

A peer-reviewed version of this preprint was published in PeerJ on 19 December 2017.

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Selwyn JD, Johnson JE, Downey-Wall AM, Bynum AM, Hamner RM, Hogan JD, Bird CE. 2017. Simulations indicate that scores of lionfish (*Pterois volitans*) colonized the Atlantic Ocean. PeerJ 5:e3996
<https://doi.org/10.7717/peerj.3996>

Estimating the number of lionfish (*Pterois volitans*) that colonized the Atlantic Ocean

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The invasion of the western Atlantic Ocean by the Indo-Pacific red lionfish (*Pterois volitans*) has had devastating consequences for marine ecosystems. Estimating the number of colonizing lionfish can be useful in identifying the introduction pathway and can inform policy decisions aimed at preventing similar invasions. It is well-established that at least ten lionfish were initially introduced. However, that estimate has not faced statistical scrutiny and is based solely on the number of haplotypes in the maternally-inherited mitochondrial control region observed in samples from the western Atlantic Ocean. To rigorously estimate the number of lionfish introduced, we used a forward-time, Wright-Fisher, population genetic model of the mitochondrial control region in concert with a demographic life-history model to simulate the invasion. The colonization of the western North Atlantic and subsequent spread to the Caribbean and Gulf of Mexico were modeled as single introductions of 1-800 females, which should conservatively estimate the most likely number of colonizing lionfish relative to a model with multiple introductions. Assuming a balanced sex ratio and no Allee effects, the simulations indicate that the Atlantic population was most likely founded by 118 (54-514, 95% HPD) lionfish from the Indo-Pacific, the Caribbean by 84 (22-328, 95% HPD) lionfish from the Atlantic, and the Gulf of Mexico by at least 114 (no upper bound on 95% HPD) lionfish from the Caribbean. Considering the additional possibilities that (1) multiple Indo-Pacific populations colonized the Atlantic and (2) Allee effects (e.g. 50% reduction in fecundity) plagued the colonists, the most likely number of lionfish increased to 180 (88-416, 95% HPD). These results, in combination with all other published data, support the hypothesis that lionfish were introduced to the Atlantic via the aquarium trade, rather than shipping. The most parsimonious introduction pathway is a single release of lionfish in one location, but it seems likely that a combination of smaller and larger releases from a variety of aquarium trade stakeholders occurred near Miami, Florida. Efforts to prevent future invasions via the

aquarium trade should focus on the education of stakeholders and the prohibition of release, with adequate rewards for compliance and penalties for violations.

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17 **Submission to:** PeerJ

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19

20 **ABSTRACT**

21 The invasion of the western Atlantic Ocean by the Indo-Pacific red lionfish (*Pterois volitans*) has
22 had devastating consequences for marine ecosystems. Estimating the number of colonizing
23 lionfish can be useful in identifying the introduction pathway and can inform policy decisions
24 aimed at preventing similar invasions. It is well-established that at least ten lionfish were initially
25 introduced. However, that estimate has not faced statistical scrutiny and is based solely on the
26 number of haplotypes in the maternally-inherited mitochondrial control region observed in
27 samples from the western Atlantic Ocean. To rigorously estimate the number of lionfish
28 introduced, we used a forward-time, Wright-Fisher, population genetic model of the
29 mitochondrial control region in concert with a demographic life-history model to simulate the
30 invasion. The colonization of the western North Atlantic and subsequent spread to the Caribbean
31 and Gulf of Mexico were modeled as single introductions of 1-800 females, which should
32 conservatively estimate the most likely number of colonizing lionfish relative to a model with
33 multiple introductions. Assuming a balanced sex ratio and no Allee effects, the simulations
34 indicate that the Atlantic population was most likely founded by 118 (54-514, 95% HPD)
35 lionfish from the Indo-Pacific, the Caribbean by 84 (22-328, 95% HPD) lionfish from the
36 Atlantic, and the Gulf of Mexico by at least 114 (no upper bound on 95% HPD) lionfish from the
37 Caribbean. Considering the additional possibilities that (1) multiple Indo-Pacific populations
38 colonized the Atlantic and (2) Allee effects (50% reduction in fecundity) plagued the colonists,
39 the most likely number of lionfish increased to 180 (88-416, 95% HPD). These results, in
40 combination with all other published data, support the hypothesis that lionfish were introduced to
41 the Atlantic via the aquarium trade, rather than shipping. The most parsimonious introduction
42 pathway is a single release of lionfish in one location, but it seems likely that a combination of

43 smaller and larger releases from a variety of aquarium trade stakeholders occurred in the vicinity
44 of Miami, Florida. Efforts to prevent future invasions via the aquarium trade should focus on the
45 education of stakeholders and the prohibition of release, with adequate rewards for compliance
46 and penalties for violations.

47 INTRODUCTION

48 The ability to successfully eradicate an alien-invasive species is directly correlated with
49 the population size (Epanchin-Niell & Hastings, 2010) and the amount of time that has passed
50 since the initial introduction and establishment (see Blackburn et al., 2011). If the alien
51 population becomes invasive and spreads after establishing, eradication becomes nearly
52 impossible without vast financial commitments (Epanchin-Niell & Hastings, 2010). Preventative
53 measures and early detection enable the effective control of alien species before exponential
54 population growth exceeds conventional control methods (Jarrad et al., 2011). Given the costs of
55 control, prevention or minimization of future invasions is the most effective approach to avoid
56 the destruction of ecosystems and protect economic interests (CBD, 2002; Pimentel, Zuniga &
57 Morrison, 2005; Simberloff et al., 2013).

58 Elucidating the likely route of transport for already established and spreading alien
59 species, and determining how many colonists established the initial population can be used to
60 inform preventative management and education efforts (Estoup & Guillemaud, 2010; Hulme,
61 2015; McGeoch et al., 2016). While the initial mechanism of introduction is rarely conclusively
62 identified (Brockerhoff et al., 2014), understanding the number of invaders and the frequency of
63 introductions (i.e., propagule pressure) can be used to infer likely invasion routes (Hulme, 2015).
64 Propagule pressure is directly correlated with the ability of an introduced species to become
65 established and spread (Yang et al., 2012; Brockerhoff et al., 2014). To determine the propagule
66 pressure and infer likely invasion routes, it is important to have probabilistic estimates of the
67 number of invaders initially present (García-Díaz et al., 2015).

68 Genetic methodologies are particularly useful in reconstructing the history of introduced
69 species (Estoup & Guillemaud, 2010; Rius et al., 2015; Cristescu, 2015). For example, biological

70 invasions that occur following an initial population bottleneck will typically exhibit founder
71 effects, which can be exploited to estimate the initial number of founders (Azzurro et al., 2006;
72 Golani et al., 2007). Integrated population genetic (Fisher, 1922; Wright, 1931) and demographic
73 models can be used to infer and predict founding population sizes and diversity (Ficetola, Bonin
74 & Miaud, 2008; Hulme et al., 2008; Tran, Hofrichter & Jost, 2012; Benson et al., 2016).

75 The invasion of the western Atlantic Ocean by Indo-Pacific lionfish (*Pterois volitans*, *P.*
76 *miles*) has been recognized as a global environmental problem (Sutherland et al., 2010; Hixon et
77 al., 2016). In the Atlantic, alien-invasive lionfish consume both economically and ecologically
78 important species (Morris & Akins, 2009; Côté et al., 2013). The net effect of adding lionfish to
79 the ecosystem has been a reduction in recruitment and biomass of native species that fall prey to
80 the lionfish. In some cases severe reductions have been observed (Albins & Hixon, 2008; Green
81 et al., 2012; however, see Hackerott et al., 2017). Lionfish have also been indirectly tied to a
82 shift from coral to algal dominated habitats through the consumption of herbivorous fishes
83 (Albins & Hixon, 2011; Lesser & Slattery, 2011). Given their range expansion and population
84 size, conventional eradication is not a viable option for controlling these effects (Côté, Green &
85 Hixon, 2013) despite the efficacy of local management (Green et al., 2014; Usseglio et al.,
86 2017).

87 Lionfish were transported from the Indo-Pacific to the Atlantic via the aquarium trade
88 and likely released by aquarists into the wild initiating the introduction (Semmens et al., 2004).
89 The initial sightings of lionfish in the western Atlantic occurred in Florida in the mid-1980s
90 (Courtenay, 1995; Morris & Akins, 2009). While initially slow to spread outside this region
91 (Whitfield et al., 2002), an indication of Allee effects (Taylor & Hastings, 2005; Tobin et al.,
92 2007), by 2007 the invasion was characterized by large populations (Whitfield et al., 2007) and

93 exponentially increasing population size (Green et al., 2012). Between 2007 and 2010, lionfish
94 spread across the Caribbean (Betancur-R. et al., 2011). In 2010 the invasion front entered the
95 Gulf of Mexico, originating from the Caribbean population (Johnson et al., 2016). The stepping-
96 stone pattern of this invasion has resulted in three genetically distinct lionfish populations, one in
97 each of these regions, with each subsequently colonized region exhibiting progressively lower
98 genetic diversity (Johnson et al., 2016).

99 The absolute minimum number of *P. volitans* individuals that could have colonized the
100 Atlantic has been estimated to be between six and 10 (Betancur-R. et al., 2011). However, this
101 estimate is based upon counts of unique haplotypes from a 674 bp fragment of mtDNA and is not
102 a robust reflection of the number of individual lionfish that were introduced because it is not
103 rooted in a rigorous probabilistic framework. Therefore, this bare-minimum estimate is not
104 useful to inform either a further understanding of the processes leading to the establishment of
105 the invasion or policy aimed at preventing similar future invasions. Further, the lower estimate of
106 six colonists postulates that four haplotypes may have originated through mutation in the
107 Atlantic, but this is unlikely. The behavior of allelic diversity in expanding populations is well
108 established (Nei, Maruyama & Chakraborty, 1975; Maruyama & Fuerst, 1985), and is
109 characterized by an excess of rare alleles. However, this pattern takes time to develop because
110 mutation rates, even in mitochondrial DNA, are relatively slow (see Lynch, 2010). The invasive
111 lionfish population has yet to exhibit the characteristic genetic signature of an expanding
112 population, despite a large documented population size and widespread expansion. Thus, it is
113 exceedingly unlikely that any of the haplotypes observed in the Atlantic are the result of
114 mutations that arose in the Atlantic. Indeed, in the well-documented invasion of Hawai'i by the
115 bluestriped snapper (*Lutjanus kasmira*), which predates the Atlantic lionfish invasion by roughly

116 20 years, no evidence of population expansion is evident in the mitochondrial control region
117 either (Gaither et al., 2010).

