

An analysis of the mechanism of aging: endogenous viral stimulus and the deleterious effect of chronic inflammation

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Background

A trivial logic suggests that aging is a product of damage induced by natural processes in the organism like reactive oxygen species (Harman, 1956) or random somatic DNA mutations leading to cell and tissue dysfunction (Failla, 1958; Szilard, 1959). These both opinions consider living organisms as if these are human-made mechanisms created once and then constantly being worn out during exploitation. Multi-cellular living organisms are effectively an unstoppable process of replacement, renewal and regeneration. Indeed, a causative relationship between damage and aging is not proved experimentally yet. The large size of the eukaryotic genome makes it difficult to assess the effect of DNA mutations on aging making yeasts as a suitable experimental model due to the well-characterized small yeast genome with significant conservation of proteins of the DNA repair pathway between yeast and higher eukaryotes (Bitterman et al., 2003). However, experiments on yeasts do not prove that DNA mutations cause aging (Kaya et al., 2015). Indeed, living organisms are open systems adjusting to keep homeostasis and aging cannot be explained as a process of “wear and tear” as it goes in inanimate closed systems suffering under the second law of thermodynamics. The internal system of repair becomes ineffective during the course of an individual life presumably due to an evolutionary purpose (Mitteldorf, 2010). Without a doubt, the cause of aging is located in the genome because aging rate and lifespan limit is highly specific for a particular species and may be fluctuated due to external environmental challenges or individual genome variability.

Transposons and evolution of aging

Limited lifespan may provide flexibility in the evolution of species but this statement was opposed by the current mainstream evolutionary theory that prefers individual selection as more significant for evolution. This statement based on an argument that natural selection is conducted only on protein-coding genes. In this case, it is not clear why evolution lets evolve genes that decrease the fitness of a single living being for benefits of the whole species. For instance, the protein-coding genes comprise merely 1.5% of the human genome. However, about half of the human genome is composed of transposable elements (transposons). Barbara McClintock believed transposons help to promote evolution and adaptations of the organism (McClintock, 1984). Contrary, others consider transposons are parasitic genetic elements using bodies as their carriers for limitless expansion (Doolittle and Sapienza, 1980; Orgel and Crick, 1980). Nowadays, we can see a multifaceted impact of transposons on the living organism and these may be both our friend and foes. Transposons increase a range of phenotypic variability facilitating the adaptation of genomes to their environment (Evsikov and Marín

de Evsikova, 2016). Also, transposons could induce the transition from unicellular to multi-cellular life in the past (Koonin, 2016) and cause reproductive isolation and speciation (Serrato-Capuchina and Matute, 2018). It reveals transposons as an important and influential driving force of evolution leading to discussions that transposons may increase the significance of higher levels of selection (Brunet and Doolittle, 2015). If it is true, transposon-driven evolution promotes fitness a whole species over an individual organism. It paradoxically merges two contrary opinions of beneficial and detrimental effects of transposons in the organism. We might consider transposons as engines of evolution that made us as we are but also made us aging.

Retrotransposons cause genome instability

Transposons are divided into two broad classes: DNA transposons and retrotransposons different in the mechanism of action. DNA transposons act as a “cut-and-paste” tool without making additional copies of these genetic elements. Retrotransposons act as a “copy-and-paste” tool using reverse transcriptase for producing additional copies of this type of transposons. This mechanism made retrotransposons the largest class in the human genome. For instance, the activity of retrotransposons increase with age in nearly every aging model and considered to promote genome instability during aging (Moskalev et al., 2013). This phenomenon is accompanied by a decrease in repressive heterochromatin with aging that suppresses the expression and mobility of retrotransposons (Moskalev et al., 2013; Wood and Helfand, 2013). High activity of retrotransposons and decrease of heterochromatin is also observed in cancer cells (De Cecco et al., 2013). Genome instability is driven mostly by L1 (LINE-1) retrotransposons by generating double-strand breaks (Belancio et al., 2010; Gasior et al., 2006). LINE transposons also drives SINE activation (Dewannieux et al., 2003). Transposons are a vital element in adaptation to various challenges of environment (Horváth et al., 2017) and any stress causes transposons activation, therefore increased mutation rate during aging may be a side effect of the stress response.

