

Genomic differentiation in an endemic Philippine genus (Aves: *Sarcophanops*) due to geographic isolation on recently disassociated islands

Luke Campillo^{Corresp., Equal first author, 1, 2}, Joseph D. Manthey^{Equal first author, 1, 3}, Robert C. Thomson², Peter A. Hosner⁴, Robert G. Moyle¹

¹ Biodiversity Institute and Department of Ecology and Evolutionary Biology, University of Kansas, Lawrence, Kansas, United States

² Biology Department, University of Hawaii at Manoa, Honolulu, Hawaii, United States

³ Department of Biological Sciences, Texas Tech University, Lubbock, Texas, United States

⁴ Center for Macroecology, Evolution, and Climate, Natural History Museum of Denmark, Copenhagen, Denmark

Corresponding Author: Luke Campillo

Email address: campillo@hawaii.edu

Phylogeographic studies of Philippine vertebrates have demonstrated that genetic variation is broadly partitioned by Pleistocene island aggregation. Contemporary island discontinuity is expected to influence genetic differentiation, but remains relatively undocumented perhaps because the current episode of island isolation started relatively recently. We investigated inter- and intra-island population structure in a Philippine endemic bird genus (*Sarcophanops*) to determine if genetic differentiation has evolved during the recent period of isolation. We sequenced thousands of genome-wide RAD markers from throughout the Mindanao group to assess fine-scale genetic structure across islands. Specifically, we investigated patterns of gene flow and connectivity within and between taxonomic and geographic bounds. A previous assessment of mitochondrial DNA detected deep structure between *Sarcophanops samarensis* and sister species, *S. steerii*, but was insufficient to detect differentiation within either species. Analysis of RAD markers, however, revealed structure within *S. samarensis* between the islands of Samar/Leyte and Bohol. This genetic differentiation likely demonstrates an effect of recent geographic isolation (post-LGM) on the genetic structure of Philippine avifauna. We suggest that the general lack of evidence for differentiation between recently isolated islands is a failure to detect subtle population structure due to past genetic sampling constraints, rather than the absence of such structure.

1 Genomic differentiation in an endemic Philippine genus (Aves: *Sarcophanops*) due to
2 geographic isolation on recently disassociated islands

3
4 Luke C. Campillo*^{1,2}, Joseph D. Manthey*^{1,3}, Robert C. Thomson², Peter A. Hosner⁴, and Robert G.
5 Moyle¹

6
7 ¹Biodiversity Institute and Department of Ecology and Evolutionary Biology, University of Kansas,
8 Lawrence, KS, USA

9 ²Biology Department, University of Hawai'i – Mānoa, Honolulu, HI, USA

10 ³Department of Biological Sciences, Texas Tech University, Lubbock, TX, USA

11 ⁴Center for Macroecology, Evolution, and Climate; Natural History Museum of Denmark, Copenhagen,
12 Denmark

13

14 Corresponding Author:

15 Luke C. Campillo

16 2538 McCarthy Mall

17 EDM 216

18 Honolulu, HI 96822

19 campillo@hawaii.edu

20

21 * These authors contributed equally

22 Abstract

23 Phylogeographic studies of Philippine vertebrates have demonstrated that genetic variation is broadly
24 partitioned by Pleistocene island aggregation. Contemporary island discontinuity is expected to influence
25 genetic differentiation, but remains relatively undocumented perhaps because the current episode of
26 island isolation started relatively recently. We investigated inter- and intra-island population structure in a
27 Philippine endemic bird genus (*Sarcophanops*) to determine if genetic differentiation has evolved during
28 the recent period of isolation. We sequenced thousands of genome-wide RAD markers from throughout
29 the Mindanao group to assess fine-scale genetic structure across islands. Specifically, we investigated
30 patterns of gene flow and connectivity within and between taxonomic and geographic bounds. A previous
31 assessment of mitochondrial DNA detected deep structure between *Sarcophanops samarensis* and sister
32 species, *S. steerii*, but was insufficient to detect differentiation within either species. Analysis of RAD
33 markers, however, revealed structure within *S. samarensis* between the islands of Samar/Leyte and Bohol.
34 This genetic differentiation likely demonstrates an effect of recent geographic isolation (post-LGM) on
35 the genetic structure of Philippine avifauna. We suggest that the general lack of evidence for
36 differentiation between recently isolated islands is a failure to detect subtle population structure due to
37 past genetic sampling constraints, rather than the absence of such structure.