118 This current best estimate of the number of *P. volitans* that colonized the Atlantic (at least
119 nine females and one male) supports the hypothesis that releases by home aquarists triggered the
120 invasion. This hypothesis is tenuous, however, because it does not account for the genetic
121 diversity of the source population or the frequencies of haplotypes in the Atlantic. We
122 hypothesized that when considering additional available genetic information, it was highly
123 improbable that only one male and nine female lionfish (with nine unique haplotypes)
124 established the Atlantic population. If many more lionfish colonized the Atlantic, then it could
125 indicate that the scale of releases by home aquarists is larger than previously thought or that
126 home aquarists alone are not sufficient to explain the mechanism by which lionfish were
127 introduced to the Atlantic.

128 Here we simulate the lionfish (*P. volitans*) invasion and estimate the most likely number
129 of lionfish to colonize the western Atlantic Ocean, Caribbean, and Gulf of Mexico using a
130 demographic-population genetic model. The results of the simulation are used to re-evaluate the
131 mechanism by which lionfish were introduced to the Atlantic and how this affects the
132 management of future invasions.

133

134 **METHODS**

135 **Characterizing Genetic Composition of Wild Populations**

136 A total of 1,294 mitochondrial control region sequences consisting of 30 unique
137 haplotypes were gathered from GenBank and published literature (Table 1; Freshwater et al.,
138 2009; Betancur-R. et al., 2011; Toledo-Hernández et al., 2014; Butterfield et al., 2015; Johnson

139 et al., 2016). The sequences were obtained from *P. volitans* in Indonesia and the western Atlantic
140 Ocean. Johnson et al. (2016) report that lionfish in the western Atlantic exhibit genetic structure
141 among the western North Atlantic, Caribbean, and the Gulf of Mexico, but not within these
142 regions. Consequently, sampling locations within these regions of the western Atlantic were
143 pooled together for this analysis.

144 Because the Indo-Pacific population that was the source of the Atlantic lionfish invasion
145 has not been sampled, we needed to estimate its genetic makeup. We used the mean number of
146 pairwise differences and the observed number of segregating sites of a population sample of
147 Indonesian lionfish (Freshwater et al., 2009) to estimate the population parameter,
148 θ ($2N_e\mu$, where N_e is the effective population size and μ is the mutation rate) and its standard
149 deviation (s) using ARLEQUIN V 3.5 (Excoffier & Lischer, 2010). The mitochondrial control
150 region from the Indonesian population sample was tested for conformation with the assumptions
151 of the infinite alleles and infinite sites models of nucleotide evolution (Kimura, 1969; Watterson,
152 1975; Tajima, 1996), which are nearly identical for linked nucleotides like those in the
153 mitochondrial control region (Hartl & Clark, 2006), using the Ewens-Watterson test (Ewens,
154 1972; Watterson, 1978), Tajima's D (Tajima, 1989), and Fu's F_s (Fu, 1997) based on 10,000
155 simulations in ARLEQUIN v 3.5 (Excoffier & Lischer, 2010).

156 Genetic diversity was used to characterize the alien-invasive lionfish populations of the
157 western North Atlantic, Caribbean, and the Gulf of Mexico, so they could be compared with
158 simulated invading populations. Haplotype richness was calculated as the number of unique
159 haplotypes that have been observed in each population. Since sample sizes from each of the
160 invasive lionfish populations were at least 188 and the maximum number of haplotypes was
161 nine, we see little justification for rarefaction of the richness values. Haplotype diversity was

162 calculated as the probability of drawing two different haplotypes at random from the population
163 with 95% confidence intervals calculated based on 1,000 bootstraps sampled with replacement
164 (Nei & Tajima, 1981).

165

166 **Simulating Colonization**

167 The colonizations of the western North Atlantic, Caribbean, and Gulf of Mexico were
168 simulated as single introduction events, which represent the minimum number of introductions
169 required to trigger invasive populations with the observed allelic richness and mitochondrial
170 haplotype diversity. The number of introduced female lionfish was varied from 1 to 800. We
171 assumed that an equal number of males accompanied the females, given the ~1:1 ratio of males
172 to females in wild populations (Fogg et al., 2013; Downey-Wall, 2016), but the model only
173 requires enough males to fertilize the eggs of the females. Females were sampled from either (1)
174 the inferred Indo-Pacific population using an estimate of θ and the Ewens' sampling formula
175 (Ewens, 1972; Crane, 2016) when simulating the invasion of the western North Atlantic or (2)
176 the observed multinomial distribution of sampled haplotype frequencies in the western North
177 Atlantic and Caribbean populations when simulating the invasions in the Caribbean and Gulf of
178 Mexico, respectively. To test for model sensitivity to error in the estimate of θ (greater or lesser
179 genetic diversity or more source populations), simulations assuming that the source population
180 was characterized by $\theta \pm s$ and $\theta \pm 2s$ were also run.

181

182 ***Demographic model***

183 An individual-based model adapted from the stage-based matrix model developed by
184 Morris, Shertzer & Rice (2011) was used to simulate lionfish population growth. The model was

185 initialized with colonizing adult females, which minimizes the number of colonists required to
186 begin an invasion, relative to the introduction of larvae or juveniles. The model proceeded in
187 monthly time-steps and divides the lionfish life history into three primary life stages (larvae,
188 juvenile, and adult). We further divided the juvenile life stage into 11 juvenile stages of one
189 month to appropriately simulate the delay in the reproductive maturity of Atlantic derived
190 lionfish, which would not be accurately simulated by the model of Morris, Shertzer & Rice
191 (2011). Life-stage-specific demographic parameters from lionfish were used, as outlined by
192 Morris Shertzer & Rice (2011; also see for additional information regarding the estimation and
193 sensitivity of these parameters).

194 Briefly, adult fecundity (194,577 eggs/month/adult) was calculated from the reproductive
195 contribution (number of eggs) per female per spawn ($R_A = 35,315$, Morris 2009) and the
196 spawning rate of 7.9/month/female. The eggs had a mortality rate of 0.31/day (McGurk, 1987)
197 before hatching after three days (Morris, 2009). The larval stage lasted for the rest of the month
198 (Ahrenholz & Morris, 2010) with a mortality rate of 0.35/day (McGurk, 1987). Ultimately, the
199 proportion of larvae that survived and metamorphosed into juveniles was 0.00003 (G_L). The 11
200 juvenile stages had a mortality rate of 0.165/day (m_J). The adult life span was controlled by
201 mortality rate, which was 0.052/month (m_A , Lorenzen, 1996).

202 To simulate the effect of a slower growth rate at the outset of the invasion due to Allee
203 effects caused by low population densities (Stephens, Sutherland & Freckleton, 1999), we
204 adjusted the monthly fecundity to 25%, 50%, 75%, and 100% (relative fecundity, r) of that
205 reported by Morris, Shertzer & Rice (2011). Allee effects can reduce the fecundity to 25-50%, or
206 lower, in natural populations (Roll et al., 1997; Morgan, 1999; Berec, Angulo & Courchamp,
207 2007).

208 A series of four discrete, stage-based equations were used to calculate the number of
 209 individuals for each haplotype at each subsequent time point ($t + 1$) from the number at time
 210 point (t):

$$211 \quad n_L(t + 1) = rR_A n_A(t) \quad (1)$$

$$212 \quad n_{J,1}(t + 1) = G_L n_L(t) + (1 - m_J) n_{J,1}(t) \quad (2)$$

$$213 \quad n_{J,s+1}(t + 1) = (1 - m_J) n_{J,s}(t) \quad 1 < s < 11 \quad (3)$$

$$214 \quad n_A(t + 1) = (1 - m_J) n_{J,11}(t) + (1 - m_A) n_A(t + 1) \quad (4)$$

215 where $n_{[L,J,A]}$ represent the number of individuals at the larval, juvenile (s represents the 10,
 216 monthly juvenile stages), and adult stages, respectively (other variables are defined in the
 217 description of the demographic model, above). The model was run for 15 years to encompass the
 218 time from the initial lionfish sighting to range expansion (Whitfield et al., 2002).

220

221 ***Genetic drift model***

222 The demographic growth model was used to determine the number of larvae and adults in
 223 a Wright-Fisher, forward-time model of genetic drift with overlapping generations (Fisher, 1922;
 224 Wright, 1931). Each month, haplotypes are drawn from an infinite pool of eggs produced by the
 225 adult females to create the new cohort of lionfish larvae. This model assumes a mutation rate of
 226 zero due to the short time scale of the invasion, and there is no evidence of mutations driving
 227 mtDNA diversity in western Atlantic lionfish populations. Running the model for 15 years was
 228 sufficient time for haplotype frequencies to stabilize without mutation due to decreased genetic
 229 drift with increasing population size.

230

231 **Assessing Simulations**

232 Following the simulation, a sample was drawn from the simulated population that was
 233 equal in size to the number sampled and reported by researchers in the destination population.

234 For example, 459 lionfish have been sampled from the western North Atlantic (Freshwater et al.,
235 2009; Betancur-R. et al., 2011; Butterfield et al., 2015; Johnson et al., 2016), therefore 459 were
236 sampled from the simulated western North Atlantic population. The haplotype richness and
237 diversity of the simulated invasive populations were calculated for each simulation in the same
238 manner as was calculated for the observed population. The conditional joint probability of
239 finding the observed haplotype richness and haplotype diversity within the 95% confidence
240 interval given the number of female colonists was calculated based on 10,000 simulations. We
241 calculated the 50% and 95% highest probability density interval (HPD) of the initial number of
242 females using the `codA` package (Plummer et al., 2006). This process was repeated for each
243 source and destination population.

244 To test the sensitivity of results to variation in the estimate of θ for the Indo-Pacific
245 source population and the fecundity of the alien population, we performed a MANOVA with
246 both θ and fecundity as potentially interacting independent variables with the point estimate,
247 lower, and upper 95% HPD estimates of the number of colonizing females as dependent
248 variables.

