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Transplantation of adipose derived stem cells overexpressing inducible nitric oxide synthase ameliorates diabetes mellitus induced erectile dysfunction in rats

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Background: Erectile dysfunction is a major complication of diabetes mellitus. Adipose derived stem cells (ADSCs) has attracted much attention as a promising tool for the treatment of diabetes mellitus induced erectile dysfunction (DMED). Inducible nitric oxide synthase (iNOS) plays an important role in protecting penile tissues from fibrosis. The aim of this study was to determine the efficacy of ADSCs overexpressing iNOS on DMED in rats.

Methods: ADSCs were isolated and infected with adenovirus overexpressing iNOS (named as ADSCs-iNOS). The expression of iNOS was detected using western blot analysis and real-time PCR. Rats were randomly assigned into five groups: control group, DMED group, ADSCs group, ADSCs-EGFP group and ADSCs-iNOS group. 5×10^5 cells were given once via the intracorporal route. Two weeks after treatment, erectile function was assessed by electrical stimulation of the cavernous nerve. Penile tissues were obtained and evaluated at histology level.

Results: We found that ADSCs-iNOS had significantly higher expression of iNOS at mRNA and protein levels and generated more nitric oxide. ADSCs-iNOS reduced collagen I and collagen IV expression of corpus cavernosum smooth muscle cells (CCSMCs) in cell co-culture model. Transforming growth factor-β1 expression and p-Smad2/3 to Smad2/3 ratio in CCSMCs reduced following co-culture with ADSCs-iNOS. Injection of ADSCs-iNOS significantly ameliorated DMED in rats and decreased collagen/smooth muscle cell ratio of penile tissues. Moreover, elevated nitric oxide and cGMP concentrations were detected in penile tissues of ADSCs-iNOS group.

Conclusion: Taken together, ADSCs-iNOS significantly improve erectile function of DMED rats. The therapeutic effect may be achieved by increased nitric oxide generation and the suppression of collagen I and collagen IV expression in the CCSMCs to decrease penile fibrosis.

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Abstract

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Results: We found that ADSCs-iNOS had significantly higher expression of iNOS at mRNA and protein levels and generated more nitric oxide. ADSCs-iNOS reduced collagen I and collagen IV expression of corpus cavernosum smooth muscle cells (CCSMCs) in cell co-culture model. Transforming growth factor-β1 expression and p-Smad2/3 to Smad2/3 ratio in CCSMCs reduced following co-culture with ADSCs-iNOS. Injection of ADSCs-iNOS significantly ameliorated DMED in rats and decreased collagen/smooth muscle cell ratio of penile tissues. Moreover, elevated nitric oxide and cGMP concentrations were detected in penile tissues of ADSCs-iNOS group.

Conclusion: Taken together, ADSCs-iNOS significantly improve erectile function of DMED rats. The therapeutic effect may be achieved by increased nitric oxide generation and the suppression of collagen I and collagen IV expression in the CCSMCs to decrease penile fibrosis.

Introduction

Erectile dysfunction (ED) is one of the most significant complications in men with diabetes mellitus (DM). Numerous studies have found that more than 50% of men with DM are afflicted with ED (*Thorve et al., 2011*). Men with DM tend to suffer from ED about 10 years earlier than the general population (*Johannes et al., 2000*). Diabetes mellitus induced erectile dysfunction (DMED) is severe and has a negative effect on the quality of life. Phosphodiesterase 5 inhibitors are the predominant medicines to treat ED. However, diabetic male patients are less responsive to phosphodiesterase 5 inhibitors (*Cheng, 2007*). Therefore, developing new therapeutic options for DMED is urgent.

Over the past years, stem cell therapy has produced positive effects on DMED in experimental animals. Adipose derived stem cells (ADSCs) are attracting considerable attention becuase they can be isolated through standardized procedures and vastly expanded, and they secret a broad range of trophic factors (*Sowa et al., 2012*). Moreover, ADSCs transplantation seems to be safe and avoid the risks of tumorigenesis (*Alagesan & Griffin, 2014*). Autologous ADSCs significantly improved

erectile function of type-II diabetic rats (*Garcia et al., 2010*). Stem cell therapy has beneficial effects to increase contents of smooth muscle and endothelium, NO-cGMP pathway, neuronal nitric oxide synthase (nNOS)-positive nerve fibers, and decrease fibrosis and apoptosis in penis (*Chen et al., 2017; Qiu et al., 2011; Ryu et al., 2012; Sun et al., 2012; Zhang et al., 2016*).

Nitric oxide (NO) is a neurotransmitter which plays an important role in penile erection. NO is synthesized by three nitric oxide synthase (NOS) isoforms, including nNOS, endothelial NOS (eNOS) and inducible NOS (iNOS). Among these synthases, iNOS can produce NO continuously independent of Ca^{2+} (*Oliveira-Paula, Lacchini & Tanus-Santos, 2014*). Recent evidence suggests that the pathogenesis of DMED may be due to impaired NO production, and iNOS acts an essential part in protecting penile tissues from the pro-fibrotic effects of hyperglycemia (*Ferrini et al., 2010*). Continuous NO production by iNOS inhibits collagen synthesis, directly reacts with ROS to produce peroxynitrite, and downregulates transforming growth factor-β1 (TGF-β1)/Smad pathway, thus reducing fibrosis level (*Gonzalez-Cadavid & Rajfer, 2010*). Gene transfer of iNOS DNA decreased TGF-β1 and plasminogen-activator inhibitor-1 expression, and regressed Peyronie's disease-like plaque (*Davila et al., 2004*). Our previous study showed that saRNA mediated iNOS expression improved DMED via endogenously generating NO (*Wang et al., 2013*).

Given the beneficial effects of ADSCs and iNOS in the treatment of ED, we hypothesized that ADSCs combined with iNOS could improve erectile function of DMED rats. In this study we aimed to determine the efficacy of ADSCs combined with iNOS for improving erectile function in DMED rats.

