A peer-reviewed version of this preprint was published in PeerJ on 7 October 2014.

<u>View the peer-reviewed version</u> (peerj.com/articles/612), which is the preferred citable publication unless you specifically need to cite this preprint.

Chandrasekera NS, Bailey MA, Files M, Alling T, Florio SK, Ollinger J, Odingo JO, Parish T. 2014. Synthesis and anti-tubercular activity of 3substituted benzo[b]thiophene-1,1-dioxides. PeerJ 2:e612 https://doi.org/10.7717/peerj.612

Synthesis and anti-tubercular activity of 3-substituted benzothiophene-1,1-dioxides

We demonstrated that the 3-substituted benzothiophene-1,1-dioxide class of compounds are effective inhibitors of Mycobacterium tuberculosis growth under aerobic conditions. We examined substitution at the C-3 position of the benzothiophene-1,1dioxide series systematically to delineate structure-activity relationships influencing potency and cytotoxicity. Compounds were tested for inhibitory activity against virulent M. tuberculosis and eukaryotic cells. The tetrazole substituent was most potent, with a minimum inhibitory concentration (MIC) of 2.6 μ M. However, cytotoxicity was noted with even more potency (Vero cell TC50 = 0.1 μ M). Oxadiazoles had good anti-tubercular activity (MICs of 3–8 μ M), but imidazoles, thiadiazoles and thiazoles had little activity. Cytotoxicity did not track with antitubercular activity, suggesting different targets or mode of action between bacterial and eukaryotic cells. However, we were unable to derive analogs without cytotoxicity; all compounds synthesized were cytotoxic (TC50 of 0.1–5 μ M). We conclude that cytotoxicity is a liability in this series precluding it from further development. However, the series has potent anti-tubercular activity and future efforts towards identifying the mode of action could result in the identification of novel drug targets.

2 Synthesis and anti-tubercular activity of 3-substituted benzo[b]thiophene-1,1-

3 dioxides

- 4 Authors
- 5 N. Susantha Chandrasekera, Mai A. Bailey, Megan Files, Torey Alling, Stephanie K. Florio,
 6 Juliane Ollinger, Joshua O. Odingo, and Tanya Parish*
- 7 Affiliations
- 8 TB Discovery Research, Infectious Disease Research Institute, 1616 Eastlake Avenue E, Seattle,
- 9 WA 98102, USA.
- 10 *Corresponding author. TB Discovery Research, Infectious Disease Research Institute, 1616
- 11 Eastlake Avenue E, Seattle, WA 98102, USA.
- 12 Tel.: 206 858 6074; Fax: 206 381 3678
- 13 *E-mail address:* tanya.parish@idri.org.
- 14 Keywords: Benzo[b]thiophene-1,1-dioxide, Tuberculosis, Antimicrobial

16 Tuberculosis (TB), which is caused by *Mycobacterium tuberculosis* is the second leading cause 17 of death from an infectious disease and is a major global health problem. In 2010, according to 18 the World Health Organization (WHO) 8.8 million new cases and 1.4 million deaths from the 19 disease were reported [1]. In addition, one third of the world population has latent TB, 10% of 20 whom are expected to develop active TB at some point in their lives. Currently the recommended 21 first-line TB treatment regimens require a minimum of 6 months of multidrug therapy, resulting 22 in challenges with patient adherence. The result of inadequate therapy and poor compliance has 23 contributed to a rise in the emergence of multidrug resistant (MDR), resistant to isoniazid and 24 rifampicin, and extensively drug-resistant (XDR) strains, resistant to a fluoroquinolone and at 25 least one injectable drug, of *M. tuberculosis* [1]. Consequently, there is an urgent need for the 26 development of novel anti-TB drugs that are effective against both drug sensitive and resistant M. 27 tuberculosis $[\underline{2}]$.

