

Advancing ecological restoration through experimental design on spatial and temporal scales relevant to wildlife

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1 **ABSTRACT**

2 Experimenting with new and unconventional restoration methods and designs is critical to
3 advancing the field of ecological restoration. Conventional methods cannot be considered
4 reliable in a future with climate change-induced shifts in weather conditions, species
5 distributions, and ecosystem processes. It is crucial that researchers and practitioners collaborate
6 to identify the most effective restoration methods, yet there remains a disturbing lack of
7 restoration experiments at the spatial or temporal scales relevant for evaluating wildlife
8 responses. We suspect that willingness to attempt such experiments is hampered by the perceived
9 difficulty of conducting these experiments combined with a fear of failure. However, we argue
10 that failure to experiment with new methods guarantees learning nothing new. Here, we address
11 many of the major challenges of designing an experiment to evaluate wildlife responses to
12 restoration, including (1) distinguishing between the goals and objectives of the restoration
13 project and the key uncertainties the experiment will address, (2) designing the experiment itself,
14 including optimizing plot size and replication, and (3) determining how and when the results will
15 be evaluated. We then illustrate how we designed an experiment to evaluate riparian bird
16 responses to restoration along the lower Cosumnes River in the Central Valley of California,
17 USA. Researchers and practitioners working together from the start of the objectives-setting
18 /process, through experimental design, implementation, and evaluation can proactively address
19 the challenges of conducting a restoration experiment and maximize the chances of successfully
20 identifying effective restoration methods, adding to the practitioners' toolbox, and accelerating
21 the rate of successful habitat restoration.

22

23 **Keywords:** California, Central Valley, Cosumnes River, restoration experiment, riparian birds

24 **Introduction**

25 Experimentation with new and unconventional restoration methods and designs is critical to
26 making advances in the field of ecological restoration (Seavy et al. 2009, Goreau and Trench
27 2012). Climate change-induced shifts in weather conditions, species distributions, and ecosystem
28 processes signal a future in which conventional methods cannot be considered reliable (Hobbs et
29 al. 2009, Perry et al. 2015). With limited time and resources, researchers and practitioners must
30 collaborate to identify the methods and designs that will be the most effective and resilient to
31 climate change. Yet, advances have been severely limited by the general lack of scientific
32 experimentation, except on relatively small scales and over short time frames (e.g. < 1m² over <
33 1 growing season; Kettenring and Adams 2011). Larger-scale and longer-term evaluations of
34 restoration success, let alone experiments, are far more rare (Osenberg et al. 2006, Dickens and
35 Suding 2013), even though they are necessary for evaluating the responses of wildlife, the
36 restoration of ecological functions and services, and the performance of restorations under
37 extreme weather conditions.

38 In the case of wildlife responses to restoration, many restoration practitioners have
39 focused on restoring habitat structure and composition, assuming that wildlife will recolonize
40 restored areas soon after suitable habitat conditions have been provided (the “Field of Dreams”
41 hypothesis; Palmer et al. 1997). Yet, when wildlife responses to restoration are evaluated, they
42 are not always successful (Shanahan et al. 2011, Cristescu et al. 2013, Calhoun et al. 2014). This
43 is perhaps not surprising since it is usually unrealistic for restoration practitioners to remove
44 human modifications to ecosystems such as dams and levees, reverse the spread of non-native
45 species, or prevent climate change-induced shifts in weather extremes, any of which can impact
46 wildlife populations and the degree to which they benefit from a restoration project.

47 Consequently, if the goals of a restoration project include benefitting wildlife populations, it is
48 essential to assess wildlife responses to restoration. However, we argue that the field of
49 ecological restoration will benefit the most from efforts to design restoration projects that allow
50 comparing the relative effectiveness of alternative restoration designs and methods.

51 Several papers have outlined general concepts and theory for developing a restoration
52 experiment to evaluate effects on wildlife, including an array of analytical approaches and a
53 multitude of pitfalls and barriers to successful experiments (Michener 1997, Chapman 1998,
54 Block et al. 2001). Despite this guidance, experiments evaluating wildlife responses to habitat
55 restoration remain rare, and likely have been hampered by financial and logistical constraints
56 combined with fear of failure (Aslan et al. 2013, Dickens and Suding 2013). We suspect that
57 researchers and practitioners fear that an imperfect experiment will result in non-significant *p*-
58 values, squandered resources, and a failure to learn anything new. However, continuing to use
59 conventional restoration methods and designs without experimentation guarantees learning
60 nothing new. Here, we address the major challenges of designing a large-scale restoration
61 experiment, with a focus on evaluating wildlife responses. As a case study, we then describe how
62 we designed an experiment to evaluate the response of riparian birds to three habitat restoration
63 treatments as part of a project enhancing floodplain connectivity along the Cosumnes River in
64 the Central Valley of California.

65

66 **Addressing the Challenges of Designing Restoration Experiments to Evaluate** 67 **Wildlife Responses**

68 The challenges of designing large-scale restoration experiments to evaluate wildlife responses
69 begin with a lack of clearly defined goals and objectives for the restoration project itself,

70 including wildlife species and response metrics that either have not been specified, are too
71 numerous, or are too impractical to study. When it comes to the experimental design itself, a
72 major challenge lies in balancing a study area that is both (a) large enough for multiple replicates
73 of each treatment on plots that are large enough to detect a wildlife response and (b) small
74 enough that the replicate plots are relatively similar to each other and that the implementation
75 and evaluation of the experiment is manageable over multiple years. The classical experimental
76 design assumptions of randomly assigned treatments and controls and independence among
77 replicate plots further add to the challenge. Finally, there is considerable uncertainty over how to
78 evaluate the results and draw conclusions, particularly when the experimental design is
79 imperfect. To help facilitate collaborations between researchers and practitioners in designing
80 successful restoration experiments to evaluate wildlife responses, we address each of these
81 challenges sequentially, making recommendations and referring readers to the relevant literature
82 for more information.

83

84 ***1. Define the Goals and Objectives***

85 Defining the goals and objectives is a critical first step to designing successful habitat restoration
86 projects and experiments. We distinguish the goals of a restoration project, which are often broad
87 and visionary statements, from the objectives of a restoration project, which are the specific,
88 measurable outcomes necessary to achieving the goals (CMP 2013). For example, to meet a goal
89 of restoring habitat to benefit migratory birds, objectives should specify which types of
90 migratory birds (e.g., ducks or warblers), what determines whether the birds have benefitted
91 (e.g., changes in abundance, survival, or reproductive success), and the size of the response that
92 will be considered a success (e.g., target population size). The objectives of the restoration

93 experiment will flow directly from the project's goals and objectives (e.g., which of these
94 alternative restoration approaches achieves the project's goals and objectives more quickly?). A
95 conceptual model is useful for illustrating the hypothesized cause-effect relationships linking
96 specific restoration actions to achieving the restoration project's objectives and goals (Figure 1;
97 Michener 1997; Holmes & Miller 2010), as well as for identifying key uncertainties in cause-
98 effect relationships that could be addressed by a restoration experiment. We recommend that
99 researchers and practitioners collaborate on the experimental design from the start, including
100 defining the project's goals and objectives and developing a conceptual model, to ensure that the
101 restoration experiment is achievable, designed to provide the research-based information needed
102 by practitioners, and has the best chances for success.