39 Introduction

40 The Philippine Archipelago is recognized as one of the most biologically diverse hotspots in the world
41 (Myers et al., 2000) largely due to a complex geologic and climatic history that has catalyzed the
42 evolution of endemic biodiversity (Brown et al., 2013). Due to cyclic sea level change, the extent of land
43 above water in the Philippine Archipelago has varied dramatically throughout its geologic history.
44 Specifically, changing climate regimes during the Last Glacial Maximum (LGM; 19-25 kyr bp) resulted
45 in lower global sea levels, consequently uncovering shallow land bridges between islands. This network
46 of shallow land bridges dramatically increased connectivity across the archipelago (Heaney, 1985),
47 forming clustered groups of interconnected islands, or Pleistocene Aggregate Island Complexes (PAICs;
48 Diesmos et al., 2002; Brown et al., 2013). Of the more than 7,000 islands found in the present-day
49 Philippine Archipelago (Kennedy et al., 2000), nearly all were reduced to six large PAICs (Luzon,
50 Palawan, Mindoro, Negros-Panay, Mindanao, Sulu; Heaney, 1985)

51 Endemic Philippine avifauna generally adheres to the patterns of geographic and phylogenetic
52 structure predicted under the PAIC model, at least when additional complexities such as topography,
53 paleoclimatic factors, and colonization history are acknowledged (Hosner, Nyári & Moyle, 2013; Hosner
54 et al., 2014; Sánchez-González, Hosner & Moyle, 2015). That is, populations present on a particular
55 PAIC (e.g. Mindanao PAIC), are likely to be closely related to one another, but genetically distinct from
56 populations confined to different PAICs during the LGM (e.g. Luzon PAIC; Sánchez-González & Moyle,
57 2011). Although broad strokes at understanding Plio-Pleistocene diversification across the archipelago
58 have been possible for some time, the power to detect more recent, fine-scale differentiation has been
59 limited by DNA sequencing depth. Furthermore, much of the work on Philippine biodiversity has focused
60 on the patterns and processes shaping diversity throughout the archipelago, despite the fact that not all
61 lineages have distributions spanning its entirety. Hence, the generation of recent population genetic
62 structure owing to Holocene isolation on individual islands within the same PAIC remains theoretical.
63 Here, we investigate the effect of individual islands in generating genetic differentiation in the endemic
64 Philippine broadbills (Aves; Family: Eurylaimidae; Genus: *Sarcophanops*), in which all extant lineages
65 occur on one previously connected landmass (the Greater Mindanao PAIC, which now comprises many

66 islands). The two species of Philippine broadbill, *Sarcophanops steerii* and *S. samarensis*, occur in non-
67 overlapping ranges on Mindanao and the Eastern Visayas (incl. Samar, Leyte, and Bohol Islands),
68 respectively. We used restriction-site associated DNA sequencing (RAD-seq) to produce a genome-wide
69 panel of thousands of single nucleotide polymorphisms (SNPs), which allows for assessment of subtle
70 population genomic structure across islands that were part of the same PAIC as recently as the LGM.
71 Inferring differentiation at this evolutionary timescale has not, to our knowledge, been documented in
72 Philippine avifauna. Focusing on a genus (*Sarcophanops*) endemic to a single PAIC enables us to: (1):
73 examine inter- or intra-island population structure within *Sarcophanops* species to get a glimpse into
74 genetic connectivity of avifauna endemic to the Mindanao PAIC, and (2) expand our understanding of the
75 population history of these enigmatic taxa.

76

77 **Materials & Methods**

78 We obtained tissue samples ($N = 22$) of *Sarcophanops* from across their distribution in the Philippines,
79 and used two individuals from *Serilophus lunatus* as outgroup (Table 1; Fig. 1). All tissue samples are
80 frozen and/or ethanol-preserved muscle tissue and have associated voucher specimens housed in the
81 Biodiversity Institute at the University of Kansas. We used a modified RAD-seq (Miller et al., 2007)
82 protocol to prepare genomic libraries of putatively-neutral loci from across the genome. Briefly, we
83 ligated custom adapters with barcodes (Andolfatto et al., 2011) to all samples, which were pooled and
84 subsequently purified with AMPure magnetic beads (Agencourt). We used a Pippin Prep (Sage Science)
85 to size select fragments between 500 and 600 bp. We then purified the library again with magnetic beads,
86 performed a brief polymerase chain reaction (PCR) in duplicate (14 cycles), and performed a final
87 purification before dual-indexing the samples (with standard Illumina indices) for multiplexing. The
88 multiplexed library was pooled with libraries from unrelated projects and sequenced across three lanes of
89 an Illumina HiSeq2500 flow cell.

