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# A comprehensive approach towards the systematics of Cervidae

Nicola S Heckeberg<sup>Corresp., 1, 2, 3</sup>, Gert Wörheide<sup>1, 2, 4</sup>

<sup>1</sup> Department of Earth and Environmental Sciences, Palaeontology & Geobiology, Ludwig-Maximilians-Universität München, Munich, Germany

<sup>2</sup> SNSB-Bayerische Staatssammlung für Paläontologie und Geologie, Munich, Germany

<sup>3</sup> Leibniz Institute for Evolution and Biodiversity Science, Museum für Naturkunde, Berlin, Germany

<sup>4</sup> Geobio-CenterLMU, Munich, Germany

Corresponding Author: Nicola S Heckeberg  
Email address: nicola.heckeberg@mfn.berlin

Systematic relationships of cervids have been controversial for decades. Despite new input from molecular systematics, consensus could only be partially reached. The initial, gross (sub)classification based on morphology and comparative anatomy was mostly supported by molecular data. The rich fossil record of cervids has never been extensively tested in phylogenetic frameworks concerning potential systematic relationships of fossil cervids to extant cervids. The aim of this work was to investigate the systematic relationships of extant and fossil cervids using molecular and morphological characters and make implications about their evolutionary history based on the phylogenetic reconstructions. To achieve these objectives, molecular data were compiled consisting of five nuclear markers and the complete mitochondrial genome of 50 extant and one fossil cervid species. Several analyses using different data partitions, taxon sampling, partitioning schemes, and optimality criteria were undertaken. In addition, the most extensive morphological character matrix for such a broad cervid taxon sampling was compiled including 168 cranial and dental characters of 41 extant and 29 fossil cervid species. The morphological and molecular data were analysed in a combined approach and other comprehensive phylogenetic reconstructions. The results showed that most of the Miocene cervids were more closely related to each other than to any other cervids. They were often positioned between the outgroup and all other cervids or as the sister taxon to Muntiacini. Two Miocene cervids were frequently placed within Muntiacini. Plio- and Pleistocene cervids could often be affiliated to Cervini, Odocoileini or Capreolini. The phylogenetic analyses of this work provide new insights into the evolutionary history of cervids. Several fossil cervids could be successfully related to living representatives, confirming previously assumed affiliations based on comparative morphology and introducing new hypotheses. New systematic relationships were observed, some uncertainties persisted and resolving

systematics within certain taxa remained challenging.

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Nicola S. Heckeberg<sup>1,2,3</sup> and Gert Wörheide<sup>1,2,4</sup>

<sup>1</sup>Department of Earth and Environmental Sciences, Palaeontology & Geobiology, Ludwig-Maximilians-Universität München, Richard-Wagner-Str. 10, 80333 Munich, Germany

<sup>2</sup>SNSB – Bayerische Staatssammlung für Paläontologie und Geologie, Richard-Wagner-Str. 10, 80333 München, Germany

<sup>3</sup>Current address: Museum für Naturkunde, Leibniz Institute for Evolution and Biodiversity Science, Invalidenstrasse 43, 10115 Berlin, Germany

<sup>4</sup>GeoBio-CenterLMU, Richard-Wagner-Str. 10, 80333 Munich, Germany

Corresponding author:  
Nicola S. Heckeberg<sup>1,2,3</sup>

Email address: nicola.heckeberg@mfu.berlin

## ABSTRACT

Systematic relationships of cervids have been controversial for decades. Despite new input from molecular systematics, consensus could only be partially reached. The initial, gross (sub)classification based on morphology and comparative anatomy was mostly supported by molecular data. The rich fossil record of cervids has never been extensively tested in phylogenetic frameworks concerning potential systematic relationships of fossil cervids to extant cervids. The aim of this work was to investigate the systematic relationships of extant and fossil cervids using molecular and morphological characters and make implications about their evolutionary history based on the phylogenetic reconstructions. To achieve these objectives, molecular data were compiled consisting of five nuclear markers and the complete mitochondrial genome of 50 extant and one fossil cervid species. Several analyses using different data partitions, taxon sampling, partitioning schemes, and optimality criteria were undertaken. In addition, the most extensive morphological character matrix for such a broad cervid taxon sampling was compiled including 168 cranial and dental characters of 41 extant and 29 fossil cervid species. The morphological and molecular data were analysed in a combined approach and other comprehensive phylogenetic reconstructions. The results showed that most of the Miocene cervids were more closely related to each other than to any other cervids. They were often positioned between the outgroup and all other cervids or as the sister taxon to Muntiacini. Two Miocene cervids were frequently placed within Muntiacini. Plio- and Pleistocene cervids could often be affiliated to Cervini, Odocoileini or Capreolini. The phylogenetic analyses of this work provide new insights into the evolutionary history of cervids. Several fossil cervids could be successfully related to living representatives, confirming previously assumed affiliations based on comparative morphology and introducing new hypotheses. New systematic relationships were observed, some uncertainties persisted and resolving systematics within certain taxa remained challenging.

## INTRODUCTION

Cervidae (deer) belong to Ruminantia together with Tragulidae (chevrotains), Antilocapridae (pronghorns), Moschidae (musk deer), Giraffidae (giraffes), and Bovidae (cattle, sheep, antelopes). Cervids are the second most diverse group of ruminants and are natively distributed in the Americas, Europe and Asia inhabiting a broad variety of habitats. Apart from the recent dispersal and radiation into South America, cervids are mainly restricted to the Northern Hemisphere (Geist, 1998; Gentry, 2000; Scott and Janis, 1987; Webb, 2000).

Despite all efforts to resolve cervid (and ruminant) systematics over the past decades, there is only partial consensus from the phylogenetic reconstructions and several problems persist. Controversial species

48 delimitations, unknown taxon affiliation, contradictory information from the data, and/or incomplete  
49 phylogenetic reconstruction were specified as possible reasons for these problems. To solve phylogenetic  
50 relationships of cervids (and ruminants), however, is of considerable interest, because of their important  
51 biological and economic role as wild and domestic animals (Cronin, 1991; Randi et al., 2001; Price et al.,  
52 2005).

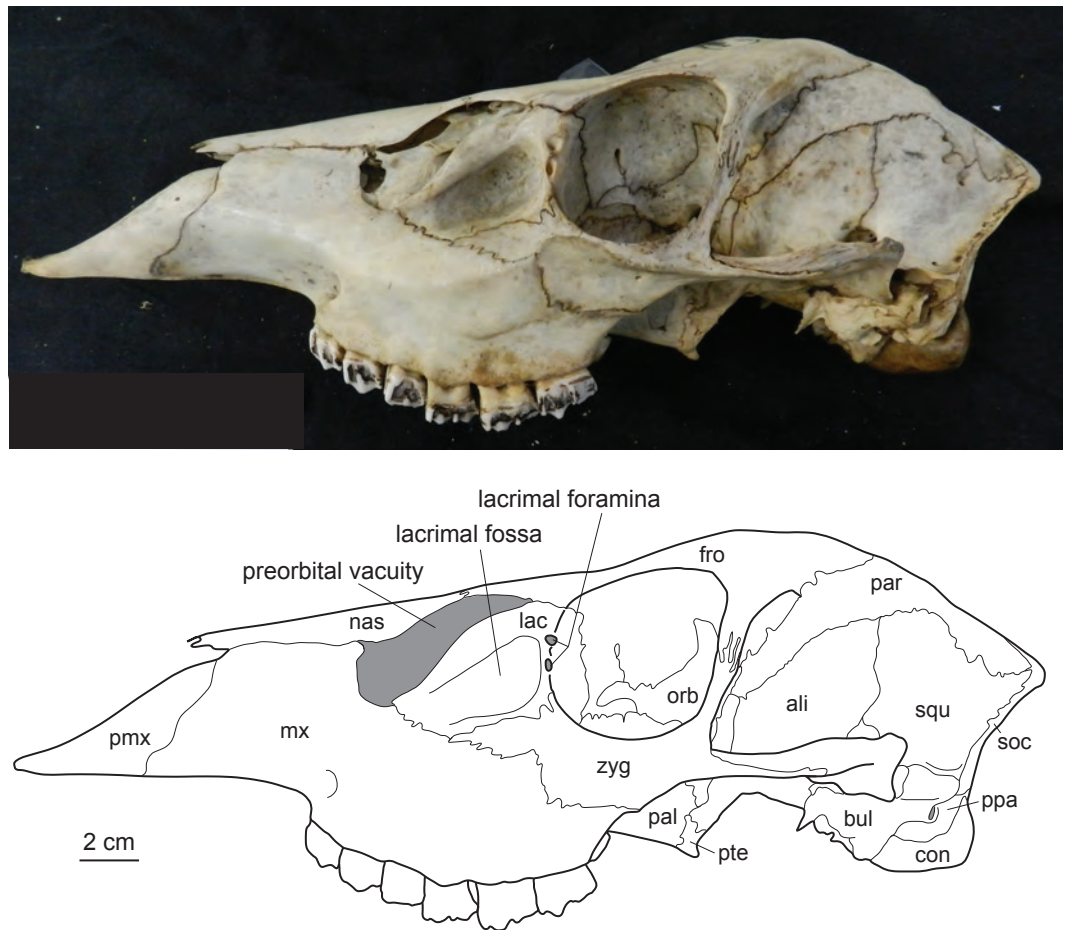
53 In contrast to early systematic studies, which were often based only on a few morphological characters,  
54 there are now numerous molecular approaches and a few supertree studies reconstructing cervid systemat-  
55 ics. However, combined or total evidence approaches are still scarce (Groves and Grubb, 1987; Groves,  
56 2014). Although the fossil record for cervids is good, systematic relationships of fossil cervids are even  
57 more uncertain than those of extant cervids. There are numerous qualitative descriptions and comparative  
58 morphological studies for fossil cervids, but there are only very few phylogenetic approaches on fossil  
59 taxa. While these were mainly based on antler characters, Mennecart et al. (2016, 2017) presented the  
60 first phylogenetic reconstructions of Miocene cervids based on inner ear morphology.

61 Various hypotheses of the intra-cervid systematic relationships have been published in the last decades.  
62 While in earlier studies up to six subfamilies of Cervidae have been recognised (Ouithavon et al., 2009),  
63 the family Cervidae now is usually classified into two subfamilies, Cervinae, consisting of Muntiacini and  
64 Cervini, and Capreolinae, consisting of Alceini, Capreolini, Odocoileini, and Rangiferini (e.g., Groves  
65 and Grubb, 1990; Miyamoto et al., 1990; Cronin et al., 1996; Randi et al., 1998, 2001; Hassanin and  
66 Douzery, 2003; Kuznetsova et al., 2005; Price et al., 2005; Gilbert et al., 2006; Hughes et al., 2006;  
67 Ouithavon et al., 2009; Hassanin et al., 2012; Heckeberg et al., 2016). This classification is supported by  
68 classical morphological concepts and molecular evidence. In some studies Muntiacini is considered as a  
69 subfamily (e.g., Cronin et al., 1996; Randi et al., 1998; Kuznetsova et al., 2005; Marcot, 2007). While the  
70 systematic relationships within Muntiacini and Cervini are resolved, with very few exceptions, systematic  
71 relationships within Capreolinae are much more controversial. The position of Capreolini and Alceini  
72 is uncertain and there are many polyphylies within Odocoileini (Heckeberg et al., 2016). The latter is  
73 the youngest clade of cervids and has a rapid diversification rate, which makes resolving the systematic  
74 relationships more difficult.

75 Diagnostic characters of cervids include for example the presence of two lacrimal foramina, a lacrimal  
76 fossa, a preorbital vacuity and brachyodont dentition (Fig. 1) Janis and Scott (1987, 1988); Bouvrain  
77 et al. (1989); Mickoleit (2004). The first classification based on morphological characters split Cervidae  
78 into Telemetacarpini and Plesiometacarpini, which is equivalent to the Cervinae-Capreolinae split (Brooke,  
79 1878). This split into Capreolinae and Cervinae was also confirmed by behavioural characters (Cap et al.,  
80 2002; Groves, 2007). Further subdivision solely based on morphological features is difficult, because  
81 most cervid characters are highly conservative, partly phylogenetically uninformative and/or prone to  
82 convergence because of ecological adaptation (Groves and Grubb, 1987; Janis and Scott, 1987; Lister,  
83 1996; Wada et al., 2007). However, there are a few morphological characters diagnosing cervid subclades  
84 (Bouvrain et al., 1989; Cronin, 1991).

85 With increasing molecular data outweighing morphological characters, morphology became less  
86 important in phylogenetic reconstructions (Huelsenbeck and Rannala, 2000). Discrepancies between  
87 morphological and molecular studies on ruminants demonstrated the need to continue combining fossil  
88 and extant species in order to reconstruct accurate phylogenies and to understand macro-evolutionary  
89 processes, which should yield better estimates than individual analyses (Hillis and Wiens, 2000; Hernández  
90 Fernández and Vrba, 2005). Several studies show the benefit of combining molecular and morphological  
91 data of fossil and living taxa in supermatrix analyses (e.g., Asher, 2007; Geisler et al., 2011; Bibi et al.,  
92 2012; Bibi, 2014). Complete species-level taxon and extensive data sampling are required to reconstruct  
93 the ecological, biological and geographical patterns of cervid and ruminant evolutionary history (Price  
94 et al., 2005).

95 Here, extensive taxon and data sampling across Cervidae was undertaken for the first time. The mor-  
96 phological data set focused on cranial and dental characters. Five nuclear markers and the mitochondrial  
97 genome were analysed and combined with the morphological data set. Several analyses were undertaken  
98 on different partitions and the combined data sets analysing fossil and extant taxa separately and together,  
99 and under different optimality criteria. Additionally, analyses using a molecular and morphological  
100 supermatrix or a constraint topology including only one fossil at a time and the Evolutionary Placement  
101 Algorithm (EPA) approach (Berger et al., 2011) were undertaken. The total evidence approaches incorpo-  
102 rated 79 fossil and living cervids covering their entire evolutionary history from the early Miocene until



**Figure 1. Diagnostic characters of cervids.** The most important anatomical features of cervids are outlined in this figure as a photograph and drawing of *Blastocerus dichotomus* (MNHN 1933-207). Note the brachyodont dentition, the preorbital vacuity, lacrimal fossa, and lacrimal foramina. Abbreviations: pmx = premaxillary, mx = maxillary, nas = nasal, lac = lacrimal, zyg = zygomaticum, pal = palatine, pte = pterygoid, orb = orbisphenoid, fro = frontal, par = parietal, ali = alisphenoid, squ = squamosal, soc = supraoccipital, ppa = paroccipital processes, bul = auditory bulla, con = condyles.

103 today.

104 We were able to investigate the strength of morphological characters to reconstruct a cervid phylogeny,  
 105 the systematic position of fossil cervids, and the influence of data partitioning and varying taxon sampling  
 106 on the phylogenetic signal. The results provide new and intriguing insights into how fossil cervids are  
 107 related to extant cervids.

## 108 METHODS

### 109 Data

#### 110 Molecular Data

111 Molecular data were compiled from GenBank ([ncbi.nlm.nih.gov/genbank/](https://ncbi.nlm.nih.gov/genbank/)). Five nuclear markers and  
 112 the mitochondrial genome were chosen for phylogenetic reconstructions based on their taxon sampling  
 113 across cervids ( $n > 10$ ). The GenBank accession numbers are in the Supplementary Material (Table S1).  
 114 The molecular data set included the nuclear non-coding markers,  $\alpha$ -lactalbumin (*Lalba*), protein kinase C  
 115 iota (*Prkci*), and the sex determining region on the Y-chromosome (*Sry*) and the nuclear coding markers

116  $\kappa$ -casein (*Csn*) and prion protein (*Prnp*) and the partially coding mitochondrial genome. The coding  
117 markers were partitioned according to codon positions 1-3. Each gene was aligned in SeaView 4.2 (Gouy  
118 et al., 2010) and Mesquite v.2.75 (Maddison and Maddison, 2011); alignments were carefully checked by  
119 eye for stop codons and/or unusual codon positions by translation into amino acids, where applicable, and  
120 were manually corrected if necessary. Some regions have been excluded from the alignment, for example  
121 the first and last couple of sites, which were not available for all taxa in the alignment. The combined  
122 molecular data set included one fossil and 50 extant cervids.

### 123 **Morphological Data**

124 In total, 41 extant cervid species, 29 fossil cervid species, six non-cervid extant ruminants, and two non-  
125 cervid fossil ruminants were measured and character-coded into the morphological matrix. The measuring  
126 distances are in the Supplementary files, the measurements in Table S3. The extant species were studied  
127 on 232 specimens, the fossil species were studied on 504 specimens (see Table S2 for complete specimen  
128 lists). Most of the fossil cervid taxa consisted of fragments of several individuals. The fossils ranged from  
129 the Miocene to the Holocene and their temporal ranges are shown in Figure 2. The character matrices and  
130 character state lists are available on morphobank (<http://morphobank.org/permalink/?P1021>).

### 131 **Phylogenetic Analyses**

132 Figure 3 is an overview of all data sets and analyses undertaken. Tragulids were chosen as the outgroup  
133 for all analyses.

### 135 **Model Choice**

136 **Molecular Data.** For each alignment we used PartitionFinder (Lanfear et al., 2012) to identify the  
137 appropriate substitution model and the optimal partitioning scheme. The Hasegawa-Kishino-Yano model  
138 (HKY; Hasegawa et al., 1985), and the Generalised Time Reversible model (GTR; Tavaré, 1986) were  
139 most commonly used.

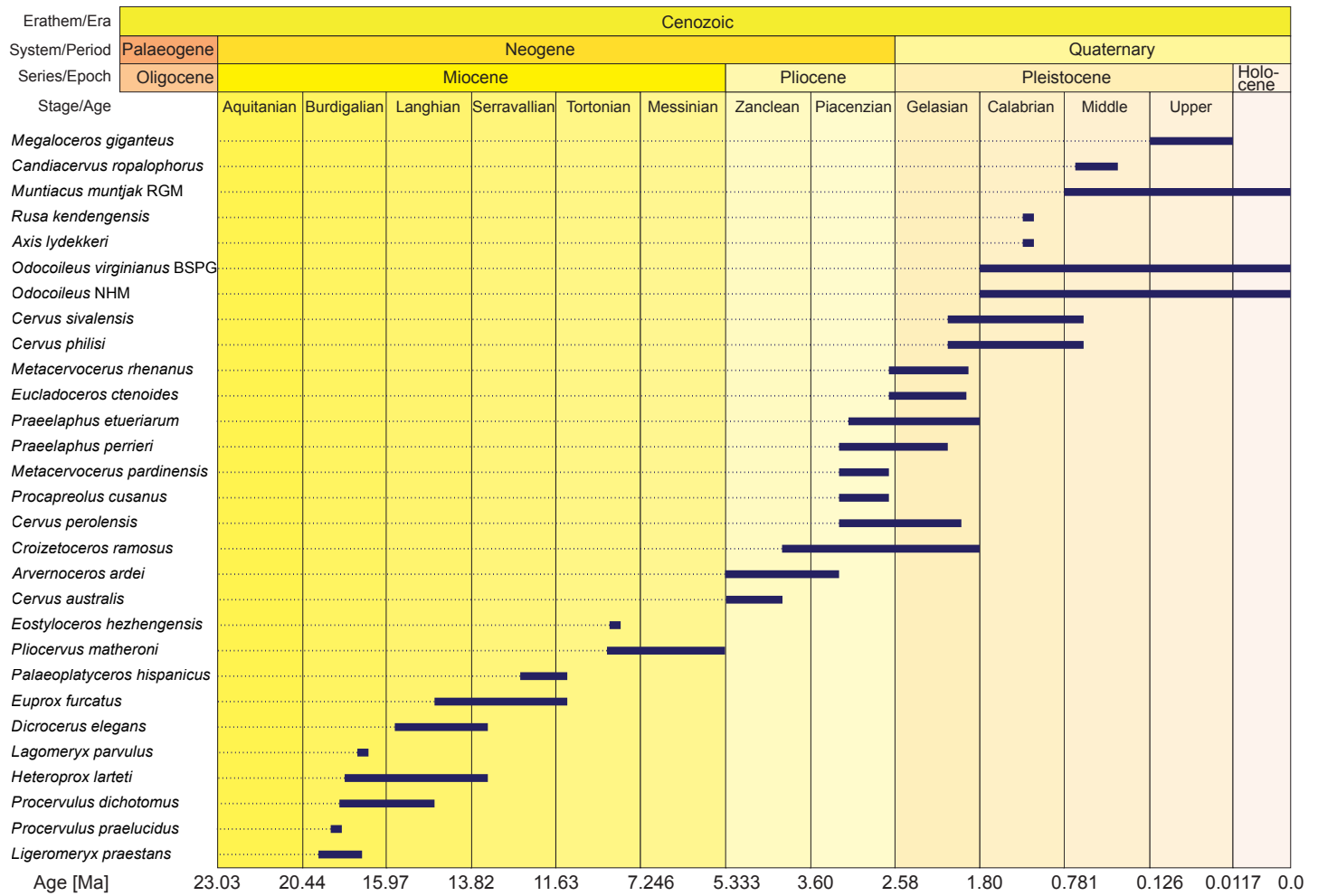
140 All analyses were run with a gamma distribution ( $\Gamma$ ) without a proportion of invariant sites (*I*), where  
141  $\Gamma$  or  $\Gamma + I$  was suggested, because combining  $\Gamma + I$  is known to cause convergence problems by creating  
142 two areas of equal probability in the tree landscape (Moyle et al., 2012). *I* was used when suggested as  
143 the sole analysis parameter.

144 After completion, the statistics of all Bayesian analyses were checked in Tracer v.1.6 ([tree.bio.ed.ac.uk](http://tree.bio.ed.ac.uk))  
145 and convergence between runs was checked using the visualisation tool AWTY (Wilgenbusch et al.,  
146 2004).

147 **Stepping Stone Analyses for Morphological Data.** The best fit of model distribution and partitioning  
148 scheme of the morphological character sets was tested using the efficient stepping stone (ss) sampling  
149 (Xie et al., 2011). The Bayes Factor (BF) was calculated as the ratio of the marginal likelihood of one  
150 model to the marginal likelihood of the competing model; BFs can then be used as the relative evidence  
151 in the data that favours one hypothesis in that respect that it predicts the observed data better than the  
152 competing hypotheses (Xie et al., 2011).

153 To test the combined morphological data set for the most suitable partitioning scheme, ordering  
154 scheme (unordered vs. ordered), and model distribution choice (gamma vs. not gamma), ss analyses were  
155 undertaken. First, the data set was tested for the partitioning scheme with an analysis of the unpartitioned  
156 data set, a maximally, and a minimally partitioned data set. Afterwards, the data set, applying the resulting  
157 partitioning scheme, was tested for the gamma ( $\Gamma$ ) distribution (Yang, 1994), and for ordering characters.

158 Each SS analysis was run for 21.5 million generations, with a diagnostic frequency of 1000 and a  
159 sample frequency of 500 and had 40 steps in total. The general settings are the same as for a normal BI  
160 analysis with MrBayes (Ronquist et al., 2012). The initial burnin of samples and the additional burnin  
161 in each step of the ss sampling were discarded. The aforementioned importance distributions are called  
162 power posterior distributions and were sampled via the Metropolis Coupled Monte Carlo Markov Chain  
163 ( $MC^3$ ) run (Ronquist et al., 2012). In MrBayes this parameter is called alpha and was left as the default  
164 setting of 0.4, because in empirical studies it was found that the accuracy is maximal with an alpha value  
165 between 0.3 and 0.5 (Ronquist et al., 2012). After completion of the ss analyses the BFs of the summary  
166 of the marginal likelihoods of all 40 steps were calculated and compared with each other to decide for the  
167 favoured hypothesis.

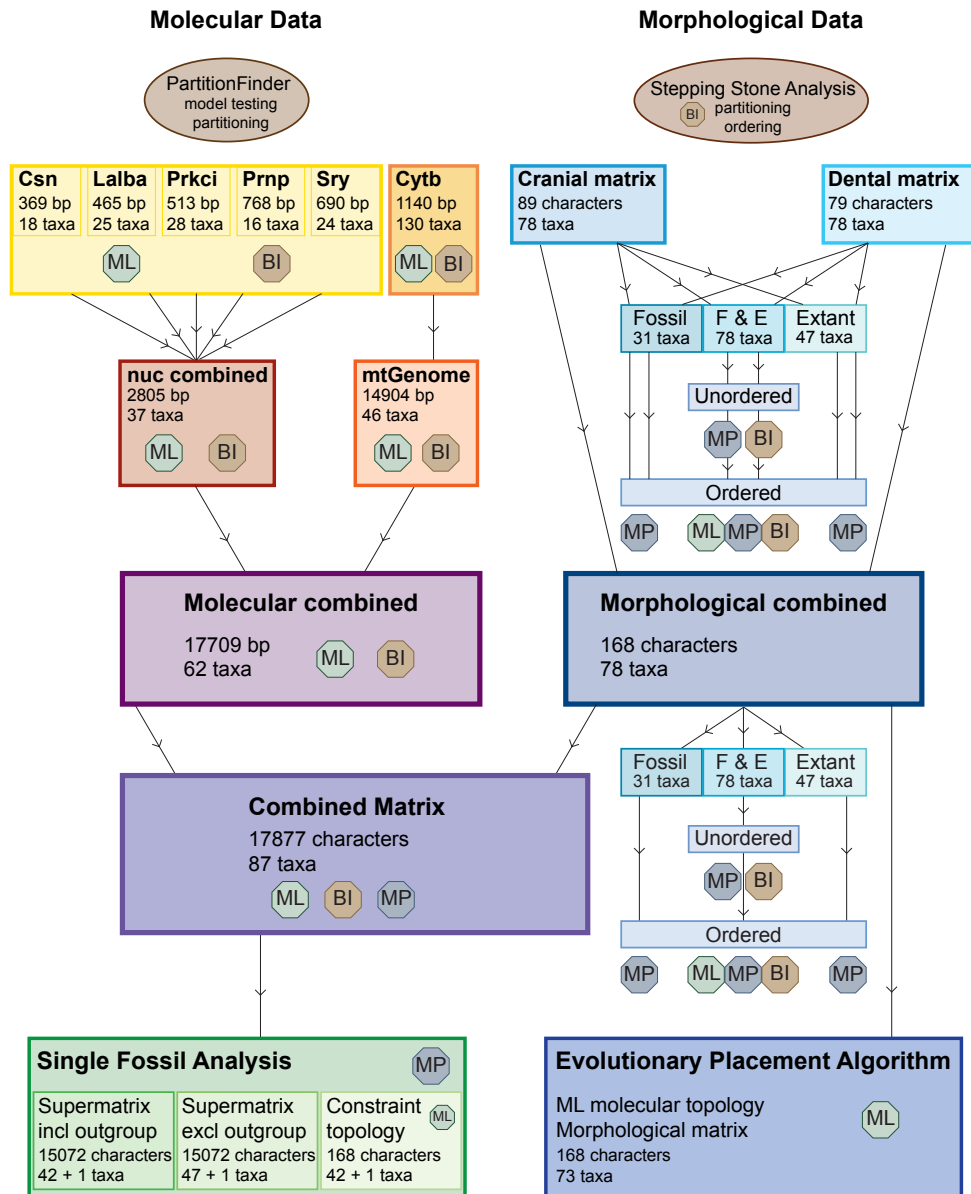


**Figure 2. Age ranges of fossil cervids.** Fossil cervids are arranged from the oldest first appearance datum (bottom) to the youngest first appearance datum (top). The stage column widths are not to scale with time. The dates were compiled from the literature (Gentry et al., 1999; Steininger, 1999; Böhme et al., 2012; Hilgen et al., 2012; Cohen et al., 2013; Croitor, 2014) and databases (NOW: [www.helsinki/science/now/](http://www.helsinki/science/now/), PBDB: [www.paleobiodb.org/](http://www.paleobiodb.org/)).

### Analyses of Morphological Data

**Data Sets.** All morphological data sets included 78 taxa, 41 extant cervid species, 29 fossil cervid species, 6 non-cervid extant ruminant species, and 2 fossil non-cervid ruminant species. In the dental ma-





**Figure 3. Overview of all analyses.** This overview shows all analyses undertaken and the optimality criteria under which they were run. Abbreviations: BI = Bayesian inference, ML = maximum likelihood, MP = maximum parsimony.

171 trix 79 characters were coded in total; 35 characters concerning upper dentition, 39 characters concerning  
 172 the lower dentition, and 5 characters concerning the upper canines and lower incisors and canines. There  
 173 were 8 discrete quantitative characters and 11 characters were suitable for ordering (6–8, 14, 21, 32, 40,  
 174 51, 59, 64, 72).

175 In the cranial matrix 89 characters were coded in total; 7 characters concerning the mandible, 65  
 176 concerning the cranium, and 17 concerning antlers and pedicles. There were 17 discrete quantitative  
 177 characters and 23 characters were suitable for ordering (2, 4, 5, 8–12, 14, 15 17–20, 23, 61, 74–79, 89).  
 178 The combined morphological data set consisted of 168 characters, of which 19 were discrete quantitative  
 179 characters and 34 were suitable for ordering (see above).

180 **Standard Phylogenetic Analyses.** The dental, cranial and morphological combined data sets were  
181 analysed each with maximum parsimony (MP) with or without character ordering and varying taxon  
182 sampling, with Bayesian inference (BI) with and without character ordering, and with maximum likelihood  
183 (ML) without character ordering (Table 1).

184 All MP analyses including bootstrap analyses were undertaken using PAUP\* v.4.0b (Swofford, 2002).  
185 The analyses used a heuristic search running 1000 replicates. Sequences were added randomly using the  
186 tree-bisection-reconnection (TBR) algorithm. Polymorphisms were treated as real polymorphisms. The  
187 strict consensus tree was calculated from all trees sampled in each analysis.

188 The BI analyses were undertaken using MrBayes 3.2.4 (Ronquist et al., 2012) and ran for 50 million  
189 generations with two runs à four chains at a temperature of 0.35; trees were sampled at every 5000<sup>th</sup>  
190 generation until the standard deviation of split frequencies was below 0.01.

191 The ML analyses were undertaken using RAxML v.8.0.26 (Stamatakis, 2014). All ML analyses  
192 started at a random number seed and were run under the Mk-model (Lewis, 2001) with the  $\Gamma$  model  
193 rate of heterogeneity without invariant sites. The analyses also included a rapid bootstrap search of 100  
194 replicates starting at a random number seed.

195 **Single Fossil Analyses (SFA).** In order to reduce missing data and noise in the data set, three sets  
196 of analyses were run, which included only one fossil taxon at a time. The first approach included the  
197 entire morphological data set and was combined with the complete mitochondrial genome (including  
198 cytochrome b (*Cytb*) only for taxa without a complete mitochondrial genome) to facilitate tree search.  
199 The data set comprised 78 taxa and 15072 characters in total. In each analysis 47 extant and one fossil  
200 species were included. The second approach was on the same data set, but excluding the 5 non-cervid  
201 ruminants; it consisted of 73 taxa and 15072 characters. In each analysis 42 extant and one fossil species  
202 were included. The third approach was based on the morphological character matrix and a constraint  
203 topology. This constraint topology was generated in an analysis of the combined molecular data including  
204 only those taxa, for which morphological data were available. The third SFA data set comprised 73  
205 (excluding 5 non-cervid ruminants) taxa and 168 morphological characters. In each analysis only 42  
206 extant and one fossil species were included. All SFA analyses were run with the PAUP\* settings specified  
207 above (Table 1).

208 **Evolutionary Placement Algorithm (EPA).** Berger et al. (2011) introduced an algorithm implemented  
209 in RAxML, which improves accurate placement of morphology-based fossils in a tree. The EPA analysis  
210 is a two step process. The first step is a morphological weight calibration, where a molecular tree is  
211 provided alongside with the morphological matrix. All taxa have to entirely overlap in this step, therefore,  
212 only extant taxa were included. The second step invokes the actual evolutionary placement algorithm  
213 using the same molecular tree as in step one, the morphological matrix, including extant and fossil taxa,  
214 and the weight vector output from step one.

215 The molecular tree used here was specifically generated in RAxML based on a data set including only  
216 the 41 cervid species for which morphological data was available, 17709 base pairs (nuc and mtDNA),  
217 and *Hyemoschus aquaticus* as outgroup. The morphological matrix for step one contained 42 species and  
218 168 morphological characters (Table 1). The second step of the EPA analysis used the same molecular  
219 tree, the morphological matrix now containing 73 taxa, and the morphological weights from the first step.

#### 220 **Analyses of Molecular Data**

221 Each nuclear gene was initially analysed separately and all five nuclear genes were analysed in a  
222 supermatrix. The combined nuclear data set comprised 2805 base pairs for 28 cervid species and nine  
223 non-cervid ruminant species (Table 1).

224 The individual nuclear gene analyses (BI) were run for five million generations at a temperature for  
225 the heated chain of 0.5 and sampled every 1000th generation using MrBayes v.3.2 (Ronquist et al., 2012).  
226 The combined nuclear data set was run for eight million generations with the same parameter settings  
227 as the individual gene analyses. The ML analyses for all single nuclear genes and the combined nuclear  
228 data set were analysed with RAxML v.2.7.3 (Stamatakis, 2006). The ML analyses also included a rapid  
229 bootstrap analysis.