103 ***Identify the species of interest.*** In many cases, the project's goals will include a broadly-
104 defined group of species (e.g., small mammals), such that we recommend carefully selecting a
105 subset of individual species that can be feasibly monitored. We recommend first considering the
106 type of monitoring required for each species, especially the investment in time and resources
107 required for gathering the appropriate data for the response metrics of interest (discussed in the
108 next section). We also recommend choosing species that are dependent on the type of habitat
109 being restored and are likely to respond soon after the habitat becomes suitable (Carignan and
110 Villard 2002), so that the lack of a response is a clear indication that the restoration was
111 unsuccessful. For example, one should consider the location of existing source populations and
112 likely dispersal distances, which can limit the ability of a species to colonize suitable restored
113 habitat (Golet et al. 2011, Grundel et al. 2014). Similarly, one should consider whether there are
114 broader factors that may limit the ability of the species to respond to a restoration project, such as
115 disease susceptibility or habitat limitation during other parts of a migratory species' annual cycle

116 (Block et al. 2001, Carignan and Villard 2002). These species may still be important to include
117 in the project's objectives, but the meaning of a lack of response to the restoration is less clear.
118 Finally, we recommend maximizing the diversity of the species chosen, in terms of trophic
119 position, life history, behavior, or even the time of year during which it relies on the type of
120 habitat being restored, so that they each reflect different aspects of habitat or ecosystem
121 condition (Carignan and Villard 2002).

122 In some cases, the species of interest will already be defined, such as restoration projects
123 designed specifically for the recovery of a threatened species. However, we caution that it can be
124 difficult to measure a response to restoration by very rare species, at least in the short term,
125 because only small numbers of animals are initially available to respond. Thus, the meaning of
126 an apparent lack of response to the restoration may be unclear. Similarly, if the species of interest
127 is a highly mobile species relative to the spatial scale of the restoration project, it may be difficult
128 to detect a measurable response within the restored area. In these cases, it may be useful to select
129 and monitor several additional species that can provide a better indication of immediate changes
130 in local habitat quality (Carignan and Villard 2002). In addition to the criteria for selecting
131 species described in the previous paragraph, we recommend considering species likely to
132 respond in a similar way to the rare or highly mobile species, such as those with similar habitat
133 requirements, life histories, or behavior.

134 ***Select response metrics that reflect project goals.*** There are many wildlife response
135 metrics that could be examined (Johnson 2007), each with its own advantages and disadvantages.
136 For example, differences in the probability of species occurrence between sites or years can be
137 estimated using species presence/absence data (Mackenzie and Royle 2005), which may require
138 as little as a few hours of active effort per plot, including methods such as point counts, camera

139 traps, or acoustical monitoring (Ralph et al. 1995, O'Farrell and Gannon 1999). Similar methods
140 can also be used to estimate changes in abundance or density (Buckland et al. 2001, Rowcliffe et
141 al. 2008), which can indicate an improvement in habitat quality (Bock & Jones 2004; but see
142 Van Horne 1983). As an alternative to species presence or abundance, changes in demographic
143 performance, including reproductive success and survival, can provide a deeper, mechanistic
144 understanding of species responses to restoration, but may require a considerably larger
145 investment of time and resources. For example, locating and monitoring songbird nests can take
146 several hours over several weeks or months for each nest, while mist-netting and banding
147 songbirds to acquire mark-recapture or mark-resight data can take hundreds of net hours over
148 several months or years for each survival estimate (Ralph et al. 1993, White and Burnham 1999).
149 Similarly, measures of body condition or behavior can also indicate habitat quality (Cooke and
150 Suski 2008, Morrison and Lindell 2011), but a considerable investment of time and resources
151 may be required to capture individuals for physiological metrics or to record individual behavior.
152 We recommend selecting wildlife response metrics that most closely reflect the goals of the
153 restoration project and fit the conceptual model linking restoration actions to changes in habitat
154 conditions and subsequent effects on the wildlife response metrics, while also considering the
155 intensity and duration of monitoring effort and the complexity of analysis each metric requires.

156 ***Define a successful response to restoration.*** In addition to identifying the species and
157 response metrics of interest, defining the specific, measurable objectives of a restoration project
158 requires quantifying the magnitude of the wildlife response that will be considered a success
159 (Osenberg et al. 2006). For example, the objectives may include an improvement over the
160 baseline conditions and/or to reach a specific benchmark, such as a desired abundance or
161 survival rate. Setting a specific benchmark can be challenging, but common approaches are to

162 base objectives on historical information or on relatively undisturbed reference sites used for
163 comparison (Rich et al. 2004, Gardali et al. 2006). In many cases, comparable reference sites
164 may not exist and objectives based on historical information may be unrealistic in landscapes
165 that have changed significantly due to factors like urbanization, altered fire regimes, or water
166 diversion (Suding 2011). In these cases, alternative approaches to setting objectives should be
167 considered (Sanderson 2006). Given the rapid pace of climate change, these objectives should be
168 informed by historical conditions, but also be forward-looking and incorporate information about
169 projected future conditions (Chornesky et al. 2015).

170 ***Identify the objectives of the experiment.*** Experiments examining wildlife responses to
171 restoration will commonly include the objectives of identifying which of several restoration
172 treatment options is most effective in achieving the project's wildlife response objectives, and
173 because restoration requires substantial financial investment, which option is most cost-effective
174 or most risky. Given the projected impacts of climate change for a region (e.g., warmer
175 temperatures and increased risk of extreme drought), the objectives of the experiment might also
176 include identifying which treatment option is able to meet the project objectives during extreme
177 weather conditions and has the highest likelihood of long-term persistence.

178

179 ***2. Design the Experiment.***

180 A large-scale restoration experiment is unlikely to meet all of the standard assumptions of
181 classical experimental design, including the randomized assignment of treatments and controls to
182 independent replicated experimental plots (Hurlbert 1984, Gotelli and Ellison 2004). Further, the
183 design of any habitat restoration experiment will be complicated by the real-world logistical
184 constraints and complications of on-the-ground restoration and monitoring that will take place

185 over multiple years. However, imperfect ‘quasi-experiments’ are not doomed to failure and will
186 still provide valuable information (Block et al. 2001), particularly if the alternative is not to
187 attempt any experiment at all. We recommend that researchers and practitioners acknowledge up
188 front that the experiment may not meet classical experimental design rules and work together to
189 maximize the value of the experiment *despite* breaking these rules.

190 ***Select restoration treatments and controls that will advance understanding.*** A critical
191 first step is to define the alternative restoration methods or treatments that will be compared and
192 contrasted, based on the objectives of the experiment defined in the previous section. The
193 simplest experiments would compare restored (treated) and unrestored (control) groups of
194 experimental plots. However, if the objective of the experiment is instead to determine whether a
195 particular restoration technique improved the wildlife response, such as irrigation of planted
196 trees, the experiment would compare plots with irrigated (treated) and unirrigated (control)
197 planted trees. If the size of the study area allows, more than one restoration treatment can be
198 included to examine the relative effectiveness of multiple methods. We recommend selecting the
199 smallest number of restoration treatments necessary to address the objectives of the experiment,
200 allowing for more replicates (plots) of each treatment.

201 ***Define the study area and, if necessary, divide it into blocks.*** We assume the study area
202 for the experiment will fall within the boundaries of a previously defined restoration project,
203 determined by property boundaries, funding, and/or logistical constraints. The study area for the
204 experiment may be identical to the project area or restricted to a subset of that area. For example,
205 consider excluding areas with unusual features and instead focus on areas that are more
206 representative of the landscape to be restored. Within the experiment’s study area, there may still
207 be important heterogeneity representative of a range of conditions in the landscape of interest,

208 such as slope, aspect, elevation, soil moisture, or prior land use. If so, we recommend dividing
209 the study area into a few blocks capturing the major patterns in this heterogeneity and/or
210 identifying important covariates to be measured on each plot (Hurlbert 1984, Gotelli and Ellison
211 2004). Focus on the primary sources of heterogeneity that are most likely to have a strong
212 interaction with the restoration treatments that will be applied, and therefore a measurable impact
213 on wildlife responses.

