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# **Placental vascular pathology and increased thrombin generation as mechanisms of disease in obstetrical syndromes**

Obstetrical complications including preeclampsia, fetal growth restriction, preterm labor, preterm prelabor rupture of membranes and fetal demise are all the clinical endpoint of several underlying mechanisms (i.e. infection inflammation, thrombosis, endocrine disorder, immunologic rejection, genetic, and environmental), therefore, they may be regarded as syndromes. Placental vascular pathology and increased thrombin generation were reported in all of these obstetrical syndromes. Moreover, elevated concentrations of thrombin-anti-thrombin III complexes and changes in the coagulation as well as anticoagulation factors can be detected in the maternal circulation prior to the clinical development of the disease in some of these syndromes. In this review, we will assess the changes in the hemostatic system during normal and complicated pregnancy in maternal blood, maternal-fetal interface and amniotic fluid, and describe the contribution of thrombosis and vascular pathology to the development of the great obstetrical syndromes.

1 **Placental Vascular Pathology and Increased Thrombin Generation as Mechanisms of**  
2 **Disease In Obstetrical Syndromes**

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20 **1. Introduction**

21 Obstetrical complications including preeclampsia, fetal growth restriction, preterm labor, preterm  
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23 mechanisms (i.e. infection inflammation, thrombosis, endocrine disorder, immunologic rejection,  
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26 maternal blood, maternal-fetal interface and amniotic fluid, and describe the contribution of  
27 thrombosis and vascular pathology to the development of the great obstetrical syndromes.

28 **2. What are the great obstetrical syndromes?**

The major obstetrical complications including preeclampsia, intrauterine growth restriction (IUGR), preterm labor (PTL), preterm prelabor rupture of membranes (PROM), fetal demise, and recurrent abortions are all syndromes, also defined as "great obstetrical syndromes". As reported in The Oxford Medical Dictionary a syndrome is 'a combination of symptoms and/or signs that form a distinct clinical picture indicative of a particular disorder'. Hence, they represent the clinical manifestation of many possible underlying mechanisms of disease<sup>1</sup>. Key features of these syndromes are<sup>2</sup>: multiple etiologies; long preclinical stage; frequent fetal involvement; clinical manifestations which are often adaptive in nature; and predisposition to a particular syndrome is influenced by gene–environment interaction and/or complex gene-gene interactions involving maternal and/or fetal genotypes. These mechanisms of disease were identified and reported in all the obstetrical complications listed above. This review is focused on the role of thrombosis and vascular pathology of the placenta in these syndromes.

### **3. What are the changes in the coagulation system during normal pregnancy?**

In term of the coagulation and hemostatic systems there are several major compartments: the maternal circulation, the fetal maternal interface (the placenta, and membranes), amniotic fluid and the fetus that each has a specific behavior during gestation. The changes in the coagulation system during gestation are considered to be adaptive mechanisms and are aimed to: 1) the prevention of bleeding at the time of trophoblast implantation and the delivery of the fetus; 2) allow the laminar flow and the intervillous space; and 3) seal amniotic fluid leak and reduce obstetrical bleeding<sup>3-7</sup>. Of interest, the fetus is somewhat less involved and its coagulation system develops during gestation, and this subject is beyond the scope of this review. Indeed, normal pregnancy has been associated with excessive maternal thrombin generation<sup>3, 8</sup> and a tendency for platelets to aggregate in response to agonists<sup>9, 10</sup>. Pregnancy is accompanied by 2 to 3-fold increase in fibrinogen concentrations and 20% to 1000% increase in factors VII, VIII, IX, X, and XII, all of which peak at term<sup>11</sup>. The concentrations of vWF increase up to 400% by term<sup>11</sup>. By contrast, those of pro-thrombin and factor V remain unchanged while the concentrations of factors XIII and XI decline modestly<sup>12</sup>. Indeed there is evidence of chronic low-level thrombin and fibrin generation throughout normal pregnancy as indicated by enhanced concentrations of pro-thrombin fragment 1.2, thrombin-antithrombin (TAT) III complexes, and soluble fibrin polymers<sup>13</sup>. Free protein S concentration declines significantly (up to 55%) during pregnancy due to increased circulating complement 4B-binding protein its molecular carrier. Protein S nadir at delivery and this reduction is exacerbated by cesarean delivery and infection<sup>11, 12</sup>. As a consequence, pregnancy is associated with an increase in resistance to activated protein

C<sup>12, 13</sup>. The concentrations of PAI-1 increase by 3 to 4-folds during pregnancy while plasma PAI-2 values, which are negligible before pregnancy reach concentrations of 160 mg/L at delivery<sup>11</sup>. Thus, pregnancy is associated with increased clotting potential, as well as decreased anticoagulant properties, and fibrinolysis<sup>14</sup>. Therefore, it can be defined as a prothrombotic state. One of the most important mediators of the hypercoagulable state of normal pregnancy is tissue factor. Indeed, there is a substantial increase in tissue factor (TF) concentrations in the decidua and myometrium<sup>15-18</sup>, as well as preventing placental abruption. The placenta is a source of TF, since trophoblast cells constitutively express it, behaving as activated endothelium, and leading to a condition of procoagulant state that, if not controlled by anticoagulant mechanisms, predisposes to thrombotic complications<sup>15</sup>. The principal anticoagulant mechanism inhibiting TF activation pathway is tissue factor pathway inhibitor (TFPI), which mRNA is highly expressed in the macrophages in the villi in term placenta<sup>19</sup>. Similarly, high TF concentrations have been detected in the fetal membranes (mainly the amnion) and amniotic fluid<sup>7, 20-23</sup>. TFPI has been found in amniotic fluid as well<sup>20</sup>, but it is not clear if the presence of TF and its natural inhibitor is related to coagulation per se or is somehow connected with embryonic development<sup>24</sup>. In contrast to the changes detected in the amniotic fluid and the decidua, the median maternal plasma immunoreactive TF concentration of normal pregnant women do not differ significantly from that of non-pregnant patients<sup>3, 25</sup>. However, labor at term increases significantly the maternal plasma immunoreactive TF concentration in comparison to the non-pregnant state<sup>20</sup>. In addition to the changes in TF, normal pregnancy is associated with increased thrombin generation<sup>3, 8</sup>, as determined by the elevation of maternal concentrations of fibrinopeptide A, prothrombin fragments (PF) 1 and 2, and thrombin–antithrombin (TAT) III complexes<sup>7, 26-28</sup>. The concentration of these complexes further increases during and after normal parturition<sup>27, 29</sup>, and subsequently decreases during the puerperium<sup>27, 29</sup>.

#### 4. What are the changes in the hemostatic system associated with the great obstetrical syndrome?

The great obstetrical syndromes are associated with changes in the hemostatic and vascular systems in the compartments mentioned above: 1) the maternal circulation; 2) the feto-maternal interface of placenta and membranes; 3) and the amniotic fluid.

##### 4.1 Changes in the hemostatic system of women with obstetrical syndromes.

The involvement of the hemostatic system in the pathophysiology of these obstetrical syndromes is becoming more and more apparent. Indeed, increased thrombin generation is reported in the

maternal circulation of women with preeclampsia<sup>30-34</sup>, IUGR<sup>30-32, 35, 36</sup>, fetal demise<sup>37</sup>, PTL<sup>8, 37, 38</sup> and preterm PROM<sup>8, 37, 39</sup>.

There are several possible explanations for the increased thrombin generation in these patients: 1) increased activation of coagulation cascade in the maternal circulation due to pathological processes including bleeding or inflammation; and 2) depletion of anticoagulation proteins that subsequently leads to increased thrombin generation (Table 1).

#### **4.1.1 Increased activation of the coagulation cascade and thrombin generation in the maternal circulation in patients with pregnancy complications**

All the obstetrical syndromes including preeclampsia<sup>30-34, 40, 41</sup>, FGR<sup>31, 32, 35, 36</sup>, fetal demise<sup>37</sup>, PTL<sup>8, 38</sup> and preterm PROM<sup>8, 37, 39</sup> are associated with a higher maternal thrombin generation than a normal pregnancy. These may be of clinical implication since in women with preterm labor, elevated maternal plasma TAT III complexes concentration was associated with a higher chance to deliver within <7 days from admission<sup>37</sup> (Fig. 1). To further understand how does thrombin affect the duration of pregnancy and the clinical phenotype of patients with the obstetrical syndromes we need to consider what are the mechanisms leading to thrombin generation and how it affects the feto-maternal unit.

Increased thrombin generation can result from the following underlying mechanisms: 1) decidual hemorrhage that leads to a retro-placental clot formation<sup>42</sup>; 2) intra-amniotic infection/inflammation which can induce decidual bleeding and sub-clinical abruption<sup>43</sup>, as well as increased intra-amniotic TAT complexes<sup>37</sup>; and 3) an increased maternal systemic inflammatory response<sup>44</sup> that may activate the extrinsic pathway of coagulation due to the expression and release of TF by activated monocytes<sup>45</sup>.

Thrombin affects many systems including also the following: 1) stimulation of decidual cell secretion of matrix metalloproteinase (MMP) (i.e. MMP-1 and MMP-3) that can degrade the extracellular matrix of the chorioamniotic membranes<sup>46, 47</sup> (as in preterm PROM); 2) myometrial activation and uterine contractions generation that may lead to preterm labor with or without rupture of membranes and a subsequent preterm delivery<sup>38, 48, 49</sup>; and 3) thrombin has an inhibitory effect on the production of TFPI by endothelial cells<sup>50</sup>, and the increased thrombin generation observed in patients with PTL may be associated with a concomitant reduction in TFPI production by the maternal vascular endothelium (the depletion of anticoagulant proteins will be discussed in the following section of this review).