249 All above analyses were performed in R v 3.3.1 using code written by the authors, with
250 figures made using the package `GGPLOT2` (Supplement B, Wickham, 2009; R Core Team, 2015).

251

252 **RESULTS**

253 **Genetic Diversity of Wild Populations**

254 Based on the mitochondrial control region haplotypes present in Indonesia, θ was
255 estimated to be 7.64 (± 2.73 SD). The population in Indonesia was generally consistent with the
256 assumptions of the infinite sites model (Tajima's $D = -1.161$, $p = 0.111$; Fu's $F = -8.008$, $p =$

257 0.004; Ewen-Watterson test $F = 0.077$, $p = 0.0807$). The invasive lionfish populations in the
258 western North Atlantic, Caribbean and Gulf of Mexico exhibited haplotype richnesses of 9, 4,
259 and 3 respectively with associated haplotype diversity estimated as 0.67, 0.47, and 0.55
260 respectively (Table 1).

261

262 **Indo-Pacific to Western North Atlantic Simulation**

263 Assuming the θ of the source population is the same as the Indonesian population and no
264 Allee effects in founders, the most likely number of *P. volitans* females to have colonized the
265 western North Atlantic was 59 (27 – 257 95% HPD, Fig. 1C), which can be doubled to 118 (54-
266 514, 95% HPD) females and males if we assume a 1:1 sex ratio. Varying the diversity of the
267 source population (θ) and the strength of Allee effects (fecundity) in the colonizing population
268 had a significant interactive effect on the estimates of colonizing lionfish (MANOVA $F_{(3,14)} \cong$
269 6.37; Pillai = 0.58; $p = 0.006$; Figs. 1, 2 & 3). Increasing the diversity of the source population by
270 increasing θ resulted in smaller estimated colonizing population sizes (Figs. 1 & 2), which reach
271 a minimum value at or before $\theta+2s$ except when fecundity is 25% of the expected value. Varying
272 the diversity of the source population with θ had little effect on the lower limit of the 95% HPD
273 of the colonizing population size, but had a large effect on the upper estimate (MANOVA $F_{(3,14)}$
274 $\cong 26.4$; Pillai = 0.85; $p < 0.0001$; Figs. 1 & 2). Similarly, as reproductive output increased, the
275 size of the colonizing population decreased (MANOVA $F_{(3,14)} \cong 14.2$; Pillai = 0.75; $p < 0.0002$;
276 Figs. 1 & 3). Including Allee effects (50% reduction in fecundity) and the additional possibility
277 that multiple source populations in the Indo-Pacific colonized the Atlantic increased the estimate
278 of introduced lionfish to ~180 (88-416, 95% HPD) females and males (Fig. 1E).

279

280 Simulations of Caribbean and Gulf of Mexico Colonizations

281 The most likely number of *P. volitans* females to have colonized the Caribbean from the
282 western North Atlantic is 42 (11-164 95% HPD, Fig. 4A; 84 females and males). The most likely
283 number of females to colonize the Gulf of Mexico from the Caribbean was 158 (57 - >800 95%
284 HPD, Fig. 4B; 316 females and males) but was only able to be estimated when the simulation
285 included the maximum fecundity. The lack of an upper bound on the number of Gulf colonists,
286 which is partially due to only simulating up to 800 colonizing females, is consistent with the low
287 diversity of the Caribbean population and its rapid colonization and expansion throughout the
288 Gulf of Mexico. Notably, the simulation results are congruent with previous research showing
289 strong genetic differentiation between the western North Atlantic and Caribbean, and relatively
290 weak genetic differentiation between the Caribbean and Gulf of Mexico (Johnson et al., 2016).

291

292 DISCUSSION

293 The simulations of the lionfish invasion performed here suggest that many more than 10
294 lionfish colonized the Atlantic Ocean. Assuming no Allee effects and a 1:1 ratio of females to
295 males, a rigorous, conservative, minimum estimate of the number of colonists is at least 54
296 lionfish (lower bound of 95% HPD). As the estimated number of female colonists increases, it
297 becomes increasingly improbable for only one male to have colonized the Atlantic given the
298 ~1:1 sex ratio observed in wild populations (2×10^{-7} for 27 female colonists based on the binomial
299 distribution). Therefore, the most likely number of colonizing males is the same as the number of
300 colonizing females, and all further discussion of the number of colonists will double the
301 estimates of females to include both sexes. It is noteworthy that the lower bound of the estimated
302 number of colonists was relatively robust to greater-than-expected genetic diversity in the source

303 population (greater θ equals greater genetic diversity), and that this estimate assumes no
304 reduction in fecundity or other Allee effects, which increase the estimated number of colonists.
305 Thus, it is unlikely that fewer than 54 lionfish established the Atlantic population. It is likely,
306 however, that the number of colonizing lionfish was greater than 54. Five hundred and fourteen
307 colonists are the upper bound of 95% HPD interval, and the most likely number of colonists was
308 118, assuming a single source population and no Allee effects (Fig. 1C).

309 Making parameters in the model more realistic improves the estimates of the number of
310 colonizing lionfish. Slower initial population growth rates due to Allee effects are suggested by
311 the lag time between the detection of lionfish and their spread (Schofield, 2009; Morris & Akins,
312 2009). The number of simulated colonists increased with slower population growth rates caused
313 by Allee effects. It is also possible that several source populations colonized the Atlantic, which
314 would increase the effective θ of the source population. Increasing θ decreases the estimated
315 number of lionfish. If we assume a 50% reduction in fecundity and multiple source populations,
316 or a greater amount of genetic diversity in the source population, 180 (88-416 95% HPD)
317 colonists are predicted (Figs. 1E & 2B). Stronger Allee effects result in a much greater number
318 of colonists, but greater genetic diversity will have little effect.

319 As the lionfish population expanded, the invasion spread beyond the western North
320 Atlantic, into the Caribbean and later the Gulf of Mexico (Johnson et al., 2016). Distinct genetic
321 structure developed among these regions, roughly matching patterns of genetic structure in other
322 species with a similar range (Taylor & Hellberg, 2006). As we have demonstrated here, each
323 expansion of lionfish across a semipermeable biogeographic barrier can be simulated as a new
324 colonization event, and it is likely that ~84 lionfish established the Caribbean population (Fig.
325 4A).

326 Due to the small founding population size of the Caribbean, there is an opportunity for
327 genetic drift to cause shifts in genetic composition that perpetuate as the population grows, with
328 a minimal influx of migrants from the founding population (Excoffier & Ray, 2008). This
329 process, termed gene surfing (Hallatschek et al., 2007; Hallatschek & Nelson, 2008), has been
330 observed in bacterial colonies experiencing a range expansion in laboratory settings and predicts
331 that rapidly expanding populations would be characterized by genetic discontinuities, as
332 observed in Atlantic-Caribbean lionfish. First reported by Johnson et al. (2016) for lionfish, to
333 our knowledge, this is the first documentation of gene surfing in a wild population. Given the
334 relatively small number of colonists likely to have spread from the western North Atlantic into
335 the Caribbean (84, Fig. 4A), the most parsimonious explanation for this expansion and shift in
336 genetic composition is simply a range expansion coupled with the phenomena of gene surfing,
337 rather than a secondary introduction as posited by Butterfield et al. (2015; see Johnson et al.,
338 2016).

339 The ‘tsunami’ of lionfish that rapidly propagated from the Caribbean was unlikely to
340 have experienced an Allee effect or to promote much additional gene surfing. Indeed, we were
341 unable to precisely estimate the number of colonists initiating the range expansion from the
342 Caribbean into the Gulf of Mexico (Fig. 4B), but it is likely that there were at least 114 colonists
343 (Fig. 4B) to produce the observed pattern of genetic diversity. This lack of convergence suggests
344 that while there is a genetic break between the Caribbean and Gulf of Mexico, it is a relatively
345 permeable barrier, which is supported by conventional population genetic studies and larval
346 tracking work (Johnson et al., 2016; Kitchens et al., 2017). We propose that the vast array of oil
347 platforms in the Gulf of Mexico likely provided ample habitat and aided in the rapid
348 establishment and spread of lionfish throughout the Gulf of Mexico (*sensu* Sheehy & Vik, 2010).

349 It is also important to note that our estimates of the number of initial colonists introduced to each
350 region increases with each additional step. An increasing number of founders in each region
351 aligns with observations of rapid establishment and spread of lionfish as they become introduced
352 to the Caribbean and Gulf of Mexico.

353

354 **Model Assumptions**

355 All models require making assumptions, and the most critical assumptions made in this
356 effort are that (1) the lionfish that colonized the Atlantic Ocean originated from a single
357 population in the Indo-Pacific, (2) the demographic model is reasonably parameterized, (3) a
358 single colonization event led to the establishment of the populations, and (4) adult lionfish
359 colonized. By manipulating θ , however, we showed that increasing diversity in the source
360 population (which would be the result of multiple populations colonizing the Atlantic) had a
361 relatively small effect on the lower estimate of the number of colonists. Additionally, it may be a
362 reasonable assumption that the lionfish came from few source populations. Indonesia and the
363 Philippines are the primary suppliers of coral reef fishes for the aquarium trade (Wood, 2001;
364 Rhyne et al., 2017; <https://www.aquariumtradedata.org/>), and these countries are in the center of
365 the range of *Pterois volitans*. With respect to the demographic parameters, the model we
366 employed was designed and parameterized for established lionfish populations, but it is likely
367 that Allee effects would result in slower population growth rates. Consequently, we varied
368 fecundity and found that Allee effects would require even more introductions to explain the
369 observed pattern of genetic diversity in the Atlantic. As pointed out previously and detailed in
370 the next section, modeling a single colonization results in the minimum number of lionfish
371 required to explain the observed pattern of genetic diversity. Finally, assuming that adult lionfish

372 were released minimizes the number of colonists required to establish a population because
373 adults are less likely to die before reproducing than larvae or juveniles. Adult females could even
374 be holding fertilized eggs upon release.

