A peer-reviewed version of this preprint was published in PeerJ on 3 July 2014.

<u>View the peer-reviewed version</u> (peerj.com/articles/432), which is the preferred citable publication unless you specifically need to cite this preprint.

Tickle PG, Paxton H, Rankin JW, Hutchinson JR, Codd JR. 2014. Anatomical and biomechanical traits of broiler chickens across ontogeny. Part I. Anatomy of the musculoskeletal respiratory apparatus and changes in organ size. PeerJ 2:e432 https://doi.org/10.7717/peerj.432

Anatomical and biomechanical traits of broiler chickens across ontogeny. 1. Anatomy of the musculoskeletal respiratory apparatus and changes in organ size. Peter G. Tickle*, Heather Paxton[§], Jeffery W. Rankin[§], John R. Hutchinson[§] and Jonathan R. Codd*1 *Faculty of Life Sciences, University of Manchester, Manchester, UK §Structure & Motion Laboratory, Department of Comparative Biomedical Sciences, The Royal Veterinary College, University of London, Hatfield, Hertfordshire, UK ¹Corresponding author: Jonathan Codd, Faculty of Life Sciences, University of Manchester, Michael Smith Building, Oxford Road, Manchester, M13 9PT, UK Tel: +44 161 275 5474; Fax: +44 161 275 3938 E-mail: jonathan.codd@manchester.ac.uk

3637

53 54

49

50

56

55

5758

59 60

616263

64

65

66 67

68

6970

. .

ABSTRACT

Genetic selection for improved meat yields, digestive efficiency and growth rates have transformed the biology of broiler chickens. Modern birds undergo a 50-fold multiplication in body mass in just six weeks, from hatching to slaughter weight. However, this selection for rapid growth and improvements in broiler productivity is also widely thought to be associated with increased welfare problems as many birds suffer from leg, circulatory and respiratory diseases. To understand growth-related changes in musculoskeletal and organ morphology and respiratory skeletal development over the standard six-week rearing period, we present data from post-hatch cadaveric commercial broiler chickens aged 0, 2, 4 and 6 weeks. The heart, lungs and intestines decreased in size for hatch to slaughter weight when considered as a proportion of body mass. Proportional liver size increased in the two weeks after hatch but decreased between 2 and 6 weeks. Breast muscle mass on the other hand displayed strong positive allometry, increasing in mass faster than the increase in body mass. Contrastingly, less rapid isometric growth was found in the external oblique muscle, a major respiratory muscle that moves the sternum dorsally during expiration. Considered together with the relatively slow ossification of elements of the respiratory skeleton, it seems that rapid growth of the breast muscles might compromise the efficacy of the respiratory apparatus. Furthermore, the relative reduction in size of the major organs indicates that selective breeding in meat-producing birds has unintended consequences that may bias these birds toward compromised welfare and could limit further improvements in meat-production and feed efficiency.

INTRODUCTION

Genetic selection in domesticated broiler chickens has brought about significant improvements in the form of increasing meat yields and growth performance. Growth rates in intensively reared industrial broiler chickens have consistently accelerated such that a 300% increase has been engineered in the past 60 years, from 25g per day in the 1950s to 100g per day in the modern bird (Knowles, et al., 2008). Consequently the optimal slaughter mass of approximately 3kg is reached in six rather than 16 weeks (Govaerts, et al., 2000; Griffin and Goddard, 1994; Havenstein, et al., 1994a). Maximising pectoral (breast) muscle mass is a primary target for selection. Compared to ancestral varieties, pectoral hypertrophy in the broiler chicken has resulted in an approximate doubling in muscle size, making up ~20% of total body mass in the modern bird (Havenstein, et al., 2003b; Schmidt, et al., 2009). Mounting evidence suggests that selection for such economically desirable traits in the modern broiler has been accompanied by reduced welfare (Julian, 1998; Knowles, et al., 2008) and increased mortality (Havenstein, et al., 2003a; Havenstein,

as the multitude of leg pathologies that may affect locomotion in broiler chickens (Bradshaw, et al., 2002; Corr, et al., 2003a; Corr, et al., 2003b; Kestin, et al., 1992; Knowles, et al., 2008; Paxton, et al., 2010), cardiac (Wilson, et al., 1988) and pectoral (Randall, 1982) myopathies, pulmonary hypertension (Wideman, 2001) and ascites (Julian, 1993; Wilson, et al., 1988). The prevalence of these conditions indicates that further improvements in efficiencies and meat production may be constrained by the physiological capabilities of broilers because skeletal, cardiac, respiratory and digestive systems appear to be close to their functional limit.

6

The relationships between body mass and organ, skeleton and muscle size are crucial to our understanding of animal physiology. Physical scaling rules determine the structural and functional consequences of changes in size and therefore exert a profound effect on organismal form (Schmidt-Nielsen, 1984). Understanding the relative growth and size of organs and muscles is important in broilers as it can help to better understand the diseases that they suffer from. For example, the broiler heart and brain become progressively smaller as a proportion of body mass over development, unlike in ancestral breeds (Jackson and Diamond, 1996; Schmidt, et al., 2009). In contrast, selection for faster growth and muscle mass is reflected in the proportionally greater intestine mass and accelerated pectoral growth in broiler chickens (Jackson and Diamond, 1996; Konarzewski, et al., 2000; Schmidt, et al., 2009). These relationships demonstrate how artificial selection in the broiler has resulted in developmental trade-offs; reallocation of resources to maximize nutrient absorption and pectoral muscle mass has coincided with a relative decrease in the size of other organs (Havenstein, et al., 2003b; Havenstein, et al., 1994b; Jackson and Diamond, 1996; Schmidt, et al., 2009). To better understand the effects of intensive selection on broilers, more information is required on how increasing body mass and growth rate has shaped their anatomy and physiology across ontogeny. Therefore, in this paper we present data on how organ and muscle growth varies with increasing body mass in a commercial broiler strain.