There is evidence to support that the extrinsic pathway of coagulation is activated in many of these pregnancy complications and it is the source of the increased thrombin generation<sup>51</sup>. Indeed, increased immunoreactive TF concentrations were reported in women with preeclampsia and

those with preterm PROM<sup>52</sup>. Moreover, the contribution of preeclampsia to elevated maternal immunoreactive TF persisted also among patients with fetal demise, while those with fetal death who were normotensive did not have higher median TF concentration than normal pregnant women. Moreover, the median TF concentration of patients with preeclampsia was also higher than in patients with fetal demise without hypertension. These findings are consistent with previous studies<sup>3, 53</sup>, suggesting that elevated TF immunoreactivity and activity may be associated with the pathophysiologic process leading to preeclampsia, rather than being a consequence of the fetal death.

In some of the obstetrical syndromes there was elevated tissue factor activity in the maternal circulation without a concomitant increase in the plasma concentration of this factor. This was the case among patients with an SGA neonate and those with preterm labor<sup>54 31</sup> (Table 1). This suggests that the increased TF activity among patients with PTL as well as those with an SGA neonate, contributes to a higher generation of factor Xa that, along with the physiologic increase in the maternal plasma concentrations of factor VII and factor X during gestation<sup>11, 55-57</sup>, may be the underlying mechanism leading to the increased thrombin generation reported these syndromes.

The differences between PTL and preterm PROM in term of maternal plasma TF concentration and activity may derive from the specific component of the common pathway of parturition, which is activated in each obstetrical syndrome<sup>58</sup>. While preterm PROM is associated with the activation of the decidua and the membranes, myometrial activation is the major component of preterm labor with intact membranes<sup>58</sup>. This is relevant because the decidua and the membranes have a high TF concentration<sup>17, 18, 59</sup>.

In summary, the evidence brought herein suggests that increased thrombin generation in patients with the great obstetrical syndromes may reflect the activation of the coagulation cascade mainly through the extrinsic arm. This activation can be attributed to various underlying mechanisms.

#### 4.1.2 Depleted or insufficient anticoagulant proteins concentration

In the normal state there is a delicate balance between the proteins activating/participating the coagulation cascade and their inhibitors. Increased thrombin generation may result, as we presented above, from activation of the coagulation cascade due to higher concentrations or activities of the proteins included in the coagulation cascade. However, thrombin generation can also result from insufficient concentration or activity of anticoagulation proteins.

Tissue factor pathway inhibitor (TFPI), a glycoprotein comprising of three Kunitz domain<sup>60</sup> that are specific inhibitors of trypsin-like proteinases<sup>61</sup>, is the main inhibitor of the extrinsic pathway of coagulation. TFPI inhibits thrombin generation through the inactivation of activated factor X

163 and the factor VIIa/TF complex<sup>60, 62</sup>. The mean maternal plasma concentrations of total TFPI  
 164 increases during the first half of pregnancy, remains relatively constant in the second half<sup>63</sup> and  
 165 decreases during labor<sup>20</sup>. There are two types of TFPI: 1) TFPI-1 is the more prevalent form in  
 166 the non-pregnant state in the maternal circulation and can also be found in the fetal blood,  
 167 platelets, endothelial cells and other organs<sup>19, 64</sup>; and 2) TFPI-2- the major form of TFPI in the  
 168 placenta<sup>65-68</sup>, also known as Placental Protein 5 (PP5)<sup>69, 70</sup>. During pregnancy, the maternal plasma  
 169 concentration of TFPI-2 increases gradually, reaches a plateau at 36 weeks and subsides after  
 170 delivery<sup>71-74</sup>.  
 171 The overall balance between the concentration and activity of the coagulation factors and the  
 172 anti-coagulation proteins is one of the determining factors of thrombin generation. In the normal  
 173 state, the immunoreactive concentrations of TFPI in the plasma are 500 to 1000 times higher than  
 174 that of TF<sup>75</sup>, suggesting that an excess of anti-coagulant proteins closely controls the coagulation  
 175 cascade activity. The median maternal plasma TFPI concentration increases during  
 176 preeclampsia<sup>53, 76</sup>, which is associated with an exaggerated maternal systemic inflammatory  
 177 response. However, the increase in the median maternal TF plasma concentration is such that the  
 178 overall balance between TF and its inhibitor is affected leading to increased thrombin generation  
 179 in this syndrome. In contrast to preeclampsia, maternal plasma TFPI concentration decreases in  
 180 patients with PTL<sup>52</sup> and preterm PROM<sup>77</sup> regardless to the presence of intra-amniotic  
 181 infection/inflammation, as well as in women with fetal demise<sup>54</sup>, and does not change in mothers  
 182 with SGA fetuses<sup>53</sup>. Overall these findings suggest that the increased thrombin generation  
 183 observed among these patients may derive not only from an increased activation of the  
 184 hemostatic system, but also from insufficient anti-coagulation, as reflected by the lower TFPI  
 185 concentrations (Fig. 2).  
 186 A possible explanation of the lower maternal plasma concentration observed in some of the  
 187 obstetrical syndromes may be that during these syndromes there is a reduction in the placental  
 188 production of TFPI<sup>65, 66, 69, 76</sup> (mainly TFPI-2), contributing to the low maternal plasma  
 189 concentrations detected in patients with PTL, in addition to the thrombin inhibitory effect to TFPI  
 190 expression on endothelial cells, as above mentioned. Indeed, patients with vascular complications  
 191 of pregnancy (preeclampsia, eclampsia, placental abruption, fetal growth restriction, and fetal  
 192 demise) have a lower placental concentration of total TFPI, and TFPI mRNA expression than in  
 193 women with normal pregnancies<sup>78, 79</sup>.  
 194 Other proteins implicated in the inhibitory control of the coagulation cascade are protein S,  
 195 protein C and protein Z. Protein S is a cofactor to protein C in the inactivation of factors  
 196 Va and VIIIa. This protein exists in two forms: a free form and a complex form bound



197 to complement protein C4b-binding protein (C4BP). Only the free form is active<sup>80</sup>. Protein S also  
 198 acts as a TFPI cofactor, in the presence of weak pro-coagulant stimuli, by enhancing the  
 199 interaction of TFPI with factor Xa while using Ca<sup>2+</sup> and phospholipids in the process<sup>81</sup> without  
 200 increasing inhibition of factor VIIa-TF by TFPI<sup>82</sup>. During pregnancy there is a physiologic  
 201 change in the relationship between the bound and the free forms of protein S in the maternal  
 202 plasma. The increase in C4BP during gestation reduces free protein S concentration in up to 55%  
 203 of its value out of pregnant state, reaching its nadir at delivery. Of interest, cesarean delivery and  
 204 infection exacerbate the reduction in free protein S concentrations<sup>11, 83</sup>. Moreover, a functional  
 205 protein S deficiency can explain a poor response to activated protein C<sup>84</sup>.  
 206 The association between the alteration of concentration and function of protein S and protein C in  
 207 the great obstetrical syndromes is not completely clear. The evidence regarding the association of  
 208 protein S and protein C deficiency and preeclampsia is controversial<sup>85, 86</sup>.  
 209 While some reported an association between protein S deficiency and an increased risk for this  
 210 syndrome (especially for early onset preeclampsia)<sup>85</sup> others could not demonstrate this effect<sup>86</sup>.  
 211 There is some evidence regarding the relation of protein S deficiency and increased risk of  
 212 stillbirth<sup>87</sup> and mid-trimester IUGR<sup>88</sup>. An increased risk of stillbirth has been reported in patients  
 213 with protein S deficiency while the risk was not significantly increased in cases of protein C  
 214 deficiency<sup>87</sup>, and Kupferminc et al<sup>88</sup> found that protein S, but not protein C deficiency, was  
 215 significantly associated with severe mid-trimester IUGR.  
 216 Protein Z, in complex with protein Z-dependent protease inhibitor (ZPI) (Fig. 3)<sup>89-91</sup>, acts as a  
 217 physiologic inhibitor of activation of prothrombin by factor Xa. Protein Z is a vitamin  
 218 K-dependent plasma glycoprotein<sup>92</sup> that is an essential cofactor for ZPI activity. In the absence of  
 219 protein Z, the activity of ZPI is reduced by more than 1000-fold<sup>91</sup>. Normal pregnancy is  
 220 characterized by an increased plasma concentration of protein Z<sup>93</sup>, probably as a compensation  
 221 for the increase of factor X concentration. Women with preterm labor without intra-amniotic  
 222 infection or inflammation and those with vaginal bleeding who delivered preterm had a lower  
 223 median maternal plasma protein Z concentration than women with a normal pregnancy and those  
 224 with vaginal bleeding who delivered at term<sup>94</sup>. The changes of protein Z concentrations in other  
 225 pregnancy complications are controversial. Some demonstrated that the median plasma  
 226 concentration of protein Z in patients with preeclampsia, IUGR, and late fetal death were not  
 227 significantly different than that of patients with a normal pregnancy<sup>95</sup>. Others reported lower  
 228 median maternal plasma protein Z concentrations in women with preeclampsia or pyelonephritis  
 229 and higher proportion of protein Z deficiency (defined as protein Z plasma concentration below  
 230 the 5<sup>th</sup> percentile) in patients with preeclampsia or fetal demise than in those with a normal

231 pregnancy<sup>96</sup>. Moreover, increased maternal plasma anti-protein Z antibodies concentrations were  
 232 associated with SGA neonates, fetal demise and preeclampsia.  
 233 The information presented above suggest that it is not only the concentration of one coagulation  
 234 factor or anticoagulation protein, but rather the overall balance between the coagulation factors  
 235 and their inhibitors that increases thrombin generation in the great obstetrical syndromes. Indeed,  
 236 although preterm labor was not associated with a significant change in the median maternal  
 237 plasma TF concentration, the TFPI/TF ratio of these patients was lower than that of normal  
 238 pregnant women, mainly due to decreased TFPI concentrations.  
 239 This observation was also reported in patients with preterm PROM<sup>77</sup>, and those with  
 240 preeclampsia<sup>53</sup>. The lower TFPI/TF ratio in patients with preeclampsia occurs despite the increase  
 241 in the median maternal plasma TFPI concentration observed in these patients. This suggests that  
 242 the balance between TF and its natural inhibitor may better reflect the overall activity of the TF  
 243 pathway of coagulation, than the individual concentrations of TF or TFPI.  
 244 Collectively, these observations suggest that our attention should be focused not only on the  
 245 coagulation protein but also on their inhibitors since an imbalance between them may contribute  
 246 to increased thrombin generation leading to the onset of the great obstetrical syndromes.

