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Author Cover Page

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Antonelli et al: Neotropical biodiversity

Abstract: The outstanding biodiversity found in the American tropics (the Neotropics) has attracted the attention of naturalists for centuries. Despite major advances in the generation of biodiversity data, many questions remain to be answered. In this review, we first summarize some of the knowns and unknowns about Neotropical biodiversity, and discuss how human impact may have drastically affected some of the patterns observed today. We then link biodiversity to landscape, and outline major advances in biogeographical research. In particular, we argue that it is crucial to test the effect of landscape and climatic evolution to biotic diversification and distribution in order to achieve a comprehensive understanding of current patterns. In this context, it is also important to consider extant and extinct taxa, as well as to use probabilistic and parametric methods that explicitly include landscape evolution models. We subsequently explore different scales in Neotropical biogeography, focusing on the intersection between biogeography and community ecology, both of which often address similar questions from different angles. The concepts of community assembly, island biogeography, neutral processes, and ecological interactions are then discussed as important components of the complex processes that determine the patterns observed today. Single-taxon and cross-taxonomic studies are complementary and greatly needed, but achieving synthesis remains challenging. Finally, we argue that phylogenetic approaches hold great potential to connect across taxonomic, spatial and temporal scales, despite current difficulties to generate and cross-analyze large volumes of molecular data. We conclude by outlining major prospects and hindrances for further advancing our knowledge on the rich Neotropical biodiversity. **Keywords:** biogeography, biotic diversification, community ecology, human impact, landscape evolution, phylogeny, scale, spatio-temporal evolution.

Page 3

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Antonelli et al: Neotropical biodiversity

I. WHAT DO WE KNOW ABOUT PATTERNS OF NEOTROPICAL BIODIVERSITY?

67 Biodiversity refers to the diversity of life across all levels of biological organization (Gaston & Spicer, 68 69 2004). The diversity of life is unevenly distributed on Earth and varies among and within geographic 70 regions, between terrestrial and aquatic ecosystems, and among different groups of organisms. 71 Biodiversity increases from the poles to the equator, reaching the highest values in tropical regions, a 72 pattern termed the latitudinal diversity gradient (Willig, Kaufman & Stevens, 2003). This pattern is 73 complex though, with numerous non-diverse tropical or diverse non-tropical areas and taxa. More 74 importantly, there are still numerous uncertainties in the underlying data and in our ability to generalize 75 overall patterns and identify their main determinants. 76 77 For many groups of organisms, the Neotropics are home to outstandingly high levels of biodiversity, when compared to other major biotic realms (Lundberg et al., 2000a; Antonelli & Sanmartín, 2011). This 78 region, extending from central Mexico to central Argentina and including the Caribbean islands 79 (Morrone, 2013), contains a vast range of biomes and habitat types, each with a particular biota and 80 81 evolutionary history (Hughes, Pennington & Antonelli, 2013) (Fig. 1). As such, understanding 82 Neotropical biodiversity patterns and the processes associated with its origin and maintenance is complex (Magurran, 2013). As result, researchers tend to focus on different aspects of biodiversity such as 83 84 taxonomic, phylogenetic, and functional diversity (Swenson, 2011). Each of these aspects of biodiversity may vary differently among regions and taxa, and each must therefore be assessed by independent criteria 85 86 (Strecker et al., 2011). 87 88 **Taxonomic diversity** 89 Taxonomic diversity refers to how many taxa can be found within a given area or higher clade, and how 90 individuals are distributed among these taxa. Taxonomic diversity can be quantified at different

taxonomic ranks (e.g., species, genera, families), with the species rank being the most popular by far.

92 Species richness – the number of distinct evolutionary lineages – is widely viewed as a fundamental 93 measure of overall biodiversity (Gotelli & Colwell, 2001). This is due to the fact that the species boundary defines the limits of genetic variation, natural selection, and adaptation (Sexton et al., 2009). 94 95 While individual organisms live and die, the stable phenotypes recognized as species may persist for 96 millions of years, serving as predictable components of the ecosystems in which all species function and 97 evolve (Eldredge, 2014). As result, species are thought to constitute the basic structural and functional 98 units in ecology and evolution (Tilman & Downing, 1996; Worm et al., 2006). 99 100 Generic and family-level taxonomic ranks are occasionally used in comparative studies, especially when 101 species identification or delimitation is difficult (Bertrand, Pleijel & Rouse, 2006). However, higher-level 102 taxa are only arbitrary constructs, reflecting little biological organization and incorporating further biases 103 and artifacts when compared. Species are thus seen as the "fundamental category of biological 104 organization" despite the multitude of species definitions available (de Queiroz, 2005). Taxonomic 105 diversity is generally measured by taxon richness, i.e., the number of taxa in a given area. However, diversity indices (e.g., Brillouin, Shannon-Wiener, and Simpson Index) that take the relative abundances 106 107 of taxa into account can also be used. These indices allow us to distinguish habitats with the same species 108 richness but different degrees of dominance. Because quantitative abundance data are so rare, few studies to date have documented patterns of diversity in the Neotropics (but see Tuomisto, 2010; Valdujo, 109 110 Carnaval & Graham, 2013; Steege et al., 2013; Jenkins et al., 2015; Moura et al., 2016; Azevedo, Valdujo 111 & C Nogueira, 2016). 112 113 For well-studied clades, relatively good estimates of taxonomic diversity are available for the Neotropics as a whole, and for each of the major biomes included therein. These estimates have been used to identify 114 115 the best predictors of diversity at large scales (Jenkins et al., 2015; Moura et al., 2016). However, the 116 observed taxonomic diversity is sensitive to sampling effort. For the Neotropics, taxonomic diversity is

117 generally underestimated, even though the level of sampling across taxa is comparable or even greater 118 than in other tropical regions (Fig. 2). 119 120 For example, in a few years of increased collection efforts, the Guide of the Ducke Reserve (Brazil) (da S 121 Ribeiro, 1999), which covers one of the most thoroughly studied areas of Amazonian forest, increased the 122 number of known vascular plants from 825 (Prance, 1990) to 2079 (Hopkins, 2005). Unless sampling is 123 thorough at a given site, species richness of any community will always be underestimated. The accuracy of estimates of taxonomic diversity depends on the number of individuals sampled, the size of the local 124 125 species pool, the size of the area, and the status of taxonomic knowledge of the groups surveyed (e.g., Tuomisto, Ruokolainen & Ruokolainen, 2012). 126 127 128 Even among Neotropical vertebrates, several examples of species-rich yet incompletely-documented 129 fauna are available, including large clades of freshwater fishes, amphibians, and some groups of reptiles. Although about 5,600 species of freshwater fishes are currently known in the Amazon, the Orinoco, and 130 adjacent river basins of tropical South and Central America, more than 100 new species are described 131 132 every year. In other words, approximately two new species are described per week, although a higher 133 number of new species would be expected if a greater amount of trained taxonomists were available. This rapid pace of species description is not slowing, and recent estimates for the total number of Neotropical 134 135 freshwater fishes exceed 8,000 species (Reis et al., 2016). This estimate is remarkable, implying that 136 more than 2,400 fish species remain to be described in the Neotropics alone, a number that exceeds the 137 combined number of rodent species currently known on Earth. This large number of expected, but still 138 hidden, lineages represents an example of the unknown unknowns of Neotropical biodiversity. 139 140 Current knowledge of taxonomic limits of Neotropical amphibians and reptiles is gradually growing. 141 Several molecular studies have shown that known diversity is underestimated in many taxonomic orders by the occurrence of high levels of cryptic diversity, i.e., the existence of two or more lineages within a 142

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Antonelli et al: Neotropical biodiversity

known species (Bickford et al., 2007; Fouquet et al., 2012) to 350% in some groups (Funk, Caminer & Ron, 2011). Even in the much more densely sampled and well-studied Atlantic Rainforest of Brazil, charismatic species of frogs are still being discovered. For instance, seven new species of *Brachycephalus* were recently described for the Atlantic Rainforest (Ribeiro et al., 2015). Likewise, intraspecific analyses of Neotropical lizards show that the occurrence of cryptic diversity is often manifested across biomes (Geurgas & Rodrigues, 2010; Domingos et al., 2014; Guarnizo et al., 2016; Domingos et al., 2017). This subdivision of broadly distributed taxa into multiple cryptic species with restricted geographic distributions increases the perception of biological diversity of a given region, as well as has numerous implications for biogeography (Werneck et al., 2012a), and conservation (Simões et al., 2014). Estimates of local taxonomic diversity can be more accurately compared among areas when based on quantitative and standardized sampling such as metrics of beta diversity, i.e., changes in species composition among sites (Tuomisto, 2010; Leprieur et al., 2011). However, this data is only available for organisms whose taxonomy is relatively well understood, such as vascular plants (e.g., trees, ferns), and some vertebrates (e.g., birds, primates). For these same organisms, we also have a general understanding about species richness gradients (e.g., Rosauer & Jetz, 2014). For most other organisms, however, little data are available, preventing accurate circumscriptions of taxa and reasonable estimates of species richness gradients (e.g., Andújar et al., 2015). Indeed, the smaller and less conspicuous the organism, the poorer the state of knowledge. For instance, very little is known about microbial and fungal diversity, and insect diversity is similarly under studied (Basset et al., 2012). Another difficulty in assessing taxonomic diversity is associated with the fact that taxa may not represent comparable units. In organisms for which we have a good understanding of distribution patterns, morphological variation, and phylogenetic relationships, more narrowly defined taxa may be recognized. On the other hand, in poorly-studied organisms, species complexes are generally circumscribed as broadly defined taxa, biasing diversity estimates. Similarly, in well-sampled areas, species are likely to be

more narrowly defined than those from poorly-known areas. As result, our understanding of taxonomic diversity and species gradients is biased by our taxonomic knowledge (Brito, 2010). Taxonomic units may also vary according to the taxonomist revising a particular group (e.g., whether a 'splitter' or a 'lumper'), and by the data and methodologies underlying taxonomic revisions and species circumscriptions. This issue becomes obvious when taxonomic treatments of the same group are produced by different researchers independently. For example, the Neotropical palm genus *Attalea* included 29 species in one monograph (Henderson & Chávez, 1995), and 65 species in another taxonomic treatment published just four years later (Glassman, 1999). Similarly, the Caribbean palm genus *Coccothrinax* included 14 species in one taxonomic treatment (Henderson, Galeano & Bernal, 1995) and 53 species in another (Dransfield *et al.*, 2008).