Lamina-associated domains are involved in normal and premature aging

Interestingly, the repressive heterochromatin is associated with peripheral areas of the nucleus and overlapped with lamina-associated domains (LADs). LADs are featured by gene scarcity and the repetitive genome. The borders of LADs are marked with the insulator protein CTCF, CpG islands, gene promoters and high expression of genes (Guelen et al., 2008). Also, the movement of these LADs away from the nuclear periphery is associated with the increase of gene expression (Peric-Hupkes et al., 2010). Nuclear lamina serves an anchor for LADs connected with repressive heterochromatin (Gibcus and Dekker, 2013). Thus, the detachment of LADs due to degradation of lamin proteins leads to massive derepression of retrotransposons. This mechanism explains how defects in lamin proteins cause premature aging due to mutation of the lamin-coding LMNA gene in Hutchinson-Gilford progeria syndrome. The same process occurs during of normal aging. For example, cell nuclei from old individuals acquire defects similar to those of Hutchinson-Gilford patient cells, including changes in histone modifications and increased DNA damage (Scaffidi and Misteli, 2006). Additionally, interactions with the nuclear lamina exhibit oscillating patterns regulating expression of circadian genes (Zhao et al., 2015).

Age-associated patterns in DNA and histone methylation

Loss of heterochromatin is also accompanied by the global loss of DNA methylation. Age-associated demethylation is correlated with a certain type of retrotransposons: Alu and HERV-K (Jintaridth and Mutirangura, 2010). Replicative senescence in human cells exhibits patterns of global DNA hypomethylation and local hypermethylation. Hypomethylation occurs preferentially in LADs. Hypermethylation arises in CpG islands including tumorigenic loci, associated with key developmental genes. Changes of hypermethylation in replicative senescence are similar to those in cancer (Cruickshanks et al., 2013). In contrast, patterns of hypomethylation are different for aging and cancer. Hypomethylation in H3K4me1 is observed preferentially in aging, while the loss of DNA methylation in cancer was mostly associated with H3K9me3 marks (Pérez et al., 2018). H3K4me1 is a mark of the chromatin remodeling complex BAF linked to the activity of p53 (Local et al., 2018). Therefore, normal p53 activity determines its antitumorigenic effect. Hypermethylated promoters are associated with Polycomb repressive complexes (Widschwendter et al., 2007) targeting bivalent chromatin (Rakyan et al., 2010). Inflammation induces expression of Jmjd3 demethylase due to the presence of inflammatory cytokines. Jmjd3 targets H3K27me3 histone and represses PcG target genes associated with the polycomb repressive complex (De Santa et al., 2007). Interestingly, the epigenetic clocks based on DNA methylation demonstrate paradoxically high correlation with chronological age (above 0.95) (Hannum et al., 2013; Horvath, 2013). Therefore, epigenetics is directly linked to aging processes. Epigenetic aging is also related to increased likelihood of cancer incidence through stabilizing stem cells features (Teschendorff et al., 2010). Moreover, epigenetic aging correlates with overall survival in several types of cancer (Lin and Wagner, 2015). Increased epigenetic age was associated with the increase of fatality and recurrence of cancer (Ren et al., 2018).

Oxidative stress and hypomethylation

Hypermethylation triggered by changes in signaling pathways, while hypomethylation looks like a result of stochastic processes (Marttila et al., 2015). The stochastic nature of global hypomethylation is likely a consequence of oxidative stress. Although there is no causal relationship between aging and oxidative stress triggered by reactive oxygen species (ROS) (Doonan et al., 2008; Stuart et al., 2014), ROS may have a role in cellular signaling and cause DNA hypomethylation (Afanas'ev, 2013). This mechanism can be explained through oxidation of 5-methylcytosine that leads to loss of markers of DNA methylation (Valinluck and Sowers, 2007). The increase of ROS during aging is presumably induced with chronic inflammation and innate immune response. Increased ROS production often is associated with mild hypoxia (Pialoux et al., 2009). Hypoxia is induced by the STAT3 signaling pathway (Niu et al., 2008) that is vital in immune system response including also the mTOR signaling (Saleiro and Platanias, 2015).