214 ***Define the minimum plot size.*** Each restoration treatment will ideally be applied to
215 multiple experimental plots within the study area and at least one per block. Selecting an
216 appropriate size for experimental plots is of the utmost importance, because if the plots are too
217 small, there may not be a measurable response by wildlife (Block et al. 2001), but if they are too
218 large, it will be difficult to replicate each treatment within each block. We recommend first
219 defining the minimum plot size appropriate to the restoration treatment, species, and response
220 metrics of interest. For example, to measure a change in the number of territories on a plot (i.e.
221 species presence or an increase in density), the plots must be large enough to accommodate
222 multiple territories. Information about the typical home range, territory size, or density of the
223 species of interest, or simply the scale of previous successful habitat restoration efforts, can
224 provide guidance. After identifying the minimum plot size required, it is straight-forward to
225 determine the maximum number of plots that can fit within the study area and/or within each
226 block. Ideally, there will be enough space for one plot of each treatment type per block and
227 multiple plots of each treatment within the study area.

228 ***Confront complications with compromises.*** This is the stage in the experimental design
229 process where the ideal experiment is confronted with the realities on the ground. When deciding
230 how to compromise, we recommend revisiting the goals and objectives to ensure that the

231 experiment will still address the key uncertainties of interest. For example, given the species of
232 interest and restoration treatments to be examined, it may not be possible to fit multiple
233 replicates of each treatment within the study area or one of each treatment per block. In this case,
234 either the blocks or study area need to be larger, the total number of treatments examined fewer,
235 or the plots smaller. While the total size of the study area is often limited by outside constraints,
236 the number of the blocks could be reduced. The objectives of the experiment could also be
237 refined to focus on fewer restoration treatments (e.g. one treatment and one control only). On the
238 other hand, if the species of interest require plot sizes so large that it will always be difficult to fit
239 multiple plots within a study area of manageable size, consider selecting different species that
240 will be more likely to respond to restoration on a smaller scale (see previous section). As an
241 alternative, consider accepting that there will not be replicates. For example, in the case of very
242 large-scale, costly, and/or opportunistic restoration treatments that cannot be replicated on
243 multiple plots, it is still possible to compare one treatment and one control plot using the before-
244 after-control-impact (BACI) design (Stewart-Oaten et al. 1992, Osenberg et al. 2006),
245 particularly if the effect of the treatment is expected to be large, and provided pre-treatment data
246 on both plots are available. Further, replication and strength of inference can come from
247 repeating experiments over time and meta-analysis (Oksanen 2001).

248 Another complication to confront is the potential lack of independence between adjacent
249 plots. Ideally, plots would be spaced far apart to minimize any influence of neighboring plots and
250 the chances that the same individual animal is sampled on multiple plots (Block et al. 2001,
251 Gotelli and Ellison 2004). However, widely dispersed plots can be more difficult and costly to
252 monitor and are also more likely to differ in other ways that can influence the outcome of the
253 experiment (Gotelli and Ellison 2004). Further, wildlife responses to a restoration plot will

254 always be influenced by the surrounding landscape, whether it's another experimental plot or
255 land outside the study area that may be restored or unrestored; no field experiments take place in
256 a vacuum. Thus, we recommend carefully considering how the landscape surrounding each plot
257 might influence wildlife responses, such as by magnifying or minimizing the apparent effect of
258 each treatment. Randomizing the assignment of each plot to a treatment can help by ensuring that
259 treatments and controls are interspersed throughout the study area.

260

261 ***3. Determine How and When the Results Will Be Evaluated.***

262 Perhaps the greatest objection to conducting an experiment such as we have outlined in the
263 previous section is that any statistical analysis of the results will be invalid if any of the classical
264 experimental design assumptions have been violated, and thus the experiment will have been a
265 waste of time and resources. This closed-minded approach is preventing practitioners from
266 attempting new or unconventional restoration methods and designs, in turn hindering advances in
267 the field of ecological restoration, even as the need to develop climate-smart restoration methods
268 continues to grow (Hobbs et al. 2009, Chornesky et al. 2015, Perry et al. 2015). Thus, we
269 recommend practitioners and researchers discuss how and when the results of the experiment
270 will be evaluated, consider the possible outcomes, and plan for the unexpected.

271 ***How.*** A conventional statistical approach would test a null hypothesis of no difference
272 between restoration treatments, such as a simple t-test comparing the wildlife response metrics in
273 treatment and control plots at the same point in time after the restoration. If the classical
274 experimental design assumptions have been violated, such as due to a lack of replication or
275 randomization among plots, the results of any significance test are meaningless (Hurlbert 1984).
276 However, in most cases, null hypothesis testing will be less important than estimating the

277 strength of evidence for an effect and the relative effect size of each restoration treatment
278 (Stewart-Oaten et al. 1992, Fidler et al. 2006, Osenberg et al. 2006). For example, information-
279 theoretic methods provide a way to evaluate the relative strength of support for alternative
280 models with or without a treatment effect and model averaging (Burnham and Anderson 2002).
281 Bayesian methods can be used to directly evaluate the probability of a particular effect size, and
282 have been used to generate a better understanding of the impacts of logging on rainforest
283 mammals (Crome et al. 1996), forest treatments on Spotted Owl productivity (Lee and Irwin
284 2005), and avian responses to prescribed fire (Russell et al. 2009). Strategies used to assess
285 environmental impacts, which are neither replicated nor randomized, may also be useful, such as
286 accounting for differences in baseline values by comparing the change in each plot before and
287 after restoration (Wiens and Parker 1995).

288 **When.** Wildlife responses to restoration treatments may differ in the total magnitude of
289 change, rate of change, persistence of the change over time, or all of the above (Figure 2), such
290 that the results of the experiment may depend on when the results are evaluated. For example, it
291 can take a decade or more for conditions at a restoration site to become suitable for certain
292 species and the full magnitude of the change to be known (Gardali et al. 2006). Similarly, the
293 ability of a site to withstand extreme weather conditions may require waiting for a number of
294 years before those extreme conditions occur. Differences in the rate of change may be a useful
295 initial indicator of relative effectiveness, if a faster response is likely to be correlated with a
296 greater magnitude of change or achieving project goals more quickly, but these differences may
297 not persist over time. Thus, depending on the objectives of the experiment and the time frame
298 available for conducting the experiment, we recommend evaluating the selected wildlife
299 response metrics for as long as possible to examine the trajectory of each plot and the long-term

300 relative effect size of each treatment, and in turn build a more complete understanding of how
301 the system works.

302 *Consider the possible outcomes.* With a better understanding of the relative effect size of
303 each treatment and the strength of evidence for a difference between treatments, an important
304 next step is to revisit the conceptual model linking restoration actions to changes in habitat
305 conditions and subsequent wildlife responses (Figure 1). We recommend evaluating whether the
306 sequence of events after restoration and resulting wildlife responses were consistent with the
307 current understanding of how the system works. In a best case scenario, the restoration
308 treatments will have resulted in large and obvious differences in habitat conditions and wildlife
309 responses between plots, even before any analysis is conducted, successfully identifying what
310 worked and what did not. Another successful outcome would be restoration treatments that
311 produced similar habitat conditions and wildlife responses; with no clear winner, such an
312 outcome would suggest that multiple restoration approaches are available to be added to the
313 practitioner's toolbox, perhaps including ones that may be more cost-effective or use fewer
314 resources and should be considered further.