375

376 **Propagule Pressure and Invasions**

377 Propagule pressure, the number and frequency of introductions (Lockwood, Cassey &
378 Blackburn, 2005), is highly correlated with the establishment of alien invasive populations, and it
379 is likely that the Atlantic was subject to substantial propagule pressure from lionfish. In the
380 context of propagule pressure, there are generally two patterns of introduction prior to
381 establishment: (1) sporadic, infrequent introductions of large numbers of individuals, or (2)
382 frequent introductions of small numbers of individuals (see Simberloff, 2009 for both supporting
383 examples and exceptions). Previous invasions that have been triggered by infrequent introduction
384 of many individuals often involve the intentional release of a species to serve some purpose, after
385 which it becomes a nuisance species (e.g. cane toads throughout the Caribbean and Pacific:
386 Eastel, 1981; various fishes in Hawai'i: Randall, 1987; Gaither et al., 2010). Unintentional
387 large-scale releases are also known to have occurred as a result of escapes from both (1)
388 aquaculture and farming (Naylor, Williams & Strong, 2001; Zapiola et al., 2008; Ramírez et al.,
389 2015) and (2) international shipping in ballast water (Lavoie, Smith & Ruiz, 1999; Drake &
390 Lodge, 2004).

391 The alternate pathway to the establishment of an alien species, where small introductions
392 occur with relatively high frequency have led to the successful establishment of several alien
393 species (Eurasian house sparrows in North America, European red deer in New Zealand;
394 biocontrol insects in Canada: in Simberloff, 2009). In marine systems this often is a result of the

395 international aquarium trade and many individuals releasing pets into the environment (Padilla &
396 Williams, 2004; Duggan, Rixon & MacIsaac, 2006; Holmberg et al., 2015; Rhyne et al., 2017).
397 These releases may occur when people, seeking a seemingly more humane option than
398 euthanasia, release unwanted pets into the environment (Courtenay & Taylor, 1986; Duggan,
399 Rixon & MacIsaac, 2006).

400

401 **Most Likely Introduction Vector**

402 While many species can survive in ballast water and be transported by shipping traffic
403 (Medcof, 1975; Carlton, 1985), the introduction of lionfish to the Atlantic via ballast water is less
404 likely than through the aquarium trade (Whitfield et al., 2002), and the results presented here
405 reinforce that. In an analysis of international shipping patterns (ballast), ornamental marine fish
406 imports, and established populations of alien fishes, Semmens et al. (2004) conclude that
407 southeast Florida reefs (the epicenter of the Atlantic lionfish invasion) host an unusually high
408 number of alien species due to aquarium releases. Indeed, lionfish introduced through the
409 aquarium trade are hardy adults that are likely to survive (Wabnitz, 2003). In contrast, fragile
410 larvae have a high probability of mortality upon entering bilge tanks (Gollasch et al., 2000),
411 during transport (Gollasch et al., 2000; Ghabooli et al., 2016) and after release (Padilla &
412 Williams, 2004). Lionfish egg masses float for ~36-72 hours prior to hatching (Fishelson, 1975,
413 Morris, 2009) and are unlikely to enter bilge tanks while floating. Larvae that survive bilge entry
414 will travel for approximately 26 days between the Indo-Pacific region and Miami, given an
415 average speed of 24 knots (Notteboom & Cariou, 2009) and two days to traverse the Panama
416 Canal. Larvae begin feeding four days post hatch, can survive six days of starvation (Thresher,
417 1984), and thus, must feed in the bilge for at least 16 days on depleted plankton populations to

418 survive the voyage (see Gollasch et al., 2000; Ghabooli et al., 2016). The population of lionfish
419 in the bilge is likely to experience an extreme bottleneck and reduced genetic diversity (Ghabooli
420 et al., 2016). If the population does not go extinct during transport, the larvae must accrue
421 enough resources to metamorphose 20-35 days after fertilization (Ahrenholz & Morris, 2010)
422 and survive for another 11 months before reaching reproductive maturity in the Atlantic
423 (Thresher, 1984). Finally, they must find a mate in order to propagate the species in their new
424 environment.

425 If ballast water was the primary vector for the lionfish invasion, then we expect that there
426 would have been several successful introductions in other western Atlantic ports between New
427 York and Brazil. It is clear, however, that the lionfish invasion began in one location (near
428 Miami, Florida) in the mid-1980s, and all genetic evidence indicates that there were not
429 successful introductions in other Atlantic locations (Johnson et al., 2016). Further, given the
430 results of the simulations presented here, the offspring of at least 27 mothers, and likely many
431 more, would have to be loaded into ballast tanks, survive the voyage and develop into
432 reproductively mature adults that successfully reproduce. Adult lionfish have been found at
433 densities up to 26.3 ha⁻¹ in the Indo-Pacific region (Kulbicki et al., 2012); thus, many ships
434 would have transported lionfish larvae to Florida, exclusively, to produce the pattern of genetic
435 diversity observed in the Atlantic. The ballast hypothesis does not hold water.

436 We conclude, as others have previously, that the aquarium trade was the most likely
437 vector for the lionfish invasion. In light of the ~180 colonizing lionfish estimated here, it seems
438 most parsimonious to conclude that a combination of small and larger releases caused the
439 lionfish invasion. If the colonization involved several introductions of a small number of fish by
440 home aquarists, then many more lionfish would have to be released to generate the observed

441 pattern of genetic diversity in the Atlantic Ocean. Lionfish are voracious predators and are prime
442 candidates to be a nuisance by consuming other fish in a tank; thus, it is not implausible that
443 many aquarists would discard lionfish from their aquaria in the 1970s and 1980s when the home
444 saltwater aquarium trade skyrocketed (Andrews, 1990; Chapman et al., 1997). These lionfish,
445 however, would need to be released in the same locations (such as ports, harbors or piers) or
446 locate each other and aggregate after introduction. Consequently, its plausible or even likely that
447 wholesalers, aquarium shops, and aquarists were releasing lionfish into the Atlantic, leading to
448 the establishment of the population.

449

450 **Preventing Future Invasions**

451 Propagule pressure is an important predictor of invasion success (Lockwood, Cassey &
452 Blackburn, 2005), and despite the destructive consequences of the lionfish invasion, there is still
453 a healthy market serviced by the port of Miami. Between 2008 and 2011 approximately $7,000 \pm$
454 $3,000$ (SD) *Pterois* spp. yr⁻¹ were imported into the port of Miami, the site of the earliest reports
455 of lionfish in the Atlantic (Schofield, 2009). Approximately 70% were visually-identified as *P.*
456 *volitans* (Rhyne et al., 2017; <http://www.aquariumtradedata.org>), meaning that 30% were other
457 lionfish species that can become established if introduced. There are several other species of
458 lionfish in the Indo-Pacific region and one other species, *Pterois miles*, has also established a
459 population in the western North Atlantic (Hamner, Freshwater & Whitfield, 2007). Hybridization
460 of other *Pterois* spp. with the existing invasive lionfish population is also possible (see Wilcox,
461 2014), providing a potential adaptive subsidy to the invasive population. A small fraction of the
462 lionfish or other species being imported into Miami and other locations in the Atlantic Ocean on
463 a yearly basis could trigger another invasion.

464 Many invasions have been triggered by very small numbers of individuals (Simberloff,
465 2009); therefore, management and enforcement should act to safeguard against a similar invasion
466 in the future by eliminating propagule pressure (Caffrey et al., 2014). One focus should be on
467 containment of live alien species to minimize the risk of an accidental large-scale release of
468 potential invaders. A second focus should be on educating wholesalers, retailers, home aquarists,
469 and the public about the dangers to the ecosystem and economy of releasing unwanted pets into
470 the environment. Release into the wild is potentially viewed as a favorable alternative to
471 euthanasia for an unwanted pet, therefore, this education effort could be coupled with some form
472 of buy-back program where unwanted pets can be traded back to a store or government agency
473 as a humane and ecosystem-friendly option (Courtenay & Taylor, 1986; Courtenay & Stauffer,
474 1990; Duggan, Rixon & MacIsaac, 2006). A third focus should be on implementing penalties for
475 violating regulations and incentives for following them to increase compliance (Furlong, 1991;
476 Keane et al., 2008; Floerl, Inglis & Diettrich, 2016). Game theory dictates that cheating is a
477 viable strategy, unless a combination of the (1) risk of getting caught and (2) the cost associated
478 with getting caught is sufficiently great (Smith, 1982). If regulatory entities and aquarium trade
479 stakeholders take the threat of biological invasions seriously, it should be possible to avoid
480 another devastating invasion like the lionfish in the Atlantic.

481

482 **ACKNOWLEDGEMENTS**

483 NSF-MRI-CNS-1429518 provided a high-performance computing platform to perform the
484 simulations. RMH was supported by a Texas Comprehensive Research Fund grant from the
485 Texas A&M University-Corpus Christi Division of Research, Commercialization and Outreach.
486 This manuscript was generated as part of the Population Genetics/Evolutionary Genetics course

487 at TAMUCC where JDS, JEJ, and AMB were students and CEB was the instructor. We dedicate
488 this manuscript in memory of Dr. Stephen A. Karl. Dr. Karl played a prominent role in educating
489 CEB and many other molecular ecologists about population genetic theory. Without his
490 influence and teachings, this research would not have been conducted.

491 REFERENCES

- 492 Ahrenholz DW., Morris JA. 2010. Larval duration of the lionfish, *Pterois volitans* along the
493 Bahamian Archipelago. *Environmental Biology of Fishes* 88:305–309. DOI:
494 10.1007/s10641-010-9647-4.
- 495 Albins MA., Hixon MA. 2008. Invasive Indo-Pacific lionfish *Pterois volitans* reduce recruitment
496 of Atlantic coral-reef fishes. *Marine Ecology Progress Series* 367:233–238.
- 497 Albins MA., Hixon MA. 2011. Worst case scenario: potential long-term effects of invasive
498 predatory lionfish (*Pterois volitans*) on Atlantic and Caribbean coral-reef communities.
499 *Environmental Biology of Fishes*:1–7.
- 500 Andrews C. 1990. The ornamental fish trade and fish conservation. *Journal of Fish Biology*
501 37:53–59. DOI: 10.1111/j.1095-8649.1990.tb05020.x.
- 502 Azzurro E., Golani D., Bucciarelli G., Bernardi G. 2006. Genetics of the early stages of invasion
503 of the Lessepsian rabbitfish *Siganus luridus*. *Journal of Experimental Marine Biology*
504 *and Ecology* 333:190–201. DOI: 10.1016/j.jembe.2005.12.002.
- 505 Benson JF., Mahoney PJ., Sikich JA., Serieys LEK., Pollinger JP., Ernest HB., Riley SPD. 2016.
506 Interactions between demography, genetics, and landscape connectivity increase
507 extinction probability for a small population of large carnivores in a major metropolitan
508 area. *Proc. R. Soc. B* 283:20160957. DOI: 10.1098/rspb.2016.0957.
- 509 Berec L., Angulo E., Courchamp F. 2007. Multiple Allee effects and population management.
510 *Trends in Ecology & Evolution* 22:185–191. DOI: 10.1016/j.tree.2006.12.002.
- 511 Betancur-R. R., Hines A., Acero P. A., Ortí G., Wilbur AE., Freshwater DW. 2011.
512 Reconstructing the lionfish invasion: insights into Greater Caribbean biogeography.
513 *Journal of Biogeography* 38:1281–1293. DOI: 10.1111/j.1365-2699.2011.02496.x.

