $\lambda = 0.45$, LRT $P = 0.0112$). Highly polymorphic base pair abundance, however, was not significantly influenced by phylogenetic history (Table 4; Fig. 5), even when considering only the subunit regions.

Of the 53 resolved clades in the phylogeny, none showed polymorphism values more extreme than expected under a Brownian motion model after correcting for multiple comparisons. The most extreme clade was that formed by *A. hypoleuca* and *A. otarioides*, which had an ancestral number of polymorphic positions exceeded by 79 of the 10,000 simulations (when excluding the spacer regions this node was exceeded by 60 simulations). The following most extreme nodes were those ancestral to *A. rosea* and *A. lemmonii* with more polymorphic positions than all but 130 (79) simulations, and ancestral to *A. boliviensis* and *A. mellodora* with more polymorphic positions than all but 246 (150) simulations. Under a Bonferroni correction a clade would require 9 or fewer simulations more extreme than the observed value ($\alpha = 0.05$) to reject a hypothesis of no divergence from Brownian motion.

Identification of Mixed Ancestry

The number of polymorphic sites for hybrid individuals, 299 for *A. albicans* $\times$ *subulata* and 161 for *A. speciosa* $\times$ *syriaca*, are less than the mean number of 333 polymorphic sites; and the number of highly polymorphic sites, 12 and 37, are less than or greater than the mean of 28. Of those positions that are highly polymorphic, 4 of 37 in *A. speciosa* $\times$ *syriaca* have a minor allele frequency of 0.3 or higher, while none of the positions in *A. albicans* $\times$ *subulata* have a minor allele frequency above 0.2.

DISCUSSION

Absolute levels of intragenomic polymorphism among the copies of nrDNA in *Asclepias* (mean=333 positions) were found to be much higher than levels reported for nematodes (<250; Bik et al., 2013), fungi (3-37; Ganley & Kobayashi, 2007), and *Drosophila* (3-18; Stage & Eickbush, 2007) when including all polymorphic positions. When considering only positions that are highly polymorphic, *Asclepias* exhibits slightly higher rates (mean=28.4 positions) than fungi and *Drosophila*, but much lower rates than nematodes. However, these comparisons may be misleading: First, the number of nrDNA copies varies greatly between these taxa, estimated to range from about 50-180 in the fungal species, 200-250 in *Drosophila melanogaster*, 56-323 in the nematodes, and about 960 in *Asclepias* (Ganley & Kobayashi, 2007; Stage & Eickbush, 2007; Straub et al., 2011; Bik et al., 2013). Second, polymorphic base pair counts are confounded by differing criteria for scoring polymorphism (i.e., methods for excluding sequencing errors). The levels listed for the fungal species include both “high-confidence” and “low-confidence” polymorphisms, based primarily on sequence quality (Ganley & Kobayashi, 2007). The levels listed for *Drosophila* are polymorphisms present in $\geq 3\%$ of loci (Stage & Eickbush, 2007). Bik et al. (2013) tallied read counts using a method similar to the method presented here, but called positions polymorphic when the count of differing reads exceeded what would be expected for a single copy locus. Nevertheless, given that *Asclepias* has absolute counts of polymorphic positions at least 33% higher than the other organisms studied, and that the sequencing depth was nearly two orders of magnitude greater in the nematodes than in *Asclepias* (6.3-10x per nrDNA copy in nematodes, vs. $\sim 0.1x$ in *Asclepias*; Bik et al., 2013), it is likely that *Asclepias* harbors greater rates of intragenomic polymorphism within the nrDNA cistron than the organisms studied to date.

Polymorphism patterns across the nrDNA cistron

Spacer regions (ITS1, ITS2) had higher frequencies of polymorphic positions than subunit regions (18S, 5.8S, 26S; Fig. 2). However, positions with low polymorphism frequencies are

distributed much more evenly across the nrDNA cistron (Fig. 2A) than highly polymorphic positions, which show strong differentiation between the subunit and spacer regions (Fig. 2B). The lower frequency of highly polymorphic positions within the subunit regions suggests that these regions are under selection to remain homogenous within individual genomes. The lower difference in low polymorphism frequency between subunit and spacer regions suggests that this selection pressure is positively correlated with the proportion of nrDNA copies that differ from the majority. These findings are in contrast to those reported for nematodes (Bik et al., 2013), where the subunit regions had much higher levels of polymorphism abundance than the spacer regions.

Positions in stem regions were more likely to be polymorphic than loop positions (Fig. 3). This was strongly significant for all polymorphic positions (Tables 2, 3A) and moderately significant for highly polymorphic positions (Table 3B). This would seem to contradict the hypothesis that stem sites in general should be more highly conserved in order to maintain a functional RNA secondary structure. Indeed, this finding agrees with those from Rhetsky (1995), who not only found that trees estimated from stem regions contained longer branch lengths than those from loop regions, but that those stem sites least likely to affect secondary structure tended to be less conserved. Loop sites, on the other hand, contain a large proportion of the sites critical to ribosomal function (Rzhetsky, 1995), and may be under stronger stabilizing selection than stem sites.

Among the subunit regions, the 26S region harbored the highest frequency of highly polymorphic positions. This agrees with Stage & Eickbush (2007) who showed the *Drosophila* 28S region to have a higher mutation rate than the other subunit regions. However, in that study the 28S region also had a lower frequency of polymorphic base pairs. Stage & Eickbush (2007) hypothesize that this is due to the action of two retrotransposable elements found in many *Drosophila* 28S copies, with instances of aborted insertions causing the cell to repair the region

using a nearby template and thereby homogenize the copies. They predict that levels of polymorphism will be higher in the 28S region of organisms lacking the retrotransposable elements, as seen here in the homologous angiosperm 26S region.

Within the subunit regions, highly polymorphic positions may be more common within expansion regions of the RNA gene (less conserved regions that tend to incorporate sequence insertions without affecting functionality; Clark et al., 1984). This is strongly implied by the recovery of clusters of highly polymorphic sites near *A. syriaca* positions 4440 and 5020, which are directly within the 26S expansion regions seven and eight, respectively (Kolosha & Fodor, 1990; Fig. 1). This agrees with results found in *Drosophila* (Stage & Eickbush, 2007). However, this may not be true for all highly polymorphic sites, as the high peak at position 4172 is not within an expansion region.