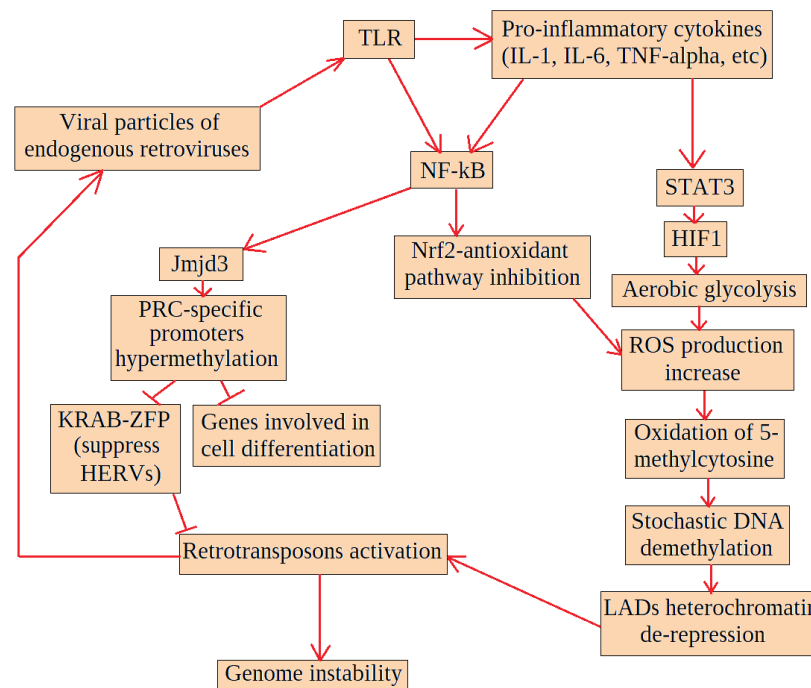


Figure 1. Interactions of signaling pathways mediated by antiviral response to endogenous retroviruses expression leads to genome instability, increased ROS production and epigenetic modification

Chronic inflammation, ROS, and epigenetics

The fact that increased ROS production is induced by inflammation and cause global hypomethylation with age is consistent with facts when accelerated epigenetic age is associated with activation pro-inflammatory and interferon pathways (Irvin et al., 2018; Levine et al., 2018). Moreover, patterns of methylation associated with mortality during aging reveal a genetic regulatory network focused on NF- κ B (Jylhävä et al., 2016). Indeed, inhibition of NF- κ B reduces oxidative stress and delayed cellular senescence leads to a suggestion that inflammation is considered as a culprit of senescence (Osorio et al., 2012; Salminen et al., 2008; Tilstra et al., 2012). NF- κ B inhibits gonadotropin-releasing hormone (GnRH) in the hypothalamus (Zhang et al., 2013) reducing the production of sex hormones and triggering reproductive fading. Moreover, NF- κ B p65 subunit represses the Nrf2-antioxidant response element (ARE) pathway (Liu et al., 2008). Thus, inflammation increases ROS production through suppression of anti-oxidant pathways and inducing hypoxia. Activation of NF- κ B reduces the activity of p53 and increases the likelihood of cancer (Gudkov et al., 2011).

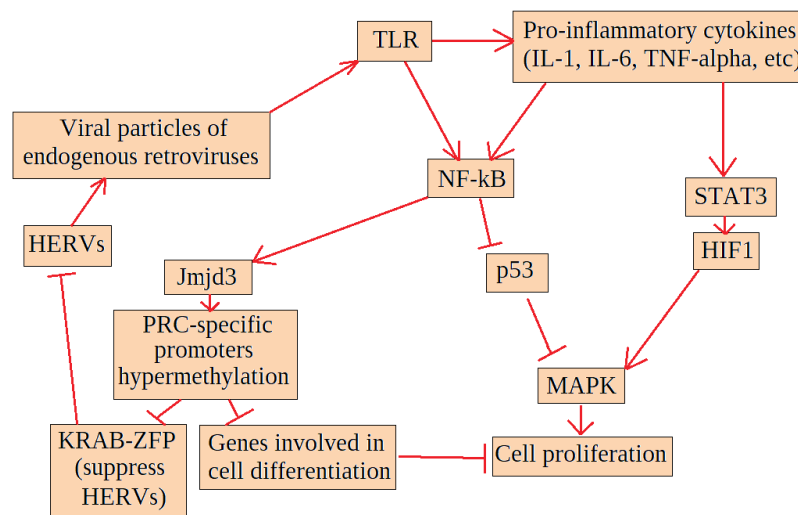


Figure 2. Endogenous retroviruses promotes NF- κ B upregulation leads to excessive cell proliferation and tumorigenesis.