Materials and methods

Rat ADSCs isolation and culture

Rat ADSCs were isolated as previously described (*Wang et al., 2015*). Briefly, adipose tissues were incised from inguen, and digested at 37°C in 0.1% collagenase type I (Sigma-Aldrich, St. Louis, USA) with vigorous shaking for 90 minutes. The digested tissues were mixed with Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal bovine serum (FBS), and centrifuged at 220 g for 10 minutes. The cells were suspended in DMEM containing 10% FBS and filtered through a 75-μm cell striner (Solarbio, Beijing, China). Then cells were plated and cultured in DMEM containing 10% FBS at 37°C in humidified incubator with 5% CO_2. Cells were passaged with trypsin/EDTA (Beyotime, Nantong, China) as required.

Flow cytometry

ADSCs were characterized by flow cytometry at passage 4. Briefly, ADSCs were harvested and washed twice with phosphate-buffered saline (PBS). Then, the cells were incubated for 30 minutes in PBS containing anti-CD29-FITC, CD34-FITC, CD45-FITC, CD49-FITC, CD73-FITC, CD105-FITC, CD31-PE, CD90-PE, CD106-PE (all from BD Biosciences, San Diego, USA). The stained cells were then subjected to flow cytometry analysis.

Differentiation of ADSCs

For adipocyte differentiation, ADSCs were cultured at a density of 1×10^4 cells/cm^2 in DMEM containing 10% FBS, 50 mg/L indomethacin, 0.5 mmol/L isobutylmethylxanthine, 10 mg/L insulin and 1 mmol/L dexamethasone (all from Sigma-Aldrich). After 21 days of induction, the cells were stained using Oil red-O staining solution.

For smooth muscle cell differentiation, ADSCs were cultured at a density of 1×10^4 cells/cm^2 in low-glucose DMEM containing 10% FBS, 50 μg/mL platelet-derived growth factor-BB and 5 μg/L TGF-β1 (all from PeproTech, Rocky Hill, USA). After 14 days of induction, the cells were stained with the antibody against α-smooth muscle actin (α-SMA, 1:200, Boster, Wuhan, China).

Transfection with adenovirus

The forth-passage ADSCs were cultured for 24 h. ADSCs were incubated with adenovirus expressing EGFP or iNOS-EGFP (GenecChem, Shanghai, China) at a multiplicity of infection (MOI) of 30. After incubation for 12 h, the medium was changed into fresh growth medium.

Real-time PCR

Real-time PCR was performed in cultured cells on the 1st, 3rd, 5th, 7th, 10th and 14th day after infection with adenovirus. RNA was extracted with a multisource RNA miniprep kit (Corning, NY, USA). Total RNA (500 ng) was reversely transcribed into cDNA using the PrimeScript™ RT reagent kit (TaKaRa, Dalian, China). Real-time PCR for each sample was performed on an MX3000P quantitative PCR system (Agilent, Santa Clara, USA) using SYBR Premix Ex Taq (TaKaRa) and the following primers: iNOS_ 5'-AAGCACATTTGGCAATGGAGAC-3', 5'-TGGAGCCCAGGCCAAATAC-3', β-actin_ 5'-GACGGTGTGCACCAACATCTA-3', 5'-TTCTTGGCTTTCAGGATGGAG-3'. PCR condition included 95°C for 30 seconds, followed by 40 cycles of 95°C for 5 seconds, 60°C for 30 seconds and 72°C for 30 seconds. Relative expression level of target genes was calculated with $2^{-\Delta\Delta Ct}$ method. β-actin was chosen as normalization control.

Western blot analysis

Protein extraction from rat penile tissues and cultured cells was performed using NP-40 lysis buffer (Beyotime). Protein concentration was detected with the BCA protein assay kit (Beyotime). 50 μg protein was electrophoresed on 12% sodium dodecyl sulfate/polyacrylamide gels and then blotted on a polyvinylidene difluoride membrane (Millipore, Billerica, USA). The membrane was blotted with primary antibodies against collagen I (1:500, Proteintech, Wuhan, China), collagen IV (1:500, Proteintech), iNOS (1:500, Santa Cruz, CA, USA), TGF-β1(1:1,000, Abcam, Cambridge, UK), Smad2/3 (1:1,000, Abcam) and phospho-Smad2/3 (p-Smad2/3, 1:1,000, Abcam) followed by secondary antibodies. Each band was quantified by densitometry with Image J software.

Cell proliferation assay

Proliferation assay was performed with cell counting kit-8 (CCK-8, Dojindo, Shanghai, China). Briefly, ADSCs, ADSCs-EGFP and ADSCs-iNOS were cultured in 96-well plates at

141 3×10^3 cells/well. After 1, 2, 3, 5, 7 and 14 days, 10 μl CCK-8 solution was added into each well,
142 and incubated at 37°C for 2 hours. The absorbance at 450 nm was detected with a multiscan MK3
143 microplate reader (Thermo Fisher Scientific, Waltham, USA).
144
145 **Measurement of NO concentration**
146 Cells were incubated in culture medium containing 10 mmol/L L-Arginine for 24 h on the 1st,
147 3rd, 5th, 7th, 10th and 14th day after infection with adenovirus. Then the supernatant was
148 harvested. NO concentrations in penile tissues or cultured cells were determined with total NO
149 assay kit (Beyotime) according to the manufacturer's instructions.
150
151 **Cyclic guanosine monophosphate (cGMP) concentration determination**
152 The cGMP concentrations in penile tissues or cultured cells were measured using an enzyme-
153 linked immunosorbent assay (ELISA) kit (Westang, Shanghai, China) according to the
154 manufacturer's instructions.
155
156 **Rat corpus cavernosum smooth muscle cell (CCSMC) isolation and culture**
157 Rat CCSMCs were obtained from corpus cavernosum tissues as described previously with
158 minor modifications(*Chen et al., 2011*). Briefly, corpus cavernosum tissues were harvested and
159 cut into fragments of 1 mm^3. These fragments were placed in an overturned 25 cm^2 culture flask.
160 Then, 4 ml DMEM containing 10% FBS was added into culture flask. After 30 minutes, the flask
161 was flipped over. Differential adherence method was used to purify CCSMCs. Immunostaining
162 with an antibody against α-SMA (1:200, Boster) was performed to identify CCSMCs.
163
164 **Transwell co-culture of ADSCs and CCSMCs**
165 ADSCs and CCSMCs were co-cultured in medium containing 30 mmol/L glucose in 0.4 μm
166 pore size Transwell inserts (Corning, NY, USA). CCSMCs were cultured in 12-well plates at
167 1×10^4 cells/well. ADSCs, ADSCs-EGFP or ADSCs-iNOS were seeded on Transwell inner
168 membrane at a density of 1×10^4 cells/well. The co-culture system allowed ADSCs and CCSMCs
169 to grow in the same medium without direct contact between them.
170
171 **Animal experiments**
172 Animal experiments were carried out according to the guidelines and regulations by the Ethical
173 Committee of Tongji Hospital, Tongji Medical College, Huazhong University of Science and
174 Technology. This study was approved by the Ethical Committee (TJ-A20131213). Sixty male SD
175 rats (8-week old) were purchased from Hunan SJA Laboratory Animal Co., Ltd. After an overnight
176 fast, fifty-one of them were randomly selected and received intraperitoneal injection with
177 streptozotocin (60 mg/kg, Sigma-Aldrich) (*Yang et al., 2013*). The remaining 9 rats were used as
178 controls. Three days after streptozotocin injection, rats with fasting blood glucose level higher than
179 16.7 mmol/L were selected as DM. After 8 weeks, DM rats were selected by the apomorphine
180 (APO) test. Briefly, APO (80 mg/kg, Sigma-Aldrich) was injected in the loose skin of neck.
181 DMED rats were identified if rats did not exhibit an erectile response within 30 minutes. ,