230 The complete mitochondrial genome (mtG) available for 33 cervid species including 39 taxa and  
231 seven non-cervid ruminants with a total of 14904 base pairs of Hassanin et al. (2012) was re-analysed.  
232 The extensive *Cytb* data set from Heckeberg et al. (2016) was combined with the mtG. For the combined  
233 mtG-*Cytb*-analyses, the original *Cytb* region of the mtG was replaced by the more taxon-rich *Cytb*

**Table 1.** Overview of all analyses undertaken. <sup>x</sup> indicates analyses that were not successful, <sup>\*</sup> indicates topologies that are figured in the main text, <sup>#</sup> only summarising topology figured in the main text; the topologies of all other analyses can be found in the Supplemental material. Abbreviations: Dent = Dental, Cran = Cranial, Combi = Combined UnO = unordered, O = ordered, MP = maximum parsimony, BI, MB = Bayesian inference, ML = maximum likelihood, noOut = excluding most outgroup taxa, nuc = nuclear marker, mt = mitochondrial marker, Opt. Crit.=Optimality Criterion, nchar = number of characters, ntax = number of taxa, E=Extant, F=Fossil.

Analysis ID	Opt. Crit.	Data Set	nchar	ntax
Dent_UnO_FE	MP	dental	79	78
Dent_O_FE	MP	dental	79	78
Dent_O_E	MP	dental	79	78
Dent_O_F	MP	dental	79	78
Dent_MB_UnO	BI	dental	79	78
Dent_MB_O	BI	dental	79	78
Dent_ML	ML	dental	79	78
Cran_UnO_FE <sup>x</sup>	MP	cranial	89	78
Cran_O_FE <sup>x</sup>	MP	cranial	89	78
Cran_O_E	MP	cranial	89	78
Cran_O_F <sup>x</sup>	MP	cranial	89	78
Cran_MB_UnO	BI	cranial	89	78
Cran_MB_O	BI	cranial	89	78
Cran_ML	ML	cranial	89	78
Combi_UnO_FE <sup>*</sup>	MP	morph. combined	168	78
Combi_O_FE <sup>*</sup>	MP	morph. combined	168	78
Combi_O_E	MP	morph. combined	168	78
Combi_O_F	MP	morph. combined	168	78
Combi_MB_UnO	BI	morph. combined	168	78
Combi_MB_O	BI	morph. combined	168	78
Combi_ML	ML	morph. combined	168	78
SFA_Supermatrix <sup>#</sup>	MP	morph. mol. combined	15072	48 (78)
SFA_Supermatrix_noOut <sup>#</sup>	MP	morph. mol. combined	15072	43 (73)
SFA_Backbone <sup>#</sup>	MP	morph. combined	168	43 (73)
EPA <sup>*</sup>	ML	morph. mol. combined	17709	42 (73)
			+ 168	
<i>Csn</i>	BI/ML	nuc molecular	369	20
<i>Lalba</i>	BI/ML	nuc molecular	465	25
<i>Prkci</i>	BI/ML	nuc molecular	513	29
<i>Prnp</i>	BI/ML	nuc molecular	768	21
<i>Sry</i>	BI/ML	nuc molecular	690	70
nucCombined	BI <sup>*</sup> /ML	nuc molecular	2805	37
mtGenome	BI/ML	mt molecular	14904	46
<i>Cytb</i>	BI/ML	mt molecular	1140	130
mtCombined	BI <sup>*</sup> /ML	mt molecular	14904	62
Molecular_Combined	BI <sup>*</sup> /ML	molecular combined	17709	62
Mor_Mol_Combined <sup>*</sup>	BI/ML/MP	morph. mol. combined	17877	87

234 alignment. The mitochondrial combined matrix included 51 cervid species across 56 cervid taxa and six  
235 non-cervid ruminants (Table 1).

236 The mtG-*Cytb* combined data set contained seven partitions according to Hassanin et al. (2012). For  
237 the BI analyses two runs à four chains sampled the tree landscape at a temperature of 0.35 until the  
238 standard deviation of split frequencies was below 0.01. Trees were sampled every 5000th generation. The  
239 ML analyses for both data sets included rapid bootstrap analyses and used the same partitioning scheme  
240 as in the BI analyses.

241 The combined molecular matrix consisted of 17709 base pairs for 56 cervid taxa including 50 extant  
242 and 1 fossil cervid species and 6 non-cervid ruminant species (Table 1). This data set was analysed using  
243 ML and BI with the same settings as above.

#### 244 **Combined Molecular and Morphology Analyses**

245 The total evidence (TE) matrix consisted of 17877 characters. The 87 taxa included two fossil and six  
246 extant non-cervid ruminant species and 29 fossil and 50 extant cervid species. This data set was run using  
247 ML, BI, and MP (Table 1).

## 248 **RESULTS**

### 249 **Morphological Data**

250 Figure 4 provides an overview of how well each species was sampled for morphological data. All fossil  
251 taxa are sampled for at least three partitions. The most incomplete fossil is *Eostyloceros hezhengensis*  
252 sampled from the literature with 70 % missing data followed by *Ligeromeryx praestans* with 68 %  
253 missing data. The most complete fossil cervids were *Megaloceros giganteus* with 0 % missing data  
254 and *Candiacervus ropalophorus* with 6 % missing data. Most of the other fossil taxa have around 50 %  
255 missing data.

### 256 **Cranium**

257 All cervids share several anatomical features, such as two lacrimal foramina, a preorbital vacuity, and  
258 a lacrimal fossa (Fig. 1). In lateral view, the dorsal outline is convex at the braincase, concave at the  
259 fronto-nasal transition and straight at the nasals. The anterior extension of the snout is moderate depending  
260 on the overall size of the cervid species. The basicranial outline in lateral view is flexed. The preorbital  
261 vacuity varies in size and form, the lacrimal fossa can be deep and round, covering a large proportion  
262 of the facial aspect of the skull, shallow, or barely visible (particularly in females). The position of the  
263 two lacrimal foramina on the orbit rim (more internally or externally) and the position to each other is  
264 variable. A detailed description of the craniodental morphology for each cervid species investigated is in  
265 Heckeberg (2017a).

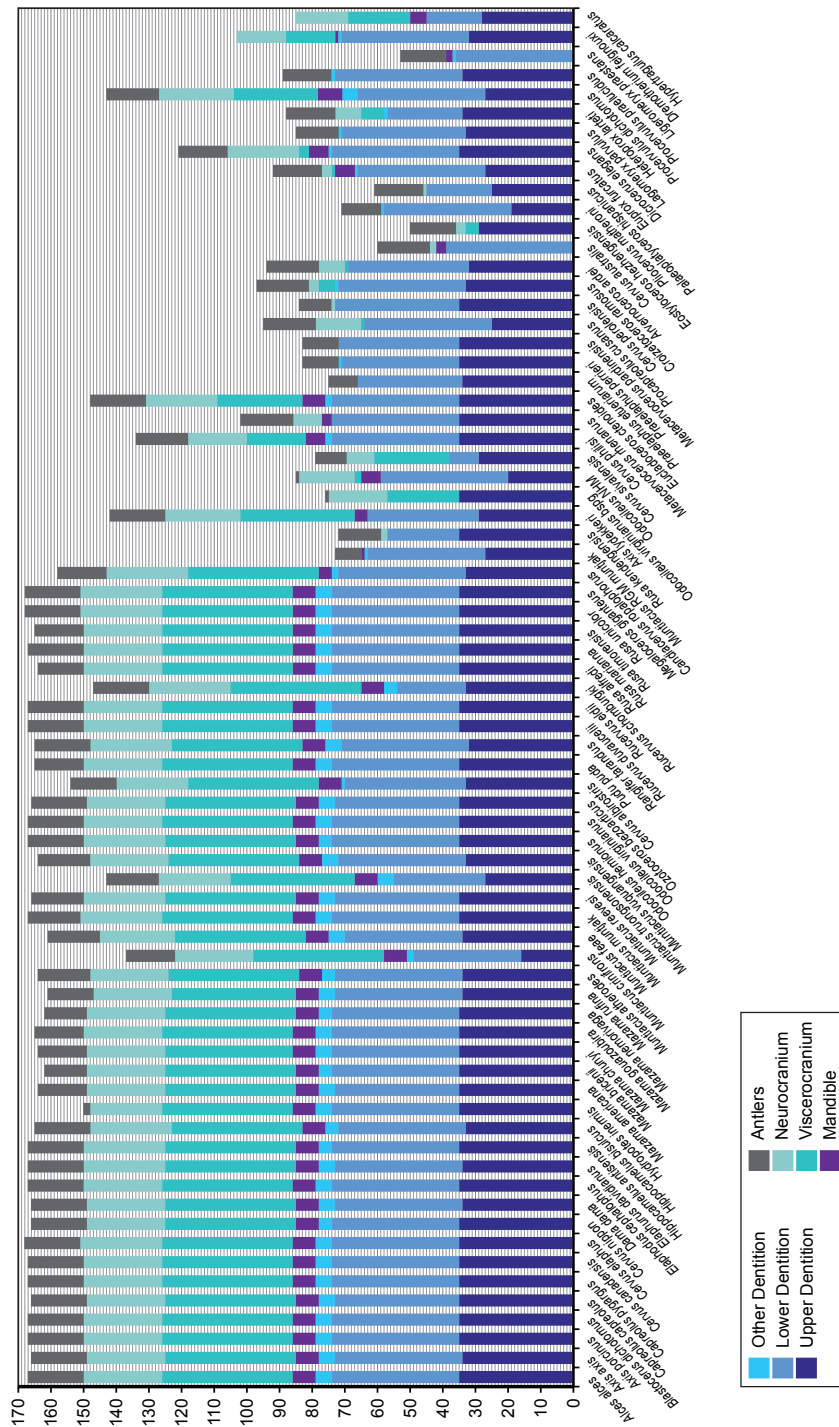
266 Some Miocene cervids have a sagittal crest (e.g., *Dicrocerus*, *Procervulus*), which is absent in all  
267 other cervids (Fig. 1). The number and size of supraorbital foramina and presence and absence of the  
268 supraorbital sulcus are variable and could potentially be features to distinguish groups of cervids; however,  
269 more specimens per species need to be investigated to confirm this. The presence of an extended vomerine  
270 septum and the division between the temporal foramina is characteristic for Capreolinae (Fig. 1). Most  
271 cervids have small, oval auditory bullae, some species have large inflated bullae (e.g., *Axis*) (Fig. 1).

272 Most Miocene cervids have long pedicles, the insertion point of the pedicle is directly above the orbit  
273 and the pedicle is upright (Fig. 6). Muntiacini, *Euprox* and *Eostyloceros* have long strongly inclined  
274 pedicles. In most other cervids the pedicles originate more posterior to the orbit, are inclined at 45–60°  
275 and short. *Mazama* and *Pudu* have strongly inclined and short pedicles.

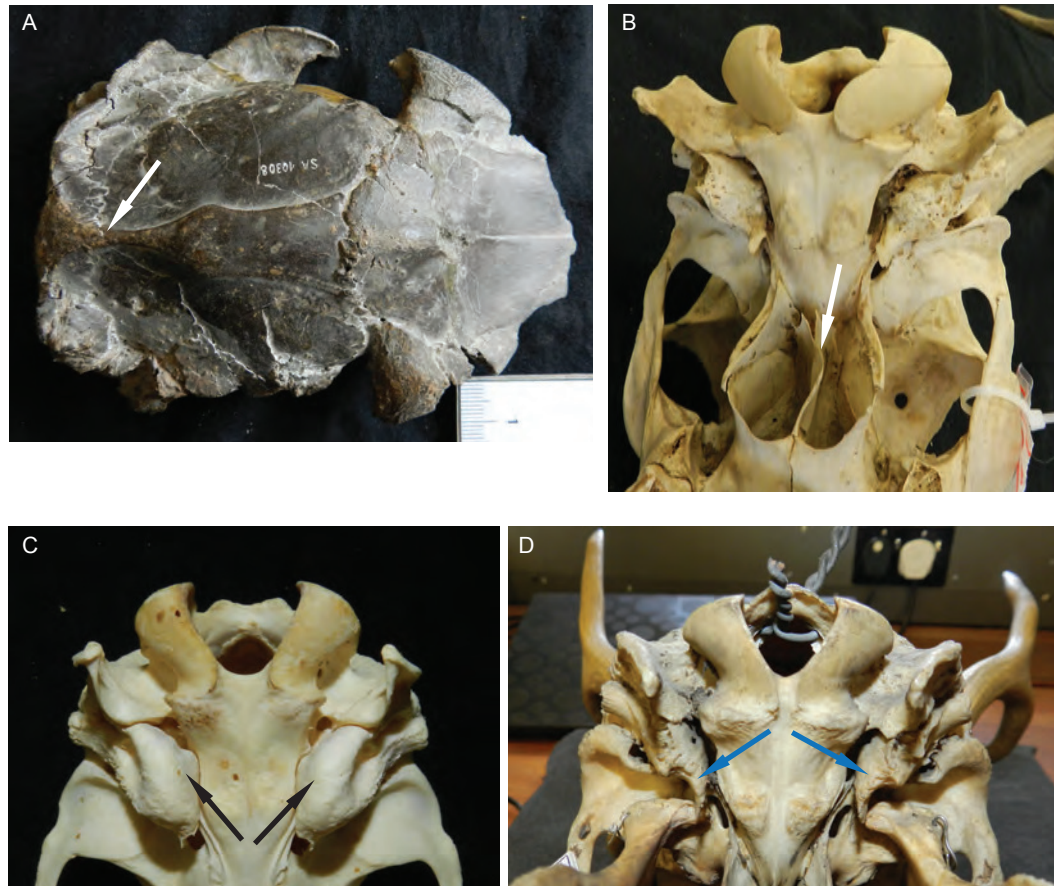
### 276 **Antlers**

277 Even though antlers are species-specific, they have a high variability, intraspecifically and ontogenetically.  
278 No antler looks exactly the same, not even the left and the right antler of the same individual are identical.  
279 Also, antlers change from one year to the next; in addition pathologies, abnormal growth, and other  
280 phenomena occur.

281 While cervid genera and most species can be qualitatively distinguished based on antler morphology,  
282 translation of these distinctions into discrete characters for quantitative or phylogenetic analyses is difficult.  
283 Convergence, which can be distinguished by eye, but is sometimes too subtle to be scored differently in  
284 the character matrix is the reason for this. Three morphotypes can be distinguished in extant cervids.



**Figure 4. Overview of the characters available for each cervid species.** Extant species are arranged in alphabetical order starting from the left, fossil cervids and the two non-cervid fossils are arranged from the youngest to the oldest following the extant taxa. Morphological characters were divided into seven partitions indicated by the different colours of each bar. The y-axis represents the absolute number of present characters.



**Figure 5. Details of the cervid cranial anatomy.** (A) Cranium of *Dicrocerus elegans* (MNHN Sa 10308) in dorsal view. The arrow indicates the sagittal crest. (B) Basicranium of *Odocoileus hemionus* (MNHN AE724). The arrow indicates the vomerine septum typical for Capreolinae. (C) Basicranium of *Axis axis* (ZSM 1958-88). The arrows indicate the large inflated auditory bullae, rarely observed in cervids. (D) Basicranium of *Ozotoceros bezoarticus* (UMZC H.18781). The arrows indicate the small flattened auditory bullae with prominent processes.

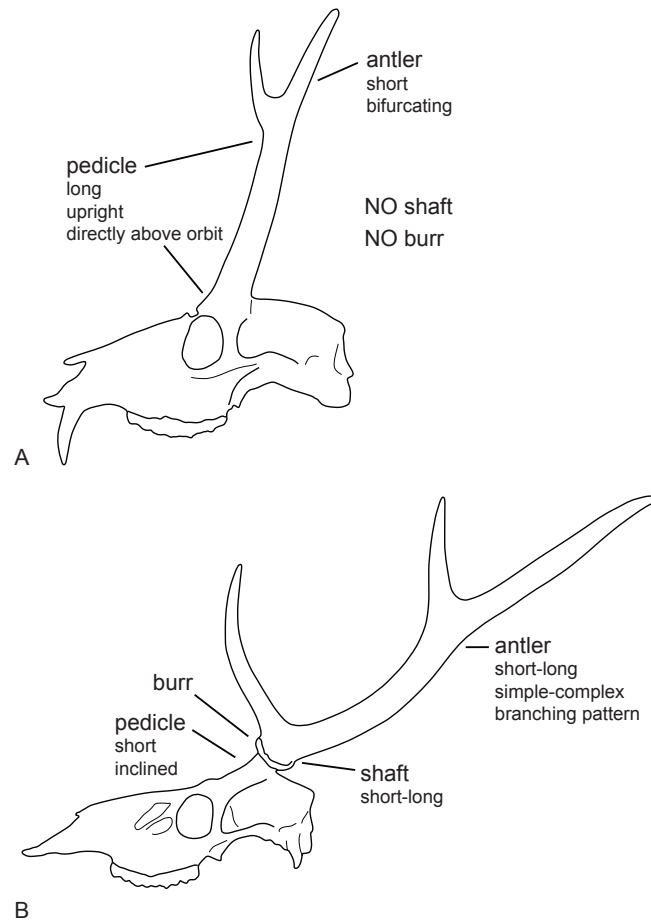
### 285 **Morphotype 1**

286 This morphotype includes all cervids with single-tined or bifurcating antlers; *Mazama* and *Pudu* have  
 287 single-tined antlers (*Pudu* antlers rarely develop a bifurcation). *Elaphodus cephalophus* has minute,  
 288 single-tined antlers. All *Muntiacus* species have bifurcating antlers on elongated inclined pedicles.  
 289 *Hippocamelus* has a bifurcating antler morphology with an open angle between the brow tine and main  
 290 tine; the main tine can have additional small tines. Fossil cervids with a bifurcating antler morphology  
 291 include *Procervulus*, *Dicrocerus*, *Heteroprox*, *Euprox*, and presumably *Cervus australis*.

### 292 **Morphotype 2**

293 This morphotype includes all cervids with antlers showing exactly three tines, e.g., *Rusa*, *Axis*, *Capreolus*,  
 294 and *Ozotoceros*. The three tines are organised either in a way, where the brow tine forms a more acute  
 295 angle to the main beam with the tip of the brow tine pointing posteriad (*Axis*, *Rusa*), or where it forms an  
 296 open angle with the tip of the brow tine pointing more upwards or forwards (*Capreolus*, *Ozotoceros*).

297 Fossil cervids of the morphotype 2 include *Axis lydekkeri*, *Rusa kendagensis*, *Metacervoceros*  
 298 *pardinensis*, '*Cervus*' *philisi*, and *Metacervoceros rhenanus* with the brow tines pointing posteriad,  
 299 *Procervulus cusanus* with the brow tines pointing upwards. *Pliocervus matheronis* antler remains are  
 300 too fragmentary to infer the direction of the brow tine unambiguously. It was also suggested that this



**Figure 6. Cervid antler evolution.** (A) Cranium of a typical Miocene cervid showing the characteristics of early pedicles and antlers. (B) Cranium of an extant cervids showing features of the pedicles and antlers seen in modern cervids.

301 species had presumably four tines (Croitor, 2014); however, as this could not be observed on the studied  
 302 specimens and literature, it was scored as possessing three tines.

### 303 **Morphotype 3**

304 This morphotype contains the more complex or palmated antlers and is present in *Alces*, *Blastoceros*,  
 305 *Cervus*, *Dama*, *Elaphurus*, *Odocoileus*, *Rangifer*, *Rucervus*. *Blastoceros dichotomus*, *Cervus albirostris*,  
 306 and *Cervus nippon* have an antler bauplan, which produces not more than four tines in adults (accessory  
 307 smaller tines not included). In *Elaphurus* it is difficult to distinguish between main tines and accessory  
 308 tines. Characteristic for *Cervus elaphus* are paired lower tines, called brow tine and bez tine, and trez tine  
 309 (Lister et al., 2010). *Dama dama* and *Rangifer tarandus* have a ramified palmated morphology, while  
 310 *Alces alces* has a palmated morphology without ramification, and thus form a subgroup within morphotype  
 311 3. The remaining eight extant cervid species develop more complex antlers with an increasing number of  
 312 tines from year to year, which is erroneously widely assumed to happen in all cervids.

313 Fossil cervids of the morphotype 3 include *Croizetoceros ramosus*, *Eucladoceros ctenoides*, *Lagomeryx*  
 314 *parvulus*, *Ligeromeryx praestans*, *Arvernoceros ardei*, *Praealaphus perrieri*, *Megaloceros giganteus*,  
 315 and *Palaeoplatyceros hispanicus*. The two lagomerycids, *Croizetoceros ramosus* and *Palaeoplatyceros*  
 316 *hispanicus* represent special cases, as their antler morphology and branching pattern is unique among  
 317 living and fossil cervids. Lagomerycids possess coronate antlers without a shaft, while *Palaeoplatyceros*  
 318 has palmated antlers without any other tines, and *Croizetoceros ramosus* shows a serial organisation of

319 small tines on the main beam. *Praeclaphus perrieri* has a distally trifurcating main beam with a basal  
320 brow tine, which is similar to the condition in *Arvernoceros ardei*, where the branching part of the main  
321 beam sometimes forms a palmation. The antler morphology of *Eucladoceros ctenoides* resembles that of  
322 *Cervus elaphus* with several short proximal tines, similar to the bez and trez tine. *Megaloceros giganteus*  
323 has enormous ramified palmated antlers similar to those of *Dama*. Also characteristic for Megacerini are  
324 flattened basal brow tines similar to *Rangifer* (Lister et al., 2010).

### 325 **Dentition**

326 Some dental characters are highly variable and thus difficult to score unambiguously. Despite convergent  
327 modifications depending on dietary requirements, a species-specific pattern underlies these adaptations in  
328 most species (pers. obs.), particularly in the lower premolars and upper molars. The difficulty is to score  
329 these species-specific patterns without scoring the convergent adaptations and the intraspecific variability.

330 **Upper premolars and molars.** The upper incisors and the P1 are absent in cervids. The upper premolar  
331 row is characterised by robust, compact, predominantly horseshoe-shaped teeth. P3 and P4 are less  
332 variable, P2 can have more rectangular or triangular outlines, particularly in early fossil cervids. All  
333 premolars have at least one prominent central fold, except for *Rangifer*, in which central folds are  
334 consistently missing (Fig. 7). Sometimes there are tiny additional folds, or the main central fold is  
335 serrated. A separation of the lingual cone into an antero- and posterolingual cone is relatively common  
336 (Fig. 7). In all Miocene cervids the P2 is longer than the P4, while in extant taxa the P4 is most often  
337 longer than the P2. Several fossil species have a well developed lingual cingulum (Fig. 7).

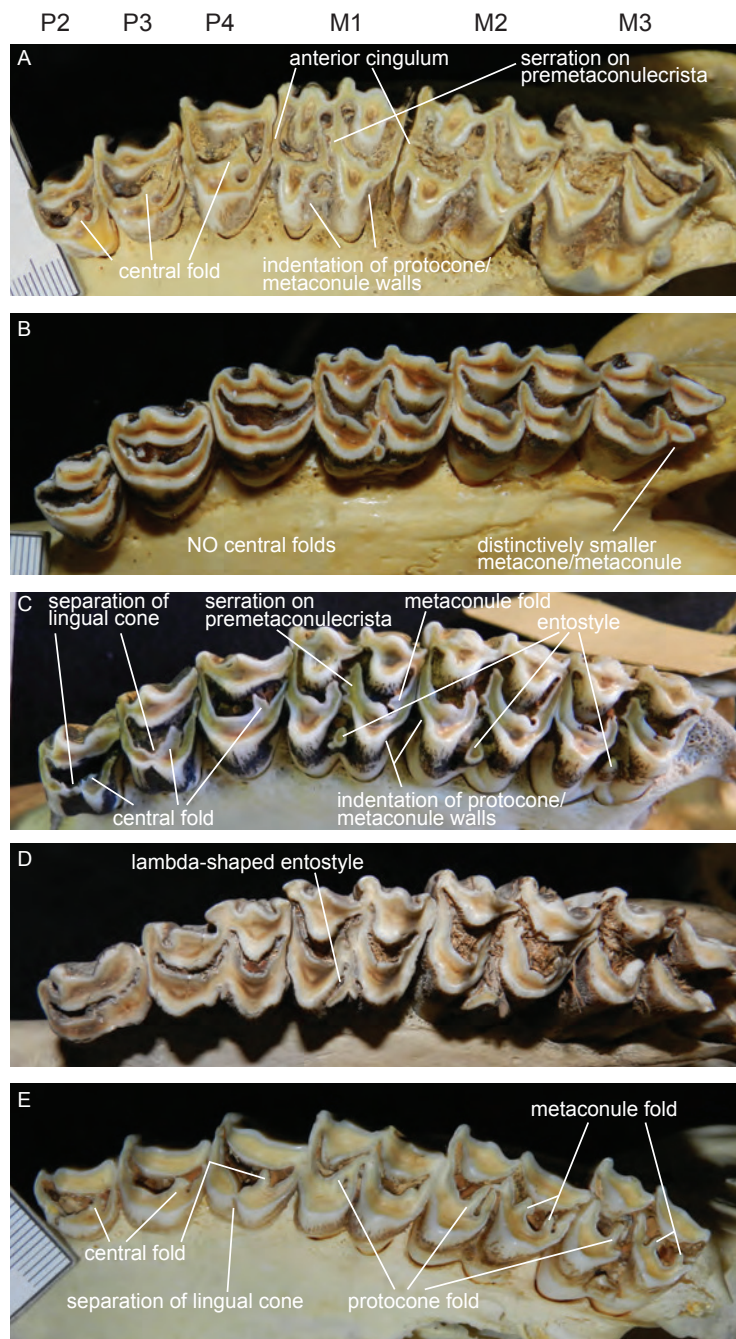
338 The upper molars are all two-lobed and quadrangular with only little variation in morphology. The  
339 posterior lobe of the M3 is distinctively smaller than the anterior one in most species. The entostyles  
340 are variably present. In some species the entostyle(s) has/have a  $\lambda$ -shaped morphology, especially in  
341 later wear (*Axis*, *Rusa*, *Rucervus* and *Elaphurus*) (Fig. 7). Metaconule folds are variably present within  
342 Cervinae and Capreolinae and are mostly small. Protocone folds are usually absent in Cervinae, while they  
343 are regularly present Capreolinae, often well developed on all molars (Fig. 7). The same applies to fossil  
344 cervids, where tiny metaconule folds are much more common than protocone folds. Only in Miocene  
345 cervids protocone folds are common. However, in these species it often looks more like a bifurcation  
346 of the postprotocrista than a fold originating from the crista, particularly when the internal part of this  
347 bifurcation is longer than the external as on M2 in *Dicrocerus*. It is not entirely evident, whether these are  
348 two independent structures or the same structure with variable characteristics. Several species have an  
349 anterior cingulum and some fossil cervids have a lingual cingulum. The protocone and metaconule folds  
350 are variably present. In a few species the premetaconulecrista is serrated. More details are in Heckeberg  
351 (2017a).

352 **Lower premolars and molars.** p1 is usually absent in cervids, although it was present in individual  
353 *Lagomeryx parvulus* specimens. The p2 has a simpler morphology with fewer elements compared to p3  
354 and p4. A strong reduction in p2 length could be observed in *Mazama* and particularly in *Ozotoceros*. In  
355 a few specimens the p2 is missing. Mesolingual cristids were variably present in p3 and p4 (absent in  
356 *Axis*, often absent in early Miocene species) (Fig. 8). p3 and p4 often show molarisation to a different  
357 extent. While p3 is molarised only in a few species and not to the same extent as p4, the p4 is molarised  
358 in many species, at least initially, and is completely molarised in *Rangifer* and *Alces* (Fig. 9). The labial  
359 incision on premolars is rarely and weakly developed in p2; it is more often developed on p3, and most  
360 often occurs on p4 (Fig. 8). p4 is the most variable tooth in cervids.

361 Some species show a spike like extension of the posterolabial conid of the p4 towards labiad; these  
362 species are *Capreolus capreolus*, *Capreolus pygargus*, *Blastocerus dichotomus*, *Hippocamelus* spp.,  
363 *Hydropotes inermis*, *Ozotoceros bezoarticus*, *Croizetoceros ramosus*, *Procapreolus cusanus*, and '*Cervus*'  
364 *philisi* (Fig. 8). Whether this feature can be used as a phylogenetic character and whether it is indicative  
365 of affiliation to a certain subclade has to be investigated in the future.

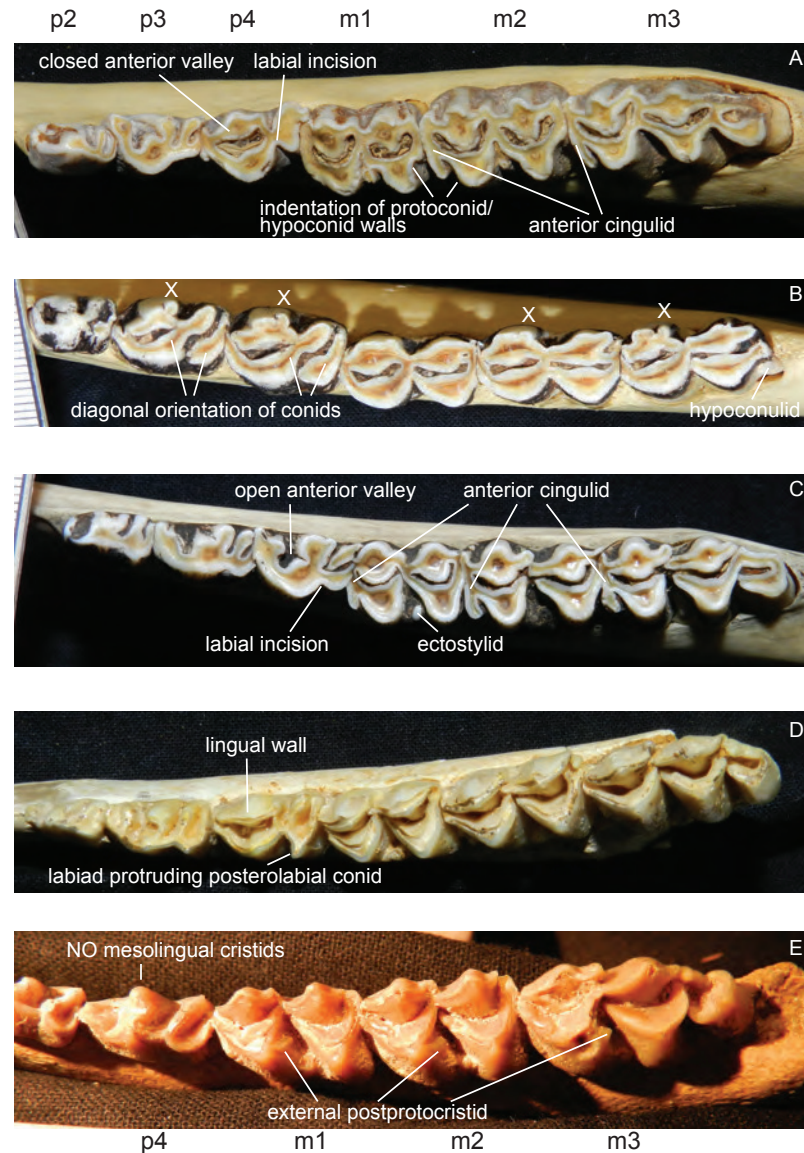
366 All lower molars have a similar morphology; m1 and m2 are two-lobed, m3 is three-lobed. The  
367 orientation of the lingual conids and cristids may be more diagonal in some species. Ectostylids are  
368 variably present on one to three molars. never high, nevertheless they become involved in wear in aged  
369 individuals (Fig. 8). In most Miocene cervids and in *Cervus australis* external postprotocristids are  
370 present on all molars (Fig. 8). Anterior cingulids are present in several species, usually more prominent  
371 on the anterior molar position(s). In *Rucervus* and *Rusa* the anterior cingulids are particularly prominent  
372 (Fig. 8). In *Rucervus* and also to a lesser extent in *Rusa* and *Axis* the anterior and posterior labial walls of





**Figure 7. Details of the upper dentition.** Close ups of the upper dentition of selected cervids showing the most striking features. (A) *Rucervus duvaucelii* (ZSM 1957-60), (B) *Rangifer tarandus* (ZSM 1959-211), (C) *Rucervus eldii* (UMZC H16194), (D) *Elaphurus davidianus* (UMZC H16235), (E) *Odocoileus hemionus* (ZSM 1971-720).

373 the lobes of the lower molars are indented (Fig. 8). The metastylids can be bent labiad in some species,  
 374 e.g., *Alces*. The third lobe on m3 is variable; most often the hypoconulid and entoconulid are connected  
 375 via the postento- and posthypoconulidcristids and form a crescent-shaped structure. Sometimes the third  
 376 lobe is reduced to one of these elements or has an additional fold on the posthypoconulidcristid. In a few



**Figure 8. Details of the lower dentition.** Close ups of the lower dentition of selected cervids showing the most striking features. (A) *Rucervus duvaucelii* (ZSM 1957-60), (B) *Rangifer tarandus* (ZSM 1959-211), (C) *Rucervus eldii* (UMZC H16194), (D) '*Cervus*' *philisi* (NMB St.V. 605), (E) *Procervulus* (MNH LRM 114).

377 individuals the third lobe is missing entirely. More details are in Heckeberg (2017a).

378 **Other dentition.** All Miocene cervids have enlarged upper canines, which are curved posteriad. From  
 379 the Pliocene onwards, the upper canines become reduced in size and are lost in some species. Extant  
 380 muntiacines have enlarged upper canines, similar to those of Miocene cervids. *Hydropotes* has strongly  
 381 elongated sabretooth-like upper canines, which differ in morphology from those in muntiacines and early  
 382 fossil cervids. In all other extant species upper canines are reduced in size or missing entirely. Most  
 383 cervines possess small upper canines. Adult capreolines rarely have upper canines, while most capreoline  
 384 juveniles have deciduous upper canines.