The benzo[b]thiophene-1,1-dioxide (BTD) series was reported to have activity against *M. tuberculosis* in a phenotypic assay [3]. Fourteen compounds were tested from this series; five of these, all of which had heteroarylthio groups, had some inhibitory activity against *M. tuberculosis.* As a part of our ongoing TB drug discovery program, we were interested in exploring the potential of the BTD series to be developed as a lead series for TB treatment. We conducted an exploratory chemistry study and evaluated the series for their activity against *M. tuberculosis* as well as cytotoxicity for eukaryotic cells.

35 Materials and Methods

36 Determination of minimum inhibitory concentration (MIC)

37 We used *M. tuberculosis* H37Rv (London Pride), a laboratory-passaged derivative of H37Rv 38 (ATCC 25618), which has been sequenced, as described in [5]. MICs were run as described [4]; 39 briefly MICs were determined against *M. tuberculosis* grown in Middlebrook 7H9 medium 40 containing 10% OADC (oleic acid, albumin, dextrose, catalase) supplement (Becton Dickinson) 41 and 0.05% w/v Tween 80 (7H9-Tw-OADC) under aerobic conditions. Compounds were prepared 42 as 10-point two-fold serial dilutions in DMSO with a starting concentration of 20 µM (lowest 43 compound concentration 40 nM). The final concentration of DMSO in the assay was 2%. 44 Bacterial growth was measured by OD₅₉₀ after 5 days of incubation at 37°C and % growth 45 measured. Growth inhibition curves were plotted and fitted using the Gompertz model. The MIC 46 was defined as the minimum concentration required for >99% growth inhibition.

47 Vero cytotoxicity assay

48 CellTiter-Glo® Luminescent Cell Viability Assay (Promega) was used to measure ATP as a indicator of cell viability. The Vero cell line (ATCC CCL81) was grown in Dulbecco's Modified 49 50 Eagle Medium (DMEM), High Glucose, GlutaMAXTM (Invitrogen), 10% FBS (Fetal Bovine 51 Serum), and 1x of Penicillin-Streptomycin Solution (100 units/mL of penicillin, 100 µg/mL of 52 streptomycin). Compounds were solubilized in DMSO (dimethyl sulfoxide) and assayed using a 53 10-point three-fold serial dilution starting at the highest concentration of 50 µM. CellTiter-Glo® 54 Reagent (Promega) was added to 96-well plates after 2 days of incubation at 37°C, 5% CO₂. 55 Relative luminescent units (RLU) were measured using Perkin Elmer Wallac 1420 Victor2 plate 56 reader. Inhibition curves were fitted using the Levenberg-Marquardt algorithm. Toxic 57 concentration (TC₅₀)was defined as the concentration of compound that gave 50% inhibition of 58 growth. Selectivity index was calculated as MIC/TC₅₀. For published data [3], SI was calculated 59 as IC₉₀/TC₅₀.

60 Analysis of compounds

61 ¹H and NMR spectral data were recorded in CDCl₃ or Acetone-d6 on a 300 MHz Bruker NMR 62 spectrometer. Column chromatography was conducted on a Revelaris flash chromatography 63 system. Reactions were monitored using thin-layer chromatography (TLC) on silica gel plates. 64 HPLC analysis was conducted on an Agilent 1100 series LC system (Agilent ChemStation 65 Rev.A.10.02; Phenomenex-Luna-C18, 4.8 mm \times 150 mm, 5 μ m, 1.0 mL/min, UV 254nm, room 66 temperature) with MeCN/H₂O (0.05% TFA or HCOOH buffer) gradient elution. HPLC-MS was 67 performed on a Gilson 321 HPLC with detection performed by a Gilson 170 DAD and a Finnigan 68 AQA mass spectrometer operating in electrospray ionisation mode using a Phenomenex Gemini 69 C18 150x4.6mm column. Compounds 3a, b, c, s, t and u were purchased from ChemBridge 70 Corporation.