The commonness of rarity. Species with low abundances and narrow geographic ranges, as well as those confined to special habits, habitats or areas, represent a sizable portion of tropical diversity. Indeed, a recent study extrapolating population size for Amazonian trees suggests that most species in the region are represented by relatively few individuals (Steege et al., 2013). Another study suggests that a considerable fraction of the rare species in the region may actually have relatively large distribution ranges (Zizka et al., 2017), although many apparently widespread species known from very few scattered records may easily turn out to contain multiple biological species. However, the characteristics of these species and their contributions to diversity patterns are difficult to quantify and remain largely obscure (Coddington et al., 2009), partly because most truly rare species will be completely unknown, and partly because rareness in the ecological sense is hard to define, depending on a variety of aspects, including the species concept adopted and the taxonomic preferences.

As in others biotas, most Neotropical species are rare, narrowly distributed, and endemic to particular regions or biomes (Steege *et al.*, 2016). Plant endemism seems to be largely related to edaphic conditions, although few studies have attempted to empirically quantify factors that cause range restrictions to



particular regions (Ficetola, Mazel & Thuiller, 2017). The geographic distributions of many riverine and floodplain taxa are limited by river basin watersheds, and opportunities for dispersal via river capture events (Albert *et al.*, 2017). Climate change velocity is also thought to be associated with restricted distribution patterns. In other words, how fast a species can expand into similar climatic conditions, can affect the species' range. For instance, most vertebrates with small ranges are restricted to areas of higher climatic stability and/or mountainous areas (Sandel *et al.*, 2011). Patterns of endemism may, however, be one of the most difficult things to document given our limited sampling of biodiversity. It is not enough to know where particular species occur, we also need to know where these species do not occur (Soria-Auza & Kessler, 2008). It is, therefore, difficult to reliably say if the biodiversity patterns known to date really reflect true patterns or simple biases in collection effort.

Additional biases and gaps. For most of the Neotropics, detailed geographical distribution information is restricted to certain well-studied taxa (e.g., primates) and well-studied areas (e.g., Barro Colorado Island in Panama, the Ducke Reserve in Brazil). Interestingly, there is also a bias towards rare species, as most scientific collectors tend to over-collect rare or uncommon taxa (Steege et al., 2011). For the vast majority of groups and areas, knowledge is still scarce. For instance, many places in tropical South America have no occurrence records available (Feeley, 2015). Furthermore, particular large areas of Amazonia remain completely unexplored (Hopkins, 2005; Guedes et al., In press). In general, knowledge of species distributions and diversity patterns are strongly biased towards areas that are more easily accessible by roads, rivers, and research stations (Hopkins, 2005; Albert & Carvalho, 2011; Meyer et al., 2015). Although bioinformatic solutions may now assist in cleaning, predicting and validating species occurrence data, taxonomic expertise is still essential but limited (Maldonado et al., 2015; Töpel et al., 2016). As a result of our limited knowledge on species distributions patterns, and large gaps in knowledge about climatic and edaphic conditions for large portions of the Neotropics, the ecological requirements for species remain only roughly or even completely unknown for most taxa.



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Antonelli et al: Neotropical biodiversity

Phylogenetic diversity

Phylogenetic diversity assesses cumulative evolutionary distinctiveness within and among areas and taxa. The use of species as universally comparable units may not always be appropriate due to differences in species concepts, operational criteria of delimitation, and circumscriptions among areas, taxa, and taxonomists. In addition, species differ widely in their evolutionary ages, geographic distributions, habitat tolerances, and degree of genetic structure. Species also differ in the biological attributes of their constituent organisms, and therefore, in the effects that these traits may have on ecological and evolutionary processes. Furthermore, species are really just the tips of larger phylogenetic trees evolving through time. Some Neotropical clades are known from just one or a few species that may represent relictual survivors of ancient and extinct groups. Examples include the leaf cacti (*Pereskia* spp; Cactaceae), the South American lungfish (*Lepidosiren paradoxa*; Lepidosirenidae), the hoatzin (Opisthocomus hoazin; Opisthocomidae), and the coral pipe snake (Anilius scytale; Aniliidae). Other species are members of species-rich Neotropical clades still in the full bloom of their diversification, like the Bignoniaceae with more than 860 species (Fischer, Theisen & Lohmann, 2004), palms with over 730 species (Dransfield et al., 2008), armoured catfishes (Loricariidae) with 680 species (Nelson & Platnick, 1980a), Cactaceae with 1400 species (Hernández-Ledesma et al., 2015), and tanagers (Thraupidae) with 371 species (Burns et al., 2014). To cope with the differences in diversity among different taxa, many researchers have turned their attention to Phylogenetic Diversity (PD) indices (Faith, 1992). The basic idea of PD is to measure the total amount of lineage evolution through time found in a particular area (Faith, 1992). Overall, PD has been shown to provide a better estimate of "feature diversity" than species richness alone (Forest et al., 2007). However, there are many ways of deriving and applying such metrics from phylogenies. As such, researchers should try to choose the most appropriate index for each situation, as well as should acknowledge these differences in cross-taxonomic comparisons (Tucker et al., 2016).

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Antonelli et al: Neotropical biodiversity

The evolutionary relationships among major groups of macroscopic organisms (i.e., up to family level) are relatively well understood (e.g., Meredith et al., 2011). Recent efforts to understand the global tree of life have improved substantially our understanding of relationships among genera and species (Hinchliff et al., 2015). For instance, recent phylogenies of birds (Jetz et al., 2012), mammals (Faurby & Svenning, 2015a) and squamate reptiles (i.e., lizards and snakes) (Tonini et al., 2016) purport to include all living species. However, a substantial fraction of the species included in these studies was placed within the phylogeny solely based on morphological features due to the lack of genetic data. This is especially common for tropical species, for which genetic data is even more limited (Reddy, 2014). In contrast, other phylogenies have been built exclusively from genetic data, such as the seed plant phylogeny for ca. 32,000 species (Zanne et al., 2014) and the Neotropical tree phylogeny (Dexter & Chave, 2016). While this approach eliminates incorrect phylogenetic placements based on morphology, it creates biases given the limited genetic data available for tropical species (Antonelli et al., 2015). More detailed knowledge on evolutionary relationships is available for selected groups of vascular plants, e.g., ferns (Lehtonen, 2011), Bignoniaceae (Lohmann, 2006; Grose & Olmstead, 2007; Olmstead et al., 2009), Orchidaceae (Chase, 2003), legumes (Azani et al., 2017) and Cactaceae (Hernández-Ledesma et al., 2015). The first trials to map phylogenetic diversity over continental and global scales were conducted for selected vertebrate groups for which phylogenies were available and for which distribution patterns are relatively well known, such as amphibians, birds, and mammals (e.g., Safi et al., 2011). Other than these, large-scale phylogenetic and functional diversity studies in the Neotropics are scarce. Some progress has been made in mapping phylogenetic diversity patterns in the Neotropics for specific clades (Rossatto, 2014; Fenker et al., 2014; Bacon et al., In press) or at the intraspecific level in the search for areas of high phylogeographic diversity and endemism (Carnaval et al., 2014; Smith et al., 2017). Several ongoing studies by independent research groups are now working to broaden our knowledge on the spatial distribution of Neotropical phylogenetic diversity.

Page 11

Complementary to phylogenetic diversity based on the relationships among taxa, patterns of genetic variation within species also represent a vital but often under-appreciated component of biodiversity.

Knowledge of intraspecific genetic variation may also improve the prediction of a species ability to adapt to changing climates, as well as can improve the understanding of the determinants of speciation. This type of information is particularly important in the light of global warming. However, our current knowledge of species genetic diversity is restricted to a few selected species, and overall patterns of intraspecific genetic diversity remain poorly understood. Even among well-studied groups (e.g., mammals), spatial patterns of genetic diversity are effectively unknown within the tropics. In one study addressing this question at a broad spatial scale (Miraldo *et al.*, 2016), found higher genetic diversity at lower latitudes, and lower genetic diversity in Europe. However, no clear pattern was recovered within the tropics.

A massive increase in the availability of genetic information is being driven by high throughput sequencing technologies. This data accumulation is likely to significantly improve our understanding of genetic diversity and evolutionary relationships amongst species (e.g., Chakrabarty *et al.*, In press). Furthermore, this data will also greatly improve our understanding of largely under-studied groups, such as soil microbes (Mahé *et al.*, 2017). However, a central challenge to understanding current patterns of evolutionary diversity is the absolute dating of phylogenies, which relies heavily on fossils. Fossils are rare and poorly explored and studied in tropical areas, complicating a detailed understanding of the ages of tropical taxa, especially those from rainforests (Wing *et al.*, 2009; Lovejoy, Willis & Albert, 2011). It is therefore crucial that more efforts are put into the exploration, digitization, and analyses of the Neotropical fossil record, in addition to further exploration and sampling of living species.

Functional diversity

Functional diversity measures differences in the physiological, behavioral, and ecological characteristics of organisms, and how biological trait values affect ecological and evolutionary processes. Knowledge

about species traits and ecological functions (including the variation within and between species) is a crucial component of biodiversity. However, this is one of the major shortcomings in current biodiversity knowledge, especially in tropical areas. Few studies to date have mapped large-scale patterns of functional diversity, although efforts in this direction are underway (see for fishes e.g., Arbour & L pez-Fern ndez, 2014; Toussaint *et al.*, 2016).