DMED rats were randomly divided into different treatment groups (9 rats in each group): DMED group received no treatment; ADSCs group received corpus cavernosum injection of 60 μl PBS containing 5×10^5 ADSCs; ADSCs-EGFP group received corpus cavernosum injection of 60 μl PBS containing 5×10^5 ADSCs-EGFP; ADSCs-iNOS group received corpus cavernosum injection of 60 μl PBS containing 5×10^5 ADSCs-iNOS.

Assessment of erectile function

After 2 weeks, intracavernous pressure (ICP) and mean arterial pressure (MAP) were detected to evaluate erectile function as described previously (*Yang et al., 2013*). The stimulus parameters were two voltages (2.5, 5.0 volts), frequency 15 Hz, duration 1 minute and pulse width 1.2 ms. Erectile response was measured at 2.5 and 5.0 voltages. Pressure was measured and recorded with a data acquisition system (AD Instruments Powerlab/4SP, NSW, Australia). Then, penile tissues were harvested for measurement of NO and cGMP concentration (n = 4/group) and masson trichrome staining (n = 5/group)

Masson trichrome staining

Penile tissues were embedded in paraffin and cut into 5 μm thick sections. Masson trichrome staining was performed as previously described (*Yang et al., 2013*). Image Pro Plus 6.0 software (Media Cybernetics Inc, Bethesda, USA) was used to quantitatively analyze smooth muscle content and collagen in five randomly selected specimens per group.

Statistical Analysis

Results were expressed as mean ± standard deviation. Data were analyzed with one-way analysis of variance followed by Tukey-Kramer test for post hoc comparisons using GraphPad Prism 5.0 (GraphPad Software, San Diego, USA). Normal distribution was determined by Kolmogorov-Smirnov test. $P<0.05$ was considered statistically significant.

Results

Isolation and characterization of ADSCs

The isolated ADSCs exhibited spindle-shaped morphology at passage 1 (Fig. 1A). ADSCs expanded *in vitro* and showed fibroblast-like shape at passage 4 (Fig. 1B). To characterize ADSCs, surface markers of the cultured cells at passage 4 were determined using flow cytometry. As shown in Fig. 1C, most of the cells expressed CD29 and CD90, while few cells were positive for CD34, CD45, CD73, CD105, CD31 and CD106. To identify the multi-lineage differentiation ability of ADSCs, we induced adipogenic and myogenic differentiation of ADSCs for 21 or 14 days in appropriate induction medium. Oil red-O staining indicated that ADSCs could differentiate to adipocytes (Fig. 1D). Immunofluorescence staining with an antibody against α-SMA indicated that ADSCs could differentiate to smooth muscle cells (Fig. 1E).

After culture for 5 to 7 days primary rat CCSMCs in spindle shape grew out of corpus cavernosum tissues. After about 2 weeks of culture, cells were passaged and purified by the