71 Synthesis of 3-bromobenzo[b]thiophene 1,1-dioxide (2)

To a solution of 1 1.62 g (7.6 mmol) in 25.0 mL in acetic acid was added 30% aqueous hydrogen
peroxide and the mixture was heated for 1 h at 100°C. The mixture was poured into ice cold water
and let it stand overnight. The resulting solid was filtered and dried to yield 2 (1.65 g, 89%). ¹H
NMR (300 MHz, CDCl₃): δ 6.98 (s, 1H), 7.58 - 7.72 (m, 4H). LCMS – ESI (M+H)⁺: 214.1.

76 General procedure for the synthesis of 3-substituted benzo[b]thiophene-1,1-dioxides

To a solution of 200 mg (0.82 mmol) of 2 in 5 mL of dimethyl formamide was added 2.0 mmol of the thiol reagent followed by 0.5 mL of triethylamine. The reaction was stirred overnight and washed with 20 mL of deionized water and extracted with 50 mL of ethyl acetate. The organic layer was dried with anhydrous sodium sulfate, filtered and concentrated *in vacuo*. The resulting residue was purified by reveleris flash chromatography system to yield the aryl/heteroaryl thio benzo[b]thiophene 1,1-dioxides. 89

3-((5-(4-methoxyphenyl)-1,3,4-oxadiazol-2-yl)thio)benzo[b]thiophene 1,1-dioxide (3d) Yield 3d: (95 mg, 31%). ¹H NMR (300 MHz, CDCl₃): 3.9 (3H, OCH₃. s); 7.0 – 8.0 (m, 9H). LCMS – ESI (M+H)⁺: 373.0.

3-((5-(4-chlorophenyl)-1,3,4-oxadiazol-2-yl)thio)benzo[b]thiophene 1,1-dioxide (3e)
Yield 3e: (122 mg, 39%). ¹H NMR (300 MHz, Methanol-d₄): 7.5 – 8.1 (m, 8H). LCMS – ESI
(M+H)⁺: 377.0.

90 Yield 3f: (87 mg, 31%). ¹H NMR (300 MHz, CDCl₃): 6.6 (1H, s); 7.6 - 8.1 (m, 6H). LCMS - ESI
91 (M+H)⁺: 282.0.

92 3-((4-phenylthiazol-2-yl)thio)benzo[b]thiophene 1,1-dioxide (3g)

3-(thiazol-2-ylthio)benzo[b]thiophene 1,1-dioxide (3f)

93 Yield 3g: (25 mg, 9%). ¹H NMR (300 MHz, CDCl₃): 7.7 - 8.0 (m, 10H). LCMS - ESI (M+H)⁺:
94 358.0.

95 3-(benzo[d]thiazol-2-ylthio)benzo[b]thiophene 1,1-dioxide (3h)

96 Yield **3h**: (65 mg, 32%). ¹H NMR (300 MHz, CDCl₃): 7.3 – 8.1 (m, 9H). LCMS – ESI (M+H)⁺:
97 332.0.

98 3-((5-chloro-3a,7a-dihydrobenzo[d]thiazol-2-yl)thio)benzo[b]thiophene 1,1-dioxide (3i)

99 Yield 3i: (25 mg, 30%). ¹H NMR (300 MHz, CDCl₃): 7.4 (1H, s); 7.5 – 8.1 (m, 7H). LCMS – ESI

100 $(2M+H_2O)^+$: 754.9.

101 3-((6-ethoxybenzo[d]thiazol-2-yl)thio)benzo[b]thiophene 1,1-dioxide (3j)

- **102** Yield **3j**: (105 mg, 28%). ¹H NMR (300 MHz, CDCl₃): 1.5 (3H, d); 4.1 (2H, t); 7.1 7.9 (m, 8H).
- **103** LCMS ESI $(M+H)^+$: 376.0.