385 The lower incisors, i1–i3, have a simple spatulate morphology. The crown width decreases from i1 to  
 386 i3, i.e., i1 typically is distinctively broader than i2 and i3. Exceptions are *Alces*, *Hippocamelus*, and *Pudu*,



















**Figure 9. Variability of p4 in cervids.** This sequence of the lower left p4 shows different degrees of molarisation starting with an open anterior valley on the left, development of mesolingual cristids, connection of mesolingual cristids to other tooth elements, closing of the anterior valley, and re-arrangement of tooth elements with a diagonal orientation.

387 where i1 is only a little broader than i2. All lower canines in Cervidae are incisiviform. More details are  
388 in Heckeberg (2017a).

### 389 Phylogenetic Analyses

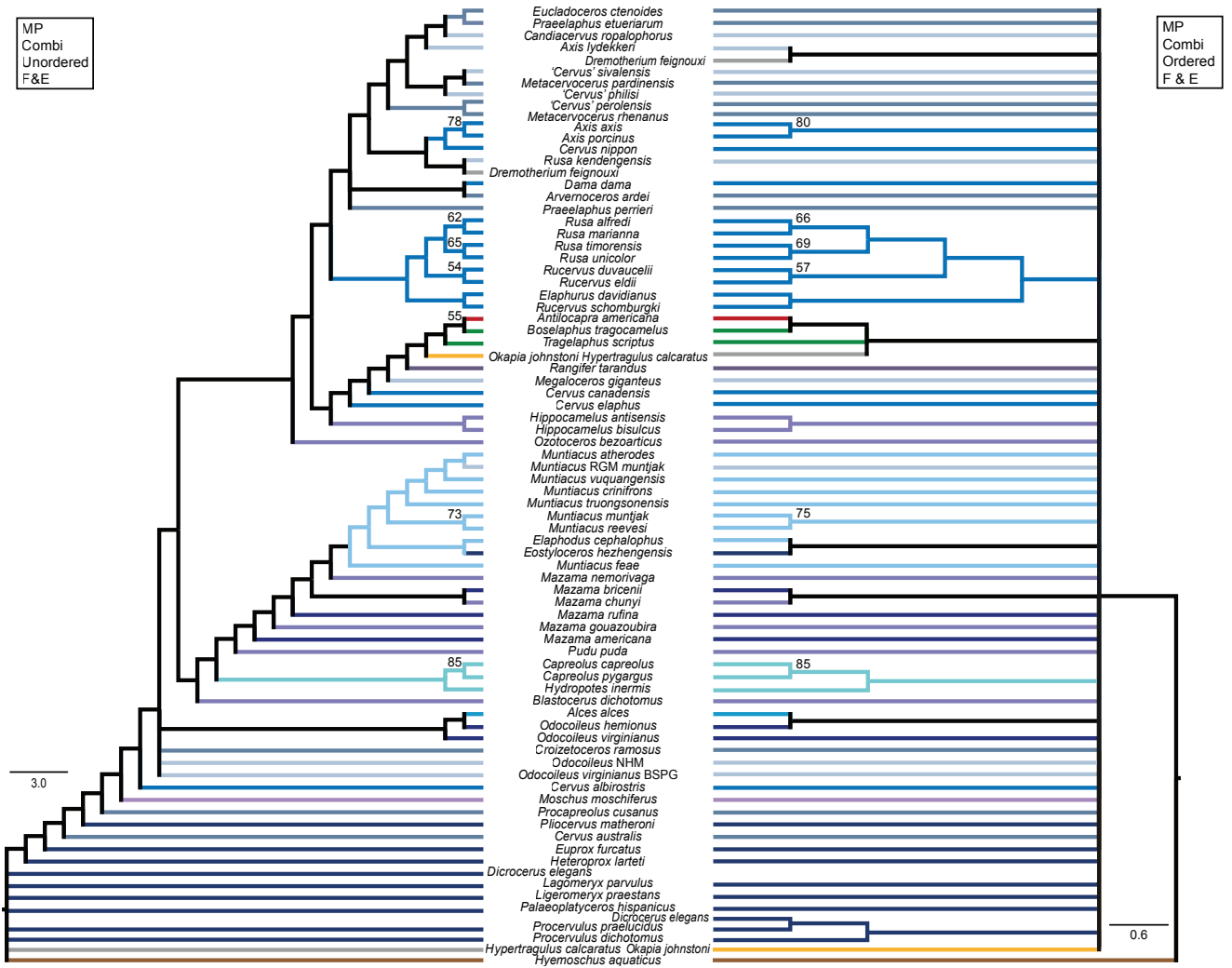
#### 390 Analyses of Morphological Data

391 **Stepping Stone Analyses.** In total, five stepping stone sampling analyses were undertaken; the first  
392 set of three analyses was used to determine the partitioning scheme, running one analysis with an  
393 unpartitioned, unordered data set with the  $\Gamma$  distribution, one with a minimal partitioning scheme, dividing  
394 the data set into a cranial and dental character set. The third data set was run with the maximal possible  
395 partitioning scheme, dividing the data set into upper post-canine dentition, lower post-canine dentition,  
396 other dentition, mandible, viscerocranium, neurocranium and antler characters. The fourth analysis was  
397 run with the unordered, unpartitioned data set, without the  $\Gamma$  distribution, and the fifth analysis was run  
398 with an ordered, unpartitioned data set with  $\Gamma$  distribution. The decision for one hypothesis is based on  
399 the Bayes Factor (BF). The results showed that the data set is best analysed unpartitioned, using the  $\Gamma$   
400 distribution and with character state ordering. However, BI and MP analyses were run unordered and  
401 ordered for each character set for comparison. See Table 1 for details. Figure 10 provides a key to the  
402 colour coding of the taxonomic groups.

	Muntiacini		Miocene cervids		Antilocapridae
	Cervini		Pliocene & Plio-/ Pleistocene cervids		Bovidae
	Capreolini		Pleistocene cervids		Giraffidae
	Alceini		Fossil outgroup		Moschidae
	Rangiferini				Tragulidae
	Blastocerina				
	Odocoileina				

**Figure 10. Colour code.** The colour code provides the key to taxonomic groups studied here and applies to all topologies within the present work.

403 **Standard Phylogenetic Analyses.** The MP topologies of the unordered and ordered morphological  
404 data set do not contradict each other (Fig. 11). The topology based on the unordered data set is more  
405 resolved. Both topologies support monophyletic Capreolini, a sister taxon relationship of *Axis axis* and



**Figure 11. Topologies from the morphological analyses.** The topologies of the maximum parsimony analyses of the combined morphological data set are shown. The left topology used the unordered data set, the topology on the right used character state ordering. Node support values are given as bootstrap support values.

406 *Axis porcinus* and *Muntiacus muntjak* and *Muntiacus reevesi*, and an *Elaphurus-Rucervus-Rusa*-clade-  
 407 clade. The *Elaphurus-Rucervus-Rusa*-clade was always recovered in the analyses based on the dental and  
 408 combined data set, in most topologies fully resolved. It consists of the *Rusa*-clade, which often has *Rusa*

409 *alfredi* as the sister taxon to the other three *Rusa*-species, of *Rucervus duvaucelii* and *Rucervus eldii* as  
410 the sister taxa to each other and to the *Rusa*-clade, and *Elaphurus davidianus* and *Rucervus schomburgki*  
411 as the sister taxa to each other and to the latter taxa. Cervini were never monophyletic in the analyses here  
412 based on the morphological data sets. The sister taxon relationships of *Rusa alfredi* and *Rusa marianna*  
413 and *Rusa timorensis* and *Rusa unicolor* are the only consistently recovered cervine clades in all topologies  
414 based on the cranial matrix. The higher hierarchical clades could not be recovered. The positions of  
415 *Dremotherium feignouxi*, *Okapia johnstoni*, *Hypertragulus calcaratus*, and *Dicrocerus elegans* differed in  
416 both topologies.

417 **Single Fossil Analyses (SFA).** Three different approaches to the single fossil analyses have been  
418 undertaken on three data sets including 31 fossil taxa each. This adds up to 93 analyses in total.  
419 Thirty-one analyses used the combined matrix of the complete mitochondrial genome and the combined  
420 morphological data set including outgroup taxa. Thirty-one analyses were undertaken using the same data  
421 set, but excluding five outgroup taxa. *Hyemoschus aquaticus* was used to root the topologies. Thirty-one  
422 analyses were undertaken with a constraint topology as a backbone; Capreolinae, Muntiacini and Cervini  
423 were constraint as monophyletic polytomous to each other. In each of the 93 analyses only one fossil  
424 taxon at the time was included. Figure 12 summarises the placements of all fossil cervid taxa in one  
425 topology. A detailed description of the analyses and the topologies of all analyses is in Heckeberg (2017a).

426 **Evolutionary Placement Algorithm (EPA).** The EPA analysis resulted in a resolved topology (Fig. 12).  
427 Cervinae, Cervini, Muntiacini, Capreolini, and Odocoileini were monophyletic. Many positions of fossil  
428 cervids were as expected from qualitative observations, e.g., those included in Cervini, whereas some  
429 were unexpected, e.g., the sister taxon position of Capreolini to Cervinae and placements of some fossil  
430 cervids, e.g., *Praeclaphus etueriarum*, *Megaloceros giganteus*, *Cervus australis*. Some Miocene cervids  
431 were included in Muntiacini, some were placed between the outgroup and all other cervids.

#### 432 **Analyses of Molecular Data**

433 **Nuclear Genes.** Although interpretations of the systematic relationships on genus and species level was  
434 difficult in the single gene topologies due to low taxon sampling and/or lack of resolution, the combined  
435 nuclear topology was well resolved and supports the higher hierarchical clades. The BI and the ML  
436 topologies were largely congruent (Fig. 13). There was no split into Odocoileina and Blastocerina as  
437 observed in the topologies based on the mitochondrial markers. The unexpected placement of *Capreolus*  
438 *capreolus* in this topology may be caused by the possibly contaminated *Sry* sequence of this species.

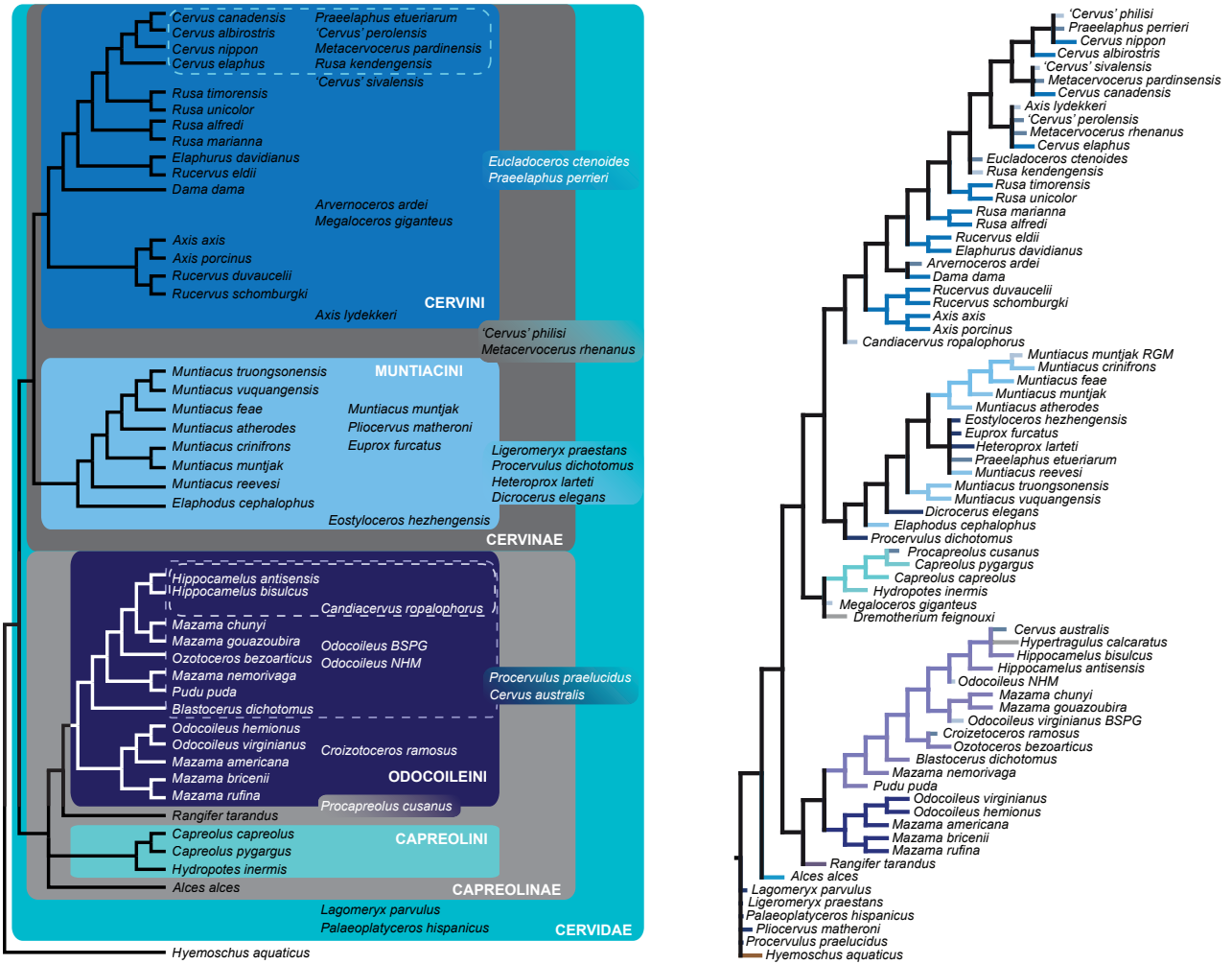
439 **Combined Mitochondrial Genes.** The BI topology of the combined mitochondrial analysis showed  
440 higher support values for the majority of nodes than the *Cytb* only topology, but lower support values for  
441 some nodes than for the mtG analysis. The ML topology differed in generally lower support values for most  
442 nodes, but was otherwise largely congruent (Fig. 13). The placement of non-cervid ruminants differed  
443 in both topologies. The main difference concerning cervid taxa is the position of *Pudu mephistophiles*  
444 (based on the correct *Cytb* sequence (Heckeberg et al., 2016)), which was the sister taxon to Blastocerina  
445 in the BI topology and the sister taxon to *Rangifer* and Odocoileini in the ML topology. This combined  
446 topology includes the polyphylies for *Rucervus*, *Hippocamelus*, *Odocoileus*, *Mazama*, and *Pudu*.

447 **Combined Molecular Analyses.** The BI and ML topologies of the combined nuclear and mitochondrial  
448 analyses were largely congruent, the support values were partly lower, particularly in the ML topology, in  
449 comparison to the topologies based on the mitochondrial markers (Fig. 13). Both topologies differed in  
450 the position of non-cervid ruminants, and the positions of *Alces alces* and *Pudu mephistophiles*, which  
451 remain uncertain. The split of Odocoileini into Blastocerina and Odocoileina was supported.

#### 452 **Combined Molecular and Morphological Analyses**

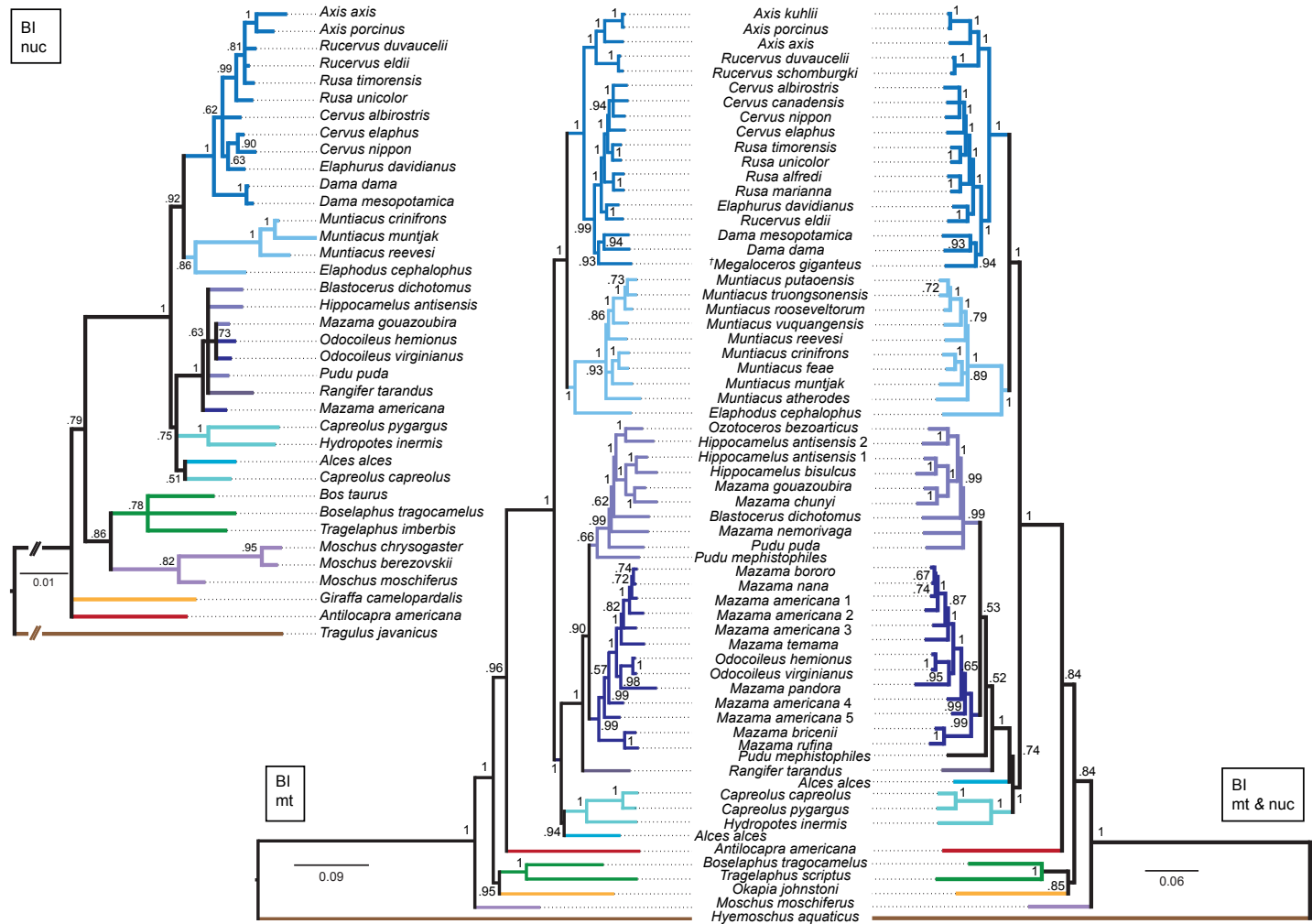
453 **Bayesian Inference.** The BI combined topology was largely unresolved (Fig. 14). Most extant cervids  
454 formed clades; the three *Axis* species and two *Rucervus* species formed a well supported clade. There was  
455 also an supported clade including eight Miocene cervids.

456 **Maximum Likelihood** In the ML combined topology the nodes were poorly or not at all supported (Fig.  
457 14). Some extant clades were recovered, e.g., Muntiacini, Odocoileina, Capreolini. Eight Miocene cervids  
458 formed a clade.

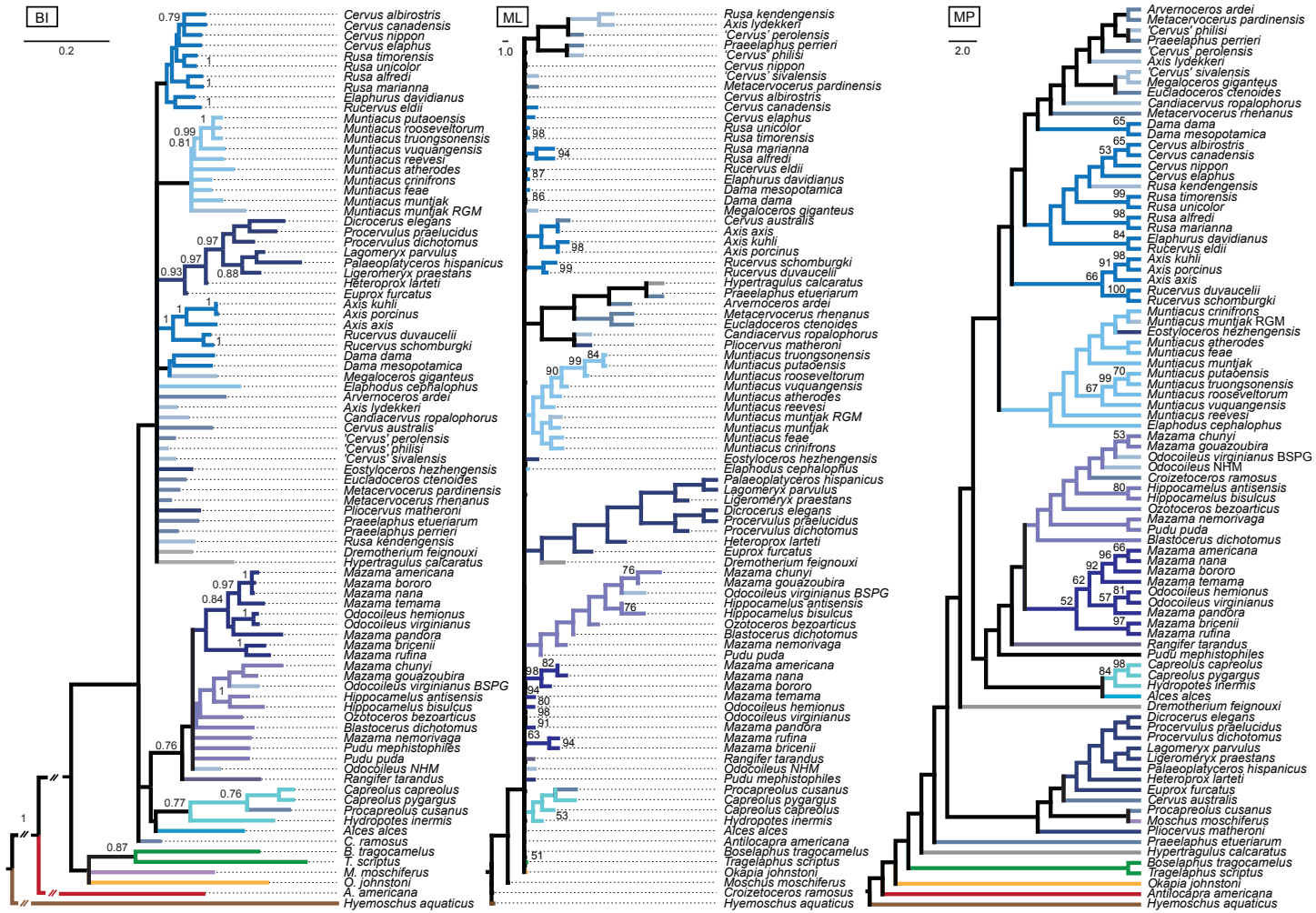


**Figure 12. Topologies from the SFA and EPA approaches.** The topology on the left summarises the systematic position of the fossils based on the SFA. The gradually shaded boxes indicate different observed positions. The topology on the right is the result of the EPA analysis.

459 **Maximum Parsimony** The nodes in the MP combined topology are largely unsupported (Fig. 14).  
 460 *Procervulus cusanus* was unexpectedly placed as the sister taxon to *Moschus*. Cervinae, Cervini,  
 461 Muntiacini, and Odocoileini form unsupported clades. Capreolini is a supported clade. All Miocene  
 462 cervids except for *Eostyloceros hezhengensis* form a clade.  
 463 Figure 15 qualitatively summarises the topologies from all analyses undertaken. The topology was  
 464 not generated by an analysis but was drawn to show the consensus of all topologies and which character  
 465 sets support the respective nodes.



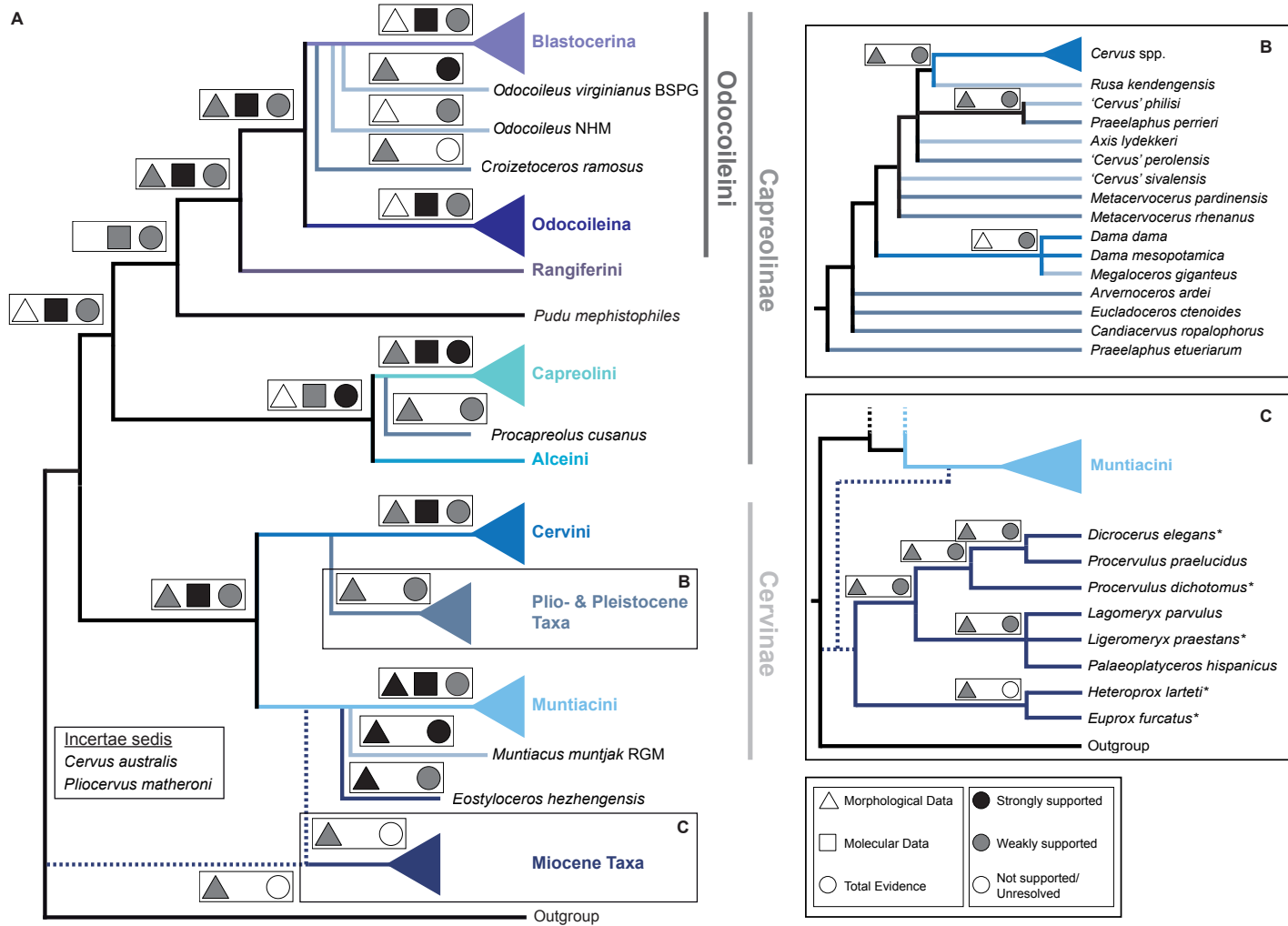
**Figure 13. Topologies resulting from the molecular data sets.** The topologies of the Bayesian inference analyses of the combined nuclear data set, the combined mitochondrial data set and the combined molecular data set (i.e., nuc + mt) are shown. Nuclear markers were available for fewer taxa than mitochondrial markers. The mitochondrial and molecular combined topologies are congruent except for the position of *Alces alces* and *Pudu mephistophiles*. Node support values are given as Bayesian posterior probabilities.



**Figure 14. Topologies from the combined molecular and morphological analyses.** The topologies of the combined molecular and morphological analyses using Bayesian inference, maximum likelihood and maximum parsimony (from left to right) are shown. Node support values are Bayesian posterior probabilities (BI) and bootstrap support values (ML, MP).



**Figure 15. The qualitative summary topology of all analyses is shown.** (A) represents the overview of the systematic relationships of higher cervid taxa including the positions of some fossil cervids. (B) shows the systematic relationship of several Plio- and Pleistocene cervids. (C) shows the systematic relationships of Miocene cervids.



466 **DISCUSSION**

467 For the first time, fossil and extant cervid species were combined in the so far most extensive data set  
468 including molecular and morphological data. Various data sets and partitions were analysed under different  
469 optimality criteria. In addition, the alternative approaches SFA and EPA were undertaken to investigate  
470 the systematic positions of fossils. The results provided new insight into the systematic relationships of  
471 fossil cervids and extant cervids. Many hypotheses about the systematic relationships of extant taxa could  
472 be confirmed; however, known controversies persisted, but could be specified in more detail. For most  
473 fossil cervids, we were able to find an affiliation to extant relatives, which has not been quantitatively  
474 tested previously.

475 Combining different data types helped to investigate the systematic relationships in detail and to  
476 reconstruct the evolutionary history of cervids. The initial separate analyses of the different data sets  
477 provided insights into the phylogenetic signal of the respective data. Some areas of the morphological  
478 topologies were congruent with the molecular topologies, some were not. However, the support of the  
479 morphological topologies did not contradict the molecular hypotheses.

480 Genotypic data partitions usually contain proportionally more characters than osteological data, which  
481 is assumed to be crucial for accuracy. On the other hand, osteological data partitions can be sampled for  
482 many more taxa, which partly cannot be sampled for molecular data (fossils) (O'Leary, 1999). Thus,  
483 morphological characters still have relevance in times of genomic analyses and serve as an independent  
484 test for molecular data, because of the relative distance between phenotype and genotype and different  
485 evolutionary dynamics of both types of data. Because selection targets on the phenotype, the resulting  
486 topology could potentially provide information on the selective history of taxa (Lee and Camens, 2009;  
487 Groves, 2014). If the same topology is supported by different data sources and reflects biological evidence  
488 at all scales (principle of consilience), it is more likely that the topology is 'correct' (Bibi et al., 2012).

489 The challenges of the data sets here were the high levels of homoplasy (particularly in the morphology)  
490 and the rapid radiations of ruminant tribes. Consensus might be difficult to achieve, because short branch  
491 lengths and/or lack of resolution potentially represent a genuine rapid diversification of clades, which  
492 may not be further solved just by increasing the sequence length or the taxon sampling. Markers that are  
493 less influenced by convergent evolution, such as rare genomic changes or cytogenomics may be useful  
494 additions in the future (Rokas and Holland, 2000; Price et al., 2005; Hernández Fernández and Vrba,  
495 2005).

496 **Models of evolution.** So far, there is no appropriate evolutionary model for morphological characters  
497 in model-based approaches such as BI and ML (O'Reilly et al., 2016). The only model of morphological  
498 evolution, which is widely used in model-based phylogenetic algorithms (BI, ML), is the Markov k (Mk)  
499 model by Lewis (2001). It is not fully understood how the standard models of molecular evolution (e.g.,  
500 HKY, GTR) translate variable rate frequencies and substitution rates to morphological data (Spencer and  
501 Wilberg, 2013). Although topologies from model-based approaches, particularly ML, are typically better  
502 resolved than strict consensus topologies from parsimony analysis, the better resolution is not necessarily  
503 meaningful. The apparent better resolution may simply be a result of an incorrect model of morphological  
504 evolution (Spencer and Wilberg, 2013).

505 **Comparison of mitochondrial vs. nuclear vs. total evidence topologies.** Previous studies demon-  
506 strated that combining mitochondrial and nuclear markers increases robustness of higher hierarchical  
507 cervid clades (Randi et al., 1998). The topologies resulting from nuclear markers often agree with  
508 morphology, but often contradict topologies resulting from mitochondrial markers (Bibi, 2014). There are  
509 few phylogenetic reconstructions for cervids based on nuclear markers (Cronin et al., 1996; Gilbert et al.,  
510 2006). Analyses of nuclear markers have the potential to characterise the distribution of genetic variation  
511 (Balakrishnan et al., 2003). Combining and interpreting nuclear and mitochondrial markers can help to  
512 uncover recent hybridisation events, as in *Elaphurus davidianus*, which takes up different positions when  
513 analysed with mitochondrial markers compared to nuclear markers (Fig. 13).

514 The nuclear topologies here, were largely congruent to those published previously. Incorporating  
515 more nuclear DNA is crucial to test relationships in ruminant systematics based on mitochondrial DNA  
516 and should be sequenced for a broader range of taxa than is available to date.

## 517 **Miocene Cervids**

518 The earliest cervids are from the mid early Miocene (MN3) represented by *Procervulus*, *Ligeromeryx*, and  
519 *Acteocemas* and became more numerous and widely distributed during the Miocene. In the late early and  
520 early middle Miocene *Stephanocemas*, *Heteroprox*, *Lagomeryx*, and *Dicrocerus* appeared (Ginsburg and  
521 Azanza, 1991; Dong, 1993). A low cervid diversity is assumed during the late Miocene and all typical  
522 Miocene cervids became extinct before the late Miocene (Ginsburg and Azanza, 1991; Böhme et al.,  
523 2012).