222 differential adherence method (Fig. 1F). The cells were identified as CCSMCs by immunostaining
223 of α-SMA (Fig. 1G).
224
225 **Overexpression of iNOS in ADSCs**
226 To overexpress iNOS in ADSCs, we infected the cells with recombinant adenovirus. 3 days
227 after infection, fluorescence microscopy was used to observe EGFP-expressing ADSCs (Fig. 2A).
228 Real-time PCR and Western blot analysis showed that the mRNA and protein expression levels of
229 iNOS were significantly higher in ADSCs-iNOS group than ADSCs-EGFP and ADSCs groups
230 (P<0.05, Fig. 2B and 2C). The expression of iNOS in ADSCs could last 14 days (Fig. 2D).
231
232 **Effects of iNOS overexpression on CCSMCs**
233 As shown in Fig. 3A, the growth rate of ADSCs-iNOS did not change significantly compared
234 with ADSCs-EGFP and ADSCs. As expected, NO concentration in supernatant of ADSCs-iNOS
235 was significantly higher than that of ADSCs and ADSCs-EGFP (P<0.05). Moreover, NO
236 concentration in supernatant of ADSCs-iNOS reached maximum level on the 7th day (P<0.05,
237 Fig. 3B). We detected the effect of ADSC-iNOS on CCSMCs using Transwell co-culture model.
238 The expression of collagen I and collagen IV in CCSMCs significantly increased in high glucose
239 concentration (P<0.05). However, significant reduction of collagen I and collagen IV expression
240 was observed in CCSMCs co-cultured with ADSCs-iNOS (P<0.05, Fig. 3C, 3D and 3E). In
241 addition, we found that TGF-β1 expression and p-Smad2/3 to Smad2/3 ratio in CCSMCs reduced
242 following co-culture with ADSCs-iNOS (P<0.05, Fig. 3F and 3G).
243
244 **Effects of ADSCs-iNOS on erectile function**
245 Finally we evaluated the effects of ADSCs-iNOS on erectile function of DMED rats. There
246 was no significant difference in initial body weight or serum glucose concentration among the 5
247 groups of rats. 14 days after the induction of diabetes, body weight was significantly lower and
248 fasting glucose concentration was significantly higher in the diabetic rats than the age-matched
249 control rats (P<0.05). However, there were no significant differences in body weight or serum
250 glucose concentration among DMED, ADSCs, ADSCs-EGFP and ADSCs-iNOS groups (Table
251 1).
252 Erectile function was measured 2 weeks after ADSCs transplantation. The ICP/MAP ratio was
253 significantly lower in DMED group than that in control group. Intracavernous injection of ADSCs
254 or ADSCs-EGFP significantly improved erectile function of DMED rats. The ICP/MAP ratio of
255 ADSCs-iNOS group was markedly elevated compared to DMED, ADSCs and ADSCs-EGFP
256 groups (all P<0.05, Fig. 4A and 4B). In addition, NO and cGMP concentrations in penile tissues
257 of ADSCs and ADSCs-EGFP groups were significantly elevated compared to DMED group,
258 although they were still significant lower compared to ADSCs-iNOS group (P<0.05, Fig. 4C and
259 4D). The mean collagen/smooth muscle cell ratio significantly increased in the DMED rats
260 compared to the control rats (P<0.05). Intracavernous injection of ADSCs or ADSCs-EGFP
261 attenuated collagen content (P<0.05). Furthermore, mean collagen/smooth muscle cell ratio in
262 ADSCs-iNOS treated rats was significantly lower than that in ADSCs or ADSCs-EGFP group

263 (P<0.05) (Fig. 4E and 4F).
264

265 Discussion

266 In this study we demonstrated that infection of ADSCs with adenovirus containing iNOS
267 expression cassette led to significantly high expression of iNOS and increased generation of NO.
268 ADSCs played a positive role in restoring DMED in rats. Furthermore, overexpression of iNOS in
269 ADSCs was proved to achieve a more significant improvement of erectile function. The
270 therapeutic effect may be achieved by increased NO generation and the suppression of collagen I
271 and collagen IV expression in the CCSMCs to decrease penile fibrosis.
272 Corpus cavernosum is composed of a loose trabecular meshwork of smooth muscle and
273 connective tissues, which are structural basis of erectile function. Ryu *et al* found that collagen
274 fiber content was significantly increased in corpus cavernosum of patients with vasculogenic ED
275 (*Ryu et al., 2004*). Further studies demonstrated that extracellular matrix such as fibronectin,
276 collagen IV and collagen I accumulated and collagen I/collagen III ratio was decreased in the
277 corpus cavernosum tissue of diabetic ED rats (*Hirata et al., 2009; Zhou et al., 2012*). Abnormal
278 extracellular matrix can cause mechanical alterations of corpus cavernosum, which may provoke
279 penile venous leakage, leading to vasculogenic ED (*Li et al., 2013*). Increased expression of TGF-
280 β1 pathway could be involved in collagen fiber accumulation and penile fibrosis (*Ryu et al., 2004*).
281 CCSMCs are the prdominant mesenchymal cell type in the corpus cavernosum. Synthesis of
282 connective tissue proteins and collagen by CCSMCs was significantly increased by exogenous
283 TGF-β1 (*Moreland et al., 1995*). In accordance with previous studies, our study showed that
284 collagen I and collagen IV expression in CCSMCs significantly increased under high glucose
285 condition and penile fibrosis significantly increased in DMED rats. Then we co-cultured CCSMCs
286 with ADSCs, and found significantly decreased synthesis of collagen I and collagen IV.
287 Furthermore, we found that TGF-β1 expression and p-Smad2/3 to Smad2/3 ratio in CCSMCs
288 significantly decreased, indicating the involvement of TGF-β1/Smad signaling pathway. However,
289 further studies are needed to elucidate precise mechanism of ADSCs' effect on CCSMCs. In *vivo*,
290 ADSCs ameliorated penile fibrosis in ED rats, maybe due to secreting exosomes enriched with
291 miR-132 and miR- let7 which inhibit the expression of tissue inhibitors of metalloproteinases and
292 stimulate the expression and activity of matrix metalloproteinases (*Gokce et al., 2014; Zhu et al.,*
293 *2017*).
294 NO/cGMP signaling pathway plays a crucial role of in erectile function. NO released from
295 nonadrenergic noncholinergic nerves and endothelium is the principal mediator of penile erection.
296 Moreover, this pathway plays an important role in ameliorating tissue fibrosis. Marta *et al* found
297 that NO inhibited TGF-β/Smad induced gene transactivation in a cGMP-dependent manner in
298 endothelial cells, leading to proteasomal degradation of phosphorylated Smad. In addition, NO
299 reduced and delayed nuclear translocation of activated Smad (*Saura et al., 2005*). cGMP
300 significantly reduced TGF-β induced upregulation of collagen I and collagen II at mRNA and
301 protein levels (*Beyer et al., 2015*). In this study, we found that iNOS overexpression in ADSCs
302 significantly enhanced NO generation and could last for 14 days. Compared to ADSCs, ADSCs-

iNOS significantly increased NO and cGMP concentrations in penile tissues, and decreased collagen I and collagen IV expression in CCSMCs to ameliorate penile fibrosis in DMED rats. These results are consistent with previous studies that the inhibition of iNOS promoted penile fibrosis. For example, Monica *et al* found that more collagen content was deposited in corpus cavernosum of iNOS knockout mouse. In addition, inactivation of iNOS gene led to exacerbated penile fibrosis in DM mouse by increasing oxidative stress and TGF-β1 expression under hyperglycemia condition (*Ferrini et al., 2010*). Similarly, L-NIL as an inhibitor of iNOS activity increased oxidative and fibrosis in the media of arteries (*Ferrini et al., 2004*).