104 3-((5-methyl-1,3,4-thiadiazol-2-yl)thio)benzo[b]thiophene 1,1-dioxide (3k)

105 Yield 3k: (26 mg, 11%). ¹H NMR (300 MHz, CDCl₃): 2.6 (3H, CH₃, s); 7.7 - 8.0 (m, 9H). LCMS
106 - ESI (M+H)⁺: 297.0.

107 3-((5-amino-1,3,4-thiadiazol-2-yl)thio)benzo[b]thiophene 1,1-dioxide (3l)

Yield 3I: (16 mg, 7%). ¹H NMR (300 MHz, CDCl₃): 6.8 (1H, s); 7.3 (2H, NH₂, s); 7.6 - 7.8 (m, 4H). Yield 20: (5 mg, 2%). ¹H NMR (300 MHz, CDCl₃): 7.1 (2H, s); 7.6 - 7.8 (m, 4H); 8.8 (1H, SH, s). LCMS - ESI (M+H)⁺: 298.0.

111 3-((5-mercapto-1,3,4-thiadiazol-2-yl)amino)benzo[b]thiophene 1,1-dioxide (3m)

- 112 Yield **3m**: (5 mg, 2%). ¹H NMR (300 MHz, CDCl₃): 7.1 (2H, s); 7.6 7.8 (m, 4H); 8.8 (1H, SH,
- 113 s). LCMS ESI (M+H)⁺: 298.0.

114 3-((1H-benzo[d]imidazol-2-yl)thio)benzo[b]thiophene 1,1-dioxide (3n)

- 115 Yield **3n**: (100 mg, 39%). ¹H NMR (300 MHz, CDCl₃): 6.7 (1H, s); 7.3 7.8 (m, 9H). LCMS –
- **116** ESI (M+H)⁺: 315.0.

117 3-((1-methyl-3a,7a-dihydro-1H-benzo[d]imidazol-2-yl)thio)benzo[b]thiophene 1,1-dioxide

- 118 **(30)**
- 119 Yield **30**: (57 mg, 21%). ¹H NMR (300 MHz, CDCl₃): 3.9 (3H, CH₃. s); 6.4 (1H, s); 7.3 7.8 (m,
- 120 8H). LCMS ESI $(M+H)^+$: 329.0.

PeerJ PrePrints

- 121 3-((5-nitro-3a,7a-dihydro-1H-benzo[d]imidazol-2-yl)thio)benzo[b]thiophene 1,1-dioxide
- 122 (**3**p)
- 123 Yield 3p: (142 mg, 48%). ¹H NMR (300 MHz, CDCl₃): 7.5 (1H, s); 7.7 7.9 (m, 4H); 8.2 (2H,
- 124 d); 8.5 (1H, s). LCMS ESI $(M+H)^+$: 360.0.
- 3-((5-methoxy-3a,7a-dihydro-1H-benzo[d]imidazol-2-yl)thio)benzo[b]thiophene 1,1-dioxide
 (3q)
- 127 Yield 3q: (97 mg, 34%). ¹H NMR (300 MHz, Methanol-d₄): 3.8 (3H, OCH₃. s); 6.6 (1H, s); 7.0 –
 128 7.7 (m, 7H).
- 129 3-((1-methyl-1H-tetrazol-5-yl)thio)benzo[b]thiophene 1,1-dioxide (3r)
- 130 Yield **3r**: (115 mg, 50%). ¹H NMR (300 MHz, CDCl₃): 4.3 (3H, 3CH₃, s); 6.9 (1H, s); 7.5 8.1
- **131** (m, 7H). LCMS ESI $(2M+H)^+$: 561.0.

132 3-(pyridin-2-ylthio)benzo[b]thiophene 1,1-dioxide (3v)

- **133** Yield **3v**: (120 mg, 53%). ¹H NMR (300 MHz, CDCl₃): 6.6 8.5 (m, 9H). LCMS ESI (M+H)⁺:
- **134** 276.0.