Broiler chickens suffer from respiratory problems that may be related to their rapid musculoskeletal development (Julian, 1998) and their potential to outgrow pulmonary capacity (Wideman, 2001). The avian respiratory system can be considered as a two-part mechanism comprising a pump (musculoskeletal) and gas exchanger (lung). The primary ventilatory mechanism in birds consists of dorsal and ventral movements of the ribs and sternum (Claessens, 2009; Zimmer, 1935) that affect air sac volume, thereby facilitating a unidirectional flow of air through the lung (Bretz and Schmidt-Nielsen, 1971; Scheid and Piiper, 1971). Uncinate processes, which are small bones that extend caudodorsally from the vertebral ribs, have been shown to function as levers that assist rib and sternal movements during breathing (Tickle, et al., 2007). Respiratory movements of the skeleton generate pressure changes within the thorax that are

respiratory muscle activity (Codd, et al., 2005). Respiratory muscle activity requires metabolic energy consumption. However, studies in guinea fowl (Numida meleagris) (Markley and Carrier, 2010) have demonstrated that breathing constitutes only 2% of whole-organism metabolism (Markley and Carrier, 2010). In contrast, research on load carrying (Tickle, et al., 2010) and behavioural energetics (Tickle, et al., 2012) indicate a higher cost of breathing in barnacle geese. In particular, carrying extra weight on the sternum, analogous to increased pectoral muscle mass seen in broiler chickens, is energetically expensive compared to an equivalent weight carried on the back (Tickle, et al., 2010). Furthermore, barnacle geese have been shown to achieve metabolic savings by changing posture; when compared to sitting, standing is associated with a 25% higher resting metabolic rate (Tickle, et al., 2012). The higher cost of standing has been in part attributed to the energetic cost of moving the heavy weight of the sternum with each breath, which does not occur while sitting (Tickle, et al., 2012). Understanding movements of the sternum are important when we consider how selection has shaped morphology in domestic birds, because of the selection for enhanced pectoral growth in meat-producing domestic ducks (Gille and Salomon, 1998; Maruyama, et al., 1999), turkeys (Swatland, 1979) and broiler chickens (Govaerts, et al., 2000; Havenstein, et al., 2003b; Havenstein, et al., 1994b; Schmidt, et al., 2009).

Only limited information is available on the developmental biology of the avian respiratory skeleton. In the domestic turkey (*Meleagris gallopavo*) a trend for delayed ossification, especially in the uncinate processes, was identified that potentially constrained respiratory performance due to the decreased rigidity of cartilaginous compared to fully ossified bones; cartilaginous uncinate processes will yield under muscle strain before comparable ossified bone would, making them less effective levers (Tickle and Codd, 2009). Furthermore, the growth trajectory of respiratory muscles in the broiler is unknown. Scaling of respiratory muscle growth in proportion to overall body size and, perhaps most importantly when we consider the dorso-ventral breathing movements of the sternum, the pectoral muscles is a factor in determining how effectively and efficiently breathing functions in broiler chickens. To better understand the respiratory problems apparent in broiler chickens, an analysis of how the skeleton develops is necessary. Here we present a description of developmental changes in the musculoskeletal elements of the broiler chicken respiratory system together with an evaluation of how organ size scales with increasing body mass.

MATERIALS AND METHODS

Specimens

Broiler chickens from a popular commercial strain were obtained from a commercial supplier.

Birds were sampled in a post-hatch growth series between days 1 and 42 (i.e. weeks 0 to 6),

For responding to a 50x range in total body mass (Ma) (Table 1)

Feel Propries 1 supplied 50x range in total body mass (Ma) (Table 1)

Feel Propries 1 supplied 50x range in total body mass (Ma) (Table 1)

Feel Propries 1 supplies 30x range in total body mass (Ma) (Table 1)

Feel Propries 1 supplies 30x range in total body mass (Ma) (Table 1)

Feel Propries 1 supplies 30x range in total body mass (Ma) (Table 1)

Feel Propries 1 supplies 30x range in total body mass (Ma) (Table 1)

Feel Propries 1 supplies 30x range in total body mass (Ma) (Table 1)

Feel Propries 1 supplies 30x range in total body mass (Ma) (Table 1)

Feel Propries 1 supplies 30x range in total body mass (Ma) (Table 1)

Feel Propries 1 supplies 2 supplies 3 suppl

Musculoskeletal growth

 $M_{\rm b}$, pectoralis major (pectoralis), pectoralis minor (supracoracoideus) and external oblique muscle mass ($M_{\rm m}$), wing and ribcage mass, fibre length ($L_{\rm f}$) and pennation angle (θ) were measured using an electronic balance (± 0.001 g), ruler (± 1 mm) and protractor ($\pm 1^{\circ}$). To account for variation within muscle architecture, $L_{\rm f}$ and θ were calculated as the mean of five measurements in each muscle. Physiological cross-sectional area (**PCSA**) was calculated for each muscle (Eq. 1; Sacks and Roy, 1982):

 $PCSA = M_{m} * \cos\theta / \rho * L_{f}$ (1)

Density of muscle tissue (ρ) was assumed to be 1.06g cm⁻³ (Mendez and Keys, 1960; Paxton *et al*, 2010). Linear measurements of the sternum and uncinate processes were recorded using a ruler (± 1 mm). Average length of the uncinate processes was calculated from those occurring on ribs 2–5 since these processes were found in all specimens. Girth was measured around the circumference of the thorax, tucked under the wings. Total mass of the wings and ribcage were also recorded. Oneway ANOVA was used to test for differences in morphology, using mean values of specimens from each developmental stage. Justification for using parametric analysis was based upon the results of a Shapiro-Wilk test, the normal quantile-quantile plot and Levene's tests that assessed assumptions of data normality and equal variances (results displayed in Appendix).