524 It was suggested to put *Lagomeryx*, *Procervulus*, *Heteroprox*, *Euprox*, *Dicrocerus*, *Stephanocemas*  
525 into a subfamily as a 'primitive' clade within Cervidae (Azanza, 1993b; Ginsburg, 1985; Rössner, 1995).  
526 Miocene cervids were usually considered to be distant from crown cervids representing a distinct group of  
527 stem cervids. They were subdivided into Lagomerycinae (-dae), Procervulinae (-dae) and Dicrocerinae  
528 (-ini). All of them were regarded as sister clades to Cervidae (Mennecart et al., 2016). It was suggested  
529 that *Lagomeryx*, *Ligeromeryx*, and *Paradicrocerus* form the lagomerycids, *Heteroprox* and *Procervulus*  
530 form the procervulines, and *Acteocemas*, *Stehlinoceros*, and *Dicrocerus* form the dicrocerines (Gentry  
531 et al., 1999). In none of the analyses here this split into three groups was distinctive. So far, not many  
532 attempts to reconstruct the phylogeny of Miocene cervids have been made (Azanza Asensio, 2000).  
533 Recently, Mennecart et al. (2016, 2017) presented the first phylogenetic analyses based on inner ear  
534 characters for several fossil cervids.

535 In the phylogenetic analyses here, Miocene cervids were most often placed either between the outgroup  
536 and all other cervids, mostly unresolved; sometimes a few taxa formed a clade. The placement between  
537 the outgroup and other cervids was expected from their temporal distribution and their shared higher  
538 proportion of plesiomorphic characters. The systematic relationships within early Miocene cervids have  
539 been and still are controversial. (Rössner, 1995; Azanza et al., 2013).

540 ***Lagomeryx parvulus* and *Ligeromeryx praestans*.** Qualitative morphological comparisons, espe-  
541 cially on antler morphology, suggest that *Lagomeryx parvulus* and *Ligeromeryx praestans* are closely  
542 related to each other. Only one analysis (cranial data set) here supports the sister taxon relationship  
543 of these two taxa. Therefore, a subfamily Lagomerycinae would be justified based on morphological  
544 qualitative comparisons, but is not supported in the topologies. Data completeness or presence of specific  
545 characters that are absent in the other taxon could be the reasons. Also, whether lagomerycids form a  
546 family as the sister taxon to Cervidae could not be entirely ruled out, but the tendency of *Ligeromeryx*,  
547 *Lagomeryx*, and *Palaeoplatyceros* to form a clade within a clade consisting of Miocene taxa indicated that  
548 lagomerycids potentially form a subfamily of Cervidae in a stem position.

549 The systematic position of lagomerycids, has always been controversial. They have been considered  
550 as a family between Giraffidae and Cervidae (Teilhard de Chardin, 1939), as part of the superfamily  
551 Cervoidea (Romer, 1966; Viret, 1961; Young, 1964), as a separate subfamily within Cervidae (Vislobokova  
552 et al., 1989), as a family of aberrant giraffoids, as a junior synonym of Palaeomerycidae (Pilgrim, 1941;  
553 Simpson, 1945; Young, 1964), as junior synonym of Muntiacini/-ae (Chow and Shih, 1978), as more  
554 closely related to Antilocapridae (Ginsburg, 1985; Solounias, 1988), or as representing an entirely  
555 independent clade (Bubenik and Bubenik, 1986; Azanza, 1993b; Azanza and Ginsburg, 1997).

556 The discussions on the taxon in the literature and the new insights resulting from the analyses here  
557 clearly show that the systematic position of Lagomerycidae represents one of the most controversial of  
558 ruminant families, so far without unambiguous consensus; however, cranial and postcranial morphology  
559 support the affiliation as stem Cervidae (Chow and Shih, 1978; Leinders and Heintz, 1980; Vislobokova  
560 et al., 1989; Azanza and Ginsburg, 1997; Mennecart et al., 2017).

561 ***Procervulus dichotomus* and *Procervulus praelucidus*.** In most analyses here, *Procervulus* was  
562 placed in a stem position and *Procervulus* and *Dicrocerus* were more closely related to each other than  
563 to other cervids. A sister taxon relationship of *Procervulus* and *Heteroprox* was not observed. In the  
564 combined morphological and TE analyses, a close relationship of *Procervulus dichotomus* and *Procervulus*  
565 *praelucidus* to *Dicrocerus elegans* was confirmed.

566 *Procervulus* was assumed to be the Miocene descendant of *Amphitragulus* and *Dremotherium* (Gentry,  
567 1994; Rössner, 1995). Presumably, transitional forms existed, which were not documented in the fossil  
568 record (Rössner, 1995). *Procervulus* has often been hypothesised to be the sister taxon to all other cervids  
569 (Janis and Scott, 1987; Groves, 2007). In previous studies, *Procervulus* was placed as the sister taxon to  
570 *Heteroprox* Azanza Asensio (2000); Mennecart et al. (2016) and both were the sister taxon to the clade

571 containing *Dicrocerus elegans*. In Mennecart et al. (2017) *Procervulus dichotomus* was the sister taxon to  
572 *Heteroprox larteti* and *Procervulus praelucidus* the sister taxon to both of them; this clade was placed  
573 between *Lagomeryx parvulus* and all other cervids, which is similar to our results.

574 ***Heteroprox larteti*.** In the analyses here, *Heteroprox larteti* was most often placed in an unresolved  
575 position, between the outgroup and cervids, as the sister taxon to *Euprox furcatus* or *Dicrocerus elegans*,  
576 or in a clade with other Miocene taxa (morphology, TE). Some topologies indicated a potential closer  
577 relationship to Muntiacini based on apomorphic characters, similar to *Euprox furcatus*.

578 *Heteroprox* was assumed to be the descendant of *Procervulus* (Rössner, 1995). In Azanza Asensio  
579 (2000) *Heteroprox* was most often placed as the sister taxon to *Procervulus* or as an (unresolved) stem  
580 lineage. Similarly, in Mennecart et al. (2017) *Heteroprox larteti* was the sister taxon to *Procervulus*  
581 *dichotomus*.

582 ***Dicrocerus elegans*.** In the analyses here, *Dicrocerus elegans* was most often placed closely related to  
583 *Procervulus*, sometimes as the sister taxon to *Heteroprox larteti*, or between the outgroup and cervids.  
584 Based on our results and discussions in the literature, *Dicrocerus* is most certainly a stem cervid with  
585 affinities primarily to *Procervulus* and secondarily to other Miocene cervids. In a few analyses a potentially  
586 closer relationship to Muntiacini was observed.

587 Azanza et al. (2011) suggested that *Dicrocerus* is a transitional form between the Procervulinae and  
588 crown Cervidae, which had also been hypothesised by Vislobokova (1990). In Azanza Asensio (2000)  
589 *Dicrocerus elegans* was placed as the sister taxon to *Acteocemas* and *Stehlinoceros* (= *Paradicrocercus*) and  
590 this clade was the sister taxon to all burr-bearing antlered cervids. In Mennecart et al. (2017) *Dicrocerus*  
591 *elegans* was the sister taxon to *Eostyloceros hezhengensis* in a sister taxon position to the crown cervids.

592 ***Euprox furcatus*.** In the TE analyses here, *Euprox furcatus* was most often placed in an unresolved  
593 position or as the sister taxon to *Heteroprox larteti*; in the TE analyses it was placed in a clade with other  
594 Miocene cervids. The results indicate that *Euprox furcatus* shares characters with other Miocene cervids,  
595 but also already had apomorphic characters, which imply a closer relationship to extant Muntiacini than  
596 to other crown cervids.

597 It was suggested that modern *Muntiacus* and fossil muntiacines such as *Eostyloceros*, *Metacervulus*,  
598 and *Paracervulus* diverged from *Euprox* (Vislobokova, 1990; Croitor, 2014). *Euprox* was the first cervid  
599 with burr-bearing antlers and a pedicle inclination similar to that of muntjacs. Therefore, it has been  
600 suggested in several studies that *Euprox* may be the earliest representative of crown cervids (Azanza,  
601 1993b; Gentry et al., 1999; Dong, 2007; Azanza et al., 2013; Mennecart et al., 2016, 2017). It was often  
602 considered as a member of Muntiacini, which would imply that Muntiacini is the sister taxon to all other  
603 cervids. In Azanza Asensio (2000), *Euprox* is variably placed closely related to *Amphiprox*, to extant  
604 *Muntiacus* and *Elaphodus*, to *Eostyloceros*, or to *Metacervulus*, or as the sister taxon to a clade containing  
605 all five of the above species or a subset thereof. In Mennecart et al. (2016), *Euprox furcatus* was placed  
606 as the sister taxon to *Cervus elaphus*. They further stated that *Dicrocerus elegans*, *Euprox furcatus*, and  
607 *Cervus elaphus* differ from the other Miocene cervids, i.e., Procervulinae, in certain inner ear characters;  
608 *Euprox furcatus* had the most derived characters among them. In Mennecart et al. (2017) *Euprox furcatus*  
609 was placed as the sister taxon to all crown cervids.

610 There is a large temporal gap in the early putative fossil muntjac-like cervid lineage between the  
611 first representatives, *Euprox*, and the presumed direct ancestors of muntiacines, e.g., *Eostyloceros*  
612 (Azanza Asensio and Menendez, 1989; Azanza, 1993b), and additionally an even larger gap between  
613 those early fossils and the first members of extant *Muntiacus*, which appear in the Pleistocene. For more  
614 certainty of the systematic relationships it would be crucial to find more fossil material that would link the  
615 early presumed muntiacines with the crown muntiacines.

616 ***Palaeoplatyceros hispanicus*.** In most analyses here *Palaeoplatyceros hispanicus* was placed between  
617 the outgroup and cervids, as the sister taxon to *Lagomeryx parvulus* or as the sister taxon to most other  
618 Miocene taxa. *Palaeoplatyceros* is highly incomplete and has a combination of plesiomorphic traits and  
619 apomorphic traits, such as 'presence of a burr'.

620 *Palaeoplatyceros hispanicus* can be distinguished from all other contemporaneous cervid species  
621 based on the palmation of antlers; however, its systematic position is problematic (Azanza Asensio, 2000).  
622 In Azanza Asensio (2000), *Palaeoplatyceros* was mostly placed as the sister taxon to all other cervids  
623 with burr-bearing antlers. Unless more material becomes available, its systematic position will remain

624 controversial. Based on the analyses here, *Palaeoplatyceros* is likely a stem cervid with burr-bearing  
625 antlers.

626 ***Pliocervus matheronis*.** *Pliocervus matheronis* is known from the Messinian (upper Turolian, MN13).  
627 In the analyses here, *Pliocervus matheronis* was most often placed in an unresolved position, mostly  
628 between the outgroup and cervids and sometimes related to other Miocene taxa.

629 Although Simpson (1945) included Pliocervinae, comprising *Cervocerus*, *Cervavitus*, *Procervus*, and  
630 *Pliocervus*, which were regarded as the immediate crown Cervini precursors (Gentry, 1994; Groves, 2007),  
631 in Cervinae, others could not find any phylogenetic relationship of *Pliocervus* with Cervini/Cervinae  
632 (Petronio et al., 2007). Gentry et al. (1999) placed *Cervavitus* and *Pliocervus* among Cervoidea, whereas  
633 Azanza and Montoya (1995) and Azanza Asensio (2000) classified *Pliocervus* as Cervinae. It was  
634 suggested to be closely related to the holometacarpal *Cervavitus* within Pliocervini, which was included  
635 in Cervinae (Czyżewska, 1968; Vislobokova, 1990; Azanza Asensio, 2000).

636 The high morphological similarity of *Pliocervus matheronis* to the late Miocene *Pavlodaria orlovi*  
637 implies that these two genera could be closely related or possibly even synonymous. It was suggested  
638 that the subfamily Pliocervinae Symeonidis 1974, containing *Pliocervus* and *Pavlodaria* is a synonym of  
639 Capreolinae. In Azanza Asensio (2000) *Pliocervus matheronis* was variably placed and seems to have the  
640 highest proportion of apomorphic characters compared to other Miocene cervids. In most recent studies  
641 *Pliocervus* was regarded as incertae sedis (Croitor, 2014).

642 A definite morphological characterisation of *Pliocervus* is still missing and its systematic position  
643 remains controversial (Godina et al., 1962; Czyżewska, 1968; Korotkevich, 1970; Azanza Asensio, 2000;  
644 Petronio et al., 2007; Croitor, 2014). More and new morphological and biometric data are needed to solve  
645 the systematic relationships of 'pliocervines' (Di Stefano and Petronio, 2002).

646 ***Eostyloceros hezhengensis*.** *Eostyloceros hezhengensis* from the late Miocene of China was used for  
647 scoring characters (Deng et al., 2014). In the analyses here, *Eostyloceros hezhengensis* was most often  
648 placed in an unresolved position or within Muntiacini, suggesting that it is probably more closely related  
649 to muntjacs than to other cervids, which would support results from comparative morphology. Thus,  
650 *Eostyloceros hezhengensis* can be considered as a direct ancestor of muntjacs.

651 *Euprox* is considered as the direct ancestor of *Eostyloceros*, *Metacervulus*, and *Paracervulus*; after a  
652 change from subtropical to more temperate climate and *Euprox*-like cervids were replaced by represen-  
653 tatives of *Eostyloceros* (Azanza Asensio and Menendez, 1989; Azanza, 1993b; Pitra et al., 2004). This  
654 lineage is assumed to lead to extant *Muntiacus* (Vislobokova, 1990; Croitor, 2014). In Azanza Asensio  
655 (2000), *Eostyloceros* was always closely related to *Muntiacus* and *Metacervulus*, while in Mennecart et al.  
656 (2017) *Eostyloceros hezhengensis* was not placed within Muntiacini but was the sister taxon to *Dicrocerus*  
657 *elegans*.

#### 658 **Pliocene and Plio-Pleistocene Cervids**

659 There is no generally accepted classification of Plio- and Plio-Pleistocene cervids Pfeiffer (1999); how-  
660 ever, for Villafranchian cervids (MN16) the following classifications were suggested: *Croizetoceros*  
661 *ramosus*, *Metacervoceros pardinensis*, '*Cervus*' *philisi*, '*Cervus*' *perolensis*, *Eucladoceros ctenoides* were  
662 considered as Cervini, *Arvernoceros ardei* as Megacerini, and *Libralces gallicus* (not included here) and  
663 *Procapreolus cusanus* were considered as Capreolinae.

664 In most morphological topologies here, Plio- and Pleistocene cervids were placed within crown  
665 cervids, sometimes forming subclades. Some Plio- and Pleistocene cervids were placed more closely  
666 related to extant Cervini. Most of them were nested in a clade together with Pleistocene cervids. In a  
667 few topologies the majority of Pliocene cervids were in an unresolved sister taxon position to all other  
668 Cervinae.

669 ***Cervus australis*.** In the phylogenetic analyses here, *Cervus australis* was most often placed in an  
670 unresolved position, sometimes closer to Muntiacini than to other cervids; it was also placed between  
671 the outgroup and cervids, as the sister taxon to *Eostyloceros hezhengensis* and *Praeclaphus etueriarum*,  
672 to *Hippocamelus bisulcus*, or *Muntiacus muntjak*. Based on qualitative morphological comparisons it is  
673 most likely a stem cervid, potentially closer to Muntiacini.

674 This species was originally described by De Serres (1832) and all known specimens are from  
675 Montpellier, France (Gervais, 1852; Czyżewska, 1959). Little further information is available in the  
676 literature concerning this species. Many entries point to muntiacines, e.g., *Paracervulus australis* (Gentry,

677 2005); however, there are no obvious similarities to muntiacines in the investigated specimens. Croitor  
678 (2018) also confirms an incertae sedis status for this cervid based on comparative morphology. Thus, the  
679 systematic position of *Cervus australis* remains uncertain.

680 ***Arvernoceros ardei*.** In our analyses *Arvernoceros ardei* was placed in an unresolved position, often  
681 close to or within Cervini. In some topologies it was placed as the sister taxon to *Metacervocerus*  
682 *pardinensis*, *Praeclaphus perrieri*, and *Metacervocerus rhenanus*. It was placed as the sister taxon to  
683 *Dama dama* in several topologies.

684 *Arvernoceros* was part of the first radiation of Cervinae/-i together with *Metacervocerus*, *Praeclaphus*,  
685 *Axis*, and *Rucervus* (Croitor, 2014). The systematic position of *Arvernoceros ardei* has been subject to  
686 speculation for decades, its definition is still incomplete and affinities to other cervids unclear. Depéret  
687 (1884) found similarity to *Axis*, but no affiliation to *Dama*; it was suggested that it is most similar to  
688 Megacerini (Heintz, 1970; Vislobokova, 1990, 2012). *Arvernoceros ardei* was considered to be closely  
689 related to modern *Elaphurus* (Teilhard de Chardin and Piveteau, 1930), declared as incertae sedis genus  
690 by (Lister, 1987), closely related to *Axis* Di Stefano and Petronio (2002), closely related to *Rucervus*  
691 (Croitor, 2009, 2018). Despite some uncertainties in the morphological analyses, a closer relationship to  
692 *Dama dama* than to other cervids was suggested here.

693 ***Croizetoceros ramosus*.** In most of the analyses here, *Croizetoceros ramosus* was placed in an un-  
694 resolved position; it was sometimes the sister taxon to *Procapreolus cusanus*, *Alces alces*, *Ozotoceros*  
695 *bezoarticus*, or *Odocoileus*. Our results suggest a placement within Capreolinae and most likely within  
696 Odocoileini.

697 The antler morphology of *Croizetoceros ramosus* does not share similarities with any extant cervid  
698 species or with other cervid species from the Villafranchian (Heintz, 1970). Unfortunately, there is not  
699 much known about its skull morphology (Croitor, 2014). In Mennecart et al. (2017) *Croizetoceros* was  
700 placed as the sister taxon to Capreolinae.

701 ***'Cervus' perolensis*.** In the analyses here, *'Cervus' perolensis* was placed in an unresolved position  
702 and as the sister taxon to several cervine taxa. Repeated placements within Cervini suggest that *'Cervus'*  
703 *perolensis* almost certainly belongs to Cervini and is likely closely related to and/or an ancestor of *Cervus*.

704 *'Cervus' perolensis*, *Metacervocerus rhenanus*, and *'Cervus' philisi* were found to be similar to each  
705 other and *'Cervus' perolensis* and *Metacervocerus pardinensis* were classified as *Pseudodama* Azzaroli  
706 (1953); Azzaroli and Mazza (1992a). Later, *'Cervus' perolensis* was considered as a descendant of  
707 *'Cervus' philisi* by Stefaniak and Stefaniak (1995). Spaan (1992), however, concluded that *'Cervus' philisi*  
708 and *'Cervus' perolensis* are junior synonyms of *Metacervocerus rhenanus* and should be renamed as such,  
709 which was supported by Pfeiffer (1999). If this were true, *'Cervus' philisi* and *'Cervus' perolensis* should  
710 come out in a similar systematic position as *Metacervocerus rhenanus*.

711 ***Procapreolus cusanus*.** In the analyses here, *Procapreolus cusanus* was placed between the outgroup  
712 and cervids, within Capreolinae, sometimes within Odocoileini, and as the sister taxon to both *Capreolus*.  
713 Thus, *Procapreolus cusanus* most likely belongs to Capreolinae and the previously suggested close  
714 relationship to *Capreolus* was confirmed in some analyses.

715 Despite the widely accepted assumption that *Procapreolus cusanus* is closely related to or even a direct  
716 ancestor of *Capreolus*, the origin of *Capreolus* within *Procapreolus* is still under debate (Lechner-Doll  
717 et al., 2001). Some authors hypothesise that it may be assigned to *Capreolus* rather than *Procapreolus*  
718 (Valli, 2010). Others place it in an intermediate position between lower Pliocene and Pleistocene  
719 *Procapreolus* species and extant *Capreolus* (Czyżewska, 1968; Heintz, 1970; Lechner-Doll et al., 2001).

720 ***Metacervocerus pardinensis*.** In the analyses here, *Metacervocerus pardinensis* was most often closely  
721 related to or within Cervini, which suggests that *Metacervocerus pardinensis* is a member of Cervini and  
722 probably a close relative and/or ancestor of *Cervus*.

723 The temporal distribution of *Metacervocerus pardinensis* suggests that it could be an ancestor of  
724 *'Cervus' philisi*. *Metacervocerus pardinensis* and *Metacervocerus rhenanus* have enough morphological  
725 differences to justify two different species (Spaan, 1992). Dietrich (1938) proposed that *Metacervocerus*  
726 *pardinensis* is synonymous with *etueriarum*, *perrieri*, *issiodorensis*, and *rhenanus*. Based on similarities  
727 to *Rusa* deer, the genus *Metacervoceros* was erected to represent European rusine deer (Croitor, 2006a).  
728 However, their systematic position remained controversial. *Metacervocerus pardinensis* was classified as  
729 *Pseudodama* by Azzaroli and Mazza (1992a), while De Vos and Reumer (1995) assigned *Metacervocerus*

730 *pardinensis* and *Metacervocerus rhenanus* to *Cervus*, Pfeiffer (1999) to *Dama*, and Di Stefano and  
731 Petronio (2002) to *Rusa*. Differences in the skull morphology suggest that *Metacervocerus* does not  
732 belong to the *Cervus-Rusa* evolutionary lineage, which needs stronger evidence from the fossil record.  
733 Croitor (2014) suggested it is more likely that *Metacervocerus pardinensis* represents an ancestor of  
734 *Dama*.

735 ***Praeelaphus perrieri*.** In the analyses here, *Praeelaphus perrieri* was placed close to or within Cervini,  
736 which suggests that *Praeelaphus perrieri* is a member of Cervini and probably closely related to and/or  
737 the ancestor of *Cervus*.

738 The teeth and postcranial material from *Praeelaphus perrieri* and *Eucladoceros* are indistinguishable;  
739 however, *Praeelaphus perrieri* and *Eucladoceros ctenoides* do not coexist in any of the known localities,  
740 although they occupy the same niches. The systematic relationships remained uncertain (Croitor, 2014).  
741 Already Portis (1920) proposed a new subgenus *Praeelaphus* for '*Cervus*' *perrieri*, as well as for *C.*  
742 *avernensis*, *C. etueriarum* from the early Villafranchian (Croitor, 2014). *Praeelaphus perrieri* was  
743 considered as the earliest representative of *Cervus* in Europe by Di Stefano and Petronio (2002), however,  
744 even though it is an early cervine, there is no clear evidence that it is directly related to *Cervus* and it more  
745 likely represents an extinct lineage within the early cervine evolution (Croitor, 2014).

746 ***Praeelaphus etueriarum*.** In the analyses here, *Praeelaphus etueriarum* was placed between *Eosty-*  
747 *loceros hezhengensis* and *Eucladoceros ctenoides*, as the sister taxon to *Metacervocerus rhenanus*,  
748 *Eostyloceros hezhengensis*, or *Eucladoceros ctenoides*. Placements as the sister taxon to the *Cervus*-clade  
749 and within Muntiacini suggest that *Praeelaphus etueriarum* belongs to Cervinae and most likely to  
750 Cervini.

751 There is consensus that *Praeelaphus* is a member of the early radiation of Cervini and *perrieri*,  
752 *warthae*, and *lyra* may be synonyms as they represent similar and contemporaneous cervids (see above)  
753 (Croitor, 2014). Heintz (1970) suggested that *Praeelaphus etueriarum* was established based on a juvenile  
754 *Praeelaphus perrieri*, which is yet to be proven.

755 ***Eucladoceros ctenoides*.** Here, *Eucladoceros ctenoides* was most often placed within Cervinae and/or  
756 Cervini, which also indicate a potentially close relationship to *Cervus*.

757 Most of the previously defined *Eucladoceros* species were synonymised with *Eucladoceros ctenoides*  
758 (Azzaroli and Mazza, 1992a; De Vos and Reumer, 1995; Pfeiffer, 1999; Croitor and Bonifay, 2001; Valli  
759 and Palombo, 2005). '*E. senezensis*' has been suggested to be an ancestor of *Megaceroides* or *Megaloceros*  
760 *giganteus* in particular (Azzaroli and Mazza, 1992a,b; Kuehn et al., 2005). Pfeiffer (2002) proposed that  
761 *Eucladoceros*, *Megaloceros*, and *Cervus* form a group. Flerov (1952) suggested that *Eucladoceros* is an  
762 ancestor of *Alces*, which is not supported by others (Heintz, 1970; Croitor, 2014). The comb-shaped antler  
763 morphology is unique and more similar to *Cervus elaphus* or *Cervus albirostris* than to any other living  
764 cervid (pers. obs.). Because upper canines in *Eucladoceros ctenoides* are absent it was interpreted that  
765 the genus most likely does not belong to the *Cervus-Rusa*-lineage (Croitor, 2014); instead, *Eucladoceros*  
766 *ctenoides* was hypothesised as a descendant of an early three-tined ancestor of *Axis* or *Metacervocerus*  
767 (Croitor, 2014). In Mennecart et al. (2017) *Eucladoceros ctenoides* was placed as the sister taxon to the  
768 *Cervus-Rusa*-clade, which confirms the results from our analyses.

769 ***Metacervocerus rhenanus*.** In the analyses here, *Metacervocerus rhenanus* was mostly placed as the  
770 sister taxon to Cervini and/or within Cervinae, which suggests that *Metacervocerus rhenanus* is a member  
771 of Cervini and potentially is either a close relative and/or ancestor of *Cervus* or *Axis*.

772 The genus *Metacervocerus* was established by Dubois (1904) as *Cervus (Axis) rhenanus* for the  
773 small sized deer from Tegelen. Spaan (1992) synonymised '*Cervus*' *philisi* from Senèze with '*C.*  
774 *rhenanus* based on dentition and antler morphology. Croitor and Bonifay (2001) assigned it to the  
775 genus *Metacervocerus*. Several three-tined cervids were described from the early Pleistocene of Europe  
776 (De Vos and Reumer, 1995); *Metacervocerus rhenanus* was considered to include '*C.* *philisi*', '*C.*  
777 *perolensis*', *C. ischnoceros*, and *Pseudodama lyra* and '*Cervus*' *philisi* was suggested to be a junior  
778 synonym of *Metacervocerus rhenanus* (Azzaroli et al., 1988; Spaan, 1992). *Metacervocerus rhenanus*  
779 was hypothesised to be an ancestor of *Dama dama* (Pfeiffer, 1999; Di Stefano and Petronio, 2002);  
780 however, this hypothesis was ruled out by the coexistence of both genera in the early Pleistocene (Croitor,  
781 2014).

782 From the analyses based on the present data sets, the synonymy of '*Cervus philisi*' and '*Cervus*'  
783 *perolensis* with *Metacervocerus rhenanus* could not be confirmed. All analyses placed the three taxa  
784 differently and not closely related to each other. This may be caused by the differing availability of  
785 characters for each taxon and should be tested based on exclusively overlapping characters.

### 786 Pleistocene Cervids

787 In the early Pleistocene, Pliocene forms were successively replaced by more modern cervids. By the  
788 middle Pleistocene, most Pliocene and some early Pleistocene cervids became extinct, while extant  
789 representatives appeared (Dong, 1993).

790 Pleistocene cervids are more similar to extant forms. In the morphological topologies, similarly to the  
791 Plio- and Plio-/Pleistocene cervids, the Pleistocene cervids were distributed across crown group clades,  
792 sometimes forming subclades. The majority of Pleistocene cervids were placed within Cervini.

793 '***Cervus philisi***. In the analyses here, '*Cervus philisi*' was most often placed within Cervinae or Cervini  
794 sometimes within the extant *Cervus*-clade, which suggests that '*Cervus philisi*' belongs to Cervini with a  
795 potentially closer relationship to *Cervus*. The results further support previous findings that '*Cervus philisi*'  
796 cannot be assigned to any extant cervid (except maybe *Cervus nippon*). '*Cervus philisi*' together with  
797 *Praeelaphus perrieri* potentially represents an extinct clade leading to *Cervus*. The suggested synonymy  
798 of *Metacervocerus rhenanus*, '*Cervus philisi*', and '*Cervus perolensis*' could not be supported in the  
799 analyses.

800 In the past, '*Cervus philisi*' was considered to be related to *Axis* (Depéret and Mayet, 1911), to  
801 *Rusa* (Stehlin, 1923; Viret, 1954), and to *Cervus nippon* (Schaub, 1941). Heintz (1970) suggested an  
802 evolutionary *Metacervocerus pardinensis*-'*Cervus philisi*'-'*Cervus perolensis*'-lineage. However, the  
803 temporal occurrence of these species in the fossil record contradicts this hypothesis. It was suggested  
804 that '*Cervus perolensis*' is the descendant of '*Cervus philisi*' (Stefaniak and Stefaniak, 1995; Croitor,  
805 2006a, 2014) and that *Metacervocerus rhenanus* from Tegelen and '*Cervus philisi*' from Senèze are  
806 synonymous and that '*Cervus philisi*' and '*Cervus perolensis*' are junior synonyms of *Metacervocerus*  
807 *rhenanus* (Spaan, 1992). Later, '*Cervus philisi*' was included in the genus *Metacervocerus* (Croitor and  
808 Bonifay, 2001; Croitor, 2006a) In Mennecart et al. (2017) '*Cervus philisi*' was placed closely related to  
809 *Axis* and *Rucervus duvaucelii*.

810 '***Cervus sivalensis***. The remains of '*Cervus sivalensis*' resemble *Rucervus duvaucelii* in morphology  
811 and size and *Rucervus eldii* in antler morphology (Azzaroli, 1954). Here, '*Cervus sivalensis*' was placed  
812 as the sister taxon to *Megaloceros giganteus* to a clade consisting of *Axis lydekkeri*, *Rusa kendagensis*,  
813 and *Metacervocerus pardinensis* to *Metacervocerus pardinensis*, to the *Elaphurus-Rucervus-Rusa*-clade,  
814 or in a polytomy with *Metacervocerus pardinensis* and *Cervus canadensis* within the *Cervus*-clade. The  
815 placements within Cervini and close to the *Cervus*-clade show that '*Cervus sivalensis*' belongs to Cervini  
816 and is most likely closely related to *Cervus*, *Rusa*, and/or *Rucervus*. Together with *Axis lydekkeri* it could  
817 belong to the ancestral group of cervids that leads to *Axis*, *Cervus*, *Rusa*, and *Rucervus*. Although the  
818 tooth morphology of '*Cervus sivalensis*' resembles that of *Rucervus* (pers. obs.), a placement closely  
819 related to *Rucervus* could not be found. There is still a lot of confusion concerning the taxonomy and  
820 systematics of this taxon and a revision is needed (Lydekker, 1884; Azzaroli, 1954; Arif et al., 1991;  
821 Samiullah and Akhtar, 2007).

822 '***Axis lydekkeri***. Even though *Axis lydekkeri* is a fairly complete fossil and despite the morphological  
823 similarities to *Axis*, *Axis lydekkeri* was not placed as closely related to extant *Axis* in our analyses. Here,  
824 *Axis lydekkeri* was mostly placed as the sister taxon to or within Cervini, or within the *Cervus*-clade,  
825 which shows that *Axis lydekkeri* belongs to Cervini.

826 *Axis lydekkeri* was suggested to be more closely related to the smaller *Axis* species of today ('*Hyela-*  
827 *phus*') than to *Axis axis*, but a clear systematic relationship to any of them could not yet be confirmed  
828 (Zaim et al., 2003; Meijaard and Groves, 2004).

829 '***Rusa kendagensis***. In the analyses here, *Rusa kendagensis* was most often placed within Cervini  
830 and sometimes as the sister taxon to the *Cervus*-clade, which shows that *Rusa kendagensis* belongs to  
831 Cervini. Even though based on comparative anatomy it is more similar to *Rusa*, the analyses placed it  
832 more closely to *Cervus*. *Rusa kendagensis* potentially belongs to an extinct group of ancestors including  
833 also *Axis lydekkeri* and '*Cervus sivalensis*', which gave rise to modern *Axis*, *Cervus*, and *Rusa*.



834 There is little information about *Rusa kendengensis* in the literature; the only study on this species  
835 reported that it belongs to *Rusa* and not to *Cervus* as previously assumed for most Pleistocene cervids  
836 from Java (Zaim et al., 2003). More material of this species is needed to further investigate its systematic  
837 relationships.

838 ***Candiacervus ropalophorus*.** In the analyses here, *Candiacervus ropalophorus* was often placed close  
839 to several fossil cervine taxa and/or within Cervinae; in the SFA it was placed within Odocoileini. The  
840 investigated *Candiacervus ropalophorus* specimens were fairly complete; therefore, it was unexpected  
841 that this taxon was difficult to place. Frequent placements as the sister taxon to Cervini or within Cervini  
842 indicated that *Candiacervus ropalophorus* belongs to Cervini. The often hypothesised close relationship  
843 to megacerine/damine deer could only be found in one topology.

844 For *Candiacervus ropalophorus*, up to six different size groups representing six taxonomic units,  
845 sometimes even eight morphotypes have been suggested, but with differing views on the actual taxonomic  
846 affiliations Simonelli (1907, 1908); Kuss (1975); Kotsakis and Palombo (1979); De Vos (1979, 1984,  
847 2000); Van der Geer et al. (2006). *Candiacervus ropalophorus* is the smallest species of the eight  
848 morphotypes. Since no cranial material can be unambiguously assigned to *Candiacervus cretensis* or  
849 *Candiacervus rethymnensis*, only *Candiacervus ropalophorus* can be considered as clearly recognisable  
850 species based on cranial and postcranial elements (De Vos, 1984).

