

**A peer-reviewed version of this preprint was published in PeerJ on 27 November 2014.**

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Williams DE, Miller MW, Bright AJ, Cameron CM. 2014. Removal of corallivorous snails as a proactive tool for the conservation of acroporid corals. PeerJ 2:e680 <https://doi.org/10.7717/peerj.680>

# Removal of corallivorous snails as a proactive conservation tool

Corallivorous snail feeding is a common source of tissue loss for the threatened coral *Acropora palmata*, accounting for roughly one-quarter of tissue loss in monitored study plots over seven years. However, corallivory by *Coralliophila abbreviata* is one of the few major sources of partial mortality (contrasting with threats such as bleaching, disease, or storm disturbances) that may be locally managed. We conducted a field experiment to explore the effectiveness and feasibility of snail removal. Long-term monitoring plots on six reefs in the upper Florida Keys were assigned to one of three removal treatments: 1) removal from *A. palmata* only, 2) removal from all host coral species, or 3) no-removal controls. During the initial removal in June 2011, 639 snails were removed from twelve 150 m<sup>2</sup> plots. Snails were removed two additional times during a seven month “removal phase”, then counted at five surveys over the next 19 months to track recolonization. At the conclusion, snails were collected, measured, and sexed. Before-After-Control-Impact analysis revealed that both snail abundance and feeding scar prevalence were reduced in removal treatments compared to the control, but there was no difference between removal treatments. Recolonization by snails to baseline abundance is estimated to be 4.3 years and did not differ between removal treatments. Recolonization rate was significantly correlated with baseline snail abundance. Maximum snail size decreased from 47.0 mm to 34.6 mm in the removal treatments. The effort required to remove snails from *A. palmata* was 30 diver minutes per 150 m<sup>2</sup> plot, compared with 51 minutes to remove snails from all host corals. Since there was no additional benefit observed with removing snails from all host species, removals can be more efficiently focused on only *A. palmata* colonies, and in areas where *C. abbreviata* abundance is high, to effectively conserve *A. palmata* in targeted areas.

1 D. E. Williams<sup>1,2</sup>, M. W. Miller<sup>2</sup>, A. J. Bright<sup>1,2</sup>, C. M. Cameron<sup>1,2</sup>

2 *Author Addresses:*

3 <sup>1</sup> Cooperative Institute for Marine and Atmospheric Studies, University of Miami, 4600 Rickenbacker

4 Cswy, Miami, FL 33149 USA

5 <sup>2</sup> National Marine Fisheries Service, Southeast Fisheries Science Center 75 Virginia Beach Dr., Miami, FL

6 33149 USA

## 7 Introduction

Predator control is most commonly considered as a management strategy for invasive predators (Baxter et al. 2008; Barbour et al. 2011; Morris Jr et al. 2011) or outbreaks of endemic predators (Yamaguchi 1986; Sanz-Aguilar et al. 2009). Previous attempts to cull corallivores, specifically *Acanthaster planci*, have largely been aimed at localized outbreaks with the goal of preserving coral tissue over a large area (Yamaguchi 1986). These efforts have been deemed ineffective due to the large numbers and migrating aggregations of these predators (Yamaguchi 1986; Johnson et al. 1990). However, removal of a relatively sedentary predator from targeted populations of a threatened coral species has not been evaluated.

Ecological theory on predator-prey dynamics can provide insight on situations when predator removal may be effective in protecting prey. Sinclair et al. (1998) present a framework whereby controlling natural predators may improve the outcome for management of declining or reintroduced populations of threatened species. In this framework, the appropriate scale of intervention depends on the functional and numerical response of predators to changing prey abundance. In cases where the effects of predation are depensatory and prey abundance is so low that they are vulnerable to stochastic events, predator control could provide benefit to prey populations (Sinclair et al. 1998). Rotjan and Lewis (2008), in a review of corallivory, suggest that the rapid pace of coral decline over the past two decades, largely from factors other than predation, may have indeed reached such a depensatory threshold such that predation is exerting undue influence, potentially compromising coral reef resilience.

On reefs in the western Atlantic, the dominant framework builder, *Acropora palmata*, is preyed upon by the corallivorous snail *Coralliophila abbreviata*. Although disease, storms, and bleaching have largely driven the range-wide decline of *A. palmata* populations, snail predation is

30 recognized as one of the top three threats to the persistence and recovery of these populations  
 31 (Bruckner 2002; Williams and Miller 2012). In the upper Florida Keys, there has been a 50%  
 32 decline in *A. palmata* tissue abundance since 2004. Although the main culprit has been disease,  
 33 feeding by *C. abbreviata* accounted for an estimated one-quarter of the observed tissue loss  
 34 (Williams and Miller 2012). It is unknown whether or not the *C. abbreviata* population is  
 35 increasing; however, typical predators of shelled gastropods are grunts, wrasses, trunkfish,  
 36 triggerfish, and pufferfish (Randall 1967). Therefore it is possible that reduced predation on  
 37 snails due to overfishing may have increased snail abundance (Burkepile and Hay 2007), though  
 38 specific data to support this hypothesis are lacking. Regardless, as *A. palmata* populations  
 39 decline, snails have been observed to become more concentrated on the remaining *A. palmata*  
 40 (Bruckner et al. 1997; Bruckner 2000; Baums et al. 2003a; Williams and Miller 2012) rather than  
 41 declining themselves, suggesting increasing per capita impact on prey.

42 *C. abbreviata* preys on multiple coral host species including acroporids, *Orbicella* spp.,  
 43 *Diploria* spp. *Colpophylia natans*, *Agaricia* spp., and occasionally on other mounding coral  
 44 species (Miller 1981). Snails found on *A. palmata* are larger, older, have higher fecundity  
 45 (Johnston and Miller 2007), and consume more coral tissue than on other coral host species  
 46 (Bruckner 2000). The snails are typically found in groups (Bruckner et al. 1997; Bruckner 2000;  
 47 Baums et al. 2003a), feeding on coral tissue and leaving a feeding scar of exposed skeleton. They  
 48 are relatively sedentary, often remaining on a prey colony until no living tissue remains, at which  
 49 point they migrate to a neighboring colony (Bruckner 2000; Williams and Miller 2012).

50 Individual snails can consume up to 16 cm<sup>2</sup> of tissue per day (Brawley and Adey 1982; Baums et  
 51 al. 2003b), though they do not feed continuously throughout the year at that rate (Bruckner et al.  
 52 1997). In addition to directly removing *A. palmata* tissue during feeding, *C. abbreviata* may  
 53 indirectly affect corals by way of vectoring disease (Williams and Miller 2005) or attracting other

54 predators such as butterflyfish (Brawley and Adey 1982) and *Hermodice carunculata* (DW, pers  
55 obs). Thus, *C. abbreviata* has substantial direct and potential indirect effects on *A. palmata*.  
56 Because this predator has low mobility and a relatively long lifespan (up to 15 years; Johnston  
57 and Miller 2007), it may be feasible to locally reduce their abundance to conserve *A. palmata*.  
58 *Acropora palmata* was listed as threatened under the US Endangered Species Act (NMFS  
59 2006) and has been proposed for uplisting to endangered (NMFS 2012) based on devastating  
60 declines throughout its range. The ESA listing carries with it a mandate to pursue management  
61 actions to foster recovery of the species (ESA, section 4f). Although predation is not the primary  
62 factor causing decline of this species, recent trajectories suggest it may be a fundamental factor  
63 inhibiting recovery and, at present, predation may be the most locally tractable threat. Even in  
64 regions where *A. palmata* is relatively rare, such as the Florida Keys, its distribution is clumped  
65 making targeted removal efforts logistically feasible. Therefore, both ecological and  
66 legal/management conditions point to removal of *C. abbreviata* as a potential conservation action  
67 that could be feasible at the local level. Earlier work (Miller 2001) showed removing *C.*  
68 *abbreviata* snails on a colony scale can conserve *A. palmata* tissue, but nothing is known about  
69 effectiveness in terms of recolonization rates or at a larger 'reef scale'. The current study utilized  
70 long-term fixed monitoring plots of *A. palmata* colonies to conduct experimental *C. abbreviata*  
71 (here after 'snail') removals to 1) determine the rate at which snails recolonize *A. palmata*  
72 colonies, 2) evaluate detectable impacts on the host *A. palmata* population, 3) compare the size  
73 distribution of recolonizing versus original snail populations, and 4) evaluate the costs of removal  
74 for resource managers.