The scaling relationships between musculoskeletal characters and body mass were determined using reduced major axis (**RMA**) linear regression, a method that is suitable since it takes into account variation in both x and y axes (Rayner, 1985; Sokal and Rohlf, 1995). All regression analyses were performed on log10-transformed data to establish allometric equations in the form:

$$\log y = \log a + b \log x \tag{1}$$

where a represents the intercept and the exponent b represents the slope of the line equation. Upper and lower 95% confidence intervals (CI) and the R² value were calculated for each regression line slope. RMA analyses were performed in the PAST statistical program (Hammer, et al., 2001). Assuming geometric similarity (i.e., isometry) over ontogeny, all dimensions are expected to scale in proportion to each other meaning that lengths should scale to $M_b^{0.33}$, areas to $M_b^{0.67}$ and masses to $M_b^{1.00}$. Isometric scaling was assumed where the regression slope $\pm 95\%$ CI did not overlap the expected value.

The ossification pattern of the thoracic skeleton over ontogeny was examined using the histochemical staining protocol of Tickle and Codd (2009). All muscle tissue was removed using dissection, the preparation cleaned by immersion in a 1% potassium hydroxide (KOH) solution and then the skeleton was treated with solutions of alcian blue (uptake corresponds to cartilage) and alizarin red (uptake corresponds to bone). Photographs of stained specimens were taken using a light microscope (Leica MZ9s; Leica Microsystems, Milton Keynes, UK) and subsequently analysed in Leica image analysis software. For comparison of structural properties, relative area of bone and cartilage was calculated for the uncinate process that projects from the fourth vertebral rib in all specimens (Tickle and Codd, 2009).

RESULTS

Organ Development

Heart and lung mass follow a negative allometric growth pattern, decreasing relative to body mass over the six-week growth period (Tables 1 and 2; Fig. 1A). Heart mass decreases from 0.74% to 0.48% of body mass over the growth period, while proportional lung mass reduces by almost half, decreasing from 1.04% to 0.54% (Tables 1 and 2; Fig. 1B). Proportional liver mass significantly increased between 0 and 2 weeks, reached a peak of 3.69% on day 14, then significantly decreased between 2 and 6 weeks when it accounted for 2.43% of M_b (Table 1; Fig. 1C). Taking all data into account indicated that overall liver mass followed an isometric growth pattern; i.e., in direct proportion to increasing M_b (Table 2). Repeating the scaling analysis for birds only aged between 0 and 2 weeks found positive allometric growth, $M_b^{1.10}$ while birds aged between 2 and 6 weeks had negative allometric growth, $M_b^{0.76}$ (Table 2; Fig. 1C). Total intestine mass was found to strongly decline as a proportion of M_b over growth with a negative allometric regression slope of $M_b^{0.75}$ (Tables 1 and 2; Fig. 1D).

Carcass Parts

No significant difference was detected between proportional wing masses with increasing age, indicating a directly proportional relationship with M_b . After accounting for variation due to body mass, girth significantly increased during growth (Table 3), reflecting a relative lateral expansion of the thorax. In contrast, as a proportion of M_b , ribcage mass was significantly lower at day 28 than at days 14 or 42 (Table 3) whereas normalised keel length displayed a trend for increased length, being highest in 42-day birds (Table 3).

Pectoralis major and minor (i.e., M. pectoralis and M. supracoracoideus) $M_{\rm m}$ increased as a proportion of $M_{\rm b}$ over development, showing strong positively allometric growth (Table 4 and 5). The growth of these muscles was defined by two phases: an initial rapid increase in $M_{\rm m}$ between weeks 0 and 2 was followed by a relatively slower increase between weeks 2 and 6 (Table 5; Fig. 2). Sternal keel length and depth developed with positive allometry, increasing above the expected geometric scaling exponent ($M_{\rm b}^{0.33}$) (Table 6), while mean uncinate process length scaled to $M_{\rm b}^{0.30}$, indicating reduced length with increasing body mass. Growth of external oblique muscle was found to increase in direct proportion to increasing body mass (Table 6).

Skeletal Development

Ossification of the uncinate processes commences at around the time of hatch; 15-day embryos have entirely chondrified processes whereas one-day old chicks show 40% ossification (Table 7; Fig. 3). Uncinate process bone synthesis proceeds from the midpoint in proximal and distal directions and overall bone area plateaus at ~77% of total uncinate process area at 40 days old. The remaining 23% remains cartilaginous, shared between tip and base (Table 7). At hatch the sternum exhibits ossification in the most proximal portion in addition to centres of ossification in the caudolateral processes. While bone growth extends distally along the sternum, at slaughter age ossification of the sternal keel remains incomplete (Fig. 3)

DISCUSSION

Organ Development

Our observations confirm a decrease in heart and lung mass relative to body mass over development (Tables 2 and 3). These findings mirror the reduction in relative heart (Govaerts, et al., 2000; Havenstein, et al., 2003b; Havenstein, et al., 1994b; Schmidt, et al., 2009; Thaxton, 2002) and lung mass (Govaerts, et al., 2000; Havenstein, et al., 2003b; Havenstein, et al., 1994b) seen over ontogeny in Ross-type broiler breeds. While absolute mass of the heart is higher than found in unselected lines, proportional mass is lower at hatching and the difference progressively increases over development (Schmidt, et al., 2009). Similarly, lung mass progressively declines as a proportion of body mass over development in our broiler strain, mirroring the negative allometric lung growth in Ross broilers ($M_b^{0.84}$ compared to $M_b^{0.86}$ in this study) (Govaerts, et al., 2000). Considering our results together with previous reports indicates that reduced circulatory and respiratory capacity is an unintended consequence of genetic selection for rapid growth and high

considered to be a contributing factor in the increased mortality and disease in modern broilers (Havenstein, et al., 2003b; Wideman, 2001). Further developments in genetic selection for increasing growth and improving breast yields may be constrained by limited respiratory and circulatory functional capacity. The increased incidence of physiological problems and mortality (Havenstein, et al., 2003a; Havenstein, et al., 1994a) in modern broilers indicates that these systems are already working at maximal levels to supply the physiological demands of growth.

