A peer-reviewed version of this preprint was published in PeerJ on 28 January 2020.

<u>View the peer-reviewed version</u> (peerj.com/articles/8225), which is the preferred citable publication unless you specifically need to cite this preprint.

Bakker FT, Antonelli A, Clarke JA, Cook JA, Edwards SV, Ericson PGP, Faurby S, Ferrand N, Gelang M, Gillespie RG, Irestedt M, Lundin K, Larsson E, Matos-Maraví P, Müller J, von Proschwitz T, Roderick GK, Schliep A, Wahlberg N, Wiedenhoeft J, Källersjö M. 2020. The Global Museum: natural history collections and the future of evolutionary science and public education. PeerJ 8:e8225 https://doi.org/10.7717/peerj.8225



The Global Museum: natural history collections and the future of evolutionary biology and public education

Freek T. Bakker ^{Corresp. 1}, Alexandre Antonelli ², Julia Clarke ³, Joseph A Cook ⁴, Scott V Edwards ⁵, Per GP Ericson ⁶, Søren Faurby ⁷, Nuno Ferrand ⁸, Magnus Gelang ⁹, Rosemary G Gillespie ¹⁰, Martin Irestedt ⁶, Kennet Lundin ⁹, Ellen Larsson ⁷, Pável Matos-Maraví ¹¹, Johannes Müller ¹², Ted von Proschwitz ⁹, George K Roderick ¹⁰, Alexander Schliep ¹³, Niklas Wahlberg ¹⁴, John Wiedenhoeft ¹³, Mari Källersjö ¹⁵

Corresponding Author: Freek T. Bakker Email address: freek.bakker@wur.nl

Natural history museums are unique spaces for interdisciplinary research and for educational innovation. Through extensive exhibits and public programming and by hosting rich communities of amateurs, students, and researchers at all stages of their careers, they provide a place-based window to focus on integration of science and discovery, as well as a locus for community engagement. At the same time, like a synthesis radio telescope, when joined together through emerging digital resources, the global community of museums (the 'Global Museum') is more than the sum of its parts, allowing insights and answers to diverse biological, environmental, and societal questions at the global scale, across eons of time, and spanning vast diversity across the Tree of Life. We argue that, whereas natural history collections and museums began with a focus on describing the diversity and peculiarities of species on Earth, they are now increasingly leveraged in new ways that significantly expand their impact and relevance. These new directions include the possibility to ask new, often interdisciplinary questions in basic and

¹ Biosystematics Group, Wageningen University and Research, Wageningen, Netherlands

² Department of Science, Royal Botanic Gardens, Kew, Richmond, United Kingdom

³ Jackson School of Geosciences, University of Texas at Austin, Austin, Texas, United States

⁴ Museum of Southwestern Biology, Department of Biology, University of New Mexico, Albuquerque, New Mexico, United States

⁵ Department of Organismic and Evolutionary Biology, Museum of Comparative Zoology, Harvard University, Cambridge, Massachusetts, United States

⁶ Dept. of Bioinformatics and Genetics, Swedish Museum of Natural History, Stockholm, Sweden

⁷ Department of Biological and Environmental Sciences, Gothenburg Global Biodiversity Centre, University of Gothenburg, Gothenburg, Sweden

⁸ Museu de História Natural e da Ciência, Universidade do Porto, Porto, Portugal

⁹ Department of Zoology, Gothenburg Natural History Museum, Gothenburg, Sweden

Essig Museum of Entomology, Department of Environmental Science, Policy and Management, University of California, Berkeley, Berkeley, California, United States

¹¹ Department of Ecology, Institute of Entomology ASCR, České Budějovice, Czechia

 $^{^{12}}$ Leibniz-Institut für Evolutions- und Biodiversitätsforschung, Museum für Naturkunde, Berlin, Germany

¹³ Department of Computer Science and Engineering, University of Gothenburg, Gothenburg, Sweden

Department of Biology, Lund University, Lund, Sweden

¹⁵ Gothenburg Botanical Garden, Göteborg, Sweden



applied science; inform biomimetic design; and even provide solutions to climate change, global health and food security challenges. As institutions, they are incubators for cutting-edge research in biology and simultaneously protect core infrastructure for present and future societal needs. In this perspective, we discuss challenges to the realization of the full potential of natural history collections and museums to serve society. After reviewing collections and types of museums, including local and global efforts, we discuss the value of specimens and the importance of observations. We then focus on mapping and modelling of museum data (including place-based approaches and discovery), and explore the main projects, platforms and databases enabling this. We also explore ways in which improved infrastructure will allow higher quality science and increased opportunities for interdisciplinary research and communication, as well as new uses of collections. Finally, we aim to improve relevant protocols for the long-term storage of specimens and tissues, ensuring proper connection with tomorrow's technologies and hence further increasing the relevance of natural history museums.



1 AUTHOR COVER PAGE

- 2 Article submission to PeerJ
- 3 Manuscript category: Literature Review Articles
- 4 Collection: "Endless forms: Advances in evolutionary analyses of biodiversity"
- 5 Article title:

8

14

- 6 The Global Museum: natural history collections and the
- 7 future of evolutionary biology and public education
- 9 Authors: Freek T. Bakker^{1,*}, Alexandre Antonelli^{2,3,4,5,11}, Julia Clarke⁶, Joseph A. Cook⁸, Scott
- 10 V. Edwards^{2,16}, Per G.P. Ericson¹⁰, Søren Faurby^{3,4}, Nuno Ferrand¹², Magnus Gelang⁹, Rosemary
- 11 G. Gillespie¹³, Martin Irestedt¹⁰, Kennet Lundin^{3,9}, Ellen Larsson^{3,4}, Pável Matos-Maraví^{3,4,17},
- Johannes Müller¹⁴, Ted von Proschwitz⁹, George K. Roderick¹³, Alexander Schliep¹⁵, Niklas
- 13 Wahlberg⁷, John Wiedenhoeft¹⁵ and Mari Källersjö⁵
- ¹Wageningen University, Biosystematics Group, Droevendaalsesteeg 1, 6708 PB Wageningen,
- 16 The Netherlands.
- ²Harvard University, Department of Organismic and Evolutionary Biology, Museum of
- 18 Comparative Zoology, 26 Oxford St., Cambridge, MA 02138, USA.
- ³Gothenburg Global Biodiversity Centre, Box 461, SE-405 30 Göteborg, Sweden.



- ⁴University of Gothenburg, Department of Biological and Environmental Sciences, Box 461, 405
- 21 30 Göteborg, Sweden.
- ⁵Gothenburg Botanical Garden, Carl Skottsbergs gata 22A, SE-41319 Göteborg, Sweden.
- ⁶Jackson School of Geosciences, The University of Texas at Austin, 2275 Speedway Stop
- 24 C9000, Austin, Texas, 78712, USA.
- ⁷Department of Biology, Lund University, Sölvegatan 37, 223 62 Lund, Sweden
- ⁸Museum of Southwestern Biology, Department of Biology, MSC03 2020, University of New
- 27 Mexico, Albuquerque, NM 87131, USA.
- ⁹Gothenburg Natural History Museum, Box 7283, SE-402 35, Göteborg, Sweden.
- ¹⁰Department of Bioinformatics and Genetics, Swedish Museum of Natural History, PO Box
- 30 50007, Stockholm 10405, Sweden
- ¹¹Royal Botanic Gardens, Kew, TW9 3AE, Richmond, Surrey, UK.
- 32 ¹²Museu de História Natural e da Ciência, Universidade do Porto, 4099-002 Porto, Portugal
- ¹³Essig Museum of Entomology, Dept Environmental Science, Policy and Management, UC
- 34 Berkeley CA 94720-3114, USA
- 35 ¹⁴Museum für Naturkunde, Leibniz-Institut für Evolutions- und Biodiversitätsforschung,
- 36 Invalidenstraße 43, 10115 Berlin, Germany
- 37 ¹⁵University of Gothenburg, Department of Computer Science and Engineering, Rännvägen 6B,
- 38 Göteborg, Sweden.



- ¹⁶Gothenburg Centre for Advanced Studies in Science and Technology, Chalmers University of
- 40 Technology and University of Gothenburg, SE-412 96, Göteborg, Sweden.
- ¹⁷Institute of Entomology, Biology Centre CAS, 370 05 České Budějovice, Czech Republic.
- * Author for correspondence. E-mail: freek.bakker@wur.nl

43 Abstract

- Natural history museums are unique spaces for interdisciplinary research and for educational
- 45 innovation. Through extensive exhibits and public programming and by hosting rich
- 46 communities of amateurs, students, and researchers at all stages of their careers, they provide a
- 47 place-based window to focus on integration of science and discovery, as well as a locus for
- 48 community engagement. At the same time, like a synthesis radio telescope, when joined together
- 49 through emerging digital resources, the global community of museums (the 'Global Museum') is
- 50 more than the sum of its parts, allowing insights and answers to diverse biological,
- environmental, and societal questions at the global scale, across eons of time, and spanning vast
- 52 diversity across the Tree of Life. We argue that, whereas natural history collections and
- museums began with a focus on describing the diversity and peculiarities of species on Earth,
- 54 they are now increasingly leveraged in new ways that significantly expand their impact and
- relevance. These new directions include the possibility to ask new, often interdisciplinary
- 56 questions in basic and applied science; inform biomimetic design; and even provide solutions to
- 57 climate change, global health and food security challenges. As institutions, they are incubators
- for cutting-edge research in biology and simultaneously protect core infrastructure for present
- and future societal needs. In this perspective, we discuss challenges to the realization of the full
- 60 potential of natural history collections and museums to serve society. After reviewing collections



67

68

69

and types of museums, including local and global efforts, we discuss the value of specimens and

the importance of observations. We then focus on mapping and modelling of museum data

63 (including place-based approaches and discovery), and explore the main projects, platforms and

databases enabling this. We also explore ways in which improved infrastructure will allow

65 higher quality science and increased opportunities for interdisciplinary research and

66 communication, as well as new uses of collections. Finally, we aim to improve relevant protocols

for the long-term storage of specimens and tissues, ensuring proper connection with tomorrow's

technologies and hence further increasing the relevance of natural history museums.

70 **Keywords:** collections, field education, Global Museum, innovation-incubator, natural history,

71 place-based, specimens, transcriptomics, epigenomics.



Introduction

72

Natural history museums, which harbour extensive records of biological diversity, have always 73 74 been meeting places for scientists, amateurs, and the public. By visiting a natural history museum and learning about nature, the lay citizen often tacitly endorses the information 75 presented and considers it worthy of consideration and reflection. As such, museums are still 76 considered trusted resources, at a time when many other institutions are bitterly mistrusted 77 (Foley, 2015). Whereas this trustworthiness is true for most types of museum collections, natural 78 history collections play a special role, in part because at the same time that they serve in public 79 education, they are also actively used and curated by professional scientists to answer pressing 80 problems in biology and beyond. 81 Museum biological collections are more than meets the eye. Each specimen harbours many kinds 82 of data, such as information on locality and collection parameters, associated pathogens, 83 biopolymers such as DNA and proteins, and metabolic compounds. This wealth of metadata 84 across many specimens turns collections into powerful research tools, enabling scientists to test 85 for historic environmental hypotheses and carry out diverse studies ranging from public health & 86 safety (as cornerstones in studies of environmental health and epidemiology; Suarez & Tsutsui, 87 2004), biomimetic design –where naturally-occurring architectures and systems inspire 88 technological innovation (Jayaram & Full, 2016; Nirodi et al., 2018), historical genomics 89 (focussing on ancient alleles or past genotypes; Bi et al. 2013; Besnard et al. 2014), to global 90 change (tracking shifts in phenotype across specimens through time; Jones & Daehler, 2018), 91 something that a database of mere species observations cannot do. But natural history collections 92 93 face challenges. They are in need of constant sustenance, funding, and curation.



Survey Methodology

This paper aims at reviewing and conceptualising how natural history museums have had and could have a continuing and increasing role in enabling discovery in evolutionary biology and beyond. We also discuss their central role in training the next generation of innovative thinkers. The paper is the result of a three-day workshop on 'The role of museums in modern evolutionary biology' organized by Chalmers University of Technology and the University of Gothenburg (Sweden), under the auspices of the Gothenburg Centre for Advanced Studies (GoCAS), and held during 7-9 June, 2017. Participants, most of whom were based at natural history museums (broadly defined to also include botanical gardens; see below) as researchers or directors, were chosen to represent diverse areas of museum-based science. They were often based in Europe, but also the United States, South Africa and South America (Peru). After several sessions of brainstorming about the current relevance of museums and roadblocks to expanding impact, we collectively drafted an outline and all authors participated in the writing and direction of the paper.

Collections & types of museums

We define natural history museums in a broad sense, as institutions containing diverse physical specimens, and sometime also including seed banks, substantial living, frozen, or dried tissue collections, and genomic data, among others. These collections include material from rapidly disappearing extant species, as well as extinct species, many of which are from the most inaccessible parts of the Earth. Collections may have innate, historic biases in taxonomic coverage and sampling design, which might need to be considered for their further development.