## 4.2 Changes in the feto-maternal interface

Normal placental development and the establishment of an adequate feto-maternal circulation are key points for a successful pregnancy. The networks of the placental vascular tree either on the maternal or fetal side are dynamic structures that can be substantially altered in cases of abnormal placentation and trophoblast invasion. The human trophoblast has properties of endothelial cells and can regulate the degree of activation of the coagulation cascade in the intervillous space<sup>97, 98</sup>. The vilous trophoblasts express heparin sulfate, protein C and protein Z on their surface that serve as anticoagulant that sustain laminar blood flow through the intervillous space. On the other hand, unlike the endothelium of other organs, the trophoblast constantly presents the active placental isoform of TF on its surface<sup>98-101</sup>. This isoform has a higher affinity for factor VIIa<sup>102</sup>, which may lead to increased activation of the coagulation cascade. One of the leading pathological processes observed in all these syndromes is thrombosis and vascular abnormality of the placenta at the maternal-fetal interface. The incidence of these pathological processes varies among the different syndromes being more prevalent in preeclampsia, IUGR, and fetal demise than in PTL and preterm PROM<sup>30, 31, 37, 38</sup>.

### 4.2.1 Placental pathology in the Great Obstetrical Syndromes

There is a range of placental vascular and thrombotic lesions that are being observed in placentas of patients with pregnancy complications. Thrombotic events of placental vessels can cause an impairment of placental perfusion, leading to FGR, preeclampsia and fetal death as well as in some extents to PTL and preterm PROM<sup>103, 104</sup>. The frequency of the specific vascular placental lesions varies among these obstetrical syndromes<sup>105</sup>. Placental vascular lesions are divided into maternal or fetal vascular origin (figure 1-2)<sup>106, 107</sup>. Lesions of the maternal vascular compartment include placental marginal and retro-placental hemorrhages, lesions related to maternal under perfusion (acute atherosclerosis and mural hypertrophy, increased syncytial knots, villous agglutination, increased intervillous fibrin deposition, villous infarcts)<sup>106</sup>. Placental fetal vascular obstructive lesions are the result of stasis, hypercoagulability and vascular damage within the fetal circulation of the placenta. Placental fetal vascular abnormalities include: cord-related abnormalities (as torsion of cord, over-coiling, strictures and tight knots<sup>108</sup>) and vascular lesions consistent with fetal thrombo-occlusive disease (thrombosis of the chorionic plate and stem villous vessels, fibrotic, hypo-vascular and avascular villi<sup>106</sup>. In addition, villitis of unknown etiology or chronic villitis, defined as lymphohistiocytic

278 inflammation localized to the stroma of terminal villi but often extending to the small vessels of  
279 upstream villi is also associated with obliterative fetal vasculopathy<sup>106</sup> (Fig. 4-5).

280 **Preeclampsia:** The classical example for an association between obstetrical syndrome and  
281 vascular placental lesions is preeclampsia. Women who develop preeclampsia have an increased  
282 rate of abnormalities of the maternal side of the placental circulation and maternal  
283 underperfusion<sup>109, 110</sup>. The frequency of these lesions is inversely related to the gestational age in  
284 which the hypertensive disorder was diagnosed. The earlier the development of  
285 hypertension/preeclampsia the more severe are the vascular lesions<sup>111, 112</sup>. Moreover, Kovo et al<sup>113</sup>  
286 reported that the presence of fetal growth restriction in women with preeclampsia increases also  
287 the frequency of fetal vascular lesions. Indeed, patients with early-onset preeclampsia  
288 complicated by FGR had a higher rate of fetal-vascular supply lesions consistent with fetal  
289 thrombo-occlusive disease than women with early-onset disease without FGR<sup>113</sup>.  
290 An assessment of the pathologic changes in placental hemostatic system has been performed in  
291 patients with preeclampsia. Teng et al<sup>114</sup> studied TF and TFPI placental levels in pregnant patients  
292 with preeclampsia, compared to normal pregnancies. They found increased TF placental  
293 expression and a reduced expression of TFPI-1 and TFPI-2, with a significant correlation  
294 between the levels of TF and TFPI-2 between maternal plasma and placenta.

295 **Fetal growth restriction:** Placentas from pregnancies complicated by FGR are smaller and have  
296 significantly increased maternal and fetal vascular lesions compared to placentas from normal  
297 pregnancies with appropriate for gestational age neonates (AGA)<sup>115, 116</sup>. Maternal vascular  
298 lesions were detected in about 50% of placentas from pregnancies complicated with FGR at term,  
299 compared to only 20% in normal pregnancies, while fetal vascular lesions were observed in 11%  
300 of FGR pregnancies compared to only 4% in placentas from normal pregnancies<sup>113</sup>.  
301 Placentas from normotensive pregnancies complicated by early-onset FGR (<34 weeks of  
302 gestation) had a higher rate of low placental weight (<10th percentile) and maternal  
303 underperfusion, as compared to placentas of women who delivered AGA neonates  $\leq$ 34 weeks of  
304 gestation<sup>115</sup>. Of interest, placentas from the late onset FGR group (after 34 weeks of gestation), in  
305 addition to the high maternal vascular abnormalities, show also more fetal vascular abnormalities,  
306 compared with AGA controls who delivered >34 weeks<sup>117</sup>.

307 **Fetal demise:** Placental disease has been recognized as an important contributor to unexplained  
308 fetal demise. Fetal vascular abnormalities<sup>105</sup> are extensively involved in early and late fetal death  
309 rather than maternal vascular lesions. In fetal death occurring prior to 34 weeks, an earlier and

extended insult in the placental development occurs. On the other hand, late fetal demise is an unpredicted event that is mostly characterized by non-thrombotic cord related lesions and less placental vascular compromise<sup>107</sup>.

**Preterm labor and preterm PROM:** Placental studies in PTL demonstrated a combination of inflammatory and vascular lesions. PTL is generally attributed to an inflammatory response involving the bacterial induction of cytokine and prostanoid production<sup>118</sup>. Finding of histological chorioamnionitis in PTL<sup>119</sup> has established infection and inflammation as a causative factor of preterm birth, moreover, noninfectious trigger may also contribute to the development of preterm labor and in some instances may be evident by placental sterile inflammatory response<sup>120</sup>. In addition, isolated placental vascular lesions, mostly of maternal supply, were reported in 20% of cases of PTL and an additional 20% had combined inflammatory and vascular lesions. Moreover, there are consistent reports describing increased rate of failure of transformation of the spiral arteries in women with preterm labor without intrauterine infection/inflammation and in those with preterm PROM than in women with normal pregnancies<sup>121</sup>. Such findings imply that an inadequate uteroplacental blood flow due to abnormal placentation plays an important role in pathogenesis of preterm parturition<sup>121, 122</sup>.

Collectively, placental vascular lesions were reported in all the great obstetrical syndromes. The severity of these lesions is associated with the timing of diagnosis of the disease. The more severe the vascular injury, the more likely these complications will become clinically evident prior to 34 weeks of gestation. Of interest, vascular lesions often come along with evidence of acute inflammation or lesions associated with chronic inflammatory processes, suggesting that sometimes more than one mechanism is involved in development of a specific obstetrical syndrome.

#### 4.3. Hemostatic changes in the amniotic fluid of women with obstetrical syndromes

During normal pregnancy, there is an increase in the amniotic fluid TF concentration<sup>7, 20-23</sup>. In order to demonstrate the association of hemostatic changes and the development of obstetrical complications, Erez et al<sup>54</sup> studied the changes in the intra-amniotic concentration of TAT III complexes, as well as TF concentration and activity, in cases of fetal demise and in normal pregnancies. Patients with a fetal demise had higher median amniotic fluid–TF concentration and activity than those with normal pregnancies. Moreover, among patients with a FD there was a significant correlation (Fig. 6) between the amniotic fluid–TF concentrations and activity ( $r = 0.88$ ,  $P$

<0.0001). The median amniotic fluid– TAT III complexes concentration did not differ significantly between the groups (normal pregnancy: median: 66.3 mg/l, range 11.4–2265.4 vs. FD: median: 59.3 mg/l, range: 13.6–15,425.3;  $P=0.7$ ). In their study, the median amniotic fluid–TF concentration in normal pregnant women was 10 fold higher than in maternal plasma. The changes in amniotic fluid thrombin generation were reported also in women with preterm parturition. Indeed, intra-amniotic infection and/or inflammation is associated with an increased amniotic fluid TAT III complexes (Fig. 7). This is important since it represents an increased thrombin generation in the amniotic cavity during infection and/or inflammation that may contribute to uterine contractility and the development of preterm birth<sup>123</sup>. Of interest, elevated intra-amniotic TAT III concentrations were associated with a shorter amniocentesis to delivery interval and an earlier gestational age at delivery only in patients with preterm labor without intra-amniotic infection or inflammation<sup>123</sup>. This observation suggests that in a subset of patients with preterm labor, activation of the coagulation system can generate preterm parturition and delivery; while in those with intra-amniotic infection and/or inflammation the activation of the coagulation and thrombin generation is a byproduct of the inflammatory process leading to preterm birth. This represents evidence of the activation and propagation of coagulation cascade, being thrombin generation the witness of the former mechanisms and the inhibitor of the initiation step<sup>54</sup>.