A complex pattern of liver growth was revealed, with organ mass growing proportionally faster than overall body mass growth in the two weeks after hatch, followed by a proportional decrease in mass between two weeks and six weeks post-hatch. Liver development in Ross-type broilers follows a similar pattern of proportionally decreasing after a period of rapid growth soon after hatch (Govaerts, et al., 2000; Schmidt, et al., 2009). Furthermore, Ross broiler peak liver mass as a proportion of M_b was reached on day 7 (3.80%) (Schmidt, et al., 2009) and day 8 (3.38%) (Govaerts, et al., 2000), although the findings presented here do not include birds sampled at these stages. Nevertheless, liver mass at day 14 is similar in our study's and Ross birds (3.15% (Govaerts, et al., 2000); or approximately 3.5% (Schmidt, et al., 2009) M_b). The relatively fast pace of growth in the first two weeks, followed by proportional decline, indicates that the liver matures rapidly, ready for the transition from fat-rich yolk stores to a predominantly carbohydrate diet (Schmidt, et al., 2009). Genetic selection for digestive efficiency in broilers may directly influence the developmental profile of the liver due to the important function it plays in carbohydrate and fat metabolism.

Total intestine mass declined relative to M_b during growth, which is similar to previous reports of proportional decline in intestine mass over development in Ross-type broilers (Iji, et al., 2001; Schmidt, et al., 2009). Absolute and proportional values of the intestine are higher in this study, potentially in part due to a larger intestine in this commercial broiler strain and also in part due to methodological differences; here we report intestine mass and contents rather than the empty intestine mass (Iji, et al., 2001). Exhaustion of the yolk sac contents presumably accounts for the large (6.9%) decline in proportional intestine mass during the first 14 days of post hatch development (Viera and Moran, 1999). After this period, relative intestine mass decreases by approximately 2-2.5% every 14 days (Table 2), which is similar to previous reports (calculations using the data provided in table 2 of Iji *et al.* (2001) and figure 5 of Schmidt *et al* (2009) indicate an approximate decrease of 2.4%). Schmidt *et al* (2009) reported complicated post-hatch intestinal growth, with positive allometric growth in the first seven days followed by a sustained period of negative allometric growth, possibly reflecting the early maturation of the digestive system (Govaerts, et al., 2000). To balance this relative mass loss and maintain high feed efficiency,

morphometric changes occur during this period to increase surface area for absorption of nutrients from ingested food (Schmidt, et al., 2009).

Carcass Parts

While proportional wing mass did not significantly change over growth the overall average yield of 7.0% is below that reported in Ross broilers (8.6%: Havenstein, et al, 1994b; 7.9%: Havenstein, et al, 2003b). The absence of significant change in wing mass is consistent with the absence of flight as a locomotor mode and the reliance on leg muscles for terrestrial locomotion, and occurs despite steep increases in pectoral muscle mass. Girth in 42-day-old broilers was very similar to a recent report for pureline and broiler birds (Paxton, et al., 2013). Presumably the rapid increase in pectoral muscle mass is a factor in the reduced proportional ribcage mass over development. Normalised sternal keel length was found to increase over growth, indicating that the area available for pectoral muscle attachment is greater as the birds reach slaughter weight, consistent with the rapid proportional increase in pectoral mass.

Thoracic Anatomy

The relationship between increasing pectoral mass and body mass was positively allometric, indicating that the breast muscles became proportionally much larger over the six-week growth period. Allometric relationships were stronger than those reported for Ross broilers (Govaerts, et al., 2000; Schmidt, et al., 2009), suggesting that the birds in this study were able to lay down breast muscle more rapidly during development. Overall breast muscle size as a proportion of M_b was similar to the values reported for Ross (Govaerts, et al., 2000; Havenstein, et al., 2003b; Schmidt, et al., 2009) and similar commercial strain (Paxton, et al., 2013) broilers.

Allometric scaling was found for sternal keel dimensions; this proportional increase in the size of the sternum over development presumably enables fast growth of the breast muscles. In contrast, average uncinate process length was found to decrease over growth, scaling with a slight negative allometry. Uncinate processes act as levers for movement of the ribs and sternum during breathing, and overall length is an important factor that determines the magnitude of this leverage (Tickle, et al., 2007). Therefore, the relative shortening of uncinate processes might correspond to a decrease in ventilatory capacity during growth.

Considering that sternal mass (i.e. breast muscle, (Table 6)) increases with positive allometry during growth it was expected that the external oblique muscle would undergo similar growth to maintain functional efficacy. Our data indicate however that the external oblique muscle becomes progressively smaller in comparison to the breast muscles as the birds age, and may be less, able to maintain adequate expiration. Further evidence is provided by the isometric growth of the property of the

 $L_{\rm f}$ and PCSA, since maximum force-generating capacity increases in direct proportion to muscle area (Lieber and Friden, 2000). No comparable data on respiratory muscle development are known, so it is difficult to establish whether this pattern of growth is unique to the broiler or more widespread among ancestral fowl breeds and other species. Coupled with the decreased proportional heart and lung masses, however, selection for meat-producing traits appears to have the undesirable effect of compromising the engine of aerobic metabolism; i.e., the respiratory system. Clearly more research into the respiratory anatomy in birds is required to understand the potential trade-offs between breast growth and breathing performance.