Phylogenetic context

Mapping the number of polymorphic positions onto the phylogeny of *Asclepias* demonstrates strong and significant phylogenetic signal when counting all polymorphisms (Fig. 4), but this signal is not significant when only counting highly polymorphic positions (Fig. 5). While this study demonstrates phylogenetic signal in polymorphism abundance, it remains unknown whether abundance within a lineage is influenced by selection or purely neutral causes. Different demographic histories across clades could create this effect via neutral causes, while selection against organisms that retain too many variant nrDNA copies could be variable across the genus. Polymorphisms present in a high proportion of nrDNA copies may be uniformly selected against, while low polymorphism sites are tolerated at varying levels. This could explain the lack of phylogenetic signal at the highly polymorphic sites, and its presence when including all polymorphic sites.

These results have implications for phylogenetic inference based on nrDNA data. As previously cautioned, it cannot be assumed that all copies of nrDNA are identical within a

genome. This is especially true for the spacer regions, but also for the subunit regions (Álvarez & Wendel, 2003). The discovery of phylogenetic signal in intragenomic polymorphism abundance demonstrates that those positions likely to lead to ambiguities are not distributed evenly across the phylogeny (Fig. 4). In addition to topology, variable polymorphism rates among lineages may affect the estimation of branch lengths on a tree. This may become especially problematic when nrDNA is used to date phylogenies. Recently developed methods of phylogenetic inference incorporating information about polymorphic positions may be able to alleviate difficulties in tree building caused by uneven levels of polymorphism abundance (Potts, Hedderson & Grimm, 2014).

Identification of mixed ancestry

Characterization of intragenomic nrDNA polymorphisms may allow for the identification of hybrid offspring between parents with differing nrDNA sequences (Zimmer, Jupe & Walbot, 1988), and may be able to provide an estimate of the number of generations since hybridization. This is especially true for early generation hybrids, as nrDNA homogenization can occur in a small number of generations (Kovarik et al., 2005), and mosaic *Saccharomyces* genomes have been identified based on intragenomic polymorphism abundance (West et al., 2014). Neither of the polymorphism profiles for the two wild-collected putative hybrid individuals in this study strongly indicate that they are early generation hybrids. This is in contrast to evidence from nuclear gene sequences that show heterozygosity consistent with inheritance of divergent alleles from the putative parents in the *A. albicans* × *A. subulata* hybrid (B. Haack and M. Fishbein, unpubl. data). Detection of mixed ancestry may be hampered in this case by a lack of fixed differences between parental haplotypes. Consensus sequences for the *A. albicans* and *A. subulata* individuals in this study show no fixed differences, while one difference is found between the *A. syriaca* and *A. speciosa* consensus sequences. Unexpectedly, this position is monomorphic in the hybrid, while the positions with minor allele frequencies >0.3 are in

positions not shown to differ between *A. syriaca* and *A. speciosa*. This likely indicates ancestry from an unsampled nrDNA haplotype.

Conclusions

Nuclear ribosomal DNA copies within individuals of *Asclepias* are not identical, with intragenomic polymorphisms present at a higher rate than reported for other organisms. Polymorphism frequencies across the genus vary by more than an order of magnitude and demonstrate strong phylogenetic signal. Stem positions of ribosomal subunits are more likely to be polymorphic than loop positions. Distribution of polymorphic sites across the nrDNA cistron are consistent with strong selection on nrDNA subunits, with polymorphic sites being more frequent in the spacer regions, and this difference being amplified for sites that are highly polymorphic. These results reinforce the need for caution when using nrDNA for phylogenetic inference, especially when using the spacer regions or for applications requiring the precise estimates of branch lengths or divergence times.

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LITERATURE CITED

Álvarez I, Wendel JF. 2003. Ribosomal ITS sequences and plant phylogenetic inference. *Plant Molecular Evolution* 29:417–434.

- Bai C, Alverson WS, Follansbee A, Waller DM. 2012. New reports of nuclear DNA content for 407 vascular plant taxa from the United States. *Annals of Botany* 110:1623–1629.
- Bainard JD, Bainard LD, Henry TA, Fazekas AJ, Newmaster SG. 2012. A multivariate analysis of variation in genome size and endoreduplication in angiosperms reveals strong phylogenetic signal and association with phenotypic traits. *New Phytologist* 196:1240–1250.
- Baldwin BG. 1992. Phylogenetic utility of the internal transcribed spacers of nuclear ribosomal DNA in plants: An example from the Compositae. *Molecular Phylogenetics and Evolution* 1:3–16.
- Baldwin BG, Sanderson MJ, Porter JM, Wojciechowski MF, Campbell CS, Donoghue MJ. 1995. The ITS region of nuclear ribosomal DNA: A valuable source of evidence on angiosperm phylogeny. *Annals of the Missouri Botanical Garden* 82:247–277.
- Bik HM, Fournier D, Sung W, Bergeron RD, Thomas WK. 2013. Intra-genomic variation in the ribosomal repeats of nematodes. *PLoS ONE* 8:e78230.
- Clark CG, Tague BW, Ware VC, Gerbi SA. 1984. *Xenopus laevis* 28S ribosomal RNA: A secondary structure model and its evolutionary and functional implications. *Nucleic Acids Research* 12:6197–6220.
- Delcher AL, Phillippy A, Carlton J, Salzberg SL. 2002. Fast algorithms for large-scale genome alignment and comparison. *Nucleic Acids Research* 30:2478–2483.
- Fishbein M, Chuba D, Ellison C, Mason-Gamer RJ, Lynch SP. 2011. Phylogenetic relationships of *Asclepias* (Apocynaceae) inferred from non-coding chloroplast DNA sequences. *Systematic Botany* 36:1008–1023.
- Fox J, Weisberg S. 2011. *An R Companion to Applied Regression*. Thousand Oaks, CA: Sage.
- Ganley ARD, Kobayashi T. 2007. Highly efficient concerted evolution in the ribosomal DNA repeats: Total rDNA repeat variation revealed by whole-genome shotgun sequence data. *Genome Research* 17:184–191.
- Garland T, Dickerman AW, Janis CM, Jones JA. 1993. Phylogenetic analysis of covariance by computer simulation. *Systematic Biology* 42:265–292.
- Gernandt DS, Liston A. 1999. Internal transcribed spacer region evolution in *Larix* and *Pseudotsuga* (Pinaceae). *American Journal of Botany* 86:711–723.

Gordon A. 2008. *FASTX toolkit*. Accessed June 6, 2011. Available at

http://hannonlab.cshl.edu/fastx_toolkit/index.html

Hamby RK, Zimmer EA. 1988. Ribosomal RNA sequences for inferring phylogeny within the grass family (Poaceae). *Plant Systematics and Evolution* 160:29–37.

