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Comparison of histomorphology and DNA preservation produced by fixatives in the veterinary diagnostic laboratory setting

Abstract: Histopathology is the most useful tool for diagnosis of a number of diseases, especially cancer. To be effective, histopathology requires that tissues be fixed prior to processing. Formalin is currently the most common histologic fixative, offering many advantages: it is cheap, readily available, and pathologists are routinely trained to examine tissues fixed in formalin. However, formalin fixation substantially degrades tissue DNA, hindering subsequent use in diagnostics and research. We therefore evaluated three alternative fixatives, TissueTek® Xpress® Molecular Fixative, modified methacarn, and PAXgene®, all of which have been proposed as formalin alternatives, to determine their suitability for routine use in a veterinary diagnostic laboratory.

This was accomplished by examining the histomorphology of sections produced from fixed tissues as well as the ability to amplify fragments from extracted DNA. Tissues were sampled from two dogs and four cats, fixed for 24-48 hours, and processed routinely. While all fixatives produced acceptable histomorphology, formalin had significantly better morphologic characteristics than the other three fixatives. Alternative fixatives generally had better DNA amplification than formalin, although results varied somewhat depending on the tissue examined. While no fixative is yet ready to replace formalin, the alternative fixatives examined may be useful as adjuncts to formalin in diagnostic practices.
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Introduction:

Histopathology is the most useful tool for diagnosis of a number of diseases, especially cancer. To be effective, histopathology requires that tissues be fixed prior to processing. The ultimate tissue fixative for histopathology would create tissue histomorphology identical to formalin, pose no hazard to human health, preserve nucleic material for an extended period of time preferably at room temperatures, and be cost effective. Formalin has been the most common histologic fixative for over 100 years. It offers many advantages: it is cheap, readily available, and pathologists are routinely trained to examine formalin-fixed tissues (Gugic et al. 2007; Srinivasan et al. 2002).

However, formalin has several disadvantages as well. It can cause respiratory irritation and is classified as a carcinogen (Bolt et al. 2010; Bosetti et al. 2008; Buesa 2008; Gugic et al. 2007). It can also degrade nucleic acids and proteins, which make formalin-fixed tissues less usable for downstream molecular diagnostics (Buesa 2008; Gugic et al. 2007). Formalin is an aldehyde-based fixative that works by cross-linking proteins, which irreversibly degrades proteins and nucleic acids (Srinivasan et al. 2002).

The most common method for preserving tissues for molecular diagnostics is freezing at -80°C, although this does not allow for histopathologic examination of tissues. This method itself has a number of issues, including requiring special equipment, difficulty in shipping samples to laboratories, and requiring duplicate samples to be taken for histopathologic examination.

This is impractical for many private veterinary practices, as these generally lack the facilities to freeze samples at -80°C and many samples are too small to duplicate samples. Therefore, alternative fixatives have been proposed to allow for both histopathologic examination and molecular diagnostics {Cox, 2006 #2; Gugic, 2007 #3; Kap, 2011 #6; Vincek, 2003 #10}. These have been shown to preserve nucleic acids with results similar to those obtained with fresh or frozen tissues, while still preserving histomorphology. Some of the more successful alternatives include Tissue-Tek® Xpress® Molecular Fixative (Gugic et al. 2007), PAXgene® (Kap et al. 2011), and modified methacarn solution (Cox et al. 2006). All of these are alcohol-based and non-cross-linking. Evaluation of histomorphology preservation has varied among the studies evaluating these fixatives and generally involve research settings using techniques that are not practical in most clinical situations. In addition, some of these fixatives are currently cost prohibitive in the veterinary clinical setting. The majority of the studies evaluating these fixatives have evaluated single organs from humans or rodents {Cox, 2006 #2} or multiple organs from humans {Kap, 2011 #6; Vincek, 2003 #10}. One study evaluated Tissue-Tek® Xpress® Molecular Fixative and formalin comparing histomorphology and RNA quality from a variety of animal tissues (small animals, rodents, lagamorphs, birds, insects, and lizards) both at room temperature and high ambient temperatures simulating field collection of samples {Gugic, 2007 #3}. They concluded that Tissue-Tek® Xpress® Molecular Fixative protected RNA and provided acceptable histomorphology that would not hinder histologic diagnosis in the species studied. Some studies evaluating multiple animal species have included limited numbers of fixatives for comparison {Gugic, 2007 #3; Vincek, 2003 #10}. 