- 514 Blackburn TM., Pyšek P., Bacher S., Carlton JT., Duncan RP., Jarošík V., Wilson JRU.,
515 Richardson DM. 2011. A proposed unified framework for biological invasions. *Trends in*
516 *Ecology & Evolution* 26:333–339. DOI: 10.1016/j.tree.2011.03.023.
- 517 Brockerhoff EG., Kimberley M., Liebhold AM., Haack RA., Cavey JF. 2014. Predicting how
518 altering propagule pressure changes establishment rates of biological invaders across
519 species pools. *Ecology* 95:594–601. DOI: 10.1890/13-0465.1.
- 520 Butterfield JSS., Díaz-Ferguson E., Silliman BR., Saunders JW., Buddo D., Mignucci-Giannoni
521 AA., Searle L., Allen AC., Hunter ME. 2015. Wide-ranging phylogeographic structure of
522 invasive red lionfish in the Western Atlantic and Greater Caribbean. *Marine Biology*
523 162:773–781. DOI: 10.1007/s00227-015-2623-y.
- 524 Caffrey JM., Baars J-R., Barbour JH., Boets P., Boon P., Davenport K., Dick JTA., Early J.,
525 Edsman L., Gallagher C., Gross J., Heinimaa P., Horrill C., Hudin S., Hulme PE., Hynes
526 S., MacIsaac HJ., McLoone P., Millane M., Moen TL., Moore N., Newman JR.,
527 O’Conchuir R., O’Farrell M., O’Flynn C., Oidtmann B., Renals T., Ricciardi A., Roy
528 HE., Shaw R., Weyl O., Williams F., Lucy FE. 2014. Tackling invasive alien species in
529 Europe: the top 20 issues. *Management of Biological Invasions* 5:1–20. DOI: Caffrey,
530 Joe M.; Baars, Jan-Robert; Barbour, Jenny H.; Boets, Pieter; Boon, Philip; Davenport,
531 Keith; Dick, Jaimie T.A.; Early, John; Edsman, Lennart; Gallagher, Cathal; Gross,
532 Jackson; Heinimaa, Petri; Horrill, Chris; Hudin, Stéphanie; Hulme, Philip E.; Hynes,
533 Stephen; MacIsaac, Hugh J.; McLoone, Paul; Millane, Michael; Moen, Toril L.; Moore,
534 Niall; Newman, Jonathan; O’Conchuir, Ruairi; O’Farrell, Martin; O’Flynn, Colette;
535 Oidtmann, Birgit; Renals, Trevor; Ricciardi, Anthony; Roy, Helen; Shaw, Richard; Weyl,
536 Olaf; Williams, Frances; Lucy, Frances E.. 2014 Tackling invasive alien species in

- 537 Europe: the top 20 issues. *Management of Biological Invasions*, 5 (1). 1-20.
538 10.3391/mbi.2014.5.1.01 <<http://dx.doi.org/10.3391/mbi.2014.5.1.01>>.
- 539 Carlton JT. 1985. Transoceanic and interoceanic dispersal of coastal marine organisms: the
540 biology of ballast water. *Oceanography and Marine Biology* 23:313–371.
- 541 CBD 2002. *Sixth Conference of the Parties, the Hague, the Netherlands. 7–19 April 2002:*
542 *Decision VI/23: Alien Species that Threaten Ecosystems, Habitats or Species to Which is*
543 *Annexed Guiding Principles for the Prevention, Introduction and Mitigation of Impacts*
544 *of Alien Species that Threaten Ecosystems, Habitats or Species.*
- 545 Chapman FA., Fitz-Coy SA., Thunberg EM., Adams CM. 1997. United States of America Trade
546 in Ornamental Fish. *Journal of the World Aquaculture Society* 28:1–10. DOI:
547 10.1111/j.1749-7345.1997.tb00955.x.
- 548 Côté IM., Green SJ., Hixon MA. 2013. Predatory fish invaders: Insights from Indo-Pacific
549 lionfish in the western Atlantic and Caribbean. *Biological Conservation* 164:50–61. DOI:
550 10.1016/j.biocon.2013.04.014.
- 551 Côté IM., Green SJ., Morris JA., Akins JL., Steinke D. 2013. Diet richness of invasive Indo-
552 Pacific lionfish revealed by DNA barcoding. *Marine Ecology Progress Series* 472:249–
553 256. DOI: 10.3354/meps09992.
- 554 Courtenay WR. 1995. Marine fish introductions in southeastern Florida. *American Fisheries*
555 *Society Introduced Fish Section Newsletter* 14:2–3.
- 556 Courtenay WR., Stauffer JR. 1990. The Introduced Fish Problem and the Aquarium Fish
557 Industry. *Journal of the World Aquaculture Society* 21:145–159. DOI: 10.1111/j.1749-
558 7345.1990.tb01017.x.

- 559 Courtenay WR., Taylor JN. 1986. Strategies for Reducing Risks from Introductions of Aquatic
560 Organisms: A Philosophical Perspective. *Fisheries* 11:30–33. DOI: 10.1577/1548-
561 8446(1986)011<0030:SFRRFI>2.0.CO;2.
- 562 Crane H. 2016. The ubiquitous Ewens sampling formula. *Statistical science* 31:1–19.
- 563 Cristescu ME. 2015. Genetic reconstructions of invasion history. *Molecular Ecology* 24:2212–
564 2225. DOI: 10.1111/mec.13117.
- 565 Downey-Wall AM. 2016. Assessment of larval dispersal and habitat utilization of the invasive
566 red lionfish, *Pterois volitans*; emphasis on species management. Thesis Thesis. Corpus
567 Christi, Texas, USA: Texas A&M University - Corpus Christi.
- 568 Drake JM., Lodge DM. 2004. Global hot spots of biological invasions: evaluating options for
569 ballast–water management. *Proceedings of the Royal Society of London B: Biological*
570 *Sciences* 271:575–580. DOI: 10.1098/rspb.2003.2629.
- 571 Duggan IC., Rixon CAM., MacIsaac HJ. 2006. Popularity and Propagule Pressure: Determinants
572 of Introduction and Establishment of Aquarium Fish. *Biological Invasions* 8:377–382.
573 DOI: 10.1007/s10530-004-2310-2.
- 574 Easteal S. 1981. The history of introductions of *Bufo marinus* (Amphibia: Anura); a natural
575 experiment in evolution. *Biological Journal of the Linnean Society* 16:93–113. DOI:
576 10.1111/j.1095-8312.1981.tb01645.x.
- 577 Epanchin-Niell RS., Hastings A. 2010. Controlling established invaders: integrating economics
578 and spread dynamics to determine optimal management. *Ecology Letters* 13:528–541.
579 DOI: 10.1111/j.1461-0248.2010.01440.x.

- 580 Estoup A., Guillemaud T. 2010. Reconstructing routes of invasion using genetic data: why, how
581 and so what? *Molecular Ecology* 19:4113–4130. DOI: 10.1111/j.1365-
582 294X.2010.04773.x.
- 583 Ewens WJ. 1972. The sampling theory of selectively neutral alleles. *Theoretical Population*
584 *Biology* 3:87–112. DOI: 10.1016/0040-5809(72)90035-4.
- 585 Excoffier L., Lischer HE. 2010. Arlequin suite ver 3.5: a new series of programs to perform
586 population genetics analyses under Linux and Windows. *Molecular ecology resources*
587 10:564–567.
- 588 Excoffier L., Ray N. 2008. Surfing during population expansions promotes genetic revolutions
589 and structuration. *Trends in Ecology & Evolution* 23:347–351. DOI:
590 10.1016/j.tree.2008.04.004.
- 591 Ficetola GF., Bonin A., Miaud C. 2008. Population genetics reveals origin and number of
592 founders in a biological invasion. *Molecular Ecology* 17:773–782. DOI: 10.1111/j.1365-
593 294X.2007.03622.x.
- 594 Fishelson L. 1975. Ethology and reproduction of pteroid fishes found in the gulf of Aqaba (Red
595 Sea), especially *Dendrochirus brachypterus* (Cuvier), (Pteroidae, Teleostei).
596 *Pubblicazioni della Stazione Zoologica, Napoli*.
- 597 Fisher RA. 1922. On the Dominance Ratio. *Proceedings of the Royal Society Edinburgh* 42:321–
598 341.
- 599 Floerl O., Inglis GJ., Diettrich J. 2016. Incorporating human behaviour into the risk–release
600 relationship for invasion vectors: why targeting only the worst offenders can fail to
601 reduce spread. *Journal of Applied Ecology* 53:742–750. DOI: 10.1111/1365-2664.12609.