## 75 **Methods**

76 Long-term *Acropora palmata* demographic monitoring plots (7 m radius) at six sites in the  
77 upper Florida Keys National Marine Sanctuary (FKNMS) were used to implement a Before-  
78 After-Control-Impact (BACI) type design (Green 1979; Smith 2006) to evaluate the effects of  
79 snail removal. This design is useful in natural settings because initial variation among individual  
80 plots can be partitioned from treatment effects by comparing each plot's trajectory over time  
81 (before vs. after a manipulation) among plots subjected to different treatments. Each site included  
82 three plots numbered one to three when initially established for monitoring at least 1 year prior to  
83 the start of the experiment. Three snail removal treatments were assigned according to plot  
84 number: 1) removal of snails from *A. palmata* colonies only ("Ap Only"), 2) removal of snails  
85 from all host corals in the plot ("All Hosts"; mainly *A. palmata*, *Diploria* spp., *Orbicella* spp.,  
86 and *Colpophylia natans*), and 3) "Control" in which snails were counted on the host corals in the  
87 plots, but were not removed.

## 88 **Removal**

89 This experiment proceeded in two phases (described in detail below); the first phase is  
90 referred to as the 'removal phase' and consisted of three removals beginning with an 'initial  
91 removal' of snails that provided the 'baseline' snail abundance and ending with a 'final removal'.  
92 The second phase is the 'recolonization phase' consisting of five surveys beginning after 'final  
93 removal' and ending at the 'final survey' when the snails were collected for analysis.

94 Removal of snails from host corals was performed by two SCUBA divers that were  
95 experienced in finding this somewhat cryptic gastropod species. Individual host colonies were  
96 searched, and when snails were found in the removal plots, the diver recorded the host species  
97 and number of snails on the colony. Snails were removed and placed in zip-top bags, pooled  
98 according to host species. Snail shell length was measured to the nearest 0.1 mm using Vernier

calipers. Due to the high abundance and small size of *Agaricia* spp. colonies, it was not feasible to systematically locate all the colonies. However, when *Agaricia* colonies were encountered in the All Hosts and Control plots, they were searched, and snails found in the All Host plots were removed. Initial removal was conducted during 14-16 June 2011. Three weeks later we returned to the treatment (Ap Only and All Hosts) plots to remove and measure any snails that were overlooked in the initial removal. The snails from these two ‘removals’ were added together as the ‘baseline’ snail abundance for each plot and will be referred to as the “initial” removal.

Snails were removed from all *A. palmata* colonies in the treatment plots again in September 2011 and January 2012 (‘removal phase’). Thereafter, snails found on *A. palmata* were counted but not removed in May 2012, September 2012, January 2013, May 2013 and August 2013 (‘recolonization phase’). At the final survey in August 2013, all host colonies were measured (length, width, height, and % live) and surveyed for snails in all plots. Snails were collected from all hosts in the All Hosts treatment plots and *A. palmata* only in the Ap Only treatment plots. All snails collected at the final survey were measured to compare the recolonized population with the initial population. Shells were crushed to determine the presence (designating males) or absence (designating females) of a penis. The time spent searching the host colonies and removing encountered snails was recorded during the final survey and the averages among plots were used to evaluate the ‘effort’ of each removal treatment.

During the removal and recolonization phases, routine monitoring of *A. palmata* in the study plots continued as described in Williams et al. (2008). Once per year (fall), all *A. palmata* colonies in each plot were counted and the length, width, and height were measured and % live tissue was visually estimated. Tissue abundance was estimated as a live area index (LAI), calculated by taking the colony’s average dimension (average of length, width, and height) squared and multiplying it by the visual estimate of % live tissue cover. The LAI is summed for



all colonies to get total tissue abundance for the 150 m<sup>2</sup> study plot. Three times per year a randomly selected subset of tagged *A. palmata* colonies was further assessed for size, % live tissue, and presence of disease, snails, and snail feeding scars. This work was done under permit numbers FKNMS-2010-033, FKNMS-2010-130 and FKNMS-2012-030 from the Florida Keys National Marine Sanctuary.

## Analyses

### *Effectiveness of Removal*

We examined the total number of snails found on *A. palmata* colonies in the plot (ApSnails), the tissue abundance (LAI) of all *A. palmata* in the study plot (ApLAI), and the prevalence of disease and feeding scars among a subset of tagged *A. palmata* colonies. We used a BACI (Before-After-Control-Impact) design to compare the statistical interaction between time and treatment for the removal treatments and control. The total number of ApSnails and the ApLAI was compared among treatments using the initial (June 2011) and final (August 2013) surveys. Because the prevalence of both disease and feeding scars vary temporally (Williams and Miller 2012), we compared the peak in prevalence from the year before the removal and the subsequent peak observed after the removal. For disease prevalence we compared fall 2010 (before) and fall 2011 (after) and for feeding scars we compared spring 2011 (survey prior to removal) and spring 2012 (first survey after final removal). Each variable (ApSnails, ApLAI, and prevalence of disease and feeding scars) was rank transformed to meet the assumption of normality and homoscedasticity, and a repeated measures ANOVA was run on the ranks to look for significant ( $p \leq 0.05$ ) within-subject interactions between time (the Before/After factor) and treatments (the Control/Impact factor) indicating that the trend in the measured parameter varied significantly between treatments.

### *Recolonization*

We examined the rate at which snails recolonized *A. palmata* in removal plots over the recolonization phase. The number of snails present following the final removal in January 2012 was assumed to be zero and the number found at each subsequent survey during this recolonization phase was plotted over time for each study plot and linear regression was used to determine the equation for the line. With y set to the baseline snail abundance for that plot, we solved for the projected date (x) that the number of snails in that plot would return to its baseline abundance observed at the initial removal. The difference between the projected date and the date of the January 2012 removal was calculated as the treatment ‘effect duration’ for that plot. A Wilcoxon Matched-Pairs test was used to compare the baseline snail abundance, recolonization rate, and effect duration between Ap Only and All Hosts removal treatments paired within site.

### *Snail size*

In order to compare the population of recolonized versus initial snails, shell length of the collected snails was measured and the data were log transformed to achieve normality. A two-way ANOVA was used to compare shell lengths between the two removal treatments and time (initial removal vs final survey). The size-frequency distributions of males and females at the final survey could not be compared between treatments due to small sample size ( $n \leq 39$ ; Table 1), but the proportion of the population that was male was calculated as the number of male snails divided by the total number of snails.

## **Results**

### **Removal**

Searching each 150 m<sup>2</sup> study plot required on average 30 person minutes ( $\pm 16$  minutes SD) for *A. palmata* and an additional 21 person minutes ( $\pm 10$  minutes SD) when the other host species were searched. A total of 279 snails were removed from *A. palmata* in the twelve 150 m<sup>2</sup>

removal treatment plots (Table 1). In the follow-up removal three weeks later, an additional 40 were found for a total of 319 snails that were removed from *A. palmata* in the removal plots at the initial survey in June 2011. A total of 157 snails were removed from other hosts in the ‘all hosts’ treatment plots.

The mean number of snails found on *A. palmata* (ApSnails) in the removal plots remained less than five per plot during the removal phase, then gradually increased after removals stopped in January 2012 (Fig. 1). The interaction between time and treatment was significant for both the total number of ApSnails per plot (Fig. 2a;  $p = 0.042$ ) and the prevalence of feeding scars (Fig. 2d;  $p = 0.004$ ); for both, the removal treatments decline significantly while the controls remained unchanged. The tissue abundance (LAI) showed no change among all treatments (Fig. 2b). The prevalence of disease was significantly higher in fall 2011 compared to fall 2010 (Fig. 2c;  $p = 0.016$ ) with no significant treatment effects or interaction.