315 Other more confusing outcomes include treatments that result in highly variable habitat
316 conditions and wildlife responses across plots, suggesting there were other important
317 environmental factors not considered by the experimental design, or outcomes that were entirely
318 unexpected. For example, two different treatments could ultimately result in similar wildlife
319 responses despite creating very different habitat conditions, or two different treatments could
320 create apparently similar habitat conditions but result in very different wildlife responses. In
321 either case, these results would suggest that the species' habitat requirements are not well
322 understood. While these outcomes may be less desirable, at the very least, these outcomes would

323 expose previously unrecognized uncertainties in the conceptual model and demonstrate a need to
324 consider alternative restoration designs or methods.

325 *Plan for the unexpected.* As with any restoration project or field experiment, unexpected
326 events or environmental variation can interfere. For example, extreme weather, fire, or flood
327 could interfere with the ability to establish new vegetation, setting back the time table for the
328 project. On the other hand, researchers and practitioners could take advantage of the opportunity
329 to see how the different restoration methods fared under extreme conditions. If environmental
330 conditions vary considerably between years, resulting in annual variation in wildlife responses, a
331 comparison of the differences between treatments in each year can provide insight into whether
332 one treatment consistently fares better than the other (Wiens and Parker 1995, Osenberg et al.
333 2006).

334 There are many other reasons why the restoration may not proceed as expected. For
335 example, vegetation may not recruit quickly or survive as well as expected, such that the wildlife
336 species of interest have not (yet) responded. In some cases, invasive plant or animal species may
337 begin to recruit instead, with potentially long-lasting negative impacts. We recommend that
338 researchers and practitioners plan for these scenarios, and identify conditions under which the
339 experiment will be allowed to go on longer or trigger points at which the experiment will end to
340 prevent harm. By working together from the start of the experimental design process through the
341 implementation and evaluation phases, restoration practitioners and researchers can address these
342 challenges proactively and creatively to maximize the experiment's chances of success.

343

344 **Lower Cosumnes River Restoration as a Case Study**

345 As an example of how these guidelines can be applied in the field, we describe our efforts to

346 establish a restoration experiment in the Central Valley of California. Thirty years ago, the
347 Cosumnes River was identified as a conservation and restoration priority in the Central Valley of
348 California, USA, due to the presence of large blocks of remnant riparian forest and grassland in
349 an otherwise agricultural and rapidly developing landscape (Swenson et al. 2003). Since then,
350 nongovernmental conservation organizations and public agencies have worked together to
351 protect and restore riparian forest. First established in 1987, the Cosumnes River Preserve now
352 protects over 18,000 ha.

353 Riparian forest restoration efforts began with direct planting of native trees and shrubs
354 into fallowed farm fields in an attempt to reconnect blocks of native riparian forests, but this
355 expensive and time-consuming work often had limited success in this highly modified landscape
356 with high levels of pressure from exotic invasive species (Swenson et al. 2012). However,
357 investigation of the landscape, historical records, and photographs revealed instances where
358 unintentional breaches in the river's levees had resulted in the development of diverse, early and
359 late successional riparian forests, presumably due to enhanced floodplain connectivity. This
360 discovery inspired a new era of using the Cosumnes River Preserve as a restoration and research
361 platform to investigate the ecological and human benefits of enhancing hydrological process to
362 restore ecosystem function and diversity.

363

364 ***1. Goals and Objectives***

365 The Lower Cosumnes River Restoration Project, led by the Nature Conservancy (TNC), is
366 enhancing floodplain hydrological processes on approximately 200 ha at the Cosumnes River
367 Preserve, and is the largest levee-breach yet implemented on this river. The goal of this project is
368 to restore riparian ecosystem function and diversity, with multiple benefits including providing

369 habitat for riparian wildlife.

370 ***Species of interest.*** We expect that many species will benefit, but birds are recognized as
371 good indicators of ecosystem condition (Temple and Wiens 1989, Carignan and Villard 2002,
372 Ortega-Álvarez and Lindig-Cisneros 2012), and monitoring techniques are well-established and
373 cost-effective (Ralph et al. 1993). Further, previous studies of riparian vegetation restoration in
374 the Central Valley have documented an immediate increase in the abundance of common bird
375 species and the return of previously absent species once the vegetation was established (Gardali
376 et al. 2006, Dybala et al. 2014), suggesting that riparian birds will respond quickly once the
377 vegetation becomes suitable. Thus, we chose to focus on a subset of riparian breeding bird
378 species that have been identified by the Central Valley Joint Venture (CVJV;
379 www.centralvalleyjointventure.org) as indicators of riparian ecosystem function (Table 1;
380 Dybala et al. in press). These species were selected because they use riparian vegetation as a
381 principal breeding habitat in the Central Valley and represent a range of life histories and
382 specific vegetation associations (Table 1), providing information about different aspects of
383 Central Valley riparian ecosystems.

384 ***Response metric that reflects project goals.*** Because our goal is to provide habitat for
385 riparian wildlife, and we have selected focal species that breed in riparian habitat, our response
386 metric of interest is the breeding density of each focal species. We will measure breeding
387 densities by spot mapping birds that are exhibiting breeding behavior (e.g., singing or carrying
388 nest material or food; Ralph et al. 1993). This method can measure the bird response at smaller
389 scales and with greater precision than the more common point count technique, and allows
390 collecting information about the density and distribution of territories on each plot.

391 ***Definition of successful response.*** We defined success as breeding densities for all 6

392 riparian focal species increasing from the pre-restoration baseline of 0 breeding territories
393 (Dettling and Seavy 2011) to territory densities that are equivalent to or higher than current
394 average breeding densities in riparian vegetation throughout the region within 10 years of
395 implementation (Table 1; Dybala et al. in press).

396 ***Objectives of the experiment.*** We expect that successfully restoring a diverse riparian
397 vegetation community will in turn support an abundant riparian breeding bird community,
398 meeting the objectives and goals of the project (Figure 1). However, the key uncertainty we
399 identified is whether it is necessary to invest substantial resources in traditional, horticultural
400 restoration (i.e. manually planting native vegetation, irrigation, and weed management) on top of
401 the process-based restoration (i.e. increasing floodplain connectivity and frequency of
402 inundation; Beechie et al. 2010), or whether a reduced effort could be just as effective. Where
403 hydrological processes are limited, intensive horticultural restoration practices may generally be
404 more effective in establishing riparian vegetation and achieving the desired wildlife response,
405 compared to a relatively low effort horticultural restoration that relies more on natural
406 recruitment but may be vulnerable to invasion by exotic species. However, we hypothesized that
407 enhancing hydrological processes would improve the effectiveness of reduced effort horticultural
408 restoration due to sediment deposition, seed dispersal and groundwater recharge (Florsheim &
409 Mount 2003; Opperman 2012).

410 To test this hypothesis, we designed a restoration experiment with the objective of
411 evaluating the effectiveness of different levels of horticultural restoration effort in increasing
412 riparian breeding bird abundance across a gradient of floodplain connectivity. We predict that
413 where floodplain connectivity is high, reduced effort horticultural restoration will result in an
414 equal (or improved) riparian breeding bird densities yet cost less and require fewer resources

415 (e.g., water for irrigation) than high effort horticultural restoration practices.

