Critical multi-stranded approach to determining the tolerance and ecological values of diatoms in unique aquatic ecosystems of anthropogenic origin

Rafał M. Olszyński Corresp., Equal first author, [1] , **Ewelina Szczepocka** [1] , **Joanna Żelazna-Wieczorek** Corresp. Equal first author, [1]

[1] Department of Algology and Mycology/Faculty of Biology and Environmental Protection, University of Lodz, Łódź, Polska

Corresponding Authors: Rafał M. Olszyński, Joanna Żelazna-Wieczorek
Email address: rafal.olszynski@biol.uni.lodz.pl, joanna.zelazna@biol.uni.lodz.pl

Background. The ecological state of surface waters is typically assessed by a multi-aspect approach based on a determination of its chemical and physical parameters by hydromorphology and the use of indicator organisms such as benthic diatoms. By assigning ecological indicator values (EIV), it is possible to create diatom indexes which serve as the basic tool in assessing the ecological status of surface waters. These ecological indicator values are set according to classification systems, such as the Van Dam et al. system, which classifies species of diatoms according to seven different ecological factors. However, recent studies on the autecology of diatoms have shown the need to verify and establish new ecological indicator values. To this end, water ecosystems are good environments to observe the range of tolerance of benthic diatoms to environmental conditions due to their unique physical and chemical parameters. The aim of the present study was to propose the establishment of new, or changed, ecological indicator values, according to Van Dam et al., of species of diatoms characteristic of three post-mining aquatic ecosystems **Methods.** In total, 36 species were identified that were characteristic of three water ecosystems: a salt water complex, a mined iron ore reservoir and a mined lignite reservoir. Their ecological indicator values were calculated using OMNIDIA software, and the environmental conditions prevailing in the studied ecosystems were determined. Of the 36 characteristic species, 16 lacking at least one assigned ecological indicator value were analysed further. The analysis identified three groups of selected characteristic species which showed a correlation, or lack of such, to the tested physical and chemical parameters. **Results.** Based on this multistage study of the autecology of characteristic diatoms, comprising an analysis of environmental conditions, literature analysis and reference indicator values of other species, it is proposed that 32 ecological indicator values be established or adjusted for 16 species, and that *Planothidium frequentissimum* be excluded from water quality assessments.

PeerJ Preprints | https://doi.org/10.7287/peerj.preprints.27739v1 | CC BY 4.0 Open Access | rec: 17 May 2019, publ: 17 May 2019

Critical multi-stranded approach to determining the tolerance and ecological values of diatoms in unique aquatic ecosystems of anthropogenic origin

Rafał M. Olszyński, Ewelina Szczepocka, Joanna Żelazna-Wieczorek

Department of Algology and Mycology, Faculty of Biology and Environmental Protection, University of Lodz, Łódź

Corresponding Author:
Rafał M. Olszyński
12/16 Banacha St, Łódź, 90-237, Poland
Email address: rafal.olszynski@biol.uni.lodz.pl

Abstract

Background. The ecological state of surface waters is typically assessed by a multi-aspect approach based on a determination of its chemical and physical parameters by hydromorphology and the use of indicator organisms such as benthic diatoms. By assigning ecological indicator values (EIV), it is possible to create diatom indexes which serve as the basic tool in assessing the ecological status of surface waters. These ecological indicator values are set according to classification systems, such as the Van Dam et al. system, which classifies species of diatoms according to seven different ecological factors. However, recent studies on the autecology of diatoms have shown the need to verify and establish new ecological indicator values. To this end, water ecosystems are good environments to observe the range of tolerance of benthic diatoms to environmental conditions due to their unique physical and chemical parameters. The aim of the present study was to propose the establishment of new, or changed, ecological indicator values, according to Van Dam et al., of species of diatoms characteristic of three post-mining aquatic ecosystems

Methods. In total, 36 species were identified that were characteristic of three water ecosystems: a salt water complex, a mined iron ore reservoir and a mined lignite reservoir. Their ecological indicator values were calculated using OMNIDIA software, and the environmental conditions prevailing in the studied ecosystems were determined. Of the 36 characteristic species, 16 lacking at least one assigned ecological indicator value were analysed further. The analysis identified three groups of selected characteristic species which showed a correlation, or lack of such, to the tested physical and chemical parameters.

Results. Based on this multistage study of the autecology of characteristic diatoms, comprising an analysis of environmental conditions, literature analysis and reference indicator values of other species, it is proposed that 32 ecological indicator values be established or adjusted for 16 species, and that *Planothidium frequentissimum* be excluded from water quality assessments.

Introduction

Diatoms (Bacillariophyta) are one of the main biotic elements used in the biological assessment of the ecological state of surface waters (*Water Framework Directive 2000*). Due to the fact that many countries are obliged to continually engage in biomonitoring, there is a clear need to develop flawlessly functioning methods based on the standardised use of diatoms as bioindicators (*Kahlert et al. 2016; Poikane, Kelly & Cantonati 2016; Szczepocka & Żelazna-Wieczorek 2018*). Diatom indexes and ecological systems based on the bioindication values of particular diatom species, derived from various environmental parameters, constitute a fundamental tool in the biological assessment of environments. Diatom indexes have been commonly used to assess flowing and standing water for over 20 years (*Kelly et al. 2008; Harding & Taylor, 2014; Szczepocka et al. 2014; Hutorowicz & Pasztaleniec 2014; Holms & Taylor 2015; Żelazna-Wieczorek & Nowicka-Krawczyk, 2015; Kolada et al. 2016*).

Currently, many countries use the OMNIDIA program (*Lecointe et al. 1993*) as a biological assessment tool. Its latest version (version 6.0.6) allows the calculation of 18 diatom indexes, and the determination of seven environmental parameters for eight ecological systems. However, the

PeerJ Preprints | https://doi.org/10.7287/peerj.preprints.27739v1 | CC BY 4.0 Open Access | rec: 17 May 2019, publ: 17 May 2019

specific ecological indicator values of many of the species given in the OMNIDIA database are absent or have not been updated in response to recent research. To complete the these missing values, and to verify existing ones, further studies are needed of the ecological tolerance of diatom species in different types of aquatic ecosystem.

Due to their specific environmental conditions, post-mining reservoirs represent an extremely valuable source of information for the study of ecological diatom tolerance ranges. Some studies of these environments have been performed, but these have addressed diatom paleoecology and their role as indicators of past climatic or environmental change (*de Haan et al. 1993; Rakowska 1996; Thomas & John 2006; Sienkiewicz & Gąsiorowski 2016*). Until now, the autecology of diatoms in post-mining reservoirs has rarely been studied (*Van Landingham 1968; de Haan et al. 1993; Rakowska 1996; Sienkiewicz and Gąsiorowski 2016*).

The present study examines the diatom assemblages present in three post-mining reservoirs of various geological origin. Due to variations in their environmental parameters, these bodies of water serve as specific and unique habitats for the development of these algae. The diatom assemblages quickly adapt to the currently prevailing conditions, which is manifested in the presence of taxa characteristic of these specific parameters. Considering their large share of the assemblage, the index values of the assemblages constitute the most important component in the calculation of diatomaceous indexes. These species are therefore of the greatest importance for surface water biomonitoring.

The aim of the present study was to identify the species of diatoms characteristic of the three studied types of post-mine reservoirs. Following this, taxa that did not have at least one ecological indicator value specified in the OMNIDIA database, according to the environmental parameters given by Van Dam et al. (*1994*), were identified. New ecological indicator values were proposed based on the relationship between the occurrence of the individual species and certain selected physical and chemical parameters, or existing ones were verified.

The Van Dam et al. (*1994*) ecological system is one of the main systems on which the OMNIDIA programme is based. It describes the indicator values of diatoms according to pH, salinity, nitrogen uptake metabolism, oxygen requirement, saprobity, trophic state and moisture aerophily. These values play a key role in calculating diatom indexes, and hence need to be kept up to date to enable accurate routine biomonitoring.

Materials & Methods

Study area

The study was performed on three hydrological objects created through exploration for mineral deposits due or were formed by the closure of mines. All three are located in the Łódzkie and Wielkopolskie voivodeships, central Poland. The first object, Pełczyska (PE), is situated in the village of Pełczyska, between Łódź and Łęczyca (Łódzkie voivodeship) (Fig. 1). As the local area is characterized by the presence of salt deposits, numerous wells were sunk in the eighteenth century to obtain brine. Currently, salt water flows out of one of them. This area has been studied by biologists and hydrobiologists since the 1960s (*Olaczek 1963; Pliński 1966; Pliński 1969; Pliński 1971a; Pliński 1971b; Pliński 1971c; Pliński 1973; Żelazna-Wieczorek 1996; Żelazna-Wieczorek 2002; Żelazna-Wieczorek, Olszyński & Nowicka-Krawczyk 2015; Żelazna-Wieczorek & Olszyński 2016*). The objects chosen for our research form the PE hydrological complex located in the vicinity of farmland; it comprises the salt water outflow, a drainage ditch and a pond, which acts as the receiver of the water.

The second object is the Łęczyca (LE) urban reservoir located within the city of Łęczyca (Łódzkie voivodeship) (Fig. 1). The reservoirs were created following the flooding the open-cast iron ore mine in the 1990s. This area is rich in syderite deposits, which are accompanied by other minerals. The complex consists of three connected reservoirs: two are directly connected to each other (LEP1 and LEP2), and the third (LEP3) is connected to LEP2 via a water drainage ditch (*Olszyński & Żelazna-Wieczorek 2018*). All three are located in an area with houses, garden plots and partly-wooded areas.

The third hydrological object is the Bogdałów reservoir (BO), created by the flooding of an opencast brown coal mine. It is located in the village of Bogdałów (Wielkopolskie voivodeship) in an area rich in lignite deposits (Fig. 1). Lignite from quaternary deposits was exploited from 1977 to a depth of 50 meters. Due to the specific construction of the open-pit area, being characterised by the thickest layer of poorly permeable boulder clay in the region, it was transformed into a storage site for quarried rocks in Koźmin. Finally, in 1993/1994, the drainage and runoff of surface waters was blocked to form a water reservoir with a depth of about 12 meters surrounded by forest (*Gabryś-Godlewska et al. 2004; Gadomska et al. 2007; Orlikowski & Szwed 2009*).

Samples

Samples of benthic diatoms and water samples were collected from each hydrological facility for the analysis of physical and chemical parameters. The water temperature, pH and electrolytic conductivity were measured *in situ* (Elmetron CP-401 and CC-401 devices). The following sampling points were established:

Pełczyska (51°58'34.47"N, 19°14'21.11"E) – outflow (PESB.), ditch (D.PEDB.) and pond (D.PEPB.); samples were collected quarterly in the period July 2013 to March 2014;

Łęczyca (52°3'5.30"N; 19°11'50.24"E) – reservoir 1 (D.LEP1.), reservoir 2 (D.LEP2.) and reservoir 3 (D.LEP3.), samples were collected quarterly in the period March 2014 to December 2015;

Bogdałów (52°2'51.29"N; 18°35'51.49"E) – reservoir (D.BOZB.), samples were collected quarterly in the period March 2015 to December 2016.

The water samples were subjected to chemical and physical analysis at the Laboratory of Geology at the Faculty of Geographical Sciences, University of Lodz.

In total, 44 benthic samples were collected. The permanent slides were created according to Żelazna-Wieczorek (*2011*).

Qualitative and quantitative analysis of diatoms was performed using a Nikon Eclipse 50i light microscope (LM) under 1000 × magnification (plan oil-immersion objective 100 × / 1.25): the diatoms were identified and counted for up to 500 valves in each preparation. Light photomicrographs were taken with an OPTA-TECH digital camera.
In the case of diatoms that were difficult to be identified using LM were subjected to scanning electron microscope (SEM) analysis utilize a Phenom Pro X (gold layer of 8 and 20 nm, at 10 kV) SEM at the Laboratory of Microscopy Imaging and Specialist Biological Techniques, Faculty of Biology and Environmental Protection, University of Lodz.

Mathematical analysis

The average percentage (AP) for a given species was determined based on the percentage contribution of the species in the samples tested for a given hydrological object (*Żelazna-Wieczorek 2011*). Species whose AP was greater than or equal to 5% for each hydrological object were identified as dominant.
The incidence was determined according to the Tümpling & Friedrich (*1999*) coefficient according to the range values: 100%–75% euconstant taxa (EC), 75%–50% constant taxa (CN), 50%–25% accessory taxa (AC) and 25%–1% accidental taxa (AD) (*Tümpling & Friedrich 1999*).
Multidimensional scaling analysis (MDS) based on Bray-Curtis similarity coefficients was used to identify natural groupings of samples. The results are given as a 3D diagram in which the degree of similarity is represented as the distances between particular points (samples), with greater distances indicating a lower degree of similarity. The reliability of the ordering of the assemblage is represented by the stress value, which reflects how well the ordination summarizes the observed distances among the samples. The best MDS fits are those with stress values near zero (*Clarke & Gorley 2015*).
The Shade Plot analysis, based on the Bray-Curtis similarity coefficient, was used to identify the diatom species that have the strongest influence on the similarities between the samples demonstrated in the MDS analysis. Shade Plot analysis compares two data matrices with each other and then groups them on two levels, according to the similarity of the samples and the factors affecting their similarity, i.e. diatom species. The results are represented graphically by shading individual cells: the intensity of the shading indicates the degree of the influence of a given factor (species) on the position of its sample within a given similarity cluster. The range of the shading was determined on the basis of log (x+1).
The SIMPER analysis was used to determine the characteristic species distinguishing the studied ecosystems. This method examines the participation of each variable in the overall similarity between groups of samples, thus indicating the species with the greatest influence on the degree of similarity, or dissimilarity, between particular samples and hydrological objects. This analysis is also based on the Bray-Curtis similarity coefficient; however, unlike the MDS method, in which one trial is compared to all the other samples, the SIMPER analysis compares a single sample to each subsequent sample (*Żelazna-Wieczorek 2011*). The results indicate the species

which most strongly differentiated a sampled site from the others, and to what extent. A species was regarded as being characteristic of the studied ecosystem if it was characterized by a mean dissimilarity greater than or equal to 2 according to the SIMPER analysis, and an mean abundance greater in one ecosystem than the other.

In total, 19 physical and chemical parameters of water were measured in the studied ecosystems. The results of the correlation analysis found 15 physical and chemical parameters to be indicative of environmental conditions. The selected parameters were subjected to principal component analysis (PCA) to determine which had the strongest effect on the studied diatom habitats.

Using the information from the OMNIDIA database, the environmental conditions for each sampling point were determined according to Van Dam et al. (*1994*). Following this, the percentage share of diatom species included in each ecological value class was indicated. For species found to be characteristic of the studied ecosystems, classes of ecological values were assembled. Taxa which had at least one value of 0 (unknown) were selected for further analysis.

The selected ecological values according to Van Dam et al. (*1994*) were verified, or new ones established, for the species found to be characteristic of the studied ecosystems according to three premises: previous literature reports about ecological indicator values of those species, chemical and physical conditions analysis, and the classification of the environmental conditions according to Van Dam et al. (*1994*).

The analyses were performed using PRIMER 7.0.13, OMNIDIA 6.0.6 and STATISTICA 13 software.

Results & Discussion

Chemical analysis of water samples

The mean values and range of all tested parameters are given in Table 1.

The PE hydrological complex was characterized by elevated values of electrolytic conductivity, reaching as high as 9230 μS L^{-1}. The pH changed with the direction of water outflow: a slightly acidic reaction was observed in the outflow and an alkaline one in the pond. Due to the geological profile of the region, the water flowing out of the well contained a high concentration of chloride ions, whose gradient decreased with the flow of water through the ditch to the pond. In addition, higher concentrations of the cations Mg^{2+}, Ca^{2+}, Na^{+} and K^{+} were observed compared to other ecosystems, as well as the anions HCO^{3-}, PO_4^{3-} and SO_4^{2-}.

Each of the sampling points in the PE complex was characterized by different chemical parameters, resulting in differences between the habitats. The highest electrolytic conductivity was noted in the outflow, which was mainly influenced by the concentrations of Cl^{-}, Na^{+} and HCO^{3-} ions. The maximum concentration of HCO^{3-} ions was recorded in D.PESB.250314; in the other locations, it did not exceed 410 mg L^{-1}.

Low concentrations of K^{+} ions were observed throughout the entire studied PE complex; however, maximum values were recorded in the locations characterised by the highest HCO^{3-} ion content. The highest concentration of Ca^{+} ions of all ecosystems was recorded in the outflow.

The ditch represented an intermediate section between the PE sampling points. However, as it is susceptible to periodic drying, limited chemical data was collected from this habitat and hence it was not possible to assess its chemical and physical nature.

The lowest electrolytic conductivity was found in the pond, which displayed lower concentrations of Cl^-, Na^+ and, to a lesser degree, HCO^{3-}. The pH of the water never dropped below 8, except in one case in March 2014. In the pond, the concentration of K^+ remained relatively unchanged, which could be related to the fact that the reservoir was also a receiver of waters flowing from the surrounding arable fields. The pond was also characterized by the lowest concentration of Ca^{2+} and Mg^{2+}. In the summer periods, a significant reduction in the water table level and occasional drying of the reservoir were noted.

The urban reservoirs in Łęczyca were characterized by a slightly alkaline water reaction, which was similar in all reservoirs during the course of the study. No elevated concentrations of Fe^{2+} / $^{3+}$ and Mn^{3+} ions were observed. The content of SO_4^{2-} anions was not higher than in other ecosystems studied. The concentration of HCO^{3-} ions was lower than that observed in BO and PE. No significant differences in chemical and physical parameters were observed between the individual sampling points constituting LE

The Bogdałów reservoir was characterized by an alkaline reaction. It K^+, Cl^- and NH^{4+} ion content was the lowest of the studied ecosystems.

Diatom samples

A total of 381 diatom taxa were identified in 44 benthic samples: 139 in PE, 192 in LE and 188 in BO. The dominant species in PE were *Navicula veneta*, and *Nitzschia frustulum*, in LE *Cyclostephanos dubius* and *Stephanodiscus hantzschii*, in BO *Achnanthidium minutissimum*, *Pantocsekiella ocellata* and *Mastogloia smithii*. In the examined ecosystems, the most commonly identified classes were accidental (PE-84; LE-111; BO-86), accessory (PE-25; LE-35) and euconstant taxa (BO-39) (Fig. 2).