Apart from the lack of data, the theory behind functional diversity is not yet consolidated. We still do not know which traits are important for different groups, how to compare traits for different sets of organisms, and how functional diversity affects forest productivity, stability, and resilience, especially in the tropics. An additional shortcoming is associated with biotic interactions. Apart from basic information on pollination and dispersal syndromes, we know surprisingly little about biotic interactions. Very few species interaction networks are available to date (see Toju *et al.*, 2017).

For the terrestrial vertebrate groups studied to date, patterns of phylogenetic diversity and species richness seem to be correlated at a global scale, generally supporting the assumption of the surrogacy of various measures (but see Pardo et al., 2016). At least for birds and mammals, global patterns of phylogenetic and functional diversity seem to be comparable (Safi *et al.*, 2011).

The human impact on biodiversity

Even through knowledge of contemporary patterns of biodiversity has improved substantially during the past years, it is still unclear whether the documented patterns are derived from natural processes or driven by human influences. Humans have occupied the Neotropics at least since the end of the Late Pleistocene and likely caused major extinctions in the diverse fauna of large mammals (Sandom *et al.*, 2014). The drastic decrease in the density and diversity of large mammals likely led to major changes in overall vegetation structure (Bond, 2005). For example, in South America, the limits between the Dry Diagonal and the adjacent forests may have shifted significantly compared to where they would have been without

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Antonelli et al: Neotropical biodiversity

any human involvement and its cascading effects (Doughty, Faurby & Svenning, 2015). In addition to anthropogenic extinctions, humans likely also caused drastic range contractions of many other species while reduced the abundance of others to the point of ecological irrelevance (Faurby & Svenning, 2015b). The human-linked reduction in Neotropical megafauna likely also affected the plants that they dispersed. This pattern was recently discussed in the context of the impact of over-hunting of primates and tapirs on the total woody biomass of Amazonia (Peres et al., 2016), and large frugivorous in the Atlantic Forest (Bello et al., 2015). Overall, it seems that the patterns observed reflect past hunting. Humans have restricted the ranges of some species, but actively or passively increased the ranges of others, such as invasives or domesticated species (Levis et al., 2017). The knowledge to date is based on the best-studied groups and still it is not clear whether substantial effects of humans will be frequent among other organisms. Clearly, we are observing just the tip of the iceberg. Apart from the effects of past human activity for the assembly of Neotropical biodiversity, current habitat loss, climate change and neglected conservation strategies also pose serious threats to natural landscapes. Indeed, these are presumably the primary drivers of the current global biodiversity crisis. Studies that quantify genetic diversity, vulnerability, and extinction risk derived from the impact of habitat loss and climate change are essential to grasp how current human activities are expected to impact the future of Neotropical diversity at multiple levels. Although we now have a fair understanding of several components of Neotropical biodiversity, for many taxonomic groups, well-defined processes remain

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II. BIOGEOGRAPHIC ADVANCES LINKING BIODIVERSITY AND LANDSCAPES

elusive and biases loom large; refining these issues will constitute an area of active scientific exploration

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Early ideas about Neotropical biogeography

for the next decade and beyond (Table 1).

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Antonelli et al: Neotropical biodiversity

The Prussian naturalist Alexander von Humboldt was among the first to realize that biotic and abiotic processes work together to constrain species distributions, and to place these influences into a geological framework. He came to this notion in the Neotropics, most famously during his study of the Chimborazo volcano in Ecuador, where he carefully documented the location of different species along elevational zones (Humboldt & Bonpland, 2010). It was in this study that he first observed that physical parameters such as topography and climate were key for floral distributions (Humboldt & Bonpland, 2010). A century later, Wegener (1912) advanced the incipient field of historical biogeography with the theory of continental drift, based in part on past geographic distributions of biotas linked by previously connected continental plates. The striking fit between the coastlines of South America and Africa was one of the pieces of evidence inspiring Wegener's theory of dynamic, non-static landmasses. In the 1960s, a geophysical mechanism for plate tectonics was proposed (Vine & Matthews, 1963; Raven & Axelrod, 1974; Rosen, 1975), placing studies of plant and freshwater fish biogeography into a plate tectonic framework. This provided historical biogeography a solid basis for further advancement. Inferring landscape evolution in the Neotropics Now, early in the 21st century, the field of historical biogeography increasingly relies on geological models that specify the landscape configurations on which species originate, disperse, and go extinct. This is especially true in the Neotropics, where understanding phylogeny and biogeography in the context of landscape evolution requires assessment of geological data, including sedimentary environments, sedimentation rates, paleontological records, and fission track ages, among others (e.g., Hoorn et al., 1995; Lundberg et al., 2000b; Figueiredo et al., 2009; Hoorn et al., 2010; Sanín et al., 2016; Jaramillo et al., 2017; Hoorn et al., 2017). Some recent reconstructions of the Neogene landscape in Amazonia are based on numerical modelling, which create reconstructions based on physical parameters such as erosion and mountain uplift. These

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Antonelli et al: Neotropical biodiversity

reconstructions are useful but often depend on arbitrary and artificial values. An example is the reconstruction of the Amazon River where Sacek, 2014 coupled different physical effects derived from the Andes uplift in a mathematical model to explain the drainage reversal in the Miocene (Sacek, 2014). However, this study ignores the synergic effects of plate movement and surface dynamics which are known to have an impact in wetland formation. Other landscape models rely on dynamic topography, in which mantle movements through time are quantified. The effects of these movements are estimated on surface subsidence and are then related to environmental and landscape changes, such as the model applied to explain the genesis of the Pebas wetland in western Amazonia (Hoorn, Wesselingh & Hovikoski, 2010) which reveals the origin of wetlands trough Amazonia. Landscape models are also extremely useful but also lack specificity. Biological data can also help improve landscape models. In recent years, integrated approaches have built landscape evolution models based on both geological and biodiversity data (Craw et al., 2016; Badgley et al., 2017). Some studies make use of geographic information systems (GIS) and combine these with well-dated palynological databases, such as Neotoma (https://www.neotomadb.org/). These models are mainly applied to reconstruct landscapes across the Quaternary timescale. For example, reconstruction of changes in connectivity across the northern Andes enabled the inference of cyclic phases of biotic dispersal and speciation versus extinction (Flantua & Hooghiemstra, 2014). Molecular phylogenetic data can also be used to statistically evaluate the likelihood of competing geological models, such as the closure of the Central American Seaway dividing South and Central America (Bacon, 2013), and the roles of the Caribbean plate margins as dispersal corridors between South and Central America (Tagliacollo et al., 2015a). Similar approaches may prove useful in several cases where geological data are insufficient or ambiguous (Baker & Couvreur, 2013).

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Effects of landscape and climatic evolution on Neotropical diversification

Given the sheer size of the Neotropical region, Neotropical biogeography and biodiversity can only properly be understood when considering the Andean uplift and the effects of this orogeny on the landscape (Fig. 3), and regional climate (Gentry, 1982; Hoorn *et al.*, 2010). The 7,000 km long Andes is

402 strategically positioned perpendicular to the principal global atmospheric currents and traps the humid air 403 of the Intertropical Convergence Zone (ITCZ). This configuration enhances precipitation along the Andean slopes and in western Amazonia, making them wetter than they would be in a low Andes setting. 404 405 Moreover, the high Andes also redirects the atmospheric flow inducing the southward deflected South 406 American Low-Level Jet (Garreaud et al., 2005; Insel, Poulsen & Ehlers, 2010; Rohrmann et al., 2016). 407 408 The situation is reversed in southern and in northwestern South America. In these regions the Andes trap 409 the humid air of the Southern Hemisphere westerlies (Garreaud et al., 2005). In contrast to the 410 Amazonian scenario, the eastern margin of the Andes at its northern and southern extremes forms a rain shadow where semi-desert conditions prevail, although this situation is partially reverse during El Niño 411 412 events. Instead, on the western flank there is increased precipitation with more humid conditions 413 (Blisniuk et al., 2005; Palazzesi, Barreda & Cuitiño, 2014). The monumental Andean barrier has thus a huge impact both on the climate and landscapes of South American lowlands, resulting in the 414 modification of river systems and drastic changes in regional climate and habitats. 415 416 The influence of South American climate on biodiversity is well illustrated in the contrasting 417 418 development of biodiversity between Amazonia and Patagonia. Over the course of the Miocene, Andean uplift led to a humidification of Amazonia and aridification of Patagonia (Blisniuk et al., 2005; Palazzesi, 419 420 Barreda & Cuitiño, 2014; Rohrmann et al., 2016). This contrast is particularly well-illustrated by the 421 history of New World monkeys (Platyrrhini), which were widely distributed in Patagonia from early to 422 middle Miocene, including the southernmost non-human primates that ever lived (Tejedor et al., 2006; Tejedor & Novo, 2017). However, those primates were later extirpated during regional aridification and 423 global cooling after the Middle Miocene. The platyrrhine record of the high Chilean Andes indicates that 424 425 the connection between Patagonia and the northern Neotropics possibly persisted on the western part of 426 South America, as the southern Andean cordillera was not an important barrier in the Middle Miocene (Flynn et al., 1995). This scenario provided primates and other animals with a migration route to the 427

428 north, facilitating faunal turnover (Tejedor & Muñoz-Saba, 2013). This connection may also have 429 contributed to the subsequent Amazonian diversification of crown platyrrhines, including some Patagonian lineages (Rosenberger et al., 2009). Three extant platyrrhine subfamilies were already present 430 in Patagonia by the early Miocene, later represented in the middle Miocene of Colombia (Tejedor & 431 432 Novo, 2017), which are diverse and widely distributed today. 433 434 The rise of the northern Andes had a major impact on Neotropical biodiversity, as documented for many taxa (e.g., Hughes & Eastwood, 2006; Santos et al., 2009; Antonelli et al., 2009; Tagliacollo et al., 435 436 2015b; Sanín et al., 2016; Chazot et al., 2016; Diazgranados & Barber, 2017). Recent studies that explicitly integrate surface uplift and climatic changes as a function of speciation and extinction include 437 438 work on the Andean bellflowers (Lagomarsino et al., 2016), Neotropical orchids (Perez-Escobar et al., In 439 press) and Neotropical hummingbirds (Condamine et al., 2017). A similar approach could be applied to many other taxa and systems, such as the recent cross-taxonomic study on the flora of the Hengduan 440 441 Mountains by Xing & Ree (2017). 442 At an intercontinental scale, the uplift of the Andes can be compared to the Eastern Arc orogeny in 443 444 Central Africa, which started during the Eocene but reached its maximum in the Pliocene (Pokorny et al., 2015). Both of these events resulted from the slow collision of two plates and contributed to the 445 446 'humidification' of Amazonia and Central Africa, respectively. However, the northeastward movement of 447 Africa brought the continent close to the Equator, closing the Thetrys Seway and precipitating an 448 aridification that remains today (Pokorny et al., 2015). Advances on climatic reconstructions via historical records and climatic modeling (Cheng et al., 2013; Wang et al., 2017) or biome paleo-distribution 449 modeling (Carnaval & Moritz, 2008; Werneck et al., 2011; 2012b; Ledo & Colli, In press) allow for 450 451 direct hypothesis testing based on independent biodiversity data. 452