## 5. Conclusion

The evidence presented herein suggests a role for increased thrombin generation and vascular placental lesions in the pathogenesis of the great obstetrical syndromes. This process can be the result of the contribution of procoagulant and vascular abnormalities as well as inflammatory and infectious mechanisms, representing the starting point for pregnancy complications based on vascular disease. As presented, these changes affect the mother, the placenta, membranes and amniotic fluid. Moreover, preliminary evidence suggest that some of the changes in the hemostatic system in the mother and in the amniotic fluid predate the clinical presentation of the disease. Suggesting that better understanding of the vascular and coagulation changes associated with the great obstetrical syndromes may assist us in earlier detection and the development or introduction of therapeutic modalities for these syndromes.





- 376 1. *Concise Medical Dictionary*: Oxford University Press, 2010.
- 377 2. ROMERO R. Prenatal medicine: the child is the father of the man. 1996. J Matern Fetal Neonatal Med 2009;22:636-9.
- 378 3. BELLART J, et al. Endothelial cell markers and fibrinopeptide A to D-dimer ratio as a measure of coagulation and fibrinolysis balance in normal pregnancy. Gynecol Obstet Invest 1998;46:17-21.
- 379 4. WALKER MC, et al. Changes in activated protein C resistance during normal pregnancy. Am J Obstet Gynecol 1997;177:162-9.
- 380 5. SØRENSEN JD, SECHER NJ, JESPERSEN J. Perturbed (procoagulant) endothelium and deviations within the fibrinolytic system during the third trimester of normal pregnancy. A possible link to placental function. Acta Obstet Gynecol Scand 1995;74:257-61.
- 381 6. YUEN PM, YIN JA, LAO TT. Fibrinopeptide A levels in maternal and newborn plasma. Eur J Obstet Gynecol Reprod Biol 1989;30:239-44.
- 382 7. DE BOER K, et al. Enhanced thrombin generation in normal and hypertensive pregnancy. Am J Obstet Gynecol 1989;160:95-100.
- 383 8. CHAIWORAPONGSA T, et al. Activation of coagulation system in preterm labor and preterm premature rupture of membranes. J Matern Fetal Neonatal Med 2002;11:368-73.
- 384 9. YONEYAMA Y, et al. Plasma adenosine levels increase in women with normal pregnancies. Am J Obstet Gynecol 2000;182:1200-3.
- 385 10. SHEU JR, et al. Mechanisms involved in the antiplatelet activity of midazolam in human platelets. Anesthesiology 2002;96:651-8.
- 386 11. BREMME KA. Haemostatic changes in pregnancy. Best Pract Res Clin Haematol 2003;16:153-68.
- 387 12. EICHINGER, et al. Prospective evaluation of hemostatic system activation and thrombin potential in healthy pregnant women with and without factor V Leiden. Thromb Haemost. 1999;82:1232-6.
- 388 13. KU DH, et al. Circulating levels of inflammatory cytokines (IL-1 beta and TNF-alpha), resistance to activated protein C, thrombin and fibrin generation in uncomplicated pregnancies. Thromb Haemost 2003;90:1074-9.
- 389 14. LOCKWOOD CJ. Pregnancy-associated changes in the hemostatic system. Clin Obstet Gynecol 2006;49:836-43.
- 390 15. ERLICH J, et al. Tissue factor is required for uterine hemostasis and maintenance of the placental labyrinth during gestation. Proc Natl Acad Sci U S A 1999;96:8138-43.
- 391 16. KUCZYŃSKI J, et al. Tissue factor (TF) and tissue factor pathway inhibitor (TFPI) in the placenta and myometrium. Eur J Obstet Gynecol Reprod Biol 2002;105:15-9.
- 392 17. LOCKWOOD CJ, KRIKUN G, SCHATZ F. Decidual cell-expressed tissue factor maintains hemostasis in human endometrium. Ann N Y Acad Sci 2001;943:77-88.
- 393 18. LOCKWOOD CJ, KRIKUN G, SCHATZ F. The decidua regulates hemostasis in human endometrium. Semin Reprod Endocrinol 1999;17:45-51.
- 394 19. EDSTROM CS, CALHOUN DA, CHRISTENSEN RD. Expression of tissue factor pathway inhibitor in human fetal and placental tissues. Early Hum Dev 2000;59:77-84.
- 395 20. USZYŃSKI M, et al. Tissue factor (TF) and tissue factor pathway inhibitor (TFPI) in amniotic fluid and blood plasma: implications for the mechanism of amniotic fluid embolism. Eur J Obstet Gynecol Reprod Biol 2001;95:163-6.
- 396 21. LOCKWOOD CJ, et al. Amniotic fluid contains tissue factor, a potent initiator of coagulation. Am J Obstet Gynecol 1991;165:1335-41.



- 422 22. OMSJØ IH, et al. Thromboplastin activity in amniotic fluid. *Gynecol Obstet Invest*  
423 1985;19:1-5.
- 424 23. CRETER D. Amnioplastin: new reagent for coagulation tests. *Lancet* 1977;2:251.
- 425 24. CARMELIET P, et al. Role of tissue factor in embryonic blood vessel development. *Nature*  
426 1996;383:73-5.
- 427 25. HOLMES VA, WALLACE JM. Haemostasis in normal pregnancy: a balancing act? *Biochem*  
428 *Soc Trans* 2005;33:428-32.
- 429 26. REBER G, AMIRAL J, DE MOERLOOSE P. Modified antithrombin III levels during normal  
430 pregnancy and relationship with prothrombin fragment F1 + 2 and thrombin-antithrombin  
431 complexes. *Thromb Res* 1998;91:45-7.
- 432 27. USZYŃSKI M. Generation of thrombin in blood plasma of non-pregnant and pregnant  
433 women studied through concentration of thrombin-antithrombin III complexes. *Eur J*  
434 *Obstet Gynecol Reprod Biol* 1997;75:127-31.
- 435 28. REINTHALLER A, MURSCH-EDLMAYR G, TATRA G. Thrombin-antithrombin III complex  
436 levels in normal pregnancy with hypertensive disorders and after delivery. *Br J Obstet*  
437 *Gynaecol* 1990;97:506-10.
- 438 29. ANDERSSON T, et al. Thrombin-inhibitor complexes in the blood during and after delivery.  
439 *Thromb Res* 1996;82:109-17.
- 440 30. SCHJETLEIN R, et al. Hemostatic variables as independent predictors for fetal growth  
441 retardation in preeclampsia. *Acta Obstet Gynecol Scand* 1999;78:191-7.
- 442 31. CHAIWORAPONGSA T, et al. Evidence of in vivo generation of thrombin in patients with  
443 small-for-gestational-age fetuses and pre-eclampsia. *J Matern Fetal Neonatal Med*  
444 2002;11:362-7.
- 445 32. HAYASHI M, et al. Blood macrophage colony-stimulating factor and thrombin-  
446 antithrombin III complex concentrations in pregnancy and preeclampsia. *Am J Med Sci*  
447 1998;315:251-7.
- 448 33. KOBAYASHI T, TERAOKA T. Preeclampsia as chronic disseminated intravascular coagulation.  
449 Study of two parameters: thrombin-antithrombin III complex and D-dimers. *Gynecol*  
450 *Obstet Invest* 1987;24:170-8.
- 451 34. HAYASHI M, et al. Characterization of five marker levels of the hemostatic system and  
452 endothelial status in normotensive pregnancy and pre-eclampsia. *Eur J Haematol*  
453 2002;69:297-302.
- 454 35. HAYASHI M, OHKURA T. Elevated levels of serum macrophage colony-stimulating factor  
455 in normotensive pregnancies complicated by intrauterine fetal growth restriction. *Exp*  
456 *Hematol* 2002;30:388-93.
- 457 36. BALLARD HS, MARCUS AJ. Primary and secondary platelet aggregation in uraemia. *Scand*  
458 *J Haematol* 1972;9:198-203.
- 459 37. EREZ O, et al. Changes in amniotic fluid concentration of thrombin-antithrombin III  
460 complexes in patients with preterm labor: evidence of an increased thrombin generation. *J*  
461 *Matern Fetal Neonatal Med* 2009;22:971-82.
- 462 38. ELOVITZ MA, BARON J, PHILLIPPE M. The role of thrombin in preterm parturition. *Am J*  
463 *Obstet Gynecol* 2001;185:1059-63.
- 464 39. ROSEN T, et al. Plasma levels of thrombin-antithrombin complexes predict preterm  
465 premature rupture of the fetal membranes. *J Matern Fetal Med* 2001;10:297-300.
- 466 40. KOBAYASHI T, et al. Coagulation/fibrinolysis disorder in patients with severe  
467 preeclampsia. *Semin Thromb Hemost* 1999;25:451-4.
- 468 41. KOBAYASHI T, et al. Coagulation index to distinguish severe preeclampsia from normal  
469 pregnancy. *Semin Thromb Hemost* 2002;28:495-500.