MDS analysis

MDS analysis identified the variation between samples for each studied hydrological object (Fig. 3). The samples taken from BO constitute a separate cloud, with the samples demonstrating high similarity with each other, whereas the samples of D.LEP1, D.LEP2 and D.LEP3 constitute a distinct group, with no clear differentiation into individual reservoirs. In the case of PE, the pond group (D.PEPB) was found to be clearly distinct from the others.

Shade Plot

Shade Plot analysis identified 50 species which had the strongest influence on the degree of similarity, or non-similarity, between the samples in the studied ecosystems. Of these taxa, the three that most strongly influenced the similarity between the samples in at least two ecosystems were *Navicula veneta*, *N. cincta*, *N. gregaria, Nitzschia frustulum, N. inconspicua, N. palea*

PeerJ Preprints | https://doi.org/10.7287/peerj.preprints.27739v1 | CC BY 4.0 Open Access | rec: 17 May 2019, publ: 17 May 2019

Planothidium frequentissimum, Amphora pediculus, Cyclotella meneghiniana, Fragilaria radians and *Achnanthidium minutissimum* (Fig. 4).

SIMPER analysis

SIMPER analysis allowed 36 species characteristic of the tested hydrological objects to be distinguished. These included *Cocconeis placentula*, *Craticula buderi, Fragilaria sopotensis, Gomphonema parvulum, Hippodonta hungarica*, *Navicula cincta, Nitzschia palea* and *Planothidium frequentissimum* for D.PEPB; *Chamaepinnularia krookiformis, C. plinskii, Craticula halophila, Navicula veneta*, *Nitzschia frustulum, N. inconspicua, N. liebethruthii, N. perminuta* and *N. tubicola* for D.PEDB and *Fragilaria famelica, Navicula veneta*, *Nitzschia frustulum, N. inconspicua, Planothidium delicatulum,* and *Tabularia fasciculata* for D.PESB Characteristic species for LA are: *Achnanthidium minutissimum, Amphora pediculus, Cyclostephanos dubius, C. invisitatus, Cyclotella meneghiniana, Navicula gregaria, N. moskalii, Nitzschia palea, Stephanodiscus binatus, S. hantzschii* and *S. parvus.* For BO: : *Achnanthidium minutissimum, Diatoma moniliformis, Encyonopsis subminuta, Mastogloia smithii, Nitzschia dissipata* var. *media, Pantocsekiella ocellata* and *P. pseudocomensis.*
.In addition, two species were found to be as characteristic of two different ecosystems: *Achnanthidium minutissimum* for LE and BO, and *Nitzschia palea* for PE and LE.

Ecological analysis based on Omnidia software

The ecological analysis of diatom assemblages based on data obtained from the OMNIDIA program database, indicated the following:
- pH requirements: while alkaliphilic species predominate in PE (63%), a large percentage in D.PEPB are unknown (24%) or neutrophilic species (23%). The LE reservoirs were dominated by alkalibiontic (45%) and alkaliphilic (24%) organisms. In D.LEP1, 25% of species were unknown. BO was dominated by alkaliphilic (39%) and neutrophilic (29%) species, and 26 % of species were unknown (Fig 5a);
- salinity: the PE complex was characterized by the occurrence of halophilic (43%), oligohalobous (30%) and mesohalobous species (16%); the greatest proportion of the mesohalobous species were found in D.PESB (28%). The LE reservoirs were dominated by oligohalobous (44%) and halophilic species (42%). BO was dominated by oligohalobous (43%), halophobe (24%) and unknown species (23%) (Fig 5b).
- nitrogen uptake: the most common species in the PE complex N-autotrophic tolerant (39%) followed by unknown (25%). The largest percentage of unknown species (28%) was recorded in D.PEPB and D.PEDB. In the LE reservoirs, the most common groups of species were N-autotrophic (57%) and unknown (25%). In BO, 51% species were unknown, 24% were N-autotrophic tolerant and 22% N-autotrophic sensitive (Fig 5c);
- oxygen requirements: in PE, the largest groups of species were low oxygen (30%), unknown (27%) and moderate oxygen (24%). In LE, oxybiontic species were most common (43%)

followed by unknown (25%). In BO, unknown (46%) and polyoxybiontic species (42%) predominated (Fig 5d);

- sensitivity to saprobity: in PE, the largest group of taxon were α-meso: polysabrobe (28%) and unknown (23%). In D.PEPB, the most abundant was α-meso-polysabrobe (34%) followed by β-mesosaprobe (31%) and unknown (27%). LE primarily included taxa from the α-mesosaprobe group (47%) and unknown (23%). In BO, unknown (34%), β-mesosaprobe (31%) and oligosaprobe taxa (28%) predominated (Fig 5e);
- trophic status: in PE, the largest group of diatoms were eutrophic (50%) and unknown taxa (25%), LE had the highest percentage (61%) of eutrophic species but also unknown (15%) and hypetrophic (13%) were present. In BO, the most abundant species were unknown (42%), indifferent (19%) and meso-eutrophic (16%) (Fig 5f);
- moisture aerophily: in PE, the largest group was aquatic to aerophilic (56%), representing 66% of species in D.PESB, 61% in D.PEDB, and 42% in D.PEPB. The second largest group was unknown (23%), constituting 32% of taxa in D.PEPB. In LE, 37% of the species were aquatic (24% of taxa in D.LEP2), 54% were occasionally aerophilic and 22% were unknown. In BO, the predominant groups of species were unknown (44%) and aquatic to aerophilic (33%) (Fig 5g).

Characteristic species: OMNIDIA and PCA analysis

The analysis of species characteristic of the tested ecosystems, determined according to Van Dam et al. (*1994*), identified 16 taxa classified as 0 in at least one category (Table 2). The next step determined the percentage contribution of each of these species classified as class 0 for the ecological parameters defined by Van Dam et al. (*1994*) at each sample point (Table 3). Based on the PCA analysis of the 16 characteristic taxa and selected physical and chemical parameters, the following relationships were demonstrated:

- Group A: *Chamaepinnularia krookiformis*, *C. plinskii*, *Nitzschia liebethruthii* and *Planothidium delicatulum* demonstrate a negative correlation with pH and a positive correlation with a decrease in the concentrations of HCO^{3-} Ca^{2+} $Fe^{2+}/^{3+}$ (Fig. 6).
- Group B: *Craticula buderi*, *Planothidium frequentissimum* and *Navicula cincta* did not demonstrate any relationship with any water parameters (Fig. 6).
- Group C: *Navicula moskalii*, *Cyclostephanos invisitatus*, *Stephanodiscus parvus*, *S. binatus*, *Diatoma moniliformis*, *Nitzschia dissipata* var. *media*, *Mastogloia smithii*, *Pantocsekiella pseudocomensis* and *Encyonopsis subminuta* demonstrated a negative correlation with a decrease in electrolyte conductivity, as well as with the concentrations of K^{+}, Mg^{2+}, Na^{+}, SO_4^{2-}, Cl^{-}, PO_4^{3-} and Mn^{3+} and water pigments (Fig. 6).

Ecological values of characteristic species

Characteristic species for PE

***Planothidium delicatulum* (PTDE) (Figs 7. A1-A6)**

Planothidium delicatulum is a euconstant taxon for PE and an accidental taxon for LE. Its mean percentage share in PE was 2%, and constituted 5% in D.PESB.
Planothidium delicatulum does not currently have six ecological values according to Van Dam et al. (*1994*).
This species was more abundant in environments such as D.PESB, which was also characterized by the highest concentration of Cl^- (up to 2976 mg L^{-1}), elevated electrolytic conductivity, and decreased K^+ concentration. The pH of the water in which this species was observed did not exceed 7.
Planothidium delicatulum was mainly recorded in salty and brackish environments with neutral or slightly alkaline conditions (*Campeau, Pienitz & Héquette 1999; Gell et al. 2005; Caballero et al. 2013; Yamamoto, Chiba & Tuji 2017; Van de Vijver et al. 2018*).
Based on our findings, we suggest changing the following ecological values in the Van Dam et al. (*1994*) classification for *Planothidium delicatulum*:
- pH requirements: 3 (neutrophilic) (changing from 5 to 3);
- salinity: 5 (brackish-marine) (changing from 4 to 5)

***Chamaepinnularia krookiformis* (CHKF) (Figs 7. B1-B5) and *Chamaepinnularia plinskii* (CHPL) (Figs 7. C1-C5)**

In 2016 *Chamaepinnularia krookiformis* was divided into two separate taxa: *C. krookiformis* and *C. plinskii* (*Żelazna-Wieczorek & Olszyński 2016*). Both species were very often recorded together in the same ecosystem. However, the publications which identified *C. krookiformis* often do not provide appropriate photographic documentation or photos of individual specimens (*Witkowski 1994, Bąk et al. 2006; Wojtal 2008; Peszek et al. 2015*). Currently available documentation is insufficient to determine whether *C. krookiformis* and *C. plinskii* are both present simultaneously in a given environment or whether just one of these species exists.
Chamaepinnularia krookiformis is an accessory taxon for PE (a constant taxon for D.PEDB), *C. plinskii* is a constant taxon for PE (a euconstant taxon for D.PEDB). The mean share of *C. krookiformis* was 1.6% in all PE samples, 4% in D.PEDB; for *C. plinskii,* this amounted to 2.7% in PE, 6% in D.PEDB.
Currently, *C. krookiformis* lacks three assigned ecological values. For PE, it constitutes 6% of the *unknown* group in nitrogen *uptake*, 6% in oxygen requirements and 6% in trophic state (respectively for D.PEDB: 16%, 16% and 15%). *C. plinskii* has no assigned ecological values and represents 26% of the *unknown* group for pH requirements, 19% for salinity, 12% for nitrogen uptake, 10% for *oxygen requirements*, 14% for saprobity, 11% for trophic state and 10% for moisture (respectively for D.PEDB: 60%, 25%, 24%, 23%, 32%, 23% and 28%).
The conditions of the environments in which both species have been recorded indicate that they are class 3 with regard to pH range (neutrophilic). Both species were the most abundant in locations subjected to periodic drying and characterised by high concentrations of chloride ions (up to 1006 mg L^{-1}) indicating a brackish environment (*Żelazna-Wieczorek, Olszyński & Nowicka-Krawczyk 2015*)

On the basis of our findings and those of previous studies (*Krammer & Lange–Bertalot 1986; Krammer 1992; Witkowski 1994; Bąk, Witkowski & Lange-Bertalot 2006; Wojtal 2009; Peszek et al. 2015; Żelazna-Wieczorek & Olszyński 2016*), we propose the following changes to the ecological values according to Van Dam et al. (*1994*) for *Chamaepinnularia krookiformis* and *C.* plinskii:
- pH requirements: 3 (neutrophilic) for both species;
- salinity: 4 (mesohalobous) established for *C. plinskii*, and changing from 3 to 4 for *C. krookiformis*;
- trophic state: 5 (eutrophic) for both species;
- moisture aerophily: 4 (aerophilic) established for *C. plinskii* and changing from 3 to 4 for *C. krookiformis*.
- saprobity: from 2 to 4 (β-mesosaprobe to α-meso-polysabrobe) for both species. Due to the specific conditions and locations of the studied objects, they were exposed to large fluctuations in the inflow of organic matter, mainly from runoff from arable fields and pollution caused by animal grazing. These impurities were manifested as elevated concentrations of K^+ ions. Therefore, we believe that classifying *C. krookiformis* as an oligosaprobe is inappropriate. Further tests are needed to determine the optimum occurrence of these species in areas subjected to organic matter loads.

Nitzschia liebethruthii (NLBT) (Figs 7. D1-D4)

Nitzschia liebethruthii is a euconstant taxon for PE. Its mean percentage share was 4% in the PE samples, and 10% in the D.PEDB samples. It was most numerous in the sample D.PEDB.301113 (19%). This species has two specific ecological values. The ecological value analysis for PE found *N. liebethruthii* to represent 17% of the *unknown* group in nitrogen uptake, 16% in oxygen requirements, 18% in saprobity, 17% in trophic state and 21% in moisture (respectively for D.PEDB: 37%, 37%, 43%, 36% and 49%).
Nitzschia liebethruthii occurred in environments subjected to periodic drying with a pH close to 7 and high concentration of chloride ions.
This species was noted in environments with increased salinity, electrolytic conductivity and high pH value (*Rumrich, Lange-Bertalot & Rumrich 2000; Witkowski, Lange-Bertalot & Metzeltin 2000; Lange-Bertalot et al. 2017; Földi et al. 2018*).
We propose the following changes to the ecological values according to Van Dam et al. (*1994*) assigned to *Nitzschia liebethruthii*:
- pH requirements: 3 (neutrophilic) (changing from 5 to 3);
- saprobity: from 2 to 4 (β-mesosaprobe to α-meso-polysabrobe) as in the case of *C. krookiformis* and *C. plinskii*;
- trophic state: 5 (eutrophic);
- moisture aerophily: 4 (aerophilic).

Craticula buderi (CRBU) (Figs 7. E1-E5)

Craticula buderi is a euconstant taxon for PE and an accidental taxon for LE. Its mean percentage share was 4.3% in all samples for PE, and 12% for D.PEPB. This species has no recorded ecological values. The ecological value analysis for the PE found *C. buderi* to constitute 19% of the *unknown* group in pH requirements, 28% in salinity, 16% in nitrogen uptake, 16% in oxygen requirements, 16% in saprobity, 19% in trophic state and 15% in moisture (respectively for D.PEPB: 53%, 73%, 46%, 46%, 46%, 54% and 41%).

Although *Craticula buderi* was classified into group B, it was found to be most abundant in environments with an elevated concentration of Cl^- ions, ranging from 685 to 1090 mg L^{-1}, (all samples from D.PEPB and one sample from D .PEDB in which the concentration of chloride ions was 1006 mg L^{-1}). However, its numbers were decreased in the D.PEPB sample, which was characterised by a chloride ion content of over 1500 mg L^{-1}. Interestingly. the concentration of K^+ ions exceeded 100 mg L^{-1} at Cl^- concentrations below 1500 mg L^{-1}; therefore, it is possible that the decline of this species could be related to the concentration of K^+ ions alone. Our observations indicate that the population of *C. buderi* from D.PEPB favours a concentration of chloride ions from 500 to 1006 mg L^{-1} which coincides with a K^+ ions concentration from 50 to 70 mg L^{-1}.

Craticula buderi is widespread throughout the world and recognized as cosmopolitan (*Rumrich, Lange-Bertalot & Rumrich 2000; Lange-Bertalot 2001; Bahls 2009; Soltanpour-Gargari, Lodenius & Hinz 2011, Żelazna-Wieczorek 2011; Cichoń 2016*). This species was found to be dominant in environments characterised by increased electrolytic conductivity and an alkaline water reaction (*Holmes & Taylor 2015*). Holmes and Taylor (*2015*) place *C. buderi* in the Bad in Water Quality class. Their recorded values of diatom indices indicate that the environment was eutrophic.

We therefore propose the following classes of ecological values according to Van Dam et al. (*1994*) for *Craticula buderi*:

- pH requirements: 4 (alkaliphilic);
- sabrobity: from 2 to 4 (β-mesosaprobe to α-meso-polysabrobe) as in the case of *C. krookiformis* and *C. plinskii*;
- trophic state: 5 (eutrophic);
- salinity: 4 (mesohalobous)
- moisture aerophily: 3 (aquatic to aerophilic).

Navicula cincta (NCCA) (Figs 7. F1-F5)

Navicula cincta is a euconstant taxon for PE, a constant taxon for LE and an accessory taxon for BO. The mean percentage share of this species for PE is 3%, of which 7% was found in D.PEPB samples.

Currently this species has been assigned two ecological values. The ecological values analysis for the PE found *N. cincta* to constitute 24% of the *unknown* group in pH requirements, 13% in nitrogen uptake, 12% in oxygen requirements, 13% in saprobity, and 12% in moisture (respectively for D.PEPB: 28%, 25%, 25%, 26% and 23%).

An analysis of the physical and chemical data and the variability of occurrence did not show any clear relationships between environmental parameters and the percentage share of *Navicula cincta* in the tested samples. This lack of dependence is also confirmed by the PCA analysis. *Navicula cincta* has been recorded in various types of ecosystems, although mainly in eutrophic ones with high conductivity. It also tolerates elevated levels of organic matter. This species was also observed in habitats subjected to periodic drying (*Lange-Bertalot & Genkal 1999; Rumrich, Lange-Bertalot & Rumrich 2000; Witkowski, Lange-Bertalot & Metzeltin 2000; Lange-Bertalot 2001; Żelazna-Wieczorek 2011; Wojtal 2013; Lange-Bertalot, Hofmann & Werum 2017*). However, several new species from the group *N. cincta* s.l. have been described, and it can be assumed that each of these individual species in this group may be associated with narrower optimal ecological conditions (*Cantonati et al. 2016*).

Based on our present findings, and those of previous studies, we believe that it is not appropriate to classify *Navicula cincta* as an oligohalobous species with regard to *salinity*: it has been recorded in fresh (*Żelazna-Wieczorek 2011; Wojtal 2013*), brackish (*Żelazna-Wieczorek, Olszyński & Nowicka-Krawczyk 2015; Żurek et al. 2018*) and salt waters (*Witkowski, Lange-Bertalot & Metzeltin 2000*). We propose the following ecological value according to Van Dam et al. (*1994*) for *Navicula cincta* s.l.:

- moisture aerophily: 3 (aquatic to aerophilic).

Shade Plot analysis found that the presence of *N. cincta* s.l. can falsely indicate high similarity between samples from different environments, thus distorting the results of any environmental analysis. Therefore, with regard to the unclear taxonomic status of *Navicula cincta* s.l. and the current lack of knowledge regarding its activities, we recommend this taxon be excluded from the biological assessment of surface water quality.

***Planothidium frequentissimum* (PLFQ) (Figs 7. G1-G5)**

Planothidium frequentissimum is a euconstant taxon for PE and LE. The mean percentage of this species for PE is 2.7%, of which 5% was found in D.PEPB samples. It was most numerous in the D.PEDB.301113 sample (19%). The species has currently six established ecological values. The ecological values analysis for PE found *P. frequentissimum* to constitute 12% of the *unknown* group in moisture aerophily (for D.PEPB 14% and D.PESB: 20%).