Autophagy and energy metabolism

Cellular senescence is determined by interactions of the p53 and mTOR signaling. The mTOR signaling pathway is vital to accumulate energy-storing substances that are necessary for intensive growth and cell proliferation. However, the p53 signaling pathway prevents cell dedifferentiation and proliferation leading to cell overload with redundant metabolites and impaired cell self-cleaning. Thus, the mTOR-induced autophagy inhibition promotes cellular senescence in p53-arrested cells (Korotchikina et al., 2010). Moreover, innate immune response induces PI3K-Akt-mTOR signaling pathway that is a major regulator of macrophages (Covarrubias et al., 2015). Therefore, the innate immune response is important in modulation the mTOR pathway. Also, mTOR activation during aging contributes to a lower induction of autophagy (Romero et al., 2016). Autophagy plays a major role in tackling cellular junk. Aspects of autophagy include lipid metabolism through lipophagy (Singh and Cuervo, 2012; Singh et al., 2009), removing dysfunctional mitochondria (Sarparanta et al., 2017) and also eliminating damaged portions of the endoplasmic reticulum (Bernales et al., 2007). The mTOR signaling pathway regulates many fundamental metabolic and physiological processes, including lipid metabolism. mTOR is a central regulator of lipid metabolism, regulating not only lipogenesis and lipolysis but also adipogenesis (Lamming and Sabatini, 2013). Pro-inflammatory cytokine TNF- α leads to increased insulin resistance (Hotamisligil, 1999), and induces mTOR (Ozes et al., 2001). Senescence-associated β -galactosidase (SA- β -gal)-stained cells is linked with mTOR activity (Sung et al., 2018). mTOR activation is mediated through TLR activation (Schmitz et al., 2008). For example, toll-like receptors 4 (TLR4) affects lipid uptake and foam cell formation through mTOR (Banerjee et al., 2018) and inflammation-induced foam cell formation results in atherosclerosis (Angelovich et al., 2015).

Extracellular matrix and stem cells

Chronic inflammation may also determine the remodeling of extracellular matrix and visible aging in the skin through the action of matrix metalloproteinases (MMPs). NF- κ B activity up-regulates MMP-1,-3 and -9 (Bond et al., 2001). Matrix metalloproteinases cause extracellular matrix remodeling (Stamenkovic, 2003) through degeneration of its structural components mostly collagen. Extracellular matrix has fundamental importance in regulating epidermal stem cells maintenance, proper mobilization, and differentiation (Chermnykh et al., 2018). Extracellular matrix controls both embryonic and adult stem cell behavior (Ahmed and French-Constant, 2016). Down-regulation of stem cell maintenance due to chronic inflammation leads to stem cells depletion (Rosengarten et al., 2011). Therefore, stem cells depletion during aging may be explained due to inflammation-mediated extracellular matrix remodeling through MMPs.

Inflammation-mediated degeneration

Chronic inflammation during aging also promotes catabolic processes leading to complex body degeneration. Pro-inflammatory cytokines TNF- α and IL-1 induce inflammation-mediated osteoporosis (Lacativa and Farias, 2010). Chronic inflammation causes frailty (Fulop et al., 2015). Sarcopenia also is associated with chronic inflammation through an inflammatory marker – C-reactive protein (Bano et al., 2017). Circulating levels of another pro-inflammatory cytokine IL-6 determines the decrease of muscle mass during cancer (Carson and Baltgalvis, 2010). Frailty, sarcopenia, and immunosenescence appear to share common inflammatory drivers (Wilson et al., 2017).