- 470 42. LOCKWOOD CJ, et al. Mechanisms of abruption-induced premature rupture of the fetal  
471 membranes: thrombin-enhanced interleukin-8 expression in term decidua. *Am J Pathol*  
472 2005;167:1443-9.
- 473 43. GÓMEZ R, et al. Idiopathic vaginal bleeding during pregnancy as the only clinical  
474 manifestation of intrauterine infection. *J Matern Fetal Neonatal Med* 2005;18:31-7.
- 475 44. GERVASI MT, et al. Maternal intravascular inflammation in preterm premature rupture of  
476 membranes. *J Matern Fetal Neonatal Med* 2002;11:171-5.
- 477 45. ØSTERUD B, BJØRKLID E. Sources of tissue factor. *Semin Thromb Hemost* 2006;32:11-23.
- 478 46. ROSEN T, et al. Thrombin-enhanced matrix metalloproteinase-1 expression: a mechanism  
479 linking placental abruption with premature rupture of the membranes. *J Matern Fetal*  
480 *Neonatal Med* 2002;11:11-7.
- 481 47. MACKENZIE AP, et al. Mechanisms of abruption-induced premature rupture of the fetal  
482 membranes: Thrombin enhanced decidual matrix metalloproteinase-3 (stromelysin-1)  
483 expression. *Am J Obstet Gynecol* 2004;191:1996-2001.
- 484 48. ELOVITZ MA, et al. The mechanisms underlying the stimulatory effects of thrombin on  
485 myometrial smooth muscle. *Am J Obstet Gynecol* 2000;183:674-81.
- 486 49. ELOVITZ MA, et al. Effects of thrombin on myometrial contractions in vitro and in vivo.  
487 *Am J Obstet Gynecol* 2000;183:799-804.
- 488 50. BILSEL AS, et al. Long-term effect of 17beta-estradiol and thrombin on tissue factor  
489 pathway inhibitor release from HUVEC. *Thromb Res* 2000;99:173-8.
- 490 51. VANWIJK MJ, et al. Enhanced coagulation activation in preeclampsia: the role of APC  
491 resistance, microparticles and other plasma constituents. *Thromb Haemost* 2002;88:415-  
492 20.
- 493 52. EREZ O, et al. High tissue factor activity and low tissue factor pathway inhibitor  
494 concentrations in patients with preterm labor. *J Matern Fetal Neonatal Med* 2010;23:23-  
495 33.
- 496 53. EREZ O, et al. Tissue factor and its natural inhibitor in pre-eclampsia and SGA. *J Matern*  
497 *Fetal Neonatal Med* 2008;21:855-69.
- 498 54. EREZ O, et al. Evidence of maternal platelet activation, excessive thrombin generation,  
499 and high amniotic fluid tissue factor immunoreactivity and functional activity in patients  
500 with fetal death. *J Matern Fetal Neonatal Med* 2009;22:672-87.
- 501 55. BELLER FK, EBERT C. The coagulation and fibrinolytic enzyme system in pregnancy and  
502 in the puerperium. *Eur J Obstet Gynecol Reprod Biol* 1982;13:177-97.
- 503 56. STIRLING Y, et al. Haemostasis in normal pregnancy. *Thromb Haemost* 1984;52:176-82.
- 504 57. BRENNER B. Haemostatic changes in pregnancy. *Thromb Res* 2004;114:409-14.
- 505 58. ROMERO R, et al. Mechanisms of preterm labor and preterm premature rupture of the  
506 membranes. In: Kurjak A, Chervenak F, eds. *Textbook of Perinatal Medicine 2nd Edition*,  
507 2006.
- 508 59. LOCKWOOD CJ, et al. The role of decidualization in regulating endometrial hemostasis  
509 during the menstrual cycle, gestation, and in pathological states. *Semin Thromb Hemost*  
510 2007;33:111-7.
- 511 60. BROZE GJ, et al. The lipoprotein-associated coagulation inhibitor that inhibits the factor  
512 VII-tissue factor complex also inhibits factor Xa: insight into its possible mechanism of  
513 action. *Blood* 1988;71:335-43.
- 514 61. LASKOWSKI M, KATO I. Protein inhibitors of proteinases. *Annu Rev Biochem*  
515 1980;49:593-626.
- 516 62. BROZE GJ, GIRARD TJ, NOVOTNY WF. Regulation of coagulation by a multivalent  
517 Kunitz-type inhibitor. *Biochemistry* 1990;29:7539-46.

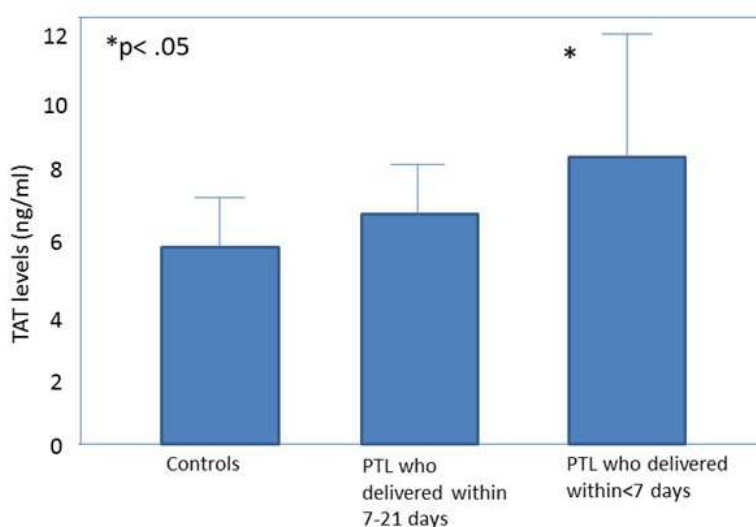
- 518 63. SARIG G, et al. Modulation of systemic hemostatic parameters by enoxaparin during  
519 gestation in women with thrombophilia and pregnancy loss. *Thromb Haemost*  
520 2005;94:980-5.
- 521 64. TAY SP, CHEONG SK, BOO NY. Circulating tissue factor, tissue factor pathway inhibitor  
522 and D-dimer in umbilical cord blood of normal term neonates and adult plasma. *Blood*  
523 *Coagul Fibrinolysis* 2003;14:125-9.
- 524 65. HUBÉ F, et al. Demonstration of a tissue factor pathway inhibitor 2 messenger RNA  
525 synthesis by pure villous cytotrophoblast cells isolated from term human placentas. *Biol*  
526 *Reprod* 2003;68:1888-94.
- 527 66. IINO M, FOSTER DC, KISIEL W. Quantification and characterization of human endothelial  
528 cell-derived tissue factor pathway inhibitor-2. *Arterioscler Thromb Vasc Biol* 1998;18:40-  
529 6.
- 530 67. SPRECHER CA, et al. Molecular cloning, expression, and partial characterization of a  
531 second human tissue-factor-pathway inhibitor. *Proc Natl Acad Sci U S A* 1994;91:3353-7.
- 532 68. UDAGAWA K, et al. Specific expression of PP5/TFPI2 mRNA by syncytiotrophoblasts in  
533 human placenta as revealed by in situ hybridization. *Placenta* 1998;19:217-23.
- 534 69. KAMEI S, et al. Genomic structure and promoter activity of the human tissue factor  
535 pathway inhibitor-2 gene. *Biochim Biophys Acta* 2001;1517:430-5.
- 536 70. KISIEL W, SPRECHER CA, FOSTER DC. Evidence that a second human tissue factor  
537 pathway inhibitor (TFPI-2) and human placental protein 5 are equivalent. *Blood*  
538 1994;84:4384-5.
- 539 71. BÜTZOW R, et al. Monoclonal antibodies reacting with placental protein 5: use in  
540 radioimmunoassay, Western blot analysis, and immunohistochemistry. *J Lab Clin Med*  
541 1988;111:249-56.
- 542 72. CHAND HS, FOSTER DC, KISIEL W. Structure, function and biology of tissue factor  
543 pathway inhibitor-2. *Thromb Haemost* 2005;94:1122-30.
- 544 73. SEPPÄLÄ M, WAHLSTRÖM T, BOHN H. Circulating levels and tissue localization of  
545 placental protein five (PP5) in pregnancy and trophoblastic disease: absence of PP5  
546 expression in the malignant trophoblast. *Int J Cancer* 1979;24:6-10.
- 547 74. OBIKWE BC, CHARD T. Placental protein 5: circulating levels in twin pregnancy and  
548 some observations on the analysis of biochemical data from multiple pregnancy. *Eur J*  
549 *Obstet Gynecol Reprod Biol* 1981;12:135-41.
- 550 75. SHIMURA M, et al. Plasma tissue factor and tissue factor pathway inhibitor levels in  
551 patients with disseminated intravascular coagulation. *Am J Hematol* 1997;55:169-74.
- 552 76. ABDEL GADER AM, et al. Total and free tissue factor pathway inhibitor in pregnancy  
553 hypertension. *Int J Gynaecol Obstet* 2006;95:248-53.
- 554 77. EREZ O, et al. A link between a hemostatic disorder and preterm PROM: a role for tissue  
555 factor and tissue factor pathway inhibitor. *J Matern Fetal Neonatal Med* 2008;21:732-44.
- 556 78. XIONG Y, et al. Changes of plasma and placental tissue factor pathway inhibitor-2 in  
557 women with preeclampsia and normal pregnancy. *Thromb Res* 2010;125:e317-22.
- 558 79. AHARON A, et al. Placental TFPI is decreased in gestational vascular complications and  
559 can be restored by maternal enoxaparin treatment. *J Thromb Haemost* 2005;3:2355-7.
- 560 80. CASTOLDI E, HACKENG TM. Regulation of coagulation by protein S. *Curr Opin Hematol*  
561 2008;15:529-36.
- 562 81. HACKENG TM, et al. Protein S stimulates inhibition of the tissue factor pathway by tissue  
563 factor pathway inhibitor. *Proc Natl Acad Sci U S A* 2006;103:3106-11.
- 564 82. NDONWI M, BROZE G. Protein S enhances the tissue factor pathway inhibitor inhibition of  
565 factor Xa but not its inhibition of factor VIIa-tissue factor. *J Thromb Haemost*  
566 2008;6:1044-6.