118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

134

135

136

137

138

Whereas historically natural history museums, in particular those in Europe, have been linked to the colonialist enterprise, increasingly specimens are held in local museums and maintained by local scientists and students, and international specimen acquisition and study are conducted in strong partnership with these institutions. International agreements, such as the Nagoya protocol, rightly mandate such participation under the terms of Access and Benefit Sharing. Additionally, citizen science increasingly contributes to collections, which today are housed all over the world, and serve as gems of diverse global centres of cutting-edge research (see Fig. 1). Natural history museums may be located at universities, sometimes without exhibits, or may include public exhibits, such as typically occurs in national, state or regional entities. In many cases, regional collections, and their exhibits, reflect and strengthen visibility, appreciation, identity and awareness of local culture and fauna & flora, therefore playing an important and confirmative role for the visiting public. Collections that span long periods of time reflect the history of science as well as changes in norms and values in society: what was sampled, how and why. This emphasis is especially visible in open, regional, collections on display. Regionality therefore, can be considered a strength of collections and fulfills an important role in sustaining regional pride in biodiversity. On the other hand, for many scientists, usually on a global research mission, regional collections could appear to be insufficient as it needs 'global collections' (see below, The Global Museum), for instance, to capture the full extent of evolutionary and biogeographic diversity of particular clades. Still, given the increasingly connected network of regional museums, the local depth provided by such regionality would be impossible to recover – if only for practical reasons - in a single, global museum, were it composed solely of physical specimens.



140

141

142

143

144

145

146

147

148

149

150

151

152

153

154

155

156

157

158

159

160

161

The distributed nature of the world's museum collections increases long term data security. Collections of natural objects will always require a specific physical location and this makes them vulnerable to theft, fire and water damage. For instance, it was thanks to this distributed nature that we still have duplicates of the tens of thousands of specimens of plants stored at the Berlin Herbarium when it was almost completely destroyed during World War II. The Butantã Museum in São Paulo had a world-renowned alcohol collection of 85,000 snakes and half a million Arachnids. After it was drastically destroyed by a fire in 2010 (*Phillips*, 2010), it became clear that there were virtually no duplicates for these two collections, either as specimens or tissues, in other museums. Along with these concrete recent examples of natural heritage loss, the infrastructure of many museums remains underfunded, exacerbating their vulnerability. A grim example is the Brazil National Museum in Rio de Janeiro, where a fire destroyed an estimated 90% of the collections in several divisions in September 2018 (*Phillips*, 2018). Creating redundancies in collections, especially for extant species and genetic resource collections, is key to ensuring the longevity of these samples and associated data. Initiatives such as the Global Genome Biodiversity Network (GGBN; Droege et al., 2016) aim to collect, catalogue, and "democratise" genomic resources across global collections, covering 50,626 species (as of 18 march 2019). Although this enterprise represents an important step in the direction of a distributed collection, it will benefit from more coordination and financial support for data security and achieving complementarity and redundancy among collections. These aims are included in the mission of the pan-European Distributed System of Scientific Collections (www.dissco.eu) initiative, which aims to extensively digitize and database diverse specimens from across the museums of Europe. In comparison, in the US, a National Science Foundationfunded major digitization initiative, named *Integrated Digitized Biocollections* (iDigBio), aims at



making data and images for millions of biological specimens available in electronic format "for the research community, government agencies, students, educators, and the general public" (*Page et al., 2015*). iDigBio serves as "the coordinating center for the national digitization effort" fostering partnerships and innovations, and developing extensive content. iDigBio is funded by grants from the NSF Advancing Digitization of Biodiversity Collections (ADBC) program. In contrast, DiSSCo started by securing government buy-in, with content being part of operational costs. Because it has become an established and recognized entity, governments can fund the infrastructure because they need the services, thereby providing DiSSCo with guaranteed political context. At the global level, the *Global Biodiversity Information Facility* (GBIF) is "an international network and research infrastructure funded by the world's governments and aimed at providing anyone, anywhere, open access to data about all types of life on Earth." Being the main global database, a large proportion of its 1 billion records comprises observations rather than specimens (see below).

The value of specimens and the importance of observations

All collections ultimately contain, or are dependent on, specimens. A specimen may consist of a complete organism (collected by naturalists over the past few centuries) or parts of a single individual organism. Increasingly, meta-data associated with the physical specimen - the 'extended specimen' (Webster, 2017) - add value and increase data richness through videos, sound recordings, information on habitat, and photographs. For example, for birds, the extended specimen may be comprised of records of the song, or recordings of behaviour of those organisms, prepared in a way that preserves them for the future. Bioacoustic tools provide unique collections that can include some of the last known evidence of extant species. Likewise, several



186

187

188

189

190

191

192

193

194

195

196

197

198

199

200

201

202

203

204

205

206

films exist (e.g. https://www.youtube.com/watch?v=nAzqGn-LHCw) portraying the behaviour of now-extinct animal species, such as the Tasmanian tiger, the golden toad, and the Hawaiian crow. Museums currently host increasingly diverse collections, which, in addition to DNA and tissue banks, may be generated by core genomic facilities or imaging labs (isotopic, x-ray computed tomography data [CT], scanning electron microscopy [SEM] images). Examples now include rich stores of high-resolution CT data generated from museum specimens, which allow investigators to look inside material in a largely non-destructive way. These require different storage resources from those that traditionally constitute museum infrastructures, namely large scale and secure long-term storage of image data. Integration of different data streams will allow bridging among disciplines and the involvement of fields underrepresented in natural history museums, such as engineering, biomedical sciences, and art. For instance, biomimetic design can benefit strongly from inspiration from natural history collections (examples from robotics see Jayaram & Hull, 2016), or solutions to global health or food security challenges can be based on exploration of natural history specimen collections (see Table 3 Specimens and pathogens). Specimens are at the heart of the discovery process and technological advances are increasing the number and diversity of possible questions that can be addressed (e.g., Schmitt et al. 2018; see below). For instance, bone fragment identification using collagen barcoding was difficult to imagine before the rise of LC-MS technology, but Welker et al. (2015) used this to identify Palaeolithic fragments of mammal bones in France. Genomic analyses of single bone fragments can inform on the evolutionary and demographic history of our own species (e.g., Slon et al., 2018). Future technologies may include more advanced chemical, biochemical, isotope or microanatomical surveys, making maintenance of specimens even more critical because they connect



key to repeatability – a core requirement of science. 208 209 Museum specimens and data are stored and made accessible for future generations, enabling them having broad reference and context. Continued field collecting secures future access to time 210 series of specimens, collected over decades or even hundreds of years. These long-term archives 211 212 provide valuable and unique information (Graham et al. 2004) on changes in the species composition in our environments and habitats, due to factors such as climate change, human-213 mediated nitrogen deposition, or other anthropogenic activities (Meineke et al. 2019a; 2019b). 214 An example is a large survey and collection of marine invertebrates from the Swedish west coast 215 from the 1920's and 30's conducted by the Gothenburg Natural History museum (GNM), in 216 which the exact sample locations could be deduced using modern methods, and consequently 217 revisited during a new survey in the 2000's, revealing a 60% loss of biodiversity (Obst et al. 218 2017). Specimens collected by researchers 200 years in the past can be compared with 219 220 contemporary (and future) sampling-- as long as these collections and institutions persist. In fact, the specimen can be seen as the outcome of a combination of genotype and past environmental 221 change or conditions (e.g., Holmes et al., 2017), and a well-curated collection captures the 222 variation in phenotype as well as genotype (see Bi et al. 2013; Rowe et al. 2011; Staats et al. 223 2013; Ruane and Christopher, 2017). For instance, Cridland et al. (2018) comparing SNP 224 patterns from historic museum and living specimens of bees, could not only infer 'rapid change' 225 in genetic composition of honey bees in California, but also identify historic genotypes in 226 candidate genes possibly involved in adaptation to new niches. As another, less-domesticated, 227 example: reconstructing the shift to C4 photosynthesis in grasses could be conducted using DNA 228 from a 100-year-old Malagasy herbarium specimen for which both its phylogenetic placement 229

all data streams and facilitate data interoperability. At the same time, maintaining specimens is



231

232

233

234

235

236

237

238

239

240

241

242

243

244

245

246

247

248

249

250

251

and the assessment of its 'genetic make-up' with regards C4 photosynthesis could be assessed (Besnard et al., 2014). Therefore, specimen collections can provide a powerful reference for functional genomics studies, in a world where phenotyping different genotypes, retrievable from the specimen, becomes more important. Increasingly, growth of museum collections is the result of their increased relevance for ecological studies, in addition to input from taxonomically-focused collecting activities, linked to specific inventories and research projects. Whereas museum staff and associated researchers and students still undertake expeditions to increase collections and make them available for future generations, many collections now come from large scale ecological studies (e.g. NEON in the US). Specimen collections enable answers to a large number of other scientific questions, some of which have not yet been posed. The earliest museums facilitated interactions among scholartravellers, to share observational data from across the planet and to help build the core of what would become natural history and modern evolutionary biology. Increasingly, museums are leveraging new data from their specimens, and this integration of data types allows training in techniques that bridge among disciplines, as well as the generation of data sets that are of relevance to disparate traditional fields such as engineering, biomedical sciences, and art. Today natural history museums serve increasingly as a *nexus* for work that disregards disciplinary boundaries and addresses questions we did not know to ask before (see Fig 2). Because collections provide the opportunity to rigorously examine diverse aspects of taxonomic, morphological, genetic, and chemical variation across vast temporal and spatial scales, they can help diverse scientists bridge the gaps between traditionally distinct disciplines. Museum spaces ideally are filled with students who learn to think anti-disciplinarily and appreciate the



253

254

255

256

257

258

259

260

261

262

263

264

265

266

267

268

269

270

271

where a next generation of critical thinkers in biology and beyond will be trained. A specimen constitutes a voucher, not only of the actual individual sampled at the time, but often also of its locality – including information about the soil and other biotic and abiotic conditions in which the specimen was collected (see below, the 'holistic specimen'). Troudet et al. (2018) describe how over the past decades the proportion of specimen-based occurrences in GBIF has decreased from 68 to 18%, in favour of observation-based occurrences, mostly from contributions by citizen science efforts such as iNaturalist and eBird (see Table 1). This will have affected repeatability and 'richness' of systematics and evolutionary studies and the authors urge that "when impossible to secure, voucher specimens can be replaced by observation-based occurrences", particularly when combined with 'ancillary' data such as recordings, pictures, DNA samples etc. In cases where ethical, conservational, or practical concerns exist, observation data instead of collected specimens provide additional (or occasionally substitute) contributions to our knowledge on where and when particular species occur. Recorded sightings, such as those from iNaturalist or e-Bird, include occurrences of diverse temporal range, and are pretty much the only observation-based data that are allowed in GBIF. In addition to such observations recorded in the field, however, collected specimens (when available) offer additional options for confirming or extending the original work using new analytical techniques. Similarly, sound recordings can be re-studied within the context of new evidence, leading to reciprocal illumination.

importance of the specimen. These spaces can therefore be considered 'Innovation Incubators'

272

273



275

276

277

278

279

280

281

282

283

284

285

286

287

288

289

290

291

292

293

294

295

296

Place-based discovery: different specimen data sets connecting to a location

As Miller (2007) puts it, "Places are not simply a semantic convenience. It is a meaningful lens for viewing the world because it is orderly with respect to geographic space." As such, 'placebased' approaches in general focus on the characteristics and meaning of particular places as a fundamental starting point for a particular development or project. Especially in charity and community development work, place-based approaches aim at "giving power to the community in guiding systemic change" and therefore "being recognised as critical to responding effectively to certain community challenges" (www.qcoss.org.au/). In education, place-based approaches are thought to "identify, recover and increase the values of local cultural specificities" (Monardo, 2018). Place-based learning and education is well developed (Gruenewald and Smith, 2014) and provides a context for local understanding and societal change. Natural history museums are well suited for hosting place-based activities, as well as making direct links between collections and associated data and societal activities and needs. The developing Island Digital Ecosystem Avatar (IDEA) project is one example (*Davies et al., 2016*), entailing "a systems ecology open science initiative to conduct the basic scientific research needed to build use-oriented simulations (avatars) of entire social-ecological systems." Many specimens will have been collected and stored, for instance for DNA barcode reference libraries, for making this possible. For biological collections and their associated and ensuing process of discovery, the place-based approach is relevant. 'Place-based' here does not ignore external, or global, evidence or connections to other geographical localities, and indeed seeks to understand how local information and processes are interconnected with those at a larger scale. Best practices for biological collections include a geographical reference for each item, as is included in the



298

299

300

301

302

303

304

305

306

307

308

309

310

311

312

313

314

315

316

317

318

Darwin Core (see https://dwc.tdwg.org/). When collected together, sets of items are necessarily place-based. In addition to standardised metadata directly associated with biological items (Kissling et al., 2018), many other types of information are place-based and can be collected at the same location and super-imposed on point specimen data. Examples include information about geology, ground and atmospheric chemistry, and archaeology. These, and other data layers - such as from GBIF (species occurrences), NCBI (DNA and amino acid sequences), Open Tree of Life (phylogenetic trees), Map of Life (abundance data), TraitBase (traits), GloBI (biotic interactions), see Table 1 - can be associated or combined with geographical location through a geographical information system (HOLOS), integrating across diverse data types and enabling testing hypotheses concerning causal impacts - the 'holistic specimen'. In a sense, this approach is comparable to correlative species distribution modelling (SDM) approaches such as using Maximum Entropy (MaxEnt;, Philips & Dudik, 2008), focusing on mostly abiotic and edaphic correlates. In addition, the place-based approach can provide a baseline for understanding changes over time (Billick et al., 2013; National Research Council, 2014). Of particular interest here is the context of historical environmental change to which current changes can be evaluated and compared. An understanding of historical processes provides a means for predicting, or forecasting, how biological systems may respond to change in the near future. For instance, Willis et al. (2008) studied how climate change may affect phenology in some angiosperm species in Concord, Massachusetts. Slingsby et al. (2017) studied the interaction between fire and climate change on species diversity in the South African Cape Floristic Region, allowing modelling of future vegetation response.