135 **3-(pyridin-4-ylthio)benzo[b]thiophene 1,1-dioxide (3w)**

- 136 Yield **3w**: (57 mg, 25%). ¹H NMR (300 MHz, CDCl₃): 6.6 8.5 (m, 9H). LCMS ESI (M+H)⁺:
- **137** 276.0.

138 3-(isoquinolin-3-ylthio)benzo[b]thiophene 1,1-dioxide (3x)

139 Yield 3x: (95 mg, 36%). ¹H NMR (300 MHz, CDCl₃): 7.3 – 8.1 (m, 6H).

140 3-(naphthalen-2-ylthio)benzo[b]thiophene 1,1-dioxide (3y)

141 Yield 3y: (110 mg, 42%). ¹H NMR (300 MHz, CDCl₃): 5.8 (1H, s); 7.6 - 8.1 (m, 11H). LCMS 142 ESI (M+2Na)⁺: 671.0.

143 **Results and Discussion**

BTD analogs were synthesized as outlined in **Scheme 1**. The oxidation of commercially available 3-bromothianaphthalene (1) with hydrogen peroxide afforded 3-bromobenzothiophene-1,1dioxide (2). This in turn was reacted with the corresponding thiols to afford the 3-substituted BTDs. To investigate the biological activity, we conducted a systematic exploration of the aryl/heteroaryl substituents linked via a thioether to the C-3 position of the benzo[b]thiophene-1,1-dioxide compound.

150 We probed the consequences of having oxazoles and oxadiazoles as substituents at the C-3 151 position. Compounds were tested for efficacy against a virulent strain of *M. tuberculosis* in liquid 152 culture under aerobic growth conditions [4]. All compounds had good activity and the minimum 153 inhibitory concentration (MIC) was very similar $(3-8 \mu M)$ (**Table 1**). The change in electronics 154 of the phenyl substituents had no effect on potency of the oxadiazole compounds. The addition of 155 the electron donating groups, methyl (3b), methoxy (3d) or an electron withdrawing Cl-group 156 (3e) to the para position of (3a) resulted in similar MIC values (Table 1). MICs were similar for 157 benzaoxazole 3c and the phenyl linked oxadiazoles (3a, b, d, and e). This confirmed that the 158 series has good anti-tubercular activity. We tested compound activity against eukaryotic cells 159 using the Vero cell line (derived from African green monkey kidney cells). All of the compounds 160 had significant cytotoxicity, with TC₅₀ values $< 0.3 \mu$ M, suggesting that these compounds are 161 even more effective against eukaryotic cells (Table 1). Of the compounds we tested, two had 162 previously been identified as having anti-tubercular activity (3a and 3i)³. In this study 3a 163 appeared to have a selectivity index (SI) of > 33. However, in our assay this compound had a SI 164 of 0.03. The compound **3a** was reported to have anti-tubercular activity with an IC₉₀ of 1.3 μ M 165 $(0.45 \ \mu g/mL)$ and a TC₅₀ of 43 μM (calculated from the published data using the equation TC₅₀= 166 SI x IC₉₀). Ananthan *et al.*, calculated IC₉₀ in their assay, representing the concentration required 167 to inhibit growth by 90%, but in our experience IC9₀ and MIC (which we used) are very similar. 168 In our case it had an MIC of 3.1 μ M and a cytotoxicity of 0.1 μ M. Therefore, the difference in SI 169 is primarily due to the difference in cytotoxicity data.