Harmon LJ, Weir JT, Brock CD, Glor RE, Challenger W. 2008. GEIGER: Investigating evolutionary radiations. *Bioinformatics* 24:129–131.

Hillis DM, Dixon MT. 1991. Ribosomal DNA: Molecular evolution and phylogenetic inference.

Quarterly Review of Biology 66:411–453.

Knaus B. 2010. Short read toolbox. Accessed January 28, 2011. Available at <http://brianknaus.com>

Kolosha VO, Fodor I. 1990. Nucleotide sequence of *Citrus limon* 26S ribosomal-RNA gene and secondary structure model of its RNA. *Plant Molecular Biology* 14:147–161.

Kovarik A, Pires JC, Leitch AR, Lim KY, Sherwood AM, Matyasek R, Rocca J, Soltis DE, Soltis PS.

2005. Rapid concerted evolution of nuclear ribosomal DNA in two *Tragopogon* allopolyploids of recent and recurrent origin. *Genetics* 169:931–944.

Kurtz S, Phillippy A, Delcher AL, Smoot M, Shumway M, Antonescu C, Salzberg SL. 2004. Versatile and open software for comparing large genomes. *Genome Biol* 5:R12.

Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows-Wheeler transform.

Bioinformatics 25:1754–1760.

Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, 1000

Genome Project Data Processing Subgroup. 2009. The Sequence Alignment/Map format and SAMtools. *Bioinformatics* 25:2078–2079.

Lorenz R, Bernhart S, Honer zu Siederdissen C, Tafer H, Flamm C, Stadler P, Hofacker I. 2011.

ViennaRNA Package 2.0. *Algorithms for Molecular Biology* 6:26.

Maddison WP, Maddison DR. 2011. *Mesquite: a modular system for evolutionary analysis*. Available at <http://mesquiteproject.org>.

Maddison WP, Maddison DR, Midford PE. 2011. *Tree Farm package for Mesquite*. Available at

<http://mesquiteproject.org>

- Nguyen P, Ma J, Pei D, Obert C, Cheng C, Geiger T. 2011. Identification of errors introduced during high throughput sequencing of the T cell receptor repertoire. *BMC Genomics* 12:106.
- Pagel M. 1999. Inferring the historical patterns of biological evolution. *Nature* 401:877–884.
- Paradis E, Claude J, Strimmer K. 2004. APE: Analyses of phylogenetics and evolution in R language. *Bioinformatics* 20:289–290.
- Potts AJ, Hedderson TA, Grimm GW. 2014. Constructing phylogenies in the presence of intra-individual site polymorphisms (2ISPs) with a focus on the nuclear ribosomal cistron. *Systematic Biology* 63:1–16.
- Ratan A. 2009. Assembly algorithms for next-generation sequence data. PhD Dissertation Thesis. University Park, Pennsylvania, USA: Pennsylvania State University.
- R Core Team. 2014. *R: A language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing.
- Revell LJ. 2012. phytools: An R package for phylogenetic comparative biology (and other things). *Methods in Ecology and Evolution* 3:217–223.
- Rzhetsky A. 1995. Estimating substitution rates in ribosomal RNA genes. *Genetics* 141:771–783.
- Schlötterer C, Tautz D. 1994. Chromosomal homogeneity of *Drosophila* ribosomal DNA arrays suggests intrachromosomal exchanges drive concerted evolution. *Current Biology* 4:777–783.
- Simon UK, Trajanoski S, Kroneis T, Sedlmayr P, Guelly C, Guttenger H. 2012. Accession-specific haplotypes of the internal transcribed spacer region in *Arabidopsis thaliana*—A means for barcoding populations. *Molecular Biology and Evolution* 29:2231–2239.
- Stage DE, Eickbush TH. 2007. Sequence variation within the rRNA gene loci of 12 *Drosophila* species. *Genome Research* 17:1888–1897.
- Straub S, Fishbein M, Livshultz T, Foster Z, Parks M, Weitemier K, Cronn R, Liston A. 2011. Building a model: Developing genomic resources for common milkweed (*Asclepias syriaca*) with low coverage genome sequencing. *BMC Genomics* 12:211.
- Straub SCK, Parks M, Weitemier K, Fishbein M, Cronn RC, Liston A. 2012. Navigating the tip of the genomic iceberg: Next-generation sequencing for plant systematics. *American Journal of Botany* 99:349–364.

Venables WN, Ripley BD. 2002. *Modern Applied Statistics with S*. New York: Springer.

Weitemier K, Straub SCK, Cronn RC, Fishbein M, Schmickl R, McDonnell A, Liston A. 2014. Hyb-Seq: Combining target enrichment and genome skimming for plant phylogenomics. *Applications in Plant Sciences*:1400042.

West C, James SA, Davey RP, Dicks J, Roberts IN. 2014. Ribosomal DNA sequence heterogeneity reflects intraspecies phylogenies and predicts genome structure in two contrasting yeast species. *Systematic Biology* 63:543–554.

Zimmer EA, Jupe ER, Walbot V. 1988. Ribosomal gene structure, variation and inheritance in maize and its ancestors. *Genetics* 120:1125–1136.

Table 1 (on next page)

Polymorphic site abundance in *Asclepias*

Polymorphic site abundance in *Asclepias* taxa. **Voucher:** Collector, collection #, [herbarium].

Poly #: Number of polymorphic positions. **Poly %:** Percentage of the assembled positions that are polymorphic. **High #, High %:** Number and percentage of highly polymorphic positions. **SRA:** NCBI Sequence Read Archive accession number.