No relationship was observed between percentage share of *Planothidium frequentissimum* and the changes in chemical and physical parameters in the tested samples. This lack of relationship was confirmed by PCA analysis.

Planothidium frequentissimum is an eurytopic species that occurs globally in a variety of habitat types, from natural springs to rivers in urban areas with high levels of pollution. Its value as an indicator is low, as confirmed by the Shade Plot analysis, which found it to significantly affect the degree of similarity observed between samples from different environments. We therefore recommend that *P. frequentissimum* be excluded from the biological assessment of surface water quality (*Siver, et al. 2005; Levkov et al. 2007; Żelazna-Wieczorek 2011; Kulikovskiy, Lange-Bertalot & Kuznestova 2015; Szczepocka, Nowicka-Krawczyk & Kruk 2018*).

Characteristic species for LE

***Cyclostephanos invisitatus* (CINV) (Figs 8. A1-A6)**

Cyclostephanos invisitatus is a euconstant taxon for LE. Its percentage share for LE was 4.8%. *C. invisitatus* currently has two ecological values assigned. The ecological values analysis for LE found it to constitute 24% of the *unknown* group in pH requirements, 17% in nitrogen uptake, 18% in oxygen requirements, 19% in saprobity, and 20% in moisture. *Cyclostephanos invisitatus* occurs in diverse environments, however, it is most frequently reported in aquatic ecosystems subjected to high human impact characterised by an alkaline reaction and increased conductivity (*Reavie & Smol 1998; Yang et al. 2005; Wojtal & Kwandrans, 2006; Kiss et al. 2012; Houk, Klee & Tanaka 2014; Reavie & Kireta 2015; Olszyński & Żelazna-Wieczorek 2018*).
We therefore propose that the following classes of ecological values according to Van Dam et al. (*1994*) be established for *Cyclostephanos invisitatus*:
- pH requirements: 4 (alkaliphilic);
- moisture aerophily: 1 (aquatic);

***Navicula moskalii* (NMOK) (Figs 8. B1-B6)**

Navicula moskalii is an accessory taxon for LE. Its mean percentage share for LE was 1.5%. Its incidence was greatest in sample D.LEP1.250315 (26%). *Navicula moskalii* has no assigned ecological values. The ecological value analysis for LE found it to constitute 5% of the *unknown* group in pH requirements, 7% in salinity, 5% in nitrogen uptake, 5% in oxygen requirements, 5% in saprobity, 6% in trophic state and 5% in moisture. The greatest occurrence of *N. moskalii* was observed in samples with the highest concentrations of Ca^{2+} (143.6 mg L^{-1}), HCO^{3-} (338.6 mg L^{-1}), SO_4^{2-} (146.9 mg L^{-1}) and with high Mg^{2+} content. *Navicula moskalii* was observed in a number of ecosystems (*Metzeltin & Witkowski 1996; Lange-Bertalot 2001; Żelazna-Wieczorek 2011; Noga et al. 2016; Lange-Bertalot, Hofmann & Werum 2017*), particularly in eutrophic waters with an elevated level of Ca^{2+} and HCO^{3-} ions. Żelazna-Wieczorek (*2011*) report a significant number of *N. moskalii* in springs with high levels of eutrophication, however with Ca^{2+}, SO_4^{2-}, HCO^{3-} and Mg^{2+} concentrations lower than those in the LE samples.
We therefore propose that the following classes of ecological values according to Van Dam et al. (*1994*) be established for *Navicula moskalii*:
- pH requirements: 4 (alkaliphilic);
- salinity: 2 (oligohalobous);
- trophic state: 7 (indifferent)

***Stephanodiscus binatus* (SBNT) (Figs 8. C1-C5)**

Stephanodiscus binatus is a euconstant taxon for LE. Its percentage share for LE was 4.3%. *S. binatus* has no recorded ecological values. The ecological value analysis for LE found it to constitute 25% of the *unknown* group in pH requirements, 47% in salinity, 18% in nitrogen uptake, 18% in oxygen requirements, 20% in saprobity, 29% in trophic state and 21% in moisture.

The largest percentage share of *S. binatus* was recorded in the spring months and the lowest in autumn. Its abundance was found to be elevated in December 2014 and 2015; the same samples demonstrated the highest concentrations of Ca^{2+}, Mg^{2+} and the highest pH (above 8).

Stephanodiscus binatus has been recorded in various water ecosystems ranging from oligotrophic to eutrophic; however, all are characterised by elevated pH value (*Stoermer & Håkansson 1984; Håkansson & Kling 1990; Houk, Klee & Tanaka 2014; Olszyński & Żelazna-Wieczorek 2018*).

We therefore propose that the following classes of ecological values according to Van Dam et al. (*1994*) be established for *Stephanodiscus binatus*:

- pH requirements: 4 (alkaliphilic);
- salinity: 2 (oligohalobous);

Stephanodiscus parvus (SPAV) (Figs 8. D1-D5)

Stephanodiscus parvus is a euconstant taxon for LE and an accidental taxon for PE. Its percentage share for LE was 2.4%. It was most abundant in the D.LEP3.260714 sample (22%). This species has three assigned ecological values. The ecological values analysis for LE found the taxon to constitute 9% of the *unknown* group in nitrogen uptake, 9% in oxygen requirements, 10% in saprobity, and 10% in moisture.

S. parvus is noted mainly in eutrophic hypertrophic ecosystems with elevated electrolytic conductivity. It is also a good indicator of waters with a strong anthropogenic impact (*Reavie & Smol 1998; Reavie & Kireta, 2015; Olszyński & Żelazna-Wieczorek 2018; Reavie & Cai 2019*).

Based on our findings and literature data, we propose the following changes in ecological values according to Van Dam et al. (*1994*) for *Stephanodiscus parvus*:

- pH requirements: from 5 to 4 (alkaliphilic);

Characteristic species for BO

Diatoma moniliformis (DMOF) (Figs 9. A1-A6)

Diatoma moniliformis is a euconstant taxon for BO. Its mean percentage share for BO was 3.9%. It currently has no assigned ecological values. According to the ecological values analysis for BO, this taxon constituted 10% of the *unknown* group in pH requirements, 11% in salinity, 8% in nitrogen uptake, 8% in oxygen requirements, 9% in saprobity, 9% in trophic state and 8% in moisture.

D. moniliformis was found in 87.5 % of samples from BO. Interestingly, it constituted 28% of the share in one sample from December 2016 (D.BOZB.091216); however, its share was below

2% in the previous season, and was not higher than 1-2% in the other samples from December 2016. The chemical and physical characteristics of D.BOZB.091216 did not differ significantly from those of the other samples.
This species is also found in fresh and salt water, as well as the Baltic and arctic areas with high conductivity (*Potapova & Snoeijs 1997; Rumrich, Lange-Bertalot & Rumrich 2000; Levkov et al. 2007; Pniewski & Sylwestrzak 2018*).
One of the factors that influences the abundance of *D. moniliformis* is the water temperature. Studies indicate that temperatures above 10-15°C (*Potapova & Snoeijs 1997; Pniewski & Sylwestrzak 2018*) are associated with population growth. However, populations have been observed in freshwater streams and lakes in arctic areas, in which the temperature of the water is below 10°C (*Antoniades Douglas & Smol 2005*). Population growth was also observed at 4.7°C in sample D.BOZB.091216; therefore, low temperature may have an influence on the abundance of this species.

Encyonopsis subminuta **(ESUM) (Figs 9. B1-B6)**

Encyonopsis subminuta is a euconstant taxon in BO, where its mean percentage share was 4.1%. Presently, *E. subminuta* has been assigned five ecological values. Ecological values analysis for BO found it to constitute 7% of the *unknown* group in *nitrogen uptake* and 8% in *moisture*.
Encyonopsis subminuta was found to be most abundant in sample D.BOZB.041115. The sample was also characterised by an elevated concentration of $Fe^{2+}/^{3+}$ ions and the lowest pH value. In subsequent samples, when the concentration of Fe ions dropped, the abundance of *E. subminuta* also decreased.
E. subminuta is regarded as a cosmopolitan taxon, occurring in the temperate and boreal zone. It is most abundant in oligo- to mesotrophic waters with electrolytic conductivity between 190- 250 $\mu S\ L^{-1}$ (*Krammer 1997; Noga et al. 2014; Novais et al. 2014; Feret, Bouchez & Rimet 2017*).
E. subminuta may be sensitive to the concentration of Fe ions; however, the increase of these ions is associated with a drop in pH. Our research confirms that the optimal pH for population size is close to 7.

Mastogloia smithii **(MSMI) (Figs 9. C1-C7)**

Mastogloia smithii is a euconstant taxon for BO. Its mean percentage share for BO was 6.3%. it was found in greatest numbers in D.BOZB.300615 (22%) and D.BOZB.261016 (15%). *M. smithii* has been assigned four ecological values. The ecological values analysis for the BO found this species to constitute 13% of the *unknown* group in nitrogen uptake, 14% in oxygen requirements and 15% in trophic state.
The environment in BO regarding *salinity* was classified according to Vam Dam et al. (*1994*) as oligohalobus (43% species); however, 7% of the mesohalobous species were represented by one species: *Mastogloia smithii*.
This species is recorded in fresh, brackish and salt water (*Witkowski, Lange-Bertalot & Metzeltin 2000; Busse & Snoeijs 2003; Weckström & Juggins 2005; Martinzes-Goss & Evangelista 2011;*

Lange-Bertalot, Hofmann & Werum 2017). Its presence in environments with varying degrees of salinity may suggest that this does not have an significant influence on population size.
Based on our present findings and literature data, we propose the following changes in the ecological values according to Van Dam et al. (*1994*) for *Mastogloia smithii*:
- salinity: from 4 to 3 (halophilic).

Nitzschia dissipata var. _media_ (NDME) (Figs 9. D1-D5)

Nitzschia dissipata var. *media* is a euconstant taxon for BO. Its mean percentage share for BO was 3.8%, and the highest proportion (20%) was found in D.BOZB.041115. *N. dissipata* var. *media* has been assigned two ecological values. Ecological values analysis for BO found it to constitute 9% of the *unknown* group in nitrogen uptake, 10% in oxygen requirements, 11% in saprobity, 11% in trophic state and 9% in moisture.
The growth in occurrence of *Nitzschia dissipata* var. *media* is associated with an increase in the level of $Fe^{2+}/^{3+}$ ions , similar to *Encyonopsis subminuta*. In addition, it was found in the ecosystem, i.e. BO, with the lowest concentrations of ions indicative of the presence of organic pollutants in the environment, such as K^+ and NH^{4+}.
Although *Nitzschia dissipata* var. *media* is found sporadically, it is commonly found in oligo- to mesotrophic waters with a pH between 7 and 8 (*Van der Vijver, Frenot & Beyens 2002; Antoniades, Douglas & Smol 2005; Żelazna-Wieczorek 2011; Lange-Bertalot, Hofmann & Werum 2017*).
Based on our findings and literature data, we propose the following ecological values according to Van Dam et al. (*1994*) for *Nitzschia dissipata* var. *media*:
- saprobity: 2 (β-mesosaprobe)

Pantocsekiella pseudocomensis (PPCS) Figs (9. E1-E7)

Pantocsekiella pseudocomensis is a euconstant taxon for BO. Its mean percentage share for BO was 4.4%. It was most abundant in D.BOZB.250315 (9%) and in D.BOZB.220616 (10%). *P. pseudocomensis* has not been assigned any ecological values according to Van Dam et al. (*1994*). The ecological values analysis for BO found it to represent 22% of the *unknown* group in pH requirements, 25% in salinity, 10% in nitrogen uptake, 11% in oxygen requirements, 18% in saprobity, 12% in trophic state and 13% in moisture.
The greatest amount of *P. pseudocomensis* was found in samples characterised by the highest levels of ammonium ions. Its percentage share was lowest in samples with the lowest water temperature, apart from D.BOZB.250315.
Currently, *Pantocsekiella pseudocomensis* is assigned to the *P. comensis* complex, with *P. comensis* and *P. costei*. We believe that assigning ecological values for particular species of the *P. comensis* complex is unjustified at the current state of knowledge, and that all species within the complex should be assigned the same provisional ecological values until their individual properties are better understood (*Houk, Klee & Tanaka 2010; Kistenich et al. 2014; Duleba et al. 2015*).

Conclusions

The water ecosystems created in the post-mining areas create a complex of conditions that are not found in other natural ecosystems, and the benthic diatom species present in such environments are very often present in higher numbers than in other habitats. The specific hydro-geological conditions prevailing in the post-production reservoirs provide a unique opportunity to observe interspecies differences and intra-species variability, allowing for the verification or isolation of new taxa and a greater insight into their autecology (*Żelazna-Wieczorek & Olszyński 2016; Olszyński & Żelazna-Wieczorek 2018*).

The identification of species characteristic of the studied ecosystems may foster further growth of ecological research and increase the reliability of surface water quality assessment, as such knowledge is needed to verify their ecological indicator values, and hence calculate diatomaceous indexes with greater accuracy.

Ecological values as set out by Van Dam et al. (*1994*) are utilized in many ecological works describing the ecological conditions of the studied ecosystems. These ecological values form the basis for calculating diatomaceous indexes describing the ecological state of surface waters. It is therefore necessary to constantly update and establish new ecological indicator values for particular diatom species.

Many authors who describe new species, or encounter existing species in new ecosystems, regularly propose updates for individual ecological indicator values. However, these findings, may not be introduced and updated in the OMNIDIA program for a number of years. The OMNIDIA system is used by state institutions in many countries around the world to assess surface water quality, (*Campeau, Pienitz & Héquette 1999; Rumrich, Lange-Bertalot & Rumrich 2000; Witkowski, Lange-Bertalot & Metzeltin 2000; Gell et al. 2005; Potapova & Ponader 2008; Caballero et al. 2013; Żelazna-Wieczorek & Olszyński 2015; Yamamoto, Chiba & Tuji 2017; Lange-Bertalot, Hofmann & Werum 2017; Földi et al. 2018; Van de Vijver, Wetzel & Ector 2018*). Clearly, if these assessments are based on incomplete or outdated data, assessments of aquatic environments may be fraught with error.

The present study used three principles to identify proposed changes in the classification of ecological indicator values for characteristic species according to the Van Dam et al. (*1994*) system, or to establish new values which were previously absent: the analysis of environmental conditions prevailing in the studied ecosystems, the analysis of relevant literature data, and references to the ecological indicator values of other species. This mode of research can serve as a model for updating databases used to assess surface water quality.

References

Antoniades D., Douglas M.S.V. & Smol J.P. 2005. Benthic diatom autecology and inference model development from the Canadian High Arctic Archipelago. *Journal of Phycology* 41: 30-45 DOI: 10.1111/j.1529-8817.2005.04049.x.

Bahls L.L. 2009. A checklist of diatoms from inland waters of the northwestern United States. *Proceedings of The Academy of Natural Sciences of Philadelphia* 158: 1-35 DOI: 10.1635/053.158.0101.

Bąk M. Witkowski A. & Lange–Bertalot H. 2006. Diatom flora diversity in the strongly eutrophicated and β–mesosaprobic waters of the Szczecin Lagoon, NW Poland, southern Baltic Sea. – In: Ognjanova–Rumenova N. & Manoylov K., eds. *Advances in Phycological Studies, Festschrift in Honour of Professor Dobrina Teminskova–Topalova*. Sofia – Moscow: Pensoft Publishers & University Publishing House, 293–317.

Busse S. & Snoeijs P. 2003. Gradient responses of diatom communities in the Bothnian Sea (northern Baltic Sea), with emphasis on responses to water movement. *Phycologia* 42(5): 451-464 DOI: 10.2216/i0031-8884-42-5-451.1.

Caballero M., Rodríguez A., Vilaclara G., Ortega B., Roy P. & Lozano S. 2013. Hydrochemistry, ostracods and diatoms in a deep, tropical, crater lake in Western Mexico. *Journal of Limnology* 72(3): 512-523 DOI: 10.4081/jlimnol.2013.e42.

Campeau S, Pienitz R. & Héquette A. 1999. Diatoms from the Beaufort Sea coast, southern Arctic Ocean (Canada). In: Lange-Bertalot H. & Kociolek P., eds. *Bibliotheca Diatomologica Vol. 42*. Berlin, Stuttgart: J. CRAMER 244 p.

Cantonati M., Angeli N. Spitale D. & Lange-Bertalot H. 2016. A new *Navicula* (Bacillariophyta) species from low–elevation carbonate springs affected by anthropogenic disturbance. *Fottea, Olomouc* 16(2): 255-265 DOI: 10.5507/fot.2016.013.

Cichoń S. 2016. *Craticula buderi* (Bacillariophyceae) in Poland. *Polish Botanical Journal* 61(2): 301–305 DOI: 10.1515/pbj-2016-0024.

Clarke K.R. & Gorley, R.N. 2015. PRIMER v7: User Manual/Tutorial. Devon, United: Kingdom: PRIMER-E Ltd 296 pp.

De Haan H., Van Liere L., Klapwijk Sj.P. & Van Donk E. 1993. The structure and function of fen lakes in relation to water table management in The Netherlands. *Hydrobiologia* 265: 155–177 http://dx.doi.org/10.1007/BF00007266.

Duleba M., Kiss K.T., Földi A., Kovács J., Borojević K.K., Molnár L.F., Plenković-Moraj A., Pohner Z., Solak C.N., Tóth B. & Ács E. 2015. Morphological and genetic variability of assemblages of *Cyclotella ocellata* Pantocsek/*C. comensis* Grunow complex (Bacillariophyta, Thalassiosirales), *Diatom Research* 30(4): 283-306 DOI: 10.1080/0269249X.2015.1101402.

European Union. 2000. Directive 2000/60/EC of the European Parliament and of the Council of 23 October 2000 establishing a framework for community action in the field of water policy. Official Journal of the European Communities L327, 1–73.

Feret L., Bouchez A & Rimet F. 2017. Benthic diatom communities in high altitude lakes: a large scale study in the French Alps. *Annales de Limnologie - International Journal of Limnology* 53: 411-423 DOI: 10.1051/limn/2017025.