90 To assemble loci de novo and create SNP datasets from our sequencing data, we used the
91 STACKS (Catchen et al., 2013) pipeline (more details available in Supp Mat). We used *ustacks* with the
92 default settings. In *cstacks*, we tested various numbers of mismatches allowed between stacks when
93 assembling loci ($N = 1-7$; Table S2). We then used the *sstacks* module with default settings. Finally, we
94 used the *populations* module of STACKS to filter SNP loci and create two datasets: (1) a 50% coverage
95 matrix (50CM; requiring a SNP to be represented in $\geq 50\%$ of individuals), and (2) a 75% coverage
96 matrix (75CM). We required all loci to have a minimum read depth of five and maximum observed
97 heterozygosity $< 50\%$ to reduce inclusion of paralogs. We also assessed how changing the minimum read
98 depth could affect population genetic estimates and insured good coverage across the genome by using
99 the BLAST+ utility (Camacho et al., 2009), requiring a minimum of 70% sequence identity across at least
100 25 bp, and a maximum e-value of 0.001 to define a match.

101 We used RAxML v8 (Stamatakis, 2014) to identify phylogenetic relationships among individuals
102 using a concatenated matrix of all full-length sequences. We first estimate an appropriate model of
103 sequence evolution (GTR + I + G in this case) based on the Bayesian Information Criterion (BIC) using
104 PAUP v.4.0.151 (Swofford, 2002). In RAxML, we estimated a maximum likelihood tree and assessed
105 support using 1000 rapid bootstrap replicates. We used the programs STRUCTURE (Pritchard et al.,
106 2000) and DAPC (Jombart, Devillard & Balloux, 2010) to investigate population genetic structure for the
107 75CM dataset. For both analyses, we subset our datasets to include only one SNP per locus (two
108 replicates each) to minimize potential linkage effects. We ran STRUCTURE initially to infer lambda with
109 the number of populations (k) limited to one. Next, we used a constant lambda, the admixture model with

110 correlated allele frequencies, and a number of likely k values ($k = 1-5$, five runs for each k). We defined
111 the burn-in period as the first 100,000 MCMC generations with a subsequent 100,000 iterations sampled.
112 To determine the most likely number of genetic clusters, we used the ΔK method of Evanno *et al.* (2005).
113 We also performed the same analysis on *S. samarensis* alone to look for population structure within the
114 Visayan islands. DAPC analyses were performed in R (R Core Team, 2013), using the package
115 ‘adegenet’ (Jombart, 2008; Jombart & Ahmed, 2011). For DAPC, the most likely number of populations
116 was determined based on BIC values.

117

118

119 Results

120 Sequencing coverage across individuals was variable (Table 1), with a median ~ 1.6 million reads per
121 individual ($sd = 822,669$ reads). From these reads we recovered $\sim 25,000$ RAD-tags per individual ($sd =$
122 8533). The 50% and 75% coverage matrices had 1,737 and 885 loci, respectively, corresponding to 4,310
123 and 2,271 SNPs (Table S1). All raw sequence data from RAD-seq are available at the NCBI Sequence
124 Read Archive, accessioned under BioProject XXX (ID #s pending). Genetic differentiation, measured by
125 F_{ST} , between sampling localities within a given species was generally low ($F_{ST} < 0.15$), but high between
126 species ($F_{ST} > 0.30$; Table 2). Genetic structure was most apparent across species (i.e. between *S.*
127 *samarensis* and *S. steeri*) in phylogenetic (Fig. 1) and population genetic (Fig. 2) analyses. Phylogenetic
128 analysis in RAxML supported a deep split between species, but relationships within *S. samarensis* were
129 largely ambiguous with respect to island. However, we did find Bohol was recovered as monophyletic in
130 the 50CM tree. Population genetic analyses recovered a similar pattern overall pattern, but some
131 differences are observed, likely due to RAxML analyses being based on a concatenated dataset. In
132 STRUCTURE, the ΔK method most strongly supported two genetic clusters, separating Visayan and
133 Mindanao individuals (Fig. 2). We also ran STRUCTURE on only the Visayan individuals and recovered
134 a strong break between Bohol and Samar + Leyte, but did not recover any further genetic partitioning (i.e.
135 no split between Samar and Leyte). When running DAPC, we observed three distinct clusters
136 corresponding to individuals from Mindanao, Bohol, and Samar + Leyte (Fig. 2).