Asclepias taxon	Voucher	Poly #	Poly %	High #	High %	SRA
<i>A. albicans</i> S. Watson	Fishbein 3146 [WS]	174	3.12	16	0.29	SRX000000
<i>A. albicans</i>	Fishbein 6463 [OKLA]	273	4.70	11	0.19	
<i>A. alticola</i> E. Fourn.	Steinmann 5243 [IEB]	668	11.45	31	0.53	
<i>A. amplexicaulis</i> Sm.	Lynch 12652 [OKLA]	248	4.25	25	0.43	
<i>A. angustifolia</i> Schweigg.	Reina 2004-1315 [ARIZ]	55	1.04	36	0.68	
<i>A. angustifolia</i>	Reina 2008-203 [OKLA]	169	2.99	26	0.46	
<i>A. arenaria</i> Torr.	Lynch 11495 [OKLA]	174	2.98	2	0.03	
<i>A. asperula</i> (Decne.) Woodson ssp. <i>asperula</i>	Lynch 12037 [OKLA]	342	5.87	7	0.12	
<i>A. asperula</i> ssp. <i>asperula</i>	Fishbein 6536 [OKLA]	391	6.73	24	0.41	
<i>A. asperula</i> ssp. <i>capricornu</i> (Woodson) Woodson	Lynch 13314 [OKLA]	474	8.15	38	0.65	
<i>A. asperula</i> ssp. <i>capricornu</i>	Fishbein 6486 [OKLA]	347	5.95	6	0.10	
<i>A. atrovioleacea</i> Woodson	Fishbein 3612 [ARIZ]	451	7.73	26	0.45	
<i>A. auriculata</i> Kunth	Lynch 1694 [OKLA]	513	8.85	33	0.57	
<i>A. auriculata</i>	Fishbein 5833 [OKLA]	378	6.54	39	0.68	
<i>A. boliviensis</i> E. Fourn.	Fishbein 6072 [OKLA]	875	15.08	111	1.91	
<i>A. brachystephana</i> Engelm. ex Torr.	Lynch 10642 [OKLA]	309	5.32	22	0.38	
<i>A. californica</i> Greene	Lynch 10779 [OKLA]	472	8.09	24	0.41	
<i>A. aff. candida</i> Vell.	Fishbein 6347 [OKLA]	245	4.20	14	0.24	
<i>A. cinerea</i> Walter	Fishbein 4793 [OKLA]	297	5.10	27	0.46	
<i>A. circinalis</i> (Decne.) Woodson	Webster 17186 [OKLA]	464	7.95	59	1.01	
<i>A. connivens</i> Baldwin ex Elliott	Lynch 12336 [OKLA]	394	6.75	16	0.27	
<i>A. cordifolia</i> (Benth.) Jeps.	Lynch 10942 [OKLA]	344	5.96	30	0.52	
<i>A. cordifolia</i>	Fishbein 5772 [OKLA]	308	5.35	13	0.23	
<i>A. coulteri</i> A. Gray	Ventura & Lopez 7986 [TEX]	471	8.09	27	0.46	
<i>A. cryptoceras</i> S. Watson ssp. <i>cryptoceras</i>	Fishbein 6504 [OKLA]	230	4.00	35	0.61	
<i>A. cryptoceras</i> ssp. <i>davisii</i> (Woodson) Woodson	Fishbein 5723 [OKLA]	350	6.00	13	0.22	
<i>A. curassavica</i> L.	Zuloaga & Morrone 7087 [OKLA]	258	4.42	24	0.41	
<i>A. cutleri</i> Woodson	Fishbein 6511 [OKLA]	167	2.90	18	0.31	
<i>A. cutleri</i>	Fishbein 6500 [OKLA]	157	2.69	8	0.14	
<i>A. emoryi</i> (Greene) Vail ex Small	Carr 12032 [TEX]	124	2.29	44	0.81	
<i>A. engelmanniana</i> Woodson	Lynch 11224 [OKLA]	141	2.54	33	0.59	

<i>A. engelmanniana</i>	Lynch 11029 [OKLA]	192	3.30	7	0.12	
<i>A. eriocarpa</i> Benth.	Lynch 10923 [OKLA]	610	10.49	32	0.55	
<i>A. eriocarpa</i>	Lynch 10799 [OKLA]	492	8.43	17	0.29	
<i>A. feayi</i> Chapm. ex A. Gray	Fishbein 5586 [OKLA]	329	5.71	42	0.73	
<i>A. fournieri</i> Woodson	Fishbein 3660 [ARIZ]	556	9.58	47	0.81	
<i>A. fournieri</i>	Lynch 1655 [OKLA]	711	12.25	49	0.84	
<i>A. glaucescens</i> Kunth	Lynch 14142 [OKLA]	190	3.33	19	0.33	
<i>A. glaucescens</i>	Lynch 1623 [OKLA]	90	1.69	85	1.60	
<i>A. glaucescens</i>	Fishbein 5097 [OKLA]	215	3.72	18	0.31	
<i>A. sp. nov. aff. glaucescens</i>	Fishbein 3671 [ARIZ]	225	3.87	25	0.43	
<i>A. hallii</i> A. Gray	Lynch 11299 [OKLA]	600	10.36	54	0.93	
<i>A. humistrata</i> Walter	Fishbein 5596 [OKLA]	331	5.67	13	0.22	
<i>A. hypoleuca</i> (A. Gray) Woodson	Lynch 11374 [OKLA]	670	11.51	44	0.76	
<i>A. incarnata</i> L.	Lynch 12567 [OKLA]	434	7.45	28	0.48	
<i>A. involucrata</i> Engelm. ex Torr.	Lynch 12050 [OKLA]	326	5.65	35	0.61	
<i>A. involucrata</i>	Fishbein 6531 [OKLA]	217	3.72	13	0.22	
<i>A. jaliscana</i> B.L. Rob.	Fishbein 2493 [ARIZ]	140	2.97	74	1.57	
<i>A. jaliscana</i>	Fishbein 3657 [WS]	165	3.00	65	1.18	
<i>A. jorgeana</i> Fishbein & S.P. Lynch	Vásquez & Alvarez 4905 [IEB]	160	2.76	12	0.21	
<i>A. lanceolata</i> Walter	Fishbein 5605 [MISSA]	249	4.27	8	0.14	
<i>A. lanuginosa</i> Nutt.	Lynch 12661 [OKLA]	343	5.92	11	0.19	
<i>A. latifolia</i> (Torr.) Raf.	Lynch 11018 [OKLA]	236	4.05	11	0.19	
<i>A. lemmonii</i> A. Gray	Lynch 11453 [OKLA]	620	10.70	23	0.40	
<i>A. leptopus</i> I.M. Johnst.	Fishbein 6263 [OKLA]	282	4.84	9	0.15	
<i>A. longifolia</i> Michx.	Lynch 12447 [OKLA]	427	7.34	24	0.41	
<i>A. lynchiana</i> Fishbein	Venable & Becerra s.n. [ARIZ]	129	2.39	31	0.57	
<i>A. macrosperma</i> Eastw.	Gierisch 4191 [ARIZ]	76	1.42	30	0.56	
<i>A. macrosperma</i>	Fishbein 6518 [OKLA]	391	6.72	21	0.36	
<i>A. macrotis</i> Torr.	Lynch 11260 [OKLA]	364	6.27	11	0.19	
<i>A. macrotis</i>	Lynch 11263 [OKLA]	260	4.46	5	0.09	
<i>A. masonii</i> Woodson	Fishbein 3101 [OKLA]	151	2.59	7	0.12	
<i>A. meadii</i> Torr. ex A. Gray	Freeman 9106 [KANU]	208	3.62	20	0.35	
<i>A. mellodora</i> A. St.-Hil.	Zuloaga & Morrone 7168 [OKLA]	377	6.47	18	0.31	
<i>A. mexicana</i> Cav.	Fishbein 3009 [ARIZ]	186	3.22	18	0.31	
<i>A. michauxii</i> Decne.	Lynch 12316 [OKLA]	458	7.87	39	0.67	
<i>A. notha</i> W.D. Stevens	Lynch 14113 [OKLA]	375	6.45	44	0.76	
<i>A. notha</i>	Fishbein 5389 [OKLA]	249	4.31	41	0.71	
<i>A. notha</i>	Nee 32966 [NY]	432	7.47	17	0.29	
<i>A. sp. nov. cf. notha</i>	Fishbein 5816 [OKLA]	376	6.45	16	0.27	