Földi A., Ács É., Grigorszky I., Ector L., Wetzel C.E., Várbíró G., Kiss K.T, Dobosy P., Trábert Z., Borsodi A.K. & Duleba M. 2018. Unexpected consequences of bombing. Community level response of epiphytic diatoms to environmental stress in a saline bomb crater pond area. *PLoS ONE* 13(10): e0205343 https://doi.org/10.1371/journal.pone.0205343.

Gabryś-Godlewska A., Gruszecki J., Kochanowska J., Kozula R., Pasieczna A. & Tomassi-Morawiec H. 2004. Arkusz Turek (550). In: Sikorska-Maykowska M., Zdanowski A. & Gabryś-Godlewska A., eds. *Objaśnienia do mapy geośrodowiskowej Polski 1:50 000*. Warszawa: Państwowy Instytut Geologiczny. Opracowanie Zamówione Przez Ministra Środowiska. p 30.

Gadomska W., Kaźmierczak J., Merdzińska M., Zielińska B., Wiśniewska-Zientara G., Gajda M., Opas Z., Cieślak J., Kończak D., Kujawa S., Lukasiewicz J., Zielińska-Adasiak S., Ratajczyk G., Michalska R. & Sobczak D. 2007. Raport o stanie gminy Brudzew lata 2002-2006. Urząd Gminy Brudzew. Brudzew. p. 15.

Gell P.A., Bulpin S., Wallbrink P., Hancock G. & Bickford S. 2005. Tareena Billabong – a palaeolimnological history of an ever-changing wetland, Chowilla Floodplain, lower Murray-Darling Basin, Australia. *Marine and Freshwater Research* 56: 441-456 DOI: 10.1071/MF04107.

Harding W.R. & Taylor J.C., 2014. Diatoms as indicators of historical water quality: A comparison of samples taken in the Wemmershoek catchment (Western Province, South Africa) in 1960 and 2008. *Water S.A*. 40 (4): 601–606.

Håkansson H. & Kling H. 1990. The current status of some very small freshwater diatoms of the genera *Stephanodiscus* and *Cyclostephanos*. *Diatom Research* 5: 273-287 DOI:10.1080/0269249X.1990.9705119.

Houk, V., Klee R. & Tanaka H. 2010. Atlas of freshwater centric diatoms with a brief key and descriptions. Part III. Stephanodiscaceae A. *Fottea (Supplement)* 10:1–498.

Houk V., Klee R. & Tanaka H. 2014. Atlas of freshwater centric diatoms with a brief key and descriptions. Part IV. Stephanodiscaceae B. *Fottea (Supplement)* 14:1–532.

Holmes M. & Taylor J.C. 2015 Diatoms as water quality indicators in the upper reaches of the Great Fish River, Eastern Cape, South Africa. *African Journal of Aquatic Science* 1-17 DOI: 10.2989/16085914.2015.1086722.

Hutorowicz A. & Pasztalenic A. 2014. Phytoplankton metric of ecological status assessment for polish lakes and its performance along nutrient gradient. *Polish Journal of Ecology* 62(3): 525-540 DOI:10.3161/104.062.0312.

Kahlert M., Ács E., Almeida S.F.P., Blanco S., Dreßler M., Ector L., Karjalainen S.M., Liess A., Mertens A., van der Wal J., Vilaste S. & Werner P. 2016. Quality assurance of diatom counts in Europe: towards harmonized datasets. *Hydrobiologia* 772: 1–14 DOI: 10.1007/s10750-016-2651-8.

Kelly M.G, Juggins S., Guthrie R., Pritchard S., Jamieson J., Rippey B., Hirst H. &Yallop M. 2008. Assessment of ecological status in U.K. rivers using diatoms. *Freshwater Biology* 53 (2): 403–422 DOI: 10.1111/j.1365-2427.2007.01903.x.

Kiss K.T, Klee R., Ector L. & Ács É. 2012. Centric diatoms of large rivers and tributaries in Hungary: morphology and biogeographic distribution. *Acta Botanica Croatica* 71: 311–363 DOI: 10.2478/v10184–011–0067–0
Kistenich S., Dreßler M., Zimmermann J., Hübener T., Bastrop R. & Jahn R. 2014. An investigation into the morphology and genetics of *Cyclotella comensis* and closely related taxa. *Diatom Research* 29(4): 423-440 DOI: 10.1080/0269249X.2014.922125.
Kolada A., Pasztaleniec A., Bielczyńska A. & Soszka H. 2016. Phytoplankton, macrophytes and benthic diatoms in lake classification: consistent, congruent, redundant? Lessons learnt from WFD-compliant monitoring in Poland. *Journal of Limnology* 59: 44–52 DOI: 10.1016/j.limno.2016.05.003.
Krammer K. 1992. Pinnularia. Eine Monographie der europäischen Taxa In: *Bibliotheca Diatomologica 26*. Berlin–Stuttgart: J. Cramer. 353 pp.
Krammer K. 1997. Die cymbelloiden Diatomeen. Eine Monographie der weltweit bekannten Taxa. Teil 2. *Encyonema* part,. *Encyonopsis* and *Cymbellopsis*. In:. Lange-Bertalot H. & Kociolek P., eds. *Bibliotheca Diatomologica Vol. 37*. Berlin, Stuttgart: J. CRAMER. 96 pp.
Krammer K. & Lange-Bertalot H. 1986. Bacillariophyceae. 1. Teil: Naviculaceae In: Ettl H.H, Gerloff J., Heynig H. & Mollenhauer D., eds. *Süsswasserflora von Mitteleuropa, Band 2/1*. Jena: Gustav Fisher Verlag. 876 pp
Kulikovskiy M.S., Lange-Bertalot H. & Kuznetsova I.V. 2015. Lake Baikal: Hotspot of Endemic Diatoms II. In: Lange-Bertalot H., ed. *Iconographia Diatomologica Vol. 26. Taxonomy-Biogeography-Diversity.* Germany: Koeltz Scientific Books. 656 pp.
Lange-Bertalot, H. 2001. *Navicula* sensu stricto. 10 Genera Separated from *Navicula* sensu lato. *Frustulia.* In: Lange-Bertalot H., ed. *Diatom of Europe Vol. 2*. Ruggell:vA.R.G. Gantner Verlag K.G. 526 pp.
Lange-Bertalot, H. & Genkal, S.I. 1999. Diatoms from Siberia I. Islands in the Arctic Ocean (Yugorsky-Shar Strait). In: Lange-Bertalot H., ed. *Iconographia Diatomologica Vol. 6. Phytogeography-Diversity-Taxonomy*. Vaduz: A.R.G. Gantner Verlag K.G. 292 pp.
Lange-Bertalot H., Hofmann G., Werum M. & Cantonati M. 2017. Freshwater Benthic Diatoms of Central Europe: Over 800 Common Species Used in Ecological Assessment. Koeltz Botanical Books. 942 pp.
Lecointe C., Coste M. & Prygiel J. 1993. "Omnidia:" a software for taxonomy, calculation of diatom indices and inventories management. *Hydrobiologia* 269/270: 509-513.
Levkov Z., Krstic S., Metzeltin D. & Nakov T. 2007. Diatoms of Lakes Prespa and Ohrid. About 500 taxa from ancient lake system. In: Lange-Bertalot H., ed. *Inconografia Datomologica Vol. 16*. Ruggell: A.R.G. Gantner Verlag K.G. 613 pp.
Martinez-Goss M.R. & Evangelista L.T. 2011. A Contribution to the Taxonomy of *Mastogloia* (Class Bacillariophyceae) in the Philippines. *Philippine Journal of Science* 140(1): 7-12.

Metzeltin D. & Witkowski A. 1996. Diatomeen der Bären-Insel. Süβwasser- und marine Arten. In: Lange-Bertalot H., ed. *Iconographia Diatomologica. Annotated Diatom Micrographs. Vol 4. Taxonomy*. Königstein/Germany: Koeltz Scientific Books 20-21

Noga T., Stanek-Tarkowska J., Pajączek A., Kochman N. & Peszek Ł. 2014. Ecological assessment of the San River water quality on the area of the San Valley Landscape Park. *Journal of Ecological Engineering* 15(4): 12-22 DOI: 10.12911/22998993.1125453.

Noga T. Stanek-Tarkowska J. Rybek M., Kochaman-Kędziora N., Peszek Ł. & Pajączek A. 2016. Diversity of diatoms in the natural, mid-forest Terebowiec Stream – Bieszczady National Park. *Journal of Ecological Engineering* 17(4): 232-247 DOI: 10.12911/22998993/64506.

Novais M.H., Morais M.M., Rosado J., Dias L.S, Hoffmann L. & Ector L. 2014. Diatoms of temporary and permanent watercourses in Southern Europe (Portugal). *River Research and Applications* 30: 1216-1232 DOI: 10.1002/rra.2818.

Olaczek R. 1963. Zbiorowiska roślinne torfowisk niskich okolic Łęczycy. Unpublished doctoral dissertation, University of Lodz, Poland.

Olszyński R.M. & Żelazna-Wieczorek J. 2018. *Aulacoseira pseudomuzzanensis* sp. nov. and other centric diatoms from post iron ore mining reservoirs in Poland. *Diatom Research* 33(2): 155-185 DOI: 10.1080/0269249X.2018.1509886.

Orlikowski D. & Szwed L. 2009. Wodny kierunek rekultywacji w KWB „Adamów" SA – inwestycją w przyszłość regionu. *Górnictwo i Geoinżynieria* 33(2): 351-361.

Peszek Ł., Noga T., Stanek-Tarkowska J., Pajączek A., Kochman-Kędziora N., Pieniążek M. 2015. The effect of anthropogenic change in the structure of diatoms and water quality of the Żołynianka and Jagielnia streams. *Journal of Ecological Engineering* 16(2): 33-51 https://doi.org/10.12911/22998993/1856.

Pliński M. 1966. Glony słonego źródła w Pełczyskach pod Ozorkowem. Unpublished master dissertation, University of Lodz, Poland.

Pliński M. 1969. Okrzemki słonego źródła w Pełczyskach pod Ozorkowem. *Zesz. Nauk. UŁ* 2(31): 123-136.

Pliński M. 1971a. Rzadkie i nowe dla flory Polski gatunki glonów z terenów solnisk w okolicach Łęczycy. *Zesz. Nauk. UŁ* 2(41): 171-184.

Pliński M. 1971b. Gatunki z rodzaju Enteromorpha (Link.) Agardh z terenu solnisk podłęczyckich. *Zesz. Nauk. UŁ*. 2(41): 159-169.

Pliński M. 1971c. System halobow w świetle współczesnych poglądów. *Wiad. Ekol*. 17(1): 18-29.

Pliński M. 1973. Glony solnisk podłęczyckich. The algae of salt marches near Łęczyca, Central Poland. *Mon. Botanic*.39.

Pniewski F. & Sylwestrzak Z. 2018. Influence of short periods of increased water temperature on species composition and photosynthetic activity in the Baltic periphyton communities. *Biologia* 73(11): 1067-1072 https://doi.org/10.2478/s11756-018-0122-6.

Poikane S., Kelly M. & Cantonati M. 2016. Benthic algal assessment of ecological status in European lakes and rivers: Challenges and opportunities. *Science of the Total Environment* 568: 603–613 DOI:10,1016/j.scitotenv.2016.02.027.

Potapova M. & Snoeijs P. 1997. The natural life cycle in wild populations of *Diatom moniliformis* (Bacillariophyceae) and its disruption in an aberrant environment. *Journal of Phycology* 33: 924-937 https://doi.org/10.1111/j.0022-3646.1997.00924.x.

Potapova M.G. & Ponader K.C. 2008. New species and combinations in the diatom genus *Sellaphora* (Sellaphoraceae) from Southeastern United States. *Harvard Papers in Botany* 13(1): 171–181 DOI: 10.3100/1043-4534(2008)13[171:NSACIT]2.0.CO;2.

Rakowska B. 1996. The benthic diatom community of a reservoir after the exploration of brow coal in Konin (central Poland). *Algological Studies* 82: 103–116 http://dx.doi.org/0342–1120/0116–103.

Reavie E.D. & Cai M. 2019. Consideration of species-specific diatom indicators of anthropogenic stress in the Great Lakes. *PLoS ONE 14(5):* e0210927 DOI: https://doi.org/10.1371/journal.pone.0210927

Reavie E.D. & Kireta A.R. 2015. Centric, Araphid and Eunotioid Diatoms of the Coastal Laurentian Great Lakes. In: Lange-Bertalot H. & Kociolek J.P., eds. *Bibliotheca Diatomologica Vol. 62.* Stuttgart: J. Cramer Gebr. Borntraeger Verlagsbuchhandlung. 184 pp.

Reavie E.D. & Smol J.P. 1998. Freshwater diatoms from the St. Lawrence river. In: Lange-Bertalot H. & Kociolek J.P., eds. *Bibliotheca Diatomologica Vol 41*. Berlin, Stuttgart: J. Cramer Gebr. Borntraeger Verlagsbuchhandlung, 184 pp.

Rumrich, U., Lange-Bertalot, H. & Rumrich, M. 200. Diatomeen der Anden. Von Venezuela bis Patagonien/Tierra del Fuego. Und zwei weitere Beiträge. In: Lange-Bertalot H., ed. *Iconographia Diatomologica Vol. 9*. Ruggell: A.R.G. Gantner Verlag K.G. 673 pp.

Sienkiewicz E & Gąsiorowski M. 2016. The evolution of a mining lake – From acidity to natural neutralization. *Science of the Total Environment* 557–558: 343–354 http://dx.doi.org/10.1016/j.scitotenv.2016.03.088.

Siver P.A., Hamilton P.B., Stachura-Suchoples K. & Kociołek J.P. 2005. The Freshwater Flora of Cape Code, Massachusetts, U.S.A. In: Lange-Bertalot H., eds. *Iconographia Diatomologica Vol. 14. Diatoms of North America*. Ruggell: A.R.G. Gantner Verlag K.G. 463 pp.

Soltanpour-Gargari A., Lodenius M. & Hinz F. 2011. Epilithic diatoms (Bacillariophycae) from streams in Ramsar, Iran. *Acta Botanica Croatica*. 70(2): 167-190 DOI: 10.2478/v10184-010-0006-5.

Stoermer E.F. & Håkansson H. 1984. *Stephanodiscus parvus*: validation of an enigmatic and widely misconstrued taxon. *Nova Hedwigia* 39: 497–511 http://www.jstor.org/stable/3226406.

Szczepocka E., Nowicka-Krawczyk P. & Kruk A. 2018. Deceptive ecological status of urban streams and rivers—evidence from diatom indices. *Ecosphere* 9(7):e02310 DOI: 10.1002/ecs2.2310

Szczepocka E., Szulc B., Szulc K., Rakowska B. & Żelazna-Wieczorek J. 2014. Diatom indices in the biological assessment of the water quality based on the example of a small lowland river. *Oceanological and Hydrobiological Studies* 43(3): 265–273 DOI: 10.2478/s13545-014-0141-z

Szczepocka E. & Żelazna-Wieczorek J. 2018. Diatom biomonitoring – scientific foundations, commonly discussed issues and frequently committed errors. *Oceanological and Hydrobiological Studies* 47(3): 313-325 DOI: 10.1515/ohs-2018-0030

Thomas E.J. & John J. 2006. Diatoms and macroinvertebrates as biomonitors of mine-like in Collie, Western Australia. *Journal of the Royal Society of Western Australia* 89: 109–117.

Tümpling W & Friedrich G. 1999. Methiden der Biologischen Wasseruntersuchung. Jena. Stutgart. Lübeck. Ulm. G. Fischer Verlag 133-153.

Weckström K. & Juggins S. 2005. Coastal diatom–environment relationships from the Gulf of Finland, Baltic Sea. *Journal of Phycology* 42: 21-35 DOI: 10.1111/j.1529-8817.2006.00166.x

Witkowski A. 1994. Recent and fossil diatom flora of the Gulf of Gdańsk, Southern Baltic Sea. In: *Bibliotheca Diatomologica Vol. 28*. Cramer, Berlin, Stuttgart. 313 pp

Witkowski A., Lange-Bertalot H. & Metzeltin D. 2000. Diatom flora of marine coasts I. In: Lange-Bertalot H., ed. *Iconographia Diatomologica. Vol. 7. Diversity-Taxonomy-Identification*. Ruggell, Liechtenstein: A.R.G. Gantner Verlag K.G. 419 pp.

Wojtal A.Z. 2009. The diatoms of Kobylanka stream near Kraków (Wyżyna Krakowsko–Częstochowska upland, S Poland). *Polish Botanical Journal* 54: 129–330.

Wojtal A.Z. 2013. Species composition and distribution of diatom assemblages in spring water from various geological formation in Southern Poland. In: Lange-Bertalot H. & Kociolek P., eds. *Bibliotheca Diatomologica Vol. 59*. Stuttgart: J. CRAMER in der Gebr. Borbtraeger Verlagsbuchhandlung. 111 p.

Wojtal A.Z. & Kwadrans J. 2006. Diatoms of the Wyżyna Krakowsko-Częstochowska Upland (S. Poland) – Coscinodiscophyceae (Thalassiositophycidae). *Polish Botanical Journal* 51: 177–207.

Van Dam H.; Mertens A. & Sinkeldam, J. 1994. A coded checklist and ecological indicator values of freshwater diatoms from the Netherlands. *Netherlands Journal of Aquatic Ecology* 28: 117–133.

Van de Vijver B., Frenot Y. & Beyens L. 2002. Freshwater diatoms from Ile de la Possession. In: Lange-Bertalot H. & Kociolek P., eds. *Bibliotheca Diatomologica Vol. 46*. Stuttgart: J. CRAMER in der Gebr. Borbtraeger Verlagsbuchhandlung 71 p.

Van de Vijver B., Wetzel C.E. & Ector L. 2018. Analysis of the type material of Planothidium delicatulum (Bacillariophyta) with the description of two new *Planothidium* species from the sub–Antarctic Region. *Fottea* 18(2): 200-211 DOI: 10.5507/fot.2018.006.

Van Landingham S.L. 1968. Investigation of a diatom population from mine Tailigns in Nye Country, Nevada, U.S.A. *Journal of Phycology* 4: 306–310.