324325

326

327

328

329

330

331332333334

335

336

337

338

339

340

341

342

343

344

345

346

347

348

349

316

317

318

319

320

321

322

323

Skeletal Development

The broiler skeleton ossifies following a pattern similar to other domestic poultry (Hogg, 1980; Maxwell, 2008; Tickle and Codd, 2009). Ossification of skeletal characters, however, is seen to develop at different time points. For example, the long bones of the leg and wing are almost entirely ossified at hatch, while the sternum, tip of the scapula and proximal and distal portions of the ribs ossify after hatch (Fig. 3). Ossification of uncinate processes begins around the time of hatch, in agreement with an earlier report in the chicken (Hogg, 1980), when bone replacement of cartilage becomes apparent in the midpoint of the shaft (Table 7; Fig. 3). Rapid synthesis of bone tissue must occur in the days immediately prior to hatch, considering that the processes are entirely chondrified six days before hatch (Table 7). Onset of ossification therefore coincides with the initial in ovo respiratory skeleton movements and transition from chorioallontoic gas exchange to pulmonary ventilation (Menna and Mortola, 2002); perhaps this mechanical stimulus prompts remodeling of uncinate process structure toward ossified tissue. Uncinate process growth in the broiler is similar to development in the domestic turkey (Tickle and Codd, 2009) in that ossification remains incomplete, with a plateau in bone growth at around 80% of total process area. An interesting difference between domestic turkey and chicken is the lack of a bony symphysis between the base of the process and adjoining rib in the broiler (Tickle and Codd, 2009). A cartilaginous base may have significant implications for the functional efficacy of the uncinate process because they act as levers for rib movement, against which muscles contract (Tickle, et al., 2007). The cartilaginous portion of uncinate processes has a lower elastic modulus, or is less stiff, than the ossified area. If it is less rigid, this might compromise the lever function of the uncinate processes (Tickle and Codd, 2009). It is possible that an ossified connection between uncinate process and rib would form later in development, as is seen in the domestic turkey between days 64 and 94 post-hatch (Tickle and Codd, 2009), suggesting that immaturity of the skeleton even at slaughter weight may have a functional impact on breathing in the broiler.

Ossification of the sternum is incomplete over the six-week period, with specifically the distal portion of the keel remaining chondrified. The cartilaginous distal portion of the sternum is common to meat-producing and laying strains (Breugelmans, et al., 2007) and presumably reflects the reduced or negligible importance of flight as a means of locomotion in domestic fowl; without need for a sturdy anchor to accommodate and dissipate the strong contractions of pectoral muscles during flapping flight, it seems the energetic resources are directed to growing muscle tissue rather than synthesis of bone for the sternum.

The rapid development of body mass in broilers is not mirrored by accelerated ossification of the respiratory skeleton. Instead the skeleton appears to develop in a pattern similar to other poultry, indicating that this process is tightly conserved within galliform birds. Given the potential compromise in respiratory performance due to cartilaginous skeletal elements coupled with a large sternal weight, selection for faster and increased ossification may provide a benefit to broiler health.

Here we have considered the how the growth trajectories of major organs and respiratory elements are affected by increasing body mass in broilers. In contrast to the strong pectoral growth, major organs become proportionally smaller with age and development of the respiratory pump is delayed. Our findings indicate that selection for rapid growth and large breast muscles could have the unintended consequence of negatively affecting broiler physiology, possibly compromising the function capacity of cardiovascular and respiratory systems.

ACKNOWLEDGMENTS

We are thankful to Cobb-Vantress Inc. for providing broiler chicken specimens. This work was supported by BBSRC grant BB/I021116/1 to JRC and JRH.

373 REFERENCES

Bradshaw, R. H., R. D. Kirkden, and D. M. Broom. 2002. A review of the aetiology and pathology of leg weakness in broilers in relation to welfare. Avian Poult Biol Rev 13:45-103. doi

- 377 10.3184/147020602783698421
- 378 Bretz, W. L., and K. Schmidt-Nielsen. 1971. Bird respiration: flow patterns in the duck lung. J.
- 379 Exp. Biol. 54:103-118.

- 380 Breugelmans, S., S. Muylle, P. Cornillie, J. Saunders, and P. Simoens. 2007. Age determination of
- poultry: a challenge for customs. Vlaams Diergen Tijds 76:423-430.
- 382 Claessens, L. P. a. M. 2009. The skeletal kinematics of lung ventilation in three basal bird taxa
- 383 (emu, tinamou, and guinea fowl). J Exp Zool A 311A:586-599. doi 10.1002/Jez.501