## Recolonization

Despite high variability among individual removal plots, linear regressions of the number of snails found at each survey (Fig. 3) yielded  $r$  values  $\geq 0.7$  within each plot. Both the baseline abundance of ApSnails and the rate of snail recolonization (regression slopes of 0.002 to 0.050 snails  $d^{-1}$ ) varied by an order of magnitude among reefs (Fig. 3). When treatment plots were paired by site, neither the baseline abundance of ApSnails nor the recolonization rate differed between treatments. If the Sand Island pair is excluded as an outlier (based on the extreme number of snails found in the Sand Island All Hosts plot, Table 1), then the baseline number of snails still does not differ between treatments, but the difference in recolonization rate between treatments becomes marginally significant (Wilcoxon,  $Z = 2.02$ ,  $n = 5$ ,  $p = 0.043$ ). The baseline snail abundance and recolonization rate were highly correlated across all removal plots (Fig. 4). Overall, calculated ‘effect durations’ ranged from 1.3 to 6.9 years (Fig. 3) with an overall average of  $4.3 \pm 1.7$  ( $\pm$  SD) years. Effect durations did not differ significantly between Ap Only removal

195 (4.5 ± 1.6 years; mean ± SD) and the All Hosts removal (5.5 ± 1.6 years; mean ± SD) treatments  
196 (Wilcoxon, Z = 0.31, n = 6, p = 0.8).

### 197 **Size frequency/Sex ratios**

198 A two-way ANOVA on log-transformed shell lengths (size) was used to compare the  
199 recolonized versus initial ApSnail populations between the two removal treatments (Fig. 5).  
200 Mean shell length was significantly larger between the initial (24.4 mm ± 7.8 mm, pooled mean ±  
201 SD;  $F_{1,465} = 8.61$ ,  $p = 0.003$ ; Fig. 5a-b) and final (22.1 mm ± 5.7 mm, pooled mean ± SD; Fig. 5c-  
202 d) surveys but not between the two removal treatments ( $F_{1,465} = 1.09$ ,  $p = 0.3$ ) and there was no  
203 significant interaction ( $F_{1,465} = 0.16$ ,  $p = 0.7$ ). A separate two-way ANOVA was also used to  
204 compare the initial and final log transformed shell lengths between ApSnails (pooled treatments)  
205 and the snails collected from other host corals in the All Hosts treatments (Fig. 6). Snails  
206 collected from other host corals were significantly smaller (18.8 mm ± 3.6 mm, mean ± SD;  $F_{1,710}$   
207 = 103.58,  $p < 0.001$ ) than those collected from *A. palmata* (24.4 mm ± 7.8 mm, pooled mean ±  
208 SD), and the snails collected at the initial survey were significantly larger (22.5 mm ± 7.2 mm,  
209 pooled mean ± SD;  $F_{1,710} = 16.83$ ,  $p < 0.001$ ) than those collected at the final survey (20.3 mm ±  
210 5.9 mm, pooled mean ± SD), but there was no significant interaction ( $F_{1,710} = 0.23$ ,  $p = 0.6$ )  
211 between host and time factors.

212 At the final survey, the proportion of males among ApSnails in the Ap Only treatment was  
213 0.66 (54 males and 28 females) and 0.74 (52 males and 18 females) in the All Hosts treatment.  
214 Unfortunately, we do not have gender ratios for the initial population, but as larger snails are  
215 known to be female in this protandrous species (Johnston and Miller 2007), the larger snail sizes  
216 of the initial population would be expected to reflect a lesser proportion of males relative to the  
217 final population.

## 218 Discussion

219 Removal was effective in significantly decreasing both corallivore abundance and the  
220 prevalence of feeding scars observed 19 months following the snail removal. Consequent  
221 declines in the prevalence of disease and a parallel enhancement of total tissue abundance (LAI)  
222 might be expected, but both of these factors are strongly influenced by a multitude of additional  
223 known and unknown factors, thereby decreasing the ability to detect these parallel changes in the  
224 present study. However, based on the significantly lower prevalence of feeding scars we can  
225 deduce that less tissue was consumed by snails (Miller 2001).

226 The recolonization rate calculations, though complicated by high site-specific variability,  
227 indicate full recolonization to the baseline abundance of snails over a 4-year period. At the final  
228 survey, the average size of recolonizing snails was smaller than at the initial removal. Though the  
229 decrease in mean snail size was modest, the larger sized ApSnails ( $\geq 35$  mm) were not observed  
230 to recolonize at all (Fig. 5). At the initial removal, the maximum snail size was 47.0 mm and at  
231 the final survey the maximum was 34.6 mm. The cumulative tissue consumption of smaller snails  
232 is expected to be less than for larger snails (Hayes 1989; Bruckner 2000; Baums et al. 2003b);  
233 thus, although this portion of the size distribution represented approximately 10% of the  
234 population, these larger snails likely were inflicting greater than 10% of the tissue loss.

235 In addition to the direct impact of feeding, fecundity is disproportionately higher in these  
236 large snails (Johnston and Miller 2007). At the initial removal there were 42 snails found with  
237 shell length  $> 34.6$  mm. Based on the relationship between shell length and veliger production  
238 (Johnston and Miller 2007), we can estimate that these 42 snails, assuming they were all females,  
239 would have produced  $> 715,000$  veligers in one reproductive cycle (clutch). For comparison, if we  
240 look at the largest 10% of the final size distribution (females ranging from 29.8 to 34.6 mm),  
241 these 17 females are expected to yield approximately 155,000 veligers in one reproductive cycle.

242 In fact, all 46 female ApSnails found at the final survey combined would be predicted to yield  
243 280,000 veligers in one clutch. Thus, the combination of fewer snails and the shift to smaller  
244 sizes could potentially decrease snail reproductive output by more than 50%.

245 In the 4.3 years projected for full recolonization, it is possible that the snail size distribution  
246 will also shift back to baseline size. Based on the size-age relationship reported by Johnston &  
247 Miller (2007), the largest individual observed at the start of the experiment (i.e., 47 mm) might be  
248 approximately 15 years old while the maximum size observed at the final survey would be  
249 expected to be approximately 9 years old. However, it is also possible that the smaller sizes are  
250 more vulnerable to predation due to thinner shells (e.g., Wainwright 1987) and will be kept from  
251 attaining the larger less vulnerable sizes.

252 Recolonizing individuals appear to be primarily migrants from surrounding reef areas rather  
253 than larval recruits. Although there were small increases in the smallest size bins (Fig. 6a and c),  
254 the smallest recolonizing individuals found (~ 8 mm) are expected to be ~ 3 years old (Johnston  
255 and Miller 2007), significantly older than the 1.5 years duration of the recolonization phase in  
256 this experiment (January 2012 - Aug 2013). However, if the other host corals were the only  
257 source of snails that recolonized *A. palmata* colonies, then we would expect a difference in size  
258 distribution and numbers of recolonizing snails between the two removal treatments (Fig. 5c-d),  
259 which was not the case. Although the snails on other host corals in the Ap Only treatment were  
260 not measured at the start of the experiment, they would likely have had the same size distribution  
261 as those collected in the All Hosts treatment at the start (Fig. 6b). Presumably, if these other host  
262 coral species were the primary source of recolonizing ApSnails, then the size distribution of  
263 ApSnails at the final survey (Fig. 5c) would be similar to the distribution of the snails collected  
264 from other host corals at the initial removal (Fig. 6b). Specifically, there were relatively few  
265 snails larger than 24 mm on the other host corals, yet more than one-third of recolonizing snails

were larger than 24 mm (Fig. 6c). Instead, the size distribution of recolonizing snails in both removal treatments looks the same (Fig. 5c-d). Although the mean snail size is overall smaller at the final survey than at the start, the mean size of ApSnails is still significantly larger (22.1 mm) in the final survey than snails collected from other hosts (18.8 mm) at the initial removal (Fig. 6).