- 567 83. FAUGHT W, et al. Changes in protein C and protein S levels in normal pregnancy. *Am J*  
568 *Obstet Gynecol* 1995;172:147-50.
- 569 84. DAHLBÄCK B, CARLSSON M, SVENSSON PJ. Familial thrombophilia due to a previously  
570 unrecognized mechanism characterized by poor anticoagulant response to activated  
571 protein C: prediction of a cofactor to activated protein C. *Proc Natl Acad Sci U S A*  
572 1993;90:1004-8.
- 573 85. RODGER MA, et al. Inherited thrombophilia and pregnancy complications revisited.  
574 *Obstet Gynecol* 2008;112:320-4.
- 575 86. YALINKAYA A, et al. The relationship between thrombophilic mutations and preeclampsia:  
576 a prospective case-control study. *Ann Saudi Med* 2006;26:105-9.
- 577 87. PRESTON FE, et al. Increased fetal loss in women with heritable thrombophilia. *Lancet*  
578 1996;348:913-6.
- 579 88. KUPFERMINC MJ, et al. Mid-trimester severe intrauterine growth restriction is associated  
580 with a high prevalence of thrombophilia. *BJOG* 2002;109:1373-6.
- 581 89. HAN X, FIEHLER R, BROZE GJ. Isolation of a protein Z-dependent plasma protease  
582 inhibitor. *Proc Natl Acad Sci U S A* 1998;95:9250-5.
- 583 90. HAN X, et al. The protein Z-dependent protease inhibitor is a serpin. *Biochemistry*  
584 1999;38:11073-8.
- 585 91. HAN X, FIEHLER R, BROZE GJ. Characterization of the protein Z-dependent protease  
586 inhibitor. *Blood* 2000;96:3049-55.
- 587 92. YIN ZF, et al. Prothrombotic phenotype of protein Z deficiency. *Proc Natl Acad Sci U S A*  
588 2000;97:6734-8.
- 589 93. TAYLOR FB, et al. Active site inhibited factor VIIa (DEGR VIIa) attenuates the coagulant  
590 and interleukin-6 and -8, but not tumor necrosis factor, responses of the baboon to LD100  
591 *Escherichia coli*. *Blood* 1998;91:1609-15.
- 592 94. KUSANOVIC JP, et al. Plasma protein Z concentrations in pregnant women with idiopathic  
593 intrauterine bleeding and in women with spontaneous preterm labor. *J Matern Fetal*  
594 *Neonatal Med* 2007;20:453-63.
- 595 95. BRETELLE F, et al. Protein Z in patients with pregnancy complications. *Am J Obstet*  
596 *Gynecol* 2005;193:1698-702.
- 597 96. NIEN JK, et al. Pyelonephritis during pregnancy: a cause for an acquired deficiency of  
598 protein Z. *J Matern Fetal Neonatal Med* 2008;21:629-37.
- 599 97. SOOD R, et al. Fetomaternal cross talk in the placental vascular bed: control of  
600 coagulation by trophoblast cells. *Blood* 2006;107:3173-80.
- 601 98. SOOD R, et al. Maternal Par4 and platelets contribute to defective placenta formation in  
602 mouse embryos lacking thrombomodulin. *Blood* 2008;112:585-91.
- 603 99. LANIR N, AHARON A, BRENNER B. Procoagulant and anticoagulant mechanisms in human  
604 placenta. *Semin Thromb Hemost* 2003;29:175-84.
- 605 100. ISERMANN B, et al. The thrombomodulin-protein C system is essential for the  
606 maintenance of pregnancy. *Nat Med* 2003;9:331-7.
- 607 101. AHARON A, et al. Tissue factor and tissue factor pathway inhibitor levels in trophoblast  
608 cells: implications for placental hemostasis. *Thromb Haemost* 2004;92:776-86.
- 609 102. BUTENAS S, et al. Tissue factor in thrombosis and hemorrhage. *Surgery* 2007;142:S2-14.
- 610 103. MIDDELDORP S. Thrombophilia and pregnancy complications: cause or association? *J*  
611 *Thromb Haemost* 2007;5 Suppl 1:276-82.
- 612 104. MARTINELLI I, et al. Risk of pregnancy-related venous thrombosis in carriers of severe  
613 inherited thrombophilia. *Thromb Haemost* 2001;86:800-3.
- 614 105. KOVO M, SCHREIBER L, BAR J. Placental vascular pathology as a mechanism of disease  
615 in pregnancy complications. *Thromb Res* 2013;131 Suppl 1:S18-21.



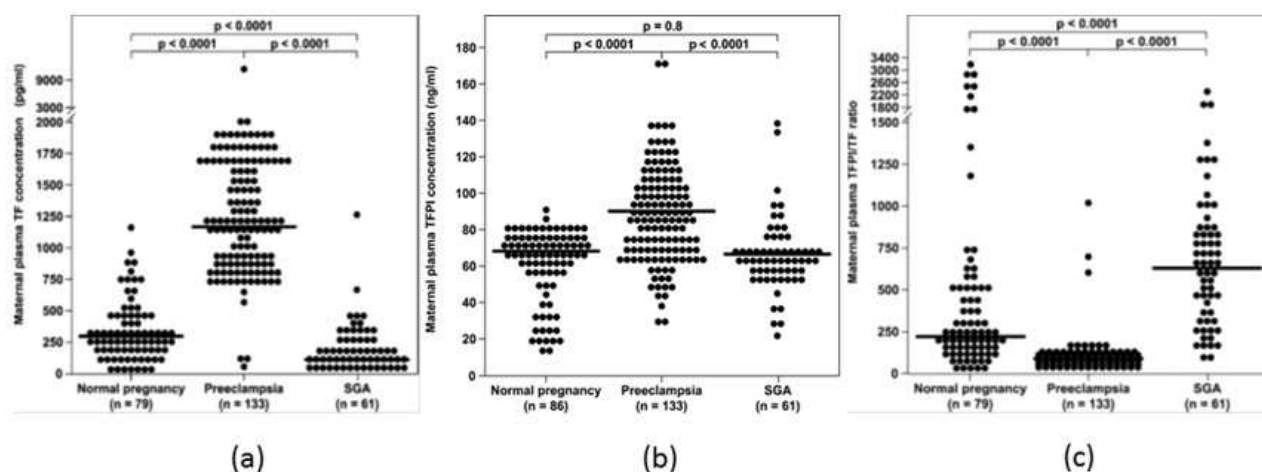
- 616 106. REDLINE RW, et al. Placental diagnostic criteria and clinical correlation--a workshop  
617 report. *Placenta* 2005;26 Suppl A:S114-7.
- 618 107. BAR J, et al. The placental vascular component in early and late intrauterine fetal death.  
619 *Thromb Res* 2012;130:901-5.
- 620 108. CROMI A, et al. Sonographic umbilical cord morphometry and coiling patterns in twin-  
621 twin transfusion syndrome. *Prenat Diagn* 2005;25:851-5.
- 622 109. SALAFIA CM, et al. Clinical correlations of patterns of placental pathology in preterm pre-  
623 eclampsia. *Placenta* 1998;19:67-72.
- 624 110. ROBERTS DJ, POST MD. The placenta in pre-eclampsia and intrauterine growth restriction.  
625 *J Clin Pathol* 2008;61:1254-60.
- 626 111. MAYHEW TM, et al. Stereological investigation of placental morphology in pregnancies  
627 complicated by pre-eclampsia with and without intrauterine growth restriction. *Placenta*  
628 2003;24:219-26.
- 629 112. OGGE G, et al. Placental lesions associated with maternal underperfusion are more  
630 frequent in early-onset than in late-onset preeclampsia. *J Perinat Med* 2011;39:641-52.
- 631 113. KOVO M, et al. Placental vascular lesion differences in pregnancy-induced hypertension  
632 and normotensive fetal growth restriction. *Am J Obstet Gynecol* 2010;202:561.e1-5.
- 633 114. TENG Y, et al. The relationship between plasma and placental tissue factor, and tissue  
634 factor pathway inhibitors in severe pre-eclampsia patients. *Thromb Res* 2010;126:e41-5.
- 635 115. REDLINE RW. Placental pathology: a systematic approach with clinical correlations.  
636 *Placenta* 2008;29 Suppl A:S86-91.
- 637 116. SALAFIA CM, et al. Intrauterine growth restriction in infants of less than thirty-two weeks'  
638 gestation: associated placental pathologic features. *Am J Obstet Gynecol* 1995;173:1049-  
639 57.
- 640 117. KOVO M, et al. The placental component in early-onset and late-onset preeclampsia in  
641 relation to fetal growth restriction. *Prenat Diagn* 2012;32:632-7.
- 642 118. ROMERO R, et al. The preterm parturition syndrome. *BJOG* 2006;113 Suppl 3:17-42.
- 643 119. ARIAS F, et al. Maternal placental vasculopathy and infection: two distinct subgroups  
644 among patients with preterm labor and preterm ruptured membranes. *Am J Obstet*  
645 *Gynecol* 1993;168:585-91.
- 646 120. NATH CA, et al. Histologic evidence of inflammation and risk of placental abruption. *Am*  
647 *J Obstet Gynecol* 2007;197:319.e1-6.
- 648 121. KIM YM, et al. Failure of physiologic transformation of the spiral arteries in the placental  
649 bed in preterm premature rupture of membranes. *Am J Obstet Gynecol* 2002;187:1137-42.
- 650 122. SALAFIA CM, et al. Placental pathologic findings in preterm birth. *Am J Obstet Gynecol*  
651 1991;165:934-8.
- 652 123. STEPHENSON CD, et al. Thrombin-dependent regulation of matrix metalloproteinase  
653 (MMP)-9 levels in human fetal membranes. *J Matern Fetal Neonatal Med* 2005;18:17-22.