- Codd, J. R., D. F. Boggs, S. F. Perry, and D. R. Carrier. 2005. Activity of three muscles associated
- with the uncinate processes of the giant Canada goose Branta canadensis maximus. J Exp Biol
- 386 208:849-857. doi 10.1242/jeb.01489
- Corr, S. A., M. J. Gentle, C. C. Mccorquodale, and D. Bennett. 2003a. The effect of morphology on
- the musculoskeletal system of the modern broiler. Anim Welfare 12:145-157.
- Corr, S. A., M. J. Gentle, C. C. Mccorquodale, and D. Bennett. 2003b. The effect of morphology on
- walking ability in the modern broiler: A gait analysis study. Anim Welfare 12:159-171.
- 391 Gille, U., and F. V. Salomon. 1998. Muscle growth in wild and domestic ducks. Brit Poultry Sci
- 392 39:500-505.
- Govaerts, T., G. Room, J. Buyse, M. Lippens, G. De Groote, and E. Decuypere. 2000. Early and
- 394 temporary quantitative food restriction of broiler chickens. 2. Effects on allometric growth and
- growth hormone secretion. Brit Poultry Sci 41:355-362. doi 10.1080/713654923
- 396 Griffin, H. D., and C. Goddard. 1994. Rapidly growing broiler (meat-type) chickens: their origin
- and use for comparative studies of the regulation of growth. Int J Biochem 26:19-28.
- Hammer, Ø., D. a. T. Harper, and P. D. Ryan. 2001. PAST: Paleontological statistics software
- package for education and data analysis. Palaeontologica Electronica 4:9pp.
- Havenstein, G. B., P. R. Ferket, and M. A. Qureshi. 2003a. Growth, livability, and feed conversion
- of 1957 versus 2001 broilers when fed representative 1957 and 2001 broiler diets. Poultry Science
- 402 82:1500-1508.
- Havenstein, G. B., P. R. Ferket, and M. A. Qureshi. 2003b. Carcass composition and yield of 1957
- versus 2001 broilers when fed representative 1957 and 2001 broiler diets. Poultry Science 82:1509-
- 405 1518.
- Havenstein, G. B., P. R. Ferket, S. E. Scheideler, and B. T. Larson. 1994a. Growth, livability, and
- feed conversion of 1957 vs 1991 broilers when fed "typical" 1957 and 1991 broiler diets. Poultry
- 408 Science 73:1785-1794.
- Havenstein, G. B., P. R. Ferket, S. E. Scheideler, and D. V. Rives. 1994b. Carcass composition and
- 410 yield of 1991 vs 1957 broilers when fed "typical" 1957 and 1991 broiler diets. Poultry Science
- 411 73:1795-1804.
- Hogg, D. A. 1980. A re-investigation of the centres of ossification in the avian skeleton at and after
- 413 hatching. J Anat 130:725-743.
- 414 Iji, P. A., A. Saki, and D. R. Tivey. 2001. Body and intestinal growth of broiler chicks on a
- commercial starter diet. 1. Intestinal weight and mucosal development. Brit Poultry Sci 42:505-513.
- 416 doi 10.1080/00071660120073151
- Jackson, S., and J. Diamond. 1996. Metabolic and digestive responses to artificial selection in
- 418 chickens Evolution 50:1638-1650, doi:10.2307/2410900 reer/ PrePrints http://dx.doi.org/10.7287/peerl.preprints.342V1 CC-BY 4.0 Open Access | received: 31 Mar 2014, published: 31 Mar 2014

- 419 Julian, R. J. 1993. Ascites in poultry. Avian Pathology 22:419-454. doi
- 420 10.1080/03079459308418934
- Julian, R. J. 1998. Rapid growth problems: ascites and skeletal deformities in broilers. Poultry
- 422 Science 77:1773-1780.
- Kestin, S. C., T. G. Knowles, A. E. Tinch, and N. G. Gregory. 1992. Prevalence of leg weakness in
- broiler chickens and its relationship with genotype. Vet Rec 131:190-194.
- Knowles, T. G., S. C. Kestin, S. M. Haslam, S. N. Brown, L. E. Green, A. Butterworth, S. J. Pope,
- D. Pfeiffer, and C. J. Nicol. 2008. Leg disorders in broiler chickens: prevalence, risk factors and
- 427 prevention. PLoS ONE 3:e1545. doi 10.1371/journal.pone.0001545
- 428 Konarzewski, M., A. Gavin, R. Mcdevitt, and I. R. Wallis. 2000. Metabolic and organ mass
- responses to selection for high growth rates in the domestic chicken (Gallus domesticus). Physiol
- 430 Biochem Zool 73:237-248. doi 10.1086/316729
- 431 Lieber, R. L., and J. Friden. 2000. Functional and clinical significance of skeletal muscle
- architecture. Muscle Nerve 23:1647-1666.
- Markley, J. S., and D. R. Carrier. 2010. The cost of ventilation in birds measured via unidirectional
- 434 artificial ventilation. Comp Biochem Physiol A 155:146-153. doi 10.1016/j.cbpa.2009.10.023
 - Maruyama, K., M. K. Akbar, and C. M. Turk. 1999. Growth pattern and carcase development in
- male ducks selected for growth rate. Brit Poultry Sci 40:233-239.
- Maxwell, E. E. 2008. Comparative embryonic development of the skeleton of the domestic turkey
- 438 (Meleagris gallopavo) and other galliform birds. Zoology 111:242.
- 439 Menna, T. M., and J. P. Mortola. 2002. Metabolic control of pulmonary ventilation in the
- developing chick embryo. Respiratory Physiology & Neurobiology 130:43.
- Paxton, H., N. B. Anthony, S. A. Corr, and J. R. Hutchinson. 2010. The effects of selective
- breeding on the architectural properties of the pelvic limb in broiler chickens: a comparative study
- 443 across modern and ancestral populations. J Anat 217:153-166. doi 10.1111/j.1469-
- 444 7580.2010.01251.x

- Paxton, H., M. Daley, S. Corr, and J. Hutchinson. 2013. The gait dynamics of the modern broiler
- 446 chicken: A cautionary tale of selective breeding. J Exp Biol 216:3237-3248. doi
- 447 10.1242/jeb.080309
- Randall, C. J. 1982. Acute pectoral myopathy in broiler breeders. Avian Pathology 11:245-252. doi
- 449 10.1080/03079458208436098
- Rayner, J. M. V. 1985. Linear relations in biomechanics: the statistics of scaling functions. J. Zool.,
- 451 Lond. (A) 206:415-439.