416

417 **2. Experimental Design**

418 ***Restoration treatments and controls.*** To examine the bird responses to additional
419 horticultural restoration effort on top of the process-based restoration, we defined 3 experimental
420 groups including 2 scenarios of horticultural restoration effort and control plots with no
421 horticultural restoration effort:

- 422 1. High effort, representative of many river restoration projects in North America, including
423 a structurally diverse planting palette and irrigation of native plantings;
- 424 2. Reduced effort, including the planting of native trees only; and
- 425 3. Control plots, which will go unplanted.

426 These 3 groups allow us to compare the bird response between high and reduced effort scenarios
427 of horticultural restoration, as well whether either of these groups is different from the control
428 plots.

429 ***Study area and blocks.*** Within the 200 ha project area, we selected 120 ha of former
430 agricultural land along the Cosumnes River as the study area for the experiment (Figure 4).
431 Because we expect the bird response in each type of plot to vary with the degree of floodplain
432 inundation, we took advantage of a natural gradient of topography and floodplain connectivity in
433 the study area and organized the study area into 3 blocks representing the degree of floodplain
434 connectivity.

435 ***Minimum plot size.*** To estimate the minimum plot size necessary for each of the six focal
436 species, we again drew on current regional average densities in riparian vegetation in the Central
437 Valley (Table 1; Dybala et al. in press). Densities ranged from 0.34 to 5.35 birds/ha, and

438 assuming two individuals per territory, we estimated that 10 ha plots could support between 1.70
439 and 26.75 territories of each focal species. Thus, we chose to aim for 12 plots that were
440 approximately 10 ha in size, with 4 plots in each of the 3 blocks.

441 ***Compromises.*** For convenience, we used existing boundaries (e.g. roads or irrigation
442 ditches) to divide the 120 ha study area into 12 plots (Figure 4), which resulted in some variation
443 in plot size. Initially, all of the plots were over 9 ha, but hydrogeomorphology monitoring close
444 to the streambed ultimately required excluding a portion of the study area, and two plots were
445 reduced to approximately 7 ha. Although we would have preferred larger plots, all plots were
446 above the minimum of 6 ha recommended for Breeding Bird Census plots (15 acres; Hall 1964).

447 We also recognized that the plots are adjacent to each other, such that neighboring plots
448 could influence an individual bird's habitat selection decisions. As a result, the differences in
449 breeding densities among plots could either be exaggerated if one plot type is consistently
450 preferred over the other adjacent plots (Gotelli and Ellison 2004), or underestimated if birds
451 establishing territories in the preferred plot attract additional birds whose territories spill over
452 into adjacent plots. However, the random assignment of treatments to plots ensures that
453 treatments are interspersed (Figure 4), and we expect that our territory mapping and behavioral
454 observations will allow us to detect if either of these cases is happening. The alternatives would
455 be to: (1) greatly reduce the plot sizes to create space between plots, which would severely limit
456 the number of territories that could be established in each plot and likely still would not ensure
457 independence between plots; (2) attempt to establish widely dispersed and isolated plots, which
458 would likely be more heterogeneous and could still be influenced by the surrounding agricultural
459 matrix; or (3) simply not conduct an experiment and guarantee learning nothing new. We felt
460 that none of these alternatives were acceptable, and that this experiment would still provide

461 valuable insights into the relative effectiveness of the restoration treatments, *despite* the potential
462 lack of independence between plots.

463

464 **3. Evaluating Results**

465 **How.** To evaluate the response of each focal species, we will map breeding territories
466 across the study area and quantify the territory density of each focal species in each plot
467 (territories/ha). Using the territory density of each species as a response variable, we can use
468 generalized linear models with an information-theoretic approach to evaluate the weight of
469 evidence for differences among treatments. By quantifying and incorporating the actual number
470 of days each plot is inundated as a covariate, we can also examine the influence of floodplain
471 connectivity.

472 **When.** We anticipate that our results will initially vary among species, as well as over
473 time. While we expect an immediate response from several of the focal species that are
474 associated with early successional riparian vegetation (i.e., within 3 years), it will take more time
475 for the area to become suitable for species associated with mature trees (Gardali et al. 2006).
476 Early results may be an important indication of whether or not all treatments are on a desirable
477 trajectory, and thus whether the experiment should continue, while continued monitoring will
478 reveal the full value of each treatment for each species. The analysis will be repeated over time
479 to re-evaluate the total magnitude, rate, and persistence of change in plots of each treatment type,
480 as well as whether the bird response to high and reduced effort treatments are converging.

481 **Possible outcomes.** The results of the Cosumnes River experiment will help evaluate how
482 much planting effort is necessary to achieve the desired outcome for riparian bird populations.
483 Based on our understanding of the rate of riparian vegetation growth and riparian bird responses,

484 we expect that 5 years after restoration focal species in the high effort plots will have achieved
485 densities >75% of the regional average densities (Table 1), while focal species in the reduced
486 effort and control plots will reach densities of <50% and <10% of regional average densities,
487 respectively (Figure 3). However, if wildlife responses to the reduced effort treatment are
488 relatively similar to the high effort treatment (e.g., >50% of regional average densities after 5
489 years and still increasing), this experiment may identify an alternative, more cost-effective option
490 for riparian restoration. If wildlife responses vary with floodplain connectivity, it will illuminate
491 the role of this connectivity and flooding in the success of restoration. Otherwise, it will
492 contribute scientific evidence for the necessity of traditional, high effort planting, even where
493 floodplain connectivity is high.

494

495 **Conclusions**

496 Nobody likes to fail, and when we consider how easy it is to sling arrows at large-scale
497 restoration experiments, it's not surprising that relatively few examples exist. Conducting
498 restoration experiments on spatial and temporal scales relevant for wildlife is challenging but not
499 impossible, and these experiments are very likely to generate new insights into effective
500 restoration methods and designs. Identifying effective methods that will persist under climate
501 change is becoming increasingly important, and is likely to require experimenting with new and
502 unconventional methods. By addressing the logistical challenges and fear of failure, we intend
503 for the recommendations and examples presented here to facilitate collaborations between
504 researchers and practitioners and accelerate the rate of relatively large-scale, long-term
505 restoration experimentation. By addressing key uncertainties, such experiments will demonstrate
506 the relative effectiveness of alternative methods, in turn helping restoration practitioners choose

507 the best course of action, accelerating the rate of successful restoration, and advancing the field
508 of ecology restoration.

509

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516

517 **References**

518 Aslan, C. E., M. L. Pinsky, M. E. Ryan, S. Souther, and K. A. Terrell. 2013. Cultivating
519 creativity in conservation science. *Conservation Biology* 28:345–353.

520 Beechie, T. J., D. A. Sear, J. D. Olden, G. R. Pess, J. M. Buffington, H. Moir, P. Roni, and M.
521 M. Pollock. 2010. Process-based principles for restoring river ecosystems. *BioScience*
522 60:209–222.

523 Block, W. M., A. B. Franklin, J. P. Ward, J. L. Ganey, and G. C. White. 2001. Design and
524 implementation of monitoring studies to evaluate the success of ecological restoration on
525 wildlife. *Restoration Ecology* 9:293–303.

526 Bock, C. E., and Z. F. Jones. 2004. Avian habitat evaluation: Should counting birds count?
527 *Frontiers in Ecology and the Environment* 2:403–410.