<i>A. nyctaginifolia</i> A. Gray	Fishbein 6268 [OKLA]	109	1.92	23	0.40
<i>A. obovata</i> Elliott	Lynch 11543 [OKLA]	708	12.75	87	1.57
<i>A. oenotheriodes</i> Schltdl. & Cham.	Fishbein 5819 [OKLA]	230	3.95	15	0.26
<i>A. oenotheroides</i>	Lynch 13339 [OKLA]	100	1.74	16	0.28
<i>A. oenotheroides</i>	Lynch 11477 [OKLA]	134	2.30	7	0.12
<i>A. otarioides</i> E. Fourn.	Bellsey 97-5 [ARIZ]	745	12.80	35	0.60
<i>A. otarioides</i>	Lynch 1533 [OKLA]	659	11.32	20	0.34
<i>A. otarioides</i>	Fishbein 5857 [OKLA]	697	11.95	48	0.82
<i>A. ovalifolia</i> Decne.	Lynch 13546 [OKLA]	333	5.91	42	0.75
<i>A. ovata</i> M. Martens & Galeotti	Laferrière 1478 [MO]	324	5.59	39	0.67
<i>A. pellucida</i> E. Fourn.	Fishbein 5136 [OKLA]	439	7.53	9	0.15
<i>A. perennis</i> Walter	Lynch 12408 [OKLA]	356	6.11	15	0.26
<i>A. pilgeriana</i> Schltr. ("flava" in Fishbein et al., 2011)	Zuloaga & Morrone 7069 [OKLA]	458	7.85	23	0.39
<i>A. pratensis</i> Benth.	Fishbein 5143 [OKLA]	410	7.11	29	0.50
<i>A. pratensis</i>	Pérez 1850 [MO]	609	10.49	34	0.59
<i>A. prostrata</i> W.H. Blackw.	Fishbein 2432 [ARIZ]	465	7.99	41	0.70
<i>A. purpurascens</i> L.	Lynch 12847 [OKLA]	233	4.05	22	0.38
<i>A. purpurascens</i>	Fishbein 5654 [MISSA]	216	3.73	7	0.12
<i>A. quadrifolia</i> Jacq.	Webb s.n. [ARIZ]	116	2.33	41	0.82
<i>A. quadrifolia</i>	Fishbein 6545 [OKLA]	204	3.55	31	0.54
<i>A. rosea</i> Kunth	Lynch 1656 [OKLA]	747	12.86	33	0.57
<i>A. ruthiae</i> Maguire	Riser 329 [WS]	31	0.61	23	0.45
<i>A. sanjuanensis</i> K.D. Heil, J.M. Porter & S.L. Welsh	Ellison s.n. [HPSU]	158	2.96	33	0.62
<i>A. sanjuanensis</i>	Fishbein 6525 [OKLA]	237	4.11	38	0.66
<i>A. sanjuanensis</i>	Riser 335 [WS]	68	1.18	26	0.45
<i>A. scaposa</i> Vail	Fishbein 2951 [ARIZ]	500	8.61	35	0.60
<i>A. schaffneri</i> A. Gray	Fishbein 5846 [OKLA]	327	5.65	36	0.62
<i>A. scheryi</i> Woodson	Fishbein 5137 [OKLA]	586	10.05	15	0.26
<i>A. scheryi</i>	Zamudio 5234 [MEXU]	272	4.88	35	0.63
<i>A. similis</i> Hemsl.	Fishbein 3000 [ARIZ]	294	5.05	8	0.14
<i>A. similis</i>	Fishbein 5148 [MISSA]	386	6.71	45	0.78
<i>A. solanoana</i> Woodson	Lynch 10884 [OKLA]	882	15.13	35	0.60
<i>A. speciosa</i> Torr.	Lynch 10981 [OKLA]	242	4.19	23	0.40
<i>A. aff. standleyi</i> Woodson	Reina 98-579 [WS]	150	2.57	19	0.33
<i>A. subaphylla</i> Woodson	Fishbein 3518 [WS]	182	3.31	23	0.42
<i>A. subaphylla</i>	Lynch 1008 [OKLA]	383	6.86	77	1.38
<i>A. subulata</i> Decne.	Fishbein 6434 [OKLA]	269	4.64	7	0.12
<i>A. subulata</i>	Fishbein 6446 [OKLA]	370	6.36	18	0.31
<i>A. subulata</i> × <i>albicans</i>	Fishbein 3142 [WS]	299	5.14	12	0.21