Yang X-D., Dong X-H., Gao G., Pan H-X. & Wu J-L. 2005. Relationship Between Surface Sediment Diatoms and Summer Water Quality in Shallow Lakes of the Middle and Lower Reaches of the Yangtze River. *Journal of Integrative Plant Biology* 4792: 153-164 https://doi.org/10.1111/j.1744-7909.2005.00035.x.

Yamamoto M., Chiba T. & Tuji A. 2017. Salinity responses of benthic diatoms inhabiting tidal flats. *Diatom Research* 32: 243-250 DOI: 10.1080/0269249X.2017.1366951

Żelazna-Wieczorek J. 1996. Studium taksonomiczne rodzaju *Vaucheria* De Candolle oraz uwagi o występowaniu zidentyfikowanych gatunków tego rodzaju. Unpublished doctoral dissertation, University of Lodz, Poland.

Żelazna-Wieczorek J. 2002. *Vaucheria* species in selected regions in Poland. *Acta Societatis Botanicorum Poloniae* 71(2): 129-139.

Żelazna-Wieczorek, J. 2011. Diatom flora in springs of Łódź Hills (Central Poland). In: Witkowski A., ed. *Diatom monographs. Vol. 13.* Ruggell, Liechtenstein: A.R.G. Gantner Verlag K.G. 419 pp.

Żelazna-Wieczorek J. & Nowicka-Krawczyk P. 2015. The cascade construction of artificial ponds as a tool for urban stream restoration – The use of benthic diatoms to assess the effects of restoration practices. *Science of the Total Environment* 538: 591–599 DOI:10.1016/j.scitotenv.2015.08.110.

Żelazna-Wieczorek J. & Olszyński R.M. 2016. Taxonomic revision of *Chamaepinnularia krookiformis* Lange-Bertalot et Krammer with a description of *Chamaepinnularia plinskii* sp. nov. *Fottea, Olomouc* 16(1): 112-121 DOI: 10.5507/fot.2016.001.

Żelazna-Wieczorek J.; Olszyński R.M. & Nowicka-Krawczyk P. 2015. Half a century of research on diatoms in athalassic habitats in central Poland. *Oceanological and Hydrobiological Studies* 441: 51–67 DOI: 10.1515/ohs-2015-0006.

Żurek R., Diakiv V., Szarek-Gwiazda E., Kosiba J. & Wojtal A.Z. 2018. Unique Pit Lake Created in an Opencast Potassium Salt Mine (Dombrovska Pit Lake in Kalush, Ukraine). *Mine Water and the Environment* 37(3): 456-469 https://doi.org/10.1007/s10230-018-0527-z

Figure 1

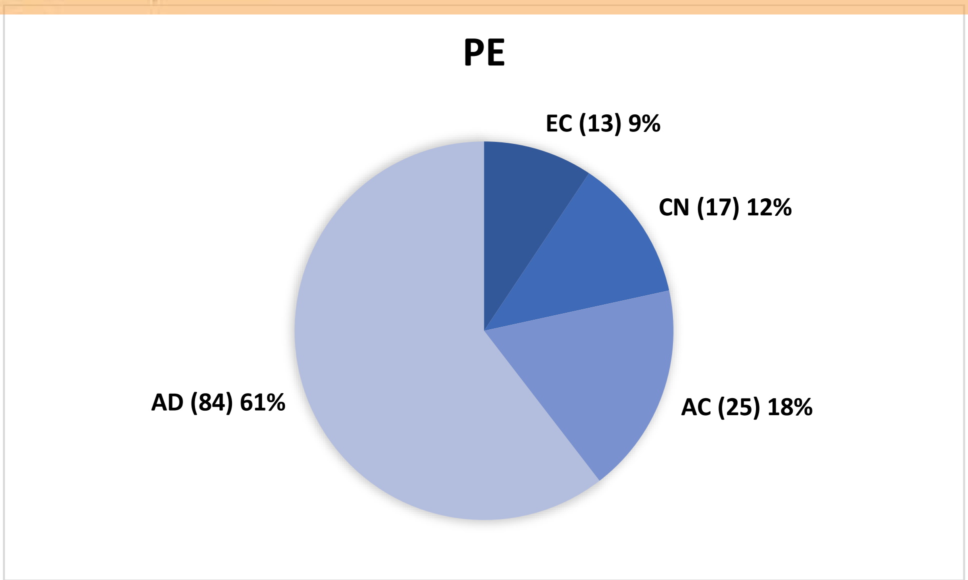
Location of sampling points in the Łódzkie and Wielkopolskie voivodships. Poland

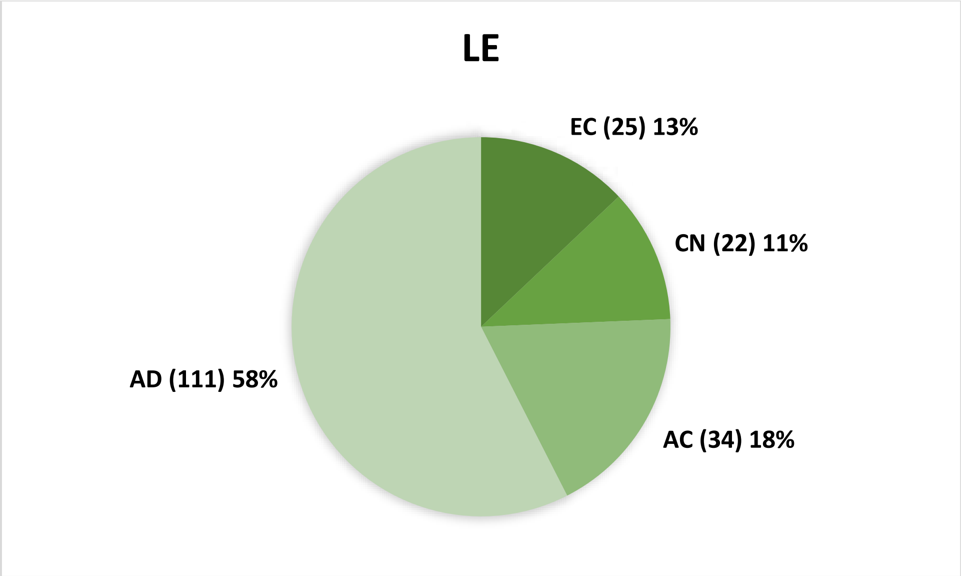
Figure 2(on next page)

Percentage share of diatoms in individual classes of prevalence according to the Tümpling & Friedrich factor (1999)

PE – Pełczyska complex, LE – Łęczyca reservoirs, BO – Bogdałów reservoir. EC – euconstant taxa, CN – constant taxa, AC – accessory taxa AD – accidental taxa. The number of species is shown in brackets

PeerJ Preprints | https://doi.org/10.7287/peerj.preprints.27739v1 | CC BY 4.0 Open Access | rec: 17 May 2019, publ: 17 May 2019


PE
EC (13) 9%
CN (17) 12%
AC (25) 18%
AD (84) 61%


LE
EC (25) 13%
CN (22) 11%
AC (34) 18%
AD (111) 58%

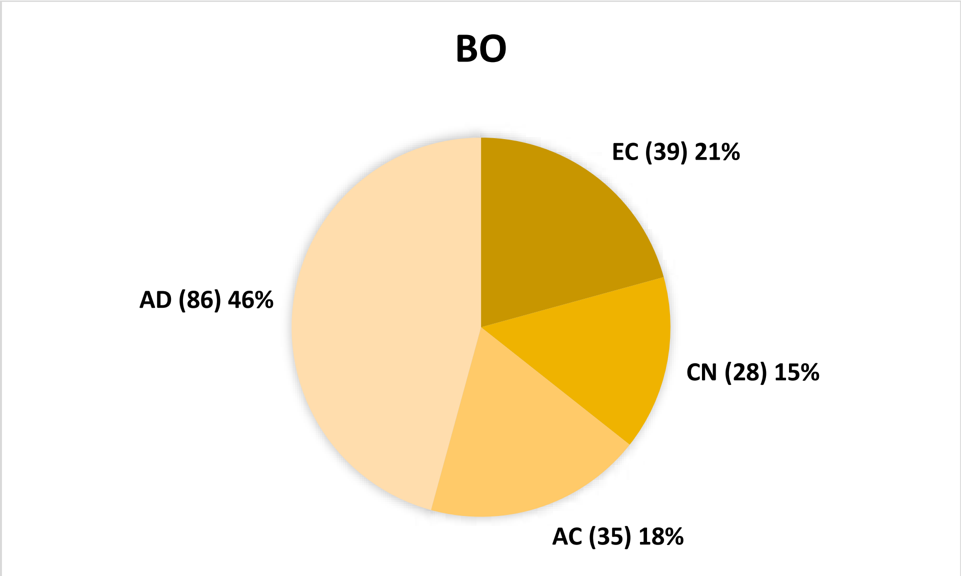

BO
EC (39) 21%
CN (28) 15%
AC (35) 18%
AD (86) 46%

Figure 3

MDS 3D analysis. The diagram shows three distinct clouds of samples which are grouping coincides with three hydrological objects

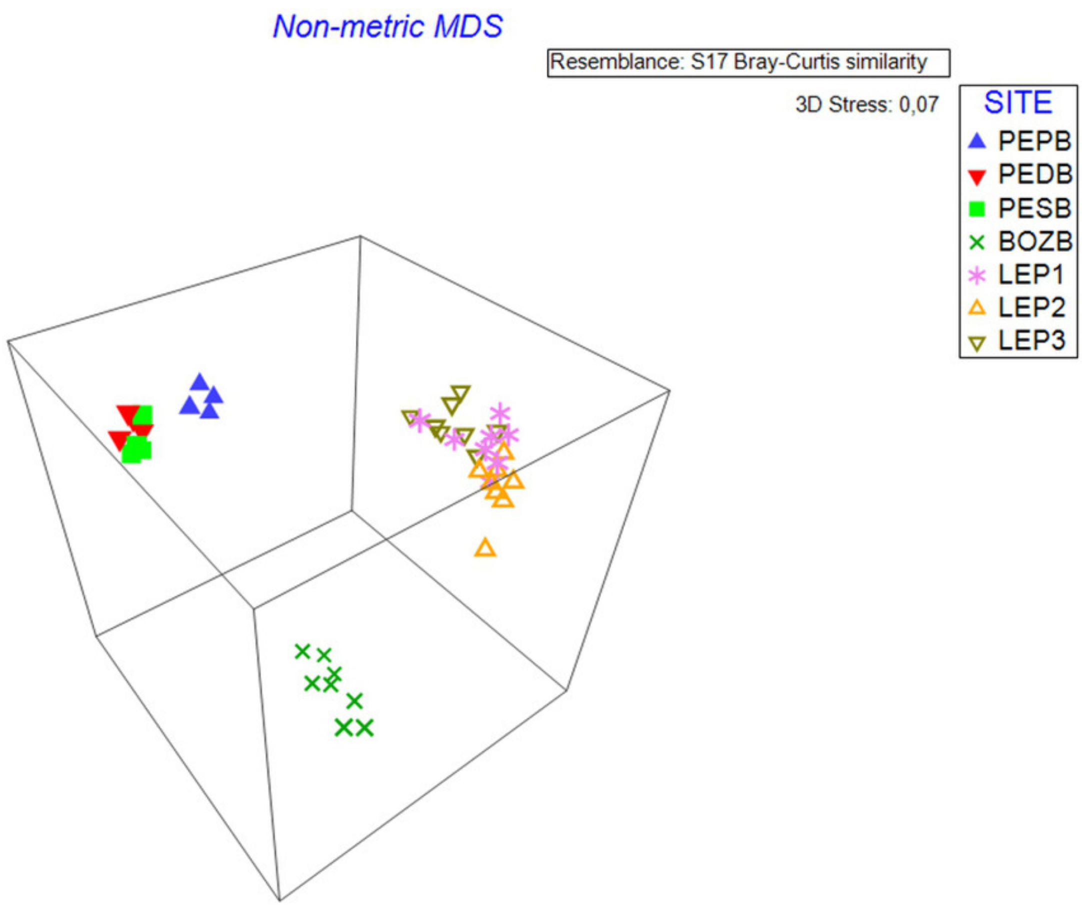
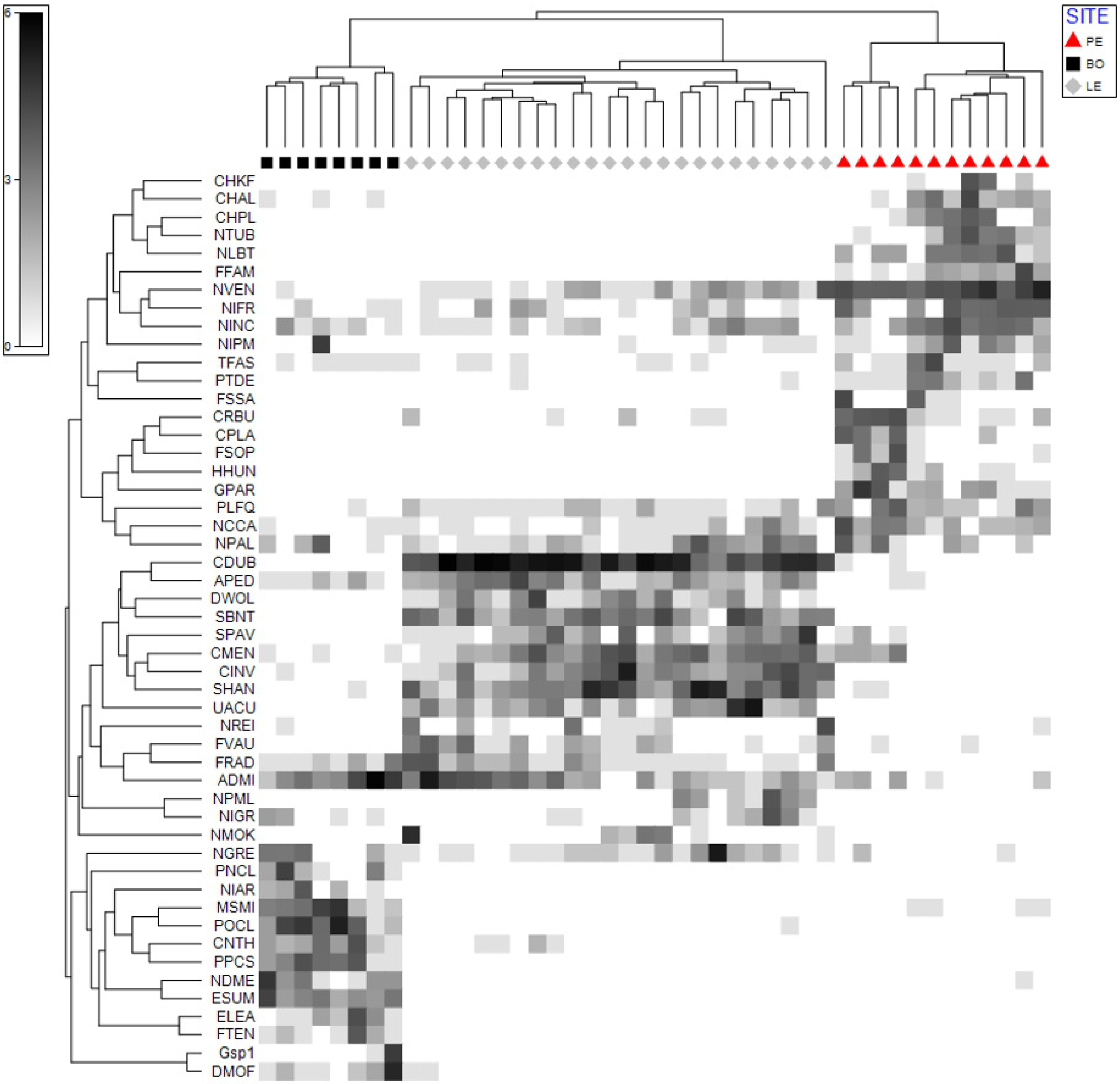


Figure 4

Shade Plot analysis. The diagram shows the strength of the factor (taxon) affecting the similarity between the samples

Upper dendrogram - samples divided according to hydrological object. Left dendrogram - 50 taxa of diatoms which have the strongest influence on the similarity between the samples