Methodological approaches for estimating biogeographic histories

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454 Inferring spatial and temporal dimensions of evolution are fraught with difficulties, especially due to the 455 lack of abundant and evenly sampled biological and geological data. This is particularly critical for the Neotropics due to the region's size, limited access, extraordinary biodiversity levels, landscape 456 457 heterogeneity, and complex evolutionary and geo-climatic histories. To tackle these problems, we 458 summarize some of the main issues associated with the analyses of biogeography and diversification, 459 focusing on how those issues affect Neotropical biodiversity research. 460 Definition and use of operational units. Defining units of study in biogeography, sometimes called "areas 461 462 of endemism," is not an easy task, especially when diverse systems are involved such as the Neotropics. Sympatry or the geographic congruence among the distribution areas of taxa, is often used as a criterion 463 to define sound units for these studies. The identification of such areas has long been based on expert 464 465 opinion, with data-driven approaches that use actual species distribution data only becoming available more recently (Holt et al., 2013; Vilhena & Antonelli, 2015; Edler et al., 2016; Antonelli, 2017). These 466 approaches to bioregionalization are of great importance as they allow for more objective, reproducible 467 and informative analyses. Areas have also been defined using geologically explicit criteria, including 468 information on the geological history of landmasses or geographic barriers, both of which are not 469 470 exclusive to the group under study (Antonelli et al., 2009; Albert & Carvalho, 2011; Töpel et al., 2016 (Bacon et al., In press). Areas defined based on species distribution patterns and geological history 471 472 are of particular interest (Perret et al., 2007; Givnish et al., 2014). 473 474 The use of areas as discrete entities is useful in parametric biogeographic models where areas are considered as biogeographic traits that evolve along the phylogeny, and whose ancestral areas are inferred 475 at speciation nodes. In these models, the spatial units of analysis are defined by the biogeographic 476 477 hypothesis under examination. For example, it is possible to determine whether diversification rates have 478 been historically higher in Andean or non-Andeantaxa (Chazot et al., 2016). However, defining areas as 479 discrete entities is difficult when there are overlapping boundaries and an excess of widespread taxa.

480 Models have been proposed to objectively define areas of endemism by overlapping taxa with "fuzzy" 481 boundaries (Szumik et al., 2002; Szumik & Goloboff, 2004). Similarly, biotic element analyses have also been proposed to test for non-random distributions of species ranges (Hausdorf & Hennig, 2003). Some 482 483 of these methods have been applied to Neotropical taxa (Casagranda M, Roig-Juñet & Szumik, 2009; 484 Noguera-Urbano & Escalante, 2015; Azevedo, Valdujo & C Nogueira, 2016). 485 Another possibility is to use vicariance -- geographic barriers -- rather than areas as units of analysis 486 (Hovenkamp, 1997; Arias, Szumik & Goloboff, 2011; Arias, In press). This approach explicitly 487 488 introduces the spatial (landscape) aspect missing from the predefined areas-as-discrete entities used in parametric biogeography. Since this approach is based on taxon-defined ranges, biogeographic 489 490 reconstructions are not dependent on different area definitions (Arias, in press). A parametric version of 491 this approach allows geographic (dispersal) barriers to evolve over time within the landscape (Albert et 492 al., 2017). 493 An alternative to using discrete areas in biogeographical analyses is the spatial diffusion approach, which 494 495 conducts spatial-temporal reconstructions under random walk models within likelihood (Lemmon & 496 Lemmon, 2008) or Bayesian (Lemey et al., 2010) frameworks. This approach has been used to study taxa from open and dry Neotropical biomes (Werneck et al., 2011; 2012b; Nascimento et al., 2013; Camargo 497 498 et al., 2013), and taxa with broad continental distributions (Gehara et al., 2014). A further development of 499 this approach has been applied to the Neotropical bird genera *Psophia* and *Cinclodes* (Quintero et al., 500 2015). The method uses georeferenced point-localities to infer ancestral areas and thus does not make assumptions about species ranges and operational units that fit many taxa. On the other hand, this method 501 suffers from the common issue of ancestral lineages occupying average values of the descendant lineages. 502 503 For instance, analyses with this method have reconstructed the ancestral of Cinclodes ovenbirds to a 504 region in-between the western and eastern margins of South America, where no such species occur today (Quintero et al., 2015). 505

Considering the complex and dynamic nature of the Neotropical region across multiple temporal and spatial scales, spatial diffusion biogeographic analyses would greatly benefit from the incorporation of landscape-explicit models that allow the reconstruction of actual paths along branches (McRae *et al.*, 2008). These developments would allow the incorporation of spatial heterogeneity via dispersal constraints, derived from estimated ecological niche models or landscape evolution models, among others.

Single lineage approaches. These provide detailed reconstructions of the temporal and spatial evolution for individual clades, species, or lineages. The focus is not on generalities but on contingencies or events that are idiosyncratic to the group under study. Methodological advances in single lineage approaches have undergone major developments with parametric methodologies (Ree & Sanmartín, 2009; Landis et al., 2013; Matzke, 2014). The explosion of molecular phylogenetics and molecular dating analyses allowed inference of time-calibrated trees, where branch lengths are measured as units of time or rates of molecular evolution. The integration of time-calibrated trees into biogeographic analyses allowed the establishment of links with external sources of temporal information such as landscape evolution, geological history, fossil record, and climate history.

Typical biogeographic analyses currently use time-trees and parametric models of biogeographic evolution to reconstruct ancestral ranges at speciation nodes, and to infer rates of biogeographic processes (e.g., dispersal, speciation, and extinction). Parametric biogeographic models are based on Continuous-Time Markov chains (CTMC). These are stochastic, memoryless processes in which transition rates between discrete states (i.e., geographic ranges) are governed by an instantaneous Q matrix, with transition probabilities obtained after exponentiating the matrix as a function of time (i.e., branch lengths) (Ronquist & Sanmartín, 2011). Transition rates are defined in terms of biogeographic processes



describing the evolution of geographic ranges, such as range expansion, jump dispersal, range contraction and local extinction.

Biogeographic hypotheses or models about the relative role of these processes in a the geographic evolution of particular groups can be compared statistically using methods for model selection in phylogenetics, such as Likelihood Ratio Tests (LRT) or Bayes Factors (Sanmartín & Meseguer, 2016). Moreover, the rates of these processes may be modified (scaled) to reflect the changing connectivity among the areas of analysis over time (Ree & Smith, 2008). These advances have contributed to the integration of landscape dynamics and geological history into taxon biogeography in the Neotropics (e.g., Perret *et al.*, 2013; Givnish *et al.*, 2014; Chazot *et al.*, 2016; Bacon *et al.*, In press). Dispersal-Extinction-Cladogenesis (DEC), a likelihood-based method derived from CTMC models, is likely the most popular parametric biogeographic method. This method infers anagenetic evolution (i.e., along branch internodes) as a function of two rate parameters: range expansion (dispersal) and range contraction (local extinction). Cladogenetic evolution (i.e., at speciation nodes) is modeled as the likelihood of alternative range inheritance scenarios that describe the division of ancestral ranges into descendant nodes: sympatric speciation, allopatric (vicariance) speciation, and peripheral isolate speciation in the case of widespread ranges (Ree & Smith, 2008).

The popularity of DEC is based on the fact that, given a time tree and associated terminal distributions, it can provide detailed biogeographic reconstructions of the ancestral origin of a clade and the history of dispersal and extinction events that helped model its spatial evolution (Sanmartín & Meseguer, 2016). A potential drawback of DEC is, however, the number of areas that it can implement. Because widespread geographic ranges (comprising two or more areas) are possible states in the Q matrix, a large number of unit areas rapidly leads to computational and convergence issues. Constraining the number of states based on biological or geological criteria, is a way to decrease model complexity (Ree & Sanmartín, 2009).