<i>A. subverticillata</i> (A. Gray) Vail	Fishbein 2948 [ARIZ]	160	3.09	92	1.77	
<i>A. syriaca</i> L.	Lynch 11138 [OKLA]	204	3.51	11	0.19	
<i>A. syriaca</i>	Fishbein 4885 [OKLA]	298	5.10	7	0.12	SRX040889
<i>A. syriaca</i> × <i>speciosa</i>	Fishbein 2810 [ARIZ]	161	2.89	37	0.66	
<i>A. tomentosa</i> Elliott	Fishbein 5608 [MISSA]	198	3.39	11	0.19	
<i>A. tuberosa</i> L. ssp. <i>interior</i> Woodson	Fishbein 2816 [ARIZ]	685	11.84	29	0.50	
<i>A. tuberosa</i> ssp. <i>interior</i>	Fishbein 4825 [MISSA]	297	5.10	31	0.53	
<i>A. tuberosa</i> ssp. <i>rolfsii</i> (Britton ex Vail) Woodson	Lynch 12526 [OKLA]	251	4.33	11	0.19	
<i>A. uncialis</i> Greene	Fishbein 6494 [OKLA]	282	4.86	19	0.33	
<i>A. variegata</i> L.	Lynch 12787 [OKLA]	375	6.50	29	0.50	
<i>A. verticillata</i> L.	Lynch 11102 [OKLA]	23	0.41	21	0.37	
<i>A. vestita</i> Hook. & Arn. ssp. <i>parishii</i> (Jeps.) Woodson	Lynch 10735 [OKLA]	506	8.68	14	0.24	
<i>A. viridis</i> Walter	Lynch 12955 [OKLA]	261	4.55	40	0.70	
<i>A. viridula</i> Chapm.	Fishbein 4806 [MISSA]	425	7.37	46	0.80	
<i>A. welshii</i> N.H. Holmgren & P.K. Holmgren	Lynch 11369 [OKLA]	364	6.53	73	1.31	
<i>A. woodsoniana</i> Standl. & Steyerl.	D. A. Neil 242 [MO]	122	2.28	30	0.56	

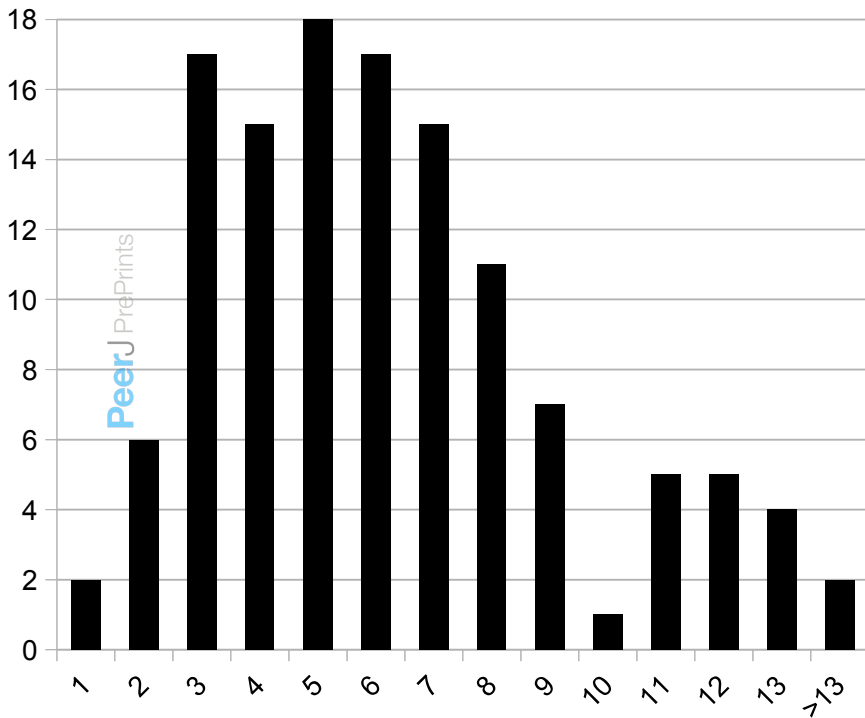
Figure 1 (on next page)

Polymorphic site frequency among species of *Asclepias*.

Histogram of polymorphic site frequency among species of *Asclepias*. Individuals contained from 0.4% to 15.1% polymorphic sites.

Number of Individuals

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Percent polymorphic positions

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Figure 2_(on next page)

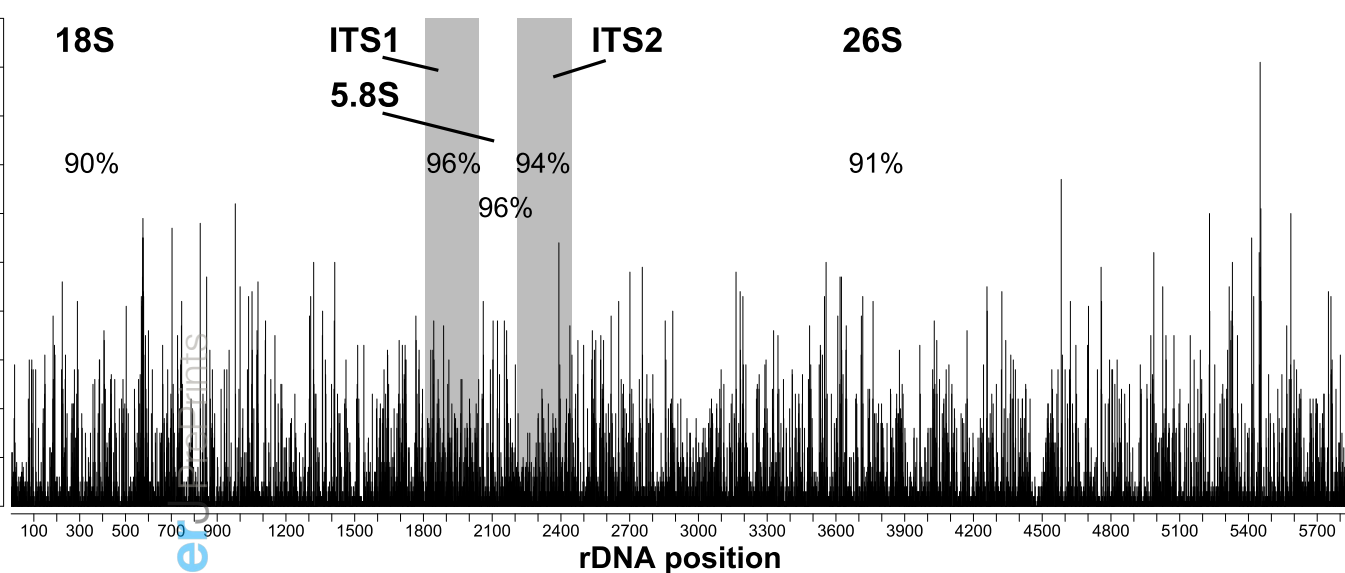
Polymorphic sites across the nrDNA cistron of *Asclepias*.

Number of individuals that are A) polymorphic and B) highly polymorphic at each position.

Polymorphic positions are those with $\geq 2\%$ of reads differing from the consensus; highly polymorphic positions are those with $\geq 10\%$ differing reads. Subunit regions, white background; spacer regions, shaded background. Numbers in each region are the percentage of sites polymorphic or highly polymorphic in at least one individual.