In general, the additional value of place-based approach for scientific discovery includes the serendipity of collecting data over periods of change identified later, as well as the interaction of researchers sharing an interest in the same geographical location or region (*Michener et al.*, 2009). Place-based initiatives associated with larger networks (see Table 1) can provide access and understanding to a diversity of communities, which is both democratic and allows broad participation in discovery. Examples of such initiatives include developing new natural history museums (Darwin Initiative, www.darwininitiative.org.uk) and DNA barcoding of local biodiversity (*van de Bank et al.*, 2008; *Janzen & Hallwachs*, 2016). In addition to natural history museums, the benefits of a place-based approach are also shared with field stations, botanical gardens, and biological reserves (*National Research Council*, 2014).

The Global Museum

As indicated above, many museums serve regional communities, and collections in such institutions usually reflect regional interests, fauna and flora, funding and research questions. Given that science is an international endeavour, the question can be asked as to whether evolutionary biology would be better served by enhanced ability to document and analyse patterns across regions, such as with the use of GBIF. For instance, for taxonomy, having the virtual, global, workbench of the Barcode Of Life Database BOLD (www.boldsystems.org; *Ratnasingham & Hebert, 2007*) allowed taxonomists globally to harmonise species delineations by collectively analyzing and interpreting DNA barcode patterns from global rather than regional data sets. Historically however, collections have been mainly curiosity-driven, emphasizing rarities (i.e. single individuals per species, a phenomenon which may actually be commonly-occurring), a pattern that still exists for certain organism groups and regions (*Novotný & Basset*,



343

344

345

346

347

348

349

350

351

352

353

354

355

356

357

358

359

360

361

362

363

364

2000; ter Steege et al., 2011), and mostly the result of general collecting efforts. This generalism has resulted in an invaluable reference specimen base in today's museum collections, allowing comparison with living specimens, identifying relatives of medically- and economicallyimportant species (for instance melon, Sebastian et al., 2010), or testing historic biological hypotheses (e.g. Délye et al. 2013). On the other hand, this patchy tradition of biological collecting has come at a cost to easily comparing organisms across large geographic regions or across temporal spans. For example, evolutionary biology would benefit from being able to analyze more common species represented in collections worldwide, because this would allow assessing phenotypic variation at much broader scales. In addition, assumptions about species ID based on morphology may be falsified by genetic data (DeSalle et al. 2005) - but also the reverse - revealing an unexpectedly high level of cryptic diversity in certain groups (e.g., Hebert et al., 2004). Such a mandate would require a concerted effort of museums globally to collect and archive specimens in a coordinated manner that would help document current biodiversity and variation of common species across the globe. Such an effort was originally planned to be conducted by NEON in the United States, but in some cases has fallen short of this goal (Cook et al., 2016). Other ventures include the above-mentioned BOLD (with iBOL extending its coverage) which holds 6.6M barcode records across 0.29M species, many of which are commonly-occurring. Future collections should continue to expand with specimens sampled widely across biodiversity, but in addition should amass commonly-occurring species, which can serve as environmental monitors, especially when sufficient metadata is also collected. Museum communities are increasingly not confined by a single, local physical space but able to distribute their reach through innovations in technology. Databases and other online tools enable international access and an array of novel platforms facilitate participation of a broad swath of



the public in discovery and documentation, from undergraduate classes to young children 365 participating in aspects of the scientific process. Examples of such activities include encouraging 366 children to make observations of butterflies in drawers, thereby building their sense of 367 biodiversity. Another example is citizen science projects in which volunteers help in interpreting 368 and digitizing information on old collection labels, as has been done for the Paris Herbarium 369 (http://lesherbonautes.mnhn.fr/) and for brachiopod fossils at the Swedish Museum of Natural 370 History. 371 From a Global Museum perspective, we may ask whether phenomena such as global change 372 have been effectively documented in collections in the past so that we can use that ensemble of 373 past collections to forecast future conditions. For instance, collections can help scientists 374 document how C4 photosynthesizing plants have spread during recent decades as a response to 375 the global increase in atmospheric CO2 concentration (Besnard et al., 2014), or how species 376 extinctions may be overrepresented in particular clades or areas (e.g., Ricklefs, 2006). Such work 377 would be impossible without having the integrated, properly digitised and databased platform 378 that a Global Museum provides. 379 Large international data sharing initiatives (e.g. LifeWatch ERIC, GBIF, Encyclopedia of Life, 380 BOLD and iBOL, see Table 1) allow access to collections by scientists and the public living far 381 from privileged historic western centres for inquiry. For instance, GBIF alone provides access to 382 now over a billion records of specimens and observations from around the world. iDigBio, 383 GBIF, and the Atlas of Living Australia (ALA) and its affiliated atlasses are the world's largest 384 and best-developed biodiversity data aggregators and mobilizers. As outlined above, DiSSCo is a 385 developing initiative of major significance that will unify natural science collections in Europe. 386 With increasingly distributed access to large datasets and online portals to large-scale 387



389

390

391

392

393

394

395

396

397

398

399

400

401

402

403

404

405

computational resources, analysis of the "big data" of biodiversity records can also go global (see Antonelli et al. (2015) for an example in global angiosperm biogeography and speciation). Digitization of collections will be increasingly important in this respect; there are many valuable but undigitized collections residing in museums. Citizen science can contribute significantly to building collections, as for instance seen in many entomological collections that grow these days by amateur entomologists donating their well curated collections for posterity. eBird and iNaturalist are excellent examples of connecting citizen with science in a highly-efficient manner (and then feeding into GBIF). The Gothenburg Natural History Museum (GNM) malacological collections have benefitted tremendously by citizen science efforts with devoted ('advanced') amateurs donating their often well-curated private collections. Based on these collections, which can be considered 'environmental archives', Bolotov et al. (2018) could infer from freshwater pearl mussel collections that morphology has changed in time correlated to environmental alteration and climate change. Based on historical and recent specimens from extensive geographical sampling, the authors concluded that the latter may well have accelerated the population decline in pearl mussels over the last 100 years. The study underlines the importance of preserving large collections (many individuals) to enable meaningful statistical analysis of morphological measurements. Table 4 lists another example from the GNM concerning garden slugs sent in by the general public.

406

407

408

409

Further increasing the relevance of museums through digitization and imaging

To facilitate the coordination of collection and databasing efforts between museums – a necessity to achieve the Global Museum – it is vital to increase awareness of what knowledge is available,



not only at regional museums but across museums globally. This is most easily achievable 410 through digitization of the collections. Moreover, such digitization also opens the collections for 411 a number of additional researchers interested in overall temporal or spatial patterns in 412 biodiversity. iDigBio provides a good example of how digitization can be successful and 413 provides outreach to a global user base. In its first 10 years, iDigBio has prioritized digitization 414 415 of specimens that can drive collaborative research and answer specific biological questions. This specialization necessarily results in only a small fraction of available specimens being digitized. 416 A major question for the future is how the community should greatly expand the scope of 417 digitized specimens. Just as haphazardly collected historical specimens often prove useful for 418 research questions not envisioned during the collecting event, it is also likely that specimens 419 digitized without a specific research question in mind will prove useful for answering scientific 420 or societal questions, especially if digitized on a large scale. But digitizing 'blindly' must of 421 course be balanced with the pressure of meagre resources; enabling citizen scientists to assist 422 423 offers a good solution (Rouhan et al., 2016; see Table 4). Large amounts of metadata remain to be digitized and would generate knowledge on biogeography (geographic data of specimens), 424 disease spread (genetic material from parasites), biological interactions (pollination data), 425 phenology, or shifts in species distributions (Suarez & Tsutsui, 2004; James & al., 2018). New 426 advances in image recognition through deep learning using neural networks are also likely to 427 enable easy identification of many species, such as already implemented in the iNaturalist 428 platform, and hence help digitization. 429 Tracking specimen taxonomy. For research on temporal patterns of global change, such as global 430 431 warming studies or analyses of movement of hybrid zones, museum records provide a unique source of historical records. Because they are backed up by physical specimens, records can be 432



identified to individual species irrespective of how many taxonomic changes have occurred since 433 specimen collection, an endeavour that is often impossible if the records were purely 434 observational. However, an important caveat of this effort is that it is rarely obvious from online 435 databases on what taxonomical opinion each identification was based. Indeed, to capitalize on 436 the inherent strength of specimen-based records a citation to the taxonomic revision behind each 437 identification would be ideal, but is not always available. 438 Combining specimen databases. Another challenge related to digitization and to the value of the 439 physical specimens is that specimens may be one of the only clear, if frequently underutilized, 440 ways to identify duplicates between different databases. For example, this challenge is often 441 encountered among mammalian fossils where several databases, including New and Old World 442 Mammals (NOW) and PaleoDB, have been started independently. Together these databases 443 completely document diversity for some groups, and out of ~1586 accepted species of carnivores 444 and relatives (Carnivora, Hyanodonta and Oxyaenidae) in the fossil record, 1460 species are 445 446 included in at least one of them (Faurby, Werdelin, Antonelli, unpublished). However, individually they are each highly incomplete (1121 species (6385 records) in NOW, 1040 species 447 (6756) in PaleoDB). Analyses attempting to summarize the complete fossil record therefore 448 needs to combine databases. For many uses, such as when estimating speciation or extinction 449 rates (Silvestro & al., 2014), it would be highly desirable to resolve redundancy between 450 databases, a challenging effort that could be made trivial if both databases contained the museum 451 specimen ids for the records. 452 Big clades, large collections. Digitization will be extremely challenging for the most diverse 453 454 taxonomic groups, such as typically large collections of insects, although there have been attempts to automate digitization of such groups (Hudson & al., 2015). Knowing the limits in 455



common groups, may suffice for further scientific analysis. Finer scale distributional data in such 457 cases can easily be supplemented by citizen science initiatives (e.g iNaturalist), especially if 458 associated with pictures or movies with smartphones that contain georeferenced and 459 timestamped records that facilitate re-validation through inspection of the images by experts (see 460 for instance iSpot; Silvertown et al., 2015). 461 Museums still harbour large amounts of undiscovered and undocumented information. The total 462 number of specimens deposited in museum collections around the world may be as large as 1-2 463 billion (Ariño, 2010), and for herbaria an estimated 350 M specimens are known to be deposited 464 in 3400 collections world-wide (Soltis, 2007). Moreover, statistical approaches to estimate the 465 size of collections agreed in 2010 that less than 5% of the universe of natural history collections 466 data is available in databases such as GBIF (Ariño, 2010), although this fraction has been 467 decreasing, with the fast increase of observation data in GBIF (Troudet et al., 2018). Wilson 468 469 (2003) noted that the smaller the organism the more poorly known the group to which it belongs, exemplified by fungi, nematodes and microbes. For instance, a random selection of specimens 470 collected in a tropical rain forest and deposited in jars at a natural history collection resulted in 471 the description of almost 200 new species of ichneumonid parasitoid wasps to science 472 (Veijalainen et al., 2012). Bebber et al. (2010) described a comparable case for angiosperm 473 species, with an estimated 35,000 undescribed species already residing in herbarium collections. 474 An abundance of undescribed species is only the tip of the iceberg on the amount of data 475 undiscovered and undocumented in the world's museums. 476 477 In terms of species diversity, DNA barcoding reference libraries such as BOLD (Ratnasingham & Hebert, 2007) and UNITE (Koljag & al., 2013) provide a good framework against which the 478

geographical and temporal distribution and the size of the collection, especially for the most



extent of diversity deposited at natural history collections can be measured. However, molecular 479 approaches to identify hidden diversity remain debatable (e.g., Brower, 2006) but can be 480 overcome in large clades such as Lepidoptera (*Hebert et al.*, 2010). In UNITE fungal species 481 hypotheses are generated and named, but also tagged with a citable digital object identifiers 482 (DOIs) so it can be unambiguously communicated, allowing harmonisation of species concepts 483 throughout communities. In comparison, BOLD allows barcode index numbers (BINs) to refer to 484 barcode clusters that have not been yet described taxonomically. 485 Programs like the UK Darwin Initiative to train observers and scientists in countries rich in 486 diversity but low in funding for conservation and science surveys can further support 487 democratization of not just specimens and data but also the knowledge for performing analyses 488 and conducting research. However, much more needs to be done in this area, especially in 489 capacity building, infrastructural development, and task distribution. 490 Reference collections. A major need for collections worldwide is to develop basic molecular data 491 associated with a given taxon. The key importance is the burgeoning use of metabarcoding in 492 ecological studies can be anchored to museum specimens, and thus linked to the associated 493 494 metadata. Many museums have embarked on such endeavors, for example in CSIRO, and efforts of multiple museums ideally come together in clade-based DNA barcode projects in BOLD. 495 To enhance the broader relevance of natural history museums it is also important to message 496 effectively to industry and policy makers. In particular, the museum community should explore 497 ways to use specimens to find novel ways to bridge the traditional chasm between the sciences, 498 arts and humanities. Shared themes include place-based research and experiential learning, both 499 encouraged in instructional efforts in science, technology, engineering and mathematics (STEM 500 fields) and the arts. Both the arts and sciences depend on inspiration, creativity, and critical 501



503

504

505

506

507

508

509

510

511

512

513

514

515

516

517

518

519

520

assessment, and museum specimens serve well as sources for both inspiration and fascination. While scientific education and research offer rigorous methods for testing hypotheses and creating new knowledge, integration of experiential art and humanities work can into science fosters non-traditional ways of exploring and messaging about our world (Balengee 2010). Natural history museums should continue their efforts to train scientists and artists to develop novel solutions to emerging problems, especially as we face an increasingly uncertain environmental future. Efforts by the Global Museum to assemble collections that will fulfil their key roles in the future require facilitating international agreement and participation. Such a massive effort cannot remain the province of a relatively few marginally resourced programs. Identifying the answers to the most pressing questions facing society and our environment require fertile spaces for cultivating innovation in the context of training in knowledge of biodiversity. This task is impossible without museum spaces and collection resources. We cannot afford to 'waste' the potential of natural specimens due to degradation, improper storage, or disposal, especially in the light of rapid biodiversity loss. They need space-efficient, climate-controlled and pest-free spaces. Innovations in these areas are likely needed to accommodate collections in the long term (hundreds to thousands of years) and to deal with preservation issues that may be exacerbated by global climate change (including increasing frequency of extreme weathers, hurricanes, flooding).