137

138 Discussion

139 When comparing diversification in *Sarcophanops* to other endemic fauna from the Mindanao PAIC, we
140 observe that many taxa show a similar pattern of differentiation. For example, in *Cyrtodactylus* geckos
141 (Welton *et al.*, 2010) and *Crocidura* shrews (Esselstyn, Timm & Brown, 2009) the Visayan and
142 Mindanao populations form independent genetic clusters, which was consistent with our phylogenomic
143 and population genetic analyses which recover a deep split between the Mindanao (*S. steerii*) and Visayan
144 (*S. samarensis*) species. Recently published findings based on Bayesian species delimitation of
145 mitochondrial DNA sequence data also revealed the same deep split between Mindanao and Visayan
146 species (Hosner *et al.*, 2018), but failed to identify a signature of divergence within *S. samarensis* as we
147 found here. The well-supported phylogenetic split between the Mindanao and Visayan species in both the
148 mtDNA and nuclear DNA suggest they remained isolated during the LGM, despite the fact all these
149 islands formed a single contiguous island, the Mindanao PAIC. Possibly, this isolation relates to the role
150 of environmental suitability. Based on paleoclimate projections, (Hosner *et al.*, 2014) found that the
151 shallow Leyte Gulf—the land bridge uniting the northern and southern islands of the Mindanao PAIC—
152 was unsuitable for most species in their study and still acted as a barrier to gene flow despite increased

153 land connectivity. Although we did not perform niche modeling in this study, the Leyte Gulf could have
154 also been unsuitable habitat for *Sarcophanops*, thus facilitating the divergence of Mindanao and Visayan
155 populations.

156 In our study, RAD-seq data revealed fine-scale inter-island diversification within the Visayan
157 broadbills, which was not evident in mtDNA alone (Hosner et al., 2018). This suggests the shallow split
158 between Bohol and Samar + Leyte is rather recent, most likely post-LGM. SNP-based genetic structure
159 (Fig. 2) revealed a high probability of two distinct populations within *S. samarensis*: Samar + Leyte, and
160 Bohol. This geographic partitioning is particularly interesting given the current taxonomic treatment of *S.*
161 *samarensis*, which contains no subspecific taxa from the Visayas. In Mindanao, there are two described
162 subspecies, but we recovered only one *S. steerii* population in the RAD-seq dataset, with slight evidence
163 to support separation of the Zamboanga population, as seen in the mtDNA dataset.

164 Although all present-day islands in the eastern Visayas were connected at one point during the LGM, the
165 narrow (0.8-1.6 km) and shallow (max. 20 m) San Juanico Strait separating Samar and Leyte probably
166 extended terrestrial connectivity between these two islands longer relative to other neighboring islands in
167 the Mindanao PAIC. Rising sea levels at the end of the Pleistocene would have isolated Bohol first, while
168 prolonged connectivity between Samar and Leyte could have promoted gene flow, thus obscuring
169 population genetic effects of inter-island diversification. Because little is known about the current
170 population status of these birds, and because little appropriate forested habitat remains on Bohol in
171 particular, understanding the genetic connectivity across the Visayan islands is an important contribution
172 to properly addressing the conservation needs of this enigmatic genus.

173

174 Conclusions

175 Numerous studies have investigated the effect of PAICs in generating endemism in the Philippines
176 (Brown et al., 2013). Yet, the nature of those studies has provided limited understanding of recent,
177 between-island differentiation. Focusing on an endemic lineage restricted to a single and well-established
178 island group, we were able to recover both deep and subtle genetic differentiation between islands.
179 Because this differentiation was not well-supported in the “fast evolving” mtDNA, we suggest the two,
180 previously undocumented Visayan lineages arose after the LGM and are therefore only detectable in a
181 deep, genome-wide scan of thousands of loci using a method such as RAD-seq. This study represents a
182 solid step forward in understanding genetic differentiation consistent with a post-LGM timeframe in a
183 single PAIC. Furthermore, our results suggest that subtle differentiation within islands groups,
184 particularly since the LGM, has been overlooked due to past genetic sampling constraints.