In our experiment, no significant added benefit was derived from the additional effort required to remove snails from all host corals (Fig. 1). Removing snails from *A. palmata* only required 30 diver minutes versus 51 diver minutes for removal from all hosts. The density of other host colonies in the plots was  $5 \pm 4.9$  colonies (mean  $\pm$  SD) and a total of  $21 \pm 15.6$  (mean  $\pm$  SD) snails per plot were found on these other host colonies. If the site with the unusually high number of snails was excluded, the recolonization rates were marginally faster in plots where the snails were not removed from other hosts. It is possible that in areas where the other hosts species are more abundant or are harboring greater numbers of snails, the additional effort to remove them would be worthwhile.

We removed snails at three surveys roughly three months apart and each requiring roughly 30 minutes of diver time. Interestingly, during these three sequential removals, the average number of snails found on the subsequent survey did not diminish (Fig. 1; mean of ~2 to 5 snails) so it does not seem that the pool of colonizing snails was depletable at these temporal and spatial scales. There is also indication that, at least in some plots, the rate of snail arrival may accelerate over time, which is consistent with the aggregating behavior of snails (DW, pers obs).

In planning snail removal as an *A. palmata* conservation effort, the cost (diver time) and benefit (reduced snail load) must be balanced. With the mean effect duration of four years, one strategy could be to perform a removal at four year intervals. However, our results showing high site variability and the strong correlation of recolonization rate with initial snail load (Fig. 4) suggest that the frequency or need for subsequent removals may be indicated by the number of

snails that are found at the initial removal. In another view, the mean rate of snail recolonization appears to increase after one year (Fig. 1), so annual removal might be a useful target, at least for areas of high snail abundance ( $>0.2 \text{ m}^{-2}$ ). Removals in the warmer months are likely to be more efficient when corallivorous snails are more actively feeding (Al-Horani et al. 2011). Additionally, although their egg production cycle is not well established, it is more common to find egg cases with mature veligers in mid-to-late summer (DW, pers obs), thus, removal prior to that may reduce larval production.

Our sites were located on spur and groove formations on the shallow fore reef. However, our study plots did not occupy the full extent of a reef 'spur', leaving contiguous reef areas populated with snails. In practice, removal from all corals on a contiguous spur may further prolong the effect duration of the removal. This may not be practical in areas where there are not natural breaks in reef structure, but removal from contiguous stands of *A. palmata* may be possible. Effect of *A. palmata* colony density on snail recolonization rate was not tested in this study; however, other studies have found that *C. abbreviata* abundance is generally lower in higher density 'thicket'-type stands (Bruckner et al. 1997; Miller et al. 2002; Baums et al. 2003a) of *A. palmata* colonies, so removal effort could be more efficiently focused on *A. palmata* stands with intermediate or low colony density rather than dense thickets. Although removing snails is not technically difficult, *C. abbreviata* is fairly well camouflaged and divers need to be trained to recognize them to ensure effective removal and to minimize collection of other non-corallivorous species such as *Thais deltoidea* that are commonly found around *A. palmata*.

Given the ecologically and legally imperiled status of *A. palmata*, and the intractability of managing or reducing many of its ongoing threats, proactive conservation measures that can be implemented at a local level are needed. This experiment demonstrates the effectiveness of snail



313 removal at a local scale with a 30 minute diver investment effectively reducing corallivore loads  
314 over an estimated four year time scale in seven meter radius plots containing *A. palmata*.

## 315 Acknowledgements

316 This project was made possible by support from the NOAA Coral Reef Conservation Program  
317 and NMFS Southeast Regional Office, Protected Resources Division and was conducted under  
318 permit # FKNMS-2010-033, FKNMS-2010-130 and FKNMS-2012-030. Field assistance by K.  
319 Lohr, C. Kiel, and L. Richter is greatly appreciated.

## 320 References

- 321 Al-Horani FA, Hamdi M, Al-Rousan SA (2011) Prey selection and feeding rates of *Drupella*  
322 *cornus* (Gastropoda: Muricidae) on corals from the Jordanian coast of the Gulf of Aqaba,  
323 Red Sea. Jordan Journal of Biological Sciences 4:191-198
- 324 Barbour AB, Allen MS, Frazer TK, Sherman KD (2011) Evaluating the potential efficacy of  
325 invasive lionfish (*Pterois volitans*) removals. PLoS ONE 6:e19666
- 326 Baums IB, Miller MW, Szmant AM (2003a) Ecology of a corallivorous gastropod, *Coralliophila*  
327 *abbreviata*, on two scleractinian hosts. I: population structure of snails and corals. Mar  
328 Biol 142:1083-1091
- 329 Baums IB, Miller MW, Szmant AM (2003b) Ecology of a corallivorous gastropod, *Coralliophila*  
330 *abbreviata*, on two scleractinian hosts. II. feeding, respiration and growth. Mar Biol  
331 142:1093-1101
- 332 Baxter PWJ, Sabo JL, Wilcox C, McCarthy MA, Possingham HP (2008) Cost-effective  
333 suppression and eradication of invasive predators. Conserv Biol 22:89-98
- 334 Brawley SH, Adey WH (1982) *Coralliophila abbreviata*: a significant corallivore. Bull Mar Sci  
335 32:595-599
- 336 Bruckner AW (2002) Proceedings of the Caribbean *Acropora* Workshop: potential application of  
337 the U.S. Endangered Species Act as a conservation strategy. NOAA Technical  
338 Memorandum NMFS-OPR-24, Silver Spring, MD, p 199
- 339 Bruckner RJ (2000) Effects of the population dynamics of the corallivorous prosobranch  
340 gastropod *Coralliophila abbreviata* (Lamarck) on coral tissue predation. M.S. thesis,  
341 University of Puerto Rico, p 113
- 342 Bruckner RJ, Bruckner AW, Williams EHJ (1997) Life history strategies of *Coralliophila*  
343 *abbreviata* Lamarck (Gastropoda: Coralliophilidae) on the southeast coast of Puerto Rico.  
344 Proc 8th Int Coral Reef Symp 1:627-632
- 345 Burkepile DE, Hay ME (2007) Predator release of the gastropod *Cyphoma gibbosum* increases  
346 predation on gorgonian corals. Oecologia 154:167-173

- 347 Green RH (1979) Sampling design and statistical methods for environmental biologists. John  
348 Wiley & Sons
- 349 Hayes JA (1989) The biology and ecology of *Coralliophila abbreviata* Lamarck (Gastropoda,  
350 Coralliophilidae) : the importance of corallivores in the ecology of coral reefs. Ph.D.  
351 thesis, University of Texas at Austin, p 177
- 352 Johnson DB, Moran PJ, Driml S (1990) Evaluation of a crown-of-thorns starfish (*Acanthaster*  
353 *planci*) control program at Grub Reef (central Great Barrier Reef). Coral Reefs 9:167-171
- 354 Johnston L, Miller MW (2007) Variation in life-history traits of the corallivorous gastropod  
355 *Coralliophila abbreviata* on three coral hosts. Mar Biol 150:1215-1225
- 356 Miller AC (1981) Cnidarian prey of the snails *Coralliophila abbreviata* and *C. caribaea*  
357 (Gastropoda: Muricidae) in Discovery Bay, Jamaica. Bull Mar Sci 31:932-934
- 358 Miller MW (2001) Corallivorous snail removal: evaluation of impact on *Acropora palmata*.  
359 Coral Reefs 19:293-295
- 360 Miller MW, Baums IB, Williams DE, Szmant AM (2002) Status of Candidate coral, *Acropora*  
361 *palmata*, and its snail predator in the upper Florida Keys National Marine Sanctuary:  
362 1998-2001. NOAA Technical Memorandum, Miami, FL, p 26
- 363 Morris Jr JA, Shertzer KW, Rice JA (2011) A stage-based matrix population model of invasive  
364 lionfish with implications for control. Biol Invasions 13:7-12
- 365 Federal Register (2006) Endangered and threatened species: final listing determinations for  
366 Elkhorn coral and Staghorn coral. Federal Register:26852-26872
- 367 Federal Register (2012) Endangered and Threatened wildlife and plants: proposed listing  
368 determinations for 82 reef-building coral species; proposed reclassification of *Acropora*  
369 *palmata* and *Acropora cervicornis* from Threatened to Endangered. Federal  
370 Register:73220-73262
- 371 Randall JE (1967) Food habits of reef fishes of the West Indies. Stud Trop Oceanog 5:665-847
- 372 Rotjan RD, Lewis SM (2008) Impact of coral predators on tropical reefs. Mar Ecol Prog Ser  
373 367:73-91
- 374 Sanz-Aguilar A, Martínez-Abraín A, Tavecchia G, Mínguez E, Oro D (2009) Evidence-based  
375 culling of a facultative predator: efficacy and efficiency components. Biol Conserv  
376 142:424-431
- 377 Sinclair ARE, Pech RP, Dickman CR, Hik D, Mahon P, Newsome AE (1998) Predicting effects of  
378 predation on conservation of endangered prey. Conserv Biol 12:564-575
- 379 Smith EP (2006) BACI Design. In: El-Shaarawi AH, Piegorisch WW (eds) Encyclopedia of  
380 Environmetrics. John Wiley & Sons, Ltd, Chichester, pp 141-148
- 381 Wainwright PC (1987) Biomechanical limits to ecological performance: mollusc-crushing by the  
382 Caribbean hogfish, *Lachnolaimus maximus* (Labridae). J Zool 213:283-297
- 383 Williams DE, Miller MW (2005) Coral disease outbreak: pattern, prevalence and transmission in  
384 *Acropora cervicornis*. Mar Ecol Prog Ser 301:119-128
- 385 Williams DE, Miller MW (2012) Attributing mortality among drivers of population decline in  
386 *Acropora palmata* in the Florida Keys (USA). Coral Reefs 31:369-382
- 387 Williams DE, Miller MW, Kramer KL (2008) Recruitment failure in Florida Keys *Acropora*  
388 *palmata*, a threatened Caribbean coral. Coral Reefs 27:697-705
- 389 Yamaguchi M (1986) *Acanthaster planci* infestations of reefs and coral assemblages in Japan: a  
390 retrospective analysis of control efforts. Coral Reefs 5:23-30