654 Figure 1. Thrombin–antithrombin III (TAT) levels in control patients, patients with preterm labor  
 655 who delivered within 3 weeks, and patients with preterm labor who delivered after 3 weeks. Open  
 656 diamonds, Mean levels; black error bars, SD. \*P <.05, Student-Newman-Keuls method (from  
 657 Elovitz MA, Baron J, Phillippe M. The role of thrombin in preterm parturition. Am J Obstet  
 658 Gynecol 2001 Nov;185(5):1059-1063. With permission).



659 Figure 2. (a) Comparison of median maternal plasma TF concentration between patients with  
 660 normal pregnancy (n=79), pre-eclampsia (n=133), and women who delivered an SGAneonate  
 661 (n=61). (b) Comparison of median maternal plasma TFPI concentration between patients with  
 662 normal pregnancy (n=86), pre-eclampsia (n=133), and women who delivered an SGA neonate

663 (n=61). (c) Comparison of maternal plasma TFPI/TF ratio between women with normal  
 664 pregnancy (n=79), pre-eclampsia (n=133), and women who delivered an SGA neonate (n=61).  
 665 (From [Erez O, Romero R, Hoppensteadt D, Than NG, Fareed J, Mazaki-Tovi S, Espinoza](#)  
 666 [J, Chaiworapongsa T, Kim SS, Yoon BH, Hassan SS, Gotsch F, Friel L, Vaisbuch E, Kusanovic](#)  
 667 [JP](#). Tissue factor and its natural inhibitor in pre-eclampsia and SGA. [J Matern Fetal Neonatal](#)  
 668 [Med](#). 2008 Dec;21(12):855-69. With permission).



669 Figure 3. Factor X activation and protein Z/protein Z-dependent protease inhibitor (ZPI)  
 670 inhibition of activated factor X. (a) Then formation of the complex of tissue factor (TF) and  
 671 factor VIIa (FVIIa) at the site of injury and activation of extrinsic coagulation cascade. (b)  
 672 Activation of circulating factor X by the TFpFVIIa complex in the presence of exposed  
 673 phospholipids and Ca<sup>2+</sup>p. (c) Inhibition of factor Xa (FXa) by the protein Z/ZPI complex by  
 674 binding to its active site. Modified from Broze JG, Lancet 2001;357:900–901.

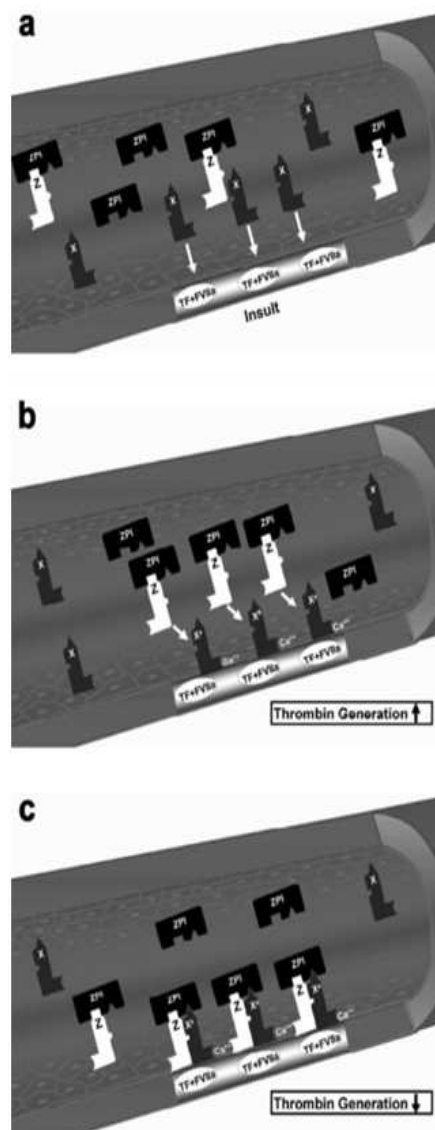
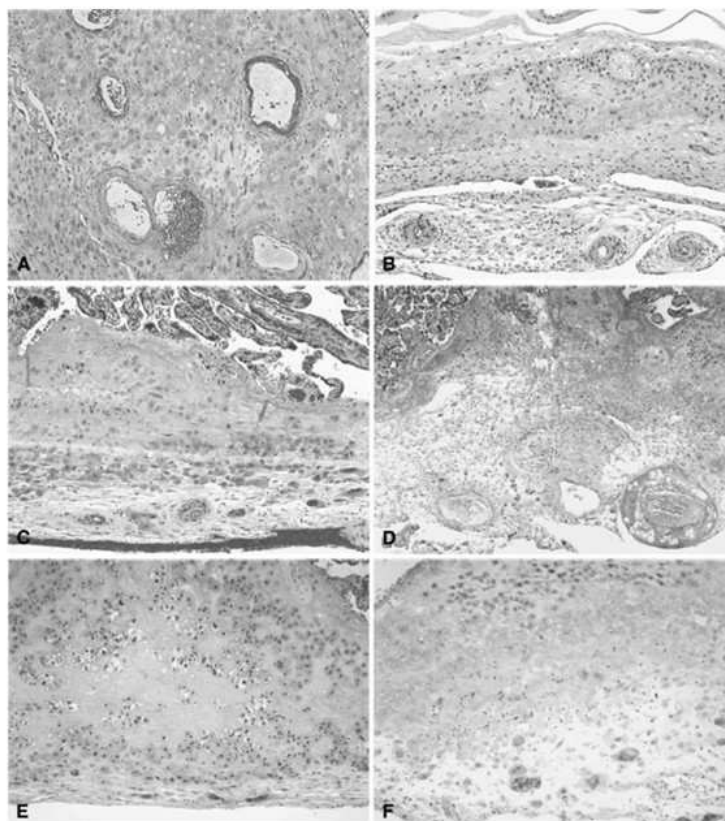


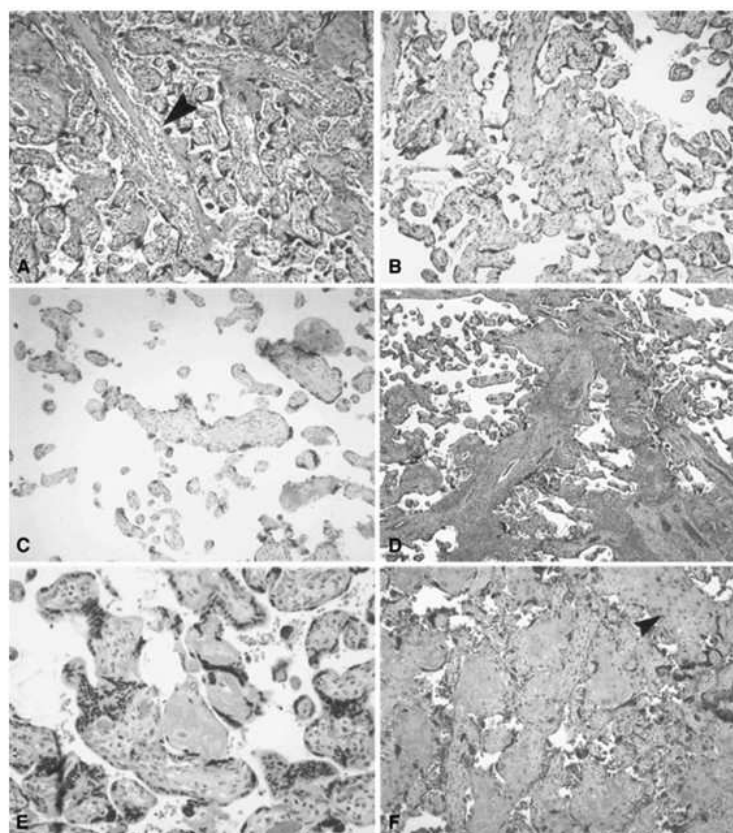
Figure 4. Histologic features of maternal vessel and implantation site reaction patterns: a. Acute atherosclerosis of maternal arterioles in the placental membranes: a cluster of decidual arterioles shows varying stages of fibrinoid necrosis. The vessel at the upper right shows full histologic expression with dark homogenous fibrinoid replacement of the vessel wall accompanied by occasional foamy macrophages ([original magnification is indicated for all panels] X 20). b. Mural hypertrophy of decidual arterioles in the placental membranes: a cluster of arterioles shows medial hypertrophy with the vessel wall occupying greater than one third of total vessel diameter (X 10). c. Muscularized basal plate arteries with accompanying implantation site abnormalities: maternal spiral arteries in the basal plate lack normal trophoblast remodeling and retain their pre-