528 Buckland, S. T., D. R. Anderson, K. P. Burnham, J. L. Laake, D. L. Borchers, and L. Thomas.
529 2001. *Introduction to Distance Sampling: Estimating Abundance of Biological*

- 530 *Populations*. Oxford: Oxford University Press.
- 531 Burnham, K. P., and D. R. Anderson. 2002. *Model selection and multi-model inference: a*
532 *practical information-theoretic approach*. Second edition. New York: Springer-Verlag.
- 533 Calhoun, A. J. K., J. Arrigoni, R. P. Brooks, M. L. Hunter, and S. C. Richter. 2014. Creating
534 successful vernal pools: A literature review and advice for practitioners. *Wetlands*
535 34:1027–1038.
- 536 Carignan, V., and M.-A. Villard. 2002. Selecting indicator species to monitor ecological
537 integrity: A review. *Environmental Monitoring and Assessment* 78:45–61.
- 538 Chapman, M. G. 1998. Improving sampling designs for measuring restoration in aquatic habitats.
539 *Journal of Aquatic Ecosystem Stress and Recovery* 6:235–251.
- 540 Chornesky, E. A., D. D. Ackerly, P. Beier, F. W. Davis, L. E. Flint, J. J. Lawler, P. B. Moyle, M.
541 A. Moritz, M. Scoonover, K. Byrd, P. Alvarez, N. E. Heller, E. R. Micheli, and S. B.
542 Weiss. 2015. Adapting California’s ecosystems to a changing climate. *BioScience*
543 65:247–262.
- 544 Conservation Measures Partnership (CMP). 2013. *The open standards for the practice of*
545 *conservation*. Third edition. Washington, D.C. www.conservationmeasures.org
- 546 Cooke, S. J., and C. D. Suski. 2008. Ecological restoration and physiology: An overdue
547 integration. *BioScience* 58:957–968.
- 548 Cristescu, R. H., J. Rhodes, C. Frère, and P. B. Banks. 2013. Is restoring flora the same as
549 restoring fauna? Lessons learned from koalas and mining rehabilitation. *Journal of*
550 *Applied Ecology* 50:423–431.
- 551 Crome, F. H. J., M. R. Thomas, and L. A. Moore. 1996. A novel Bayesian approach to assessing
552 impacts of rain forest logging. *Ecological Applications* 6:1104–1123.

- 553 Dettling, M. D., and N. E. Seavy. 2011. Cosumnes River Preserve Denier and Oneto property
554 pre-restoration avian monitoring 2011 progress report. Petaluma, CA: PRBO
555 Conservation Science.
- 556 Dickens, S., and K. Suding. 2013. Spanning the science-practice divide: Why restoration
557 scientists need to be more involved with practice. *Ecological Restoration* 31:134–140.
- 558 Dybala, K. E., N. Clipperton, T. Gardali, G. H. Golet, R. Kelsey, S. Lorenzato, R. Melcer Jr., N.
559 E. Seavy, J. G. Silveira, and G. S. Yarris. In press. Population and habitat objectives for
560 avian conservation in California's Central Valley riparian ecosystems. *San Francisco*
561 *Estuary and Watershed Science*.
- 562 Dybala, K. E., N. E. Seavy, M. D. Dettling, M. M. Gilbert, and R. Melcer. 2014. Does restored
563 riparian habitat create ecological traps for riparian birds through increased Brown-headed
564 Cowbird nest parasitism? *Ecological Restoration* 32:239–248.
- 565 Fidler, F., M. A. Burgman, G. Cumming, R. Buttrose, and N. Thomason. 2006. Impact of
566 criticism of null-hypothesis significance testing on statistical reporting practices in
567 conservation biology. *Conservation Biology* 20:1539–1544.
- 568 Florsheim, J. L., and J. F. Mount. 2003. Changes in lowland floodplain sedimentation processes:
569 pre-disturbance to post-rehabilitation, Cosumnes River, CA. *Geomorphology* 56:305–
570 323.
- 571 Gardali, T., A. L. Holmes, S. L. Small, N. Nur, G. R. Geupel, and G. H. Golet. 2006. Abundance
572 patterns of landbirds in restored and remnant riparian forests on the Sacramento River,
573 California, U.S.A. *Restoration Ecology* 14:391–403.
- 574 Golet, G. H., T. Gardali, J. W. Hunt, D. A. Koenig, and N. M. Williams. 2011. Temporal and
575 taxonomic variability in response of fauna to riparian restoration. *Restoration Ecology*

- 576 19:126–135.
- 577 Goreau, T. J. and R. K. Trench (eds). 2012. *Innovative methods of marine ecosystem restoration*.
578 Boca Raton, FL: CRC Press.
- 579 Gotelli, N. J., and A. M. Ellison. 2004. *A primer of ecological statistics*. Sunderland, MA:
580 Sinauer Associates, Inc.
- 581 Grundel, R., D. A. Beamer, G. A. Glowacki, K. J. Frohnapple, and N. B. Pavlovic. 2014.
582 Opposing responses to ecological gradients structure amphibian and reptile communities
583 across a temperate grassland-savanna-forest landscape. *Biodiversity and Conservation*
584 24:1089–1108.
- 585 Hall, G. A. 1964. Breeding-bird censuses - why and how. *Audubon Field Notes* 18:413–416.
- 586 Hobbs, R. J., E. S. Higgs, and J. A. Harris. 2009. Novel ecosystems: Implications for
587 conservation and restoration. *Trends in Ecology and Evolution* 24:599–605.
- 588 Holmes, A. L., and R. F. Miller. 2010. State-and-transition models for assessing grasshopper
589 sparrow habitat use. *Journal of Wildlife Management* 74:1834–1840.
- 590 Van Horne, B. 1983. Density as a misleading indicator of habitat quality. *Journal of Wildlife*
591 *Management* 47:893–901.
- 592 Hurlbert, S. H. 1984. Pseudoreplication and the design of ecological field experiments.
593 *Ecological Monographs* 54:187–211.
- 594 Johnson, M. D. 2007. Measuring habitat quality: A review. *Condor* 109:489–504.
- 595 Kettenring, K. M., and C. R. Adams. 2011. Lessons learned from invasive plant control
596 experiments: A systematic review and meta-analysis. *Journal of Applied Ecology*
597 48:970–979.
- 598 Lee, D. C., and L. L. Irwin. 2005. Assessing risks to spotted owls from forest thinning in fire-

- 599 adapted forests of the western United States. *Forest Ecology and Management* 211:191–
600 209.
- 601 Mackenzie, D. I., and J. A. Royle. 2005. Designing occupancy studies: General advice and
602 allocating survey effort. *Journal of Applied Ecology* 42:1105–1114.
- 603 Michener, W. K. 1997. Quantitatively evaluating restoration experiments: Research design,
604 statistical analysis, and data management considerations. *Restoration Ecology* 5:324–337.
- 605 Morrison, E. B., and C. A. Lindell. 2011. Active or passive forest restoration? Assessing
606 restoration alternatives with avian foraging behavior. *Restoration Ecology* 19:170–177.
- 607 O’Farrell, M. J., and W. L. Gannon. 1999. A comparison of acoustic versus capture techniques
608 for the inventory of bats. *Journal of Mammalogy* 80:24–30.
- 609 Oksanen, L. 2001. Logic of experiments in ecology: is pseudoreplication a pseudoissue? *Oikos*
610 94:27–38.
- 611 Opperman, J. J. 2012. A conceptual model for floodplains in the Sacramento-San Joaquin Delta.
612 *San Francisco Estuary and Watershed Science* 10:jmie_sfews_11155.
- 613 Ortega-Álvarez, R., and R. Lindig-Cisneros. 2012. Feathering the scene: The effects of
614 ecological restoration on birds and the role birds play in evaluating restoration outcomes.
615 *Ecological Restoration* 30:116–127.
- 616 Osenberg, C. W., B. M. Bolker, J.-S. S. White, C. M. St. Mary, and J. S. Shima. 2006. Statistical
617 issues and study design in ecological restorations: lessons learned from marine reserves.
618 Pages 280–302 in D. A. Falk, M. A. Palmer, and J. B. Zedler (eds), *Foundations of*
619 *Restoration Ecology*. Washington: Island Press.
- 620 Palmer, M. A., R. F. Ambrose, and N. L. R. Poff. 1997. Ecological theory and community
621 restoration ecology. *Restoration Ecology* 5:291–300.