A

Number of polymorphic individuals

**B**

Number of highly polymorphic individuals

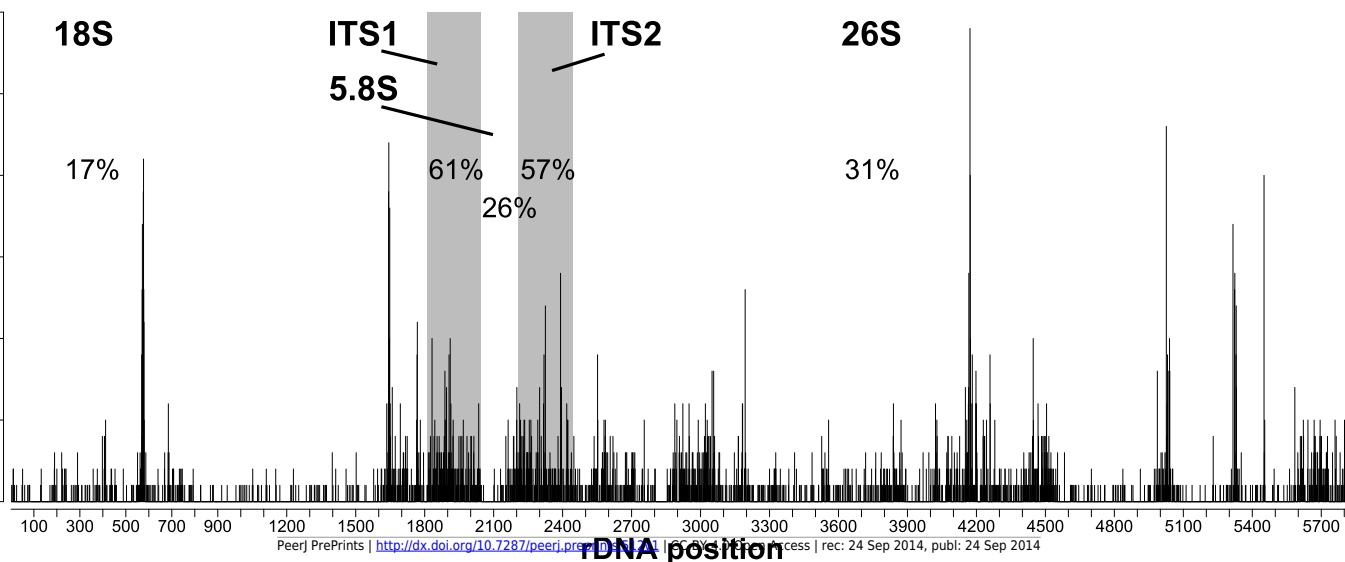


Figure 3_(on next page)

Polymorphism probability by region and structure.

Probability that at least one individual is A) polymorphic or B) highly polymorphic at a position that is either within a spacer (ITS1, ITS2) or subunit region (18S, 5.8S, 26S), and either paired (stems) or unpaired (loops). Error bars indicate 95% confidence intervals. Values derived from two-factor multiple logistic regressions (Table 3).

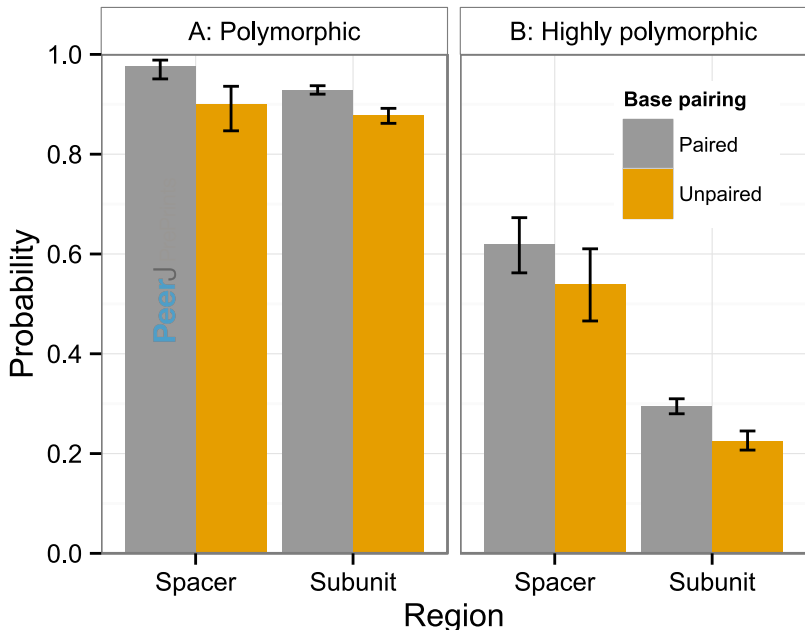


Table 2_(on next page)

ANOVA of the number of polymorphic individuals at nrDNA positions, by position type.

Two-way ANOVA of the number of polymorphic individuals at nrDNA positions categorized as either subunit (18S, 5.8S, 26S) or spacer regions (ITS1, ITS2), and as either paired (stems) or unpaired (loops). More individuals are likely to be polymorphic at sites that are in spacer regions over subunit regions, and that are paired over unpaired. **Bold** values indicate categories that significantly affect polymorphism abundance ($P < 0.05$). *df* = degrees of freedom.

Source of variation	Sum of Squares	<i>df</i>	<i>F</i> value	<i>P</i>
(Intercept)	8310.6	1		
Paired	109.1	1	53.5887	<0.0001
Subunit	16.4	1	8.0664	0.0045
Paired*Subunit	0.4	1	0.1859	0.6663
Residuals	11878.9	5835		

Table 3_(on next page)

Multiple logistic regression of the likelihood of nrDNA position polymorphism, by position type.

Two-factor multiple logistic regression of the likelihood of nrDNA positions being A) polymorphic or B) highly polymorphic in at least one individual. Positions are categorized as either within a subunit (18S, 5.8S, 26S) or spacer region (ITS1, ITS2), and as either paired (stem) or unpaired (loop). Odds ratios indicate whether a category decreases (<1) or increases (>1) the likelihood a position is polymorphic or highly polymorphic. The intercept represents paired, spacer positions. Categories that significantly affect polymorphism likelihood are indicated by *italics* ($P<0.1$) or **boldface** ($P<0.05$). CI = confidence interval, Std. Error = standard error. Polymorphism probabilities for each category are presented in Fig. 3.