851 The systematic position of *Candiacervus* is controversial; a close relationship to *Megaceros*, *Prae-*  
852 *megaceros*, *Eucladoceros*, *Cervus*, or *Croizetoceros*, as has been suggested before (Kuss, 1975; De Vos,  
853 1984). It remains difficult to determine the ancestor of the Greek island deer, and data are still insufficient  
854 to establish robust phylogenetic relationships of Cretan deer (Van der Geer et al., 2006).

855 ***Megaloceros giganteus*.** In the morphological analyses here, *Megaloceros giganteus* was placed in  
856 varying positions, within Cervinae, as the sister taxon to *Dama dama*, and often closely related to *Rangifer*  
857 *tarandus* (presumably due to similarities in antler morphology). A close relationship to *Dama*, as strongly  
858 suggested by molecular analyses (Lister et al., 2005), is also supported in the TE BI and ML topologies.  
859 Together with the evidence from comparative morphology a close relationship of *Megaloceros giganteus*  
860 to *Dama* is almost certain.

861 There is a broad consensus today that *Megaloceros* consists of only one species, *Megaloceros giganteus*  
862 (Vislobokova, 1990, 2012, 2013; Azzaroli and Mazza, 1993; Croitor et al., 2006; Croitor and Bonifay,  
863 2001; Croitor, 2014). All recent phylogenetic analyses consistently placed *Megaloceros giganteus* within  
864 Cervinae (Lister et al., 2005; Hughes et al., 2006; Vislobokova, 2009). In some studies *Megaloceros*  
865 *giganteus* was placed closely related to *Cervus elaphus* based on molecular data (Kuehn et al., 2005)  
866 and morphological data (Geist, 1998; Pfeiffer, 1999, 2002; Vislobokova, 2009). Lönnberg (1906) put  
867 it close to *Rangifer* because of a completely ossified vomer and palmated brow tines; however, it was  
868 found that the division of the nasal cavity is only ossified in the anterodorsal part of the vomerine  
869 septum, which is different from the condition in Capreolinae and presumably is a side effect of the cranial  
870 pachyostosis (Lister, 1994; Croitor, 2006b, 2014). Already Lydekker (1898) suggested an affiliation of  
871 *Megaloceros giganteus* to the damine group, which was supported in several subsequent studies using  
872 morphological, molecular or both types of data (Gould, 1974; Kitchener, 1987; Lister, 1994; Lister et al.,  
873 2005; Vislobokova, 2009). In the topology of Marcot (2007) *Megaloceros giganteus* was the sister taxon  
874 to all cervine taxa, and in Pfeiffer (2002) it was the sister taxon to two extant *Cervus*. In Mennecart et al.  
875 (2017) *Megaloceros giganteus* was the sister taxon to *Dama*.

876 ***Odocoileus*.** In the analyses here, both fossil *Odocoileus* specimens were most often placed as the sister  
877 taxon to odocoileine taxa, within Blastocerina, and sometimes to the other fossil *Odocoileus*.

878 The results for both fossil *Odocoileus* suggest that they are included within Capreolinae and within  
879 Odocoileini. However, only a few analyses placed them as sister taxa or closely related to their presumed  
880 living descendants *Odocoileus virginianus* and *Odocoileus hemionus*. Particularly the BSPG specimen  
881 was more often placed closely related to *Mazama* species. In Mennecart et al. (2017) the fossil *Odocoileus*  
882 BSPG specimen was placed in a trichotomy with the extant *Odocoileus* species.

883 ***Muntiacus*.** The fossil *Muntiacus muntjak* was often placed within Muntiacini, mostly as the sister taxon  
884 to *Muntiacus atherodes*. The results show that the fossil *Muntiacus* is certainly a member of Muntiacini.

## 885 Extant Cervidae

886 Until recently, there were no comprehensive studies investigating the phylogenetic relationships of extant  
887 cervids based on morphology. Due to the highly conservative craniodental features of cervids, implications  
888 from the topologies based on morphology alone were limited. In the molecular topologies here, the  
889 systematic relationships of most clades above genus level were consistently recovered and well supported  
890 by different data sets. Many systematic relationships at genus- and/or species-level were also stable  
891 and were consistently placed on the same positions in topologies based on various molecular data sets.  
892 However, even though molecular data contributed to delimiting cervid clades and helped understanding  
893 the morphological evolution, some nodes remain unresolved or unstable. In the molecular and combined  
894 topologies, apart from a very few exceptions, Cervidae, Capreolinae, and Cervinae were monophyletic;  
895 Cervini, Muntiacini, Odocoileini including *Rangifer* most often were monophyletic, too. The unstable  
896 position of Capreolini and Alceini questioned the monophyly of Capreolinae.

## 897 Cervini

898 The phylogenetic relationships of Cervini here, were similar to the results of recent molecular studies  
899 including Cervini; (Randi et al., 1998, 2001; Meijaard and Groves, 2004; Pitra et al., 2004; Hernández  
900 Fernández and Vrba, 2005; Gilbert et al., 2006; Hughes et al., 2006; Marcot, 2007; Ouithavon et al.,  
901 2009; Hassanin et al., 2012; Heckeberg et al., 2016). The relationships within the subclades vary slightly  
902 depending on the taxon and character sampling.

903 There has been a long ongoing discussion about the genus and subgenus status of cervine taxa. In this  
904 study and in most of the recent literature (e.g., IUCN, 2016; Mattioli, 2011) six genera were distinguished:  
905 *Axis*, *Cervus*, *Dama*, *Elaphurus*, *Rucervus*, and *Rusa*. *Przewalskium* was often listed as a seventh separate  
906 genus; however, extensive morphological investigation did not find enough difference for a separate genus  
907 status (pers. obs.). *Elaphurus*, *Rucervus*, and *Rusa* are often considered as subgenera (Meijaard and  
908 Groves, 2004; Pitra et al., 2004; Gilbert et al., 2006; Hassanin et al., 2012), but have many morphological  
909 distinctive features that justify separate genera (pers. obs.).

910 **Axis.** The study of Meijaard and Groves (2004) was so far the only one to include the three species,  
911 *Axis axis*, *Axis porcinus* and *Axis kuhli*, for which molecular data was available. In the supertree analysis  
912 of Hernández Fernández and Vrba (2005) all four *Axis* species were included. *Axis* was not monophyletic  
913 in some studies (Pitra et al., 2004; Marcot, 2007; Agnarsson and May-Collado, 2008). This is most likely  
914 caused by re-analysing the same misidentified sequences (see discussion in Gilbert et al. (2006)).

915 In the analyses here *Axis* formed a well supported clade. *Axis axis* was always the sister taxon to  
916 the other two *Axis* species. Based on craniometrics and morphological similarities *Axis calamianensis*,  
917 *Axis kuhli*, *Axis porcinus* were considered to be closely related to each other and distinct from *Axis axis*  
918 (Meijaard and Groves, 2004). This was confirmed by our molecular and combined topologies. In most  
919 of the topologies here *Axis* was closely related to *Rucervus*, which differs from the results in Pitra et al.  
920 (2004) and the supertree analysis in Hernández Fernández and Vrba (2005).

921 **Cervus.** The morphological analyses here, resulted in varying positions for the four *Cervus* species. All  
922 of them have a very similar cranial and dental morphology (pers. obs.). In the nuclear analyses, *Cervus*  
923 *elaphus*, *Cervus canadensis*, and *Cervus nippon* were more closely related to each other than to *Cervus*  
924 *albirostris*. In the mtG analyses *Cervus albirostris* and *Cervus nippon* formed a clade and *Cervus elaphus*  
925 was the sister taxon to them; if *Cervus canadensis* was included it was the sister taxon to *Cervus nippon*  
926 (and *Cervus albirostris*, if it was a trichotomy) and *Cervus elaphus* was the sister taxon to all of them.  
927 This was also the case in the combined molecular and TE analyses. This difference between mitochondrial  
928 and nuclear genes may indicate an ancient hybridisation event.

929 In previous studies, *Cervus elaphus* was the sister taxon to *Cervus nippon* (Lister, 1984; Randi et al.,  
930 1998). or *Cervus nippon* was the sister taxon to *Cervus canadensis*, with *Cervus elaphus* and *Rusa* as  
931 the sister taxa to them (Randi et al., 2001; Pitra et al., 2004; Hughes et al., 2006). *Cervus canadensis*  
932 was the sister taxon to *Cervus nippon* with *Cervus albirostris* and *Cervus elaphus* as the sister taxon  
933 to all of them in Kuwayama and Ozawa (2000); Groves (2006); Zachos et al. (2014). This contradicts  
934 results from traditional morphology, where *Cervus elaphus* and *Cervus canadensis* were usually sister  
935 taxa (Kuwayama and Ozawa, 2000). However, Polzheim and Strobeck (2002) stated that the divergence of  
936 mtDNA noted for *Cervus nippon*, *Cervus canadensis*, and *Cervus elaphus* is congruent with geographical,  
937 morphological, and behavioural distinctions.

938 In some studies, *Cervus albirostris* was the sister taxon to the other *Cervus* species (Hernández  
939 Fernández and Vrba, 2005); it was the sister taxon to *Cervus nippon*, with *Cervus canadensis* as the  
940 sister taxon to both and *Cervus elaphus* the sister taxon to all of them (Marcot, 2007), as in Hassanin  
941 et al. (2012), excluding *Cervus canadensis*. In Agnarsson and May-Collado (2008) *Cervus albirostris*  
942 was the sister taxon to *Cervus elaphus*, and *Cervus nippon* to both of them. *Cervus albirostris* was  
943 the sister taxon to *Cervus canadensis* and *Cervus nippon* in some studies or to *Cervus nippon* (Polziehn  
944 and Strobeck, 2002; Liu et al., 2002; Groves, 2006), which is also confirmed in the analyses here. In  
945 contrast to this, Flerov (1952) suggested that *Cervus albirostris* diverged from *Rusa* in the late Pliocene  
946 and Koizumi et al. (1993) considered it more closely related to *Rucervus*. However, all recent molecular  
947 studies placed it closer to the *Cervus* species (Leslie, 2010). *Cervus albirostris* almost certainly evolved in  
948 temperate northern Eurasia; *Epirusa hiltzheimeri* or *Eucladoceros* may have been its Pleistocene ancestors  
949 (Di Stefano and Petronio, 2002; Flerov, 1952; Zdansky, 1925; Geist, 1998; Grubb, 1990; Leslie, 2010).

950 It is known that hybridisation between *Cervus nippon* and *Cervus elaphus* (mainly *Cervus elaphus*  
951 females and *Cervus nippon* males) occurs and that hybrids are fertile. Hybridisation may lead to extensive  
952 introgression (Zachos and Hartl, 2011). Studies on population genetics and subspecies of red deer  
953 exclusively used mtDNA, which may suggest relationships that are not reproducible when using paternal  
954 genes. Hybridisation could have occurred frequently in *Cervus*. The topologies here suggested varying  
955 sister taxon relationships across the four *Cervus* species.

956 **Dama.** In the analyses here, *Dama dama* and *Dama mesopotamica* were always sister taxa to each other  
957 and in most cases placed as the sister taxon to a clade consisting of *Cervus*, *Rusa*, *Elaphurus davidianus*,  
958 and *Rucervus eldii*. In previous studies, both *Dama* species were also sister taxa to each other (Randi  
959 et al., 2001; Lister et al., 2005; Hughes et al., 2006; Hassanin et al., 2012).

960 **Elaphurus.** In the nuclear analyses here, *Elaphurus davidianus* was mostly placed close to *Cervus*,  
961 while it was consistently placed as the sister taxon to *Rucervus eldii* in all mitochondrial, molecular  
962 combined, and TE analyses. In the morphological analyses it was placed closer to *Cervus* based on cranial  
963 characters and closer to *Rucervus* and *Rusa*, particularly *Rucervus schomburgki*, based on the dentition  
964 and the morphological combined data set.

965 The oldest known fossils of the *Elaphurus davidianus* lineage are known from the late Pliocene or  
966 slightly earlier (Taru and Hasegawa, 2002) and the first certain *Elaphurus davidianus* fossils date from the  
967 mid Pleistocene (Ji, 1985). The speciation of *Elaphurus* has been discussed as an ancient (late Pliocene or  
968 earlier) hybridisation event (Meijaard and Groves, 2004). *Cervus canadensis* or a closely related ancestor  
969 supposedly was the male parent and *Rucervus eldii* or a very close ancestral relative the female parent  
970 (Taru and Hasegawa, 2002; Meijaard and Groves, 2004; Pitra et al., 2004; Groves, 2006). The unique  
971 antler morphology and the overall phenotype of *Elaphurus davidianus* is distinct from all other cervids  
972 (Lydekker, 1898; Emerson and Tate, 1993; Meijaard and Groves, 2004; Pitra et al., 2004). Although some  
973 similarities to *Rucervus eldii* were stated (e.g., Meijaard and Groves, 2004), morphological scrutiny does  
974 not necessarily support that. The morphology of *Elaphurus* contains apomorphic character states and  
975 is not intermediate between its two parent taxa (Groves, 2014, ; own observations). This phenomenon  
976 is called transgressive segregation and the new phenotypes may be favoured in the new hybridogenetic  
977 population (Rieseberg et al., 1999; Groves, 2014).

978 Because of this hybridisation molecular phylogenetic analyses result in conflicting systematic positions  
979 as clearly shown here, but also in earlier studies. Analyses of mitochondrial data placed *Elaphurus*  
980 *davidianus* as the sister taxon to *Rucervus eldii* (Randi et al., 2001; Pitra et al., 2004), while Electrophoretic  
981 patterns of 22 proteins and  $\kappa$ -casein DNA, and the karyotype placed *Elaphurus* closer to *Cervus* (Emerson  
982 and Tate, 1993; Cronin et al., 1996; Meijaard and Groves, 2004).

983 **Rucervus.** *Rucervus* species have a unique antler morphology and their teeth are uniquely folded  
984 indicating a specialisation for graminivory (Grubb, 1990; Meijaard and Groves, 2004); both provide  
985 useful morphological characters. The hypothesis that *Rucervus* is more closely related to *Rusa* than to  
986 *Cervus* was partly supported in the nuclear analyses and the morphological analyses here, while in the  
987 mitochondrial, molecular combined, and TE analyses *Rucervus* was polyphyletic with *Rucervus eldii*  
988 more closely related to *Elaphurus davidianus* and the other two species more closely related to *Axis*.  
989 Based on this it was suggested that *Rucervus eldii* may represent a different evolutionary lineage than the  
990 other two *Rucervus* species (Meijaard and Groves, 2004) and was sometimes put into a separate genus  
991 *Panolia* (Pocock, 1943; Groves, 2006). It is now widely regarded as *Rucervus eldii* (Wilson and Reeder,

2005; Timmins et al., 2008; Angom and Hussain, 2013). This is also supported by the topologies here, particularly the morphological topologies show the close relationship to the other two *Rucervus* species. The placement of *Rucervus eldii* separate from its two congeners in molecular topologies (especially mtDNA) is most likely artificially caused by the hybridisation of *Rucervus eldii* and *Cervus canadensis* in the past.

*Rucervus duvaucelii* and *Rucervus schomburgki* were sister taxa to each other in the analyses here and were mostly the sister taxon to *Axis*. The last specimen of *Rucervus schomburgki* became extinct in 1938. The first accounts on the species were by Blyth (1863), who noted the distinctive antler pattern. According to Gähler (1936), the geographical distribution of *Rucervus schomburgki* was restricted to Siam. It was assumed to be closely related to *Rucervus duvaucelii* and potentially interbreeding with *Rucervus eldii* in its natural habitat. The earliest fossils of *Rucervus* date back to 2.9 mya (Azzaroli et al., 1988; Meijaard and Groves, 2004).

**Rusa.** In the morphological analyses here, *Rusa* was more closely related to *Rucervus* (rarely to *Axis*). In the nuclear analyses, it was close to *Rucervus* or within Cervini, while it was more closely related to *Cervus* in the mitochondrial, combined molecular, and TE analyses. When all four *Rusa* were included, *Rusa timorensis* and *Rusa unicolor* were sister taxa and *Rusa marianna* and *Rusa alfredi* were sister taxa.

Despite some new insights into the systematic relationships of *Rusa*, uncertainties remain (Heckeberg et al., 2016). The Philippine *Rusa alfredi* and *Rusa marianna* share morphological similarities, and are distinct from the other two *Rusa* because of the overall smaller size. *Rusa unicolor* and *Rusa timorensis* from the mainland and Indonesia were considered to be more derived (Groves and Grubb, 2011), which is in contrast to the assumption that based on the high similarity of *Rusa unicolor* to pliocervines, an extinct lineage of Pliocene cervids, it is the most ancestral of the four extant rusine deer (Petronio et al., 2007; Leslie, 2011).

Although the monophyly of *Rusa* has been controversial based on morphological and molecular evidence (Meijaard and Groves, 2004; Hernández Fernández and Vrba, 2005; Randi et al., 2001; Leslie, 2011), in our analyses *Rusa* is more often supported to be monophyletic than not.

The first appearance of *R. unicolor* was recorded from the middle Pleistocene (Zong, 1987; Dong, 1993; Meijaard and Groves, 2004). The oldest *R. timorensis* is reported from the late Pleistocene (Van Mourik and Stelmasiak, 1986; Dong, 1993) and suggested to have then dispersed south-eastwards to Taiwan and Java (Meijaard and Groves, 2004).

#### 1022 **Muntiacini**

1023 In the recent literature, muntiacines have been included in phylogenetic reconstructions to a different  
1024 extent (Randi et al., 1998; Wang and Lan, 2000; Randi et al., 2001; Pitra et al., 2004; Hernández Fernández  
1025 and Vrba, 2005; Gilbert et al., 2006; Hughes et al., 2006; Marcot, 2007; Outhavon et al., 2009; Hassanin  
1026 et al., 2012). The systematic relationships within Muntiacini vary mostly depending on the taxon sampling,  
1027 but do not contradict each other. The monophyly of Muntiacini uniting *Muntiacus* and *Elaphodus* has  
1028 never been questioned Gilbert et al. (2006) and is supported by our analyses.

1029 **Elaphodus.** *Elaphodus cephalophus* was always the sister taxon to the other muntiacine species in all  
1030 molecular and TE analyses presented here, which is also widely supported in the literature (e.g., Wang  
1031 and Lan, 2000; Hernández Fernández and Vrba, 2005; Agnarsson and May-Collado, 2008; Hassanin  
1032 et al., 2012). In contrast, in Marcot (2007) *Elaphodus cephalophus* is the sister taxon to all cervids.

1033 *Elaphodus cephalophus* has the smallest known antlers, which are completely covered by tufts  
1034 (Leslie et al., 2013). Groves and Grubb (1990) considered *Elaphodus cephalophus* as the most primitive  
1035 representative of living muntiacines. However, this is in contrast to the absence of fossils with such  
1036 diminutive antlers. The first *Elaphodus* fossils are known from the Pleistocene of China, which were  
1037 larger than *Elaphodus cephalophus*; therefore, the decrease in size can be considered as evolutionary  
1038 trend in this species (Leslie et al., 2013).

1039 **Muntiacus.** All muntjacs have long pedicles, facial crests, and bifurcating antlers (pers. obs.; e.g., Ma  
1040 et al., 1991). In the morphological analyses here, muntiacine taxa were placed as the sister taxa to most  
1041 other cervids or in an unresolved position. In most of the combined morphological analyses Muntiacini  
1042 was monophyletic except for the BI analyses. In the MP analyses, Muntiacini were placed more closely  
1043 related to other small cervids, such as *Mazama* and *Pudu*.

1044 The earliest fossil of the *Muntiacus* lineage is *Muntiacus leilaoensis* from Yunnan, China and was  
1045 dated to the late Miocene 9–7 mya (Dong et al., 2004). All *Muntiacus* species consistently formed a clade  
1046 as the sister taxon to Cervini in the mitochondrial, molecular combined, and TE analyses here. A clade  
1047 consisting of *Muntiacus crinifrons*, *Muntiacus feae*, and *Muntiacus muntjak* and a clade consisting of  
1048 *Muntiacus putaoensis*, *Muntiacus truongsongensis*, *Muntiacus rooseveltorum*, *Muntiacus vuquangensis*,  
1049 and *Muntiacus reevesi* were recovered in the mitochondrial and combined molecular analyses. *Muntiacus*  
1050 *atherodes* was placed in a polytomy with these clades. In the TE analyses *Muntiacus reevesi* was placed  
1051 between *Elaphodus cephalophus* and the other muntjacs and *Muntiacus atherodes* was the sister taxon to  
1052 *Muntiacus feae*.

1053 Several new muntiacine species have been discovered in the 1990s; subsequently, five to possibly  
1054 six new muntjac species were established, *Muntiacus gongshanensis*, *Muntiacus crinifrons*, *Muntiacus*  
1055 *feae*, *Muntiacus reevesi*, *Muntiacus muntjak* (Lan et al., 1995). Ma et al. (1986b,a) stated that *Muntiacus*  
1056 *crinifrons* and *Muntiacus rooseveltorum* derived from *Muntiacus reevesi*, whereas *Muntiacus feae* and  
1057 *Muntiacus muntjak* derived from a different lineage. The species status of *Muntiacus rooseveltorum* has  
1058 been controversial for decades (Amato et al., 1999b); for example, Groves and Grubb (1990) suggested  
1059 that *Muntiacus rooseveltorum* is the synonym of *Muntiacus feae* and that *Muntiacus feae* is the sister taxon  
1060 to *Muntiacus muntjak* and *Muntiacus crinifrons*. This is supported by most molecular studies and the  
1061 topologies of this work. Sometimes, *Muntiacus crinifrons* and *Muntiacus gongshanensis* are considered  
1062 as a single species (Amato et al., 1999b). It was proposed that *Muntiacus atherodes* should be included  
1063 in *Muntiacus muntjak* based on morphological evidence, because the holotype of *Muntiacus atherodes*  
1064 is a subadult male with single-tined antlers (Ma et al., 1986b). The two specimens investigated here  
1065 were indeed subadult individuals with not yet fully developed antlers (pers. obs.). However, molecular  
1066 topologies here and in the literature indicate a separate species status for *Muntiacus atherodes* (Heckeberg  
1067 et al., 2016). The genus status of *Megamuntiacus* is not justified demonstrated by the sequence divergence  
1068 estimated for the mitochondrial variation and by morphological comparisons; therefore, it is referred to as  
1069 *Muntiacus* (Schaller, 1996; Gao et al., 1998; Amato et al., 1999a; Rabinowitz et al., 1999; Wang and Lan,  
1070 2000). Apart from the larger size, there are no morphological features that would justify a separate genus  
1071 (pers. obs.).

### 1072 **Alceini**

1073 **Alces.** *Alces* has a highly derived skull morphology with an elongated viscerocranial proportion and  
1074 antlers that protrude horizontally. The dentition shows similar modifications as in *Rangifer*. In the  
1075 morphological analyses here, *Alces alces* was in an unresolved position or placed as the sister taxon to  
1076 *Odocoileus hemionus*, *Mazama chunyi*, *Ozotoceros bezoarticus* or *Cervus canadensis*. In the mitochon-  
1077 drial, combined molecular and TE analyses *Alces alces* was consistently placed as the sister taxon to  
1078 Capreolini, except for the BI combined molecular topology, where it was placed between Capreolini and  
1079 Odocoileini plus *Rangifer*.

1080 The first *Alces alces* is known from the Riss glaciation 200-100 kya; those late Pleistocene moose  
1081 were larger than their extant representatives (Franzmann, 1981). In most recent studies, *Alces* was placed  
1082 as the sister to Capreolini (Randi et al., 1998; Pitra et al., 2004; Hughes et al., 2006; Agnarsson and  
1083 May-Collado, 2008; Hassanin et al., 2012) or as the sister taxon to *Capreolus* (Hernández Fernández and  
1084 Vrba, 2005). In Marcot (2007) *Alces* was the sister taxon to Capreolini and Odocoileini and Rangiferini,  
1085 while it was in a polytomy with Odocoileini plus *Rangifer* and Capreolini or the sister taxon to Odocoileini  
1086 plus *Rangifer* in Gilbert et al. (2006). More controversial positions included *Alces* as the sister taxon  
1087 to Cervini or *Dama dama* in Kuehn et al. (2005) and the sister taxon position to *Rangifer* in Pfeiffer  
1088 (2002). *Alces* was in a polytomy with Odocoileini and Rangiferini in Lister (1984) and took up variable  
1089 positions in previous studies as summarised in Lister (1998). Thus, the systematic position of *Alces*  
1090 remains unresolved.

### 1091 **Capreolini**

1092 Most analyses based on the combined morphological data set supported monophyletic Capreolini. How-  
1093 ever, the systematic position of Capreolini varied and could not be determined with certainty using  
1094 morphological data only. In the molecular analyses here, Capreolini was always monophyletic and mostly  
1095 placed closely related to or in most cases as the sister taxon to Odocoileini plus *Rangifer*.

1096 Miyamoto et al. (1990) suggested that Capreolini probably originated in the late Miocene in the Old  
1097 World. The assumption of a late Miocene Old World origin of Capreolinae is in congruence with our

1098 findings considering the placement of *Procapreolus*. Cronin (1991) hypothesised that *Alces* and *Rangifer*  
1099 split earlier than the *Capreolus* lineage, but after the separation of Cervinae and Capreolinae.

1100 **Capreolus.** In the morphological, molecular, and TE topologies *Capreolus capreolus* and *Capreolus*  
1101 *pygargus* both species were consistently placed as sister taxa. In the mitochondrial, molecular combined  
1102 and TE topologies, *Capreolus* was always the sister taxon to *Hydropotes* with strong support. Molecular  
1103 studies of the past decades support the consistent placement of *Hydropotes* as the sister taxon to *Capreolus*  
1104 forming monophyletic Capreolini (Douzery and Randi, 1997; Randi et al., 1998; Hassanin and Douzery,  
1105 2003; Pitra et al., 2004; Hughes et al., 2006; Gilbert et al., 2006; Marcot, 2007; Agnarsson and May-  
1106 Collado, 2008; Hassanin et al., 2012; Heckeberg et al., 2016).

1107 **Hydropotes.** Here, *Hydropotes* and *Capreolus* were sister taxa in the morphological combined, mt,  
1108 molecular combined and TE analyses. In the past, *Hydropotes* was considered as a separate subfamily  
1109 Hydropotinae as the sister taxon of all other cervids (e.g., Groves and Grubb, 1987; Janis and Scott, 1987;  
1110 Hernández Fernández and Vrba, 2005; Kuznetsova et al., 2005). Already Bouvrain et al. (1989) favoured  
1111 the hypothesis that *Hydropotes* and Capreolini are sister taxa. The first molecular studies indicated that  
1112 *Hydropotes* is included in monophyletic Cervidae (Kraus and Miyamoto, 1991). From this follows that  
1113 *Hydropotes* lost the antlers secondarily and developed enlarged upper canines as compensation (Douzery  
1114 and Randi, 1997; Randi et al., 1998; Hassanin and Douzery, 2003).

1115 Randi et al. (1998) demonstrated that the two *Capreolus* species and *Hydropotes* share a G at position  
1116 525 of *Cytb*, which occurs only rarely in other mammal species and stated that ‘this replacement represents  
1117 a nearly exclusive synapomorphy for the *Hydropotes-Capreolus*-clade. Further, the telemetacarpal  
1118 condition and a large medial opening of the temporal canal are morphological features that *Hydropotes*  
1119 shares with other Capreolinae (Bouvrain et al., 1989; Douzery and Randi, 1997; Randi et al., 1998).  
1120 Behavioural characters also suggested that *Hydropotes inermis* is closely related to *Capreolus* (Cap et al.,  
1121 2002).

1122 In contrast to the opinion stated in the extensive review of *Hydropotes inermis* (Schilling and Rössner,  
1123 2017) more and more evidence (mitochondrial and nuclear DNA, morphology, behaviour) point to a sister  
1124 taxon relationship of *Hydropotes* and *Capreolus*.

#### 1125 **Rangiferini**

1126 **Rangifer.** The systematic position of *Rangifer* was variable in the morphological analyses here. *Rangifer*  
1127 has some apomorphic characters, not shared by other cervids, which is likely the cause of the difficulties  
1128 to place the taxon based on morphology only. In the molecular and TE topologies *Rangifer tarandus*  
1129 was consistently placed as the sister taxon to Odocoileini. This is supported by the most recent literature,  
1130 (Randi et al., 1998; Hassanin and Douzery, 2003; Pitra et al., 2004; Hernández Fernández and Vrba, 2005;  
1131 Gilbert et al., 2006; Hughes et al., 2006; Agnarsson and May-Collado, 2008; Duarte et al., 2008; Hassanin  
1132 et al., 2012). *Rangifer* was in a polytomy with Odocoileini and Alceini in Lister (1984). Pfeiffer (2002)  
1133 found that *Rangifer* is the sister taxon to *Alces* based on morphological characters.

1134 *Rangifer* appeared in the fossil record in the Pleistocene; based on its arctic specialisations it is  
1135 hypothesised that it dispersed to America during the Pleistocene contemporaneously with *Alces* (Gilbert  
1136 et al., 2006).

#### 1137 **Odocoileini**

1138 In the morphological topologies here most odocoileine taxa were in unresolved and/or variable posi-  
1139 tions. In several topologies the small odocoileine cervids were in a clade with muntiacine taxa. In the  
1140 nuclear topologies, systematic relationships within Odocoileini were partly or entirely unresolved. In the  
1141 mitochondrial, combined molecular, and TE topologies here, Odocoileini split into the two subclades  
1142 Blastocerina and Odocoileina (Heckeberg et al., 2016).

1143 In previous phylogenetic studies, the taxon sampling for Odocoileini varied greatly, therefore, it  
1144 is difficult to compare the topologies (Douzery and Randi, 1997; Randi et al., 1998; Pitra et al., 2004;  
1145 Hernández Fernández and Vrba, 2005; Hughes et al., 2006; Gilbert et al., 2006; Marcot, 2007; Agnarsson  
1146 and May-Collado, 2008; Duarte et al., 2008; Hassanin et al., 2012). In these studies, Odocoileini usually  
1147 formed a monophyletic group with Rangiferini as the sister taxon to them. *Blastocerus dichotomus*,  
1148 *Ozotoceros bezoarticus*, and *Pudu puda* were particularly unstable across studies with comparable taxon  
1149 sampling. In the topologies here, they were sensitive to changes in the analysis parameters. Odocoileina  
1150 and Blastocerina were sister taxa in several recent studies (Pitra et al., 2004; Hughes et al., 2006; Gilbert

1151 et al., 2006; Marcot, 2007; Agnarsson and May-Collado, 2008; Hassanin et al., 2012; Heckeberg et al.,  
1152 2016). This is also the case in Duarte et al. (2008), but *Pudu puda* was in a polytomy to those clades.  
1153 In addition, the results here and those of previous studies showed polyphyly for three odocoileine  
1154 genera *Hippocamelus*, *Mazama*, and *Pudu* and for both species of *Odocoileus* (Pitra et al., 2004; Gilbert  
1155 et al., 2006; Agnarsson and May-Collado, 2008; Duarte et al., 2008; Hassanin et al., 2012; Heckeberg  
1156 et al., 2016). It remains uncertain, whether *Pudu* is monophyletic, polyphyletic within Blastocerina or  
1157 polyphyletic with one species in Blastocerina and one species in Odocoileina. More morphological and  
1158 molecular, particularly nuclear markers, and cytogenetic data are needed to reconstruct the complex  
1159 evolutionary history of Odocoileini (Duarte et al., 2008; Hassanin et al., 2012).

1160 **Blastocerus.** In the analyses here, *Blastocerus dichotomus* was positioned in an unresolved position  
1161 based on morphological data and consistently placed within Blastocerina in the molecular and TE analyses.  
1162 Most often it was positioned between *Pudu puda* (sometimes also *Mazama nemorivaga*) and the other  
1163 Blastocerina. In previous studies *Blastocerus* took up variable positions, most likely depending on the  
1164 taxon sampling. For example, as the sister taxon to *Hippocamelus bisulcus* plus *Mazama gouazoubira*  
1165 (Duarte et al., 2008), as the sister taxon to *Mazama gouazoubira* (Agnarsson and May-Collado, 2008), in  
1166 a polytomy with *Mazama gouazoubira*, *Pudu puda*, *Hippocamelus antisensis* (Gilbert et al., 2006), as the  
1167 sister taxon to *Pudu puda* (Hughes et al., 2006), and as sister taxon to *Mazama nemorivaga* (Hassanin  
1168 et al., 2012). Studies with a more extensive taxon sampling (Heckeberg et al., 2016) and the analyses of  
1169 this work indicated a systematic position of *Blastocerus* as the sister taxon to most blastocerine species,  
1170 with *Mazama nemorivaga* as the sister taxon to them and *Pudu puda* as the sister taxon to all other  
1171 Blastocerina. A few analyses placed *Blastocerus* as the sister taxon to all other Blastocerina. These  
1172 differing placements of *Blastocerus* most likely resulted from a differing taxon sampling.

1173 The first *Blastocerus* fossils are known from the Pleistocene of Brazil and Paraguay. The populations  
1174 in central Brazil most likely expanded between 28–25 kya and it was assumed that there were no  
1175 geographical barriers until about 300 years ago (Merino and Rossi, 2010).

1176 **Hippocamelus.** In several of the morphological topologies, both *Hippocamelus* species were mono-  
1177 phyletic, sometimes with *Ozotoceros* as the sister taxon. Two of the four sequences for *Hippocamelus*  
1178 *antisensis* formed a clade with *Hippocamelus bisulcus*, while the other two formed a clade with *Ozotoceros*  
1179 *bezoarticus* (Heckeberg et al., 2016). This makes it almost certain that two of the four sequences are  
1180 misidentified or mislabelled; a less likely possibility is that this polyphyly represents a valid split within  
1181 the genus. Without knowing the exact provenance of the samples it cannot be determined which sequences  
1182 are truly *Hippocamelus antisensis*. In the molecular combined and TE analyses here, we included those  
1183 *Hippocamelus antisensis* mt-sequence(s), with which the genus is monophyletic (Heckeberg et al., 2016).  
1184 *Hippocamelus* was the sister taxon to *Mazama gouazoubira* (plus *Mazama chunyi*, if included).