391 **Figure captions**

392 Figure 1 Number of *Coralliophila abbreviata* found on *Acropora palmata* per plot (mean  $\pm$  SE).  
393 The initial removal occurred in June 2011, remaining snails were removed through January 2012  
394 (removal phase, gray dots) after which they were only counted and left in place during the survey  
395 phase (solid dots)

396 Figure 2 Before-After-Control-Impact analysis of a) the total number of *Coralliophila*  
397 *abbreviata* found on *Acropora palmata* in a study plot, b) the *A. palmata* tissue abundance as  
398 measured by the live area index (LAI, see text), c) the prevalence of white disease on a random  
399 subset of *A. palmata* colonies during the seasonal peak in disease before and after the initial  
400 removal, and d) the prevalence of *C. abbreviata* feeding scars on this random subset of *A.*  
401 *palmata* colonies at the survey prior to the initial removal and one year later. All points are mean  
402  $\pm$  standard error. Data were rank transformed for analysis and the p-values based on the  
403 transformed data are shown

404 Figure 3 Number of *Coralliophila abbreviata* snails found on *Acropora palmata* in each plot  
405 where they were removed from *Acropora palmata* only (Ap Only) and from all host coral species  
406 following the removal phase of the experiment. The dotted line indicates the baseline number of  
407 *C. abbreviata* that were removed from that treatment plot at the start of the experiment in June  
408 2011. 'Effect duration' is the estimated time for the number of snails to reach the baseline  
409 (recolonization), according to the regression for each plot

410 Figure 4 Recolonization rate (number of snails per day) based on the slope of the linear  
411 regressions (Fig. 3) versus the number of snails found in each study plot at the initial removal

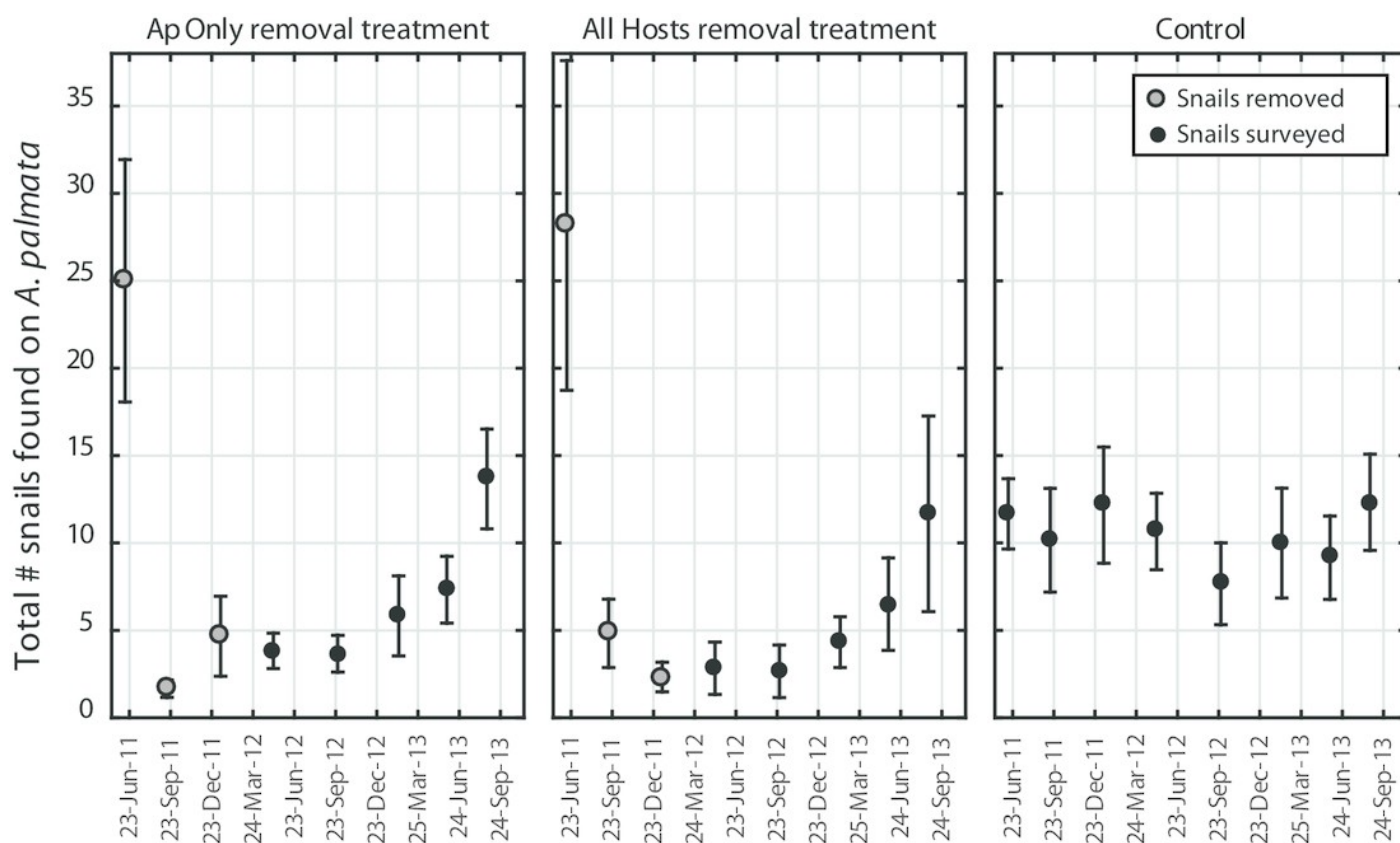
412 Figure 5 Size and gender (Sept 2013 only) frequency distribution for the *Coralliophila*  
413 *abbreviata* snails collected from *Acropora palmata* host colonies in the a) Ap Only (snails  
414 removed from *A. palmata* only) and b) All Hosts (snails removed from all host coral species)  
415 treatments at the initial removal (June 2011) and at the final survey (Sept 2013) for c) Ap Only  
416 and d) All Hosts treatments

417 Figure 6 Size and gender (Sept 2013 only) frequency distribution for the *Coralliophila*  
418 *abbreviata* snails collected from a) *Acropora palmata* host colonies and b) other host species at  
419 the initial removal (June 2011) and at the final survey (Sept 2013) from c) *A. palmata* and d) and  
420 other host coral species