521

522

523

Public perception of natural history museums



525

526

527

528

529

530

531

532

533

534

535

536

537

538

539

540

541

542

543

544

545

546

We argue that natural history museums should be regarded as 'Innovation Incubators', places where 'anti-disciplinary' science is thriving by building bridges between otherwise or so far improbable disciplines, and scientists from various 'disciplines' meet, an Academic Nexus of Integration. Because natural history collections can facilitate examining diverse aspects of taxonomic, morphological, genetic, and chemical variation across vast temporal and spatial scales, they can help diverse scientists bridge the gaps between traditional disciplines. In places where this situation is not yet in place the way to get there would be to enable ready access to both collections and research facilities, an effort that has been highly successful, for example, under the European SYNTHESYS Access scheme for the last decade. The K-12 education project "Exploring California Biodiversity" at the University of California, Berkeley (http://gk12calbio.berkeley.edu; Mitchell & Gillespie, 2007), which takes grade school students and teachers into the field, provides an excellent example of natural history museum collections broadening access and opportunities for education. Efforts such as AIM-UP! combined the expertise of educators, curators, collection managers, database managers, and others in undergraduate education (Cook et al., 2016; Lacey et al., 2017). In terms of public perception of natural history museums, it is important to safeguard their role in society and justify long-term funding by continuing outreach and engaging the general public by proper messaging, for instance by initiating citizen science projects. In some sectors of the US it is still clear that the public misunderstands the mission of museums and does not appreciate the need for continued responsible collecting. One recent example is the unwarranted overreaction against scientific collecting of a bird specimen from the Solomon Islands, information about which was placed on the web by well-meaning media directors at the American Museum of Natural History (Johnson, 2018). This sad event, which resulted in death threats and cyber-



harassment of the scientist involved, shows that many people see only the destructive, indeed amoral, aspects of collecting of individual specimens, and do not connect this act with the many beneficial services of museums to science and society. Additionally, the public in this case did not appreciate the relative insignificance of scientific collecting as an agent of species loss as compared with habitat loss and introduced or feral predators, such as house cats. In this case, the public seemed to place undue emphasis on the loss of life incurred by collection of a single bird, suggesting much more relevance of an emotional response rather than a scientific appraisal of the true impact of collecting data on species biology. Clearly, museum curators and scientists need to join forces in working proactively with the public to increase their awareness of, and appreciation for, the practice of rigorous biological sampling.

Collecting for the future: integrated analysis of museum specimens for evolutionary

biology

Museums need room to grow in targeted ways that will allow us to address scientific issues critical to looming societal issues such as emerging pathogens and food security (*Morrison et al.*, 2017; Schindel & Cook, 2018) (see Box 2). Specimen-based field work should aim to preserve extensive sets of natural history material at a particular time and place that would represent multiple individuals of each species, multiple species per collecting locale, and multiple diverse aspects of individual specimens. For example, collection of mammals and their associated ectoparasites and digestive tracts has led to detailed understanding of co-evolution of hosts and parasites (Cook et al., 2017) and can fuel future studies of the role of the microbiome in such processes (Roggenbuck et al., 2014, Greiman et al., 2018). Such holistic collection events can better capture the complex interactions of biotic communities and, if repeated, over time could



can best leverage collecting activities across the Global Museum and that planning should lead to 571 a global effort to more rigorously inventory biodiversity. 572 Genomics is a key source of information and rapidly changing area in which the scope and 573 potential of future applications are particularly promising. Nonetheless, there are several factors 574 575 known to limit the utility of specimens for genomic analyses. For example, using 'methylated spirit' (methanol containing alcohol) instead of pure alcohol for field preservation of animal 576 tissue can severely hamper retrieval of usable DNA later on (Post & al., 1998) (see Box 1). Heat 577 treatment of plants, as typically applied in most historic herbarium collections, was found to 578 lower genomic copy numbers but not cause significant miscoding lesions (Bakker, 2015; Staats 579 et al., 2011). Conventional x-rays (as opposed to x-ray computed tomography with digital 580 imaging) of mummies and bone or using pesticides on insect collections all negatively affect or 581 destroy DNA (Gotherstrom et al., 1995). Use of formalin to preserve specimens limits extraction 582 583 of usable DNA from both animal or plant tissues as it causes cross-links among DNA molecules, preventing PCR (Ruane and Christopher, 2017; McGuire et al., 2018). 584 For historic samples, significant progress in securing biopolymers has been made and museum 585 and ancient genomics has attracted considerable interest, from researchers and industry 586 (Hofreiter & al., 2015; Lindqvist & Rajora, 2019). Still, although some DNA sequencing 587 technologies work well with degraded DNA, such as in herbarium DNA using Illumina 588 sequencing (Staats & al., 2013; Bakker et al., 2016; Hart et al., 2016), single-molecule, '3rd 589 generation', genome sequencing will never be applicable for most museum-preserved specimens 590 591 given the fragmented nature of their DNA. In contrast, for cryo-specimens preserved in liquid nitrogen in tissue banks, 3rd generation sequencing may well be highly successful. But even here, 592

provide key insights into changing conditions. Discussions should be held that address how we



some of us have noted poor DNA retrieval from tissues collected with standard field-protocols 593 and ultimately preserved cryogenically (S. Edwards, pers. obs.). For birds, best practices for 594 genome sequencing may not include freezing in the short term, which can fragment DNA, but 595 rather unfrozen archiving of blood, which will preserve the longest DNA fragments. On the 596 other hand, whether or not EDTA or 95% EtOH was used for DNA sample storage can be 597 important too for successful long read sequencing (MI, pers. obs.). 598 'Re-sequencing', i.e. sequencing and mapping reads against a related reference genome 599 sequence, has been successful in museum plants, fungi and insects (Bi et al., 2014; Staats et al., 600 2013). For organisms with relatively small genome sizes, such as birds the price for re-601 sequencing a genome from a study skin has become so low that curators of bird collections may 602 consider to actually requiring complete genome sequencing for tissue from old museum samples. 603 In this case, all parties ideally would benefit, the user for having access to the specimen, the 604 museum for putting a halt to further specimen deterioration (as the genome sequence has been 605 606 generated), and the next user for have both specimen and genome sequence available. That said, it is difficult to predict how DNA extraction techniques may evolve, and perhaps require 607 considerably lower tissue amounts to produce higher DNA yields, meaning that high-throughput 608 DNA extraction without an immediate use (DNA banking) is not an obvious choice for 609 museums. 610 611 Often scientists endeavour to see inside museum specimens. Previous approaches such as dissection or histology are invasive techniques that necessarily result in the destruction of other 612 data. Although recent imaging techniques (diceCT; Gignac et al., 2016) enable largely non-613 614 destructive work on these questions in non-model organisms preserved in alcohol, they do not completely ameliorate data loss due to selectivity in field materials preserved. Specimen field 615



617

618

619

620

621

622

623

624

625

626

627

628

629

630

631

632

633

634

635

636

637

638

preparations may include freezing fresh tissue for DNA, preserving skeletons and skins but removing most internal organs and muscles. The latter obviously limits the kinds and diversity of research that can ultimately be performed on such specimens. For example, the vocal organ of birds was often not collected in birds despite the perceived importance of bird song and other vocalization. Some now broadly used imaging techniques (e.g. diceCT) have not been studied for their effects on DNA/RNA amplification from formalin or alcohol-preserved specimens, and it is unknown if they further inhibit downstream molecular work involving these specimens. For most large multicellular organisms, it is challenging to collect large numbers of tissues. However, more portions of an organism can be feasibly preserved before discarding tissues when making new collections, particularly of common, easily accessed species. For example, at the Museum of Comparative Zoology at Harvard University, a typical avian specimen is now accompanied by 7-10 cryovials filled with DNA- and RNA-ready tissues from different organs, as well as at least one tube of unfrozen but refrigerated blood for genome sequencing. Such sampling will no doubt pose space challenges for long-term storage (which could be partially solved through the use of space-efficient biobanks), but is essential for a deep understanding of the effects of anthropogenic change on biodiversity (Schmitt et al., 2018). Integrating new imaging techniques into museum work flows will increase documentation prior to destruction (e.g. for genomic work). For instance, the Thermal Age Web Tool (http://thermal-age.eu/) was developed to help collections managers and users to quantify the risks associated with destructive analysis of specimens, based on calculated 'thermal ages' (Smith et al., 2003). The Synthesis of Systematic Resources programme (see http://www.synthesys.info/joint-researchactivities/) provides further recommendations for non-destructive sampling of museum specimens and decision analysis as to how to best sample specimens for genomic research.



Lower price points for acquisition of genome data and some imagining techniques makes defining these best practices more urgent.

641

642

643

644

645

646

647

648

649

650

651

652

653

654

655

656

657

658

659

660

Conclusions

For hundreds of years, natural history museums around the world have provided the general public and scientists with numerous opportunities to learn more about our natural world. Taken together, this 'Global Museum' must be seen as one of the most valuable assets of modern society and culture, providing the material to address challenges facing humanity today – such as baseline information against which to test hypotheses of local and global environmental change – and a critical regional cultural touchstone for the public. Natural history museums can function as inter-disciplinary meeting places, or innovation incubators, where questions are addressed that we did not consider asking before. The core of these institutions are the specimens. To maximise their use, it is therefore imperative to carefully consider how to best sample, preserve, handle, and store specimens in ways that not only meet today's demands but also new, unforeseen needs. Viewing natural history museums as critical infrastructure for scientific inquiry and public understanding may help raise their profile and awareness, facilitating continued support. Despite their immense value, natural history museums are facing grand challenges. Taxonomic expertise is decreasing for many organism groups or is not represented in the curation of some collections. Funding often relies on public sources and may be adversely affected by political and socio-economic changes, comprising the long-term continuity of a museum's activities. New international regulations on the collection, export and use of specimens for non-commercial and commercial purposes are now increasing administrative burdens and may prevent further



661	development of collections. Cross-institutional, international coordination of secure data
662	standards has not yet been fully realised. We urge scientists, citizens, and policy-makers to give
663	natural history museums the proper recognition they deserve as data archives, innovation hubs,
664	and Academic Nexūs of Integration.
665	
666	Acknowledgements
667	This paper is the result of a Workshop on 'The role of museums in modern evolutionary biology
668	Workshop' in June 2017, organized by Chalmers University of Technology and the University of
669	Gothenburg (Sweden), under the auspices of the Gothenburg Centre for Advanced Studies
670	(GoCAS). We are very thankful to Karin Hårding, Gunnar Nyman, and Mattias Marklund for
671	their continuous support and assistance in the program as GoCAS organizers.
672	
673	Funding
674	We acknowledge funding from the following sources: the Swedish Research Council
675	(B0569601), the European Research Council under the European Union's Seventh Framework
676	Programme (FP/2007-2013, ERC Grant Agreement 331024), the Swedish Foundation for
677	Strategic Research, a Wallenberg Academy Fellowship, the Faculty of Sciences at the University
678	of Gothenburg, the Wenner-Gren Foundations, and the David Rockefeller Center for Latin
679	American Studies at Harvard University to A.A.; the Marie Sklodowska-Curie research
680	fellowship (European Commission, project MARIPOSAS-704035) to P.MM.





683	Figure legends
684	Figure 1. The centrality of natural history collections to evolutionary biology and public
685	understanding. Users, contributors and stakeholders of natural history collections are indicated;
686	yellow arrows represent data flow, green arrows the flow of specimens.
687	
688	Figure 2. Natural history museum specimens drive the cycle between inspiration and innovation,
689	which spans across exhibitions and collections.
690	
691	Figure 3. Word cloud illustrating relative abundance of topics and themes covered in this study.
692	



694	References

695 Alivisatos AP, Blaser MJ, Brodie EL, Chun M, Dangl JL, Donohue TJ, Dorrestein PC, Gilbert JA, Green JL, Jansson JK, Knight R, Maxon ME, McFall-Ngai MJ, Miller JF, Pollard KS, Ruby EG, Taha SA, Unified 696 Microbiome Initiative Consortium. 2015. A unified initiative to harness Earth's microbiomes. Science 697 350:507-508 DOI: 10.1126/science.aac8480. 698 699 Ariño AH. 2010. Approaches to estimating the universe of natural history collections data. Biodiversity Informatics 700 7:81-92. 701 Bakker FT. 2015. DNA sequences from plant herbarium tissue. In: Hörandl E, Appelhans M, eds. Next-generation 702 sequencing in plant systematics, p271-284. International Association for Plant Taxonomy (IAPT) 703 Bakker FT, Lei D, Yu J, Mohammadin S, Wei Z, Van de Kerke S, Gravendeel B, Nieuwenhuis M, Staats M, 704 Alquezar-Planas DE, Holmer R. 2016. Herbarium genomics: Plastome sequence assembly from a range of 705 herbarium specimens using an terative organelle genome assembly (IOGA) pipeline. Biological Journal of the 706 Linnean Society. 117:33–43 http://dx.doi.org/10.1111/bij.12642. 707 Balengee, B. 2010. Malamp, the occurrence of deformities in amphibians, Arts Catalyst, London and Yorkshire 708 Sculpture Park, Wakefield, England. 709 Bank M van de, Olivier M, Duthoit S, Savolainen V. 2008. A DNA barcode for the flora of the Kruger National 710 Park (South Africa). South African Journal of Botany 74:370–371 10.1016/j.sajb.2008.01.073. Bebber DP, Carine MA, Wood JRI, Wortley AH, Harris DJ, Prance GT, Davidse G, Paige J, Pennington TD, 711 712 Robson NKB, Scotland RW. 2010. Herbaria are a major frontier for species discovery. Proceedings of the 713 National Academy of Sciences of the United States of America 107:22169-22171. 714 Berenbaum MR and Zan. 1998. Chemical phenotype matching between a plant and its insect herbivore. 715 Proceedings of the National Academy of Sciences of the United States of America 95: 13743–13748. 716 Besnard G, Christin P-A, Malé P-JG, L'huillier E, Lauzeral C, Coissac E, Vorontsova MS. 2014. From 717 museums to genomics: old herbarium specimens shed light on a C3 to C4 transition. Journal of Experimental 718 Botany DOI 10.1093/jxb/eru395 Bi K, Linderoth T, Vanderpool D, Good JM, Nielsen R, Moritz C. 2013. Unlocking the vault: next-generation 719 720 museum population genomics. *Molecular ecology* **22:**6018-6032.