- 622 Perry, L. G., L. V. Reynolds, T. J. Beechie, M. J. Collins, and P. B. Shafroth. 2015.
623 Incorporating climate change projections into riparian restoration planning and design.
624 *Ecohydrology* 8:863–879.
- 625 Ralph, C. J., S. Droege, and J. R. Sauer. 1995. Managing and monitoring birds using point
626 counts: Standards and applications. Pages 161–169 in C. J. Ralph, J. R. Sauer, and S.
627 Droege (eds), *Monitoring Bird Populations by Point Counts*. Gen. Tech. Report PSW-
628 GTR-149. Albany, CA: U.S. Department of Agriculture, Forest Service, Pacific
629 Southwest Research Station.
- 630 Ralph, C. J., G. R. Geupel, P. Pyle, T. E. Martin, and D. F. DeSante. 1993. *Handbook of Field*
631 *Methods for Monitoring Landbirds*. Albany, CA: Pacific Southwest Research Station,
632 Forest Service, U.S. Department of Agriculture.
- 633 Rich, T. D., C. J. Beardmore, H. Berlanga, P. J. Blancher, M. S. W. Bradstreet, G. S. Butcher, D.
634 W. Demarest, E. H. Dunn, W. C. Hunter, E. E. Iñigo-Elias, J. A. Kennedy, A. M. Martell,
635 A. O. Panjabi, D. N. Pashley, K. V. Rosenberg, C. M. Rustay, J. S. Wendt, and T. C.
636 Will. 2004. *Partners in Flight North American Landbird Conservation Plan*. Ithaca, NY:
637 Cornell Lab of Ornithology.
- 638 Rowcliffe, J. M., J. Field, S. T. Turvey, and C. Carbone. 2008. Estimating animal density using
639 camera traps without the need for individual recognition. *Journal of Applied Ecology*
640 45:1228–1236.
- 641 Russell, R. E., J. A. Royle, V. A. Saab, J. F. Lehmkuhl, W. M. Block, and J. R. Sauer. 2009.
642 Modeling the effects of environmental disturbance on wildlife communities: Avian
643 responses to prescribed fire. *Ecological Applications* 19:1253–1263.
- 644 Sanderson, E. W. 2006. How many animals do we want to save? The many ways of setting

- 645 population target levels for conservation. *BioScience* 56:911–922.
- 646 Seavy, N. E., T. Gardali, G. H. Golet, F. T. Griggs, C. A. Howell, R. Kelsey, S. L. Small, J. H.
647 Viers, and J. F. Weigand. 2009. Why climate change makes riparian restoration more
648 important than ever: Recommendations for practice and research. *Ecological Restoration*
649 27:330–338.
- 650 Shanahan, S. A., S. M. Nelson, D. M. Van Dooremolen, and J. R. Eckberg. 2011. Restoring
651 habitat for riparian birds in the lower Colorado River watershed: An example from the
652 Las Vegas Wash, Nevada. *Journal of Arid Environments* 75:1182–1190.
- 653 Stewart-Oaten, A., J. R. Bence, and C. W. Osenberg. 1992. Assessing effects of unreplicated
654 perturbations: No simple solutions. *Ecology* 73:1396–1404.
- 655 Suding, K. N. 2011. Toward an era of restoration in ecology: Successes, failures, and
656 opportunities ahead. *Annual Review of Ecology, Evolution, and Systematics* 42:465–487.
- 657 Swenson, R. O., R. J. Reiner, M. D. Reynolds, and J. Marty. 2012. River floodplain restoration
658 experiments offer a window into the past. Pages 218–231 in J. A. Wiens, G. D. Hayward,
659 H. D. Safford, and C. M. Giffen (eds), *Historical Environmental Variation in*
660 *Conservation and Natural Resource Management*. Oxford: John Wiley & Sons, Ltd.
- 661 Swenson, R. O., K. Whitener, and M. Eaton. 2003. Restoring floods on floodplains: Riparian and
662 floodplain restoration at the Cosumnes River Preserve. Pages 224–229 in P. M. Faber
663 (ed), *California Riparian Systems: Processes and Floodplain Management, Ecology,*
664 *Restoration, 2001 Riparian Habitat and Floodplains Conference Proceedings*.
665 Sacramento, CA: Riparian Habitat Joint Venture.
- 666 Temple, S. A., and J. A. Wiens. 1989. Bird populations and environmental changes: Can birds be
667 bio-indicators? *American Birds* 43:260–270.

- 668 White, G. C., and K. P. Burnham. 1999. Program MARK: survival estimation from populations
669 of marked animals. *Bird Study* 46:120–138.
- 670 Wiens, J. A., and K. R. Parker. 1995. Analyzing the effects of accidental environmental impacts:
671 Approaches and assumptions. *Ecological Applications* 5:1069–1083.

Table 1. Riparian bird focal species life history traits, vegetation associations, regional average density estimates, and the resulting expected number of territories per 10 ha plot.

Species	Migratory status	Nest substrate	Habitat & vegetation associations	Regional average density (individuals/ha) ^a	Expected territories per 10 ha plot
Nuttall's Woodpecker (<i>Picoides nuttallii</i>)	Resident	Tree, 1° cavity	Mature riparian woodland	1.34 (1.10-1.64)	6.70 (5.50-8.20)
Ash-throated Flycatcher (<i>Myiarchus cinerascens</i>)	Resident	Tree, 2° cavity	Mature, open riparian woodland	2.14 (1.76-2.59)	10.70 (8.80-12.95)
Common Yellowthroat (<i>Geothlypis trichas</i>)	Migrant	Herb, Shrub	Dense understory and ground cover, esp. near river edges and wetlands	0.34 (0.24-0.49)	1.70 (1.20-2.45)
Spotted Towhee (<i>Pipilo maculatus</i>)	Resident	Ground	Dense understory and ground cover	5.35 (4.56-6.29)	26.75 (22.80-31.45)
Song Sparrow (<i>Melospiza melodia</i>)	Resident	Herb, Shrub	Dense understory	3.33 (2.65-4.20)	16.65 (13.25-21.00)
Black-headed Grosbeak (<i>Pheucticus melanocephalus</i>)	Migrant	Tree	Complex habitat with large trees and dense understory	0.81 (0.63, 1.04)	4.05 (3.15-5.20)

^aAverage density estimates for the Delta planning region of the Central Valley Joint Venture (Dybala et al. n.d. in press).

Figure 1. Conceptual model for the lower Cosumnes River restoration project. Boxes represent different phases of vegetation, and arrows represent the ecosystem processes (solid) or management actions (dashed) that facilitate transitions between phases. The levee breach along the Cosumnes River has enhanced floodplain connectivity and the ecosystem processes facilitating the transition from bare ground to diverse riparian vegetation, which is the phase during which riparian bird density is expected to be highest. In the absence of these ecosystem processes, horticultural restoration is essential to establishing diverse riparian vegetation, but in the presence of these ecosystem processes, the effectiveness of investing in additional horticultural restoration is uncertain.