However, other studies suggested that erectile function was impaired by iNOS expression. The induction of iNOS accompanying penil fibrosis was demonstrated in rat models of DMED, Peyronie's disease and aging-related ED (*Bivalacqua et al., 2000; Ferrini et al., 2001; Usta et al., 2003*). In addition, iNOS inhibition improved erectile function in Peyronie's disease or DMED rat. However, the application of aminoguanidine as an iNOS inhibitor was not appropriate, because aminoguanidine inhibited not only all three kinds of NOS, but also advanced glycation end products involved in tissue fibrosis. Moreover, further studies are needed to determine if elevated iNOS expression is possibly a compensatory reflection when eNOS and nNOS expression is decreased under pathological conditions.

There are some limitations in the current study. First, we did not investigate the effect of iNOS expression on the stemness of ADSCs. In our study, more NO were released from ADSCs-iNOS than ADSCs. Tapia-Limonchi *et al* found that NO maintained embryonic stem cells (ESC) pluripotency and delayed ESC differentiation by regulating Gsk3-β/β-catenin and PI3K/Akt signaling pathways (*Tapia-Limonchi et al., 2016*). The effect of NO released from ADSCs iNOS on ADSC stemness required further investigation. Second, ADSCs were only transplanted into corpus cavernosum once. Repeated injection of ADSCs may achieve better therapeutic effects but need to be confirmed in future studies. Third, we did not compare therapeutic effect of ADSCs-iNOS to that of iNOS alone. Although adenovirus act as efficient agents for gene transfer, they can activate cellular and humoral immune response in the hosts, which limits the safety and efficacy *in vivo*. The innate immune response is mediated by the adenovirus particle, but not viral transcription (*Muruve, 2004*) . In contrast, transplanted stem cells could suppress excessive immune response(*Lin, Lin & Lue, 2012*). ADSCs overexpressing iNOS may be safer and more efficient than iNOS alone.

Conclusions

In conclusion, intracavernous administration of ADSCs-iNOS improved erectile function of DMED rats. Injection of ADSCs-iNOS in a rat model of DM significantly decreased penile fibrosis, mainly due to increased NO generation and suppressed expression of collagen I and collagen IV in the CCSMCs.

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Figure 1

Figure 1. Primary culture and characterization of rat ADSCs and CCSCMs.

(A) Morphological features of ADSCs at passage 1 (primary magnification: ×100) . (B) Morphological features of ADSCs at passage 4 (primary magnification: ×100). (C) ADSCs were identified by flow cytometry at passage 4. (D) Adipogenic differentiation of ADSCs assessed by oil red-O staining (primary magnification: ×200). (E) Myogenic differentiation of ADSCs assessed by immunofluorescence staining with α-SMA antibody (green, primary magnification: ×200). (F) Morphological features of CCSMCs after purification (primary magnification: ×100) . (G) CCSMCs were assessed by immunostaining with α-SMA antibody (red, primary magnification: ×100). The nuclei were labeled with DAPI (blue).

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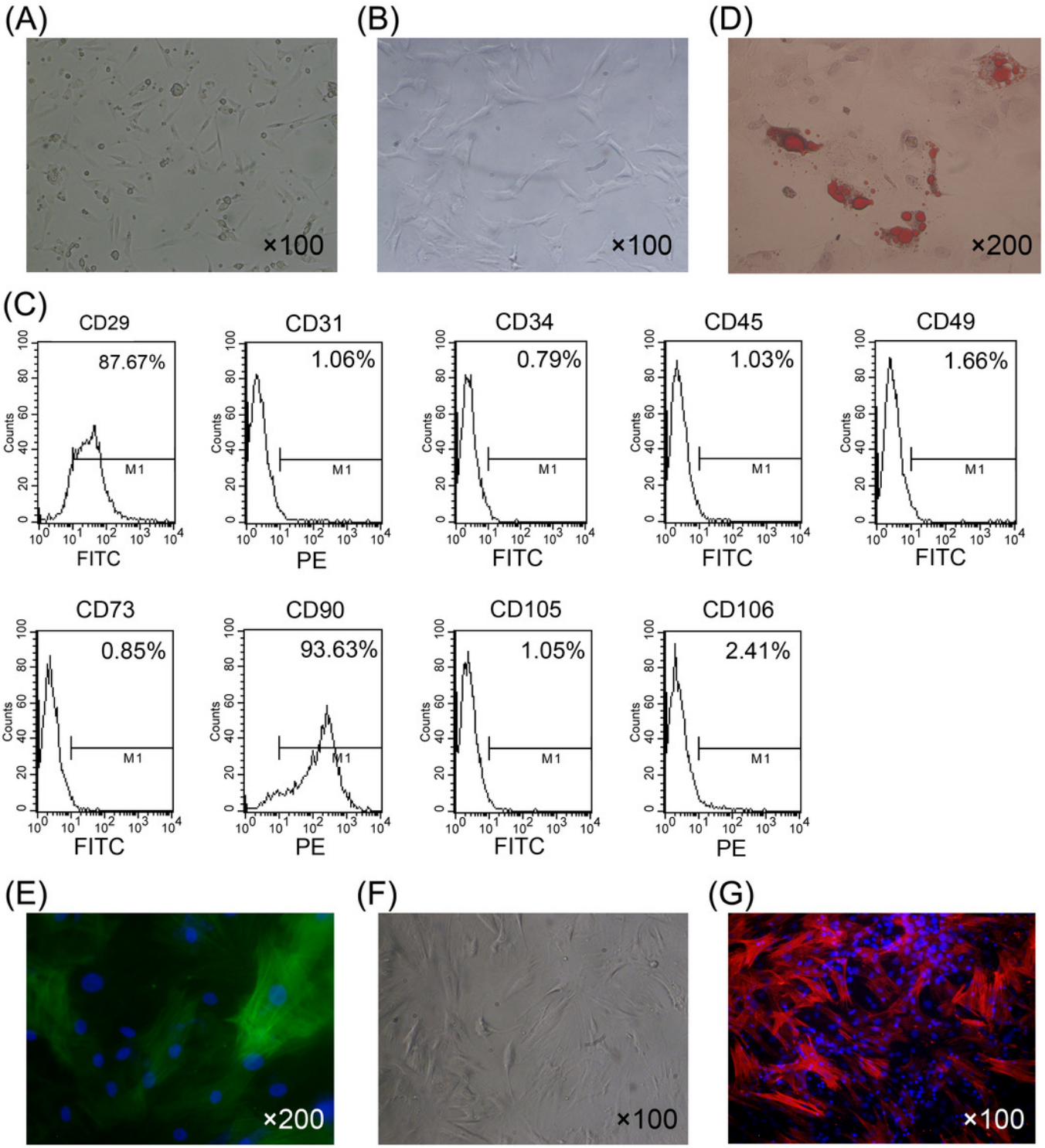
(A)
×100
(B)
×100
(D)
×200
(C)
CD29
87.67%
CD31
1.06%
CD34
0.79%
CD45
1.03%
CD49
1.66%
CD73
0.85%
CD90
93.63%
CD105
1.05%
CD106
2.41%
Counts
M1
FITC
PE
(E)
×200
(F)
×100
(G)
×100