721 Bolotov IN, Makhrov AA, Yu M, Gofarov, Aksenova OV, Aspholm PE, Bespalaya YV, Kabakov MB, 722 Kolosova YS, Kondakov AV, Ofenböck T, Ostrovsky AN, Yu Popov I, von Proschwitz T, Rudzīte M, 723 Rudzītis M, Sokolova SE, Valovirta I, Vikhrev IV, Vinarski MV & AA Zotin. 2018. Climate Warming as a Possible Trigger of Keystone Mussel Population Decline in Oligotrophic Rivers at the Continental Scale. 724 725 Scientific Reports 8:35 DOI:10.1038/s41598-017-18873-y Brower AVZ. 2006. Problems with DNA barcodes for species delimitation: "Ten species" of Astraptes fulgerator 726 reassessed (Lepidoptera: Hesperiidae). Systematics and Biodiversity 4:127-132 727 728 Billick, I., I. Babb, B. Kloeppel, J. C. Leong, J. Hodder, J. Sanders, and H. Swain. 2013. Field Stations and 729 Marine Laboratories of the Future: A Strategic Vision. National Association of Marine Laboratories and 730 Organization of Biological Field Stations. Available at http://www.obfs.org/fsml-future Colegate SM, Welsh SL, Gardner DR, Betz JM, and KE Panter. 2014. Profiling of Dehydropyrrolizidine 731 732 Alkaloids and their N-Oxides in Herbarium-Preserved Specimens of Amsinckia Species Using HPLCesi(+)MS. J Agric Food Chem. 62(30):7382–7392. 733 Cook JA, Edwards SV, Lacey EA, Guralnick RP, Soltis PS, Soltis DE, Welch CK, Bell KC, Galbreath KE, 734 735 Himes C, Allen JM, Heath TA, Carnaval AC, Cooper KL, Liu M, Hanken J, Ickert-Bond S. 2014. Natural 736 History Collections as Emerging Resources for Innovative Education. *BioScience* **64:**725–734. 737 Cook, J.A., Galbreath, K.E., Bell, K.C., Campbell, M.L., Carrière, S., Colella, J.P., Dawson, N.G., Dunnum, J.L., Eckerlin, R.P., Greiman, S.E., Fedorov, V., Hass, G.M.S., Haukisalmi, V., Henttonen, H., Hope, 738 739 A.G., Jackson, D., Jung, T., Koehler, A.V., Kinsella, M., Krejsa, D., Kutz, S.J., Liphardt, S., MacDonald, 740 S.O., Malaney, J.L., Makarikov, A., Martin, J., McLean, B.S., Mulders, R., Nyamsuren, B., Talbot, S.L., Tkach, V.V., Tsyetkova, A., Toman, H.M., Waltari, E.C., Whitman, J.L., Hoberg, E.P. 2017. The 741 742 Beringian Coevolution Project: Holistic Collections of Mammals and Associated Parasites Reveal Novel 743 Perspectives on Evolutionary and Environmental Change in the North. Arctic Science 3:585-617. 744 Cook JA, Greiman S, Agosta RP, Anderson BS, Arbogast RJ, Baker W, Boeger RD, Bradley DR, Brooks R, 745 Cole JR, Demboski AP, Dobson JL, Dunnum RP, Eckerlin J, Esselstyn K, Galbreath J, Hawdon H, 746 Hoekstra S, Kutz J, Light L, Olson BD, Patterson JL, Patton AJ, Phillips E, Rickart DS, Rogers M, Siddall V, Tkach and EP Hoberg. 2016. Transformational principles for NEON sampling of mammalian 747 parasites and pathogens: a response to Springer and colleagues. *BioScience* 66:917-919. 748



749	Cook JA, Lacey EA, Ickert-Bond SM, Hoberg EP, Galbreath KE, Bell KC, Greiman SE, McLean BS,
750	Edwards SV. 2016. From Museum Cases to the Classroom: Emerging Opportunities for Specimen-Based
751	Education. Pp 787-799 in Pavlinov I, ed. "Aspects of Biodiversity", Volume 54, Archives of Zoological Museum
752	of Moscow State University, Moscow, Russia.
753	Darwin Initiative, https://www.gov.uk/government/groups/the-darwin-initiative
754	Davies N, Field D, Gavaghan D, Holbrook SJ, Planes S, Troyer M, Bonsall M, Claudet J, Roderick G,
755	Schmitt RJ. 2016. Simulating social-ecological systems: the Island Digital Ecosystem Avatars (IDEA)
756	consortium. GigaScience 5, 1. http://doi.org/10.1186/s13742-016-0118-5
757	Délye C, Deulvot C, Chauvel B. 2013. DNA Analysis of Herbarium Specimens of the Grass Weed Alopecurus
758	myosuroides Reveals Herbicide Resistance Pre-Dated Herbicides. PLoS ONE 8(10), e75117
759	Droege G, Barker K, Seberg O, Coddington J, Benson E, Berendsohn WG, Bunk B, Butler C, Cawsey EM,
760	Deck J, Döring M, Flemons P, Gemeinholzer B, Güntsch A, Hollowell T, Kelbert P, Kostadinov I,
761	Kottmann R, Lawlor RT, Lyal C, Mackenzie-Dodds J, Meyer C, Mulcahy D, Nussbeck SY, Ó Tuama É,
762	Orrell T, Petersen G, Robertson T, Söhngen C, Whitacre J, Wieczorek J, Yilmaz P, Zetzsche H, Zhang Y
763	Zhou X. 2016. The Global Genome Biodiversity Network (GGBN) Data Standard specification. Database.
764	baw125. DOI: -10.1093/database/baw125
765	Dunnum JL, Yanagihara R, Johnson KM, Armien B, Batsaikhan N, Morgan L, Cook JA. 2017. Biospecimen
766	repositories and integrated databases as critical infrastructure for pathogen discovery and pathobiology
767	research. PloS Neglected Tropical Diseases11: e0005133
768	Foley, 2015. the_Macroscope.org
769	Gitzendanner MA, PS Soltis, GK-S Wong, BR Ruhfel, and DE Soltis. 2018. Plastid phylogenomic analysis of
770	green plants: A billion years of evolutionary history. American Journal of Botany 0(0): 1–11.
771	doi:10.1002/ajb2.1048
772	Gokhman D, Lavi E, Prüfer K, Fraga MF, Riancho JA, Kelso J, Pääbo S, Meshorer E, Carmel L. 2014.
773	Reconstructing the DNA Methylation Maps of the Neandertal and the Denisovan. Science 344: pp. 523-527
774	doi: 10.1126/science.1250368



775 Graham CH, Ferrier S, Huettman F, Moritz C, Peterson AT. 2004. New developments in museum-based 776 informatics and applications in biodiversity analysis. Trends in ecology & evolution, 19(9), pp.497-503. 777 Grayson P, Sin SYW, Sackton TB, Edwards SV. 2017. Comparative genomics as a foundation for evo-devo 778 studies in birds, Pages 11-46 in G. Shen, ed. Methods in Molecular Biology: Avian and Reptilian Developmental Biology. New York, Humana Press. 779 Gruenewald, D.A. and Smith, G.A. eds. 2014. Place-based education in the global age: Local diversity. Taylor and 780 781 Francis Press, New York. Greiman, S., J. A. Cook, V. V. Tkach, E. P Hoberg, D. Menning, A. G. Hope, S. A Sonsthagen, and S. L. 782 783 Talbot. 2018. Museum metabarcoding: a novel method revealing gut helminth communities of small mammals across space and time. International Journal of Parasitology. 48:1061-1070. 784 785 Hart ML, Forrest LL, Nicholls JA & CA Kidner. 2016. Retrieval of hundreds of nuclear loci from herbarium 786 specimens. Taxon 65(5): 1081-1092. Hebert PDN, Penton EH, Burns JM, Janzen DH, Hallwachs W. 2004. Ten species in one: DNA barcoding 787 788 reveals cryptic species in the neotropical skipper butterfly Astraptes fulgerator. PNAS 101: 14812-14817. Hebert PDN, deWaard JR, Landry J-F. 2010. DNA barcodes for 1/1000 of the animal kingdom. Biol. Lett. 789 790 (2010) 6, 359–362 doi:10.1098/rsbl.2009.0848 791 Hess CM, Wang Z, and SV Edwards. 2007. Evolutionary genetics of Carpodacus mexicanus, a recently colonized 792 host of a bacterial pathogen, Mycoplasma gallisepticum. Genetica 129:217-225. 793 Hofreiter M, Paijmans JLA, Goodchild H, Speller CF, Barlow A, Fortes GG, Thomas JA, Ludwig A, Collins MJ. 2015. The future of ancient DNA: Technical advances and conceptual shifts. *BioEssays* 37: 284–293, doi: 794 795 10.1002/bies.201400160 796 Holmes MW, Hammond TT, Wogan GOU, Walsh RE, LaBarbera K, Wommack EA, Martins FM, Crawford 797 JC, Mack KL, Bloch LM, MW Nachman. 2018. Natural history collections as windows on evolutionary 798 processes. Molecular Ecology https://doi.org/10.1111/mec.13529 Holos, Ecoengine, UC Berkeley, https://holos.berkeley.edu/about/ 799



800 Hudson LN, Blagoderov V, Heaton A, Holtzhausen P, Livermore L, Price BW, van der Walt S, Smith VS. 801 **2015.** Inselect: Automating the Digitization of Natural History Collections. 802 https://doi.org/10.1371/journal.pone.0143402 803 **IBOL** (International Barcode of Life), 2014. What is iBOL? Available at: http://ibol.org/about-us/what-is-ibol/ 804 Ingala MR, Simmons NB, Wultsch C, Krampis K, Speer KA, Perkins SL. 2018. Comparing Microbiome 805 Sampling Methods in a Wild Mammal: Fecal and Intestinal Samples Record Different Signals of Host Ecology, 806 Evolution. Front. Microbiol., 01 May 2018 | https://doi.org/10.3389/fmicb.2018.00803 James SA, Soltis PS, Belbin L, Chapman AD, Nelson G, Paul DL, Collins M. 2018. Herbarium data: Global 807 808 biodiversity and societal botanical needs for novel research: Global. Applications in Plant Sciences. John Wiley 809 and Sons Inc. https://doi.org/10.1002/aps3.1024 Janzen DH & Hallwachs W. 2016. DNA barcoding the Lepidoptera inventory of a large complex tropical 810 811 conserved wildland, Area de Conservacion Guanacaste, northwestern Costa Rica. Genome 59(9):641-60. doi: 10.1139/gen-2016-0005. 812 813 Javaram K & Full RJ. 2016. Cockroaches traverse crevices, crawl rapidly in confined spaces, and inspire a soft, legged robot. PNAS 113, 23 February 2016, Pages E950-E957 814 815 Johnson KW. 2018. The Ornithologist the Internet Called a Murderer. The New York Times, June 15, 2018. 816 Jones CA, Daeler CC. 2018. Herbarium specimens can reveal impacts of climate change on plant phenology; a 817 review of methods and applications. PeerJ 6:e4576 https://doi.org/10.7717/peerj.4576 Kissling WD, Ahumada JA, Bowser A, Fernandez M, Fernández N, García EA, Guralnick RP, Isaac NJB, 818 Kelling S, Los W, McRae L, Mihoub JB, Obst M, Santamaria M, Skidmore AK, Williams KJ, Agosti D, 819 820 Amariles D, Arvanitidis C, Bastin L, De Leo F, Egloff W, Elith J, Hobern D, Martin D, Pereira HM, 821 Pesole G, Peterseil J, Saarenmaa H, Schigel D, Schmeller DS, Segata N, Turak E, Uhlir PF, Wee B, AR 822 Hardisty. 2017. Building essential biodiversity variables (EBVs) of species distribution and abundance at a 823 global scale. *Biological Reviews* https://doi.org/10.1111/brv.12359 Kõljalg U, Nilsson RH, Abarenkov K, Tedersoo L, Taylor AFS, Bahram M, Bates SB, Bruns TD, Bengtsson-824 825 Palme J, Callaghan TM, Douglas B, Drenkhan T, Eberhardt U, Dueñas M, Griffith TGW, Hartmann M, Kirk PM, Kohout P, Larsson E, Lindahl BD, Lücking R, Martín RMP, Matheny PB, Nguen NH, 826