185

186 Acknowledgements

187 We would like to thank Mark Robbins from the University of Kansas Natural History Museum. We thank
188 the KU Genome Sequencing Core (supported by NIH grant 5P20GM103638 to E.A. Lundquist) and the
189 KU Advanced Computing Facility (partially funded by NSF grant CNS 1337899 to A. T. Peterson). The
190 National Science Foundation (DEB-0743491; DEB-1418895), American Ornithologists' Society,
191 American Museum of Natural History Chapman Fund, and the University of Kansas Panorama Fund
192 supported fieldwork; The National Science Foundation (DEB-1110619; DEB-1557053) and the
193 University of Kansas Graduate Student Research Fund supported lab work.

194

195 References

- 196 Andolfatto P, Davison D, Erezylmaz D, Hu TT, Mast J, Sunayama-Morita T, Stern DL. 2011.
197 Multiplexed shotgun genotyping for rapid and efficient genetic mapping. *Genome Research*
198 21:610–617. DOI: 10.1101/gr.115402.110.
- 199 Brown RM, Siler CD, Oliveros CH, Esselstyn JA, Diesmos AC, Hosner PA, Linkem CW, Barley AJ,
200 Oaks JR, Sanguila MB, Welton LJ, Blackburn DC, Moyle RG, Townsend Peterson A, Alcalá AC.
201 2013. Evolutionary Processes of Diversification in a Model Island Archipelago. *Annual Review of*
202 *Ecology, Evolution, and Systematics* 44:411–435. DOI: 10.1146/annurev-ecolsys-110411-
203 160323.
- 204 Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+:
205 architecture and applications. *BMC Bioinformatics* 10:421. DOI: 10.1186/1471-2105-10-421.
- 206 Catchen J, Hohenlohe PA, Bassham S, Amores A, Cresko WA. 2013. Stacks: an analysis tool set for
207 population genomics. *Molecular Ecology* 22:3124–3140. DOI: 10.1111/mec.12354.
- 208 Diesmos AC, Brown RM, Alcalá AC, Sison RV, Afuang LE, Gee GVA. 2002. Philippine amphibians and
209 reptiles. Philippine biodiversity conservation priorities: a second iteration of the National
210 Biodiversity Strategy and Action Plan.
- 211 Esselstyn JA, Timm RM, Brown RM. 2009. Do geological or climatic processes drive speciation in
212 dynamic archipelagos? The tempo and mode of diversification in southeast asian shrews.
213 *Evolution* 63:2595–2610. DOI: 10.1111/j.1558-5646.2009.00743.x.
- 214 Heaney L. 1985. *Zoogeographic evidence for Middle and Late Pleistocene landbridges to the Philippine*
215 *Islands*.
- 216 Hosner PA, Campillo LC, Andersen MJ, Sánchez-González LA, Oliveros CH, Urriza RC, Moyle RG.
217 2018. An integrative species delimitation approach reveals fine-scale endemism and substantial
218 unrecognized avian diversity in the Philippine Archipelago. *Conservation Genetics*. DOI:
219 10.1007/s10592-018-1085-4.
- 220 Hosner PA, Nyári ÁS, Moyle RG. 2013. Water barriers and intra-island isolation contribute to
221 diversification in the insular *Aethopyga* sunbirds (Aves: Nectariniidae). *Journal of Biogeography*
222 40:1094–1106. DOI: 10.1111/jbi.12074.
- 223 Hosner PA, Sánchez-González LA, Peterson AT, Moyle RG. 2014. Climate-driven diversification and
224 pleistocene refugia in philippine birds: evidence from phylogeographic structure and
225 paleoenvironmental niche modeling: philippine avian phylogeography. *Evolution* 68:2658–2674.
226 DOI: 10.1111/evo.12459.
- 227 Jombart T, Devillard S, Balloux F. 2010. Discriminant analysis of principal components: a new method
228 for the analysis of genetically structured populations. *BMC Genetics* 11:94. DOI: 10.1186/1471-
229 2156-11-94.
- 230 Kennedy RS, Gonzales PC, Dickinson EC, Miranda HC, Fisher TH. 2000. *A guide to the birds of the*
231 *Philippines*. Oxford ; New York: Oxford University Press.
- 232 Miller MR, Dunham JP, Amores A, Cresko WA, Johnson EA. 2007. Rapid and cost-effective
233 polymorphism identification and genotyping using restriction site associated DNA (RAD)
234 markers. *Genome Research* 17:240–248. DOI: 10.1101/gr.5681207.
- 235 Myers N, Mittermeier RA, Mittermeier CG, da Fonseca GA, Kent J. 2000. Biodiversity hotspots for
236 conservation priorities. *Nature* 403:853–858. DOI: 10.1038/35002501.
- 237 R Core Team. 2013. *R: A Language and Environment for Statistical Computing*. Vienna, Austria: R
238 Foundation for Statistical Computing.
- 239 Sánchez-González LA, Hosner PA, Moyle RG. 2015. Genetic Differentiation in Insular Lowland
240 Rainforests: Insights from Historical Demographic Patterns in Philippine Birds. *PLOS ONE*
241 10:e0134284. DOI: 10.1371/journal.pone.0134284.
- 242 Sánchez-González LA, Moyle RG. 2011. Molecular systematics and species limits in the Philippine
243 fantails (Aves: Rhipidura). *Molecular Phylogenetics and Evolution* 61:290–299. DOI:
244 10.1016/j.ympev.2011.06.013.
- 245 Stamatakis A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large
246 phylogenies. *Bioinformatics*:btu033.