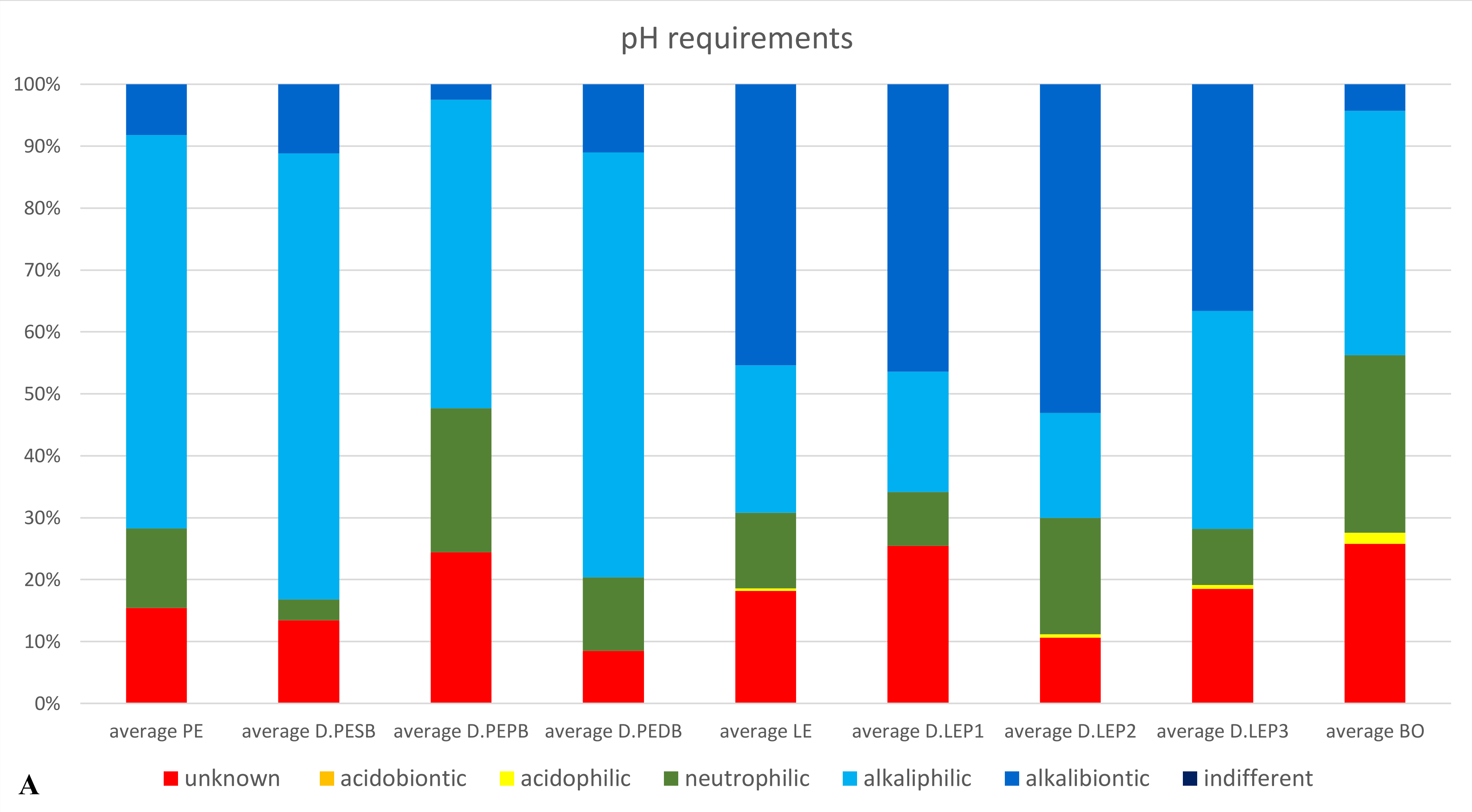
6
3
0
SITE
PE
BO
LE
CHKF
CHAL
CHPL
NTUB
NLBT
FFAM
NVEN
NIFR
NINC
NIPM
TFAS
PTDE
FSSA
CRBU
CPLA
FSOP
HHUN
GPAR
PLFQ
NCCA
NPAL
CDUB
APED
DWOL
SBNT
SPAV
CMEN
CINV
SHAN
UACU
NREI
FVAU
FRAD
ADMI
NPML
NIGR
NMOK
NGRE
PNCL
NIAR
MSMI
POCL
CNTH
PPCS
NDME
ESUM
ELEA
FTEN
Gsp1
DMOF

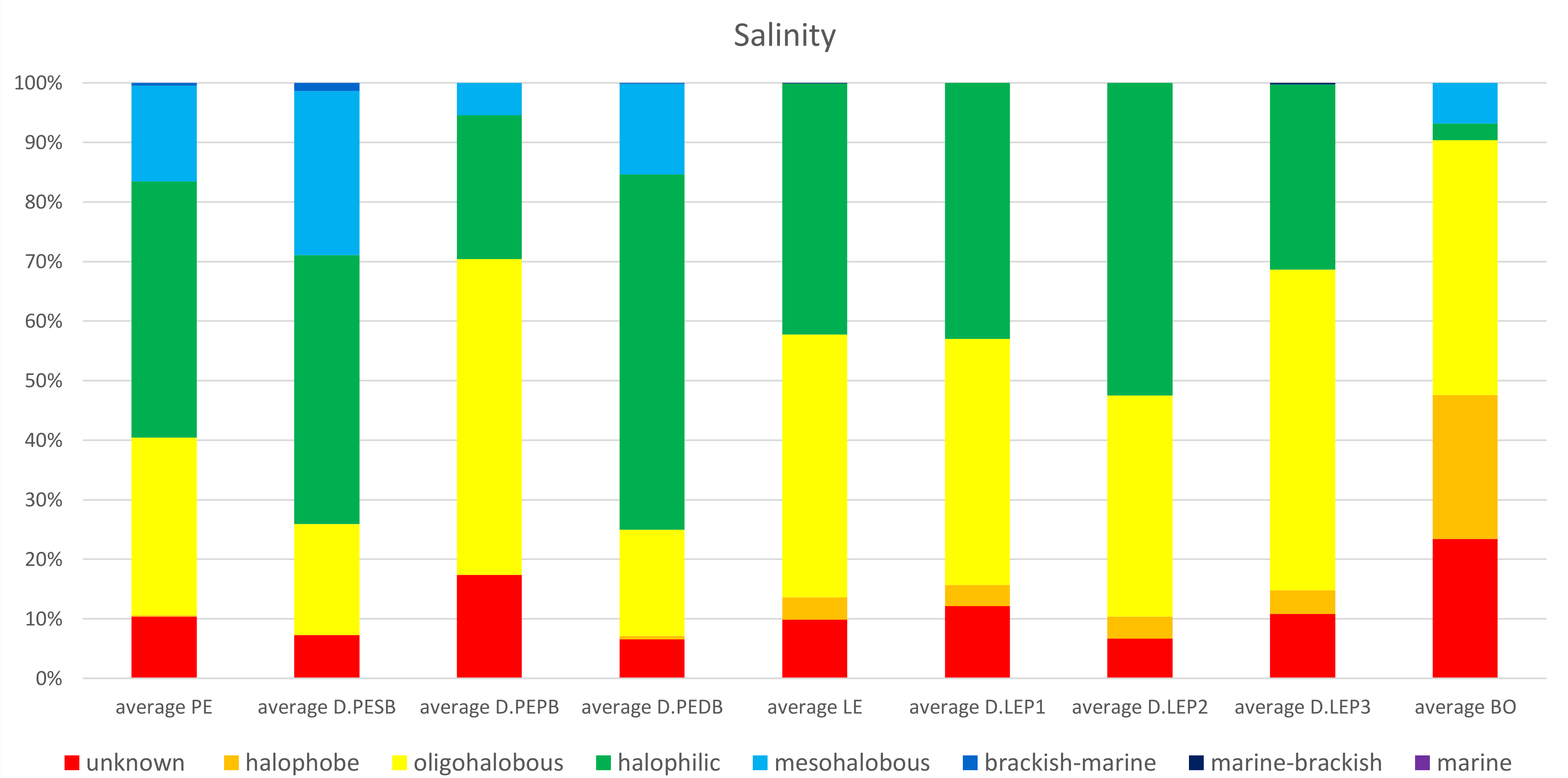
Figure 5(on next page)

Average percentages of diatom species in individual classes of ecological values according to Van Dam et al. 1994 based on OMNIDIA 6.0.6 software.

A. pH requirements. B. Salinity. C. Nitrogen uptake. D. Oxygen requirements. E. Saprobity. F. Trophic state. G. Moisture aerophily

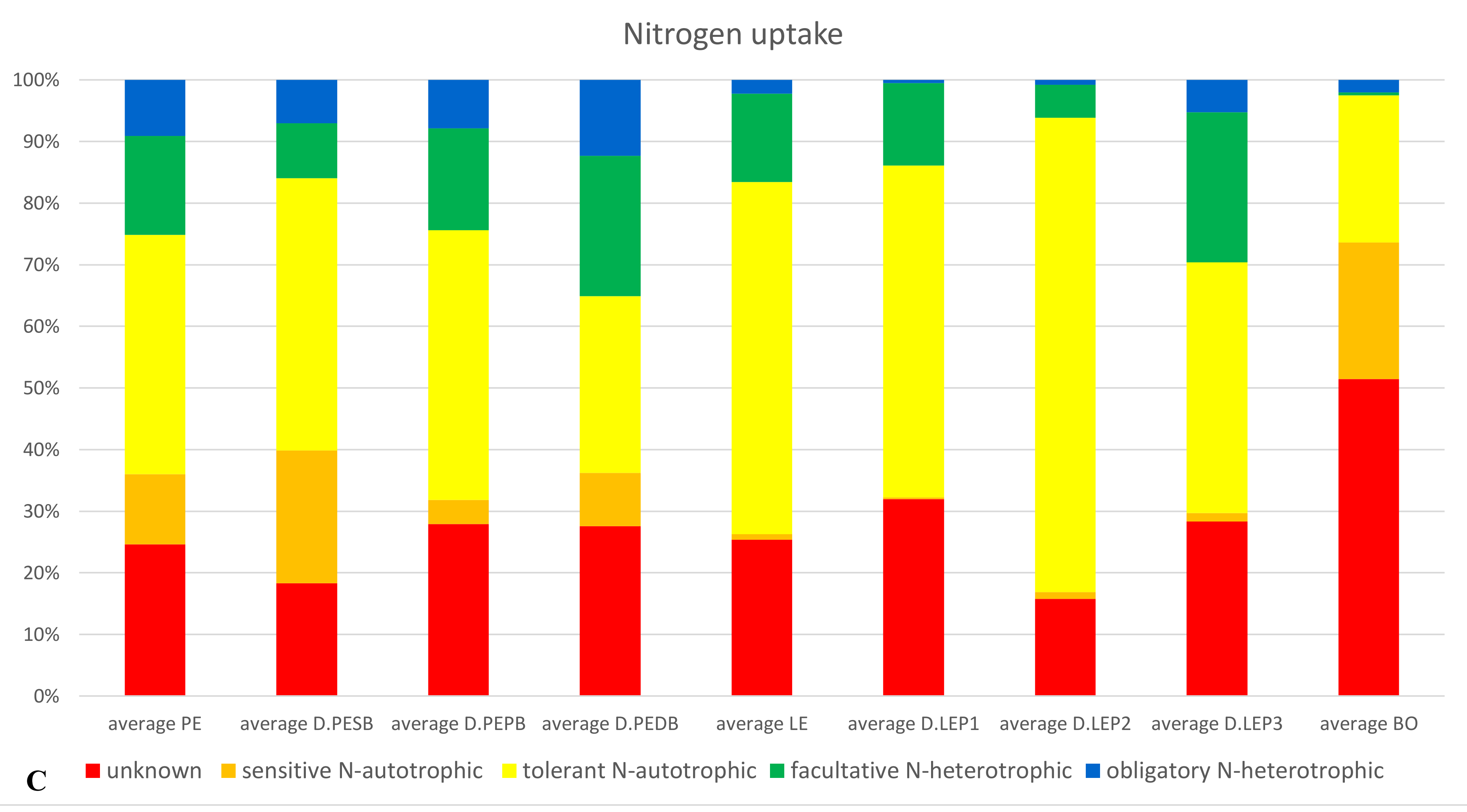
PeerJ Preprints | https://doi.org/10.7287/peerj.preprints.27739v1 | CC BY 4.0 Open Access | rec: 17 May 2019, publ: 17 May 2019


pH requirements
100%
90%
80%
70%
60%
50%
40%
30%
20%
10%
0%
average PE
average D.PESB
average D.PEPB
average D.PEDB
average LE
average D.LEP1
average D.LEP2
average D.LEP3
average BO
unknown
acidobiontic
acidophilic
neutrophilic
alkaliphilic
alkalibiontic
indifferent
A

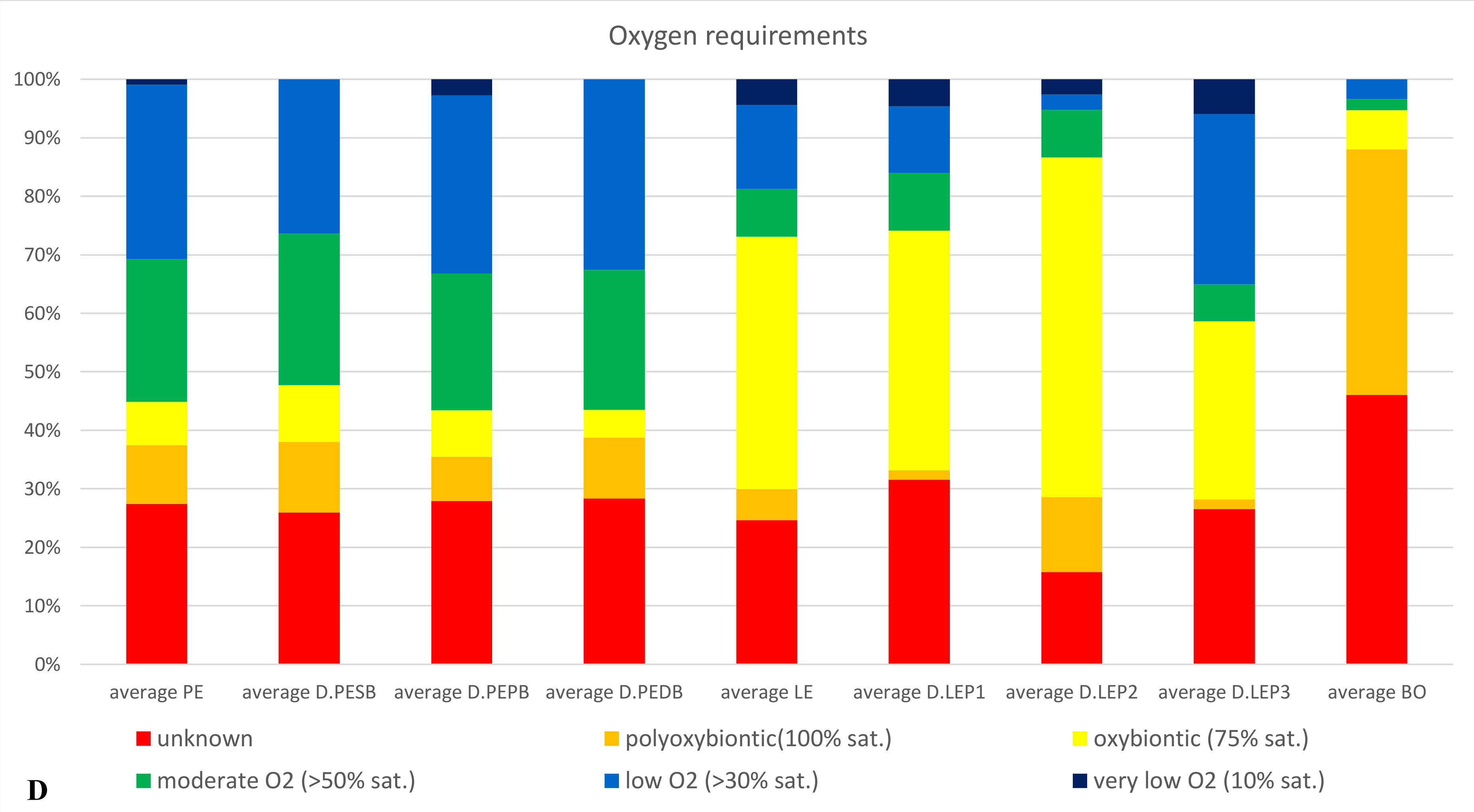


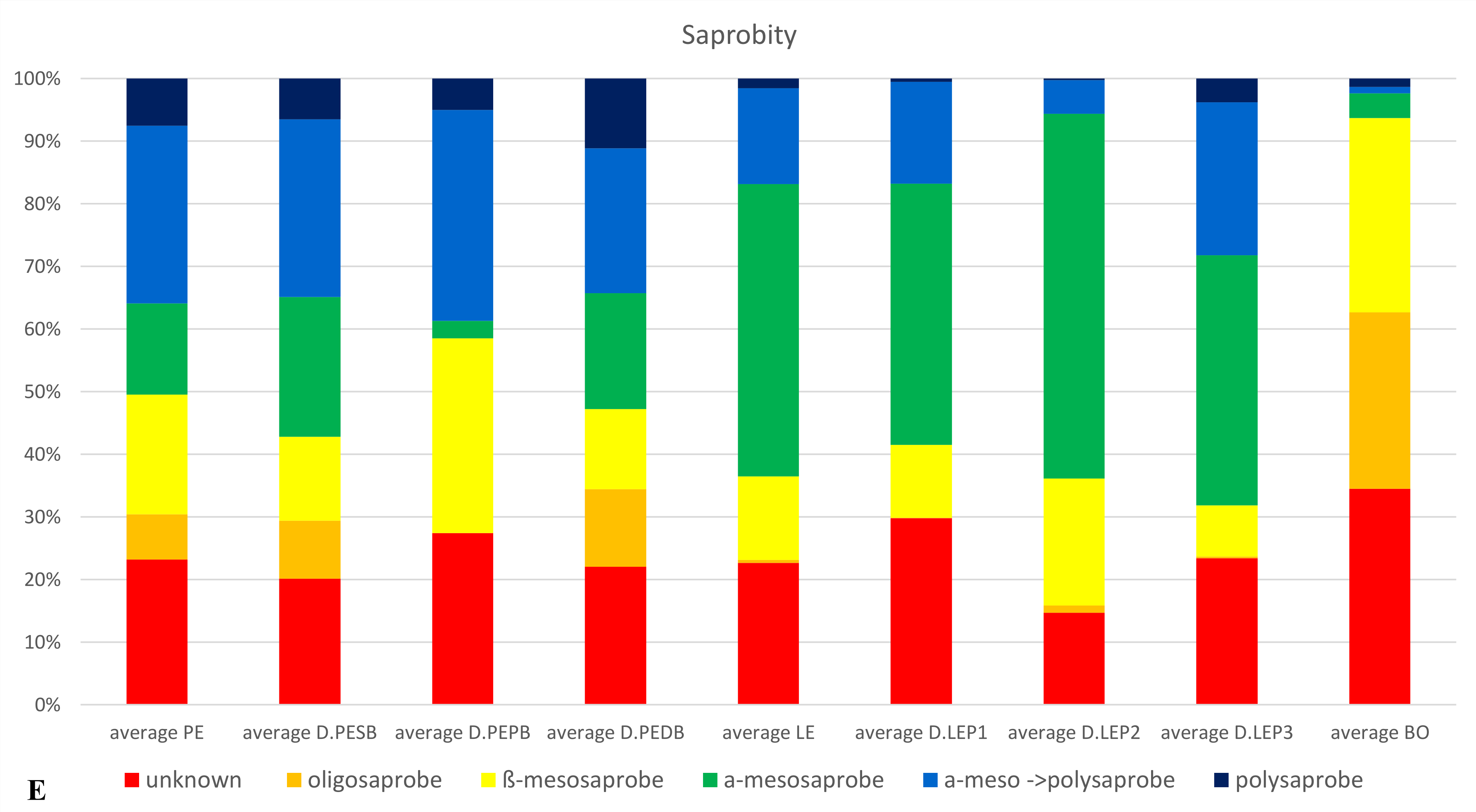
B

PeerJ Preprints | https://doi.org/10.7287/peerj.preprints.27739v1 | CC BY 4.0 Open Access | rec: 17 May 2019, publ: 17 May 2019