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Antonelli et al: Neotropical biodiversity

Bay-Area, a data augmentation approach based on stochastic mapping and which extends the DEC model to deal with a large number of unit areas, has been proposed to tackle the limited number of areas allowed in DEC (Landis et al., 2013). Furthermore, the parameter ("J") was introducted to model "jump dispersal" or founder-event speciation (Matzke, 2014). This extra parameter effectively reduces the contribution of range expansion, and therefore of widespread range evolution, in biogeographic likelihood estimations (Matos-Maraví et al., 2013; Matzke, 2014; Ree & Sanmartin, in prep.). The DEC+J model is implemented in the package BioGeoBEARS (Matzke, 2013) implemented in R (R Development Core Team, 2017), and is now widely used in Neotropical biogeography (e.g., Matos-Maraví et al., 2014; Espeland et al., 2015; Chomicki & Renner, 2016). Cross-taxonomic (multi-clade) approaches. Unlike single taxon biogeographic approaches, crosstaxonomic approaches aim to extract generalities on the evolution of a biogeographic region or a whole biota, or generalities on the relationships among biogeographic regions or biotas, by reconstructing the history of their individual components. The focus of this approach is not on obtaining detailed reconstructions of each individual lineage (although these reconstructions inform the model), but on inferring shared biogeographic histories, such as general patterns of colonization and diversification or a common response to extinction events. This approach was traditionally known as "area biogeography" and was the focus of the cladistic biogeographic school for decades (Nelson & Platnick, 1980b; Humphries & Parenti, 1999b). The first methods used for cross-taxonomic biogeographic approaches were based on parsimony, which does not allow the integration of a temporal dimension (Crisci et al., 1991; Marshall & Liebherr, 2000; Sanmartín, 2016). Further approaches attempted to solve this issue by adopting an event-based approach to recover the relative sequence of biogeographic events (Sanmartín, 2007). However, these methods are subject to "biogeographic pseudo-congruence," when the same biogeographic pattern originates in two clades at different times and therefore, not as a result of a shared biogeographic history (Donoghue &

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Antonelli et al: Neotropical biodiversity

Moore, 2003). Ad hoc introduction of time into event-based methods allow the identification of reticulate history or the cyclical formation and disappearance of dispersal corridors and barriers. This is the case of the North Atlantic and Beringian Land Bridges in the migration of Holarctic faunas and floras (e.g., Sanmartín, Enghoff & Ronquist, 2001; Donoghue & Smith, 2004). Parametric approaches such as the biogeographic CTMC models (Ronquist & Sanmartín, 2011) offer a more powerful way to obtain generalities about patterns of dispersal and diversification in biotas, allowing us to test between alternative geological or spatial scenarios. One such example is the Bayesian Island Biogeographic (BIB) model of Sanmartín et al. (2008), which uses a hierarchical Bayesian model to infer common rates of colonization and area carrying capacities from phylogenetic and distribution data across multiple clades. Because phylogenies are co-estimated with the model (and not fixed as in DEC), and each clade is allowed to evolve under its own molecular rate, BIB can be used across different, unrelated organisms that differ in biological traits such as the age of origin or dispersal ability, but which inhabit the same set of oceanic (Sanmartín, van der Mark & Ronquist, 2008) or continental islands (Sanmartín et al., 2010). Landscape evolution models (LEMs) and biotic diversification. A potential problem with single-taxon and cross-taxonomic parametric analyses is that areas are treated as traits of organisms evolving along phylogenetic trees. Geology is often used to inform the model but does not form its core. For instance, area connectivity is often used in parametric methods to constrain or scale migration rates but not as an actual part of the model. At first, the explanatory power of vicariance biogeography was the ability to predict biogeographic distributions of individual taxa and that of whole biotas from knowledge of how landscapes changed through time (Rosen, 1978). The paradigmatic example is the geological fragmentation of the Gondwana supercontinent, and the resulting fragmentation of the resident Gondwanan biotas. The vicariance biogeography approach satisfies the scientific impulse of systematists and biogeographers for general

explanations of organismal distributions, rather than ascribing each distribution to the vagaries of

609 idiosyncratic dispersal and extinction events (Humphries & Parenti, 1999a). However, ecologists have 610 long understood dispersal to be a perennial and pervasive process influencing biogeographic distributions (Cowie & Holland, 2006), and evidence for the action of long-distance, overseas dispersal has now been 611 612 documented in the formation of many biotas worldwide (Bell et al., 2015) including those in the 613 Neotropics (Smith et al., 2014; Tagliacollo et al., 2015a; Hawlitschek, Ramírez Garrido & Glaw, 2017). 614 Similarly, palaeontologists have long understood extinction as an important driver of patterns in 615 biodiversity and biogeography (Jablonski, 2008). The challenge to vicariance biogeography as a general theory is the commonplace observation that vicariant cladogenesis (i.e., speciation) is only one of three 616 617 general macroevolutionary processes, along with dispersal and extinction (Ree & Smith, 2008). 618 619 A new generation of methods is now being developed using the power of landscape evolution models to 620 study the full panoply of evolutionary processes, at both microevolutionary (population) level (Byrne & 621 Hopper, 2008; Morlon, 2014) and macroevolutionary (interspecific) scales (e.g., Tagliacollo et al., 2015b; Badgley et al., 2017). For example, uplift of a dissected landscape and river capture are two landscape 622 evolution processes with great power to generate high species richness. Both of these processes 623 624 simultaneously and continuously merge and separate portions of adjacent landscape areas, allowing biotic 625 dispersal and larger geographic ranges, vicariant speciation and smaller geographic ranges, and extinction when range sizes are subdivided below a minimum persistence threshold (Albert et al., 2017). 626 627 628 Integrating fossils into biogeography. One important shortcoming of molecular-based biogeographic 629 analyses in general, and parametric models of range evolution in particular, is the fact that it is almost always based on extant data alone. Because of the effects of extinction, the pattern of geographic 630 distribution we observe today may be a poor representation of the actual biogeographic history, especially 631 632 if extinction rates have been unequal among areas (Meseguer et al., 2015) and taxa (Silvestro et al., 633 2016). One way to solve this is to include extinct lineages in biogeographic analyses (Mao et al., 2012), or to use their distribution (the fossil record) to constrain inferences of ancestral ranges (Meseguer et al., 634

2015). This has in many cases revealed a new, different biogeographic history for the study group (Mao *et al.*, 2012; Meseguer *et al.*, 2015). An alternative approach is a modification of DEC, termed DES (Dispersal-Extinction-Sampling), to infer rates of dispersal and area extinction exclusively from fossil data (Silvestro *et al.*, 2014; 2015). An additional sampling parameter is used to account for the unevenness of the fossil record both spatially and temporally. Advantages of this approach are that the fossil record provides a more accurate measuring of changes in rates of geographic evolution and unbiased extinction rates, than if only extant taxa are used (Silvestro *et al.*, 2015).

III. FROM LARGE TO FINE SCALES: BIOGEOGRAPHY MEETS COMMUNITY ECOLOGY

While biogeographic studies are key to identify large-scale patterns and processes, different processes operate at regional and local levels. A major question in tropical biodiversity is how ecological communities have been assembled over time and how abiotic factors and species' interactions have influenced this process. Approaches for the study of tropical communities have employed a wide range of models with diverse conceptual roots. Over the last 20 years, there has been an expansion from studies focusing on contemporary community structure and spatial patterns of physical properties of ecosystems, to studies focusing on historical aspects of community structure and evolution (Leite & Rogers, 2013; Smith *et al.*, 2017). Early approaches (e.g., Margalef, 2015) focused on indexes of diversity, descriptions of community membership, as well as flow charts of energy and nutrients through the community. Key variables influencing community structure consistently emphasized classic Hutchinsonian processes such as resource use, competition, and niche partitioning. However, as ecologists adopted new techniques, the resolution of the niche increased from simple variables exclusively to also include high-resolution data on climate, soil chemistry, microbiomes and other physical and biological properties. These approaches have guided several recent analyses of tropical groups, including micro-organisms and palms (Costa *et al.*, 2009a; Mendes *et al.*, 2015). Additionally, the availability of spatially explicit online global datasets of

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Antonelli et al: Neotropical biodiversity

climate and environmental parameters has helped spawn a generation of studies using large-scale spatial biodiversity surveys and inventories, analyzed with statistical approaches, allowing the interpolation between sampled sites and estimated diversity in unsampled areas (Costa et al., 2007; 2009b; Steege et al., 2010; 2013). Such studies yield important data for interpreting community processes. However, progress in this area is often based on the important assumption that ecosystems and community structures are at equilibrium, or nearly so. Analyzing such data sets under non-equilibrium assumptions is an important challenge for the future. The theory of island biogeography (TIB) (MacArthur & Wilson, 2016) introduced parameters such as colonization, immigration, and extinction within a mathematical framework, allowing the prediction of community structure, dynamics, and diversity (Losos & Ricklefs, 2009; Warren et al., 2015). The TIB has inspired models that attempted to integrate additional parameters, such as speciation and island ontogeny (Whittaker, Triantis & Ladle, 2008), abundance (Rosindell & Harmon, 2013), and trophic interactions (Gravel et al., 2011). Whereas the TIB maintained a focus on species as the unit of analyses, another strand of theory introduced individual-based models that assumed ecological or functional equivalence of individuals in communities inspired by Hubbell's neutral theory of biodiversity (NTB) (Hubbell, 1997). In the ecological sense, both the TIB and NTB are not-equilibrium models, however they are often invoked as equilibrium models in macroevolutionary perspectives. Their predictions are based on conditions of systems states (i.e., values of parameters such as dispersal, colonization, speciation, extinction, and the strength of species interactions) in which all competing influences are balanced (dynamic equilibrium), or have not changed over time (static equilibrium). Alternatively, other models have dynamic equilibria in which the parameter values balance one another,

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Alternatively, other models have dynamic equilibria in which the parameter values balance one another, giving a constant result, such as the number of species. In the last 15 years, community ecology, and an expanded TIB, have adopted a more historical approach by integrating phylogenetic data to the study of community assembly and dynamics, including the role of *in situ* adaptation or speciation versus dispersal

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Antonelli et al: Neotropical biodiversity

in community assembly, the temporal sequence of species interactions, or the role of abiotic and biotic factors in diversification of specific lineages (Webb et al., 2003; Sanmartín, van der Mark & Ronquist, 2008; Valente, Etienne & Phillimore, 2014; Valente, Phillimore & Etienne, 2015; Cabral, Valente & Hartig, 2017). By adopting a more historical focus, these methods get around the equilibrium assumption by explicitly attempting to reconstruct the sequence of events leading to modern-day communities. These approaches relax the assumption of ecological neutrality, and focus on the uniqueness of individual lineages, historical contingency, and particularities of present-day outcomes (Emerson & Kolm, 2005; Sanmartín, van der Mark & Ronquist, 2008). In their most recent forms, these models incorporate ecological parameters such as competition and species interactions (Clarke, Thomas & Freckleton, 2016) or landscape dynamics (Aguilée, Claessen & Lambert, 2013). The diversity of theoretical approaches has enriched the field of tropical biology, particularly in the Neotropics. Below we provide some examples of how different approaches to community ecology have been applied to (Neo)tropical systems. Island biogeography studies. The Amazon basin is highly heterogeneous although this heterogeneity is rarely structured in ways that are amenable to the application of island biogeography theory. However, 'white-sand' habitats in the interior of the Amazon seem to represent 'islands' with savanna-like vegetation and distinctive plants and animals, that often achieve differentiation or endemism due to their isolation and environmental uniqueness compared to the surrounding lowland rainforest (Anderson, 1981; Alonso, Metz & Fine, 2013). Although a similar situation is found for the fragmented and understudied Amazonian savannas (de Carvalho & Mustin, 2017), open areas are characterized by lower richness for some taxa (e.g., amphibians and reptiles) that may harbor species with restricted distributions missing

from adjacent Amazonian mainland forests (Borges et al., 2016). These islands are known as "campinas"

and are the focus of active research programs (see Fine & Bruna, 2016). Bird diversity in these white

sands islands has been shown to conform well to the predictions of TIB, with larger and more connected