Figure 2

Figure 2. iNOS expression in ADSCs.

(A) EGFP-expressing ADSCs were observed by fluorescence microscopy 3 days after infection (primary magnification: ×100). (B) iNOS expression at mRNA level was detected by real-time PCR and the relative ratio of iNOS/β-actin measured in ADSCs was arbitrarily presented as 1. (C) iNOS expression at protein level was detected by Western blot analysis. (D) Densitometry analysis of relative ratio of iNOS/β-actin in ADSCs and the relative ratio of iNOS/β-actin measured on 1st day was arbitrarily presented as 1. $^{*}P<0.05$ *vs.* ADSCs, $^{\#}P<0.05$ *vs.* 1d

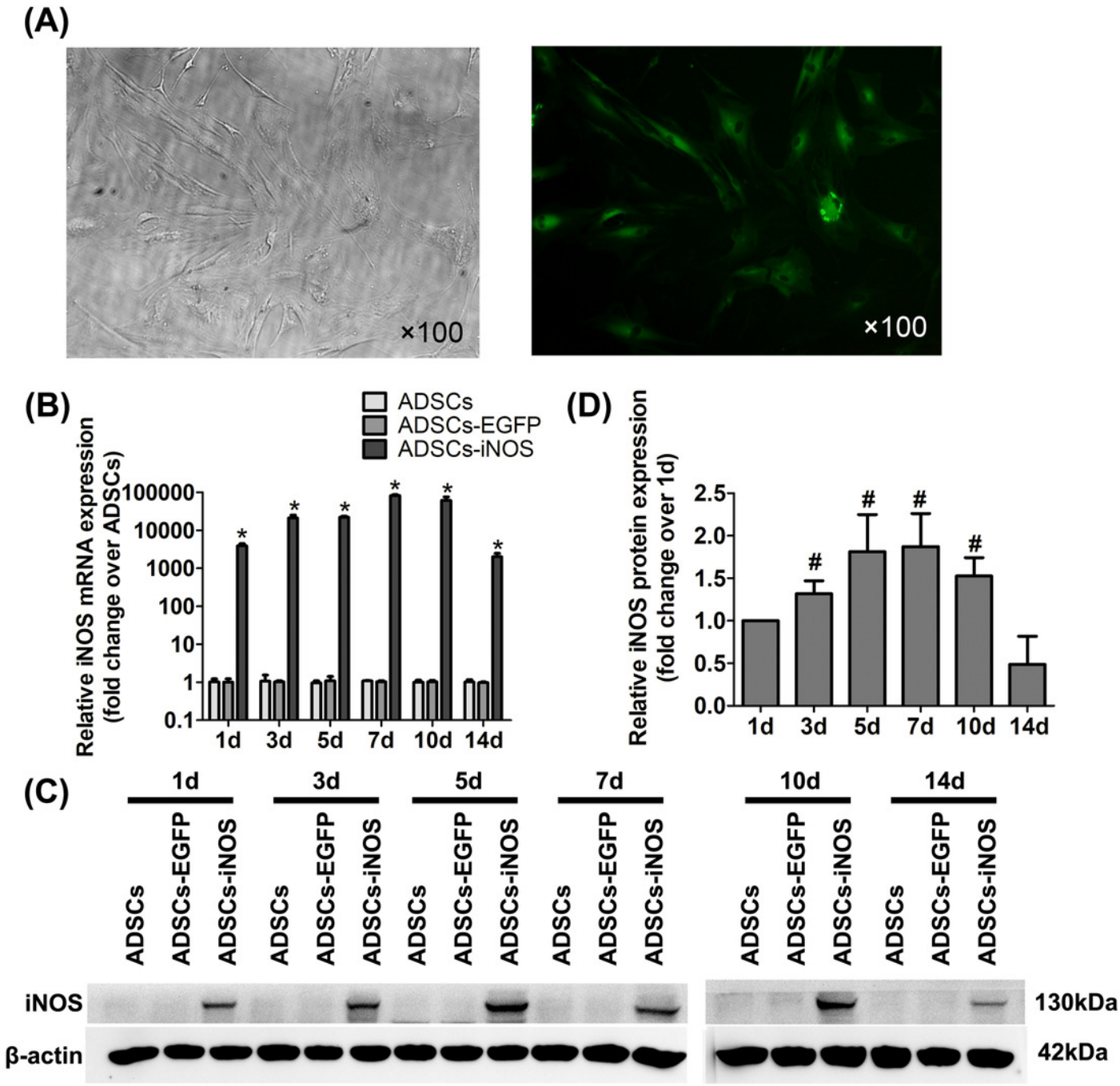
(A)
×100
×100
(B)
ADSCs
ADSCs-EGFP
ADSCs-iNOS
Relative iNOS mRNA expression (fold change over ADSCs)
100000
10000
1000
100
10
1
0.1
1d
3d
5d
7d
10d
14d
(D)
Relative iNOS protein expression (fold change over 1d)
2.5
2.0
1.5
1.0
0.5
0.0
(C)
iNOS
β-actin
130kDa
42kDa

Figure 3

Figure 3. Characterization of ADSCs-iNOS.