- 602 Fogg AQ., Hoffmayer ER., Driggers III WB., Campbell MD., Pellegrin GJ., Stein W. 2013.
603 Distribution and length frequency of invasive lionfish (*Pterois sp.*) in the northern Gulf of
604 Mexico. *Gulf and Caribbean research* 25:111–115.
- 605 Freshwater DW., Hines A., Parham S., Wilbur A., Sabaoun M., Woodhead J., Akins L., Purdy
606 B., Whitfield PE., Paris CB. 2009. Mitochondrial control region sequence analyses
607 indicate dispersal from the US East Coast as the source of the invasive Indo-Pacific
608 lionfish *Pterois volitans* in the Bahamas. *Marine Biology* 156:1213–1221. DOI:
609 10.1007/s00227-009-1163-8.
- 610 Fu Y-X. 1997. Statistical tests of neutrality of mutations against population growth, hitchhiking
611 and background selection. *Genetics* 147:915–925.
- 612 Furlong WJ. 1991. The Deterrent Effect of Regulatory Enforcement in the Fishery. *Land*
613 *Economics* 67:116–129. DOI: 10.2307/3146490.
- 614 Gaither MR., Bowen BW., Toonen RJ., Planes S., Messmer V., Earle J., Ross Robertson D.
615 2010. Genetic consequences of introducing allopatric lineages of Bluestriped Snapper
616 (*Lutjanus kasmira*) to Hawaii. *Molecular Ecology* 19:1107–1121. DOI: 10.1111/j.1365-
617 294X.2010.04535.x.
- 618 García-Díaz P., Ross JV., Ayres C., Cassey P. 2015. Understanding the biological invasion risk
619 posed by the global wildlife trade: propagule pressure drives the introduction and
620 establishment of Nearctic turtles. *Global Change Biology* 21:1078–1091. DOI:
621 10.1111/gcb.12790.
- 622 Ghabooli S., Zhan A., Paolucci E., Hernandez MR., Briski E., Cristescu ME., MacIsaac HJ.
623 2016. Population attenuation in zooplankton communities during transoceanic transfer in
624 ballast water. *Ecology and Evolution* 6:6170–6177. DOI: 10.1002/ece3.2349.

- 625 Golani D., Azzurro E., Corsini-Foka M., Falautano M., Andaloro F., Bernardi G. 2007. Genetic
626 bottlenecks and successful biological invasions: the case of a recent Lessepsian migrant.
627 *Biology Letters* 3:541–545. DOI: 10.1098/rsbl.2007.0308.
- 628 Gollasch S., Lenz J., Dammer M., Andres H-G. 2000. Survival of tropical ballast water
629 organisms during a cruise from the Indian Ocean to the North Sea. *Journal of Plankton*
630 *Research* 22:923–937. DOI: 10.1093/plankt/22.5.923.
- 631 Green SJ., Akins JL., Maljković A., Côté IM. 2012. Invasive Lionfish Drive Atlantic Coral Reef
632 Fish Declines. *PLoS One* 7:e32596.
- 633 Green SJ., Dulvy NK., Brooks ALM., Akins JL., Cooper AB., Miller S., Côté IM. 2014. Linking
634 removal targets to the ecological effects of invaders: a predictive model and field test.
635 *Ecological Applications*. DOI: 10.1890/13-0979.1.
- 636 Hackerott S., Valdivia A., Cox CE., Silbiger NJ., Bruno JF. 2017. Invasive lionfish had no
637 measurable effect on prey fish community structure across the Belizean Barrier Reef.
638 *PeerJ* 5:e3270. DOI: 10.7717/peerj.3270.
- 639 Hallatschek O., Hersen P., Ramanathan S., Nelson DR. 2007. Genetic drift at expanding frontiers
640 promotes gene segregation. *Proceedings of the National Academy of Sciences*
641 104:19926–19930. DOI: 10.1073/pnas.0710150104.
- 642 Hallatschek O., Nelson DR. 2008. Gene surfing in expanding populations. *Theoretical*
643 *Population Biology* 73:158–170. DOI: 10.1016/j.tpb.2007.08.008.
- 644 Hamner RM., Freshwater DW., Whitfield PE. 2007. Mitochondrial cytochrome b analysis
645 reveals two invasive lionfish species with strong founder effects in the western Atlantic.
646 *Journal of Fish Biology* 71:214–222. DOI: 10.1111/j.1095-8649.2007.01575.x.

- 647 Hartl DL., Clark AG. 2006. *Principles of Population Genetics, Fourth Edition*. Sunderland,
648 Mass: Sinauer Associates, Inc.
- 649 Hixon MA., Green SJ., Albins MA., Akins JL., Morris Jr JA. 2016. Lionfish: a major marine
650 invasion. *Marine Ecology Progress Series* 558:161–165.
- 651 Holmberg RJ., Tlusty MF., Futoma E., Kaufman L., Morris JA., Rhyne AL. 2015. The 800-
652 Pound Grouper in the Room: Asymptotic Body Size and Invasiveness of Marine
653 Aquarium Fishes. *Marine Policy* 53:7–12. DOI: 10.1016/j.marpol.2014.10.024.
- 654 Hulme PE. 2015. Invasion pathways at a crossroad: policy and research challenges for managing
655 alien species introductions. *Journal of Applied Ecology* 52:1418–1424. DOI:
656 10.1111/1365-2664.12470.
- 657 Hulme PE., Bacher S., Kenis M., Klotz S., Kühn I., Minchin D., Nentwig W., Olenin S., Panov
658 V., Pergl J., Pyšek P., Roques A., Sol D., Solarz W., Vilà M. 2008. Grasping at the routes
659 of biological invasions: a framework for integrating pathways into policy. *Journal of*
660 *Applied Ecology* 45:403–414. DOI: 10.1111/j.1365-2664.2007.01442.x.
- 661 Jarrad FC., Barrett S., Murray J., Parkes J., Stoklosa R., Mengersen K., Whittle P. 2011.
662 Improved design method for biosecurity surveillance and early detection of non-
663 indigenous rats. *New Zealand Journal of Ecology* 35:132–144.
- 664 Johnson J., Bird CE., Johnston MA., Fogg AQ., Hogan JD. 2016. Regional genetic structure and
665 genetic founder effects in the invasive lionfish: comparing the Gulf of Mexico, Caribbean
666 and North Atlantic. *Marine Biology* 163:216. DOI: 10.1007/s00227-016-2981-0.
- 667 Keane A., Jones JPG., Edwards-Jones G., Milner-Gulland EJ. 2008. The sleeping policeman:
668 understanding issues of enforcement and compliance in conservation. *Animal*
669 *Conservation* 11:75–82. DOI: 10.1111/j.1469-1795.2008.00170.x.

- 670 Kimura M. 1969. The Number of Heterozygous Nucleotide Sites Maintained in a Finite
671 Population Due to Steady Flux of Mutations. *Genetics* 61:893–903.
- 672 Kitchens LL., Paris CB., Vaz AC., Ditty JG., Cornic M., Cowan JH., Rooker JR. 2017.
673 Occurrence of invasive lionfish (*Pterois volitans*) larvae in the northern Gulf of Mexico:
674 characterization of dispersal pathways and spawning areas. *Biological Invasions*:1–9.
675 DOI: 10.1007/s10530-017-1417-1.
- 676 Kulbicki M., Beets J., Chabanet P., Cure K., Darling E., Floeter S., Galzin R., Green A.,
677 Harmelin-Vivien M., Hixon M., Letourneur Y., de Loma T., McClanahan T., McIlwain
678 J., MouTham G., Myers R., O’Leary J., Planes S., Vigliola L., Wantiez L. 2012.
679 Distributions of Indo-Pacific lionfishes *Pterois spp.* in their native ranges: implications
680 for the Atlantic invasion. *Marine Ecology Progress Series* 446:189–205. DOI:
681 10.3354/meps09442.
- 682 Lavoie DM., Smith LD., Ruiz GM. 1999. The Potential for Intracoastal Transfer of Non-
683 indigenous Species in the Ballast Water of Ships. *Estuarine, Coastal and Shelf Science*
684 48:551–564. DOI: 10.1006/ecss.1999.0467.
- 685 Lesser MP., Slattery M. 2011. Phase shift to algal dominated communities at mesophotic depths
686 associated with lionfish (*Pterois volitans*) invasion on a Bahamian coral reef. *Biological*
687 *Invasions*:1–14.
- 688 Lockwood JL., Cassey P., Blackburn T. 2005. The role of propagule pressure in explaining
689 species invasions. *Trends in Ecology & Evolution* 20:223–228. DOI:
690 10.1016/j.tree.2005.02.004.

- 691 Lorenzen K. 1996. The relationship between body weight and natural mortality in juvenile and
692 adult fish: a comparison of natural ecosystems and aquaculture. *Journal of Fish Biology*
693 49:627–642. DOI: 10.1111/j.1095-8649.1996.tb00060.x.
- 694 Lynch M. 2010. Evolution of the mutation rate. *Trends in Genetics* 26:345–352. DOI:
695 10.1016/j.tig.2010.05.003.
- 696 Maruyama T., Fuerst PA. 1985. Population Bottlenecks and Nonequilibrium Models in
697 Population Genetics. Ii. Number of Alleles in a Small Population That Was Formed by a
698 Recent Bottleneck. *Genetics* 111:675–689.
- 699 McGeoch MA., Genovesi P., Bellingham PJ., Costello MJ., McGrannachan C., Sheppard A.
700 2016. Prioritizing species, pathways, and sites to achieve conservation targets for
701 biological invasion. *Biological Invasions* 18:299–314. DOI: 10.1007/s10530-015-1013-1.
- 702 McGurk MD. 1987. Natural mortality and spatial patchiness: Reply to Gulland. *Marine ecology*
703 *progress series. Oldendorf* 39:201–206.
- 704 Medcof JC. 1975. Living marine animals in a ship's ballast water. In: *Proceedings of the*
705 *National Shellfisheries Association*. UK (Plymouth), 11–12.
- 706 Morgan JW. 1999. Effects of Population Size on Seed Production and Germinability in an
707 Endangered, Fragmented Grassland Plant. *Conservation Biology* 13:266–273. DOI:
708 10.1046/j.1523-1739.1999.013002266.x.
- 709 Morris J. 2009. *The Biology and Ecology of the Invasive Indo-Pacific Lionfish*. Raleigh: North
710 Carolina State University.
- 711 Morris JA., Akins JL. 2009. Feeding ecology of invasive lionfish (*Pterois volitans*) in the
712 Bahamian archipelago. *Environmental Biology of Fishes* 86:389–398. DOI:
713 10.1007/s10641-009-9538-8.