170 Since we had seen good activity with the compounds, but significant cytotoxicity, we determined 171 whether we could separate the two activities to generate potent, non-toxic compounds. We 172 examined the influence of thiazoles and thiadiazoles on the biological activity and cytotoxicity of 173 these BTD compounds. Anti-tubercular activity was diminished by the replacement of an 174 oxazole with either a thiazole or a thiadiazole; these compounds showed MICs $\geq 20 \mu M$ (Table 175 2). The only exception was compound 3k which showed good activity (9 μ M), where the 176 addition of an electron donating ethoxy group to the benzothiazole compound improved its 177 potency to 5 µM (3i). In contrast, addition of an electron-withdrawing group diminished activity 178 in compound 3j (MIC >20 μ M, Table 2). Cytotoxicity was also reduced by 10-100-fold, and 179 although the selectivity index (SI) was also improved the compounds were still more active 180 against eukaryotic cells with SI of < 0.2 (Table 2). The benzothiazole compound 3i has 181 previously been reported³, but in contrast to our results, it had a SI > 150, whereas our data 182 indicate that the SI = 0.5. The compound **3i** was reported to have a TB IC₉₀ of $< 0.3 \mu M$ (< 0.1183 μ g/mL) and a TC₅₀ of 45 μ M. In our assays it gave a MIC of 20 μ M and a cytotoxicity of 1 μ M. 184 In this case the difference in SI is due to both the difference in activity and cytotoxicity data.

We then investigated the effect of C-3 imidazoles to see if we could improve the SI. Similar to the thiazoles and thiadiazoles, this resulted in diminished activity (MIC > 20 μ M) (3n - 3q) (Table 3). Cytotoxicity was similar to those seen with the thiazole and thiadiazole groups. Methylation of the N-1 of the imidazole (30) had no effect on activity (3n, 3p and 3q). The tetrazole compound (3r) showed the best activity of all the compounds synthesized (MIC = 2.6 μ M), but also had significant cytotoxicity (Table 3).

Finally, we explored the influence of having six membered heterocycles in the C-3 position. We synthesized compounds with pyrimidyl (3t), pyridyl (3v, 3w), quinolinyl (3u), or isoquinolinyl (3x) groups and a non-heterocyclic compound with a naphthyl group (3y). All these analogs were inactive suggesting that the BTD series requires a five membered heterocyclic substituent at the C-3 position linked via a thioether for its activity against *M. tuberculosis* (Table 4).

196 Conclusions

197 We conducted a systematic exploration of the aryl/heteroaryl thioether substituents at the C-3 198 position of the benzo[b]thiophene-1,1-dioxide compound series for its inhibitory activity against 199 *M. tuberculosis*. The series exhibited encouraging activity with some MIC values $<10 \mu$ M. The 200 tetrazole, oxazole and the oxadiazoles were the most potent compounds tested, whereas 201 compounds bearing six-membered aromatic substituents at the C-3 position were inactive. 202 However, the BTD series was also active against eukaryotic cells showing significant toxicity 203 against the Vero cell line; in fact cytotoxicity was more pronounced than the anti-mycobacterial 204 activity. Our data are in contrast to that previously reported in which cytotoxicity was not 205 observed in selected members of the series [3]. Differences in cytotoxicity could be due to the 206 exact assay method and the cell line used; in this case we used the same Vero cell line. 207 However, the assays conditions were different; we used passaged cells which were actively

replicating as opposed to cells recovered directly from frozen. Since the majority of cytotoxicity is manifested during cell division, this may account for our increased sensitivity. In any case, we found that the series as a whole was cytotoxic. We were unable to reduce cytotoxicity in this series, even after significant modifications of the third position substituent. On this basis we concluded that the series lacks further potential for drug development. However, the target of the series may still be of interest, since one might find alternative scaffolds with specificity. Thus, in future, we are interested in finding the target of these compounds.