827	Niskanen 1, Oja J, Peay KG, Peintner U, Peterson M, Polumaa K, Saag L, Saar 1, Schussier A, Scott JA,
828	Senés C, Smith ME, Suija A, Taylor DL, Telleria T, Weiss M, Larsson K-H, 2013. Towards a unified
829	paradigm for sequence-based identification of fungi. Molecular Ecology 22: 5271–5277.
830	Lacey EA, Hammond TT, Walsh RE, Bell KC, Edwards SV, Ellwood ER, Guralnick R, Ickert-Bond SM,
831	Mast AR, McCormack JE, Monfils AK, Soltis PS, Soltis DE, Cook JA. 2017. Climate change, collections
832	and the classroom: using big data to tackle big problems. Evo Edu Outreach 10:2; doi 10.1186/s12052-017-
833	0065-3
834	Llamas B, Holland ML, Chen K, Cropley JE, Cooper A, Suter CM. 2012. High-Resolution Analysis of Cytosine
835	Methylation in Ancient DNA. PLoS ONE 7, e30226 34.
836	Liede-Schumann S, Khanum R, Mumtaz AS, Gherghel I & Pahlevani A. 2016. Going west – A subtropical
837	lineage (Vincetoxicum, Apocynaceae: Asclepiadoideae) expanding into Europe. Molecular Phylogenetics and
838	Evolution 94: 436–446.
839	Lindqvist C, Rajora OP (eds.). 2019. Paleogenomics, Genome-Scale Analysis of Ancient DNA. Springer.
840	Maldonado C, Barnes CJ, Cornett C, Holmfred E, Hansen SH, Persson C, Antonelli A, Rønsted N. 2016.
841	Phylogeny Predicts the Quantity of Antimalarial Alkaloids Within the Iconic Yellow Cinchona Bark
842	(Rubiaceae: Cinchona calisaya). Frontiers in Plant Science 8:391.
843	Matos-Maraví P, Duarte Ritter C, Barnes CJ, Nielsen M, Olsson U, Wahlberg N, Marquina D, Sääksjärvi I,
844	Antonelli A. 2019. Biodiversity seen through the perspective of insects: 10 simple rules on methodological
845	choices and experimental design for genomic studies. PeerJ Preprints 7:e26661v3; doi
846	10.7287/peerj.preprints.26661v3
847	McGuire JA, Cotoras DD, O'Connell B, Lawalata SZS, Wang-Claypool CY, Stubbs A, Huang X, Wogan
848	GOU, Hykin SM, Reilly SB, et al. 2018. Squeezing water from a stone: high-throughput sequencing from a
849	145-year old holotype resolves (barely) a cryptic species problem in flying lizards. PeerJ 6, e4470.
850	(DOI:10.7717/peerj.4470).
851	Meineke EK, Davies TJ, Daru BH, Davis CC. 2019a. Biological collections for understanding biodiversity in the
852	Anthropocene. Philosophical Transactions of the Royal Society B, 2019; 374: 20170386. doi:
853	10.1098/rstb.2017.0386.



854 Meineke EK & Davies TJ. 2019b. Museum specimens provide novel insights into changing plant-herbivore 855 interactions. Philosophical Transactions of the Royal Society B, 2019; 374: 20170393. doi: 856 10.1098/rstb.2017.0393. Michener WK, Bildstein KL, McKee A, Parmenter RR, Hargrove WW, McClearn D, Stromberg M. 2009. 857 858 Biological field stations: research legacies and sites for serendipity. *BioScience* **59**, 300-310. http://dx.doi.org/10.1525/bio.2009.59.4.8 859 Mitchell BJ, Gillespie RG. 2007. Graduate Students Take to the Field in K-12 Education. *PLoSBiology* 5(6): e162. 860 861 doi:10.1371/journal.pbio.0050162 862 Monardo B. 2019. Innovation districts as turbines of smart strategy policies in US and EU. Boston and barcelona 863 experience. In Smart Innovation, Systems and Technologies, 100, pp. 322-335 doi: 10.1007/978-3-319-92099-3 38 864 Morrison SA, Sillett TS, Funk WC, Ghalambor CK, Rick TC. 2017. Equipping the 22nd-Century Historical 865 Ecologist. Trends in Ecology & Evolution 32, 578-588. 866 National Research Council. 2014. Enhancing the Value and Sustainability of Field Stations and Marine 867 868 Laboratories in the 21st Century The National Academies Press, Washington, DC. 869 http://doi.org/10.17226/18806 870 Nirody JA, Jinn J, Libby T, Lee TJ, Jusufi A, Hu DL, Full RJ. 2018. Geckos Race Across the Water's Surface 871 UsingMultiple Mechanisms. Current Biology28, 4046–4051 doi.org/10.1016/j.cub.2018.10.064 Obst M, Vicario S, Lundin K, Berggren M, Karlsson A, Haines R, Williams A, Goble C, Mathew C, Güntsch 872 873 A. 2017. Marine long-term biodiversity assessment suggests loss of rare species in the Skagerrak and Kattegat region. Marine Biodiversity, doi: 10.1007/s12526-071-0749-5 874 Page LM, MacFadden BJ, Fortes JA, Soltis PS, Riccardi G. 2015. Digitization of biodiversity collections reveal 875 876 biggest data on biodiversity. Bioscience 65(9): 841–842. 877 Pauli JN, Newsome SD, Cook JA, Harrod C, Steffan SA, Baker C, Ben-David M, Bloom D, Brown G, Cerling T, Cicero C, Cook C, Dohm M, Eherlinger J, Federhen S, Frey B, Ghosh P, Graves G, Gropp R, Hobson 878 879 K, Jordan C, O'Connell T, Pilaar Birch S, Poelen J, Ratnasingham S, Russell L, Stricker C, Uhen M, 880 Yarnes C, Hayden B. 2017. Opinion: Why we need a centralized repository for isotopic data. *Proceedings* National Academy Sciences, USA 114: 2997-3001. 881



882 Phillips SJ, Dudik M. 2008. Modeling of species distributions with Max-Ent: new extensions and a comprehensive 883 evaluation. *Ecography* **31:**161–175 Phillips T. 2010. Sao Paulo fire destroys one of the largest collections of dead snakes. The Guardian 16 May 2010. 884 Phillips T. 2018. Brazil National Museum: as much as 90% of collection destroyed in fire. The Guardian 4 885 886 September 2018. Portugal SJ, Cooper HJ, Zampronio CG, Wallace LL, Cassey P. 2010. Can museum egg specimens be used for 887 888 proteomic analyses? Proteome Science 8:40 889 Post RJ, Flook PK, Millest AL. 1993. Methods for the preservation of insects for DNA studies. Biochemical 890 Systematics and Ecology 21:85–92, https://doi.org/10.1016/0305-1978(93)90012-g 891 von Proschwitz T. 1997. Arion lusitanicus MABILLE and A. rufus (L.) in Sweden: A comparison of occurrence, 892 spread and naturalization of two alien slug species. Heldia 4 Sonderheft 5: 137-138. 893 Raffauf RF, von Reis Altschul S. 1968. The Detection of Alkaloids in Herbarium Material. Economic Botany 22:267-269 894 Ramos-Madrigal J, Smith BD, Moreno-Mayar JV, Gopalakrishnan S, Ross-Ibarra J, Gilbert MTP, Wales N. 895 896 2016. Genome Sequence of a 5,310-Year-Old Maize Cob Provides Insights into the Early Stages of Maize 897 Domestication. Current Biology 26: 3195-3201, https://doi.org/10.1016/j.cub.2016.09.036 898 Ratnasingham & Hebert. 2007. BOLD: The Barcode of Life Data System (www.barcodinglife.org). Molecular 899 Ecology Notes, doi: 10.1111/j.1471-8286.2006.01678. Ricklefs RE. 2006. Global variation in the diversification rate of passerine birds. Ecology 87(10):2468-78 900 901 Roggenbuck-Wedemayer M, Bærholm Schnell I, Blom N, Bælum J, Bertelsen MF, Sicheritz-Pontén T, Johannes Sørensen S, Gilbert MTP, Graves GR, Hansen LH. 2014. The microbiome of New World 902 vultures, Nature Communications 5 5498 (2014), doi:10.1038/ncomms6498. 903 904 Rouhan, Germinal, et al. 2016. "The herbonauts website: Recruiting the general public to acquire the data from 905 herbarium labels." Botanists of the Twenty-First Century: Roles, Challenges and Opportunities. United Nations Educational, Scientific and Cultural Organisation: 143-148. 906 907 Rowe KC, Singhal S, Macmanes MD, Ayroles JF, Morelli TL, Rubidge EM, Bi KE, Moritz CC. 2011. 908 Museum genomics: low-cost and high-accuracy genetic data from historical specimens. Molecular Ecology 909 Resources, 11(6), pp.1082-1092.



910	Ruane S, Christopher A, C. 2017. Phylogenomics using formalin-fixed and 100+ year-old intractable natural
911	history specimens. <i>Molecular Ecology Resources</i> 17, 1003-1008. doi:10.1111/1755-0998.12655.
912	Schindel DE, Cook JA. 2018. The next generation of natural history collections. PLoS Biol 16(7): e2006125.
913	https://doi.org/10.1371/journal.pbio.2006125
914	Schmitt CJ, Cook J, Zamudio K, Edwards SV. 2018. Museum specimens of terrestrial vertebrates are sensitive
915	indicators of environmental change in the Anthropocene. Phil. Trans. R. Soc. Lond. B. in review.
916	Sebastian P, Schaefer H, Telford IRH, Renner SS. 2010. Cucumber (Cucumis sativus) and melon (C. melo) have
917	numerous wild relatives in Asia and Australia, and the sister species of melon is from Australia. PNAS 2010
918	107: 14269–14273
919	Shultz AJ, Baker AJ, Hill GE, Nolan PM, Edwards SV. 2016. SNPs across time and space: population genomic
920	signatures of founder events and epizootics in the House Finch (Haemorhous mexicanus). Ecology and
921	Evolution 6 , 7475–7489.
922	Silvertown J, Harvey M, Greenwood R, Dodd M, Rosewell J, Rebelo T, Ansine J, McConway K. 2015.
923	Crowdsourcing the identification of organisms: A case-study of iSpot. ZooKeys 480: 125-146. doi:
924	10.3897/zookeys.480.8803
925	Silvestro D, Salamin N, Schnitzler J. 2014. PyRate: a new program to estimate speciation and extinction rates
926	from incomplete fossil data. Meth. Ecol. Evol. 5:1126-1131 doi: 10.1111/2041-210X.12263
927	Slingsby JA, Merow C, Aiello-Lammens M, Allsopp N, Hall S, Kilroy Mollmann H, Turner R, Wilson AM
928	and JA Silander Jr. 2017. Intensifying postfire weather and biological invasion drive species loss in a
929	Mediterranean-type biodiversity hotspot. PNAS vol. 114(18): 4697–4702
930	Slon V, Mafessoni F, Vernot B, de Filippo C, Grote S, Viola B, Hajdinjak M, Peyrégne S, Nagel S, Brown S,
931	Douka K, Higham T, Kozlikin MB, Shunkov MV, Derevianko AP, Kelso J, Meyer M, Prüfer K, Pääbo S.
932	2018. The genome of the offspring of a Neanderthal mother and a Denisovan father. Nature
933	https://doi.org/10.1038/s41586-018-0455-x
934	Smith CI, Chamberlain AT, Riley MS, Stringer C, Collins M. 2003. The thermal history of human fossils and
935	the likelihood of successful DNA amplification. Journal of Human Evolution 45, 203-217
936	Smith, O. et al. 2014. Genomic methylation patterns in archaeological barley show de-methylation as a time-
937	dependent diagenetic process. Sci. Rep. 4, 5559



938	Smith, R.W. et al. 2015. Detection of cytosine methylation in ancient DNA from five Native American populations
939	using bisulfite sequencing. PLoS ONE 10, e0125344 35.
940	Staats M, Cuence A, Richardson JE, Vrielink-van Ginkel R, Petersen G, Seberg O, Bakker FT. 2011. DNA
941	damage in plant herbarium tissue. PLoS ONE 6: e28448
942	Staats M, Erkens RHJ, van de Vossenberg B, Wieringa JJ, Kraaijeveld K, Stielow B, Geml J, Richardson JE,
943	Bakker FT. 2013. Genomic Treasure Troves: Complete Genome Sequencing of Herbarium and Insect Museum
944	Specimens. PLoS ONE 8(7): e69189. doi:10.1371/journal.pone.0069189
945	Suarez AV, Tsutsui ND. 2004. The Value of Museum Collections for Research and Society. Bioscience 54:66-74
946	Troudet J, Vignes-Lebbe R, Grandcolas Ph, Legendre F. 2018. The Increasing Disconnection of Primary
947	Biodiversity Data from Specimens: How Does It Happen and How to Handle It?, Systematic Biology, 67, Issue
948	6, 1 November, Pages 1110–1119, https://doi.org/10.1093/sysbio/syy044
949	Veijalainen A, Wahlberg N, Broad GR, Erwin TL, Longino JT, Sääksjärvi, IE. 2012. Unprecedented
950	ichneumonid parasitoid wasp diversity in tropical forests. https://doi.org/10.1098/rspb.2012.1664
951	Webster MS. 2017. The Extended Specimen: Emerging Frontiers in Collections-Based Ornithological Research.
952	Boca Raton, FL, USA: CRC Press.
953	Welker F, Soressi M, Rendu W, Hublin J-J, Collins M. 2015. Using ZooMS to identify fragmentary bone from
954	the Late Middle/Early Upper Palaeolithic sequence of Les Cottés, France. Journal of Archaeological Science
955	54, pp. 279-286
956	Wen J, Egan AN, Dikow RB, Zimmer EA. 2015. Utility of transcriptome sequencing for phylogenetic inference
957	and character evolution. in: Hörandl, E. & Appelhans, M.S. (eds.), Next-Generation Sequencing in Plant
958	Systematics. Chapter 2, http://dx.doi.org/10.14630/000003© International Association for Plant Taxonomy
959	(IAPT)
960	Wilson EO. 2003. The encyclopedia of life. Trends in Ecology and Evolution 18: 77-80
961	Yanagihara R, Gu SH, Song J-W. 2015. Expanded host diversity and global distribution of hantaviruses:
962	Implications for identifying and investigating previously unrecognized hantaviral diseases. In: Shapshak P,
963	Sinnott JT, Somboonwit C, Kuhn J, eds. Global Virology. Identifying and Investigating Viral Diseases. New
964	York: Springer-Verlag. 161±198



965	Yates TL. et al. 2002. The ecology and evolutionary history of an emergent disease: Hantavirus pulmonary
966	syndrome. Bioscience, 52: 989-998.
967	Yoshida K, Burbano HA, Krause J, Thines M, Weigel D et al. 2014. Mining Herbaria for Plant Pathogen
968	Genomes: Back to the Future. PLoS Pathog 10(4): e1004028, doi:10.1371/journal.ppat.1004028
969	Yoshida K, Sasaki E, Kamoun S. 2015. Computational analyses of ancient pathogen DNA from herbarium
970	samples: Challenges and prospects. Frontiers in Plant Science 6, Article number 771, 6p
971	Zeyl E, Marcotegui P, Harris PD, Bakke TA, Bachmann L. 2012. Nye parasitter fra gamle fisk / New parasites
972	from old fish. Naturen 136, 274-280
973	Zhang Q, Hill GE, Edwards SV, Backström N. 2014. A house finch (Haemorhous mexicanus) spleen transcriptome
974	reveals intra- and interspecific patterns of gene expression, alternative splicing and genetic diversity in passerines
975	BMC Genomics 15:305.