- Scheid, P., and J. Piiper. 1971. Direct measurement of the pathway of respired gas in duck lungs.
- 453 Respiration Physiology 11:308-314.
- Schmidt, C. J., M. E. Persia, E. Feierstein, B. Kingham, and W. W. Saylor. 2009. Comparison of a
- 455 modern broiler line and a heritage line unselected since the 1950s. Poultry Science 88:2610-2619.
- 456 doi Doi 10.3382/Ps.2009-00055
- 457 Schmidt-Nielsen, K. 1984. Scaling: why is animal size so important? CUP.
- Sokal, R. R., and F. J. Rohlf. 1995. Biometry: the principles and practice of statistics in biological
- research. 3rd ed. ed. Freeman, New York; Oxford.
- Swatland, H. J. 1979. Development of shape in turkey carcasses. J Agr Sci 93:1-6.
- 461 Thaxton, J. P. 2002. Heart growth in broilers. Brit Poultry Sci 43:24-27. doi 462 10.1080/00071660120109827
- Tickle, P. G., and J. R. Codd. 2009. Ontogenetic development of the uncinate processes in the
- domestic turkey (Meleagris gallopavo). Poultry Science 88:179-184. doi Doi 10.3382/Ps.2008-
- 465 00349
- Tickle, P. G., A. R. Ennos, L. E. Lennox, S. F. Perry, and J. R. Codd. 2007. Functional significance
- 467 of the uncinate processes in birds. J Exp Biol 210:3955-3961. doi 10.1242/jeb.008953
- Tickle, P. G., R. L. Nudds, and J. R. Codd. 2012. Barnacle geese achieve significant energetic
- savings by changing posture. PLoS ONE 7:e46950. doi 10.1371/journal.pone.0046950
- 470 Tickle, P. G., M. F. Richardson, and J. R. Codd. 2010. Load carrying during locomotion in the
- barnacle goose (*Branta leucopsis*): The effect of load placement and size. Comp Biochem Phys A
- 472 156:309-317. doi Doi 10.1016/J.Cbpa.2010.01.022
- Viera, S. L., and E. T. Moran, Jr. 1999. Effects of egg of origin and chick post-hatch nutrition on
- broiler live performance and meat yields. World's Poult. Sci. J. 55:126-142.
- Wideman, R. F. 2001. Pathophysiology of heart/lung disorders: pulmonary hypertension syndrome
- 476 in broiler chickens. World Poultry Sci J 57:289-307. doi Doi 10.1079/Wps20010021
- Wilson, J. B., R. J. Julian, and I. K. Barker. 1988. Lesions of right heart failure and ascites in broiler
- 478 chickens. Avian Dis 32:246-261.
- Zimmer, K. 1935. Beitrage zur Mechanik der Atmung bei den Vögeln in Stand und Flug. Aufgrund
- anatomischer-physiologisher und experimenteller Studien. Zoologica 88:1-69.
- 481 482

Figure 1

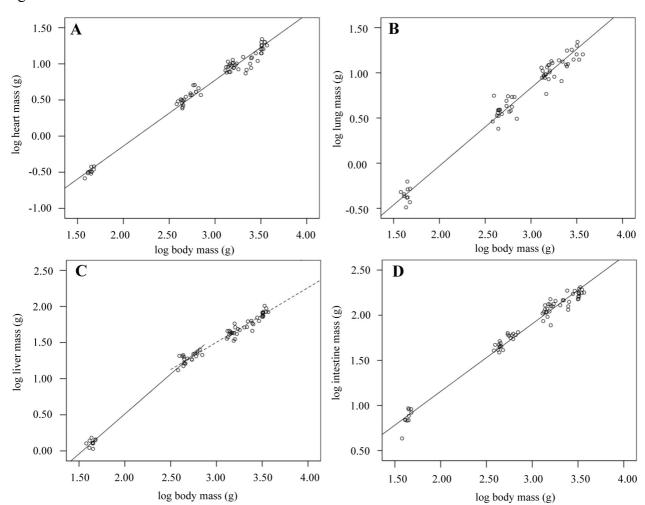


Figure 2

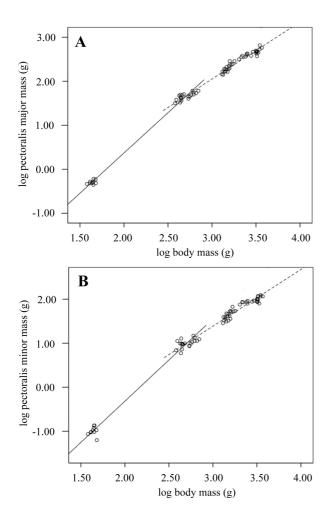


Figure 3

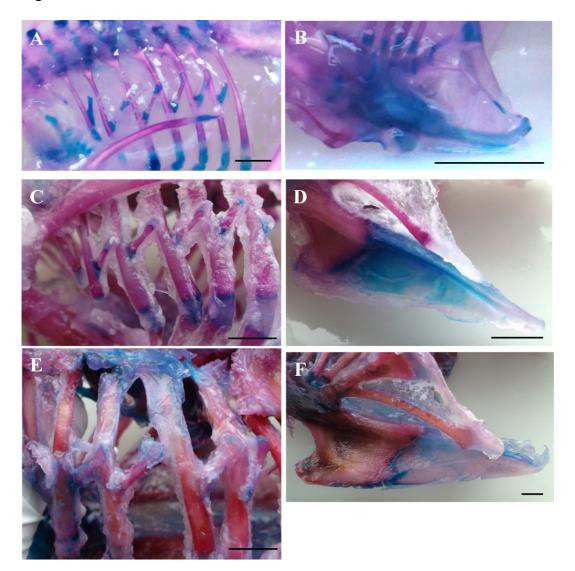


Table 1
Internal organ mass as a proportion of total body mass (M_b) over development. Data are mean \pm standard deviation. Asterisk (*) denotes significant differences at the 0.05 level.