1185 Duarte et al. (2008) stated that it is surprising that members of morphologically cohesive genera such  
1186 as *Hippocamelus*, *Mazama*, or *Pudu* were not monophyletic based on molecular data. *Hippocamelus*  
1187 *antisensis* and *Hippocamelus bisulcus* were found to be osteologically nearly indistinguishable (Flueck  
1188 and Smith-Flueck, 2011, pers. obs.). Based on this, a monophyly for *Hippocamelus* is more likely  
1189 than a polyphyly as suggested by some of the molecular data. Thus, the potential polyphyly within  
1190 *Hippocamelus* cannot be confirmed or ruled out yet; new sequences and more investigations are needed to  
1191 clarify which of the available sequences genuinely belong to *H. antisensis*.

1192 The first *Hippocamelus bisulcus* is known from the late Pleistocene of Chile, Argentina, and Bolivia  
1193 (Canto et al., 2010; Merino and Rossi, 2010). *Odocoileus lucasi* is considered to be the ancestor of  
1194 *Hippocamelus bisulcus*.

1195 **Mazama.** In the morphological analyses here most *Mazama* species were placed as closely related  
1196 to each other most likely because of their small size and because they are morphologically almost  
1197 indistinguishable (González et al., 2009, ; own observations). In Gutiérrez et al. (2015), the suggested  
1198 potential morphological difference of *Mazama bricenii* and *Mazama rufina* referring to the degree of  
1199 concavity of the dorsal outline in lateral view is controversial, as both individuals seem to differ greatly in  
1200 age based on the tooth crown height. The second character, the lacrimal fossa, can generally be highly  
1201 variable among species. In the specimens scrutinised here, all *Mazama bricenii* skulls show a weak  
1202 concavity in the dorsal outline, not as deep as in the figure of Gutiérrez et al. (2015). One of the two  
1203 *Mazama rufina* specimens (NHMW 528) has a more clearly concave outline, the other one (ZSM 1927/41)  
1204 has a straight outline. In the most recent studies (Gutiérrez et al., 2015; Heckeberg et al., 2016) and the

1205 molecular analyses here, *Mazama bricenii* consistently was the sister taxon to *Mazama rufina*. Gutiérrez  
1206 et al. (2015) suggested that *Mazama bricenii* is not a valid taxon, but a junior synonym of *Mazama*  
1207 *rufina*. The systematic relationships of *Mazama* were discussed in detail in Heckeberg et al. (2016) and  
1208 polyphylies persist throughout different molecular and TE data sets. The complex taxonomy of *Mazama*  
1209 needs a thorough revision.

1210 While the monophyly of *Mazama* has never been questioned based on morphological characters,  
1211 molecular studies repeatedly suggested polyphyletic relationships (Gilbert et al., 2006; Duarte et al.,  
1212 2008; Gutiérrez et al., 2015; Escobedo-Morales et al., 2016; Heckeberg et al., 2016). Duarte et al. (2008)  
1213 suggested that *Mazama gouazoubira* and *Mazama nemorivaga* should be assigned to a different genus.  
1214 The low morphological diversity among *Mazama* is not correlated with the genotypic diversification,  
1215 which leads to the problematic taxonomy; thus, a varying number of species were established based on  
1216 different types of data (Groves and Grubb, 1987, 1990; Duarte and Merino, 1997; Duarte et al., 2008).

1217 Only little is known about rare *Mazama* species (and neotropical cervids in general), which represent  
1218 the least studied organisms and many aspects of their life history are poorly understood (Duarte et al.,  
1219 2012e,d,b,a,f; Lizcano et al., 2010; Gutiérrez et al., 2015).

1220 Previous molecular studies and the topologies here showed polyphylies of *Mazama americana*, which  
1221 suggested that it comprises several evolutionary units. The genetic distance between the two *Mazama*  
1222 *americana*-clades was higher than the genetic difference of *Mazama bororo* and *Mazama nana*. Therefore,  
1223 at least two species were assumed to be within the *Mazama americana*-complex, with a separate evolution  
1224 of the two clades starting 1 mya and 2 mya, respectively (Duarte et al., 2008; Abril et al., 2010). The  
1225 first fossil *Mazama* are known from the Pleistocene of Argentina, Ecuador, Peru, and Brasil (Merino and  
1226 Rossi, 2010).

1227 **Odocoileus.** In the morphological analyses based on the combined data set here *Odocoileus hemionus*  
1228 is the sister taxon to *Alces alces*, and in several topologies *Odocoileus virginianus* is the sister taxon  
1229 to them. In all other morphological topologies, odocoileine taxa are placed in unresolved or varying  
1230 positions. In the analyses including mitochondrial markers and a broad taxon sampling, both species were  
1231 polyphyletic. In the analyses based on the nuclear markers, polyphylies of the species were not observed.

1232 Despite all the research undertaken on the genus, the taxonomy remains difficult. There are numerous  
1233 subspecies (8–10 for *O. hemionus*, 37–38 for *O. virginianus*; Wilson and Reeder (2005); Mattioli (2011)),  
1234 which possibly, at least partly, represent separate species (Groves and Grubb, 2011).

1235 Latch et al. (2009) demonstrated that there are two different morphotypes of *O. hemionus*, the  
1236 mule deer and black-tailed deer, which is supported by a strong genetic discontinuity across the spatial  
1237 distribution. Early investigations of mtDNA data demonstrated that *O. hemionus* is polyphyletic because  
1238 the sequences of the mule deer (*O. hemionus*) and *O. virginianus* are more similar than the DNA of the  
1239 black-tailed deer (*O. hemionus columbianus*) is to both of them (5–7 % different) (Carr et al., 1986;  
1240 Cronin et al., 1988, 1996; Latch et al., 2009).

1241 Similarly, the genetic divergence within *O. virginianus* is remarkably high, even higher than the  
1242 genetic distance between other subspecies and between *O. virginianus* and mule deer. This led to the  
1243 classification of white tailed deer into two distinct groups, the *carriacou*-division and the *virginianus*-  
1244 division (Wilson et al., 1977; Smith et al., 1986; Groves and Grubb, 1987; Grubb, 1990). Some topologies  
1245 here (Figs 13, 14) and the literature (Heckeberg et al., 2016) most likely show the two distinct genetic  
1246 groups in each of the *Odocoileus* species. *Odocoileus virginianus* is a highly plastic species occupying a  
1247 great variety of geographically and ecologically extensive habitats between Canada and Peru, however,  
1248 extreme habitat differences do not necessarily lead to large morphological divergence (Smith et al., 1986;  
1249 Moscarella et al., 2003; Merino and Rossi, 2010; Duarte et al., 2012c). Introgression seems to be the  
1250 likely explanation because natural hybridisation and interbreeding between both species of *Odocoileus*  
1251 have been documented (Groves and Grubb, 2011; Hassanin et al., 2012).

1252 The first *Odocoileus* is from the early Pliocene (3.5 mya) of North America, where they were the  
1253 most common cervids until the Pleistocene. *Odocoileus virginianus* appeared 2 mya presumably as the  
1254 descendant of *O. brachyodontus*, which originated in Central America and dispersed to higher latitudes  
1255 only recently (Hershkovitz, 1972; Smith, 1991; Merino and Rossi, 2010). It has been assumed that  
1256 *Odocoileus virginianus* evolved in North America; it was further suggested that all South American cervid  
1257 fossils belong to *Odocoileus* and that *Mazama* later diverged as a consequence of isolation within South  
1258 America (Smith et al., 1986; Moscarella et al., 2003). This is in contrast with the most recent molecular  
1259 topologies (e.g., Escobedo-Morales et al., 2016; Heckeberg et al., 2016) and this work (Figs 13), from



1260 which it appears that *Odocoileus* originated from the odocoileine *Mazama*-clade.

1261 **Ozotoceros.** Similar to *Blastocerus*, the systematic position of *Ozotoceros* varied with the taxon  
1262 sampling. With an extensive taxon sampling *Ozotoceros bezoarticus* was relatively consistently placed as  
1263 the sister taxon to *Hippocamelus*, *Mazama gouazoubira* and *Mazama chunyi* (if included) in the analyses  
1264 here.

1265 The origin of *Ozotoceros bezoarticus* possibly dates back to 2.5 mya coinciding with a substantial  
1266 cooling event; fossils are known from the late Pleistocene and Holocene of Brazil, the late Pleistocene of  
1267 Uruguay, and the Holocene of Argentina (Gonzalez et al., 1998; Merino and Rossi, 2010).

1268 **Pudu.** Both *Pudu* species are almost indistinguishable based on morphology, but do not evidently form a  
1269 monophyletic group based on molecular data (Heckeberg et al., 2016). *Pudu puda* was placed as the sister  
1270 taxon to all Blastocerina in almost all of the analyses here and in previous studies with a sufficient taxon  
1271 sampling (Hassanin et al., 2012). The systematic position of its congener, unfortunately, is much less  
1272 certain. *Pudu mephistophiles* was most often placed as the sister taxon to all Odocoileini plus *Rangifer* or  
1273 to Odocoileini. Only in one topology there *Pudu mephistophiles* was included within Blastocerina.

1274 The spatial and chronological origin of *Pudu* is unknown. *Pudu* most likely diverged from an  
1275 odocoileine lineage, which existed in America since the Miocene-Pliocene-boundary (Merino and Rossi,  
1276 2010; Gonzalez et al., 2014). *Pudu* was probably restricted to South America since the Pliocene (Escamilo  
1277 et al., 2010).

## 1278 **Aspects of the Evolution of Cervidae**

### 1279 **Morphological Evolution**

1280 **Cranium** The cranial morphology of cervids is highly conservative (Lister, 1996; Merino and Rossi,  
1281 2010). Also, some morphological characters in ruminants likely are the results of convergent evolution and  
1282 thus are homoplastic, which may cause difficulties in reconstructing phylogenetic relationships (Bouvrain  
1283 et al., 1989; Douzery and Randi, 1997). Despite the homoplasy, some clades were well defined and  
1284 re-occurring across different data sets in the topologies here.

1285 Differences in the size of the praeorbital vacuity are primarily species specific, but have also an  
1286 ontogenetic component, since they are often smaller in aged individuals. Similarly, the lacrimal fossa  
1287 varies in size and depth in different species, presumably depending on the presence, size, and usage of  
1288 the lacrimal gland. Also, there is a difference between males and females. The position of the lacrimal  
1289 foramina to each other and on the orbit rim can potentially be used to distinguish groups of cervids. The  
1290 consistent presence of two lacrimal foramina is typical for cervids, but is also present in some bovid  
1291 species. In *Dremotherium feignouxi* sometimes only one lacrimal foramen is present (Costeur, 2011). The  
1292 contact of the lacrimal and the frontal at the orbit rim without interlocking sutures was first observed in  
1293 Rössner (1995). This trait is most likely an intraspecific variability and could be an effect of ageing.

1294 Evolutionary trends observed in Pliocene cervids include an increase of the overall body size, a  
1295 decrease of the pedicle length relative to the antler length and an associated increase of the antler length  
1296 (Heintz, 1970). The degree of inclination of the pedicles changes through time and is presumably a result  
1297 adapting to rich vegetation. With the stronger inclination the insertion point of the pedicle on the skull  
1298 moved posteriad. The pedicle in early Miocene cervids is entirely above the supraorbital process and not  
1299 in contact with the braincase; the pedicles are vertical in lateral view, parallel or converging in frontal  
1300 view. The shortening of the pedicles could be related to the increasing size of antlers, because a longer  
1301 and heavier set of antlers would put a biomechanically unfavourable leverage on the pedicles.

1302 Basicranial and ear region characters were not yet widely used when inferring morphological phylogene-  
1303 nies, but were assumed to have strong potential to provide characters, which are less prone to convergent  
1304 evolution caused by climatic change (Janis and Theodor, 2014). Recently, it has been shown that traits of  
1305 the inner ear provide useful characters with phylogenetic signal (Mennecart et al., 2016, 2017).

1306 **Antlers** There is broad consensus that antlers originated only once (Loomis, 1928; Azanza and Morales,  
1307 1989; Azanza, 1993a,b; Azanza et al., 2011; Heckeberg, 2017b). The antlers of most Miocene cervids  
1308 have a simple bifurcating pattern, sometimes with an additional tine, or are coronate (Azanza et al.,  
1309 2011). These antlers are relatively short, do not have a shaft and the bifurcation originates directly from  
1310 a broad antler base. From the late Miocene onwards, more complex branching patterns developed, the  
1311 length of antlers increased and antlers developed a shaft below the first bifurcation. Evolution of size and

1312 complexity of antlers is associated with reduction or loss of upper canines (Scott, 1937; Beninde, 1937;  
1313 Geist, 1966; Brokx, 1972).

1314 In extant cervids, short and simple antlers and long and more complex or palmated antlers are present.  
1315 Many extant cervids develop exactly three tines (Heckeberg, 2017b). The three antler morphotypes  
1316 have previously been associated with ecological habitats: simple antlers for the tropics, a three-tined  
1317 antler plan for woodland areas typical in East Eurasia or India, and the large and complex display organs  
1318 in temperate regions (Pitra et al., 2004). The simple antlers in *Mazama* and *Pudu* are considered as a  
1319 secondary adaptation to dense vegetation.

1320 There is a lot of inter- and intraspecific variation in antlers (Goss, 1983; Heckeberg, 2017b). The  
1321 high variability of antlers is a problem particularly in fossil taxa, where the entire intraspecific variation  
1322 cannot always be observed due to the lack of a sufficient number of specimens or the incompleteness of  
1323 ontogenetic stages. The taxonomy of fossil cervids is often based on antler morphology, because antlers  
1324 are easy to identify and numerous in the fossil record antler morphology having more distinctive, features  
1325 than other anatomical characters (Kurtén, 1968; Fry and Gustafson, 1974; Lister et al., 2010; Merino and  
1326 Rossi, 2010). Thus, the validity of some fossil cervid taxa is doubtful. To base classifications just on  
1327 antler morphology is problematic for the given reasons.

1328 In contrast to Loomis (1928), Gentry et al. (1999) stated that cranial appendage morphology proved  
1329 to be more suitable than tooth morphology to distinguish species. This applies in general to Pecora and  
1330 specifically to Cervidae. It is true that different cervid species can be easily identified based on their  
1331 antler morphology (branching pattern, orientation, size). Antler characters were often used to solve  
1332 intra-subfamily relationships, but they are problematic because of convergent development and subsequent  
1333 homoplasy in antler characters (Pitra et al., 2004).

1334 Since Cervidae is diagnosed by the presence of antlers (Janis and Scott, 1987; Pitra et al., 2004),  
1335 the reason for the absence of antlers in *Hydropotes inermis* species was controversially discussed; a  
1336 primitive condition and secondary loss have been suggested. To solve this issue, thorough research on the  
1337 process(es), which trigger the growth of the first set of antlers in antler-bearing species and when how and  
1338 why these processes/prerequisites are absent in *Hydropotes inermis* needs to be undertaken. The more  
1339 widely accepted hypothesis that *Hydropotes inermis* secondarily lost its antlers was applied here and the  
1340 presence of antlers is the synapomorphy of Cervidae.

1341 **Dentition** Variations of accessory dental elements in combination with the degree of molarisation of  
1342 premolars can be used to identify genera or species. Widely accepted evolutionary trends in cervids  
1343 concerning the dentition are increasing hypsodonty, the reduction of the premolar row length and the  
1344 reduction or loss of upper canines (Heintz, 1970; Dong et al., 2004). However, the hypsodonty index,  
1345 although widely used in ruminant phylogeny, has been considered to be a misleading character due to its  
1346 ambiguous definition and convergent evolution among all large herbivorous mammals (Janis and Scott,  
1347 1987; Hassanin and Douzery, 2003).

1348 The first deer had brachyodont dentition and were considered as leaf-eaters; recent dental analyses  
1349 generally support these findings, but also showed that *Procervulus ginsburgi* likely was a seasonal mixed  
1350 feeder. Based on this a facultative leaf-grass mixed feeding strategy with preference for leaf-eating is  
1351 likely the primitive dietary state in cervids and ruminants (DeMiguel et al., 2008).

1352 Ginsburg and Heintz (1966) regarded the bifurcation of the postprotocrista into an internal and external  
1353 crista as a derived cervid character based on its presence in *Dicrocerus* and *Euprox*. *Amphimoschus*  
1354 is the only other non-cervid pecoran species that shows this trait (Janis and Scott, 1987). The bifurcated  
1355 postprotocrista was regarded as an advanced cervoid character in Janis and Scott (1987), while later this  
1356 character is referred to as 'primitive presence of bifurcated protocone'. In extant cervids this feature is  
1357 present in *Odocoileus*, *Blastocerus*, *Alces*, *Mazama*, *Pudu*, and *Capreolus* (Janis and Scott, 1987). These  
1358 observations could be confirmed here by morphological comparisons. One specimen of *Palaeoplatyceros*  
1359 *hispanicus* (MNCN 39181) shows both a bifurcating postprotocrista and a tiny protocone fold on the  
1360 preprotocrista. This indicates that both structures may in fact be developmentally independent, however,  
1361 as this could only be observed in one specimen, it remains speculation.

1362 Throughout the evolutionary history of cervids the lingual cingulum, regularly present on molars and  
1363 sometimes even on premolars of fossil cervids, becomes reduced and eventually lost in extant cervids. In  
1364 *Rucervus*, *Rusa*, and *Axis* the anterior and posterior lingual walls of the molars tend to be indented; this is  
1365 also observed in *Axis lydekkeri*, *Rusa kendagensis*, and '*Cervus*' *sivalensis*.

1366 p2 is the tooth with the fewest changes in occlusal morphology throughout cervid evolution; only a  
1367 shortening is observed in most extant taxa and in a few individuals p2 is lost entirely.

1368 The elongated upper canines in *Hydropotes inermis* are actively used in intraspecific fights. It is likely  
1369 that the presence and/or size of upper canines is somehow genetically linked with the antlers; this brings  
1370 up the question, why female deer have upper canines, too (Brokx, 1972). Even though they are often  
1371 much smaller, especially in species, where males have enlarged upper canines, they are present without  
1372 any obvious function. In other ungulates, where males use their canines in intraspecific fights, for example  
1373 in equids, upper and lower canines are lost in almost all females. Much more research is needed to find  
1374 this link and associated interactions and effects on behaviour.

### 1375 **Systematics of Ruminant Families**

1376 Despite decades of research the systematic relationships of the six ruminant families, especially among  
1377 the pecoran families have been proven to be difficult (Kraus and Miyamoto, 1991; Cronin et al., 1996;  
1378 Randi et al., 1998; Cap et al., 2002; Hassanin and Douzery, 2003; Hassanin et al., 2012). Particularly,  
1379 the position of Moschidae, Antilocapridae, and Giraffidae remained problematic. Hassanin and Douzery  
1380 (2003) and Price et al. (2005) presented an overview of the systematic relationships of ruminants dating  
1381 back to 1934.

1382 Most recent molecular studies relatively consistently showed that the clade consisting of Moschidae  
1383 plus Bovidae was the sister taxon to Cervidae, which was the sister taxon to Giraffidae, then Antilocapridae;  
1384 Tragulidae was the sister taxon to all of them (Kuznetsova et al., 2005; Marcot, 2007; Agnarsson and  
1385 May-Collado, 2008; Hassanin et al., 2012).

1386 In the molecular topologies here, the systematic relationships among the six ruminant families varied.  
1387 Most variation was observed in the nuclear markers; Cervidae was sometimes unresolved as the sister  
1388 taxon to Antilocapridae, Giraffidae and Bovidae, with Moschidae as the sister taxon to all of them. Most  
1389 often, however, Moschidae and Bovidae were sister taxa to each other with Cervidae as the sister taxon,  
1390 and Antilocapridae and Giraffidae as sister taxa to that clade, either unresolved or as clade.

1391 This demonstrates that the supposed consensus about the systematic relationships among ruminant  
1392 families is an artefact of repeatedly re-analysing identical data sets with similar parameters. More  
1393 and different types of data are needed to solve this problem in a more sophisticated and consistent way,  
1394 particularly because of the potential implications for conservation in some genera (Price et al., 2005). Also,  
1395 further work is needed to investigate the impact of inclusion of fossil taxa (Agnarsson and May-Collado,  
1396 2008; O'Leary and Gatesy, 2008).

### 1397 **Evolutionary History**

1398 During the Eocene, selenodont artiodactyls diversified and ruminants were the only successful descendants  
1399 from this radiation. Subsequent rapid radiations of ruminants resulted in the most diverse group of large  
1400 mammals today (Hernández Fernández and Vrba, 2005).

1401 Collision of the African and Indian continents with Eurasia around 40 mya caused drastic environ-  
1402 mental changes triggering artiodactyl evolution. The expansion and diversification of grasslands at the  
1403 Eocene-Oligocene-boundary (34 mya) coincided with climate changes from warm and humid to colder  
1404 and drier conditions (Prothero and Heaton, 1996; Meng and McKenna, 1998; Hassanin and Douzery,  
1405 2003). The divergence of major ruminant lineages has occurred within a very short period of time since  
1406 their origination and ruminant evolution rates were not constant through time (DeMiguel et al., 2013).  
1407 From the Oligocene to the mid Pliocene global climatic and vegetational changes led to several successive  
1408 rapid radiations within Pecora with additional short-termed diversification events within Bovidae and  
1409 Cervidae (Hernández Fernández and Vrba, 2005). This rapid cladogenesis and parallel evolution may  
1410 explain the lack of resolution or taxon instability in ruminant topologies and the plethora of convergent  
1411 morphological developments (Hernández Fernández and Vrba, 2005; Janis and Theodor, 2014).

1412 From the Oligocene to the Miocene cooler and more arid climate led to the replacement of forest  
1413 habitats with open grasslands in Asia favouring the diversification and dispersal of many pecoran groups  
1414 (Meijaard and Groves, 2004; Lorenzini and Garofalo, 2015). C3 grass dominated habitats occurred around  
1415 22 mya, C4 grass expanded around 17.5 mya (DeMiguel et al., 2013). These conditions were perfect  
1416 for the origin and diversification of Cervidae and other ruminant groups. The resulting competition of  
1417 overlapping habitats of grazers and browsers must have played a crucial role in the evolution of Cervidae  
1418 (Gilbert et al., 2006). At the Oligocene-Miocene boundary, the first cervoids appeared diverging from  
1419 Oligocene taxa like *Dremotherium* or *Bedenomeryx* (Ludt et al., 2004). The antlerless *Dremotherium*

1420 from the early Miocene of Europe has been suggested as the earliest member of cervids (Brooke, 1878;  
1421 Ginsburg and Heintz, 1966; Vislobokova, 1983). *Dremotherium* was consistently found to be most similar  
1422 to cervids and together with *Amphitragulus* is now widely considered to be an early cervoid (Heintz et al.,  
1423 1990; Gentry et al., 1999). The exact systematics of *Dremotherium feignouxi* remain problematic as it  
1424 shares morphological traits with cervids and moschids (Pomel, 1853; Costeur, 2011). In the analyses here,  
1425 *Dremotherium feignouxi* was most often placed in an unresolved position, confirming its controversial  
1426 affinities.

1427 Although Central Asia/Eastern Eurasia has been long regarded as the centre of origin and evolution of  
1428 Cervidae (Vislobokova, 1990; Groves, 2006), evidence from the fossil record indicated that the origin of  
1429 cervids may be in Europe (Heckeberg, 2017b). Their past diversity is known from around 26 fossil genera  
1430 (Dong, 1993). Gilbert et al.'s (2006) reconstruction of the ancestral cervine, which was reconstructed to  
1431 have had antlers with three tines, sexual dimorphism, moderately sized upper canines (smaller than in  
1432 muntjacs), and a deep lacrimal fossa, cannot be confirmed by the fossil record.

1433 In the early Miocene geographical changes played an important role by opening migration routes  
1434 in Europe, Asia, and Africa. This had an rapid increase of ungulate diversity as a consequence, which  
1435 remained like that during the warm climate of the Miocene Climatic Optimum throughout the middle  
1436 Miocene. During the Miocene forest habitats were replaced by grasslands, which favoured the greatest  
1437 radiation of ruminants (Hassanin and Douzery, 2003). Stadler (2011) showed that there was a slight but  
1438 not significant increase in the diversification rate of mammals 15.85 mya. Around 15 mya, the sea-levels  
1439 fell due to cooling climate in the high latitudes and forming ice sheets in the Eastern Antarctic; the fallen  
1440 dry areas became grasslands (Haq et al., 1987; Flower and Kennett, 1994; Miller et al., 1991; Ludt et al.,  
1441 2004).

1442 The climate further cooled causing colder winters and drier summers when the circulation of warm  
1443 deep water between the Mediterranean and the Indo-Pacific was interrupted. Subsequently grasslands  
1444 spread over Europe and Asia between 8 and 7 mya providing perfect conditions for ruminants to further  
1445 diversify (Ludt et al., 2004).

1446 The cooling climate and increased seasonality in the late Miocene likely played a crucial role in the  
1447 decline of large mammal diversity and causing endemism to occur in the climate belts. The lower diversity  
1448 and the endemism of today may have originated already in the late Miocene (12 mya) and may be more  
1449 complex than assumed (to lay in the Quaternary Climatic Cycles) (Costeur and Legendre, 2008). In the  
1450 late Miocene the temperature gradient from equator to pole was weak and higher latitudes were warmer  
1451 than today (Micheels et al., 2011).

1452 During the Late Miocene of Asia environmental changes and uplift of the Tibetan plateau (11–7.5  
1453 mya; Amano and Taira (1992)) coincided with a global increase in aridity, seasonality and subsequent  
1454 spread of grassland in Asia (Flower and Kennett, 1994; Gilbert et al., 2006). A glaciation period at the  
1455 Miocene/Pliocene boundary caused a drop in sea levels triggering further diversification particularly  
1456 within cervids (Ludt et al., 2004). A crucial factor for South East Asian cervid evolution was the split of the  
1457 Indochinese and Sundaic faunistic subregions caused by high sea levels, which cut through the Thai/Malay  
1458 Peninsula during the Early Pliocene separating faunas for the duration of around 1 my (Woodruff, 2003;  
1459 Meijaard and Groves, 2004). After the warm Middle Pliocene, the Pliocene-Pleistocene boundary was  
1460 characterised by drastic cooling (2.4–1.8 Ma) (Meijaard and Groves, 2004).

1461 There is broad consensus that ancestral odocoileine cervids entered America from Siberia via the  
1462 Bering Strait in the late Miocene/early Pliocene (Gustafson, 1985; Webb, 2000; Merino et al., 2005).  
1463 The Bering land bridge disappeared around 9000 years ago with rising sea levels and the formation of  
1464 the Bering Sea ending the faunal exchange between American and North Asia (Ludt et al., 2004). It is  
1465 assumed that their ancestors were Eurasian Pliocene deer with three-tined antlers, such as *Cervavitus*  
1466 (Fry and Gustafson, 1974; Gustafson, 1985). The first (presumed) odocoileine taxa were *Eocoileus* from  
1467 Florida and *Bretzia* from Nebraska (around 5 my old), which are similar to *Pavlodaria* from Northeastern  
1468 Kazakhstan (Fry and Gustafson, 1974; Vislobokova, 1980; Webb, 2000; Gilbert et al., 2006).

1469 The split between Odocoileini and *Rangifer* was suggested to have occurred in the middle Miocene  
1470 between 15.4 and 13.6 mya, although their origins and relationships are unknown; the presence of close  
1471 relatives of *Rangifer* among South American odocoileine fossils from the Pleistocene has been suggested  
1472 (Groves and Grubb, 1987; Douzery and Randi, 1997). Cervids migrated from North to South America via  
1473 the Panamanian bridge 2.5 mya (Plio-Pleistocene boundary) (Webb, 2000; Merino et al., 2005). The split  
1474 of Odocoileini into Blastocercina and Odocoileina was dated to around 3.4 mya. It was hypothesised that

1475 there was a diversification within Odocoileini in North America 5.1 mya, which is also supported by the  
1476 fossil record (Vrba and Schaller, 2000; Gilbert et al., 2006; Hassanin et al., 2012). The first unambiguous  
1477 adult antler fragment of *Odocoileus* is from 3.8–3.4 mya (Gustafson, 1985). The polyphyletic split of the  
1478 *Mazama* species into the two subclades, Blastocerina and Odocoileina, led to the interpretation that South  
1479 America was colonised at least twice. First, by the ancestor of Blastocerina in the Early Pliocene (4.9–3.4  
1480 mya), although this cannot yet be confirmed by the fossil record nor by a certain presence of a connection  
1481 between North and South America. However, a much earlier closure of the Panama Isthmus between 15  
1482 and 13 mya was recently suggested (Montes et al., 2015). The second colonisation was by the ancestor  
1483 of *Mazama americana* and *Odocoileus virginianus* around the Plio-/Pleistocene boundary Gilbert et al.  
1484 (2006). Stadler (2011) reported a significant rate shift of speciation to a decreasing diversification rate at  
1485 3.35 mya, which coincides with high tectonic activity.

1486 Hershkovitz (1982) assumed a small odocoileine ancestor living in North, Central, or South America  
1487 during the Miocene-Pliocene-boundary from which *Mazama* and *Pudu* diverged. This hypothesis sug-  
1488 gested an increase in body size over time in other odocoileines, which is in contrast to the traditional view  
1489 of secondarily dwarfed *Mazama* and *Pudu*. As a logical consequence, the existence of medium sized  
1490 forms during the late Miocene and Pliocene of Asia and North America was assumed, which would be  
1491 the ancestors of the small odocoileines. This is also supported by the fossil record (Webb, 2000). Slightly  
1492 differently, Merino and Rossi (2010) hypothesised that the first deer entering South America were medium  
1493 sized with branched antlers; these presumably diverged into *Mazama* and *Pudu* with simpler antlers, most  
1494 likely independently from each other.

1495 Six fossil cervid genera are known from South America; they include *Agalmaceros* (1.8–0.8 mya),  
1496 *Charitoceros* (1.8–subrecent), *Antifer* (1.2–subrecent), *Epieuryceros* 1.2–subrecent, *Morenelaphus* 0.5–sub-  
1497 recent, and *Paraceros* (0.5–0.2 mya) (Hoffstetter, 1952; Tomiati and Abbazzi, 2002; Merino et al., 2005;  
1498 Merino and Rossi, 2010; Gonzalez et al., 2014). Their fossil record is scarce and thus, the validity of  
1499 some of the species is doubtful (Alcaraz and Zurita, 2004; Menegaz, 2000; Merino and Rossi, 2010). So  
1500 far, there are only few studies on extinct neotropical cervids and even fewer attempting to reconstruct the  
1501 phylogeny of fossil and extant neotropical deer.

1502 Neotropical cervids underwent a rapid radiation after migration into South America, where they  
1503 filled niches, which are occupied by bovids on other continents, making them the most diverse group of  
1504 ungulates in South America (Gilbert et al., 2006; Merino and Rossi, 2010). The low resolution among  
1505 Odocoileini haplotypes also suggests a rapid radiation event dating to about 2.5 mya, which coincides  
1506 with the land mammal invasion from North to South America (Webb, 2000; Gilbert et al., 2006). Today's  
1507 South American cervids are adapted to a wide range of ecological habitats (Merino et al., 2005). The  
1508 radiation most likely was influenced by the absence of other ruminant artiodactyls and appears to be  
1509 the opposite scenario as in Africa, where bovids dominated. Morphology, physiology, adaptation of the  
1510 digestive system, temporal and spatial distribution of vegetation, and physicochemical properties of plants  
1511 triggered the diversification, thus making the evolutionary patterns very complex (Merino and Rossi,  
1512 2010).

1513 The origination of living cervids of South America was estimated to 200 kya for *Hippocamelus*,  
1514 *Blastoceros*, *Ozotoceros*, 65 kya for *Mazama*, 48 kya for *Odocoileus*, and 16 kya for *Pudu* (Merino et al.,  
1515 2005). These recent dates document the rapid radiation of South American cervids, which is probably the  
1516 reason for the difficulties in resolving their relationships. After decades of research, the taxonomy and  
1517 evolutionary history of South American cervids remains enigmatic, partly because of the scarce Plio- and  
1518 Pleistocene fossil record (Fry and Gustafson, 1974; Webb, 2000).

## 1519 CONCLUSION

1520 The comprehensive data collection and results from the phylogenetic analyses provided new insights  
1521 into the systematic relationships of fossil and extant cervids. These relationships were investigated using  
1522 molecular and morphological characters separately and combined.

1523 The morphological data sets were partly informative for extant taxa and gave new insights into the  
1524 systematic relationships of fossil taxa. There were some consistent splits within the morphological  
1525 topologies, for example the *Elaphurus-Rucervus-Rusa*-clade, Muntiacini, and Capreolini. The SFA and  
1526 EPA approaches were particularly useful for investigating the placement of fossil taxa.

1527 In most of the molecular and combined analyses, extant clades on subfamilial and tribal level  
1528 were monophyletic. While systematic relationships within Cervinae were relatively stable, with many

1529 consistently recovered subclades, systematic relationships within Capreolinae were more variable. Even  
1530 the monophyly of this subfamily could not be confirmed in all topologies.