- 714 Morris JA., Shertzer KW., Rice JA. 2011. A stage-based matrix population model of invasive
715 lionfish with implications for control. *Biological Invasions* 13:7–12. DOI:
716 10.1007/s10530-010-9786-8.
- 717 Naylor RL., Williams SL., Strong DR. 2001. Aquaculture--A Gateway for Exotic Species.
718 *Science* 294:1655–1656. DOI: 10.1126/science.1064875.
- 719 Nei M., Maruyama T., Chakraborty R. 1975. The Bottleneck Effect and Genetic Variability in
720 Populations. *Evolution* 29:1–10. DOI: 10.2307/2407137.
- 721 Nei M., Tajima F. 1981. DNA Polymorphism Detectable by Restriction Endonucleases. *Genetics*
722 97:145–163.
- 723 Notteboom T., Cariou P. 2009. Fuel surcharge practices of container shipping lines: Is it about
724 cost recovery or revenue making. In: *Proceedings of the 2009 International Association*
725 *of Maritime Economists (IAME) Conference*. IAME, 24–26.
- 726 Padilla DK., Williams SL. 2004. Beyond ballast water: aquarium and ornamental trades as
727 sources of invasive species in aquatic ecosystems. *Frontiers in Ecology and the*
728 *Environment* 2:131–138. DOI: 10.1890/1540-9295(2004)002[0131:BBWAAO]2.0.CO;2.
- 729 Pimentel D., Zuniga R., Morrison D. 2005. Update on the environmental and economic costs
730 associated with alien-invasive species in the United States. *Ecological Economics*
731 52:273–288. DOI: 10.1016/j.ecolecon.2004.10.002.
- 732 Plummer M., Best N., Cowles K., Vines K. 2006. CODA: convergence diagnosis and output
733 analysis for MCMC. *R News* 6:7–11.
- 734 R Core Team 2015. *R: A Language and Environment for Statistical Computing*. Vienna, Austria:
735 R Foundation for Statistical Computing.

- 736 Ramírez B., Ortega L., Montero D., Tuya F., Haroun R. 2015. Monitoring a massive escape of
737 European sea bass (*Dicentrarchus labrax*) at an oceanic island: potential species
738 establishment. *Journal of Aquaculture Research & Development* 6:1.
- 739 Randall JE. 1987. Introductions of Marine Fishes to the Hawaiian Islands. *Bulletin of Marine*
740 *Science* 41:490–502.
- 741 Rhyne AL., Tlusty MF., Szczebak JT., Holmberg RJ. 2017. Expanding our understanding of the
742 trade in marine aquarium animals. *PeerJ* 5:e2949. DOI: 10.7717/peerj.2949.
- 743 Rius M., Turon X., Bernardi G., Volckaert FAM., Viard F. 2015. Marine invasion genetics: from
744 spatio-temporal patterns to evolutionary outcomes. *Biological Invasions* 17:869–885.
745 DOI: 10.1007/s10530-014-0792-0.
- 746 Roll J., Mitchell RJ., Cabin RJ., Marshall DL. 1997. Reproductive Success Increases with Local
747 Density of Conspecifics in a Desert Mustard (*Lesquerella fendleri*). *Conservation*
748 *Biology* 11:738–746. DOI: 10.1046/j.1523-1739.1997.96013.x.
- 749 Schofield PJ. 2009. Geographic extent and chronology of the invasion of non-native lionfish
750 (*Pterois volitans* [Linnaeus 1758] and *P. miles* [Bennett 1828]) in the Western North
751 Atlantic and Caribbean Sea. *Aquatic Invasions* 4:473–479.
- 752 Semmens BX., Buhle ER., Salomon AK., PattengillSemmens CV. 2004. A hotspot of non-native
753 marine fishes: evidence for the aquarium trade as an invasion pathway. *Marine Ecology*
754 *Progress Series* 266:239–244. DOI: 10.3354/meps266239.
- 755 Sheehy DJ., Vik SF. 2010. The role of constructed reefs in non-indigenous species introductions
756 and range expansions. *Ecological Engineering* 36:1–11. DOI:
757 10.1016/j.ecoleng.2009.09.012.

- 758 Simberloff D. 2009. The Role of Propagule Pressure in Biological Invasions. *Annual Review of*
759 *Ecology, Evolution, and Systematics* 40:81–102. DOI:
760 10.1146/annurev.ecolsys.110308.120304.
- 761 Simberloff D., Martin J-L., Genovesi P., Maris V., Wardle DA., Aronson J., Courchamp F., Galil
762 B., García-Berthou E., Pascal M., Pyšek P., Sousa R., Tabacchi E., Vilà M. 2013. Impacts
763 of biological invasions: what's what and the way forward. *Trends in Ecology & Evolution*
764 28:58–66. DOI: 10.1016/j.tree.2012.07.013.
- 765 Smith JM. 1982. *Evolution and the Theory of Games*. Cambridge University Press.
- 766 Stephens PA., Sutherland WJ., Freckleton RP. 1999. What Is the Allee Effect? *Oikos* 87:185–
767 190. DOI: 10.2307/3547011.
- 768 Sutherland WJ., Clout M., Côté IM., Daszak P., Depledge MH., Fellman L., Fleishman E.,
769 Garthwaite R., Gibbons DW., Lurio JD., Impey AJ., Lickorish F., Lindenmayer D.,
770 Madgwick J., Margerison C., Maynard T., Peck LS., Pretty J., Prior S., Redford KH.,
771 Scharlemann JPW., Spalding M., Watkinson AR. 2010. A horizon scan of global
772 conservation issues for 2010. *Trends in Ecology & Evolution* 25:1–7. DOI:
773 10.1016/j.tree.2009.10.003.
- 774 Tajima F. 1989. Statistical method for testing the neutral mutation hypothesis by DNA
775 polymorphism. *Genetics* 123:585–595.
- 776 Tajima F. 1996. Infinite-allele model and infinite-site model in population genetics. *Journal of*
777 *Genetics* 75:27. DOI: 10.1007/BF02931749.
- 778 Taylor CM., Hastings A. 2005. Allee effects in biological invasions. *Ecology Letters* 8:895–908.
779 DOI: 10.1111/j.1461-0248.2005.00787.x.

- 780 Taylor MS., Hellberg ME. 2006. Comparative phylogeography in a genus of coral reef fishes:
781 biogeographic and genetic concordance in the Caribbean. *Molecular Ecology* 15:695–
782 707.
- 783 Thresher RE. 1984. *Reproduction in reef fishes*. Neptune City, New Jersey: T.F.H. Publications.
- 784 Tobin PC., Whitmire SL., Johnson DM., Bjørnstad ON., Liebhold AM. 2007. Invasion speed is
785 affected by geographical variation in the strength of Allee effects. *Ecology Letters* 10:36–
786 43. DOI: 10.1111/j.1461-0248.2006.00991.x.
- 787 Toledo-Hernández C., Vélez-Zuazo X., Ruiz-Diaz CP., Patricio AR., Mège P., Navarro M.,
788 Sabat AM., Betancur-R R., Papa R. 2014. Population ecology and genetics of the
789 invasive lionfish in Puerto Rico. *Aquatic Invasions* 9:227–237.
- 790 Tran TD., Hofrichter J., Jost J. 2012. An introduction to the mathematical structure of the
791 Wright–Fisher model of population genetics. *Theory in Biosciences* 132:73–82. DOI:
792 10.1007/s12064-012-0170-3.
- 793 Usseglio P., Selwyn JD., Downey-Wall AM., Hogan JD. 2017. Effectiveness of removals of the
794 invasive lionfish: how many dives are needed to deplete a reef? *PeerJ* 5:e3043. DOI:
795 10.7717/peerj.3043.
- 796 Wabnitz C. 2003. *From Ocean to Aquarium: The Global Trade in Marine Ornamental Species*.
797 Cambridge, United Kingdom: UNEP/Earthprint.
- 798 Watterson GA. 1975. On the number of segregating sites in genetical models without
799 recombination. *Theoretical Population Biology* 7:256–276. DOI: 10.1016/0040-
800 5809(75)90020-9.
- 801 Watterson GA. 1978. The Homozygosity Test of Neutrality. *Genetics* 88:405–417.

- 802 Whitfield PE., Gardner T., Vives SP., Gilligan MR., Courtenay WR., Ray GC., Hare JA. 2002.
803 Biological invasion of the Indo-Pacific lionfish *Pterois volitans* along the Atlantic coast
804 of North America. *Marine Ecology Progress Series* 235:289–297.
- 805 Whitfield PE., Hare JA., David AW., Harter SL., Munoz RC., Addison CM. 2007. Abundance
806 estimates of the Indo-Pacific lionfish *Pterois volitans/miles* complex in the Western
807 North Atlantic. *Biological Invasions* 9:53–64.
- 808 Wickham H. 2009. *ggplot2: elegant graphics for data analysis*. Springer New York.
- 809 Wilcox CL. 2014. Molecular investigations of the Pteroinae: insights into the invasive lionfishes
810 from the native range. Thesis Thesis. Honolulu, Hawai‘i, USA: University of Hawai‘i at
811 Mānoa.
- 812 Wood EM. 2001. Collection of coral reef fish for aquaria: global trade, conservation issues and
813 management strategies. *Marine Conservation Society, Ross-on-Wye, UK* 80.
- 814 Wright S. 1931. Evolution in Mendelian populations. *Genetics* 16:97–159.
- 815 Yang C-C., Ascunce MS., Luo L-Z., Shao J-G., Shih C-J., Shoemaker D. 2012. Propagule
816 pressure and colony social organization are associated with the successful invasion and
817 rapid range expansion of fire ants in China. *Molecular Ecology* 21:817–833. DOI:
818 10.1111/j.1365-294X.2011.05393.x.
- 819 Zapiola ML., Campbell CK., Butler MD., Mallory-Smith CA. 2008. Escape and establishment of
820 transgenic glyphosate-resistant creeping bentgrass *Agrostis stolonifera* in Oregon, USA: a
821 4-year study. *Journal of Applied Ecology* 45:486–494. DOI: 10.1111/j.1365-
822 2664.2007.01430.x.
- 823

Figure 1 (on next page)

Figure 1: Colonization of western North Atlantic from the Indo-Pacific.