Age (days)	n	$M_{\rm b}({ m kg})$	Heart (% M _b)	Lung (% M _b)	Liver (% $M_{\rm b}$)	Intestine (% M _b)
1	10	$0.04 \pm 0.003*$	0.74 ± 0.05	$1.04 \pm 0.20*$	2.99 ± 0.34	$17.1 \pm 2.70*$
14	20	$0.51 \pm 0.09*$	0.69 ± 0.09	0.77 ± 0.13	3.69 ± 0.36 *	$10.2 \pm 1.00*$
28	19	1.53 ± 0.18*	$0.62 \pm 0.08*$	0.68 ± 0.22	2.87 ± 0.39	$7.70 \pm 1.10*$
42	19	$2.97 \pm 0.46*$	$0.48 \pm 0.09*$	0.54 ± 0.11 *	2.43 ± 0.26 *	5.6 ± 0.84 *

Table 2

Scaling relationship between organ and body mass. The regression slope indicates the proportional change of organ mass with increasing body mass, and 95% confidence intervals are shown (95% CL). Isometric (=) and negative allometric (-) growth are indicated by symbols.

	N	Slope	Lower 95% CL	Upper 95% CL	R^2
Heart	69	0.91 (-)	0.89	0.94	0.98
Lungs	62	0.86 (-)	0.83	0.90	0.97
Liver (wks. 0 - 6)	69	0.95 (=)	0.92	1.00	0.98
Liver (wks. 0 - 2)	30	1.10 (+)	1.06	1.15	0.99
Liver (wks. 2 - 6)	59	0.76 (-)	0.72	0.80	0.95
Intestine	69	0.75 (-)	0.72	0.78	0.98

Table 3

Morphological examination of the external body. Data represented are mean \pm standard deviation. Following the principles of geometric similarity

(Alexander et al., 1981), girth and keel length are normalised by body mass^{1/3} to negate the effect of body size on our data. Asterisk (*) denotes significant differences at the 0.05 level.

Age (days)	n	Wings ($\% M_{\rm b}$)	Girth	Keel length	Rib Cage (% M _b)
14	10	7.5 ± 0.6	0.21 ± 0.01 *	0.08 ± 0.004 *	10.0 ± 1.1
28	9	7.8 ± 0.5	0.23 ± 0.01 *	0.07 ± 0.004 *	$7.49 \pm 0.9*$
42	12	5.8 ± 0.4	0.25 ± 0.01 *	$0.10 \pm 0.005 *$	8.9 ± 0.7



Table 4

Breast muscle mass (% body mass) over development. Data are mean \pm standard deviation.

Age (days)	Pectoralis major (% M _b)	Pectoralis minor (% $M_{\rm b}$)			
1	0.58 ± 0.06	0.12 ± 0.02			
14	8.65 ± 0.98	1.88 ± 0.27			
28	12.1 ± 1.12	2.51 ± 0.31			
42	14.5 ± 1.70	3.10 ± 0.37			

Table 5 Positive allometric growth of breast $M_{\rm m}$.

	Age (weeks)	N	Slope	Lower 95% CL	Upper 95% CL	\mathbb{R}^2
pectoralis major						
(M. pectoralis)	0 – 6	69	1.60	1.56	1.66	0.98
	0 - 2	30	1.83	1.77	1.88	0.99
	2 - 6	59	1.29	1.24	1.34	0.98
pectoralis minor (M. supracoracoideus)	0 - 6	68	1.62	1.57	1.70	0.98
	$0 - 2_{(1)}$	30	1.87	1.79	1.95	0.98
	2 - 6	58	1.29	1.22	1.36	0.96



Scaling relationships of thoracic musculoskeletal parameters and body mass. The regression slope indicates proportional change with increasing body mass. Isometric (=), positive (+) and negative (-) allometric growth are indicated by symbols.

		n	Slope	Lower 95% CL	Upper 95% CL	\mathbb{R}^2
pectoralis major	$M_{ m m}$	69	1.680 (+)	1.56	1.66	0.98
	$L_{ m f}$	37	0.46 (+)	0.42	0.50	0.95
	PCSA	37	1.23 (+)	1.19	1.27	0.99
pectoralis minor	$M_{ m m}$	68	1.62 (+)	1.57	1.70	0.98
	$L_{ m f}$	34	0.55 (+)	0.50	0.62	0.89
	PCSA	29	1.17 (+)	1.09	1.25	0.97
external oblique	$M_{ m m}$	25	0.97 (=)	0.84	1.09	0.89
	$L_{ m f}$	15	0.31 (=)	0.12	0.48	0.32
	PCSA	15	0.90 (=)	0.72	1.09	0.86
sternal keel	length	34	0.48 (+)	0.44	0.51	0.97
	depth	34	0.55 (+)	0.51	0.59	0.95
uncinate process	length	31	0.30 (-)	0.28	0.32	0.94

Table 7
Structural changes in uncinate process 4 over development from embryo (6 days before hatch) to slaughter age; presence of cartilage and bone as derived from stained specimens. Data presented as mean \pm standard deviation (SD).

	Bone		Cartilage at base		Cartilage at tip	
Age (days)	% total area SE		% total area	SD	% total area	SD
-6	0	0	100	0	100	0
1	39.52	16.76	35.02	9.40	25.46	9.64
13	72.59	8.36	23.51	8.14	3.90	0.22
29	72.55	7.37	13.44	7.68	14.00	0.31
40	76.76	8.61	9.08	5.14	14.16	2.45