The main limitation of all of these previous studies is that they have evaluated fixatives in a research setting. There has not been a systematic evaluation of these to determine their utility in the veterinary diagnostic setting. Alternative fixatives would have a number of benefits for veterinary diagnostic laboratories, including (depending on the nature of the fixative) decreasing hazardous waste disposal costs, decreasing health risks to laboratory workers, and enhancing the power of retrospective studies. Therefore, we conducted this study to determine how alternative fixatives would function in a standard diagnostic laboratory setting by evaluating histomorphology of a variety of tissues from dogs and cats, as well performing a quantitative evaluation of recoverable DNA from tissues.

**Materials and Methods:**

All study protocols were approved by the University of Florida Institutional Animal Use and Care Committee (approval #201105654), and all animals were euthanized for reasons unrelated to this project. Necropsies were performed on four cats and two dogs within four hours of euthanasia. Replicate 10 x 10 x 5 mm samples from the liver, brain, lung, lymph node, kidney and spleen were collected. One sample of each tissue was frozen at -80°C. The remaining samples were placed into 10% neutral buffered formalin, Tissue-Tek® Xpress® Molecular Fixative (TT-XMF), modified methacarn, and PAXgene®, with a minimum of 1:10 tissue to fixative volume. Samples were allowed to fix for 24-48 hours at room temperature with the exception of tissues in PAXgene®, which were fixed and preserved according to the manufacturer’s protocol.

All tissues were processed using a Tissue-Tek processor with a standard overnight protocol (excluding formalin steps) followed by paraffin embedding and hematoxylin and eosin staining. The 10% neutral buffered formalin, Tissue-Tek® Xpress® Molecular Fixative, and PAXgene® fixatives were purchased commercially (ThermoFisher Scientific, Waltham, MA). Modified methacarn was prepared as previously described, using 8 parts methanol and 1 part glacial acetic acid (Cox et al. 2006).

Histomorphology was evaluated by two blinded board-certified veterinary anatomic pathologists (MJD, JAC) and one blinded anatomic pathology resident (WFC). Histomorphology of nuclear, cytoplasmic, and cellular membrane detail were evaluated on a 1-4 scale (table 1). Sample scores were averaged between all three evaluators. For one cat, the formalin-fixed lymph node sample was lost from the block; therefore, formalin fixation histomorphometry scores for lymph node are based on the remaining five samples. Both the individual components of the histomorphometry score as well as the total score were evaluated using a Kruskal-Wallis test (Lowry 2012) to determine if there was a difference between any of the four groups. If a significant difference was found (p < 0.05), the Mann-Whitney test was used to compare each group to each other.
group, to determine significant differences between each individual fixative. Fixatives were considered significantly different if the one-tailed Mann Whitney p value was less than 0.05. The minimum, 25th quartile, median, 75th quartile, and maximum were calculated for each tissue as well as for all tissues combined using Microsoft Excel (v14.3.9, Microsoft Corp., Seattle, WA). Graphs were generated using GNUplot (v.4.6, patchlevel 3).

Tissue scrolls were obtained from the paraffin blocks one week after processing and DNA was extracted using the QIAamp DNA FFPE Tissue kit (Qiagen Inc., Valencia, CA). As a control, DNA was extracted from tissues frozen at -80°C using the QIAamp DNA Mini Kit (Qiagen Inc.). Primers were designed by aligning the sequences of the retinol-binding protein 3, interstitial gene (IRBP) from dog, mouse, rat, and human, and selecting regions that were relatively conserved, to generate 100, 200, 300, 500 and 750 base pair long amplicons (table 2). Extracted DNA was amplified via PCR on an Applied Biosystems Veriti Thermal Cycler with the following conditions: 96°C for 3 minutes, followed by 35 cycles of 96°C for 1 minute, 60°C for 1 minute, then 72°C for 1 minute. This was followed by 7 minutes at 72°C, with a final hold at 4°C until the next morning. Samples were examined on a 1.25% agarose gel via electrophoresis.

The presence or absence of bands for all sizes was noted. The Kruskal-Wallis test was used to determine if there was a significant difference between the maximum band size for any of the fixatives. If significant (p<0.05), the Mann-Whitney test was used to compare each fixative against each other to determine which had significant differences (one-tailed p<0.05). The minimum, 25th quartile, median, 75th quartile, and maximum calculated for each fixative and for each tissue using Microsoft Excel. Graphs were generated using GNUplot (v.4.6, patchlevel 3).