Table 1(on next page)

Main global and local aggregators of biodiversity data.



 Table 1. Main global and local aggregators of biodiversity data.

				Volume of
Acronym	Mission	Funding; scope	Type of data	records (M)
ADBC	Advancing Digitization of Biodiversity Collections	US		
ALA	Atlas of Living Australia. https://www.ala.org.au/	Australia	Observations, specimens	84.8
BOLD	Barcode of Life Database	Canada; global		6.8
DiSSCo	Distributed System of Scientific Collections; digitization and databasing of european specimen collections	Europe	Specimens	1500
eBird	Citizen science: the world largest biodiversity-related citizen science project, gathering information on bird sightings, archive it, and "freely share it to power new data-driven approaches to science, conservation and education." https://ebird.org/home	Global	Observations	100 'yearly'
EOL	Encyclopedia of Life; Global access to knowledge about life on Earth	Australia, Egypt, US; global	Species descriptions	<1,9
GBIF	Global Biodiversity Information Facility	Global	Observations, specimens	1300



GGBN	Global Genome Biodiversity Network	Global	DNA, tissues,	
			environmental samples	3.8
			environmental samples	
GloBI	Global Biotic Interactions; species interaction data by combining existing open	US; global	Species interaction data	
	datasets. https://www.globalbioticinteractions.org/		e.g., predator-prey,	. 0.7
			pollinator-plant, pathogen-	>0.7
			host, parasite-host	
			nost, parasite nost	
HOLOS	Berkeley Ecoinformatics Engine: accessing and visualizing integrated	US; global	Different kinds of	
	biological and environmental datasets to address questions of global change		biological and	n.a.
	biology. https://holos.berkeley.edu/		environmental datasets	
iBOL	International Barcode of Life; extending BOLD's coverage. iBOL's	Canada; global	DNA barcodes and	
	forthcoming BIOSCAN will activate a biomonitoring system for half the		metadata	
	world's ecoregions, metabarcoding assemblages and studying species			see BOLD
	interactions from 2,500 sites. iBOL.org			
	moraciono from 2,000 sitos. IDOL.OIS			
IDEA	Island Digital Ecosystem Avatar; place-based systems ecology for building	US; Moorea	Specimens, observations	0
	simulations of social-ecological systems			?



iDigBio	Integrated Digitized Biocollections; digitisation and databasing of US specimen collections	US	Specimens	117.5
iNaturalist	Citizen science: one of the world's most popular nature apps, sharing observations globally; https://www.inaturalist.org/	US; global	Observations	<1
LifeWatch	Biodiversity research, -management and -conservation priority setting	Europe	Research tools	n.a.
iSpot	Citizen science: experts helping citizen community to identify its wildlife observations. https://www.ispotnature.org/	UK; global	Species identifications	0.030
MoL	Map of Life; providing species range and dynamics information and species lists for any geographic area. https://mol.org/	Global	Occurrences, observations	8.8
NCBI	National Center for Biotechnology Information	Global	Nucleotide and amino acid sequences; genome annotations	0.37 species covered
NEON	National Ecological Observatory Network; continental-scale environmental data, infrastructure for research, educational tools to work with large data. https://www.neonscience.org/	US	Observations	?



OToL	Open Tree of Life; construct a comprehensive, dynamic and digitally-available	US; global	Phylogenetic trees and	2.6
	tree of life by synthesizing published phylogenetic trees along with taxonomic data. https://tree.opentreeoflife.org/		taxonomies	OTU's in taxonomy
Traitbase	Ecological species characteristics, individual level species information. https://traitbase.info/whatis	Spain; global	Specific characteristics e.g. body size, diet or fecundity	?



Table 2(on next page)

Specimens and best curation practices for the future.



- 1 **Table 2.** Specimens and best curation practices for the future.
- 2 Collecting. Recommendations for best preservation techniques for new specimens during field
- 3 collection are as important as the final storage conditions for improving specimen long-term
- 4 utility for genomics (*Matos-Maraví et al., 2019*). Documenting treatment practices is also key to
- 5 facilitating future analyses enabled by as yet undiscovered technologies. The plethora of
- 6 technological uses of museum specimens calls for a re-evaluation of how specimens are
- 7 preserved. For centuries, plants have been pressed, animals mounted, marine specimens ethanol-
- 8 or formalin-fixed and fungi dried. Although these standard preservation methods should still
- 9 continue, if only because they constitute the bulk of biological collections thus far and have a
- proven track record of fostering discovery, whenever possible researchers should try to sample
- additional types of specimen parts, and organs and meta-data.
- 12 Storing. More studies need to be undertaken to improve relevant protocols for the long-term
- storage of specimens and tissues. Like digitization, banking of genetic resources by museums is
- an area of rapid innovation, particularly as next-generation sequencing methods have become
- more common. As museum tissue collections are accessed more frequently for genome projects,
- it has also become clear that the preservation standards and types of tissues preserved in
- museums are often inadequate for supporting the genomics enterprise. For example, a typical
- museum tissue sample from a bird, even if frozen in nitrogen hours after sacrifice in the field,
- 19 yields DNA qualities and lengths unsuitable for 3rd generation long-read sequencing platforms
- such as PacBio and Oxford Nanopore. Such technologies rely on the use of long DNA fragments
- 21 to start with, requiring specimen tissues be frozen immediately (within > 10-15 minutes) upon
- collection. Although it may be difficult to use liquid nitrogen in the field, one solution is to use
- 23 so called Dry Shippers, which are dewars designed for safe transportation of tissues at the same



temperature as liquid nitrogen but without actually containing any free liquid nitrogen. Such shippers are routinely allowed for transportation back to the lab by airlines and can often hold 25 cold temperatures for ~3 weeks. Innovations in cryogenics are likely to transform collecting of 26 genomic resources by museums in the future. Still, we can take comfort that even from dried, 27 centuries-old traditional specimens, valuable genetic information can readily be obtained by 28 29 hybrid-capture and other approaches (Bi et al., 2013; Staats et al., 2013). At the same time, some genome sequencing strategies may alleviate the use of unwieldy 30 cryogenic methods altogether, at least for accessing DNA. What seems to be most important is 31 that DNA (and RNA) degradation is stopped as fast as possible after collection. For example, for 32 birds, one way to achieve long DNA fragments for next-generation sequencing is to use unfrozen 33 blood stored in Queen's lysis buffer, which has been used by ornithologists for decades and takes 34 advantage of birds' nucleated red blood cells. Blood stored in this way, with minimal shaking 35 that will cause shearing, is a reliable source of high molecular weight DNA and has been shown 36 37 to yield better genome assemblies than DNA retrieved from museum-grade frozen tissues (S. Edwards, G. Bravo, pers. obs). An alternative could be to store collections in the field in DMSO, 38 although this appears to prevent RNA sequencing (Irestedt, unp. data). Such best-practices for 39 fieldwork and storage of genetic resources needs to be shared more widely and rapidly among 40 the museum community. A useful platform for identifying both repositories and tissues for a 41 wide range of taxa, often called biobanks, is provided by the Global Genomic Biodiversity 42 Network mentioned above (GGBN see http://www.ggbn.org/ggbn_portal/). Below we outline 43 various updates in storage of genetic resources for both animals and plants, highlighting issues 44 facing museum curators and collection managers looking at the future: 45



Transcriptomics and Epigenomics. The ever-increasing number of genome sequences becoming 46 available can be efficiently explored in terms of gene function through transcriptomics - the 47 sequencing of all transcribed mRNA expressed at a certain time, physiological or developmental 48 state for a particular tissue. In this way, the 1Kite (http://www.1kite.org/) and 1kp 49 (https://sites.google.com/a/ualberta.ca/onekp/) projects, assembling 1000 transcriptomes of 50 51 insects and plants, respectively, have expedited progress in both comparative and functional genomics and a better understanding of gene function across these clades (see for instance 52 Gitzendanner & al., 2018). We would expect future specimens to play an increasing role in this 53 respect, but only if we make sure to store our specimens in such a way that RNA is preserved, 54 for instance by rapid cryogenic storage of use of RNA-friendly buffers like RNA-later. 55 Additionally, a diversity of epigenomics approaches, such as methylation, Chip-seq and ATAC-56 seq, are emerging and potentially of great use to the field of evolutionary biology (Grayson et 57 al., 2017). Epigenomics is already commonly applied in evolutionary studies of adaptation and 58 development, and has recently made headway in examining epigenetic maps of extinct human 59 and plant populations (Llamas et al., 2012; Gokhman et al., 2014; Smith et al., 2014; 2015). Best 60 practices for preservation of biomaterials for epigenetics has not yet been discussed in the 61 literature, and will be an important additional consideration for museum curators in the future. 62 *Proteomics.* Given future technological developments, it is likely that proteomes will be 63 determined and used for further functional studies across the Tree of Life. Additionally, collagen 64 from bone tissues have been demonstrated to give species-level amino acid variation from 65 specimens several millions of years old using a ZooMS approach (Welker et al., 2015). Portugal 66 et al. (2010) report on proteomics in museum egg specimens and conclude that current 67 approaches to proteomics in such specimens may be limited in coverage of the proteome. In any 68



case, storing tissues in the best possible ways for proteomics, ideally, cryogenically in order to 69 stop proteases from working, now ensures that such analyses can be conducted in the near future. 70 71 Secondary metabolites. Compounds such as alkaloids, glucosinolates, furanocoumarins, flavonoids or terpenes can be measured in museums tissues, especially from plants (Berenbaum 72 & Zan 1998; Colegate & al., 2014; Mithen et al., 2010; Raffauf & Von Reis Altschul, 1968). 73 74 Access to such metabolites enables testing historic biological hypotheses such as past responses to change in herbivores and climate; but also in case of invasive species and testing what 75 secondary compounds may have driven invasive success in species such as Vincetoxicum nigrum 76 (Asclepiadaceae) (Liede-Schuman & al., 2016). 77 Stable isotopes. Advancement of techniques for studying specimens include measuring of stable 78 isotopes, allowing monitoring environmental/atmospheric changes over time, given a time series 79 of museum specimens (reviewed in Schmitt et al., 2018). Because elements are not expected to 80 degrade over time like biopolymers do, proper specimen storage conditions for isotope analysis 81 is probably not critical. Limiting factors in such studies now is the availability of robust spatial 82 sampling and time series of specimens for analysis. Properly tracking the vast quantities of data 83 that are generated in these analyses directly to the specimens is also a challenge (Pauli et al., 84 2017). 85 Non-standard tissues. Classical botanical specimens comprise branches with leaves and fertile 86 organs (flowers, fruits). For some vertebrates, such as birds and mammals, it is primarily the 87 external morphology that is preserved in collections, whereas for amphibians, reptiles and fish, 88 preservation in formalin and/or alcohol can yield DNA sequences with some effort (Ruane et al., 89 2017; McGuire et al., 2018). Many biobanks, particularly in US museums, now also include 90

samples of frozen blood and tissue from vertebrates, typically heart, liver and muscle. However,



many other parts of organisms not conventionally stored in museums are becoming important in the effort to monitor global change. For instance, there is great interest among diverse scientists in investigating the microbiome of species - including the bacteria present in the digestive system, and what roles they may have to the species' adaptations to the local environments (Roggenbuck et al., 2014; Alvisatos et al., 2015; Ingala et al., 2018). Similarly, tree bark may provide important information on chemical defenses of plants and hold implications for medical applications (Maldonado et al., 2016). Transcriptome studies in vertebrates are becoming increasingly common as a means to study species' ability to adapt to changing environments and anthropogenic change (e.g., Zhang et al., 2014) and are widely used in phylogenomics of invertebrates and plants (Wen et al., 2015). Such studies encourage careful sampling and preservation of whole organisms or all organs separately when appropriate.



Table 3(on next page)

Specimens and Pathogens.