247 Welton LJ, Siler CD, Linkem CW, Diesmos AC, Brown RM. 2010. Philippine Bent-Toed Geckos of the
248 *Cyrtodactylus agusanensis* Complex: Multilocus Phylogeny, Morphological Diversity, and
249 Descriptions of Three New Species. *Herpetological Monographs* 24:55–85. DOI:
250 10.1655/HERPMONOGRAPHS-D-10-00005.1.
251

Figure 1(on next page)

RAxML trees for the 50CM (left) and 75CM (right) datasets.

Bootstrap support was assessed based on 1000 rapid bootstrap replicates; nodes with less than 50% bootstrap support have no node label. Visayan samples are shown in shades of blue, and Mindanao samples are shown in shades of yellow.

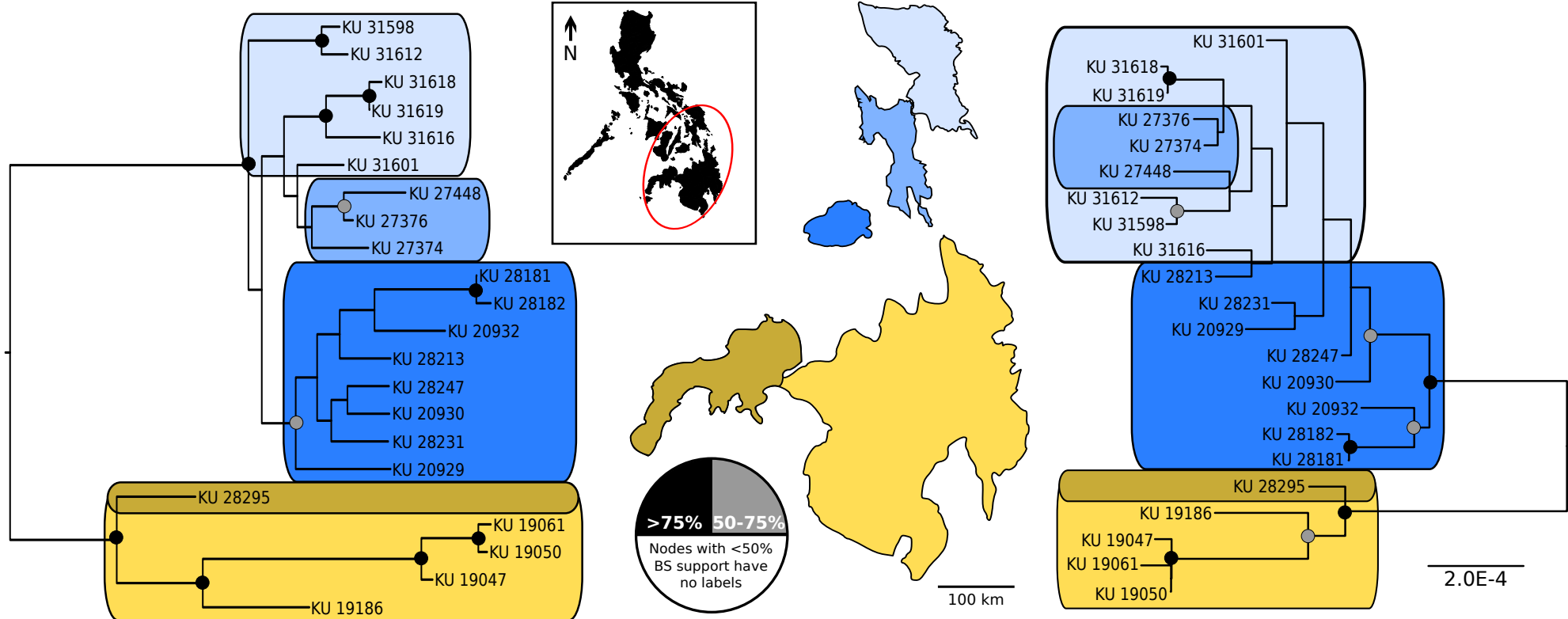


Figure 2(on next page)

DAPC and STRUCTURE results for 75CM.