A: Polymorphic

Source	Odds Ratio	95% CI	Coefficient estimate	Std. Error	Z-value	P
(Intercept)			3.7136	0.3825		
Subunit	0.320	0.150 - 0.686	-1.1382	0.3881	-2.933	0.0034
Unpaired	0.220	0.090 - 0.537	-1.5163	0.4561	-3.324	0.0009
<i>Subunit*Unpaired</i>	2.488	0.998 - 6.206	0.9115	0.4663	1.955	0.0506

B: Highly polymorphic

Source	Odds Ratio	95% CI	Coefficient estimate	Std. Error	Z-value	P
(Intercept)			0.48551	0.1201		
Subunit	0.257	0.201 - 0.329	-1.35881	0.12564	-10.815	<0.0001
<i>Unpaired</i>	0.719	0.494 - 1.047	-0.32964	0.19178	-1.719	0.0856
Subunit*Unpaired	0.970	0.651 - 1.444	-0.03061	0.20315	-0.151	0.8802

Table 4_(on next page)

Phylogenetic signal tests across *Asclepias* for the number of polymorphic or highly polymorphic sites.

Tests for phylogenetic signal across *Asclepias* for the number of polymorphic or highly polymorphic positions across the entire nrDNA cistron (Subunits + spacers) or just the subunits (Subunits only). **Parsimony permutations**: The proportion of permutations with a shorter tree length than the true data. **Lambda**: The maximum likelihood estimate of lambda. **logL**: The log-likelihood ratio of the unconstrained model including the estimated lambda over the constrained model with lambda=0. **P**: The probability of obtaining a likelihood ratio this small or smaller by chance alone.

		Parsimony permutations	Lambda	$\log L$	P
Polymorphic sites	Subunits + spacers	<0.0012	0.51	$\frac{-594.192}{-597.863}$	0.0067
	Subunits only	<0.0200	0.45	$\frac{-587.547}{-590.765}$	0.0112
Highly polymorphic sites	Subunits + spacers	>0.6050	<0.0001	$\frac{-387.226}{-387.225}$	1
	Subunits only	>0.5300	<0.0001	$\frac{-376.150}{-376.149}$	1

Figure 4_(on next page)

Ancestral state reconstruction of polymorphic site abundance.

Ancestral state reconstruction of the number of polymorphic positions in nrDNA in *Asclepias* obtained with squared-change parsimony. The tree topology is that pruned from Fig. 2 of Fishbein et al. (2011) with clades indicated by letters, following that study.

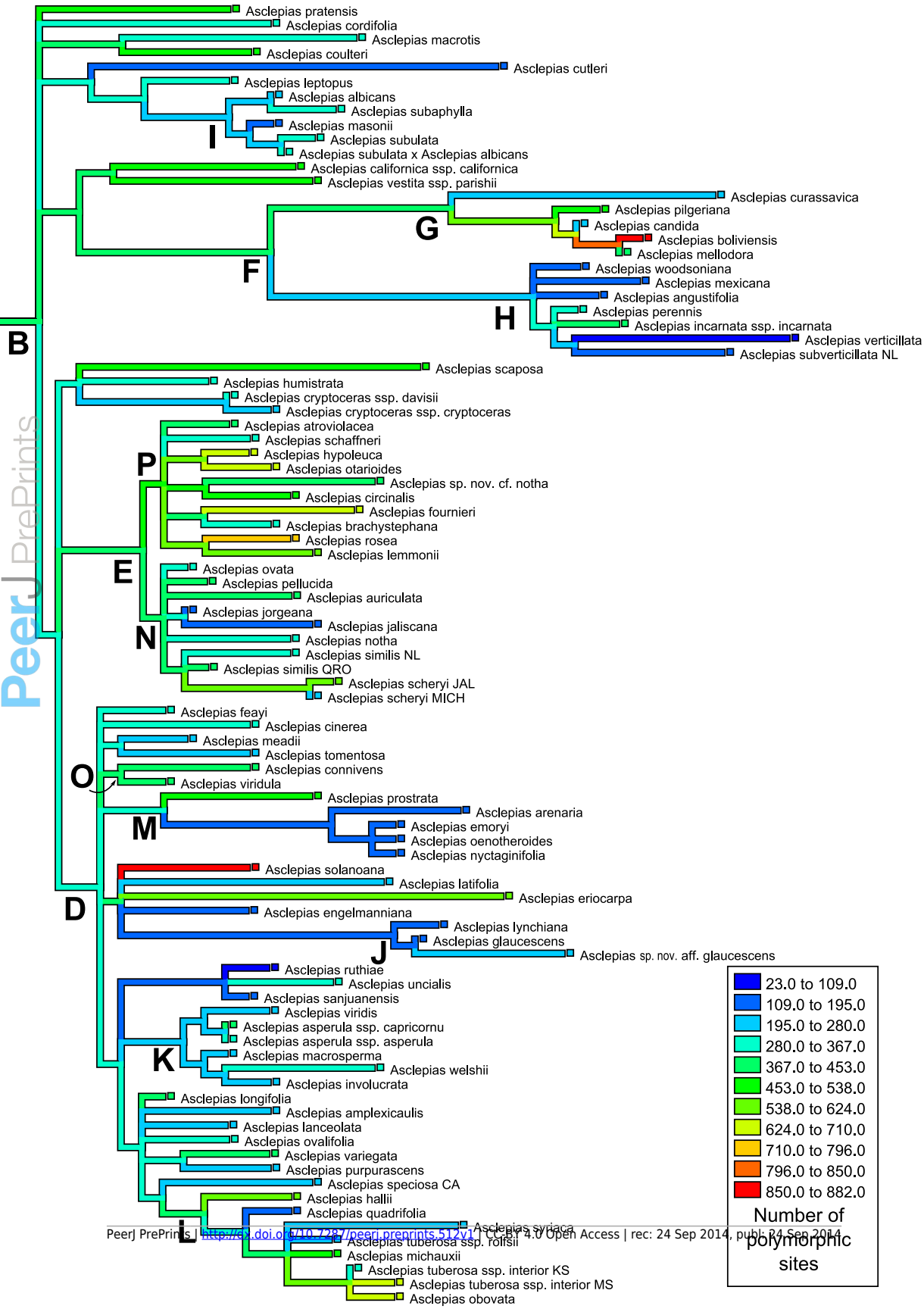


Figure 5_(on next page)

Ancestral state reconstruction of highly polymorphic site abundance.

Ancestral state reconstruction of the number of highly polymorphic positions in nrDNA in *Asclepias* obtained with squared-change parsimony. The tree topology is that pruned from Fig. 2 of Fishbein et al. (2011) with clades indicated by letters, following that study.

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