1531 No link between particularly incomplete taxa and phylogenetic instability was observed. For the  
1532 Miocene cervids, a placement in a stem position between the outgroup and all other cervids, or in a sister  
1533 position to Muntiacini was suggested in the analyses here. Most of the Miocene cervids were more closely  
1534 related to each other than to other cervids. Plio- and Pleistocene cervids, were most often placed within or  
1535 close to extant cervids and the majority of them within Cervini, some within Capreolinae. or Muntiacini.

1536 We extensively tested the systematic positions of extant and especially fossil cervids for the first time  
1537 under a comprehensive phylogenetic approach. Inclusion of more fossil cervids, postcranial characters,  
1538 soft anatomy and life history data, and cytogenetics would be useful in future analyses. Further, rare  
1539 genomic changes, such as gene duplication and genetic code changes, intron indels, and mitochondrial  
1540 gene order changes, and SNP chips have become more popular as complementary markers and should be  
1541 included as addition to the molecular partition in cervids.

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## 1550 REFERENCES

- 1551 Abril, V. V., Carnelossi, E. a. G., González, S., and Duarte, J. M. B. (2010). Elucidating the evolution of  
1552 the red brocket deer *Mazama americana* complex (Artiodactyla; Cervidae). *Cytogenetic and Genome*  
1553 *Research*, 128:177–187.
- 1554 Agnarsson, I. and May-Collado, L. J. (2008). The phylogeny of Cetartiodactyla: The importance of  
1555 dense taxon sampling, missing data, and the remarkable promise of cytochrome b to provide reliable  
1556 species-level phylogenies. *Molecular Phylogenetics and Evolution*, 48:964–985.
- 1557 Alcaraz, M. A. and Zurita, A. E. (2004). Nuevos registros de cérvidos poco conocidos: *Epieuryceros* cf.  
1558 *proximus* Castellanos y *Antifer* sp. (Mammalia, Artiodactyla, Cervidae). *Revista del Museo Argentino*  
1559 *de Ciencias Naturales, n. s.*, 6(1):41–48.
- 1560 Amano, K. and Taira, A. (1992). Two-phase uplift of higher himalayas since 17 ma. *Geology*, 20(5):391–  
1561 394.
- 1562 Amato, G., Egan, M. G., and Rabinowitz, A. (1999a). A new species of muntjac, *Muntiacus putaensis*  
1563 (artiodactyla: Cervidae) from northern myanmar. *Animal Conservation*, 2(1):1–7.
- 1564 Amato, G., Egan, M. G., Schaller, G. B., Baker, R. H., Rosenbaum, H. C., Robichaud, W. G., and  
1565 DeSalle, R. (1999b). Rediscovery of roosevelt's barking deer (*Muntiacus rooseveltorum*). *Journal of*  
1566 *Mammalogy*, 80(2):639–643.
- 1567 Angom, S. and Hussain, S. A. (2013). A review on genetic status of eld's deer *Rucervus eldii*: With notes  
1568 on distribution, population status and future perspectives. *Octa Journal of Environmental Research*,  
1569 1(2).
- 1570 Arif, M., Shah, S., and Vos, J. (1991). *Cervus rewati* sp. nov (mammalia, cervidae) from the upper  
1571 siwaliks of pakistan. *Geological Survey of Pakistan, Memoirs*, 17.
- 1572 Asher, R. J. (2007). A web-database of mammalian morphology and a reanalysis of placental phylogeny.  
1573 *BMC evolutionary biology*, 7:108.
- 1574 Azanza, B. (1993a). Sur la nature des appendices frontaux des cervidès (artiodactyla, mammalia) du  
1575 miocène inférieur et moyen. remarques sur leur systématique et leur phylogeny. *Comptes Rendus de*  
1576 *l'Académie des Sciences de Paris, Série II*, 316:1163–1169.
- 1577 Azanza, B. (1993b). Systématique et évolution du genre *Procervulus*, cervidé (artiodactyla, mammalia)  
1578 du miocène inférieur d'europe. *Comptes Rendus de l'Académie des Sciences de Paris, Série II*,  
1579 316:717–723.

- 1580 Azanza, B., DeMiguel, D., and Andrés, M. (2011). The antler-like appendages of the primitive deer  
1581 *Dicrocerus elegans*: morphology, growth cycle, ontogeny, and sexual dimorphism. *Estudios Geológicos*,  
1582 67(2):579–602.
- 1583 Azanza, B. and Ginsburg, L. (1997). A revision of the large lagomerycid artiodactyls of Europe. *Palaeon-*  
1584 *tology*, 40:461–485.
- 1585 Azanza, B. and Montoya, P. (1995). A new deer from the Lower Turolian of Spain. *Journal of Paleontology*,  
1586 69(6):1163–1175.
- 1587 Azanza, B. and Morales, J. (1989). Los artiodáctilos de huélago, huéscar-1 y cullar de baza-1 (cuenca de  
1588 guadix-baza, granada). *Trabajos sobre el Neógeno-Cuaternario*, 11:289–316.
- 1589 Azanza, B., Rössner, G. E., and Ortiz-Jaureguizar, E. (2013). The early Turolian (late Miocene) Cervidae  
1590 (Artiodactyla, Mammalia) from the fossil site of Dorn-Dürkheim 1 (Germany) and implications on the  
1591 origin of crown cervids. *Palaeobiodiversity and Palaeoenvironments*, 93:217–258.
- 1592 Azanza Asensio, B. (2000). Los cervidae (artiodactyla, mammalia) del mioceno de las cuencas del duero,  
1593 tajo, calatayud-teruel y levante. *Memorias del Museo Paleontológico de la Universidad de Zaragoza*,  
1594 8:376.
- 1595 Azanza Asensio, B. and Menendez, E. (1989). Los ciervos fósiles del neógeno español. *Paleontologia i*  
1596 *evolució*, 23:75–82.
- 1597 Azzaroli, A. (1953). The deer of the Weybourn Crag and Forest Bed of Norfolk. *Bulletin of the British*  
1598 *Museum (Natural History)*, 2(i):1–96.
- 1599 Azzaroli, A. (1954). Critical observations upon siwalik deer. *Proceedings of the Linnean Society of*  
1600 *London*, 165(1):75–83.
- 1601 Azzaroli, A., De Giuli, C., Ficarelli, G., and Torre, D. (1988). Late pliocene to early mid-pleistocene  
1602 mammals in Eurasia: faunal succession and dispersal events. *Palaeogeography, Palaeoclimatology,*  
1603 *Palaeoecology*, 66(1-2):77–100.
- 1604 Azzaroli, A. and Mazza, P. (1992a). The cervid genus *Eucladoceros* in the early pleistocene of tuscany.  
1605 *Palaeontographia Italica*, 79:43–100.
- 1606 Azzaroli, A. and Mazza, P. (1992b). On the possible origin of the giant deer genus *Megaceroides*.  
1607 *Rendiconti Lincei*, 3(1):23–32.
- 1608 Azzaroli, A. and Mazza, P. (1993). Large early pleistocene deer from Pietrafitta lignite mine, central Italy.  
1609 *Palaeontographia Italica*, 80:1–24.
- 1610 Balakrishnan, C. N., Monfort, S. L., Gaur, A., Singh, L., and Sorenson, M. D. (2003). Phylogeography  
1611 and conservation genetics of eld's deer (*Cervus eldi*). *Molecular Ecology*, 12(1):1–10.
- 1612 Beninde, J. (1937). Der Merkmalswert des Hirschgeweihs für die paläontologische Systematik der Genus  
1613 *Cervus*. *Paläontologische Zeitschrift*, 19(1-2):52–56.
- 1614 Berger, S. A., Krompass, D., and Stamatakis, A. (2011). Performance, accuracy, and web server for  
1615 evolutionary placement of short sequence reads under maximum likelihood. *Systematic biology*,  
1616 60(3):291–302.
- 1617 Bibi, F. (2014). Assembling the ruminant tree: combining morphology, molecules, extant taxa, and fossils.  
1618 *Zitteliana B*, 32(September 2013):197–212.
- 1619 Bibi, F., Vrba, E., and Fack, F. (2012). A new African fossil caprin and a combined molecular and mor-  
1620 phological Bayesian phylogenetic analysis of Caprini (Mammalia: Bovidae). *Journal of Evolutionary*  
1621 *Biology*, 25:1843–1854.
- 1622 Blyth, E. (1863). On some horns of ruminants. *Proceedings of the Zoological Society London*, page 155.
- 1623 Böhme, M., Aiglstorfer, M., Uhl, D., and Kullmer, O. (2012). The antiquity of the Rhine River:  
1624 Stratigraphic coverage of the Dinotheriensande (Eppelsheim Formation) of the Mainz basin (Germany).  
1625 *PLoS ONE*, 7(5).
- 1626 Bouvrain, G., Geraads, D., and Jehenne, Y. (1989). Nouvelles données relatives à la classification des  
1627 Cervidae (Artiodactyla, Mammalia). *Zoologischer Anzeiger*, 223:82–90.
- 1628 Brox, P. A. (1972). The superior canines of *Odocoileus* and other deer. *Journal of Mammalogy*,  
1629 53(2):359–366.
- 1630 Brooke, V. (1878). On the classification of Cervidae with a synopsis of the existing species. *Proc. Zool.*  
1631 *Soc. London*, 1878(1878):883–928.
- 1632 Bubenik, G. and Bubenik, A. B. (1986). Phylogeny and ontogeny of antlers and neuro-endocrine regulation  
1633 of the antler cycle - a review. *Säugetierkundliche Mitteilungen*, 33:97–123.
- 1634 Canto, J., Yáñez, J., and Rovira, J. (2010). Estado actual del conocimiento de los mamíferos fósiles de

- 1635 Chile. *Estudios Geológicos*, 66(2):255–284.
- 1636 Cap, H., Aulagnier, S., and Deleporte, P. (2002). The phylogeny and behaviour of Cervidae (Ruminantia  
1637 Pecora). *Ethology Ecology & Evolution*, 14:199–216.
- 1638 Carr, S. M., Ballinger, S. W., Derr, J. N., Blankenship, L. H., and Bickham, J. W. (1986). Mitochondrial dna  
1639 analysis of hybridization between sympatric white-tailed deer and mule deer in west texas. *Proceedings*  
1640 *of the National Academy of Sciences*, 83(24):9576–9580.
- 1641 Chow, B.-s. and Shih, M.-c. (1978). A skull of *Lagomeryx* from the middle miocene of linchu, shantung.  
1642 *Vertebrata Palasiatica*, 16(2):111–122.
- 1643 Cohen, K., Finney, S., Gibbard, P., and Fan, J.-X. (2013). The ics international chronostratigraphic chart.  
1644 *Episodes*, 36(3):199–204.
- 1645 Costeur, L. (2011). A partial skull of *Dremotherium feignouxi* from the aquitanian of france (mn2,  
1646 saint-gérard-le-puy, allier). *Acta Geologica Slovaca*, 3(2):105–112.
- 1647 Costeur, L. and Legendre, S. (2008). Spatial and temporal variation in European Neogene large mammals  
1648 diversity. *Palaeogeography, Palaeoclimatology, Palaeoecology*, 261:127–144.
- 1649 Croitor, R. (2006a). Early Pleistocene small-sized deer of Europe. *Hellenic Journal of Geosciences*,  
1650 41:89–117.
- 1651 Croitor, R. (2006b). Taxonomy and systematics of large-sized deer of the genus *Praemegaceros* Portis,  
1652 1920 (Cervidae, Mammalia). *CFS Courier Forschungsinstitut Senckenberg*, pages 91–116.
- 1653 Croitor, R. (2009). Systematical Position and Evolution of the genus *Arvernoceros* (Cervidae, Mammalia)  
1654 from Plio-Pleistocene of Eurasia. *Muzeul Olteniei Craiova. Oltenia. Studii si comunicari. S tiintele*  
1655 *Naturii*, 25:375–382.
- 1656 Croitor, R. (2014). Deer from Late Miocene to Pleistocene of Western Palearctic: matching fossil record  
1657 and molecular phylogeny data. *Zitteliana B*, 32:115–154.
- 1658 Croitor, R. (2018). *Plio-Pleistocene Deer of Western Palearctic: Taxonomy, Systematics, Phylogeny*.  
1659 Institute of Zoology of the Academy of Sciences of Moldova.
- 1660 Croitor, R. and Bonifay, M.-F. (2001). Étude préliminaire des cerfs du gisement pléistocène inférieur de  
1661 ceysaguet (haute-loire). *PALEO*, 13:129–144.
- 1662 Croitor, R., Bonifay, M.-F., and Bonifay, E. (2006). Origin and evolution of the Late Pleistocene island  
1663 deer *Praemegaceros (Nesoleipoceros) cazioti* (Dépéret) from Corsica and Sardinia. *Bulletin du Musée*  
1664 *d'Anthropologie préhistorique de Monaco*, 46:35–68.
- 1665 Cronin, M. A. (1991). Mitochondrial-DNA Phylogeny of Deer (Cervidae). *Journal of Mammalogy*,  
1666 72:553–566.
- 1667 Cronin, M. A., Stuart, R., Pierson, B. J., and Patton, J. C. (1996). K-casein gene phylogeny of higher  
1668 ruminants (pecora, artiodactyla). *Molecular Phylogenetics and Evolution*, 6(2):295–311.
- 1669 Cronin, M. A., Vyse, E. R., and Cameron, D. G. (1988). Genetic relationships between mule deer and  
1670 white-tailed deer in montana. *The Journal of Wildlife Management*, pages 320–328.
- 1671 Czyżewska, T. (1959). *Cervus (Rusa) sp. z plioceńskiej brekacji kostnej z weżów*. *Acta Palaeontologica*  
1672 *Polonica*, 4(4):389–429.
- 1673 Czyżewska, T. (1968). Deer from weże and their relationship with the pliocene and recent eurasiatic  
1674 cervidae. *Acta Palaeontologica Polonica*, 8(4):537–593.
- 1675 De Serres, M. M. (1832). *Recherches sur les ossements humatiles de la caverne de Lunel-Viel*. Boehm,  
1676 Montpellier.
- 1677 De Vos, J. (1979). The endemic pleistocene deer of crete. *Proceedings of the Koninklijke Nederlandse*  
1678 *Akademie van Wetenschappen Amsterdam, Series B*, 82(1):59–90.
- 1679 De Vos, J. (1984). The endemic pleistocene deer of crete. *Verhandelingen der Koninklijke Nederlandse*  
1680 *Akademie van Wetenschappen, afd. Natuurkunde, eerste reeks*, 31:1–110.
- 1681 De Vos, J. (2000). Pleistocene deer fauna in crete: Its adaptive radiation and extinction. *Tropics*,  
1682 10(1):125–134.
- 1683 De Vos, J. and Reumer, J. W. F. (1995). Early Pleistocene Cervidae (Mammalia, Artiodactyla) from the  
1684 Oosterschelde (the Netherlands), with a revision of the cervid genus *Eucladoceros* Falconer, 1868.  
1685 *Deinsea*, pages 95–121.
- 1686 DeMiguel, D., Azanza, B., and Morales, J. (2013). Key innovations in ruminant evolution: a paleontologi-  
1687 cal perspective. *Integrative zoology*, pages 1–52.
- 1688 DeMiguel, D., Fortelius, M., Azanza, B., and Morales, J. (2008). Ancestral feeding state of ruminants  
1689 reconsidered: earliest grazing adaptation claims a mixed condition for Cervidae. *BMC evolutionary*



- 1690 *biology*, 8:13.
- 1691 Deng, T., Lu, X.-K., Shi, Q.-Q., Sun, B.-Y., and Wang, S.-Q. (2014). A new species of crown-antlered  
1692 deer *Stephanocemas* (Cervidae, Artiodactyla) from the Middle Miocene of the Linxia Basin in Gansu,  
1693 China. *Vertebrata Palasiatica*, 52(2):171–182.
- 1694 Depéret, C. (1884). Nouvelles études sur les ruminants pliocènes et quaternaires d’auvergne. *Bulletin de*  
1695 *la Société Géologique de France, Série 3*, T12:247–284.
- 1696 Depéret, C. and Mayet, L. (1911). Le gisement de senèze et sa faune paléomammalogique. *Association*  
1697 *Française pour l’Avancement des Sciences, Comptes rendus 39e session, Toulouse, 1910, Notes et*  
1698 *Mémoires*, 2:261–263.
- 1699 Di Stefano, G. and Petronio, C. (2002). Systematics and Evolution of the Eurasian Plio-Pleistocene tribe  
1700 Cervini (Artiodactyla, Mammalia). *Geologica Romana*, 36:311–334.
- 1701 Dietrich, W. O. (1938). Zur kenntnis der o-pliozänen echten hirsche. *Zeitschrift der Deutschen Geologis-*  
1702 *chen Gesellschaft*, 90.
- 1703 Dong, W. (1993). The Fossil Records of Deer in China. In Ohtaishi, N. and Sheng, H.-L., editors, *Deer of*  
1704 *China*, pages 95–102. Elsevier Science Publishers.
- 1705 Dong, W. (2007). New material of Muntiacinae (Artiodactyla, Mammalia) from the Late Miocene of the  
1706 northeastern Qinghai-Tibetan Plateau, China. *Comptes Rendus - Palevol*, 6:335–343.
- 1707 Dong, W., Pan, Y., and Liu, J. (2004). The earliest *Muntiacus* (Artiodactyla, Mammalia) from the Late  
1708 Miocene of Yuanmou, southwestern China. *Comptes Rendus Palevol*, 3:379–386.
- 1709 Douzery, E. and Randi, E. (1997). The mitochondrial control region of cervidae: evolutionary patterns  
1710 and phylogenetic content. *Molecular Biology and Evolution*, 14(11):1154–1166.
- 1711 Duarte, J. M. B., Abril, V. V., Vogliotti, A., dos Santos Zanetti, E., de Oliveira, M. L., Tiepolo, L. M.,  
1712 Rodrigues, L. F., and de Almeida, L. B. (2012a). Avaliação do risco de extinção do veado-cambuta  
1713 *Mazama nana* hensel, 1872, no brasil. *Biodiversidade Brasileira*, 2(3):59–67.
- 1714 Duarte, J. M. B., González, S., and Maldonado, J. E. (2008). The surprising evolutionary history of South  
1715 American deer. *Molecular phylogenetics and evolution*, 49:17–22.
- 1716 Duarte, J. M. B. and Merino, M. (1997). Taxonomia e evolução. In Duarte, J. M. B., editor, *Biologia e*  
1717 *conservacao de Cêrvidos Sul-americanos: Blastocerus, Ozotoceros e Mazama*, pages 2–21. FUNEP,  
1718 Jaboticabal, Brasil.
- 1719 Duarte, J. M. B., Vogliotti, A., dos Santos Zanetti, E., de Oliveira, M. L., Tiepolo, L. M., Rodrigues,  
1720 L. F., and de Almeida, L. B. (2012b). Avaliação do risco de extinção do veado-catingueiro *Mazama*  
1721 *gouazoubira* g. fischer [von waldhein], 1814, no brasil. *Biodiversidade Brasileira*, 2(3):50–58.
- 1722 Duarte, J. M. B., Vogliotti, A., dos Santos Zanetti, E., de Oliveira, M. L., Tiepolo, L. M., Rodrigues,  
1723 L. F., and de Almeida, L. B. (2012c). Avaliação do risco de extinção do veado-galheiro *Odocoileus*  
1724 *virginianus* zimmermann, 1780, no brasil. *Biodiversidade Brasileira*, 2(3):15–19.
- 1725 Duarte, J. M. B., Vogliotti, A., dos Santos Zanetti, E., de Oliveira, M. L., Tiepolo, L. M., Rodrigues, L. F.,  
1726 and de Almeida, L. B. (2012d). Avaliação do risco de extinção do veado-mateiro-pequeno *Mazama*  
1727 *bororo* duarte, 1996, no brasil. *Biodiversidade Brasileira*, 2(3):42–49.
- 1728 Duarte, J. M. B., Vogliotti, A., dos Santos Zanetti, E., de Oliveira, M. L., Tiepolo, L. M., Rodrigues, L. F.,  
1729 and de Almeida, L. B. (2012e). Avaliação do risco de extinção do veado-mateiro *Mazama americana*  
1730 *erxleben*, 1777, no brasil. *Biodiversidade Brasileira*, 2(3):33–41.
- 1731 Duarte, J. M. B., Vogliotti, A., dos Santos Zanetti, E., de Oliveira, M. L., Tiepolo, L. M., Rodrigues, L. F.,  
1732 and de Almeida, L. B. (2012f). Avaliação do risco de extinção do veado-roxo *Mazama nemorivaga*  
1733 *cuvier*, 1817, no brasil. *Biodiversidade Brasileira*, 2(3):68–73.
- 1734 Dubois, E. (1904). Over een equivalent van het cromer forest-bed in nederland. *Verslag van de Gewone*  
1735 *Vergadering der Wis- en Natuurkunde Afdeling van de Koninklijke Akademie van Wetenschappen te*  
1736 *Amsterdam*, 13:243–251.
- 1737 Emerson, B. and Tate, M. (1993). Genetic analysis of evolutionary relationships among deer (subfamily  
1738 cervinae). *Journal of Heredity*, 84(4):266–273.
- 1739 Escamilo, L., Barrio, J., Benavides, J., and Tirira, D. (2010). Northern pudu *Pudu mephistophiles* (De  
1740 Winton 1896). *Neotropical cervidology. Biology and medicine of Latin American deer.*, (1975):133–139.
- 1741 Escobedo-Morales, L. A., Mandujano, S., Eguarte, L. E., Rodríguez-Rodríguez, M. A., and Maldonado,  
1742 J. E. (2016). First phylogenetic analysis of mesoamerican brocket deer *Mazama pandora* and *Mazama*  
1743 *temama* (cetartiodactyla: Cervidae) based on mitochondrial sequences: Implications on neotropical  
1744 deer evolution. *Mammalian Biology-Zeitschrift für Säugetierkunde*, 81(3):303–313.

- 1745 Flerov, K. K. (1952). *Fauna of the USSR, Mammals 1(2). Musk deer and deer*. The Academy of Sciences  
1746 of the USSR, Moscow.
- 1747 Flower, B. P. and Kennett, J. P. (1994). The middle miocene climatic transition: East antarctic ice sheet  
1748 development, deep ocean circulation and global carbon cycling. *Palaeogeography, palaeoclimatology,*  
1749 *palaeoecology*, 108(3-4):537–555.
- 1750 Flueck, W. T. and Smith-Flueck, J. M. (2011). Osteological comparisons of appendicular skeletons:  
1751 A case study on Patagonian huemul deer and its implications for conservation. *Animal Production*  
1752 *Science*, 51:327–339.
- 1753 Franzmann, A. W. (1981). *Alces alces. Mammalian Species*, 154:1–7.
- 1754 Fry, W. and Gustafson, E. (1974). Cervids from the Pliocene and Pleistocene of central Washington.  
1755 *Journal of Paleontology*, 48(2):375–386.
- 1756 Geisler, J. H., McGowen, M. R., Yang, G., and Gatesy, J. (2011). A supermatrix analysis of genomic,  
1757 morphological, and paleontological data from crown Cetacea. *BMC evolutionary biology*, 11(1):112.
- 1758 Geist, V. (1966). The evolution of horn-like organs. *Behaviour*, 27:175–214.
- 1759 Geist, V. (1998). *Deer of the world: their evolution, behaviour, and ecology*. Stackpole books, Mechan-  
1760 icsburg.
- 1761 Gentry, A., Rössner, G., and Heizmann, E. P. J. (1999). Suborder Ruminantia. In Rössner, G. and  
1762 Heissig, K., editors, *The Miocene Land Mammals of Europe*, pages 225–258. Verlag Dr. Friedrich Pfeil,  
1763 München.
- 1764 Gentry, A. W. (1994). The Miocene differentiation of old world Pecora (Mammalia). *Historical Biology*,  
1765 7:115–158.
- 1766 Gentry, A. W. (2000). The ruminant radiation. In Vrba, E. S. and Schaller, G. B., editors, *Antelopes, deer,*  
1767 *and relatives*, pages 11–25. Yale University Press, New Haven.
- 1768 Gentry, A. W. (2005). Ruminants of Ruabánya. *Palaeontographica Italica*, 90:283–303.
- 1769 Gervais, M. P. (1848–1852). *Zoologie et paléontologie françaises (animaux vertébrés) ou nouvelles*  
1770 *recherches sur les animaux et fossiles de la France*, volume t.1. Arthus Bertrand, Paris.
- 1771 Giao, P. M., Tuoc, D., Dung, V., Wikramanayake, E., Amato, G., Arctander, P., and MacKinnon, J. (1998).  
1772 Description of *Muntiacus truongsoneis*, a new species of muntjac (artiodactyla: Muntiacidae) from  
1773 central vietnam, and implications for conservation. *Animal Conservation*, 1(01):61–68.
- 1774 Gilbert, C., Ropiquet, A., and Hassanin, A. (2006). Mitochondrial and nuclear phylogenies of Cervidae  
1775 (Mammalia, Ruminantia): Systematics, morphology, and biogeography. *Molecular Phylogenetics and*  
1776 *Evolution*, 40:101–117.
- 1777 Ginsburg, L. (1985). Essai de phylogénie des eupecora (ruminantia, artiodactyla, mammalia). *Comptes*  
1778 *Rendus de l'Académie des Sciences de Paris, Série II*, 301(17):319–322.
- 1779 Ginsburg, L. and Azanza, B. (1991). Présence de bois chez les femelles du cervidé miocène *Dicrocerus*  
1780 *elegans* et remarques sur le problème de l'origine du dimorphisme sexuel sur les appendices frontaux  
1781 des Cervidae. *Comptes Rendus de l'Académie des Sciences de Paris, Série II*, 313:121–126.
- 1782 Ginsburg, L. and Heintz, E. (1966). Sur les affinités du genre *Palaeomeryx* (ruminant du miocène  
1783 européen). *Comptes rendus de l'Académie des sciences, Série D*, 262:979–982.
- 1784 Godina, A. Y., Gromova, V. I., Sokolov, I. I., Trofimov, B. A., Flerov, K. K., and Haveson, Y. I. (1962).  
1785 Order artiodactyla, even-toed ungulates. In Gromova, V., editor, *Basics of Palaeontology: Mammals*,  
1786 pages 337–341. Moscow.
- 1787 Gonzalez, E., Labarca, R., Chavez-Hoffmeister, M., and Pino, M. (2014). First Fossil Record of the  
1788 Smallest Deer cf . *Pudu* Molina , 1782 (Artiodactyla, Cervidae), in the Late Pleistocene of South  
1789 America. *Journal of Vertebrate Paleontology*, 1782(March):483–488.
- 1790 Gonzalez, S., Maldonado, J. E., Leonard, J. A., Vilà, C., Barbanti Duarte, J. M. B., Merino, M., Brum-  
1791 Zorrilla, N., and Wayne, R. (1998). Conservation genetics of the endangered pampas deer (*Ozotoceros*  
1792 *bezoarticus*). *Molecular Ecology*, 7(1):47–56.
- 1793 González, S., Maldonado, J. E., Ortega, J., Talarico, A. C., Bidegaray-Batista, L., Garcia, J. E., and  
1794 Duarte, J. M. B. (2009). Identification of the endangered small red brocket deer (*Mazama bororo*) using  
1795 noninvasive genetic techniques (mammalia; cervidae). *Molecular ecology resources*, 9(3):754–758.
- 1796 Goss, R. J. (1983). *Deer antlers: regeneration, function and evolution*. Academic Press, Inc., New York.
- 1797 Gould, S. J. (1974). The origin and function of 'bizarre' structures: antler size and skull size in the 'irish  
1798 elk', *Megaloceros giganteus*. *Evolution*, 28(2):191–220.
- 1799 Gouy, M., Guindon, S., and Gascuel, O. (2010). Seaview version 4: a multiplatform graphical user

- 1800 interface for sequence alignment and phylogenetic tree building. *Molecular biology and evolution*,  
1801 27(2):221–224.
- 1802 Groves, C. (2006). The genus *Cervus* in eastern Eurasia. *European Journal of Wildlife Research*, 52:14–22.
- 1803 Groves, C. (2014). Current taxonomy and diversity of crown ruminants above the species level. *Zitteliana*  
1804 B, 32:5–14.
- 1805 Groves, C. and Grubb, P. (1987). Relationships of living deer. In CM, W., editor, *Biology and management*  
1806 *of the Cervidae*, pages 21–59. Smithsonian Institution Press, Washington, DC.
- 1807 Groves, C. P. (2007). Family Cervidae. In Prothero, D. R. and Foss, S. E., editors, *The Evolution of*  
1808 *Artiodactyls*, pages 249–256. Johns Hopkins University Press, Baltimore and London.
- 1809 Groves, C. P. and Grubb, P. (1990). Muntiacidae. In Bubenik, G. and Bubenik, A., editors, *Horns,*  
1810 *pronghorns, and antlers: Evolution, morphology, physiology, and social significance*, pages 134–168.  
1811 Springer.
- 1812 Groves, C. P. and Grubb, P. (2011). *Ungulate taxonomy*. Johns Hopkins University Press, Baltimore.
- 1813 Grubb, P. (1990). Cervidae of southeast asia. In Bubenik, G. and Bubenik, A., editors, *Horns, pronghorns,*  
1814 *and antlers: Evolution, morphology, physiology, and social significance*, pages 169–179. Springer.
- 1815 Gühler, U. (1936). Beitrag zur geschichte von *Cervus (Rucervus) schomburgki* blyth. *Zeitschrift für*  
1816 *Säugetierkunde*, 11:155.
- 1817 Gustafson, E. P. (1985). Antlers of *Bretzia* and *Odocoileus* (mammalia, cervidae) and the evolution of  
1818 new world deer. *Transactions of the Nebraska Academy of Sciences*, 13:83–92.
- 1819 Gutiérrez, E. E., Maldonado, J. E., Radosavljevic, A., Molinari, J., Patterson, B. D., Martínez-C, J. M.,  
1820 Rutter, A. R., Hawkins, M. T., Garcia, F. J., and Helgen, K. M. (2015). The taxonomic status of *Mazama*  
1821 *bricenii* and the significance of the táchira depression for mammalian endemism in the cordillera de  
1822 mérida, venezuela. *PLoS ONE*, 10(6):e0129113.
- 1823 Haq, B. U., Hardenbol, J., and Vail, P. R. (1987). Chronology of fluctuating sea levels since the triassic.  
1824 *Science*, 235(4793):1156–1167.
- 1825 Hasegawa, M., Kishino, H., and Yano, T. (1985). Dating of human-ape splitting by a molecular clock of  
1826 mitochondrial dna. *Journal of Molecular Evolution*, 22(2):160–174.
- 1827 Hassanin, A., Delsuc, F., Ropiquet, A., Hammer, C., Jansen Van Vuuren, B., Matthee, C., Ruiz-Garcia,  
1828 M., Catzeflis, F., Areskoug, V., Nguyen, T. T., and Couloux, A. (2012). Pattern and timing of  
1829 diversification of Cetartiodactyla (Mammalia, Laurasiatheria), as revealed by a comprehensive analysis  
1830 of mitochondrial genomes. *Comptes Rendus - Biologies*, 335:32–50.
- 1831 Hassanin, A. and Douzery, E. J. P. (2003). Molecular and morphological phylogenies of ruminantia and  
1832 the alternative position of the moschidae. *Systematic biology*, 52(2):206–228.
- 1833 Heckeberg, N. S. (2017a). *A Comprehensive Approach towards the Systematics and Evolution of Cervidae*.  
1834 PhD thesis, Ludwig-Maximilians-Universität München.
- 1835 Heckeberg, N. S. (2017b). Origination of antlerogenesis. *Journal of morphology*, 278(2):182–202.
- 1836 Heckeberg, N. S., Erpenbeck, D., Wörheide, G., and Rössner, G. E. (2016). Systematic relationships of  
1837 five newly sequenced cervid species. *PeerJ*, 4:e2307.
- 1838 Heintz, E. (1970). Les cervidés villafranchiens de france et d'espagne. *Mémoires du Muséum national*  
1839 *d'histoire naturelle. Série C, Sciences de la terre*, 22(1–2):1–303, 1–206.
- 1840 Heintz, E., Brunet, M., Battail, B., and Jehenne, Y. (1990). The main features of the Cervid palaeobio-  
1841 geography. *Quartärpaläontologie*, 8:79–82.
- 1842 Hernández Fernández, M. and Vrba, E. S. (2005). A complete estimate of the phylogenetic relationships  
1843 in Ruminantia: a dated species-level supertree of the extant ruminants. *Biological reviews of the*  
1844 *Cambridge Philosophical Society*, 80:269–302.
- 1845 Hershkovitz, P. (1972). The recent mammals of the neotropical region: a zoogeographic and ecological  
1846 review. In Keast, A., Erk, F., and Glass, B., editors, *Evolution, mammals and southern continents*,  
1847 pages 311–431. University of New York Press, Albany.
- 1848 Hershkovitz, P. (1982). Neotropical deer (cervidae) part i. pudus, genus *Pudu* gray. *Fieldiana. Zoology,*  
1849 *New Series*, 11:1–86.
- 1850 Hilgen, F., Lourens, L., Van Dam, J., Beu, A., Foyes, A., Cooper, R., Krijgsman, W., Ogg, J., Piller, W.,  
1851 and Wilson, D. (2012). The neogene period. In Gradstein, F., Ogg, J., Schmitz, M., and Ogg, G.,  
1852 editors, *The Geologic Time Scale 2012*, volume 2, pages 923–978. Elsevier, Amsterdam.
- 1853 Hillis, D. M. and Wiens, J. J. (2000). Molecules versus morphology in systematics: conflicts, artifacts,  
1854 and misconceptions. In Wiens, J. J., editor, *Phylogenetic analysis of Morphological data*, pages 1–19.