Again, Mindanao samples are shown in shades of yellow, and Visayan samples are shown in shades of blue. The STRUCTURE plot on the bottom (blue and yellow), shows the break between *S. steerii* and *S. samarensis*. The STRUCTURE plot on the right only includes individuals from *S. samarensis* and shows the clear break between Bohol (top, dark blue) and Samar + Leyte (bottom, light blue). For both STRUCTURE plots, each bar represents the probability of population assignment for a single individual.

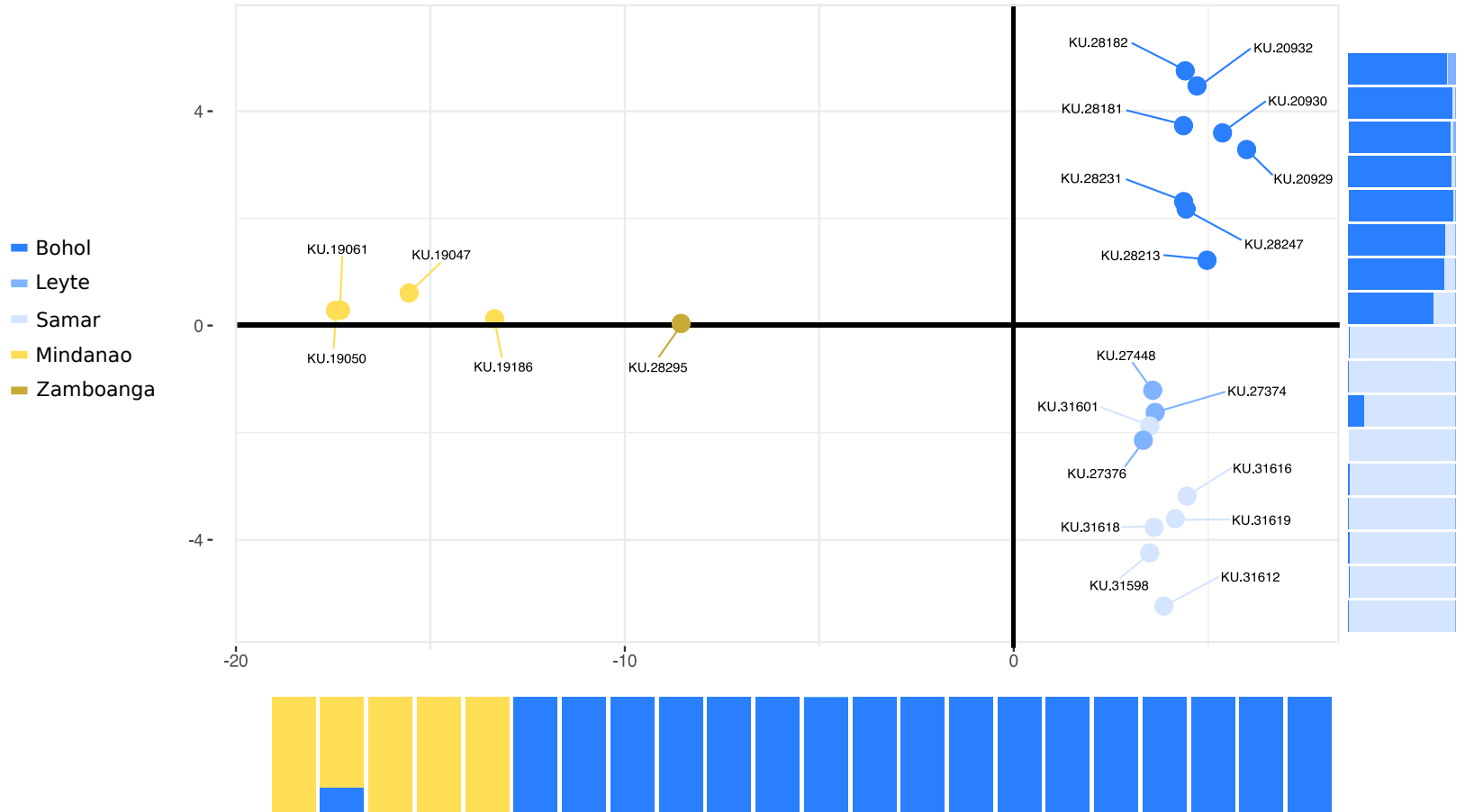


Table 1 (on next page)

List of samples used in this study and their associated sequencing statistics.