Figure 2. Potential differences in wildlife responses to two alternative restoration treatments. A) Treatments do not differ in rate of response, but in total magnitude of wildlife response. B) Treatments differ in rate but not total magnitude of wildlife response. C) Treatments do not differ in initial rate or magnitude of response, but in persistence and variance of the response over time. In all 3 cases, we assume control plots show no change in wildlife response metric over time.

Figure 3. Hypothesized responses of breeding bird densities to the Lower Cosumnes River restoration experiment: high effort (solid), reduced effort (dashed), and control (dotted), in comparison to regional average densities (horizontal line). After 5 years, we hypothesize that after 5 years, plots in the high treatment will have reached >75% of average densities, while plots in the reduced and control treatment plots will have densities <50% and <10% of regional average densities, respectively.

Figure 4. Lower Cosumnes River Restoration experimental study area, showing restoration treatments, blocks, and plot sizes (ha). The Cosumnes River flows north to south.

Figure 1

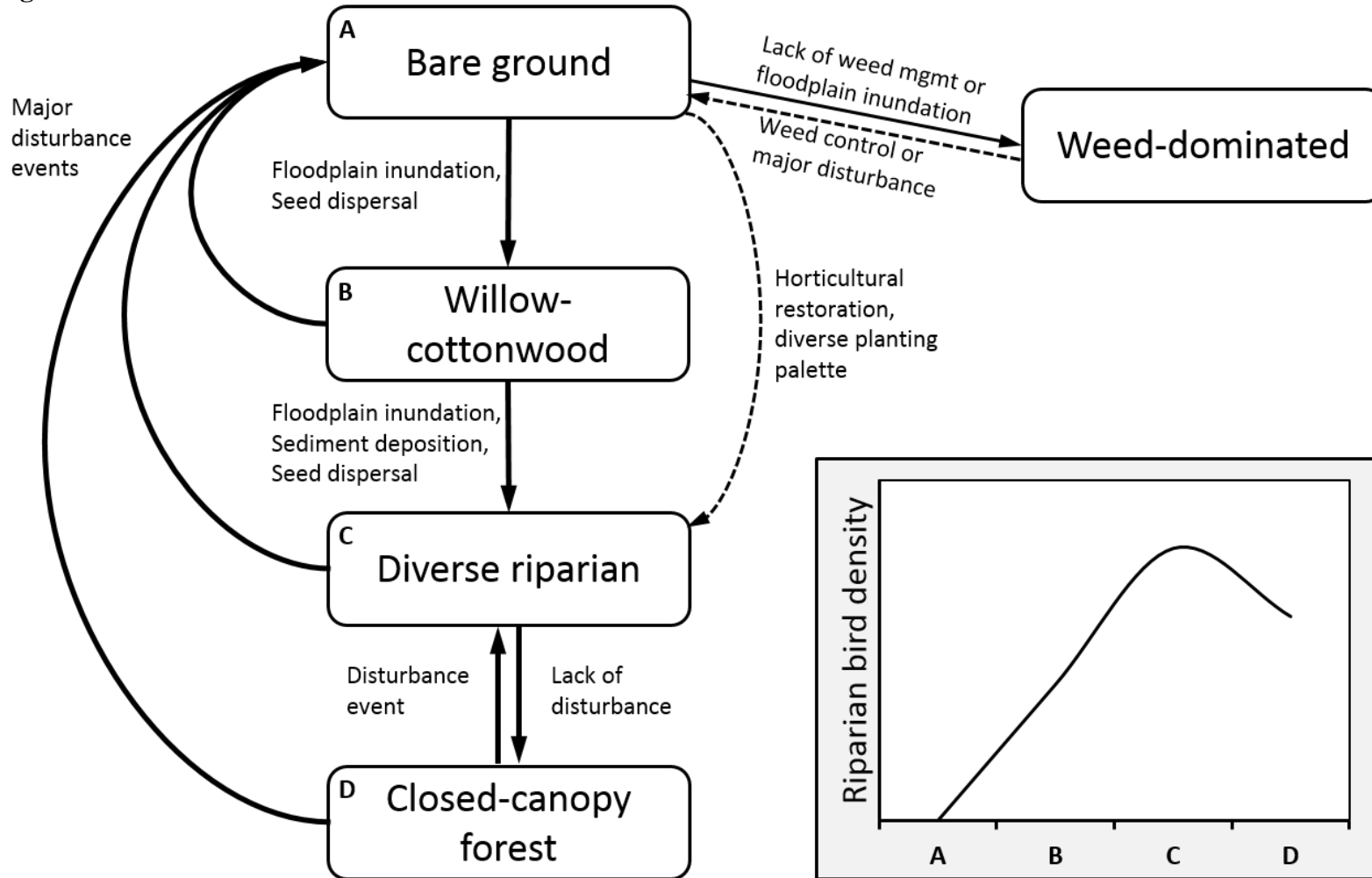


Figure 2

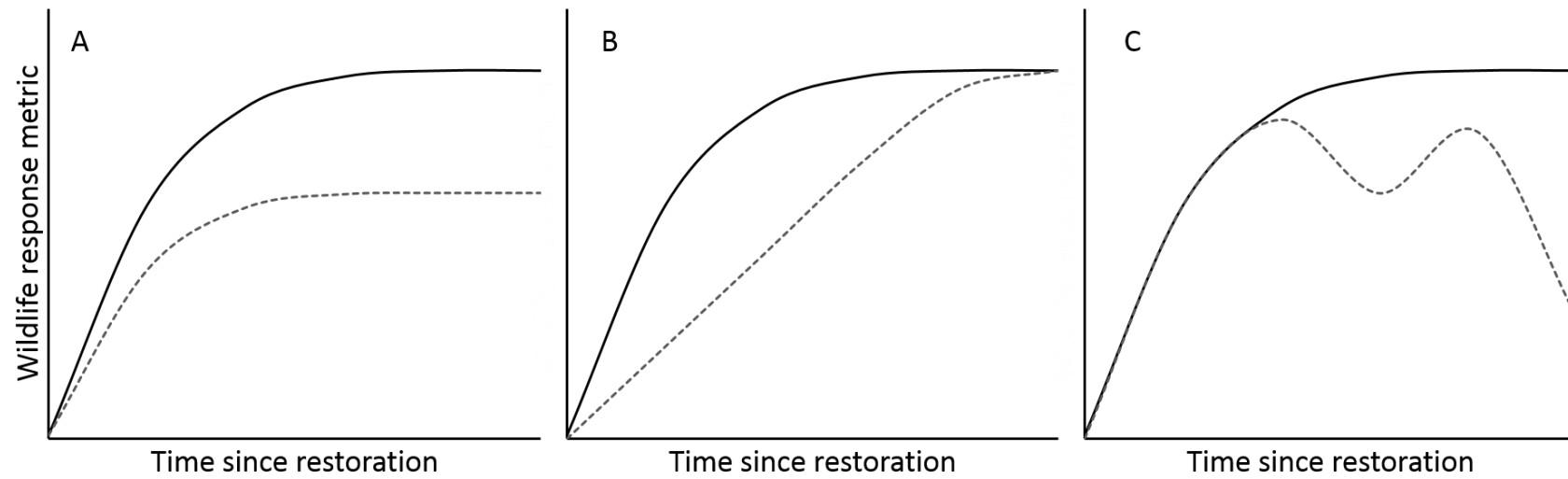


Figure 3



Figure 4