# Figure 1

## *Coralliophila abbreviata* abundance in experimental plots

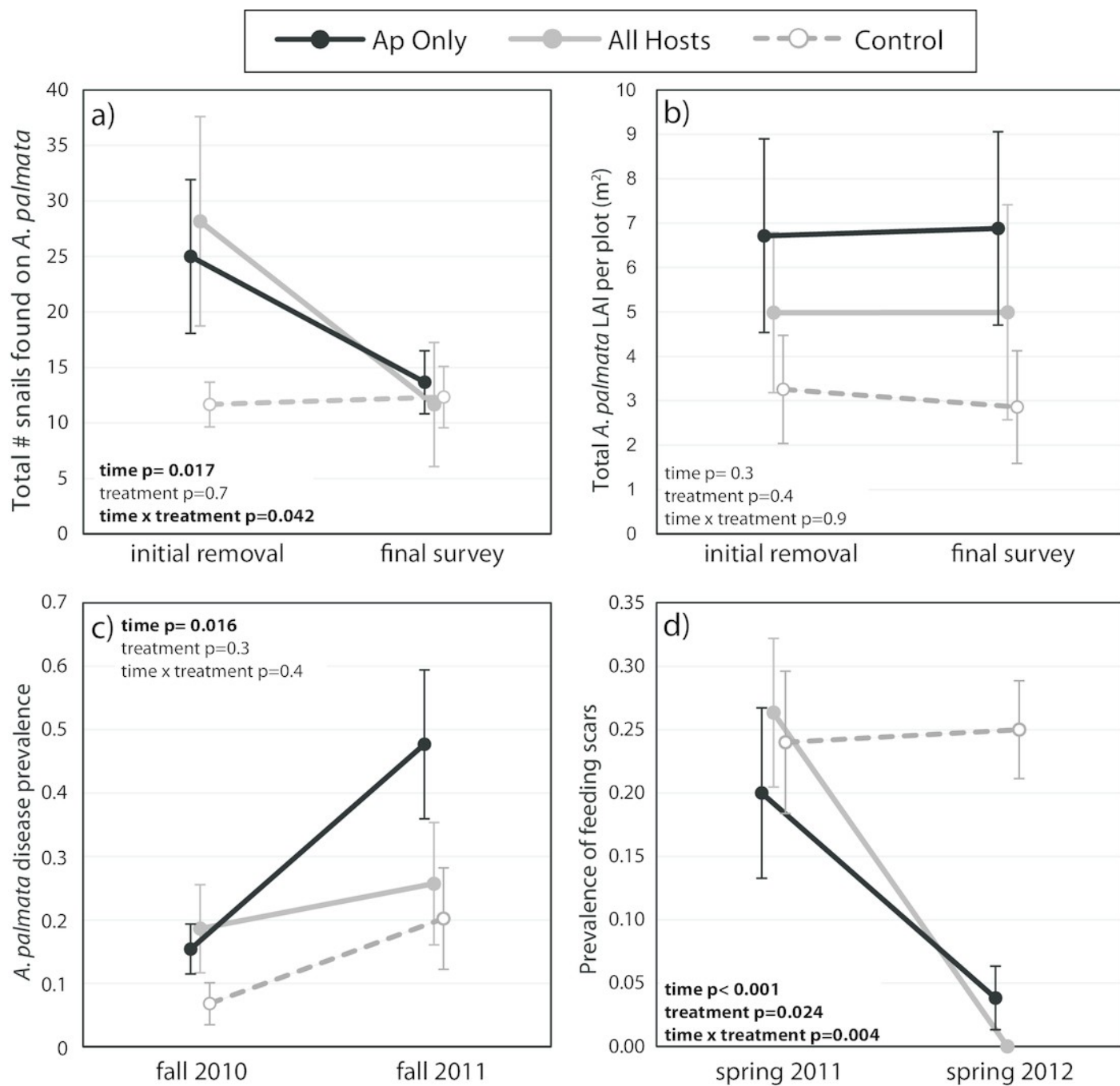
Number of *Coralliophila abbreviata* found on *Acropora palmata* per plot (mean  $\pm$  SE). The initial removal occurred in June 2011, remaining snails were removed through January 2012 (removal phase, gray dots) after which they were only counted and left in place during the survey phase (solid dots).



## Figure 2

### Before-After-Control-Impact analysis

Before-After-Control-Impact analysis of a) the total number of *Coralliophila abbreviata* found on *Acropora palmata* in a study plot, b) the *A. palmata* tissue abundance as measured by the live area index (LAI, see text), c) the prevalence of white disease on a random subset of *A. palmata* colonies during the seasonal peak in disease before and after the initial removal, and d) the prevalence of *C. abbreviata* feeding scars on this random subset of *A. palmata* colonies at the survey prior to the initial removal and one year later. All points are mean  $\pm$  standard error. Data were rank transformed for analysis and the p-values based on the transformed data are shown.