Table 1 – List of samples used in this study and their associated sequencing statistics.

Species	Museum #	Locality	# Reads	RAD-tags	Cov. Median	Cov. St.Dev.	% Missing 50% CM / 70% CM
<i>Sarcophanops steerii</i>	KU 19047	Mindanao	874532	20185	30	33.29	15.31 / 3.75
<i>Sarcophanops steerii</i>	KU 19050	Mindanao	991797	20902	35	37.70	13.46 / 3.52
<i>Sarcophanops steerii</i>	KU 19061	Mindanao	2408639	31817	72	83.53	3.46 / 0.48
<i>Sarcophanops steerii</i>	KU 28295	Mindanao	2600919	39135	74	80.55	14.20 / 13.43
<i>Sarcophanops steerii</i>	KU 19186	Zamboanga	1782408	26521	53	67.01	0.00 / 0.00
<i>Sarcophanops samarensis</i>	KU 20929	Bohol	2180969	32211	65	72.52	5.99 / 1.45
<i>Sarcophanops samarensis</i>	KU 20930	Bohol	1638944	23861	53	62.67	6.38 / 0.70
<i>Sarcophanops samarensis</i>	KU 20932	Bohol	1371645	26271	43.5	49.98	8.98 / 1.14
<i>Sarcophanops samarensis</i>	KU 28181	Bohol	712002	17718	26	29.03	15.85 / 5.02
<i>Sarcophanops samarensis</i>	KU 28182	Bohol	2285682	28961	65	78.28	3.25 / 0.84
<i>Sarcophanops samarensis</i>	KU 28213	Bohol	1197631	31888	37	38.54	13.53 / 3.57
<i>Sarcophanops samarensis</i>	KU 28231	Bohol	1981016	32462	60	68.49	4.69 / 0.97
<i>Sarcophanops samarensis</i>	KU 28247	Bohol	1465002	31170	44	51.05	6.26 / 0.48
<i>Sarcophanops samarensis</i>	KU 27374	Leyte	1722020	23775	56	65.06	3.62 / 0.00
<i>Sarcophanops samarensis</i>	KU 27376	Leyte	2833494	30055	88	98.85	3.36 / 0.00
<i>Sarcophanops samarensis</i>	KU 27448	Leyte	1525375	25388	46	58.30	3.81 / 0.00
<i>Sarcophanops samarensis</i>	KU 31598	Samar	450539	22184	15	14.55	23.32 / 2.03
<i>Sarcophanops samarensis</i>	KU 31601	Samar	808201	25697	23	28.23	14.32 / 0.31
<i>Sarcophanops samarensis</i>	KU 31612	Samar	1793799	39214	54	55.63	5.87 / 1.23
<i>Sarcophanops samarensis</i>	KU 31616	Samar	3347326	44974	87	100.42	6.17 / 1.50
<i>Sarcophanops samarensis</i>	KU 31618	Samar	889378	24240	26	31.44	9.35 / 0.35
<i>Sarcophanops samarensis</i>	KU 31619	Samar	986355	25079	32	35.91	7.87 / 0.13
<i>Serilophus lunatus</i>	KU 23405	Vietnam	3515364	55899	63	91.42	4.52 / 0.00
<i>Serilophus lunatus</i>	KU 23552	Vietnam	1302599	27169	35	43.52	12.44 / 0.00

Table 2 (on next page)

Pairwise estimates of F_{ST} for the 75% and 50% coverage matrices above and below the diagonal, respectively.

Table 2 – Pairwise estimates of F_{ST} for the 75% and 50% coverage matrices above and below the diagonal, respectively.

	Bohol	Leyte	Samar	Mindanao	Zamboanga
Bohol		0.114	0.122	0.323	0.321
Leyte	0.116		0.106	0.369	0.452
Samar	0.114	0.116		0.340	0.355
Mindanao	0.310	0.346	0.333		0.204
Zamboanga	0.309	0.447	0.357	0.214	