The conditional joint probability of simulations resulting in the observed haplotype richness and diversity is plotted against the number of colonizing females. Each panel represents a different estimated value of θ in the Indo-Pacific population, ranging from (A) $\theta - 2s$ to (E) $\theta + 2s$, where θ was estimated from Indonesian *P. volitans* and s is the standard deviation. Colors indicate the relative fecundity of individual female lionfish in the model. Solid and dashed vertical bars above the plots represent the 50% and 95% HPD intervals, respectively. The square point along the HPD line indicates the point estimate of the most likely number of females to have generated the observed pattern of genetic diversity. Note that for $\theta - 2s$ and 25% relative fecundity (panel A), the most likely number of female colonists is much greater than 800 and the HPD intervals cannot be calculated.

Indonesia to Western North Atlantic

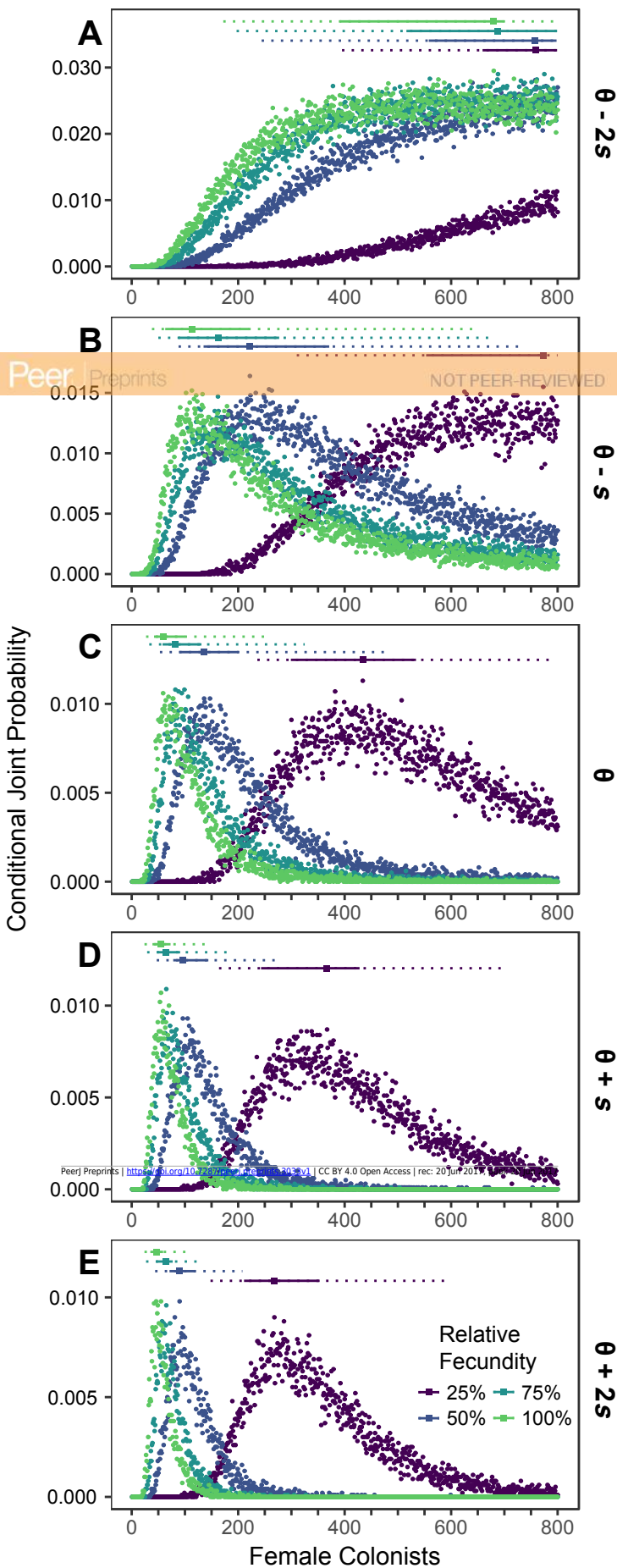
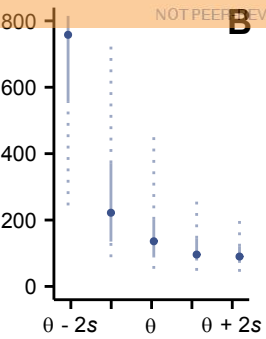
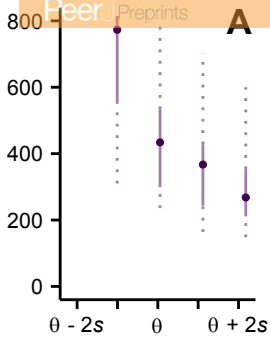


Figure 2 (on next page)

Figure 2: Effect of θ on the number of western North Atlantic female colonists.

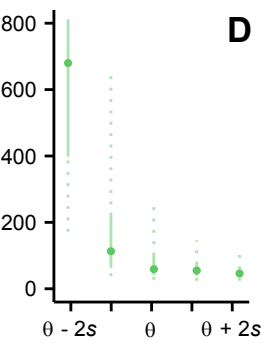
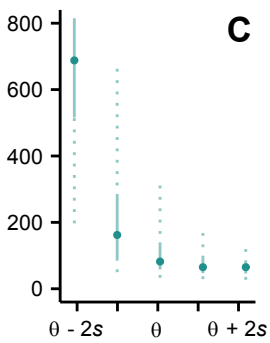
The most likely number of Atlantic female colonists is plotted against the θ of the Indo-Pacific source population, where θ was estimated from Indonesian *P. volitans* and s is the standard deviation of the estimate. Solid and dashed vertical bars represent the 50% and 95% HPD intervals, respectively. Colors indicate the relative fecundity of individual female lionfish in the model. Note that for panel A, the most likely number of female colonists at $\theta - 2s$ is not shown because it is much greater than 800.

Number of Female Colonists



Relative Fecundity

- 25%
- 50%
- 75%
- 100%



Source population θ

Figure 3(on next page)

Impact of Allee effects on the number of western North Atlantic female colonists.

The most likely number of Atlantic female colonists is plotted against the relative fecundity of female lionfish. The source population depicted has the same θ as estimated from a population of Indonesian lionfish. Solid and dashed vertical bars represent the 50% and 95% HPD intervals, respectively. Colors indicate the relative fecundity of individual female lionfish in the model.

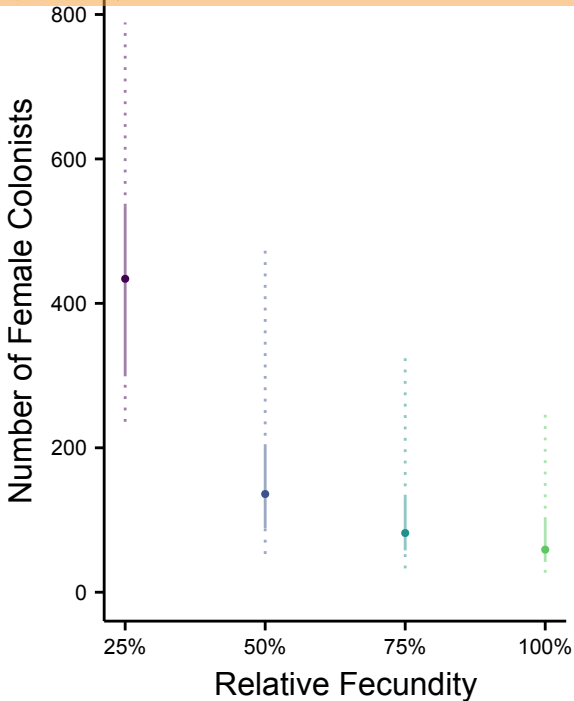
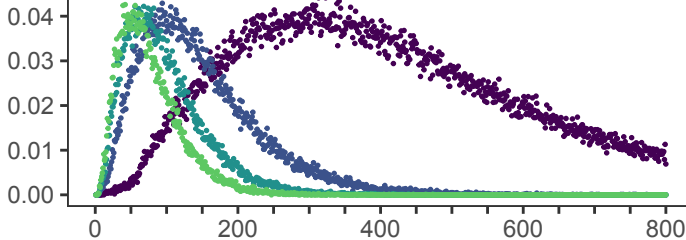


Figure 4(on next page)

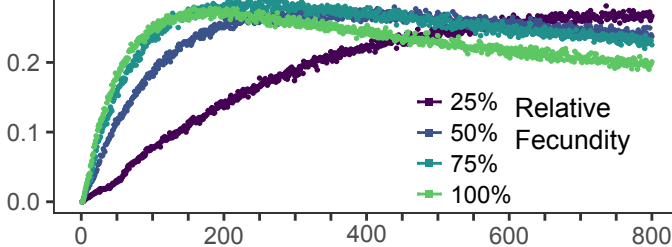
Range expansions from the western North Atlantic to the Caribbean to the Gulf of Mexico

The conditional joint probability of simulations resulting in the observed haplotype richness and diversity is plotted against the number of colonizing females (A) from the western North Atlantic into the Caribbean and (B) from the Caribbean into the Gulf of Mexico. Each point represents 10,000 simulations. Colors indicate the relative fecundity of individual female lionfish in the model. Solid and dashed vertical bars above the plots represent the 50% and 95% HPD intervals, respectively. The square point along the HPD line indicates the point estimate of the most likely number of females to have generated the observed pattern of genetic diversity.

Western North Atlantic to Caribbean

A

Caribbean to Gulf of Mexico

B

Number of Female Colonists

Table 1 (on next page)

Source material and population summaries

Pterois volitans mitochondrial d-loop haplotype data utilized in the present study.

Region	Number of individuals	Haplotype Richness	Haplotype Diversity (95% CI)	References	Accession Numbers
Indonesia	36	21	0.97 (0.95 - 1.00)	1	FJ516418- FJ516438
North Atlantic	459	9	0.67 (0.64 – 0.70)	1, 2, 4	FJ516409- FJ516417
Caribbean	601	4	0.47 (0.43 – 0.50)	2-4	FJ516409- FJ516412
Gulf of Mexico	188	3	0.55 (0.49 – 0.59)	5	FJ516409, FJ516410, FJ516412

1

2 (1-Freshwater et al., 2009; 2-Betancur-R. et al., 2011; 3-Toledo-Hernández et al., 2014; 4-

3 Butterfield et al., 2015; 5-Johnson et al., 2016)

4