Nitrogen uptake
100%
90%
80%
70%
60%
50%
40%
30%
20%
10%
0%
average PE
average D.PESB
average D.PEPB
average D.PEDB
average LE
average D.LEP1
average D.LEP2
average D.LEP3
average BO
unknown
sensitive N-autotrophic
tolerant N-autotrophic
facultative N-heterotrophic
obligatory N-heterotrophic

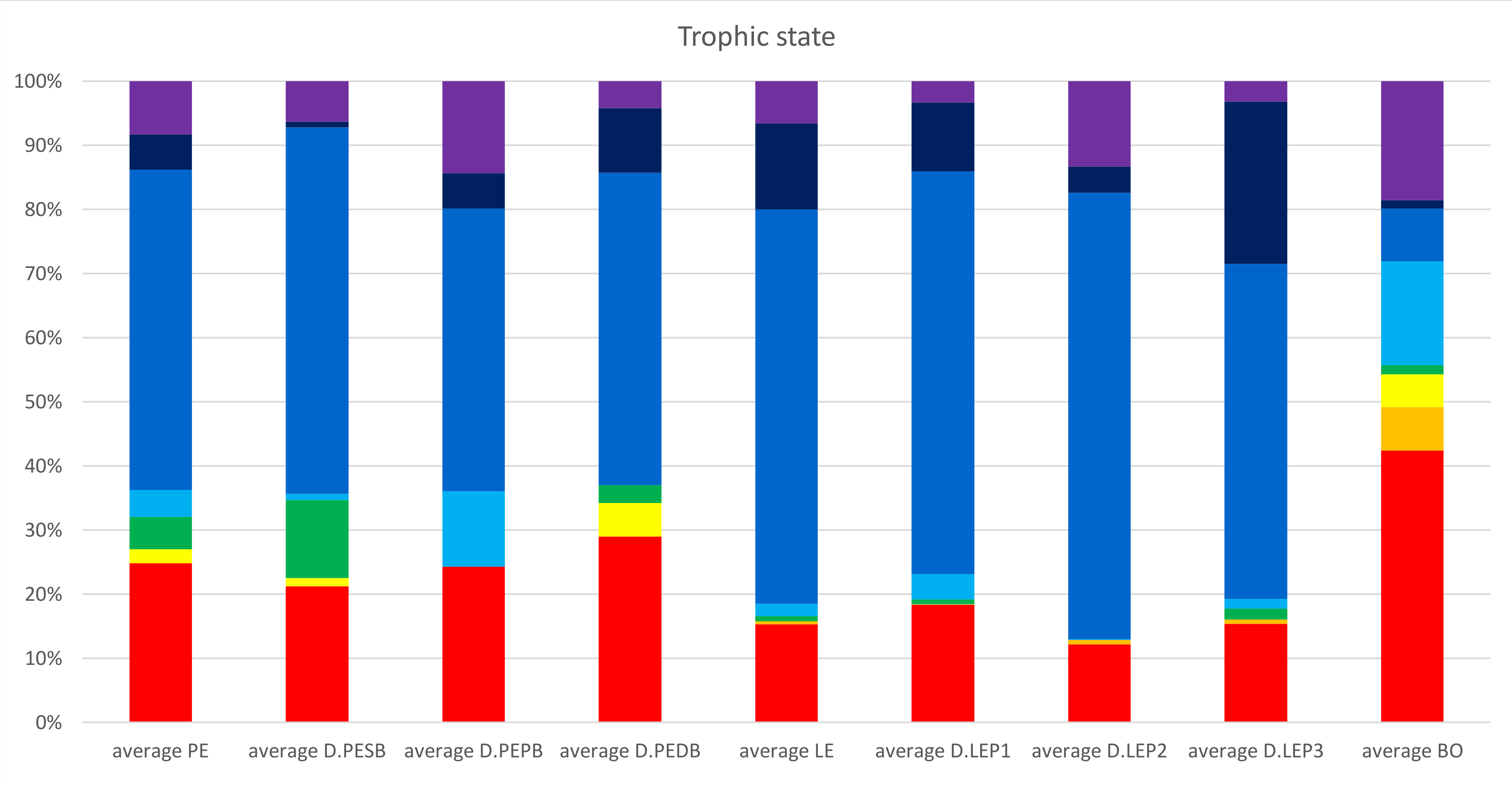
C


Oxygen requirements
100%
90%
80%
70%
60%
50%
40%
30%
20%
10%
0%
average PE
average D.PESB
average D.PEPB
average D.PEDB
average LE
average D.LEP1
average D.LEP2
average D.LEP3
average BO
unknown
polyoxybiontic(100% sat.)
oxybiontic (75% sat.)
moderate O2 (>50% sat.)
low O2 (>30% sat.)
very low O2 (10% sat.)
D



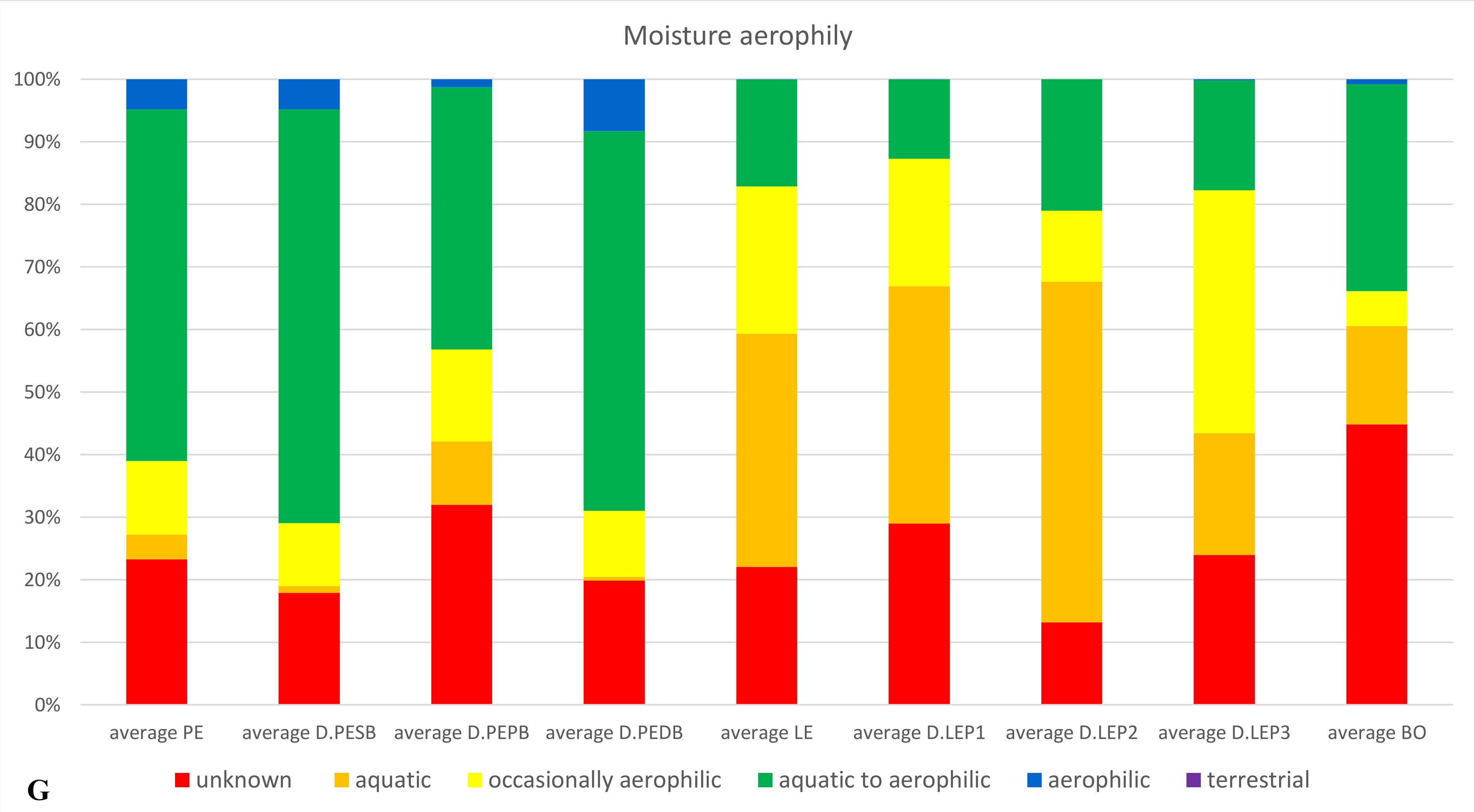
E

PeerJ Preprints | https://doi.org/10.7287/peerj.preprints.27739v1 | CC BY 4.0 Open Access | rec: 17 May 2019, publ: 17 May 2019



F

PeerJ Preprints | https://doi.org/10.7287/peerj.preprints.27739v1 | CC BY 4.0 Open Access | rec: 17 May 2019, publ: 17 May 2019


Moisture aerophily
100%
90%
80%
70%
60%
50%
40%
30%
20%
10%
0%
average PE
average D.PESB
average D.PEPB
average D.PEDB
average LE
average D.LEP1
average D.LEP2
average D.LEP3
average BO
unknown
aquatic
occasionally aerophilic
aquatic to aerophilic
aerophilic
terrestrial

G

Figure 6

PCA analysis. The diagram presents three groups of species A, B and C, whose occurrence can be correlated with selected physical and chemical factors

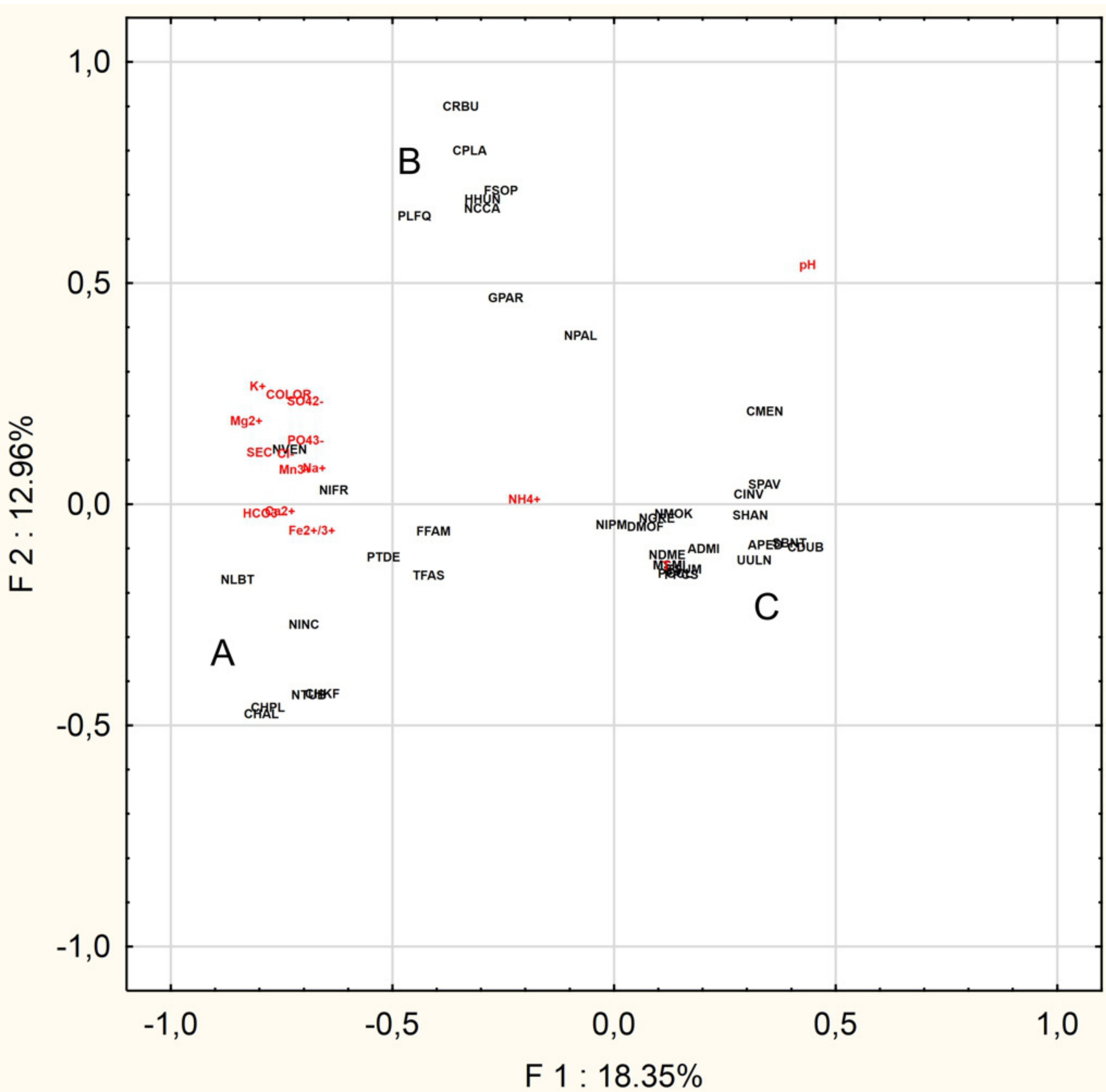


Figure 7

LM microphotographs of characteristic species for Pełczyska

A1-A6. *Planothidium delicatulum*. B1-B5. C*hamaepinnularia krookiformis*. C1-C5. *Chamaepinnularia plinskii*. D1-D4. *Nitzschia liebethruthii*. E1-E5. *Craticula buderi.* F1-F5. *Navicula cincta.* G1-G5. *Planothidium frequentissimum*. Scale bar = 10 µm

PeerJ Preprints | https://doi.org/10.7287/peerj.preprints.27739v1 | CC BY 4.0 Open Access | rec: 17 May 2019, publ: 17 May 2019

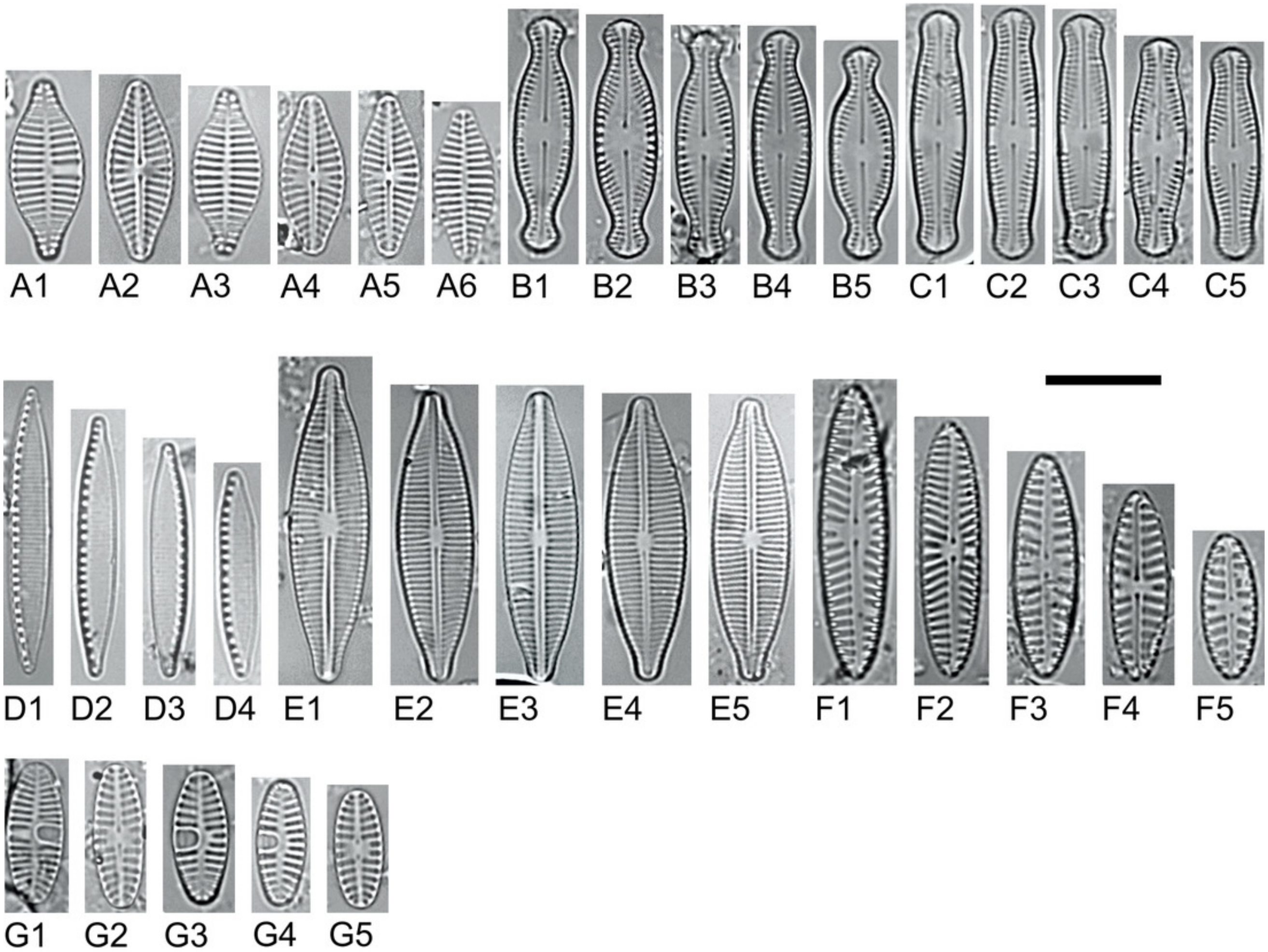

A1 A2 A3 A4 A5 A6 B1 B2 B3 B4 B5 C1 C2 C3 C4 C5
D1 D2 D3 D4 E1 E2 E3 E4 E5 F1 F2 F3 F4 F5
G1 G2 G3 G4 G5

Figure 8

LM microphotographs of characteristic species for Łęczyca

A1-A6. *Cyclostephanos invisitatus*. B1-B6. *Navicula moskalii*. C1-C5. *Stephanodiscus binatus*. D1-D5. *Stephanodiscus parvus*. Scale bar = 10 µm