**Results:**

**Histomorphology**

While the majority of the alternative fixatives produced adequate histomorphology in the tissues examined, formalin fixed tissues consistently resulted in superior histomorphology. There was no statistically significant difference between mean histomorphology scores comparing dog and cat tissues, and these were combined for subsequent analysis. Mean total, nuclear, cytoplasmic, and cellular membrane scores (figs. 1A-D) for formalin fixed tissues were higher than for all other fixatives (p<0.0001), although there is substantial variation with all fixatives (figs. 2A-D).

While a number of minor artifacts were noted, the primary difference noted between formalin and the other fixatives was in erythrocytes. This is likely reflected in
the significantly higher scores for formalin vs. other fixatives in the spleen (p=0.0026), an organ made up in large part by erythrocytes.

**DNA Preservation**

Formalin has significantly shorter total maximum DNA band sizes than TT-XMF (p=0.0158), modified methacarn solution (p < 0.0001), and PAXgene (p=0.0004) (fig. 3). In particular, the bands obtained from lymph nodes were significantly smaller with formalin than with TT-XMF (p=0.0179) and modified methacarn (p=0.004) (fig. 4). Overall, modified methacarn solution performed as well or better than the other fixatives for all tissues, with the best score in brain (median amplicon length of 750bp).

**Discussion:**

While alternative fixatives have been found to work well in research settings (Cox et al. 2006; Kap et al. 2011; Vincek et al. 2003), these are not ready to replace formalin for routine tissue processing in the veterinary laboratory. All of the fixatives require tissues be prevented from contacting formalin to benefit from their nucleic acid preserving qualities, which would require laboratories to either maintain separate tissue processors or bar submission of formalin-fixed tissues. Neither of these is practical in veterinary practice. Several fixatives produce excellent histomorphology with alternative processing techniques; this is also impracticable in most veterinary diagnostic laboratories, as it would require separate processing runs.

However, while no fixative is ideal from the standpoint of replacing formalin, all fixatives produced interpretable slides. Therefore, using alternative fixatives may be useful in specific circumstances where subsequent DNA isolation may be required. For example, tissue samples from neoplasms may be saved separately to generate a tissue bank for subsequent research projects. The specific alternative chosen should be based on the tissue selected, as well as predicted needs for DNA amplification and preservation of histomorphology. For example, while TT-XMF had better histomorphology scores in the kidney than either modified methacarn or PAXgene, it had a lower median DNA amplicon size.

One characteristic observed with alternative fixatives was that bloody or congested tissues often had unfixed areas, which could result in missing lesions and inaccurate diagnoses. This has not been found in previous studies (Cox et al. 2006), and may be due to a number of factors. First, the size of sample taken will greatly influence fixation. For most veterinary diagnostic laboratories, 1 cm thick samples are considered standard for histopathologic examination. In many previous studies, samples taken for fixation were substantially thinner; for example, the study by Cox et al. used 15 mm x 8 mm x 3 mm samples. Other possibilities include differences in processing; microwave
fixation (Cox et al. 2006) or rapid tissue processing (Vincek et al. 2003) techniques have
been used. Tuning the processing technique for the fixative selected would likely
improve fixation and the ultimate histomorphology.

Finally, our evaluation of macromolecule preservation of was limited to DNA.
Additional analysis would be required to determine whether these fixatives preserve
RNA equally well. Other variables require investigation to determine the best fixative for
a particular application. These include the effects of fixation time on nucleic acid quality,
as many samples will sit longer than 24 hours before processing, as well as the effect of
storage time after tissue processing but before sectioning for nucleic acid isolation, since
many blocks will be stored for a period of time between the evaluation of histopathology
and nucleic acid isolation. The latter is especially important if laboratories set up tissue
banks, as samples would be expected to be stored for prolonged periods.

Conclusions:
While no fixative is ideal to replace formalin, alternative fixatives have generally
acceptable histomorphologic characteristics in most tissues and are valuable adjuncts to
standard formalin fixation. Projects proposing to use an alternative fixative for a research
project should first evaluate the project requirements and ideally test the fixative with
samples of the target organ to determine the best fixative, processing characteristics, and
histomorphology compromises before actual sample collection begins.