(A) CCK-8 assay was performed to detect the proliferation of ADSCs-iNOS. (B) NO concentration in supernatant of ADSCs was measured after incubation in 10 mmol/L L-Arginine for 24 hours. (C) The protein expression of collagen I and collagen IV in CCSMCs was detected by Western blot analysis after 7 days of co-culture. (D) Data were shown as the relative density values of collagen I to β-actin as loading control. (E) Data were shown as the relative density values of collagen IV to β-actin as loading control. (F) Data were shown as the relative density values of TGF-β1 to β-actin as loading control. (G) Data were shown as the relative density values of p-Smad2/3 to Smad2/3. $^{*}P<0.05$ *vs.* ADSCs, $^{\#}P<0.05$ *vs.* CCSMCs, $^{\&}P<0.05$ *vs.* CCSMCs cultured in DMEM containing 30mM glucose, $^{\S}P<0.05$ *vs.* CCSMCs co-cultured with ADSCs.

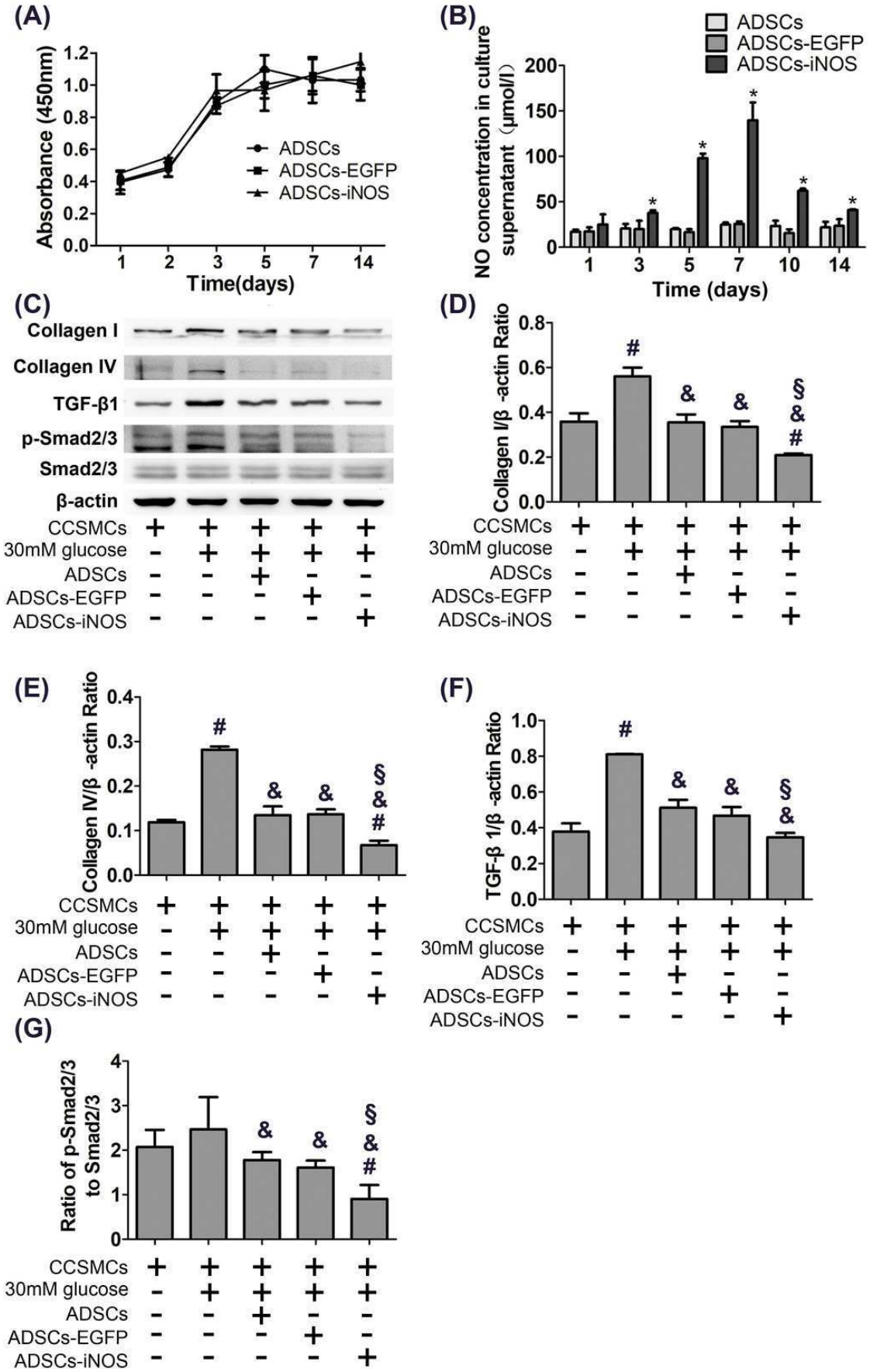
(A)
Absorbance (450nm)
Time(days)
ADSCs
ADSCs-EGFP
ADSCs-iNOS
(B)
NO concentration in culture supernatant（μmol/l）
Time (days)
ADSCs
ADSCs-EGFP
ADSCs-iNOS
(C)
Collagen I
Collagen IV
TGF-β1
p-Smad2/3
Smad2/3
β-actin
CCSMCs
30mM glucose
ADSCs
ADSCs-EGFP
ADSCs-iNOS
(D)
Collagen I/β -actin Ratio
(E)
Collagen IV/β -actin Ratio
(F)
TGF-β 1/β -actin Ratio
(G)
Ratio of p-Smad2/3 to Smad2/3

Figure 4

Figure 4. The transplantation of ADSCs-iNOS improved erectile function of DMED rats.