713 islands including a higher number of species than smaller and more isolated patches (Oren, 1982). 714 Furthermore, assembly studies at the regional scale have shown that lizard local richness is explained by patch size, degree of isolation, and stochastic extinction following isolation (Gainsbury & Colli, 2009). 715 716 However, Alonso et al. (2013) and Borges et al. (2016) suggested that more regional effects, perhaps 717 overshadowing patch size, also seem to be at play. For instance, patches south of the Marañón/Amazon 718 region seem to be depauperate when compared to northerly patches (Alonso, Metz & Fine, 2013; Borges 719 et al., 2016). Nonetheless, white sands provide a compelling example of island biogeography theory in 720 the context of a continental biota. 721 Niche-based studies. The general idea that species are adapted to their environment (i.e., have different 722 723 niches) has two important consequences. First, species distributions are expected to reflect the distribution of suitable habitats. Second, species composition in local communities should reflect the environmental 724 characteristics of the site, as unsuitable environmental characteristics or biotic interactions make it 725 726 impossible for a species to establish and/or survive. Along these lines, many studies have aimed to characterize the edaphic associations of tropical plant species (Tuomisto & Poulsen, 1996; Tuomisto et 727 al., 2003; Phillips et al., 2003; Costa, Magnusson & Luizao, 2005; Roncal, 2006; Zuquim et al., 2009; 728 729 Kristiansen et al., 2012; Cámara Leret et al., 2017) and the elevational ranges of many taxa (Kluge, Bach & Kessler, 2008). If there are more species adapted to some environmental conditions than others and 730 731 dispersal limitations are unimportant, a species richness gradient should result. However, it is also 732 possible that some environmental conditions may allow more species to coexist than others. Several 733 studies have analyzed species richness gradients along environmental gradients such as elevation (Kluge, Kessler & Dunn, 2006; Brehm, Colwell & Kluge, 2007), rainfall (Clinebell et al., 1995; Esquivel 734 Muelbert et al., 2017), and soil fertility (Costa, Magnusson & Luizao, 2005; Steege et al., 2006; 735 736 Tuomisto, Zuquim & Cárdenas, 2014). In general, these studies have shown that Neotropical species 737 richness tends to be highest in warm, humid, and aseasonal environmental niches at low to middle elevations. 738

Neutral and non-neutral perspectives. In contrast to niche-based processes, spatial patterns in abundance in communities of anurans from Central Amazonia have been shown to follow Hubbell's neutral biodiversity theory (Diniz-Filho et al., 2011). However, Manceau et al. (2015) demonstrated that, the incorporation of population genetic dynamics into NBT supported the hypothesis that biodiversity dynamics are out of equilibrium, and that these types of models can predict macroevolutionary patterns (Manceau, Lambert & Morlon, 2015). Additional research is still needed to assess the relative roles of niche constraints, neutral, and non-neutral processes in explaining and predicting Neotropical biodiversity.

Ecological interactions. It is now evident that species interactions (mutualistic or antagonistic) play a major role in the spatial distribution of taxa, the evolution of communities (Wiens, 2011), and species boundaries (Pigot & Tobias, 2013). Biotic interactions can be a source of ecological divergent selection, with interspecific competition representing a major contributor to ecological character displacement (Rundle & Nosil, 2005). In turn, this suggests that competition may play an important role in shaping ecological speciation at meta-community scale. Geography can also affect gene flow among populations, and the ecological sources of divergent selection (Räsänen et al., 2012). Large-scale biogeographical studies based on interspecies interactions are still lacking, reflecting a general paucity of interaction data as well as difficulties in estimating past interactions and processes. However, speciation resulting from species interactions may represent a main driver of biotic radiations (Elias et al., 2012; Correa Restrepo et al., 2016). Such a model seems to apply to both plants (Xu, Schl ter & Schiestl, 2012) and animals, such as the South American opossums, where biotic interactions may have led to allopatric speciation (Gutiérrez, Boria & Anderson, 2014).

Exploring ecological interactions among species can help improve our understanding of the evolution of biotic associations. For arboreal ants (*Pseudomyrmex*) and their host plant *Vachellia* (Mimosoidae),

765 historical associations and new colonizations (*Platymiscium, Tachigali*, and *Vachellia*) have been 766 documented (Chomicki, Ward & Renner, 2015). This study highlights how new interactions, formed by a group of ants colonizing a new plant, can contribute to an interaction shift (from parasitic to mutualistic). 767 768 The macroevolutionary assembly of ant-plant symbiosis in the Neotropics seems to be highly dynamic 769 and underlines convergent evolution of complex multispecies interactions. 770 771 The integration of phylogenetic inference coupled with network theory has shed light on the importance of interaction dependence (mutual dependence vs. asymmetric dependence) in the radiation of interacting 772 773 species. For instance, Ramirez et al. (2011) unraveled the evolutionary processes involved in the specialized association between Neotropical stingless bees (Euglossini) and Euglossinni-pollinated 774 Orchids using this approach (Ramírez et al., 2011). More specifically, through a combination of 775 phylogenetic inference, network analysis, and chemical data, this study revealed that selection on orchids 776 777 for their specialized pollinators triggered their radiation, whereas a similar radiation was not observed for the bees. 778 779 Phylogenetic approaches. Community phylogenetic approaches have been used to test hypotheses 780 781 involving multiple historical and ecological factors controlling phylogenetic diversity over time. Kissling et al., (2011) showed that global diversity in palms (Arecaceae) has strong phylogenetic clustering on 782 783 islands and in the Neotropics (Kissling et al., 2012). Recently, a "Historical Assembly Analysis" was 784 proposed (Weeks, Claramunt & Cracraft, 2016) as a conceptual framework for integrating evolutionary 785 history and ecological processes into studies of biotic assembly. Using this approach, the assembly 786 history of Amazonian Pteroglossus toucans was reconstructed through time and space using a combination of phylogenetic and biogeographical tools (Weeks, Claramunt & Cracraft, 2016). 787 788 789 Thus far, most community phylogenetics approaches have been applied to single lineages, within which 790 competition and other ecological processes are thought to dominate. Because it is unlikely that ecological

and community processes operate within individual lineages, cross-taxonomic community assembly analyses would be especially relevant. For instance, important insights were gained from a study on the phylogenetic assembly of Amazonian tree communities within a comparative framework (Dexter *et al.*, 2017).

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Scaling up community ecology approaches

The original goals of community ecology, as established in the early 20th century, were to predict species distributions and abundances, species richness and equitability, community productivity, food web structure, predator-prey dynamics, succession, and community assembly. However, as noted by leaders in this field, this discipline has not yet succeeded in meeting most of these goals (Ricklefs, 2008; Ritchie, 2009; Vellend, 2010; Ricklefs & Jenkins, 2011; Weber & Strauss, 2016). The reasons are many, but may be especially associated to the non-equilibrium of most local assemblages, in which the effects of historical contingencies of dispersal, extirpations, and other stochastic processes override the equilibrium expectations generated by local functional processes such as predation and competition (Fig. 4). In other words, the species composition and equitability of most local assemblages are more strongly governed by regional and historical factors than by local ecological interactions (Mittelbach & Schemske, 2015; Manceau, Lambert & Morlon, 2015; Fukami, 2015; Weeks, Claramunt & Cracraft, 2016). This crisis in community ecology has fueled the rise of alternative functionally-neutral theories, like the TIB (MacArthur & Wilson, 2016), the NTB (Hubbell, 2011), and the metacommunity theory (Leibold et al., 2004). However, neutral theories have been criticized for their simplistic assumptions and lack of predictive power under the non-neutral conditions frequently observed in nature (e.g., McGill et al., 2006). In general, the field of community ecology appears to be ripe for a paradigm shift (DeAngelis & Grimm, 2017).

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While many studies conducted at the global scale aim to test broad hypotheses about drivers of biodiversity gradients (Fine, 2015), others rely on analyses of region-wide field data collected over

decades. For example, Amazonian tree diversity catalogued across 1170 forest plots has been used to estimate tree populations across one-degree cells (Steege *et al.*, 2013). In addition to documenting hyperdominance of Amazon tree communities by only 227 species, tree diversity was also shown to be highly heterogeneous across the Amazonian landscape and at local scales (Steege *et al.*, 2013). These surveys set the stage for analyses of the environmental correlates of diversity (e.g., Benavides *et al.*, 2005; Steege *et al.*, 2013). Detailed explanations of the heterogeneity found at multiple scales remains as a major challenge for Neotropical biodiversity research.