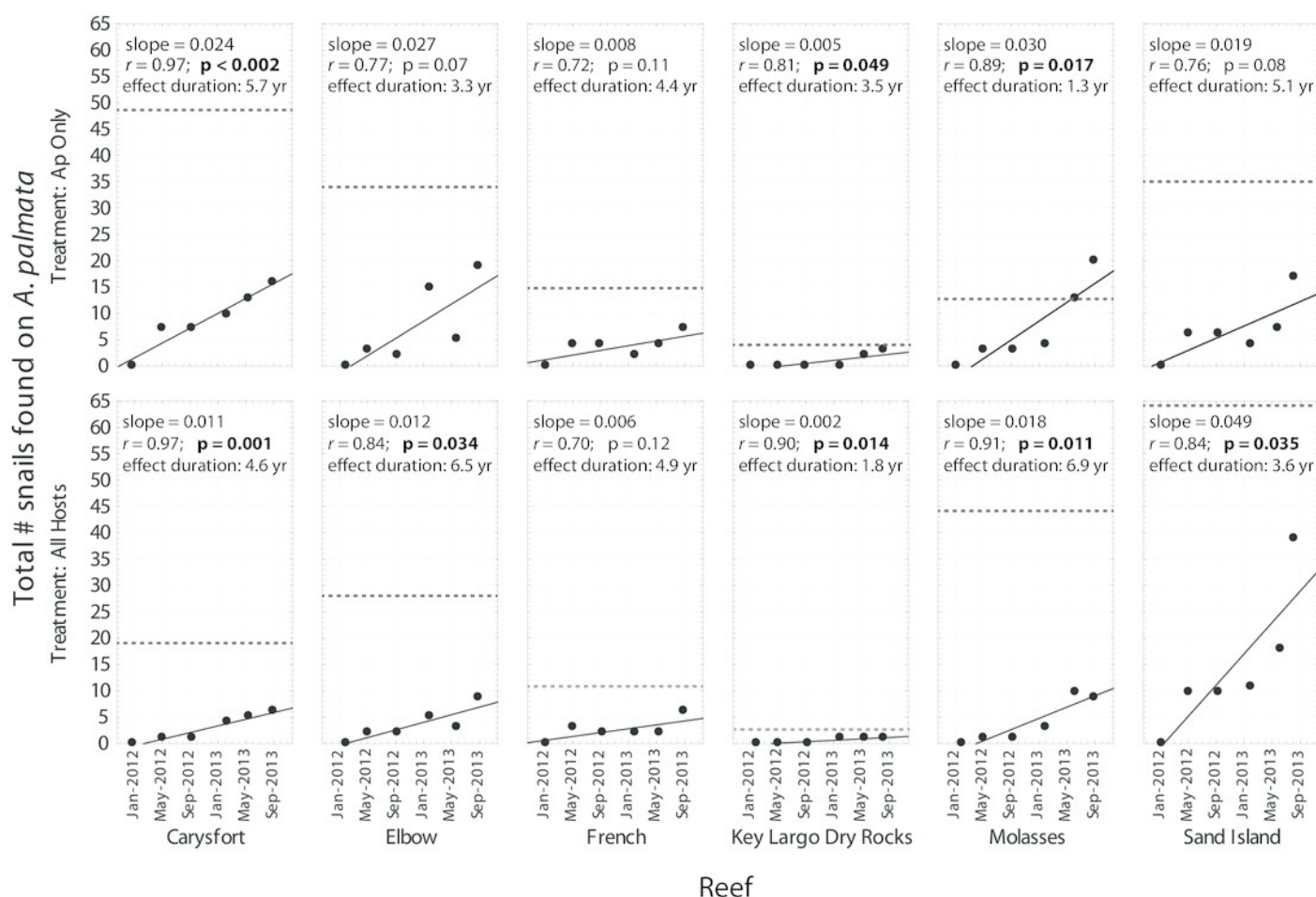




# Figure 3

## *Coralliophila abbreviata* recolonization

Number of *Coralliophila abbreviata* snails found on *Acropora palmata* in each plot where they were removed from *Acropora palmata* only (Ap Only) and from all host coral species following the removal phase of the experiment. The dotted line indicates the baseline number of *C. abbreviata* that were removed from that treatment plot at the start of the experiment in June 2011. 'Effect duration' is the estimated time for the number of snails to reach the baseline (recolonization), according to the regression for each plot.

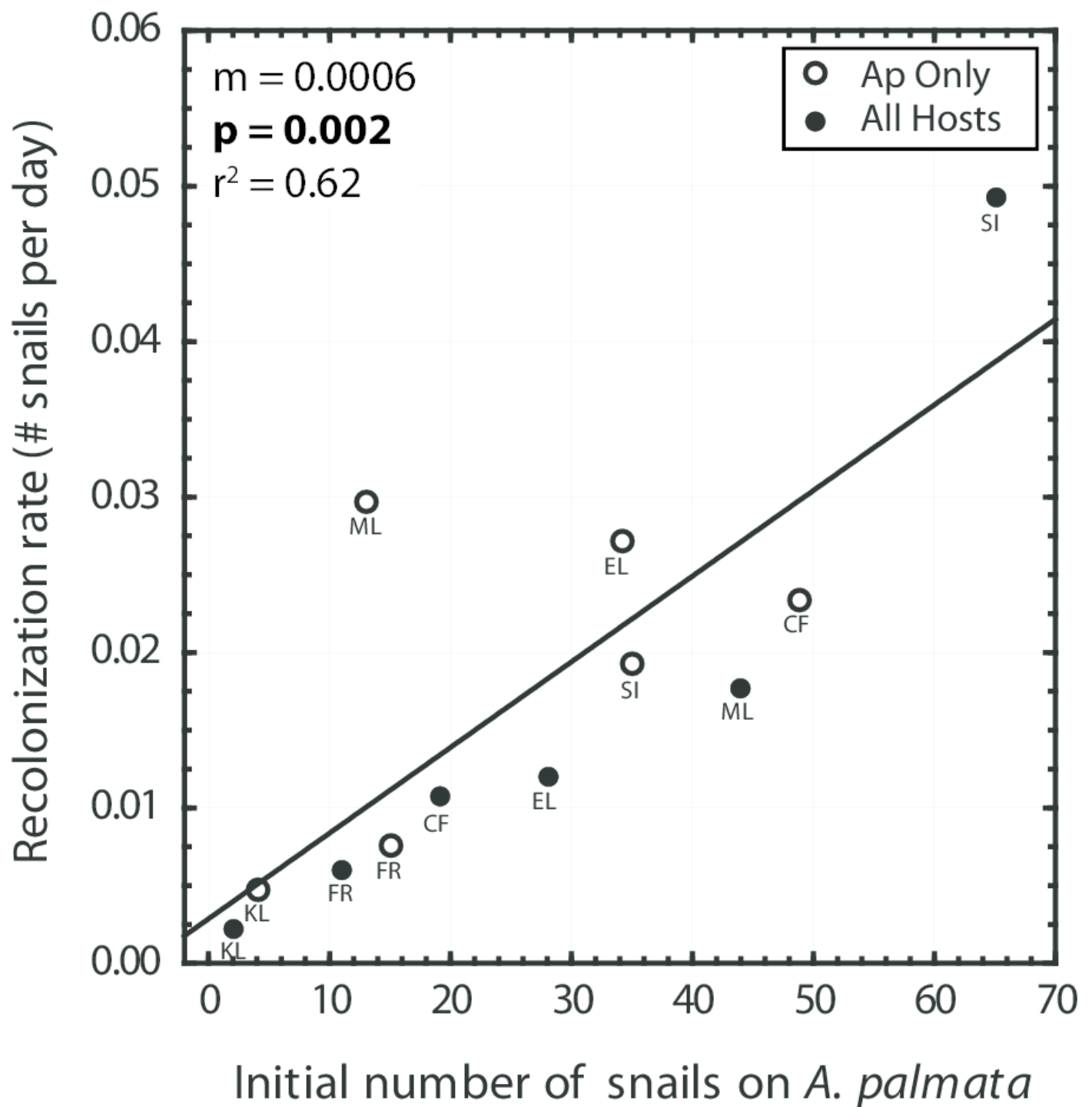




# Figure 4

*Coralliophila abbreviata* recolonization rate

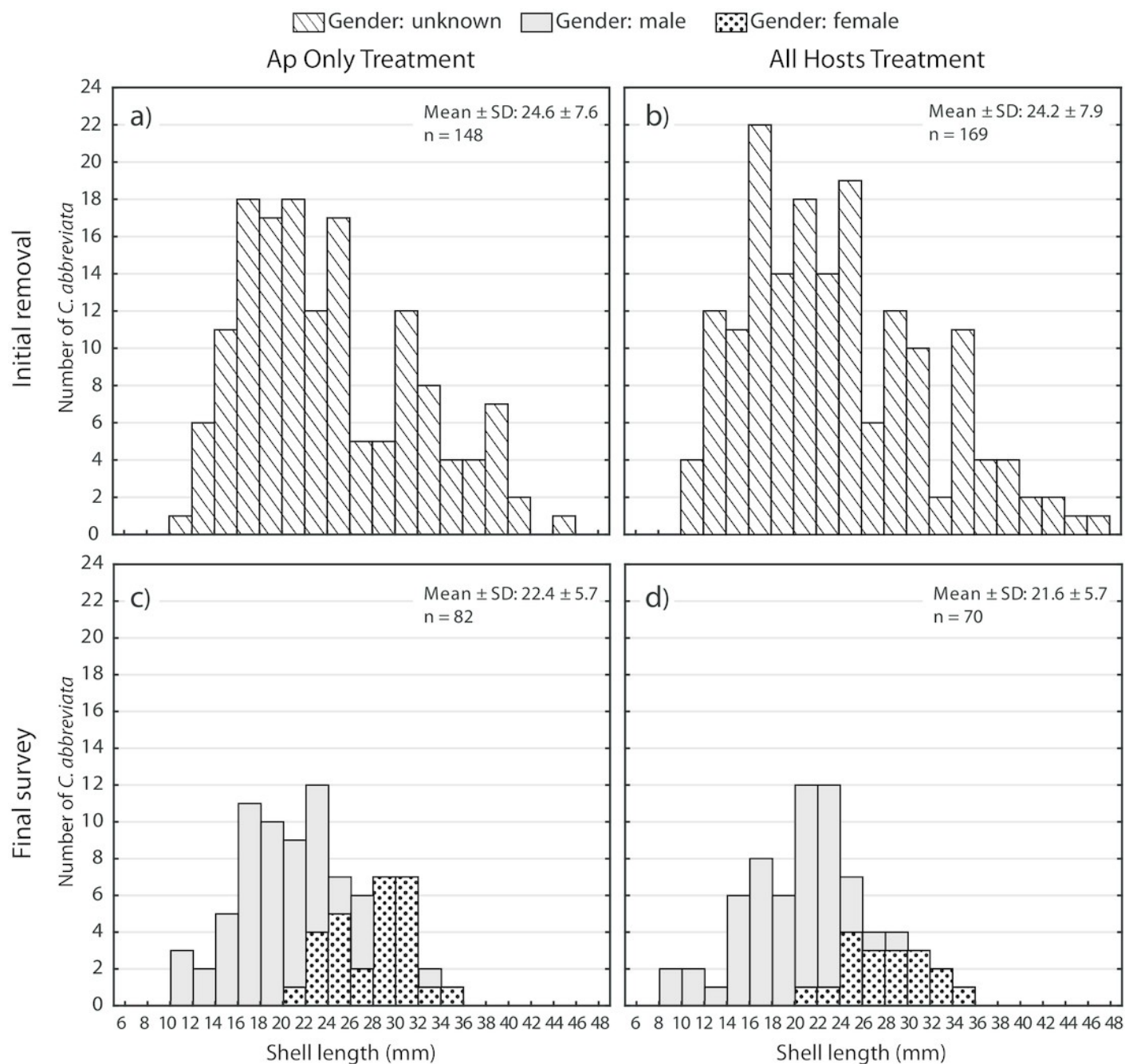
Recolonization rate (number of snails per day) based on the slope of the linear regressions (Fig. 3) versus the number of snails found in each study plot at the initial removal.