(A) MAP and ICP response to electrostimulation of cavernous nerves (5 Volts, 1 minute). (B) ICP/MAP ratio in control, DMED, ADSCs, ADSCs-EGFP and ADSCs-iNOS groups. (C) NO concentration in penile tissues of each group. (D) ELISA assay of cGMP concentration in penile tissues of each group. (E) Penile tissues were stained with Masson trichrome in all groups. Collagen fibers were stained blue, while smooth muscle was stained red (primary magnification: ×100). (F) The collagen to smooth muscle ratio in penile tissues of each group. $^{*}P<0.05$ *vs.* control group, $^{\#}P<0.05$ *vs.* DMED group, $^{\&}P<0.05$ *vs.* ADSCs group.

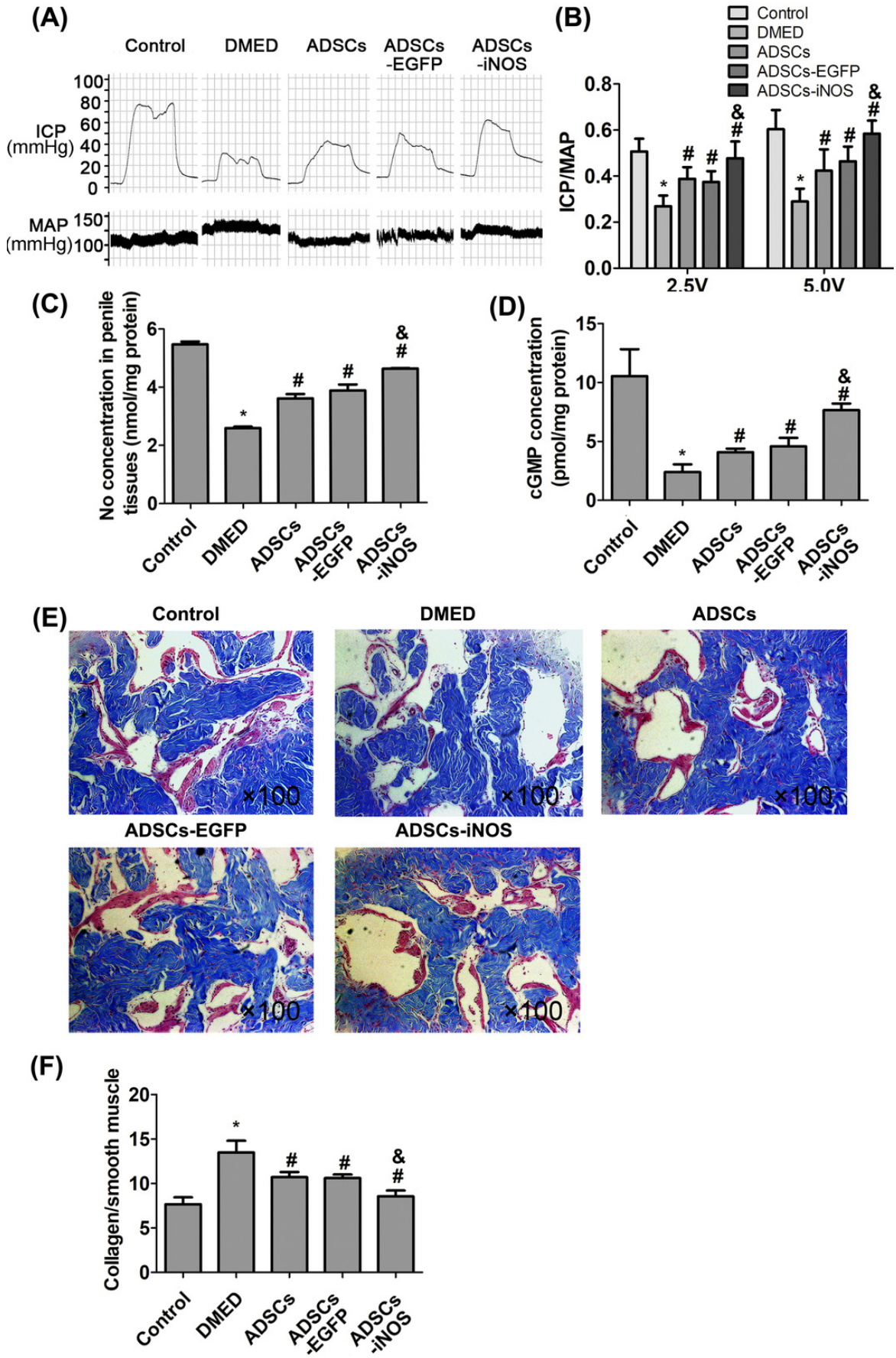
(A)
Control
DMED
ADSCs
ADSCs -EGFP
ADSCs -iNOS
ICP (mmHg)
MAP (mmHg)
(B)
Control
DMED
ADSCs
ADSCs-EGFP
ADSCs-iNOS
ICP/MAP
2.5V
5.0V
(C)
No concentration in penile tissues (nmol/mg protein)
(D)
cGMP concentration (pmol/mg protein)
(E)
×100
(F)
Collagen/smooth muscle

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Table 1 (on next page)

Table 1. Metabolic and physiological variables of experimental rats

Variable	Control	DMED	ADSCs	ADSCs-EGFP	ADSCs-iNOS
Initial weight (g)	268.0 ± 12.7	268.9 ± 8.6	268.4 ± 7.9	266.6 ± 12.9	270.7 ± 11.3
Final weight (g)	596.0 ± 54.5	274.8 ± 18.1*	284.5 ± 22.3*	281.8 ± 17.3*	285.1 ± 18.8*
Initial fasting glucose (mmol/L)	6.3 ± 0.5	6.2 ± 0.3	6.2 ± 0.4	6.2 ± 0.5	6.2 ± 0.4
Final fasting glucose (mmol/L)	6.2 ± 0.3	30.8 ± 2.0*	29.4 ± 2.1*	30.3 ± 1.6*	30.0 ± 2.8*
MAP (mmHg)	104.4 ±5.7	105.1 ± 7.7	105.8 ± 6.3	102.0 ± 7.7	108.4 ± 6.4

Note: DMED, diabetes mellitus induced erectile dysfunction; ADSCs, adipose derived stem cells; iNOS, inducible nitric oxide synthase; MAP, mean arterial pressure.*$P<0.05$ *vs.* control group. Data expressed as mean ± standard deviation.