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IV. PHYLOGENY (OUR PRIMARY EVOLUTIONARY TOOL), AND ITS CAVEATS

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The power of phylogenies to inform evolutionary processes is large and not fully realized (Eiserhardt et al., 2011). Phylogenetic data from different organisms are commonly applied in community and evolutionary analyses (Forest et al., 2007; Rull, 2008; Verboom et al., 2009; Pokorny et al., 2015; Bacon et al., 2015). For community analyses, researchers often rely on smaller scale phylogenies (rather than one 'super tree'), due to denser taxon sampling and more appropriate computational models. An inherent challenge of this approach is that different trees are often based on different priors and models, making it difficult to directly compare them, especially in terms of the distribution of node heights (ages). Several approaches have been developped to address this issue, incuding Supersmart (Antonelli et al., 2016) and msBayes (Hickerson, Stahl & Takebayashi, 2007). The Supersmart approach reconstruct large calibratedspecies phylogenies in a comparable tree framework by allowing the assemblage of vast molecular data with fossil data in three major steps. First, a backbone calibrated-tree is built using sequences from known species and fossils. Then, the backbone tree is partitioned into subclades which in a second step, descendent taxa with well coverage of genetic markers are integrated to infer a calibrated tree under an appropriate model, such as the multispecies-multilocus coalescent model (Edwards, 2009). Third, the resulting species-calibrated trees are implanted to the backbone tree. The msBayes approach, on the other hand, employs a hierarchical Approximate Bayesian Computation algorithm (hABC) to estimate

individual phylogenies and shared patterns such as the proportion of simultaneous change in a community, or the time of divergence among co-distributed species pairs across a barrier. This framework has been successfully applied to study the Neotropical biota (Carnaval *et al.*, 2009; Werneck *et al.*, 2012a; Thomé *et al.*, 2016; Gehara *et al.*, in press). Furthermore, the BIB model (Sanmartín, van der Mark & Ronquist, 2008) also draws generalities on the evolution of communities by taking the idiosyncrasies of individual clades into account.

Many large phylogenies were published in recent years at higher taxonomic levels, ranging from phylogenies of complete classes (Jetz *et al.*, 2012; Faurby & Svenning, 2015a; Figueroa *et al.*, 2016) to attempt to build the complete Tree of Life (Hinchliff *et al.*, 2015). Other studies have focused on reconstructing the phylogeny of complete regions such as the Cape of South Africa (Forest *et al.*, 2007), the Californian Floristic Province (Baldwin *et al.*, 2017), the Australian flora (Thornhill *et al.*, 2016). In contrast, we lack well-sampled phylogenies for most Neotropical groups. In addition, many researchers have expressed their justified concerns about too simplistic assumptions and problematic data downloaded from public databases. A common challenge is the compromise between taxon sampling, character sampling (i.e., the number of loci; Edwards, 2009), and computational capacity. Additionally, branch lengths and node-ages in 'super trees' are often estimated secondarily, rather than being coestimated with the phylogeny through the application of appropriate clock and birth-death models. A recently developed analytical platform allows the estimation of large phylogenies through a multi-step process (Antonelli *et al.*, 2016).

From single to many loci. Sequences of only a few loci are available for most Neotropical taxa sequenced to date. Even when multiple loci are available, these loci are often concatenated to enable more time-efficient analyses. Using single locus data to estimate phylogenies is problematic for multiple reasons (e.g., Maddison, 1997; Edwards, 2009; Liu et al., 2015), especially because a single line of evidence is provided by that individual genealogy. Analyzing multilocus sequence data requires more complex

869 computational models that are able to account for expected differences in gene tree topologies under the 870 same species tree. 871 872 Bayesian Multispecies Coalescent (MSC) methods jointly infer topology and node-heights for gene trees 873 and the underlying species tree from multilocus sequence data (Liu et al., 2009; Heled & Drummond, 874 2010; (Xu & Yang, 2016) while taking the stochastic variation of gene genealogies into account. Even though theoretically appropriate, it is often not computationally feasible to co-estimate the species tree 875 876 and all genealogies, particularly when the number of taxa and loci are high. Instead, many researchers 877 apply summary coalescent methods that split the estimation of gene trees as well as split of the species tree into two consecutive steps. Under this approach, gene trees are estimated separately for each locus 878 879 and then used in separate analyses that estimate the most likely species trees (Kubatko, Carstens & 880 Knowles, 2009; Liu et al., 2009; Mossel & Roch, 2010; Larget et al., 2010; Mirarab et al., 2014). 881 Computation limitations still prevent us from estimating multilocus trees using Bayesian MSC methods. 882 There is high demand for removing this computational bottleneck, which would provide an excellent data 883 884 source for many downstream analyses. Recent improvements of existing Bayesian MSC methods with 885 more streamlined tree searching algorithms are a step in the right direction (e.g., STACEY; Jones, 2017). Nonetheless, species divergence times based on the molecular clock model infers the original population 886 887 split rather than the last stage of the speciation process, which may not have an effect on tree topologies, but has implications for diversification analyses. In addition, using such methods for molecular species 888 889 delimitation are not exempt of controversy, as multispecies coalescence has been proposed to delimit 890 populations rather than species (Sukumaran & Knowles, 2017). 891 892 Another promising approach is SDVQuartets, which does not require a priori inference of individual gene 893 trees, but uses mathematical shortcuts (singular value decomposition of the matrix of site pattern frequencies) to infer a species tree under the coalescent framework (Chifman & Kubatko, 2014; Kubatko 894

& Chifman, 2015b). This method has been expanded to incorporate horizontal gene transfer or hybridization (Kubatko & Chifman, 2015a).

Finally, a different family of methods use gene birth-death (BD) processes to model the evolution of gene trees within species trees. These models can incorporate events such as gene paralogy (duplication) and gene loss (Boussau *et al.*, 2013) or horizontal gene transfer HGT (Szöllősi *et al.*, 2013), or a combination of both (Szöllősi *et al.*, 2015). Coalescent methods of phylogenetic analysis are only slowly making inroads into Neotropical biology, despite the demonstrated superiority over concatenation in handling large-scale multilocus data (Liu et al., 2009).

V. CONCLUSIONS AND PROSPECTS

Are we living in a unique time?

A long-standing question is the time of origin of the outstanding biodiversity we encounter today in the Neotropics. The answer to this question is inevitably linked to the search of so-called 'special periods of time' and 'overarching theories' for Neotropical diversification. The present day era likely is distinct from previous time periods, as manifested in the myriad geological and ecological footprints of the Anthropocene. However, there are often mixed definitions and questions related to the timing and mode of biotic evolution. Indeed, the origin of the Neotropical biodiversity encapsulates two contrasting subjects, the timing of origin of the hyperdiversity and the actual age of extant species (Hoorn *et al.*, 2011). It is clear there have been extraordinary periods of time throughout the geological history, both in terms of biodiversity and abiotic aspects (i.e., geology, climate) (e.g., Jaramillo, 2006; Hoorn *et al.*, 2010; Jaramillo *et al.*, 2010). However, all periods of time have contributed to the current biodiversity, and it seems unlikely that all species have entered the scene in a very 'special' time for Neotropical diversification. Instead, current diversity has deep origins in geological time, with different events (e.g., Neogene rise of the northern Andes, formation of the modern Amazon drainage system, rise of the

921 Isthmus of Panama, and Cenozoic cooling culminating in Pleistocene glacial-interglacial cycles) playing 922 different roles at different times. 923 924 Examples of meta-analyses that seek for 'special' periods of time often come from time-calibrated 925 molecular phylogenies. For instance, butterfly species-pairs seem to be relatively young in origin (i.e., < 2 Ma), suggesting that the Pleistocene and Holocene may have represented 'extraordinary times' for 926 927 Neotropical butterfly speciation (Garzón-Orduña, Benetti Longhini & Brower, 2014). However, time-928 calibrated phylogenies may not fully address the potential impact of extinction and species duration 929 (Hoorn et al., 2011). In other words, if we were able to travel back in time to any period and sequence species around us, the odds are that most species alive would also be around 2 Ma old. This potential 930 931 intrinsic bias of these types of studies remains as a challenge to be addressed in the future. 932 933 Incorporating fossil taxa in molecular diversification analyses could help resolve those issues. However, the fossil record is still limited for most Neotropical clades. In simulated phylogenies, the resulting shape 934 of lineage-through-time plots vary significantly when the fossil record is added as compared to 935 936 phylogenies that incorporate extant taxa exclusively (Matos-Maraví et al., 2014; Xing et al., 2014). The 937 inferred macroevolutionary dynamics estimated from molecular phylogenies may thus be misleading if 938 fossil taxa are neglected, or when macroevolutionary tools do not acknowledge the rare sampling of fossil 939 lineages. New methodological developments to directly integrate fossil (extinct) lineages into phylogeny 940 reconstruction (Ronquist et al., 2012; Heath, Huelsenbeck & Stadler, 2014; Zhang et al., 2016; Silvestro 941 et al., 2016) offer new hope in the quest to retrieve more accurate depictions of evolutionary patterns. 942 Variable species concepts and adequate sampling of extant and extinct taxa, represent a serious barrier for 943 944 our understanding of Neotropical biodiversity. Highly structured populations with considerable genetic divergences may be seen as "incipient species" that have not yet completed the speciation process. For 945 946 instance, excluding "incipient species" (i.e., highly structured populations that may not have yet

completed the speciation process) from phylogenies may incorporate artifacts that may lead to incorrect inferences of a diversification slowdown towards the present (Cusimano & Renner, 2010). Standardized species definitions and broad sampling of taxa are crucial while inferring macroevolutionary dynamics. Indeed, it is very likely that several broadly distributed Neotropical species may be composed of multiple narrowly distributed taxa once studied in further detail.

Operational hindrances

Comparative biology has experienced advancements in the theory and practice of biogeography and molecular phylogenetics during the past decades. However, we still need to increase sampling of Neotropical organisms drastically in order to advance our knowledge on the patterns and processes underlying Neotropical biodiversity. Despite this great need, fieldwork in the Neotropics, especially in pristine areas, is time consuming, expensive, and logistically demanding. Research funding for exploratory inventory projects is also becoming increasingly harder to obtain, despite the fact that highly successful projects (i.e., sequencing the first human genome and creating the Amazon Tree Diversity Network) were initially discovery-driven, rather than focused on testing specific hypotheses. Furthermore, obtaining permits to collect and export biological samples is also challenging, involving many differences across national legislations. Finally, fieldwork might be seen as unnecessary given the wealth of specimens already available in biological collections around the world, and their potential for biodiversity and genomic analyses (Buerki & Baker, 2016; Zedane et al., 2016).