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Vincek V, Nassiri M, Nadji M, and Morales AR. 2003. A tissue fixative that protects macromolecules (DNA, RNA, and protein) and histomorphology in clinical samples. *Laboratory investigation; a journal of technical methods and pathology* 83:1427-1435.

Figure Legends:

Figure 1. Histomorphology scores for all animals and tissues combined. The median is represented by a red diamond, the box represents the 25th and 75th quartiles, and the whiskers represent 1.5 x interquartile range.

Figure 2. Histomorphology scores for individual tissues. The median is represented by a red diamond, the box represents the 25th and 75th quartiles, and the whiskers represent 1.5 x interquartile range.

Figure 3. DNA amplicon size ranges for all animals and tissues combined. The median is represented by a red diamond, the box represents the 25th and 75th quartiles, and the whiskers represent 1.5 x interquartile range.

Figure 4. DNA amplicon sizes for different tissue samples. The median is represented by a red diamond, the box represents the 25th and 75th quartiles, and the whiskers represent 1.5 x interquartile range.

Figure 5. Representative fixative histomorphology. Samples are from the liver of a single cat. A – formalin, B – TT-XMF®, C – modified methacarn, D – PAXgene®.

Table Legends:

Table 1. Histomorphology scoring characteristics.

Table 2. DNA primers used in this study.
Fig 1.
Fig 2.
Fig. 3
Fig. 4
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<th>Characteristic</th>
<th>Score</th>
<th>Criteria</th>
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<tr>
<td>Nuclear</td>
<td>4</td>
<td>Sharp nuclear membrane; chromatin pattern and nucleolus, when present, are distinct</td>
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<tr>
<td></td>
<td>3</td>
<td>Slight degradation in chromatin pattern</td>
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<tr>
<td></td>
<td></td>
<td>Nucleolus, when present, less distinct but discernable, sharp nuclear membrane</td>
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<tr>
<td></td>
<td>2</td>
<td>Less distinct nuclear membrane, fuzzy chromatin pattern</td>
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<tr>
<td></td>
<td></td>
<td>Nucleolus, when present, is difficult to discern</td>
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<tr>
<td></td>
<td>1</td>
<td>Fuzzy nuclear membrane</td>
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<tr>
<td></td>
<td></td>
<td>Chromatin pattern difficult to determine</td>
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<tr>
<td></td>
<td></td>
<td>Nucleoli indetectable</td>
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<td>0</td>
<td>Nucleus not able to be differentiated from cytoplasm</td>
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<tr>
<td>Cytoplasm</td>
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<td>Normal cellular morphology easily determined</td>
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<tr>
<td></td>
<td>3</td>
<td>Intracytoplasmic details fuzzy</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>Only rare evidence of normal intracellular structures</td>
</tr>
<tr>
<td></td>
<td>1</td>
<td>Increased cytoplasmic pallor, increased cytoplasmic eosinophilia</td>
</tr>
<tr>
<td></td>
<td>0</td>
<td>Cytoplasm homogenously pale eosinophilic with no evidence of organelles</td>
</tr>
<tr>
<td>Cell Membranes</td>
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<td>Cells have distinct intracellular borders</td>
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<td></td>
<td>Normal substructures, if present, are easily distinguished</td>
</tr>
<tr>
<td></td>
<td>3</td>
<td>Loss of substructures in some cells</td>
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<tr>
<td></td>
<td></td>
<td>Slight loss of intracellular details</td>
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<tr>
<td></td>
<td>2</td>
<td>Loss of substructures in most cells</td>
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<td></td>
<td></td>
<td>Obvious fuzzing of many cellular borders</td>
</tr>
<tr>
<td></td>
<td>1</td>
<td>No substructures detected</td>
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<tr>
<td></td>
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<td>Significant fuzzing of most cellular borders</td>
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<tr>
<td></td>
<td>0</td>
<td>Cells unable to be distinguished from adjacent cells</td>
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Table 1
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<th>Primer Name</th>
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<td>IRBP_F</td>
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<tr>
<td>IRBP_R1_100bp</td>
<td>CTC TTG ATG GCC TGC TC</td>
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<td>GGC TCA TAG GAG ATG ACC AG</td>
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<tr>
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<tr>
<td>IRBP_R6_750bp</td>
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Table 2