215 Acknowledgements

216 We thank Alfredo Blakeley, David Roberts and Yulia Ovechkina for technical assistance.

217 References

- **218** 1. WHO 2011 Report: global tuberculosis control. Geneva: World Health Organization.
- 219 2. Ginsberg A. 2010. Drugs in development for tuberculosis. *Drugs* 70:2201-2204.
- 220 3. Ananthan S, Faaleolea ER, Goldman RC, Hobrath JV, Kwong CD, Laughon BE, Maddry AJ,
- 221 Mehta A, Rasmussen L, Reynolds RC, Secrist III JA, Shindo N, Showe DN, Sosa MI, Suling WJ,
- 222 White EL. 2009. High-throughput screening for inhibitors of Mycobacterium tuberculosis
- **223** H37Rv. *Tuberculosis* 89: 334-353.
- 4. Ollinger J, Bailey M, Moraski GC, Casey A, Florio S, Alling T, Miller MJ, Parish T. 2013. A
- 225 dual read-out assay to evaluate the potency of compounds active against Mycobacterium
- tuberculosis. PLOS One 8: e60531.
- 227 5. Ioerger TR, Feng Y, Ganesula K, Chen X, Dobos KM, Fortune S, Jacobs WR, Mizrahi V,
- 228 Parish T, Rubin E, Sassetti C, Sacchettini JC. 2010. Variation among genome sequences of

- 229 H37Rv strains of Mycobacterium tuberculosis from multiple laboratories. J. Bacteriology 192:
- **230** 3645-3653.

PeerJ PrePrints | http://dx.doi.org/10.7287/peerj.preprints.502v1 | CC-BY 4.0 Open Access | rec: 17 Sep 2014, publ: 17 Sep 2014

Table 1 (on next page)

Activity of oxazole and oxadiazole analogs of the BTD series against *M. tuberculosis* and Vero cell line.

a = MIC is the minimum concentration required to inhibit growth completely of *M. tuberculosis* in liquid culture⁴. MICs of active compounds are the average of two independent experiments \pm standard deviation. b = TC₅₀ is concentration required to inhibit growth of Vero cells by 50%. TC₅₀ is the average of two runs \pm standard deviation. c = SI is the selectivity index. Selectivity index is calculated as MIC/ TC₅₀. For comparison, MIC of rifampicin is 0.003 µM and isoniazid is 0.2 µM [4] .

Compound	R-gro up	a MIC (μM)	Ъ ТС ₅₀ (μМ)	SI °
3a	N·N C	3.1 ± 0.07	0.1 ± 0	0.03
3b	N-N O Me	8.2 ± 0.6	0.2 ± 0	0.02
3c		5.7 ± 2.9	0.2 ± 0.07	0.04
3d	N-N OMe	7.2 ± 0.3	0.3 ± 0.3	0.04
3e	CI	3.9 ± 1.7	0.3 ±0.2	0.08

Table 1. Activity of oxazole and oxadiazole analogs of the BTD series against M. tuberculosis and Vero cell line.

a = MIC is the minimum concentration required to inhibit growth completely of *M. tuberculosis* in liquid culture⁴. MICs of active compounds are the average of two independent experiments \pm standard deviation.

 $b = TC_{50}$ is concentration required to inhibit growth of Vero cells by 50%. TC₅₀ is the average of two runs \pm standard deviation.

c = SI is the selectivity index. Selectivity index is calculated as MIC/ TC₅₀.

For comparison, MIC of rifampicin is 0.003 μ M and isoniazid is 0.2 μ M [4].

Table 2(on next page)

Activity of thiazole and thiadiazole analogs of the BTD series against *M. tuberculosis* and Vero cell line.

a = MIC is the minimum concentration required to inhibit growth completely of *M. tuberculosis* in liquid culture⁴. MICs of active compounds are the average of two independent experiments \pm standard deviation. b = TC₅₀ is concentration required to inhibit growth of Vero cells by 50%. TC₅₀ is the average of two runs \pm standard deviation. c = SI is the selectivity index. Selectivity index is calculated as MIC/ TC₅₀. For comparison, MIC of rifampicin is 0.003 µM and isoniazid is 0.2 µM [4] .