Despite all these obstacles, fieldwork remains absolutely essential for data generation and monitoring biodiversity changes (Albert, 2002). Fieldwork also provides students and researchers with a deeper understanding and inspiration of their study systems (often providing new ideas and questions), while facilitates the establishment of new collaborations, enables the exchange of knowledge, fuels the development of new methods, and increases the possibilities of major discoveries (Fleischner *et al.*).

973 2017). We should seriously consider new strategies for the generation of new biodiversity data, as well as 974 for the syntheses of the already available data. Multi-taxon field campaigns could provide unique opportunities for intensive sampling, while optimizing resources, bureaucratic and logistic efforts. 975 976 Investments on these activities would be mutually beneficial and worthwhile from a global perspective. 977 The future of Neotropical biodiversity research depends on extensive collaborations and coordinated 978 efforts (Baker et al., 2017). 979 980 REFERENCES 981 982 Aguilée R, Claessen D, Lambert A 2013. Adaptive radiation driven by the interplay of eco-evolutionary 983 and landscape dynamics. Evolution 67:1291–1306. DOI: 10.1111/evo.12008. Albert J 2002. Eternal vigilance on an Amazon floodplain. Current Biology 12:R442-R443. DOI: 984 985 10.1016/S0960-9822(02)00937-5. Albert J. Carvalho TP 2011. Neogene Assembly of Modern Faunas. In: Historical Biogeography of 986 987 Neotropical Freshwater Fishes. University of California Press, 118–136. DOI: 10.1525/california/9780520268685.003.0007. 988 Albert J, Schoolmaster DR, Tagliacollo V, Duke-Sylvester SM 2017. Barrier Displacement on a Neutral 989 990 Landscape: Toward a Theory of Continental Biogeography. Systematic Biology 66:167–182. DOI: 991 10.1093/sysbio/syw080. 992 Alonso JÁ, Metz MR, Fine PVA 2013. Habitat Specialization by Birds in Western Amazonian White-993 sand Forests. *Biotropica* 45:365–372. DOI: 10.1111/btp.12020. Anderson AB 1981. White-Sand Vegetation of Brazilian Amazonia. Biotropica 13:199. 994 995 Andújar C, Arribas P, Ruzicka F, Crampton-Platt A, Timmermans MJTN, Vogler AP 2015. Phylogenetic 996 community ecology of soil biodiversity using mitochondrial metagenomics. Molecular Ecology 997 24:3603-3617. DOI: 10.1111/mec.13195.

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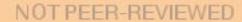
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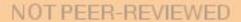


1945	AUTHOR CONTRIBUTIONS
1946	A.A. and S.E. initiated and led the project; A.Z. and S.F. compiled the data and carried out the analyses
1947	needed for producing Fig. 2; M.A. formatted references and merged the manuscript revisions; all authors
1948	participated in the discussions held under the 'Origin of Biodiversity' Workshop in Göteborg (Sweden) in
1949	the spring of 2017, wrote the paper jointly, read and approved the final version submitted for publication.
1950	
1951	
1952	CONFLICT OF INTERESTS
1953	The authors declare no conflict of interests.
1954	
1955	
1956	

1957 FIGURES



Fig. 1 The Neotropical region and examples of landscapes and taxa. Map generated through the remote-sensing ESA GlobCover 2009 project and colored by biome assignments (© ESA 2010 and UCLouvain; http://due.esrin.esa.int/page_globcover.php). a) Eastern slopes of the Bolivian Andes, where the Amazonian and Andean biotas meet; b) Patagonian mountains of southern Chile, which despite being in the temperate zone of South America is home to many Neotropical-derived lineages; c) Iguazu waterfalls, where increased humidity create gallery forests within the Open Diagonal domain; d) southern grasslands of the Pampas, a naturally open habitat now largely influenced by human activity; e) one of the ca. 338 known species of hummingbirds, a conspicuous clade currently restricted to the American continent and particularly diverse in the Andes; f) *Epidendrum ibaguense*, a widespread species in the orchid family in which many new Neotropical species are discovered each year; g) an unidentified fly in





Antonelli et al: Neotropical biodiversity

the inselbergs of southern French Guiana, where basaltic rocks emerge several hundred meters above the
surrounding Amazonian rainforest: h) <i>Hydrolycus scomberoides</i> , exemplifying the world's richest
ichthyofauna in the Amazon drainage basin; i) Ameerega flavopicta, a rock-dwelling frog species adapted
to a region of high seasonality of precipitation; j) a columnar cactus of central Mexico, near the
northwestern limits of the Neotropical region where low-canopy forests and succulent vegetation build
vegetation mosaics across the landscape. [Photo credits: a-g, i-j: A.A.; h: J.A.].

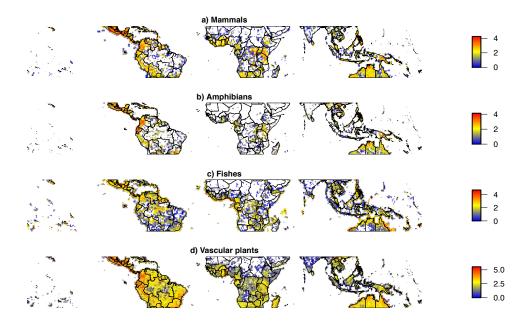
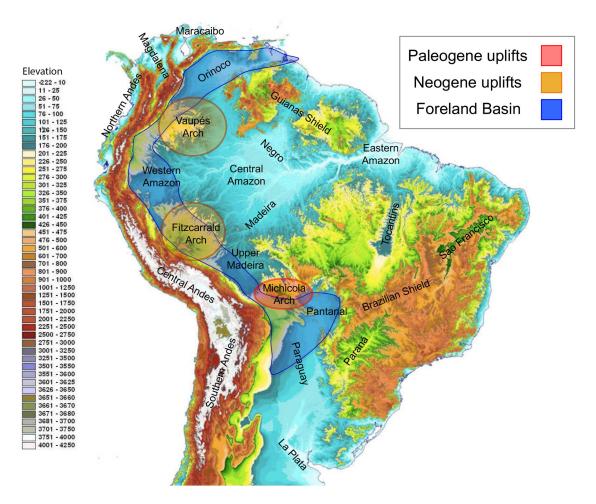


Fig. 2 Taxonomic sampling across the world's tropics. Density maps for geo-referenced species occurrences available from the Global Biodiversity Information Facility for four organism groups between the Tropics of Cancer and Capricorn (23.5 °S to 23.5 °N), showing the main spatial biases of taxonomic sampling. All records were cleaned using SpeciesGeoCoder. The figure is shown on a cylindrical equal area projection with standard parallels of 11.75 °S and 11.75 °N. The width of each cell is consistently 1 degree while the height of each cell is1 degree at the standard parallels, slightly lower at the equator and slightly higher at the Tropics of Cancer and Capricorn. The scale is in log10 number of records.



1987

1988

1989

1990

1991

1992

Fig. 3 The complex topography of South America. This map highlights the topographic differences across the continent, major mountain chains and river basins. Areas colored in blue are sedimentary basins in the Andean foreland; areas colored in yellow are topographic barriers (or 'highs') that divide these basins and define the Amazon drainage basin in its higher reaches. Base map created by Paulo Petry from the Shuttle Radar Topography Mission with elevations in meters. Note that the scale exaggerates differences at lower elevations.

1994

1993

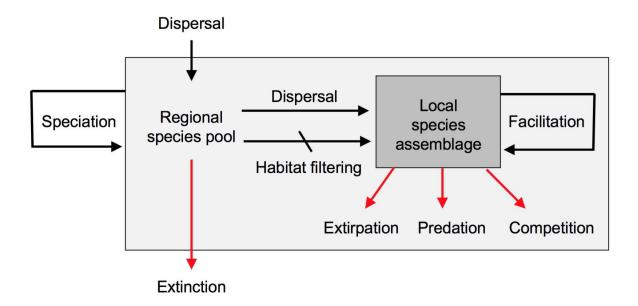


Fig. 4 Main evolutionary and ecological processes contributing to the formation of species richness.

The regional species pool (light gray box) is defined as the sum of all the local species assemblages (darker gray box). Black arrows indicate processes that increase species richness, red arrows processes that reduce species richness. Note the hierarchical organization of processes resulting in species richness, with evolutionary processes occurring over regional to continental spatiotemporal scales and ecological processes occurring over local scales. Speciation and dispersal contribute new species to the regional pool, while extinction removes species. Dispersal mediated by abiotic habitat filtering and biotic facilitation (Kraft et al., 2014) increase the richness of local assemblages by enhancing establishment of species preadapted to local conditions, or aiding in the establishment of other species. Biotic interactions such as predation and competition may serve to reduce local richness. Diagram modified from Ricklefs and Schluter (1993).



2007 TABLES

Biodiversity components	Known knowns	Known unknowns	Unknown unknowns
Taxonomic diversity (i.e., species numbers)	Approximate species numbers for macroscopic organisms; human impact tends to decrease overall diversity	Large portions of biodiversity are unexplored (i.e., microbes, invertebrates, fungi)	Taxonomic units used in biodiversity studies may be unequal
Genetic diversity (within species)	Patterns of genetic diversity for very selected taxa	Overall patterns of genetic diversity	How generalizable are conclusions drawn by such limited patterns of genetic diversity
Phylogenetic diversity	General understanding of the Tree (or Network) of Life	Drivers of diversification	Potential biases in phylogeny reconstruction and time-calibration
Spatial patterns of diversity	Hotspots and general patterns of species richness and diversity; large scale species ranges for charismatic taxa	Areas of endemism; known patterns of biodiversity distributions are biased; ecological preferences of species; drivers of spatial patterns of diversity	Human impact to overall spatial patterns
Functional diversity (traits)	Large scale productivity patterns	Biotic interactions	Relevance of current functional diversity measures; equivalency in functional traits; relationship between current and future functional diversity

2008

2009

Table 1. The various components of Neotropical biodiversity, major aspects known about them, and key

2010 topics that remain to be understood.