- 1 **Table 3.** Specimens and Pathogens.
- 2 Museum collections have provided fundamental infrastructure for identifying and mitigating
- 3 emerging pathogens and zoonotic diseases by public health agencies (Dunnum et al., 2017) such
- 4 as the Centers for Disease Control (CDC). When a new pathogen emerged in 1993 in the
- 5 southwestern United States, rapidly killing 7 people, authorities needed to know: had this
- 6 pathogen accidently been released into the wild, or was it a newly evolved pathogen, or had the
- virus always been present and simply not previously identified? Without the availability of
- 8 specimen archives, scientists would not have been able to efficiently determine the pathogen
- 9 source and answer these fundamental questions. Large tissue archives from the Museum of
- Southwestern Biology and other museums (*Yates et al., 2002*) enabled virologists to quickly
- identify that this previously unknown zoonotic hantavirus pathogen was hosted by the locally
- common deer mouse (*Peromyscus maniculatus*). This virus is apparently transmitted to humans
- through inhalation of viral infected feces. Subsequent emergence of other human cases elsewhere
- in the Americas (e.g., Chile, Argentina, Panama), but with higher mortality, mobilized other
- specimen-based research efforts that identified other new strains of hantaviruses in many rodent
- species over the next 2 decades and on multiple continents. More recently, museum specimens of
- other groups of mammals were screened, leading to a radically reshaped understanding of
- hantavirus evolution, ecology and host occurrence (Yanagihara et al., 2015). Not only were
- more rodent host species for these viruses identified, but numerous species of shrews, moles, and
- 20 bat species worldwide also harbor their own hantaviruses. These specimens originated from
- 21 multiple continents and the new discoveries significantly expand the potential risk to humanity
- of these pathogens and increase the burden on public health systems worldwide.



24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

Other examples of pathogen outbreaks have examined historical progression of diseases often decades or centuries after the outbreak (Schmitt et al., 2018). The impact of an invasive bacterial pathogen from poultry on native songbirds has been studied using avian tissue samples collected just prior to the outbreak in the eastern US, albeit without any foreknowledge of the impending epizootic (Hess et al., 2007; Shultz et al., 2016). The Norwegian fish fauna is well documented in the scientific collection of the Natural History Museum, University of Oslo and collection material was screened for monogenean ectoparasitic flatworms of the genus Gyrodactylus that were (unintendedly) collected along with the fish (Zeyl et al. 2012). This yielded 13 flatworm species that are new to science, and an additional seven parasite species new to Norway. Three Gyrodactylus species were also recorded from new fish hosts, and in particular G. ptervgialis appeared to have a broad range of host species, helping fishery biologists tremendously in understanding and managing fish populations. From plants, Yoshida et al. (2014; 2015) used potato herbarium in order to determine the genotype of the *Phytophtera infestans* strain that caused the great Irish potato famine in the 19th century (and concluded it was a 'one-off' type, never seen again). Herbarium DNA was also crucial in discovering ancient alleles in the grass Alopecurus myosuroides that are relevant to herbicide resistance but pre-dating human influence (Délye et al., 2013). Studies using genomic data of a 5,310-year-old maize cob (Ramos-Madrigal et al., 2016) have shown that our understanding of the process of domestication and early selection pressures needs adjusting.

42



Table 4(on next page)

Citizen Science.

Citizen Science 'Man-spread molluscs' project: *send in the slugs from your garden to the museum for identification*.



- Table 4. Citizen Science 'Man-spread molluscs' project: send in the slugs from your garden to
- 2 the museum for identification.
- 3 Since 1986 the Gothenburg Natural History Museum (GNM) has offered a slug-identification
- 4 service to the public. The project was initiated as the invasive Spanish slug (*Arion vulgaris*
- 5 Moquin-Tandon) began to spread rapidly over the country, prompting the need to establish a way
- 6 to follow the spread and the colonisation process. As the slug spreads passively, mainly by the
- 7 trade with ornamental plants and also with garden soil, it easily establishes in residential areas,
- 8 where it mass-propagates and causes severe damage to vegetables and plants. A proper
- 9 determination of these species requires dissection, a service offered by GNM. The project was
- advertised on TV, radio, in the newspapers and on museum web pages and GNM even offered to
- send out a transportation box which could easily be returned by mail. The response from the
- public was immense, and up to today GNM has received > 6000 samples with slugs from all
- over the country. Together with the box, GNM sent out a questionnaire asking information about
- 14 first year of occurrence, possible way of introduction etc. After determination of the specimens
- the senders got information of species identity, and in case A. vulgaris was concerned also advice
- for control measurements. Hence the project has been beneficial for both the gardeners and the
- museum. The colonisation process could be followed in detail and much information on the
- biology and behaviour of the species, as well as on the garden fauna of snails and slugs, was
- obtained. The latter included several other invasive species, from different parts of the country,
- and the development of this fauna over more than four decades could be monitored both
- 21 geographically and chronologically. Furthermore, as at least one specimen from each species in
- 22 the samples was preserved in EtOH, extensive material is available for DNA analysis, which has

- proved highly useful as the taxonomy of the Spanish slugs is complicated, involving
- 24 hybridisation with other native and introduced species (von Proschwitz, 1997).



Figure 1(on next page)

The centrality of natural history collections to evolutionary biology and public understanding.

Users, contributors and stakeholders of natural history collections are indicated; yellow arrows represent data flow, green arrows the flow of specimens.

Sample providers

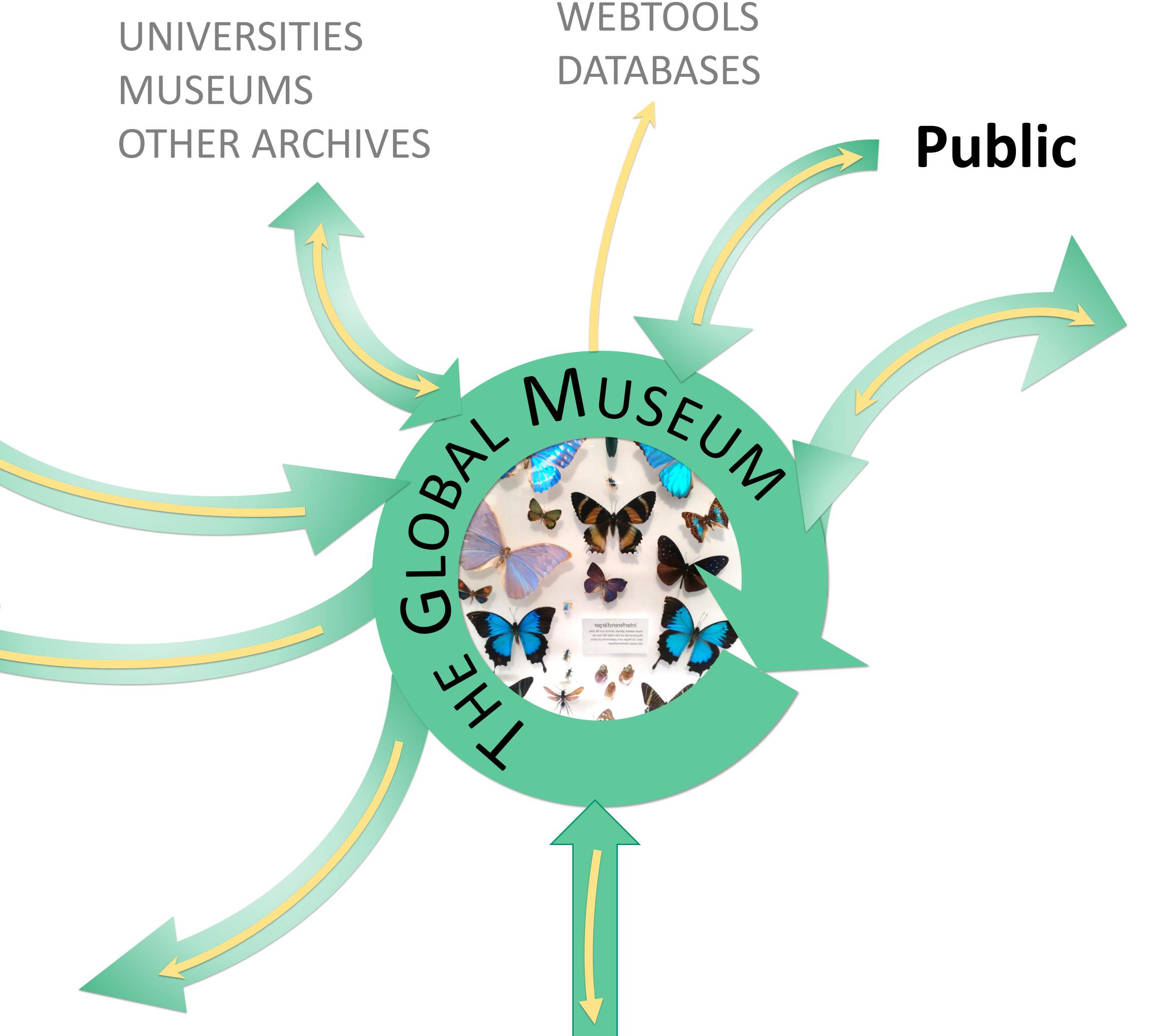
- Biodiversity Inventories
- Pathogen Identification & Monitoring
- Government Wildlife
 Agencies

Policy initiatives

- CITES
- CAFF
- NET-BIOME

Educational outreach

- Exhibits, Tours and Programs
- Curricula and Material for University
 Courses
- Web-accessible Biodiversity Data
- Internships (Field & Museum)



Teaching & Research

- Spatial and Temporal Perspectives
- Environmental Change
- Evolutionary Genomics
- Health and Economic Dimensions
- Systematics & Population Biology

Science Conservation Biology

Population Status & Structure, Abundance, Bottlenecks, Invasives

Ecology

Distribution, Migration, Food Webs, Dispersal, Invasives, Stable Isotopes

Emerging Pathogen Detection

Identification, Monitoring
Host Switching, Range Expansion

Evolution

Response to Climate Change, Genome Evolution, Hybridization, Demography



Figure 2(on next page)

Natural history museum specimens.

Natural history museum specimens drive the cycle between inspiration and innovation, which spans across exhibitions and collections.

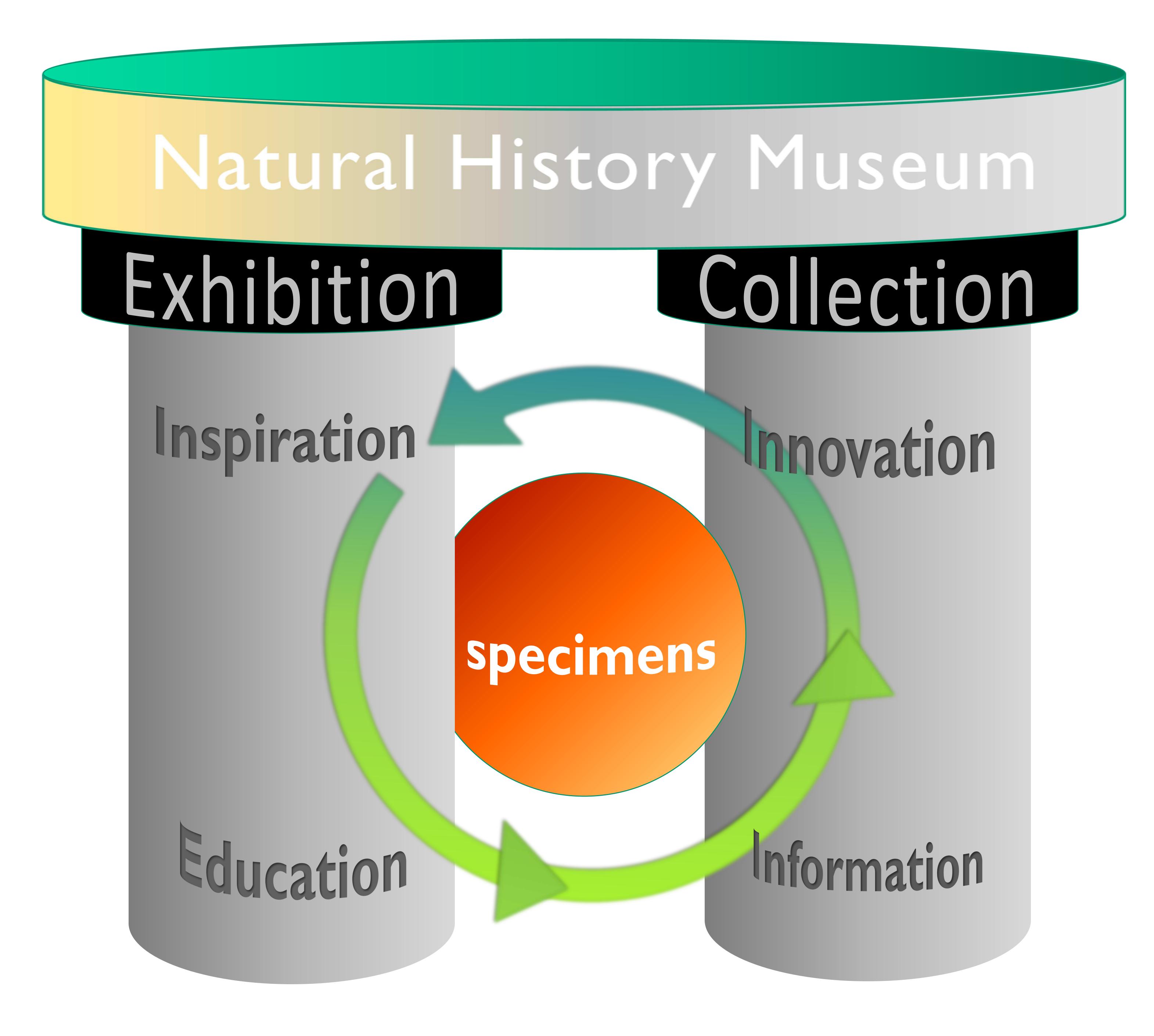




Figure 3(on next page)

Relative abundance of topics and themes.

Word cloud illustrating relative abundance of topics and themes covered in this study.