- 1855 Smithsonian Institution Press, Washington and London.
- 1856 Hoffstetter, R. (1952). Les mammifères pléistocènes de la république de l'équateur. *Mémoires de la*  
1857 *Société Géologique de France Nouvelle Serie*, 11(1-4):1–391.
- 1858 Huelsenbeck, J. P. and Rannala, B. (2000). Using stratigraphic information in phylogenetics. In Wiens,  
1859 J. J., editor, *Phylogenetic analysis of Morphological data*, pages 165–191. Smithsonian Institution  
1860 Press, Washington and London.
- 1861 Hughes, S., Hayden, T. J., Douady, C. J., Tougaard, C., Germonpré, M., Stuart, A., Lbova, L., Carden, R. F.,  
1862 Hänni, C., and Say, L. (2006). Molecular phylogeny of the extinct giant deer, *Megaloceros giganteus*.  
1863 *Molecular Phylogenetics and Evolution*, 40:285–291.
- 1864 IUCN (2016). The iucn red list of threatened species. version 2016.2. Available at [http://](http://www.iucnredlist.org)  
1865 [www.iucnredlist.org](http://www.iucnredlist.org). (accessed on 09 September 2016).
- 1866 Janis, C. M. and Scott, K. M. (1987). The origin of the higher ruminant families with special reference  
1867 to the origin of Cervioidea and relationships within the Cervioidea. *American Museum Novitates*,  
1868 2893:1–85.
- 1869 Janis, C. M. and Scott, K. M. (1988). The phylogeny of the Ruminantia (Artiodactyla, Mammalia). In  
1870 Benton, M. J., editor, *The Phylogeny and Classification of the Tetrapods. Vol. 2. Mammals*, pages  
1871 273–282. Clarendon Press, Oxford.
- 1872 Janis, C. M. and Theodor, J. M. (2014). Cranial and postcranial morphological data in ruminant  
1873 phylogenetics. *Zitteliana B*, 32:1–17.
- 1874 Ji, H. (1985). Geographical history of genus *Elaphurus*. *Vertebrata Palasiatica*, 23(3):215–222.
- 1875 Kitchener, A. (1987). Fighting behaviour of the extinct irish elk. *Modern Geology*, 11(1.28).
- 1876 Koizumi, T., Ohtaishi, N., Kaji, K., Yu, Y., and Tokida, K. (1993). Conservation of white-lipped deer  
1877 in china. In Ohtaishi, N. and Sheng, H.-I., editors, *Deer in China: Biology and Management*, pages  
1878 309–318. Elsevier Science Publishers, Amsterdam, The Netherlands.
- 1879 Korotkevich, E. L. (1970). *Late Neogene Deer of the North Black Sea Area*. Naukova Dumka, Kiev.
- 1880 Kotsakis, T. and Palombo, M. (1979). Vertebrata continentali e paleogeografia della sardegna durante il  
1881 neogene. *Annales Géologiques des Pays Héliéniques, tome hors serie*, 2:621–630.
- 1882 Kraus, F. and Miyamoto, M. M. (1991). Rapid cladogenesis among the pecoran ruminants: evidence from  
1883 mitochondrial dna sequences. *Systematic Biology*, 40(2):117–130.
- 1884 Kuehn, R., Ludt, C. J., Schroeder, W., and Rottmann, O. (2005). Molecular phylogeny of *Megaloceros*  
1885 *giganteus*—the giant deer or just a giant red deer? *Zoological science*, 22:1031–1044.
- 1886 Kurtén, B. (1968). *Pleistocene mammals of Europe*. Weidenfeld and Nicolson, London, UK.
- 1887 Kuss, S. E. (1975). Die pleistozänen hirsche der ostmediterranean inseln kreta, kamos, karpathos und rhodos  
1888 (griechenland). *Berichte der naturforschenden Gesellschaft zu Freiburg im Breisgau*, 65(1-2):25–79.
- 1889 Kuwayama, R. and Ozawa, T. (2000). Phylogenetic relationships among european red deer, wapiti,  
1890 and sika deer inferred from mitochondrial dna sequences. *Molecular phylogenetics and evolution*,  
1891 15(1):115–123.
- 1892 Kuznetsova, M. V., Kholodova, M. V., and Danilkin, a. a. (2005). Molecular phylogeny of deer (Cervidae:  
1893 Artiodactyla). *Russian Journal of Genetics*, 41(7):742–749.
- 1894 Lan, H., Wang, W., and Shi, L. (1995). Phylogeny of *Muntiacus* (cervidae) based on mitochondrial dna  
1895 restriction maps. *Biochemical genetics*, 33(11-12):377–388.
- 1896 Lanfear, R., Calcott, B., Ho, S. Y., and Guindon, S. (2012). Partitionfinder: combined selection of  
1897 partitioning schemes and substitution models for phylogenetic analyses. *Molecular biology and*  
1898 *evolution*, 29(6):1695–1701.
- 1899 Latch, E. K., Heffelfinger, J. R., Fike, J. a., and Rhodes, O. E. (2009). Species-wide phylogeogra-  
1900 phy of North American mule deer (*Odocoileus hemionus*): Cryptic glacial refugia and postglacial  
1901 recolonization. *Molecular Ecology*, 18:1730–1745.
- 1902 Lechner-Doll, M., Lason, K., Lang, D., and Behrend, A. (2001). Evolutionary aspects of dietary selection  
1903 and digestion in the European roe deer. *Mitteilungen des Museum für Naturkunde Berlin, Zoologische*  
1904 *Reihe*, 77(February):223–227.
- 1905 Lee, M. S. Y. and Camens, A. B. (2009). Strong morphological support for the molecular evolutionary  
1906 tree of placental mammals. *Journal of Evolutionary Biology*, 22:2243–2257.
- 1907 Leinders, J. J. M. and Heintz, E. (1980). The configuration of the lacrimal orifice in pecorans and  
1908 tragulids (artiodactyla; mammalia) and its significance for the distinction between bovidae and cervidae.  
1909 *Beaufortia*, 30:155–160.

- 1910 Leslie, D. M. (2010). *Przewalskium albirostre* (Artiodactyla: Cervidae). *Mammalian Species*, 42(849):7–  
1911 18.
- 1912 Leslie, D. M. (2011). *Rusa unicolor* (Artiodactyla: Cervidae). *Mammalian Species*, 43(1):1–30.
- 1913 Leslie, D. M., Lee, D. N., and Dolman, R. W. (2013). *Elaphodus cephalophus* (Artiodactyla: Cervidae).  
1914 *Mammalian Species*, 904(904):80–91.
- 1915 Lewis, P. O. (2001). A likelihood approach to estimating phylogeny from discrete morphological character  
1916 data. *Systematic biology*, 50(6):913–925.
- 1917 Lister, A. (1987). Diversity and evolution of antler form in quaternary deer. In Wemmer, C., editor,  
1918 *Biology and Management of the Cervidae*, pages 81–98. Smithsonian Institution, Washington.
- 1919 Lister, A. (1996). The morphological distinction between bones and teeth of fallow deer (*Dama dama*)  
1920 and red deer (*Cervus elaphus*). *International Journal of Osteoarchaeology*, 6(August 1995):119–143.
- 1921 Lister, A. (1998). The age of early pleistocene mammal faunas from the 'weybourne crag' and cromer  
1922 forest-bed formation (norfolk, england). *Mededelingen Nederlands Instituut voor Toegepaste Ge-*  
1923 *owetenschappen*, 60:271–280.
- 1924 Lister, A. M. (1984). Evolutionary and ecological origins of British deer.
- 1925 Lister, A. M. (1994). The evolution of the giant deer, *Megaloceros giganteus* (Blumenbach).
- 1926 Lister, A. M., Edwards, C. J., Nock, D. A. W., Bunce, M., van Pijlen, I. A., Bradley, D. G., Thomas,  
1927 M. G., and Barnes, I. (2005). The phylogenetic position of the 'giant deer' *Megaloceros giganteus*.  
1928 *Nature*, 438(December):850–853.
- 1929 Lister, A. M., Parfitt, S. A., Owen, F. J., Collinge, S. E., and Breda, M. (2010). Metric analysis of ungulate  
1930 mammals in the early Middle Pleistocene of Britain, in relation to taxonomy and biostratigraphy. II:  
1931 Cervidae, Equidae and Suidae. *Quaternary International*, 228:157–179.
- 1932 Liu, X.-H., Wang, Y.-Q., Liu, Z.-Q., and Zhou, K.-Y. (2002). Phylogenetic relationships of cervinae based  
1933 on sequence of mitochondrial cytochrome b gene. *Zoological Research*, 24(1):27–33.
- 1934 Lizcano, D. J., Álvarez, S. J., Delgado-V, C. A., Duarte, J. M. B., and Gonzales, S. (2010). Dwarf red  
1935 brocket *Mazama rufina* (pucheran 1951). In Duarte, J. M. B. and Gonzalez, S., editors, *Neotropi-*  
1936 *cal Cervidology: Biology and Medicine of Neotropical Deer*, pages 177–180. FUNEP and IUCN,  
1937 Jaboticabal, Brazil.
- 1938 Lönnberg, E. (1906). Which is the taxonomic position of the irish giant deer and allied races? *Arkiv för*  
1939 *Zoologi*, 3(14):1–8.
- 1940 Loomis, F. B. (1928). Phylogeny of the deer. *American Journal of Sciences*, 16:531–542.
- 1941 Lorenzini, R. and Garofalo, L. (2015). Insights into the evolutionary history of *Cervus* (Cervidae, tribe  
1942 Cervini) based on Bayesian analysis of mitochondrial marker sequences, with first indications for a  
1943 new species. *Journal of Zoological Systematics and Evolutionary Research*, 53(4):340–349.
- 1944 Ludt, C. J., Schroeder, W., Rottmann, O., and Kuehn, R. (2004). Mitochondrial DNA phylogeography of  
1945 red deer (*Cervus elaphus*). *Molecular Phylogenetics and Evolution*, 31:1064–1083.
- 1946 Lydekker, R. (1884). Rodents and new ruminants from the siwaliks, and synopsis of mammalia. *Paleon-*  
1947 *tologica Indica*, 10(3):1–5.
- 1948 Lydekker, R. (1898). *Deer of all lands: A history of the family Cervidae living and extinct*. Rowland  
1949 Ward, Ltd., London.
- 1950 Ma, S., Wang, Y., and Xu, L. (1986a). Taxonomic and phylogenetic studies on the genus *Muntiacus*. *Acta*  
1951 *Theriological Sinica*, 6(3):006.
- 1952 Ma, S., Wang, Y., and Xu, L. (1986b). Taxonomic and Phylogenetic Studies on the Genus *Muntiacus*.  
1953 *Acta Theriological Sinica*, VI:190–209.
- 1954 Maddison, W. and Maddison, D. (2011). Mesquite: a modular system for evolutionary analysis. version  
1955 2.75. URL <http://mesquiteproject.org>.
- 1956 Marcot, J. D. (2007). Molecular phylogeny of terrestrial artiodactyls. conflicts and resolution. In DR,  
1957 P. and SE, F., editors, *The evolution of artiodactyls*, pages 4–18. Johns Hopkins University Press,  
1958 Baltimore.
- 1959 Mattioli, S. (2011). Family cervidae (deer). In Wilson, D. E. and Mittermeier, R. A., editors, *Handbook*  
1960 *of the mammals of the world Volume 2*, page 886. Lynx Edicion, Barcelona.
- 1961 Meijaard, E. and Groves, C. (2004). Morphometrical relationships between Southeast Asian deer  
1962 (Cervidae, tribe Cervini): Evolutionary and biogeographic implications. *Journal of Zoology*, 263:179–  
1963 196.
- 1964 Menegaz, A. N. (2000). *Los camélidos y cévidos del cuaternario del sector bonaerense de la Región*

- 1965 *Pampeana*. PhD thesis, Universidad Nacional de La Plata, Argentina.
- 1966 Meng, J. and McKenna, M. C. (1998). Faunal turnovers of palaeogene mammals from the mongolian  
1967 plateau. *Nature*, 394(6691):364–367.
- 1968 Mennecart, B., DeMiguel, D., Bibi, F., Rössner, G. E., Métais, G., Neenan, J. M., Wang, S., Schulz, G.,  
1969 Müller, B., and Costeur, L. (2017). Bony labyrinth morphology clarifies the origin and evolution of  
1970 deer. *Scientific reports*, 7(1):13176.
- 1971 Mennecart, B., Rössner, G. E., Métais, G., DeMiguel, D., Schulz, G., Müller, B., and Costeur, L. (2016).  
1972 The petrosal bone and bony labyrinth of early to middle miocene european deer (mammalia, cervidae)  
1973 reveal their phylogeny. *Journal of Morphology*, 277(10):1329–1338.
- 1974 Merino, M. and Rossi, R. V. (2010). Origin, systematics and morphological radiation. In Duarte, J. M. B.  
1975 and Gonzalez, S., editors, *Neotropical Cervidology: Biology and Medicine of Neotropical Deer*, pages  
1976 2–11. FUNEP and IUCN, Jaboticabal, Brazil.
- 1977 Merino, M. L., Milne, N., and Vizcaíno, S. F. (2005). A cranial morphometric study of deer (Mammalia,  
1978 Cervidae) from Argentina using three-dimensional landmarks. *Acta Theriologica*, 50(1):91–108.
- 1979 Micheels, A., Bruch, A. A., Eronen, J., Fortelius, M., Harzhauser, M., Utescher, T., and Mosbrugger,  
1980 V. (2011). Analysis of heat transport mechanisms from a Late Miocene model experiment with a  
1981 fully-coupled atmosphere-ocean general circulation model. *Palaeogeography, Palaeoclimatology,*  
1982 *Palaeoecology*, 304(3-4):337–350.
- 1983 Mickoleit, G. (2004). *Phylogenetische Systematik der Wirbeltiere*. Pfeil Verlag, München.
- 1984 Miller, K. G., Wright, J. D., and Fairbanks, R. G. (1991). Unlocking the ice house: Oligocene-miocene oxy-  
1985 gen isotopes, eustasy, and margin erosion. *Journal of Geophysical Research: Solid Earth*, 96(B4):6829–  
1986 6848.
- 1987 Miyamoto, M. M., Kraus, F., and Ryder, O. A. (1990). Phylogeny and evolution of antlered deer  
1988 determined from mitochondrial dna sequences. *Proceedings of the National Academy of Sciences*,  
1989 87(16):6127–6131.
- 1990 Montes, C., Cardona, a., Jaramillo, C., Pardo, a., Silva, J. C., Valencia, V., Ayala, C., Pérez-Angel, L. C.,  
1991 Rodriguez-Parra, L. a., Ramirez, V., and Niño, H. (2015). Middle Miocene closure of the Central  
1992 American Seaway. *Science*, 348(6231):226–229.
- 1993 Moscarella, R. a., Aguilera, M., and Escalante, A. a. (2003). Phylogeography, Population Structure, and  
1994 Implications for Conservation of White-Tailed Deer (*Odocoileus virginianus*) in Venezuela. *Journal of*  
1995 *Mammalogy*, 84(4):1300–1315.
- 1996 Moyle, R. G., Andersen, M. J., Oliveros, C. H., Steinheimer, F. D., and Reddy, S. (2012). Phylogeny and  
1997 biogeography of the core babblers (aves: Timaliidae). *Systematic Biology*, 61(785):631–651.
- 1998 O’Leary, M. a. (1999). Parsimony Analysis of Total Evidence from Extinct and Extant Taxa and the  
1999 Cetacean – Artiodactyl Question (Mammalia, Ungulata). *Cladistics*, 15:315–330.
- 2000 O’Leary, M. A. and Gatesy, J. (2008). Impact of increased character sampling on the phylogeny of  
2001 certatiiodactyla (mammalia): combined analysis including fossils. *Cladistics*, 24(4):397–442.
- 2002 O’Reilly, J. E., Puttick, M. N., Parry, L., Tanner, A. R., Tarver, J. E., Fleming, J., Pisani, D., and Donoghue,  
2003 P. C. (2016). Bayesian methods outperform parsimony but at the expense of precision in the estimation  
2004 of phylogeny from discrete morphological data. *Biology letters*, 12(4):20160081.
- 2005 Ouithavon, K., Bhumpakphan, N., Denduangboripant, J., Siriaroonrat, B., and Trakulnaleamsai, S. (2009).  
2006 An analysis of the phylogenetic relationship of thai cervids inferred from nucleotide sequences of  
2007 protein kinase C iota (PRKCI) intron. *Kasetsart Journal - Natural Science*, 43:709–719.
- 2008 Petronio, C., Krakhmalnaya, T., Bellucci, L., and Di Stefano, G. (2007). Remarks on some Eurasian  
2009 pliocervines: Characteristics, evolution, and relationships with the tribe Cervini. *Geobios*, 40:113–130.
- 2010 Pfeiffer, T. (1999). Die stellung von *Dama* (cervidae, mammalia) im system plesiometa carpaler hirsche  
2011 des pleistozäns. phylogenetische rekonstruktion, metrische analyse. *Senckenbergische Naturforschende*  
2012 *Gesellschaft*, 211:1–218.
- 2013 Pfeiffer, T. (2002). The first complete skeleton of *Megaloceros verticornis* (Dawkins, 1868), Cervidae,  
2014 Mammalia, from Bilshausen (Lower Saxony, Germany): description and phylogenetic implications.  
2015 *Mitteilungen aus dem Museum für Naturkunde in Berlin, Geowissenschaftliche Reihe*, 5(July):289–308.
- 2016 Pilgrim, G. E. (1941). The relationship of certain variant fossil types of ‘horn’ to those of the living  
2017 pecora. *The Annals and Magazine of Natural History*, 7(38):172–184.
- 2018 Pitra, C., Fickel, J., Meijaard, E., and Groves, C. P. (2004). Evolution and phylogeny of old world deer.  
2019 *Molecular Phylogenetics and Evolution*, 33:880–895.

- 2020 Pocock, R. I. (1943). The larger deer of British India. Part II. *Journal of the Bombay Natural History*  
2021 *Society*, 43:553–572.
- 2022 Polziehn, R. O. and Strobeck, C. (2002). A phylogenetic comparison of red deer and wapiti using  
2023 mitochondrial dna. *Molecular phylogenetics and evolution*, 22(3):342–356.
- 2024 Pomel, A. (1853). *Catalogue méthodique et descriptif des Vertébrés fossiles du bassin de la Loire*.  
2025 Baillièrre, Paris.
- 2026 Portis, A. (1920). Elenco delle specie di cervicorni fossili in roma e attorno a roma. *Bollettino della*  
2027 *Società Geologica Italiana*, 39:132–139.
- 2028 Price, S. A., Bininda-Emonds, O. R. P., and Gittleman, J. L. (2005). A complete phylogeny of the whales,  
2029 dolphins and even-toed hoofed mammals (Cetartiodactyla). *Biological reviews of the Cambridge*  
2030 *Philosophical Society*, 80(April 2005):445–473.
- 2031 Prothero, D. R. and Heaton, T. H. (1996). Faunal stability during the early oligocene climatic crash.  
2032 *Palaeogeography, Palaeoclimatology, Palaeoecology*, 127(1):257–283.
- 2033 Rabinowitz, A., Myint, T., Khaing, S. T., and Rabinowitz, S. (1999). Description of the leaf deer  
2034 (*Muntiacus putaensis*), a new species of muntjac from northern myanmar. *Journal of Zoology*,  
2035 249(4):427–435.
- 2036 Randi, E., Mucci, N., Claro-Hergueta, F., Bonnet, A., and Douzery, E. J. P. (2001). A mitochondrial  
2037 DNA control region phylogeny of the Cervinae: speciation in *Cervus* and implications for conservation.  
2038 *Animal Conservation*, 4:1–11.
- 2039 Randi, E., Mucci, N., Pierpaoli, M., and Douzery, E. (1998). New phylogenetic perspectives on the  
2040 Cervidae (Artiodactyla) are provided by the mitochondrial cytochrome b gene. *Proceedings. Biological*  
2041 *sciences / The Royal Society*, 265:793–801.
- 2042 Rieseberg, L. H., Archer, M. A., and Wayne, R. K. (1999). Transgressive segregation, adaptation and  
2043 speciation. *Heredity*, 83(4):363–372.
- 2044 Rokas, A. and Holland, P. W. H. (2000). Rare genomic changes as a tool for phylogenetics. *Trends in*  
2045 *Ecology and Evolution*, 15(11):454–459.
- 2046 Romer, A. S. (1966). *Vertebrate Paleontology*. University of Chicago Press, Chicago, Illinois, third  
2047 edition.
- 2048 Ronquist, F., Klopfstein, S., Vilhelmsen, L., Schulmeister, S., Murray, D. L., and Rasnitsyn, A. P. (2012).  
2049 A total-evidence approach to dating with fossils, applied to the early radiation of the hymenoptera.  
2050 *Systematic Biology*, 61(June):973–999.
- 2051 Rössner, G. E. (1995). Odontologische und schädelanatomische untersuchungen an *Procervulus* (cervidae,  
2052 mammalia). *Münchener Geowissenschaftliche Abhandlungen (A)*, 29:127.
- 2053 Samiullah, K. and Akhtar, M. (2007). An evidence of *Cervus punjabensis* from the lower siwaliks of the  
2054 punjab, pakistan. *Punjab University Journal of Zoology*, 22(1-2):63–68.
- 2055 Schaller, G. B. (1996). Description of the giant muntjac (*Megamuntiacus vuquangensis*) in laos. *Journal*  
2056 *of Mammalogy*, 77(3):675–683.
- 2057 Schaub, S. (1941). Die kleine hirschart aus dem oberpliocen von senèze (haute-loire). *Eclogae geologicae*  
2058 *Helveticae*, 34(2):264–272.
- 2059 Schilling, A.-M. and Rössner, G. E. (2017). The (sleeping) beauty in the beast—a review on the water deer,  
2060 *Hydropotes inermis*. *Hystrix, the Italian Journal of Mammalogy*, 28(2).
- 2061 Scott, K. and Janis, C. (1987). Phylogenetic relationships of the cervidae, and the case for a superfamily  
2062 ‘cervoidea’. In Wemmer, C., editor, *Biology and management of the Cervidae*, pages 3–20. Smithsonian  
2063 Institution, Washington, London.
- 2064 Scott, W. B. (1937). *A History of Land Mammals in the Western Hemisphere*. Macmillan Co., New York.
- 2065 Simonelli, V. (1907). Mammiferi quaternari dell’isola di candia. *Memoire, Accademia delle Scienze dell*  
2066 *Instituto di Bologna, Classe di Scienze Fisiche, Serie VI*, 4:251–267.
- 2067 Simonelli, V. (1908). Mammiferi quaternari dell’isola di candia ii. *Memorie, Accademia delle Scienze.*  
2068 *Instituto di Bologna Classe di Scienze Fisiche, Ser. VI*, 5:103–111.
- 2069 Simpson, G. G. (1945). Principles of classification and a classification of mammals. *Bulletin of the*  
2070 *American Museum of Natural History*, 85:1–350.
- 2071 Smith, M. H., Branan, W. V., Marchinton, R. L., Johns, P. E., and Wooten, M. C. (1986). Genetic and  
2072 morphologic comparisons of red brocket, brown brocket, and white-tailed deer. *Journal of Mammalogy*,  
2073 67(1):103–111.
- 2074 Smith, W. P. (1991). *Odocoileus virginianus*. *Mammalian species*, 388:1–13.

- 2075 Solounias, N. (1988). Evidence from horn morphology on the phylogenetic relationships of the pronghorn  
2076 (*Antilocapra americana*). *Journal of mammalogy*, 69(1):140–143.
- 2077 Spaan, A. (1992). A revision of the deer from Tegelen (Province of Limburg, The Netherlands). *Scripta*  
2078 *Geologica*, 98:1–85.
- 2079 Spencer, M. R. and Wilberg, E. W. (2013). Efficacy or convenience? Model-based approaches to  
2080 phylogeny estimation using morphological data. *Cladistics*, 29:663–671.
- 2081 Stadler, T. (2011). Mammalian phylogeny reveals recent diversification rate shifts. *Proceedings of the*  
2082 *National Academy of Sciences of the United States of America*, 108:6187–6192.
- 2083 Stamatakis, A. (2006). Raxml-vi-hpc: maximum likelihood-based phylogenetic analyses with thousands  
2084 of taxa and mixed models. *Bioinformatics*, 22(21):2688–2690.
- 2085 Stamatakis, A. (2014). Raxml version 8: a tool for phylogenetic analysis and post-analysis of large  
2086 phylogenies. *Bioinformatics*, 30(9):1312–1313.
- 2087 Stefaniak, K. and Stefaniak, K. (1995). Late pliocene cervids from weze 2 in southern poland. *Acta*  
2088 *Palaeontologica Polonica*, 40:327–340.
- 2089 Stehlin, H. G. (1923). Die oberpliocaeene fauna von senèze (haute-loire). *Eclogae Geologicae Helvetiae*,  
2090 18(2):268–281.
- 2091 Steininger, F. F. (1999). Chronostratigraphy, geochronology and biochronology of the miocene “european  
2092 land mammal mega-zones”(elmmz) and the miocene “mammal-zones (mn-zones)”. In Rössner, G. E.  
2093 and Heissig, K., editors, *The Miocene land mammals of Europe*, pages 9–24. Verlag Dr. Friedrich Pfeil,  
2094 München.
- 2095 Swofford, D. L. (2002). Paup\*. phylogenetic analysis using parsimony (\*and other methods). version 4.  
2096 Sinauer Associates, Sunderland, Massachusetts.
- 2097 Symeonidis, N. K. (1974). Ein vollständiges geweih von *Pliocervus pentelici* (gaudry) aus pikermi  
2098 (griechenland). *Annales Géologiques des Pays Helléniques*, 23:308–316.
- 2099 Taru, H. and Hasegawa, Y. (2002). The plio-pleistocene fossil mammals from the kasumi and tama hills.  
2100 *Memoirs of the National Science Museum, Tokyo*, 38:43–56.
- 2101 Tavaré, S. (1986). Some probabilistic and statistical problems in the analysis of dna sequences. *Lectures*  
2102 *on Mathematics in the Life Sciences. American Mathematical Society*, 17:57–86.
- 2103 Teilhard de Chardin, P. (1939). The miocene cervids from shantung. *Bulletin of the Geological Society of*  
2104 *China*, 19:269–278.
- 2105 Teilhard de Chardin, P. and Piveteau, J. (1930). *Les mammifères fossiles de Nihowan (Chine)*. Masson et  
2106 Cie, Paris.
- 2107 Timmins, R., Steinmetz, R., Sagar Baral, H., Samba Kumar, N., Duckworth, J., Anwarul Islam, M.,  
2108 Gimán, B., Hedges, S., Lynam, A., Fellowes, J., Chan, B., and Evans, T. (2008). *Rusa unicolor*. *The*  
2109 *IUCN red list of threatened species. Version 2014.3*. Available at [www.iucnredlist.org](http://www.iucnredlist.org) (accessed on 17  
2110 April 2015).
- 2111 Tomiati, C. and Abbazzi, L. (2002). Deer fauna from Pleistocene and Holocene localities of Ecuador  
2112 ( South America ) Cervidés du Pléistocène et Holocène d ’ Équateur ( Amérique du Sud ). *Geobios*,  
2113 35:631–645.
- 2114 Valli, A. (2010). Dispersion of the genus *Procapreolus* and the relationships between *Procapreolus*  
2115 *cusanus* and the roe deer (*Capreolus*). *Quaternary International*, 212(2):80–85.
- 2116 Valli, a. M. F. and Palombo, M. R. (2005). Le régime alimentaire du Cervidae (Mammalia) *Eucladoceros*  
2117 *ctenoides* (Nesti 1841) reconstitué par la morphologie du crâne et par l’usure dentaire. *Eclogae*  
2118 *Geologicae Helvetiae*, 98:133–143.
- 2119 Van der Geer, A., De Vos, J., Lyras, G., and Dermitzakis, M. (2006). New data on the Pleistocene Cretan  
2120 deer *Candiacervus* sp. II (Cervinae, Mammalia). *CFS Courier Forschungsinstitut Senckenberg*, pages  
2121 131–137.
- 2122 Van Mourik, S. and Stelmasiak, T. (1986). *Rusa* deer - from past to present. *Zoologischer Anzeiger*,  
2123 217:309–320.
- 2124 Viret, J. (1954). Le loess à bancs durcis de saint-vallier (drôme) et sa faune de mammifères villafranchiens.  
2125 *Nouvelles Archives du Muséum d’Histoire Naturelle de Lyon*, 4:1–200.
- 2126 Viret, J. (1961). Artiodactyla. In Piveteau, J., editor, *Traité de Paléontologie*, volume VI(1), pages  
2127 887–1084. Masson et Cie, Paris.
- 2128 Vislobokova, I. (2013). Morphology, taxonomy, and phylogeny of megacerines (megacerini, cervidae,  
2129 artiodactyla). *Paleontological Journal*, 47(8):833–950.



- 2130 Vislobokova, I. A. (1980). On systematical position of the deer from pavlodar and the origin of neocervines.  
2131 *Paleontological Journal*, 3:91–106.
- 2132 Vislobokova, I. A. (1983). The fossil deer of mongolia. *Transaction of the Joint Soviet-Mongolian*  
2133 *Paleontological Expedition*, 23:1–78.
- 2134 Vislobokova, I. A. (1990). The fossil deer of eurasia. *Transactions of the Palaeontological Institute*,  
2135 240:1–206.
- 2136 Vislobokova, I. a. (2009). The most ancient megacerine deer from the Late Miocene of Siberia and its  
2137 implications to the evolution of the group. *Palaeoworld*, 18(2008):278–281.
- 2138 Vislobokova, I. A. (2012). The history of giant deer (meracerini, cervidae, artiodactyla). *Transactions of*  
2139 *Paleontological Institute*, 293:1–102.
- 2140 Vislobokova, I. A., Changkang, H., and Sun, B. (1989). On the systematic position of the lagomerycinae.  
2141 *Vertebrata Palasiatica*, 27:130–132.
- 2142 Vrba, E. and Schaller, G. (2000). *Antelopes, Deer, and Relatives: Fossil Record, Behavioral Ecology,*  
2143 *Systematics, and Conservation*. Yale University Press, Yale.
- 2144 Wada, K., Nishibori, M., and Yokohama, M. (2007). The complete nucleotide sequence of mitochondrial  
2145 genome in the Japanese Sika deer (*Cervus nippon*), and a phylogenetic analysis between Cervidae and  
2146 Bovidae. *Small Ruminant Research*, 69:46–54.
- 2147 Wang, W. and Lan, H. (2000). Rapid and parallel chromosomal number reductions in muntjac deer  
2148 inferred from mitochondrial DNA phylogeny. *Molecular biology and evolution*, 17:1326–1333.
- 2149 Webb, S. (2000). Evolutionary history of new world cervidae. In Vrba, E. and Schaller, G., editors,  
2150 *Antelopes, deer, and relatives: fossil record, behavioral ecology, systematics, and conservation*, pages  
2151 38–64. Yale University Press, New Haven.
- 2152 Wilgenbusch, J., Warren, D., and D.L. S. (2004). *AWTY: A system for graphical exploration of MCMC*  
2153 *convergence in Bayesian phylogenetic inference*.
- 2154 Wilson, A. C., Carlson, S. S., and White, T. J. (1977). Biochemical evolution. *Annual review of*  
2155 *biochemistry*, 46(1):573–639.
- 2156 Wilson, D. E. and Reeder, D. M. (2005). *Mammal species of the world: a taxonomic and geographic*  
2157 *reference*, volume 1. JHU Press.
- 2158 Woodruff, D. S. (2003). Neogene marine transgressions, palaeogeography and biogeographic transitions  
2159 on the thai–malay peninsula. *Journal of Biogeography*, 30(4):551–567.
- 2160 Xie, W., Lewis, P. O., Fan, Y., Kuo, L., and Chen, M. H. (2011). Improving marginal likelihood estimation  
2161 for bayesian phylogenetic model selection. *Systematic Biology*, 60:150–160.
- 2162 Yang, Z. (1994). Maximum likelihood phylogenetic estimation from dna sequences with variable rates  
2163 over sites: approximate methods. *Journal of Molecular Evolution*, 39(3):306–314.
- 2164 Young, C. (1964). On a new *Lagomeryx* from lantian, shensi. *Vertebrata Asiatica*, 4:336–340.
- 2165 Zachos, F., Mattioli, S., Ferretti, F., and Lorenzini, R. (2014). The unique mesola red deer of italy:  
2166 taxonomic recognition (*Cervus elaphus italicus nova ssp.*, cervidae) would endorse conservation.  
2167 *Italian Journal of Zoology*, 81(1):136–143.
- 2168 Zachos, F. E. and Hartl, G. B. (2011). Phylogeography, population genetics and conservation of the  
2169 European red deer *Cervus elaphus*. *Mammal Review*, 41:138–150.
- 2170 Zaim, Y., De Vos, J., Huffman, O. F., Aziz, F., and Rizal, Y. (2003). A new antler specimen from the  
2171 1936 Perring hominid site, East Jawa, Indonesia, attributable to *Axis lydekkeri*. *Journal of Mineral*  
2172 *Technology*, X(1):1–9.
- 2173 Zdansky, O. (1925). Fossile hirsche chinas. *Palaeontologia Sinica, Series C*, 2(3):1–90.
- 2174 Zong, G.-F. (1987). Note on some mammalian fossils from the early pleistocene of di-qing county, yunnan.  
2175 *Vertebrata Palasiatica*, 25:69–76.