Compound	R-group	MIC (µM) ^a	TC ₅₀ (μΜ) ^b	SI °
3g	N S	20	1	0.05
3h	S N	20	3±1.2	0.2
3i	N S	20	1	0.05
3j	N CI	>20	1 ± 0.4	NC
3k	N Do	9.0 ± 4.7	1 ± 0.1	0.1
31	N-N Ks Me	>20	1 ± 0.4	NC
3m	N-N ───────────────────────────────────	>20	3 ± 1	NC
3n	N-N HN√s∕SH	20	3±0.7	0.2

Table 2. Activity of thiazole and thiadiazole analogs of the BTD series against M. tuberculosis and Vero cell line.

a = MIC is the minimum concentration required to inhibit growth completely of *M. tuberculosis* in liquid culture⁴. MICs of active compounds are the average of two independent experiments \pm standard deviation.

 $b = TC_{50}$ is concentration required to inhibit growth of Vero cells by 50%. TC₅₀ is the average of two runs \pm standard deviation.

c = SI is the selectivity index. Selectivity index is calculated as MIC/ TC_{50.}

NC = not calculated.

Table 3(on next page)

Activity of imidazole and tetrazole analogs of the BTD series against *M. tuberculosis* and Vero cell line.

a = MIC is the minimum concentration required to inhibit growth completely of *M. tuberculosis* in liquid culture⁴. MICs of active compounds are the average of two independent experiments \pm standard deviation. b = TC₅₀ is concentration required to inhibit growth of Vero cells by 50%. TC₅₀ is the average of two runs \pm standard deviation. c = SI is the selectivity index. Selectivity index is calculated as MIC/ TC₅₀. NC = not calculated. ND = not determined.

Compound	R-group	MIC (μ M) ^a	TC ₅₀ (μM) ^b	SI °
3n		>20	1	NC
30	N N	>20	5	NC
3р		>20	ND	NC
3q		>20	0.3 ± 0.07	NC
3r	H N∙N Me Me	2.6	0.1 ± 0	0.004

Table 3. Activity of imidazole and tetrazole analogs of the BTD series against M. tuberculosis and Vero cell line.

a = MIC is the minimum concentration required to inhibit growth completely of *M. tuberculosis* in liquid culture⁴. MICs of active compounds are the average of two independent experiments \pm standard deviation.

 $b = TC_{50}$ is concentration required to inhibit growth of Vero cells by 50%. TC₅₀ is the average of two runs \pm standard deviation.

c = SI is the selectivity index. Selectivity index is calculated as MIC/ TC₅₀.

NC = not calculated.

ND = not determined.

Table 4(on next page)

Activity of six membered heterocyclic analogs of the BTD series against *M. tuberculosis* and Vero cell line.

a = MIC is the minimum concentration required to inhibit growth completely of *M. tuberculosis* in liquid culture⁴. MICs of active compounds are the average of two independent experiments \pm standard deviation. b = TC₅₀ is concentration required to inhibit growth of Vero cells by 50%. TC₅₀ is the average of two runs \pm standard deviation. c = SI is the selectivity index. Selectivity index is calculated as MIC/ TC₅₀. NC = not calculated. ND = not determined.

Compound	R-group	a MIC (µM)	ΤC ₅₀ (μΜ) ^b	SI °
3s	O S=0	>20	10 ± 1	NC
3t	N T Me	>20	ND	NC
3u		>20	ND	NC
3v		>20	ND	NC
3w	- N	>20	ND	NC
3x		>20	ND	NC
Зу		>20	ND	NC

Table 4. Activity of six membered heterocyclic analogs of the BTD series against M.tuberculosis and Vero cell line.

a = MIC is the minimum concentration required to inhibit growth completely of *M. tuberculosis* in liquid culture⁴. MICs of active compounds are the average of two independent experiments \pm standard deviation.

 $b = TC_{50}$ is concentration required to inhibit growth of Vero cells by 50%. TC₅₀ is the average of two runs \pm standard deviation.

c = SI is the selectivity index. Selectivity index is calculated as MIC/ TC₅₀.

NC = not calculated.

ND = not determined.

Table 5(on next page)

Synthesis of 3-substituted benzo[b]thiophene-1,1-dioxides.



Scheme 1. Synthesis of 3-substituted benzo[b]thiophene-1,1-dioxides.