# Figure 5

## *Coralliophila abbreviata* size and gender frequency distribution

Size and gender (Sept 2013 only) frequency distribution for the *Coralliophila abbreviata* snails collected from *Acropora palmata* host colonies in the a) Ap Only (snails removed from *A. palmata* only) and b) All Hosts (snails removed from all host coral species) treatments at the initial removal (June 2011) and at the final survey (Sept 2013) for c) Ap Only and d) All Hosts treatments.



# Figure 6

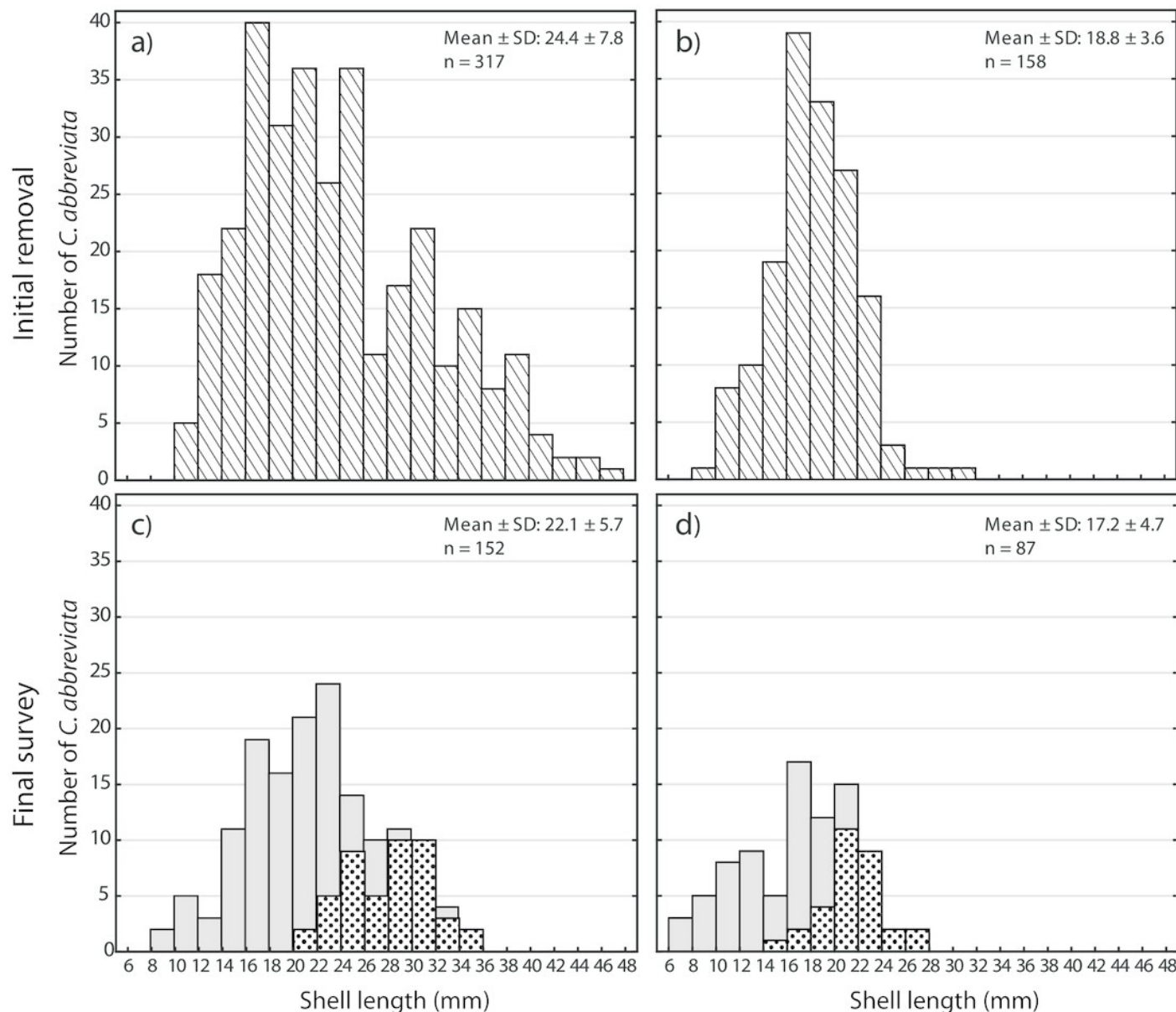
*Coralliophila abbreviata* size and gender frequency distribution

Size and gender (Sept 2013 only) frequency distribution for the *Coralliophila abbreviata* snails collected from a) *Acropora palmata* host colonies and b) other host species at the initial removal (June 2011) and at the final survey (Sept 2013) from c) *A. palmata* and d) and other host coral species.

Gender: unknown Gender: male Gender: female

ApSnails (pooled treatments)

Other host snails (All Hosts treatment only)



## Table 1 (on next page)

Summary of coral host colonies and *Coralliophila abbreviata* snails found in the study plots in the initial and final survey.

Observations at each study plot in three experimental treatments: removal of *Coralliophila abbreviata* snails from *Acropora palmata* only (Ap Only), from all coral host species (All Hosts) and controls in which snails were counted but not removed. Coral colonies and snails counted at the initial survey in June 2011 and the final survey in August 2013. *A. palmata* LAI (live area index) is calculated based on colony measurements described in the text.

Treatment	Reef	<i>A. palmata</i> colonies		<i>A. palmata</i> Snails		<i>A. palmata</i> LAI (m <sup>2</sup> )		Other host colonies		Other host snails	
		Initial	Final	Initial	Final	Initial	Final	Initial	Final	Initial	Final
Ap Only	Carysfort	32	21	49	16	8.4	8.4	0		4	
	Elbow	55	76	34	19	15.2	14.8	1		0	
	French	42	57	15	7	9.7	10.5	19		29	
	Key Largo Dry Rocks	11	11	4	3	1.6	0.7	5		9	
	Molasses	11	15	13	20	2.5	2.3	0		0	
	Sand Island	18	21	35	17	2.8	4.7	1		1	
All Hosts	Carysfort	8	13	19	6	6.7	2.3	3	3	61	34
	Elbow	28	31	28	9	5.0	4.8	2	1	11	3
	French	29	44	11	6	1.4	2.1	6	5	39	21
	Key Largo Dry Rocks	22	15	2	1	0.5	0.2	2	2	19	0
	Molasses	29	40	44	9	3.6	3.9	7	8	19	25
	Sand Island	41	51	65	39	12.7	16.6	1	1	9	4
Control	Carysfort	13	9	16	15	0.9	0.1	6	4	23	4
	Elbow	22	29	16	18	4.2	3.4	4	4	14	6
	French	24	20	8	5	1.9	1.2	15	15	29	31
	Key Largo Dry Rocks	10	9	11	10	3.0	2.5	6	6	10	10
	Molasses	22	31	15	21	8.7	8.7	7	6	7	8
	Sand Island	10	9	4	5	0.8	1.2	6	4	10	8