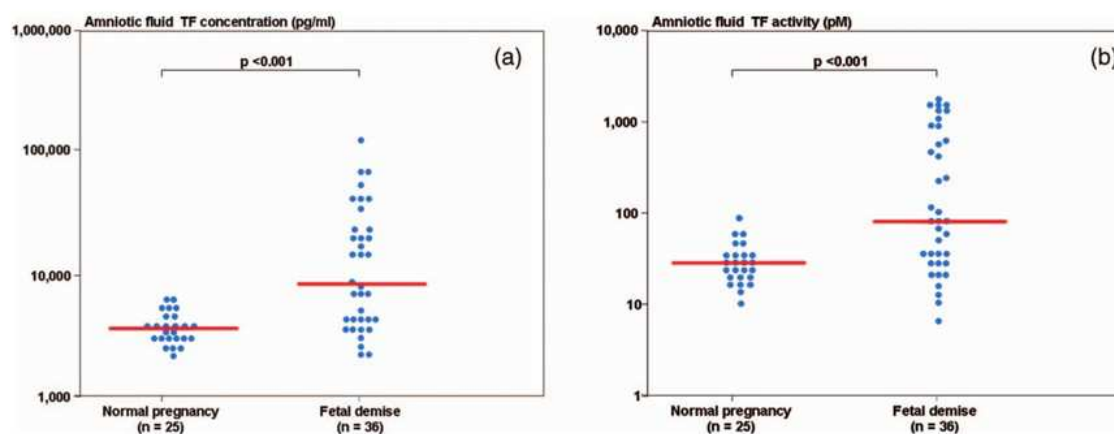
684 pregnancy muscular media. Clusters of immature intermediate trophoblast and increased  
 685 placental giant cells are seen above and below the muscular arteries, respectively (X 10). d. Acute  
 686 atherosclerosis of muscularized basal plate arteries with accompanying implantation site abnormalities:  
 687 three cross sections of a basal plate artery are seen. The two on the left show persistence of the  
 688 muscular media while the one on the right has undergone fibrinoid necrosis of the media with  
 689 foamy macrophages (acute atherosclerosis). Clusters of immature intermediate trophoblast are also  
 690 seen overlying the arteries (X 4). e. Immature intermediate trophoblast: clusters of abnormally  
 691 small intermediate trophoblast with focal vacuolation are surrounded by an excessive amount of  
 692 basal plate fibrin. Increased placental site giant cells are also seen at the lower margin (X 10). f.  
 693 Increased placental site giant cells: numerous multinucleate placental site giant cells, not usually  
 694 seen in the delivered placenta, are scattered in loose decidual tissue which is devoid of normal  
 695 intermediate trophoblast and fibrinoid (X 10).



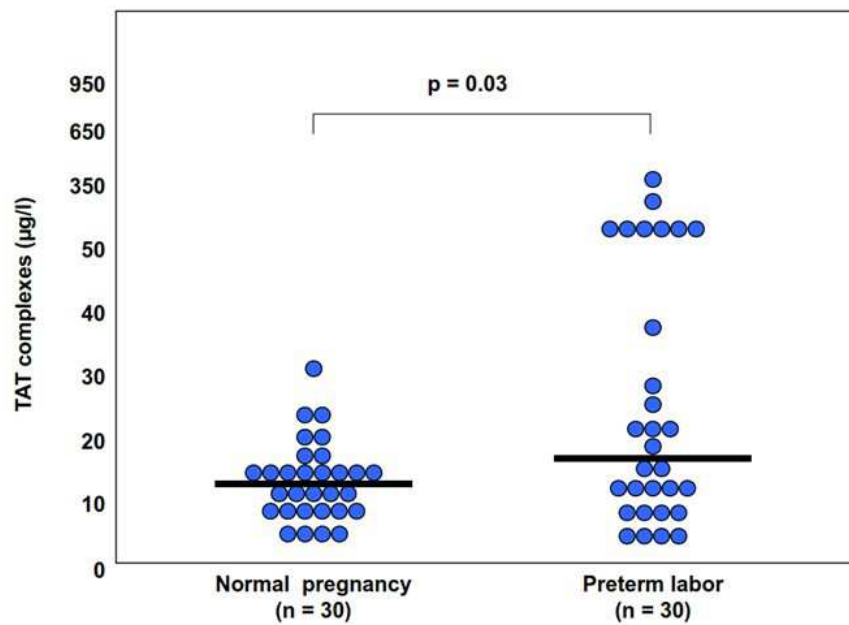
696 Figure 5. Histologic features of villous and intervillous lesions; a. Increased syncytial knots:  
 697 aggregates of syncytiotrophoblast nuclei cluster at one or more poles of distal villi in the vicinity  
 698 of larger stem villi (arrowhead) at the periphery of the lobule ([original magnification is indicated  
 699 for all panels] X 10). b. Villous agglutination: clusters of degenerating distal villi are adherent to  
 700 one another and focally enmeshed in fibrin (X 4). c. Distal villous hypoplasia: a long, thin, non-  
 701 branching stem villus is surrounded by a markedly reduced number of small hypoplastic distal  
 702 villi (X 10). d. Increased intervillous fibrin: stem villi are surrounded by a mantle of fibrin-type  
 703 fibrinoid that does not extend to distal villi at the center of the lobule (X 2). e. Nodular  
 704 intervillous (and intravillous) fibrin: small aggregates of intervillous fibrin adhere to, and are  
 705 focally reepithelialized by, distal villous trophoblast (X 20). f. Increased intervillous fibrin with  
 706 intermediate trophoblast (X-cells): stem and distal villi are enmeshed in a matrix of fibrin and  
 707 fibrinoid containing prominent intermediate trophoblast (arrowhead) (X 10).



708 Figure 6. Amniotic fluid tissue factor concentration among women with normal pregnancies  
 709 (median 3710.4 pg/ml, range 2198.8–6268) and patients with a fetal demise (median 8535.4  
 710 pg/ml, range 2208.2–125,990.0); (b) Amniotic fluid tissue factor activity among women with  
 711 normal pregnancies (median 28.4 pM, range 10.2–84.9) and patients with a fetal demise (median  
 712 81.6 pM, range 7.2–1603.4). From EREZ O, GOTSCH F, MAZAKI-TOVI S, et al. Evidence of  
 713 maternal platelet activation, excessive thrombin generation, and high amniotic fluid tissue factor  
 714 immunoreactivity and functional activity in patients with fetal death. J Matern Fetal Neonatal  
 715 Med 2009;22:672-87, with permission.



716 Figure 7. Maternal plasma TAT III concentration in women with preterm labor (PTL) and those  
717 with a Normal pregnancy (From Chaiworapongsa T, Espinoza J, Yoshimatsu J, Kim YM, Bujold  
718 E, Edwin S, et al.  
719 Activation of coagulation system in preterm labor and preterm premature rupture of membranes.  
720 J Matern Fetal Neonatal Med 2002 11(6):368-373, with permission)





## **Table 1** (on next page)

Table 1

Concentration and activity in maternal plasma of coagulating and anticoagulating factors and their relation with thrombin generation in the great obstetrical syndromes.

Table 1. Concentration and activity in maternal plasma of coagulating and anticoagulating factors and their relation with thrombin generation in the great obstetrical syndromes.

	<b>TF concentration and/or activity</b>	<b>TFPI concentration and/or activity</b>	<b>TAT III complexes concentration</b>	<b>Protein Z concentration</b>	<b>Thrombin generation</b>	<b>References</b>
Premature rupture of membranes	Activity ↑ Concentration ↑	Concentration ↓	Concentration ↑	Concentration ↓	↑	1-5
Preterm labor	Activity ↑ Concentration =	Activity = Concentration ↓	Concentration ↑	Concentration ↓	↑	1-5
Fetal demise	Activity = Concentration =	Activity = Concentration ↓	Concentration ↑	Concentration ↓	↑	1-5
Preeclampsia	Activity ↑ Concentration ↑	Concentration ↓	Concentration ↑	Concentration ↓	↑	1-5
Intrauterine growth retardation	Concentration ↓	Concentration =	Concentration ↑	Concentration ↓	↑	1-5

1. Erez O, Romero R, Vaisbuch E, Kusanovic JP, Mazaki-Tovi S, Chaiworapongsa T, Gotsch F, Fareed J, Hoppensteadt D, Than NG, Yoon BH, Edwin S, Dong Z, Espinoza J, Mazor M, Hassan SS. High tissue factor activity and low tissue factor pathway inhibitor concentrations in patients with preterm labor. *J Matern Fetal Neonatal Med.* 2010 Jan;23(1):23-33
2. Erez O, Gotsch F, Mazaki-Tovi S, Vaisbuch E, Kusanovic JP, Kim CJ, Chaiworapongsa T, Hoppensteadt D, Fareed J, Than NG, Nhan-Chang CL, Yeo L, Pacora P, Mazor M, Hassan SS, Mittal P, Romero R. Evidence of maternal platelet activation, excessive thrombin generation, and high amniotic fluid tissue factor immunoreactivity and functional activity in patients with fetal death. *J Matern Fetal Neonatal Med.* 2009 Aug;22(8):672-87
3. Kusanovic JP, Espinoza J, Romero R, Hoppensteadt D, Nien JK, Kim CJ, et al. Plasma protein Z concentrations in pregnant women with idiopathic intrauterine bleeding and in women with spontaneous preterm labor. *J Matern Fetal Neonatal Med* 2007 Jun;20(6):453-463.
4. Gris JC, Quere I, Dechaud H, Mercier E, Pincon C, Hoffet M, Vasse M, Mares P. High frequency of protein Z deficiency in patients with unexplained early fetal loss. *Blood* 2002;99:2606–2608
5. Paidas MJ, Ku DH, Lee MJ, Manish S, Thurston A, Lockwood CJ, Arkel YS. Protein Z, protein S levels are lower in patients with thrombophilia and subsequent pregnancy complications. *J Thromb Haemost* 2005;3:497–501.