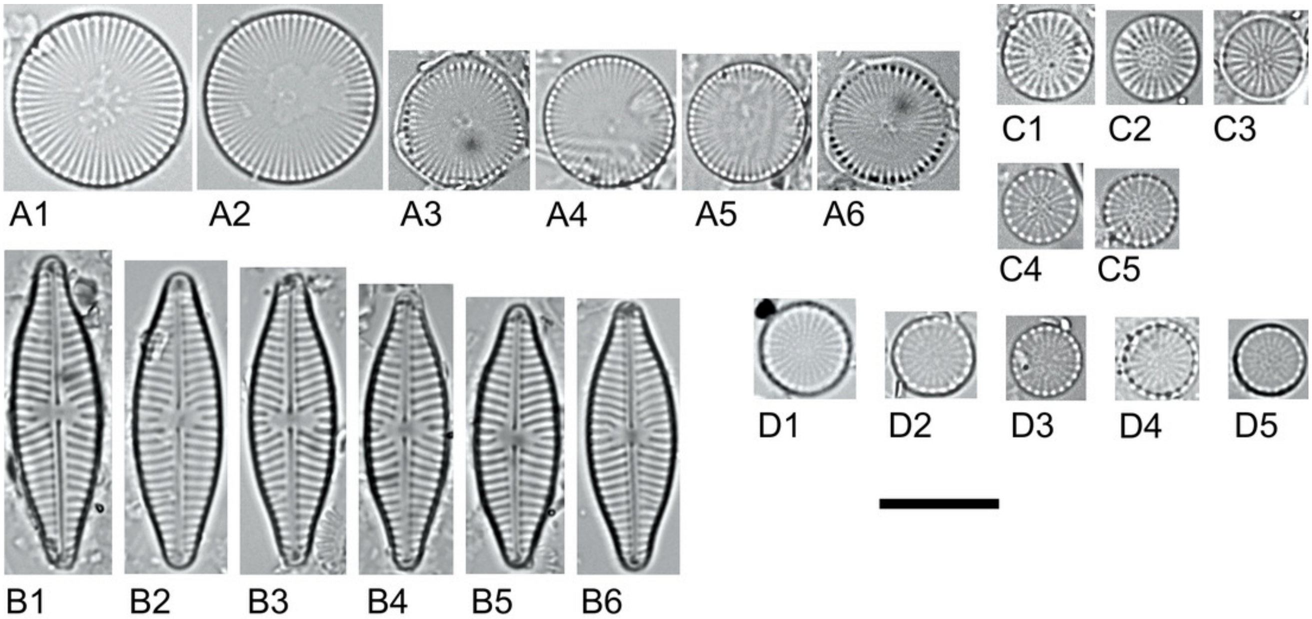

A1
A2
A3
A4
A5
A6
C1
C2
C3
C4
C5
B1
B2
B3
B4
B5
B6
D1
D2
D3
D4
D5

Figure 9

LM microphotographs of characteristic species for Bogdałów

A1-A6. *Diatoma moniliformis*. B1-B6. *Encyonopsis subminuta*. C1-C7. *Mastogloia smithii* (C5-C7. Same specimen, different focal plane). D1-D5. *Nitzschia dissipata* var. *media*. E1-E7. *Pantocsekiella pseudocomensis*

PeerJ Preprints | https://doi.org/10.7287/peerj.preprints.27739v1 | CC BY 4.0 Open Access | rec: 17 May 2019, publ: 17 May 2019

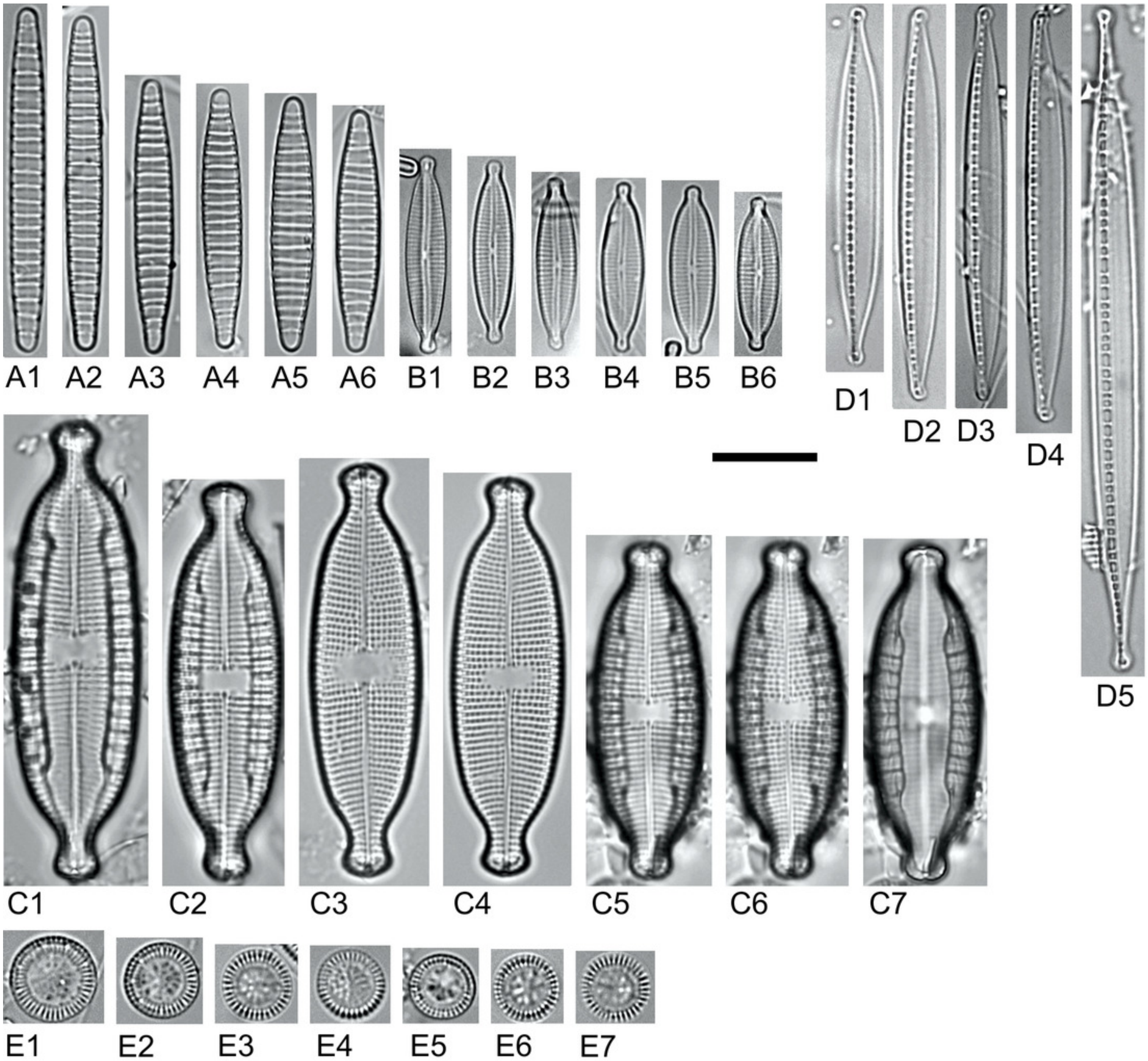

A1
A2
A3
A4
A5
A6
B1
B2
B3
B4
B5
B6
D1
D2
D3
D4
D5
C1
C2
C3
C4
C5
C6
C7
E1
E2
E3
E4
E5
E6
E7

Table 1(on next page)

Physical and chemical parameters in the examined habitats. The minimum, maximum and mean values

Table 1. Physical and chemical parameters in the examined habitats. The minimum, maximum and mean values.

	Pełczyska (PE)									Łeczyca (LE)									Bogdałów (BO)		
	outflow (D.PESB)			ditch (D.PEDB)			pond (D.PEPB)			L1 (D.LEP1)			L2 (D.LEP2)			L3 (D.LEP3)			reservoir (D.BOZB)		
	min	max	avar.	min	max	avar.	min	max	avar.	min	max	avar.	min	max	avar.	min	max	avar.	min	max	avar.
pH	6,6	7,7	7,0	6,4	6,4	6,4	7,8	9,6	8,7	7,9	8,6	8,1	7,5	8,6	8,1	7,5	8,8	8,3	7,9	8,3	8,1
SEC [µs]	4450	9230	6699	5170	5170	5170	2645	5150	3646	657	865	758	558	836	719	472	778	659	505	734	623
T [ºc]	6,8	13,9	9,1	7,8	7,8	7,8	1,1	17,3	7,1	1,6	20,6	10,3	0,6	21,1	10,3	2,2	15,2	7,8	4,7	22,7	11,4
HCO^{3-}	345	744	475	610	610	610	284	451	352	211	339	260	168	275	210	183	290	241	174	369	270
$CO_2^{[HCO3-]}$	124	268	171	220	220	220	102	163	126	76	122	94	61	99	75	66	105	87	63	133	96
Cl^-	1585	2976	2426	1006	1006	1006	685	1524	1053	57	93	78	60	93	76	67	106	78	41	52	45
N_{NH4}	0,02	1,63	0,45	0,15	0,15	0,15	0,00	0,36	0,11	0,02	0,71	0,22	0,03	0,53	0,26	0,00	1,16	0,32	0,00	0,07	0,02
NH^{4+}	0,03	2,09	0,71	0,19	0,19	0,19	0,00	0,46	0,14	0,03	0,55	0,22	0,03	0,54	0,28	0,00	0,90	0,31	0,00	0,09	0,03
PO_4^{3-}	0,60	12,46	3,73	8,57	8,57	8,57	0,85	9,10	3,99	0,22	0,53	0,36	0,05	0,57	0,41	0,09	0,57	0,34	0,27	0,45	0,34
P_{PO4}	0,20	4,11	1,23	2,83	2,83	2,83	0,28	3,00	1,32	0,07	0,17	0,12	0,02	0,19	0,13	0,03	0,19	0,11	0,09	0,15	0,12
SO_4^{2-}	176	198	188	165	165	165	151	197	176	71	147	107	67	122	101	63	91	75	110	147	123
S_{SO4}	58,8	66,2	62,9	55,1	55,1	55,1	50,3	65,7	58,7	23,5	49,0	35,8	22,3	40,8	33,8	21,0	30,3	25,1	37,9	49,2	41,5
COLOR	25	160	81	140	140	140	50	120	78	10	60	27	9	60	27	12	60	31	4	10	6
Mn^{3+}	0,17	0,51	0,36	0,14	0,14	0,14	0,03	0,25	0,13	0,00	0,05	0,02	0,01	0,03	0,02	0,01	0,07	0,03	0,00	0,01	0,00
$Fe^{2+/3+}$	0,25	0,41	0,32	0,25	0,25	0,25	0,05	0,24	0,11	0,03	0,27	0,09	0,00	0,13	0,04	0,01	0,07	0,03	0,01	0,09	0,02
Mg^{2+}	39,5	48,7	45,0	35,9	35,9	35,9	25,6	37,0	32,1	11,6	19,1	15,2	10,3	18,3	15,1	9,5	15,3	12,5	9,6	15,7	11,7
Ca^{2+}	171,7	216,2	195,2	165,0	165,0	165,0	75,8	139,8	121,1	58,0	143,6	89,0	59,5	117,6	79,1	52,6	86,4	65,5	77,7	117,6	86,3
Na^+	500,7	1537,4	1227,1	453,3	453,3	453,3	277,3	681,8	455,9	15,6	40,4	30,9	20,3	42,4	28,3	22,0	42,4	33,0	24,0	43,1	32,6
K^+	8,5	124,8	42,4	109,8	109,8	109,8	58,6	68,8	63,5	4,5	9,5	7,2	5,1	9,3	7,3	5,6	10,7	8,2	0,1	3,6	1,5

Table 2 (on next page)

Species of diatoms in the studied hydrological objects and index values assigned to them according to Van Dam et al. 1994, on the basis of OMNIDIA 6.0.6 data

Table 2. Species of diatoms in the studied hydrological objects and index values assigned to them according to Van Dam et al. 1994, on the basis of OMNIDIA 6.0.6 data.

Species	Code	Moisture aerophity	Nitrrogen uptake	pH requirements	Oxygen requirements	Salinity	Saprobity	Trophic state
Achnanthidium minutissimum	ADMI	3	2	3	1	2	2	7
Amphora pediculus	APED	3	2	4	2	2	2	5
Chamaepinnularia krookiformis	CHKF	3	0	3	0	3	1	0
Chamaepinnularia plinskii	CHPL	0	0	0	0	0	0	0
Cocconeis placentula	CPLA	2	2	4	3	2	2	5
Craticula buderi	CRBU	0	0	0	0	0	0	0
Craticula halophila	CHAL	2	2	4	2	4	3	5
Cyclostephanos dubius	CDUB	1	2	5	2	3	3	5
Cyclostephanos invisitatus	CINV	0	0	0	0	2	0	5
Cyclotella meneghiniana	CMEN	2	3	4	5	3	4	5
Diatoma moniliformis	DMOF	0	0	0	0	0	0	0
Encyonopsis subminuta	ESUM	0	0	3	1	1	1	1
Fragilaria famelica	FFAM	3	1	4	1	2	1	3
Fragilaria sopotensis	FSOP	1	2	4	1	2	2	4
Gomphonema parvulum	GPAR	3	3	3	4	2	4	5
Hippodonta hungarica	HHUN	3	2	4	3	2	2	4
Mastogloia smithii	MSMI	3	0	4	0	4	2	0
Navicula cincta	NCCA	0	0	0	0	2	0	7
Navicula gregaria	NGRE	3	2	4	4	3	3	5
Navicula moskalii	NMOK	0	0	0	0	0	0	0
Navicula veneta	NVEN	3	2	4	4	3	4	5
Nitzschia dissipata var. *media*	NDME	0	0	4	0	2	0	0
Nitzschia frustulum	NIFR	3	4	4	3	3	2	5
Nitzschia inconspicua	NINC	3	3	4	3	3	3	5
Nitzschia liebethruthii	NLBT	0	0	5	0	4	0	0
Nitzschia palea	NPAL	3	4	3	4	2	5	6
Nitzschia perminuta	NIPM	3	1	4	1	2	1	2
Nitzschia tubicola	NTUB	2	3	4	4	3	5	6
Pantocsekiella ocellata	POCL	1	1	4	1	1	1	4
Pantocsekiella pseudocomensis	PPCS	0	0	0	0	0	0	0
Planothidium delicatulum	PTDE	3	1	5	0	4	5	3
Planothidium frequentissimum	PLFQ	0	2	4	3	2	4	7
Stephanodiscus binatus	SBNT	0	0	0	0	0	0	0
Stephanodiscus hantzschii	SHAN	2	3	5	4	2	4	6
Stephanodiscus parvus	SPAV	0	0	5	0	2	0	6
Tabularia fasciculata	TFAS	3	2	4	3	4	3	5

Table 3(on next page)

Percentage of characteristic species in the unknown (total) class according to Van Dam et al. (1994) at each sample point

Percentage of individual characteristic species constituting at least 10% of the 'unknown' class at each sampling point

Table 3. Percentage of characteristic species in the unknown (total) class according to Van Dam et al. (1994) at each sample point. Percentage of individual characteristic species constituting at least 10% of the 'unknown' class at each sampling point.

	pH	Salinity	Nitrogen uptake	Oxygen requirements	Saprobity	Trophic state	Moisture aerophily
D.PESB.	total 33%	total 44%	total 34%	total 48%	total 30%	total 22%	total 43%
	CHPL (19%) NCCA (11%)	CHPL (33%) CHKF (10%)	CHPL (11%) NCCA (10%)	PTDE (25%)	CHPL (11%)	CHPL (10%)	PLFQ (20%)
D.PEDB.	total 92%	total 25%	total 82%	total 82%	total 80%	total 74%	total 85%
	CHPL (60%) NCCA (32%)	CHPL (25%)	NLBT (37%) CHPL (25%) CHKF (16%)	NLBT (37%) CHPL (16%) CHKF (16%)	NLBT (43%) CHPL (32%)	NLBT (365) CHPL (23%) CHKF (15%)	NLBT (49%) CHPL (285)
D.PEPB.	total 81%	total 73%	total 78%	total 78%	total 79%	total 60%	total 84%
	CRBU (53%) NCCA (27%)	CRBU (73%)	CRBU (46%) NCCA (25%)	CRBU (46%) NCCA (25%)	CRBU (46%) NCCA (26%)	CRBU (54%)	CRBU (41%) NCCA (23%) PLFQ (14%)
D.LEP1.	total 76%	total 79%	total 67%	total 68%	total 71%	total 54%	total 78%
	CINV (36%) SBNT (23%) NMOK (16%)	SBNT (56%) NMOK (21%)	CINV (29%) SBNT (19%) NMOK (14%)	CINV (29%) SBNT (19%) NMOK (14%)	CINV (31%) SBNT (20%) NMOK (14%)	SBNT (36%) NMOK (17%)	CINV (32%) SBNT (22%) NMOK (14%)
D.LEP2.	total 39%	total 44%	total 35%	total 35%	total 28%	total 23%	total 44%
	SBNT (27%) CINV (11%)	SBNT (44%)	SBNT (18%)	SBNT (18%)	SBNT (20%)	SBNT (23%)	SBNT (22%) SPAV (11%) CINV (11%)
D.LEP3.	total 57%	total 41%	total 50%	total 53%	total 60%	total 29%	total 57%
	SBNT (25%) CINV (24%)	SBNT (41%)	SBNT (17%) CINV (15%) SPAV (13%)	SBNT (17%) CINV (16%) SPAV (14%)	SBNT (20%) CINV (18%) SPAV (15%)	SBNT (29%)	SBNT (20%) CINV (17%) SPAV (15%)
D.BOZB.	total 33%	total 36%	total 47%	total 44%	total 38%	total 47%	total 38%
	PPCS (22%) DMOF (10%)	PPCS (25%) DMOF (11%)	MSMI (13%) PPCS (10%)	MSMI (14%) PPCS (11%) NDME (10%)	PPCS (18%) NDME (11%)	MSMI (15%) PPCS (12%) NDME (11%)	PPCS (13%)

3

PeerJ Preprints | https://doi.org/10.7287/peerj.preprints.27739v1 | CC BY 4.0 Open Access | rec: 17 May 2019, publ: 17 May 2019