Chronic inflammation interactions with endogenous retroviruses

The effect of chronic inflammation induced by innate immune response explains epigenetic alterations, increased ROS production, mild hypoxia and genome instability due to retrotransposons derepression. Among retrotransposons, production of viral proteins by endogenous retroviruses provide a clue for the innate immune response system. Viral proteins can be detected by toll-like receptors (TLRs) that recognize pathogen-associated molecular patterns (PAMPs) leading to activation of the NF- κ B and interferon pathways (Kawasaki and Kawai, 2014). In adult tissues, endogenous retroviruses are suppressed by KRAB-ZFP proteins (Hurst and Magiorkinis, 2017). Hypermethylation of a Cluster of KRAB-ZFP genes on Chromosome 19 in cancer (Lleras et al., 2011) reveals a role of endogenous retroviruses in cancer and aging because both have similar patterns of hypermethylation. For example, activation of HERV-K envelope protein is essential for tumorigenesis and metastasis of breast cancer cells (Zhou et al., 2016) due to stimulation of the innate immune system by endogenous viruses and therefore initiating the autoimmune response (Nexø et al., 2016). Transmembrane unit in the envelope of endogenous viruses also induces immunosuppression to evade the adaptive immune response promoting inability of adaptive immune system to suppress tumors (Morozov et al., 2013). HERV-K viral proteins may be biomarkers and/or tumor-associated antigens (Li et al., 2017). In contrary, the transcription of HERV-K was observed in normal human cell physiology (Schmitt et al., 2015). This collision may be explained through mutual antagonism between NF- κ B and steroid hormones (McKay and Cidlowski, 1998). In a healthy organism, normal level of steroid hormones prevents inflammation induced by viral particles from endogenous viruses.

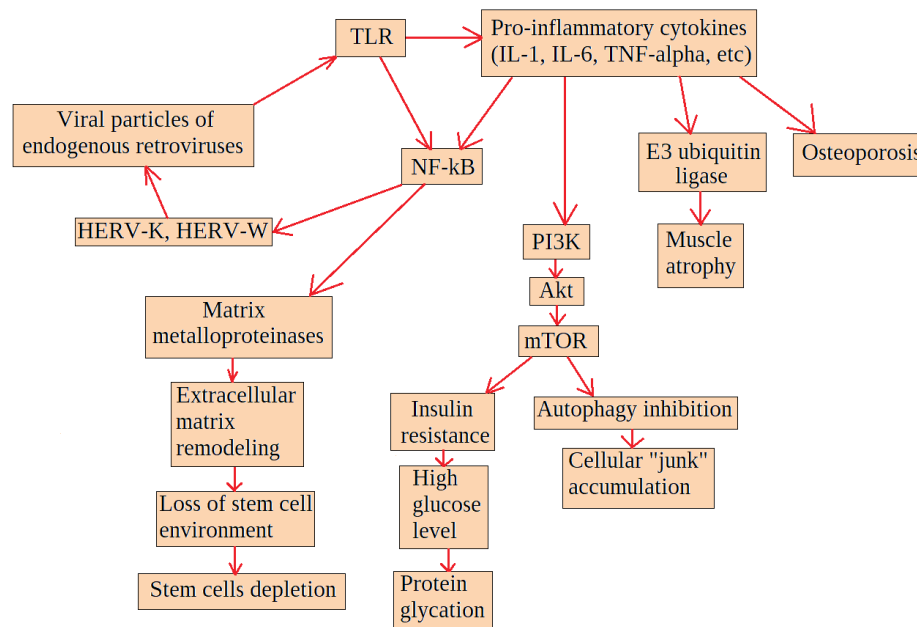


Figure 3. Endogenous retroviruses may provide senescent phenotype by inducing chronic inflammation

Dynamics of endogenous retroviruses and the endocrine system

Endogenous retroviruses induce the innate immune system and prolonged chronic inflammation (Hurst and Magiorkinis, 2015). Human endogenous retroviruses could trigger an innate immune response by producing viral particles that are similar the pathogen-associated molecular patterns (PAMPs) of exogenous viruses (Tang et al., 2012). Viral proteins stimulate the production of pro-inflammatory cytokines (IL-1 β , IL-6, and TNF-alpha) (Ariza and Williams, 2011; Rolland et al., 2006; Saito et al., 2017). These pro-inflammatory cytokines can induce NF- κ B, which could then bind to the LTR regulatory elements of HERVs. This was exhibited for HERV-W (Mameli et al., 2007) and HERV-K (Manghera and Douville, 2013). This establishes a positive feedback loop between NF- κ B and HERV-K and HERV-W expression that drives inflammation. HERV-K could additionally be induced by sex hormones (estrogen, progesterone, testosterone) (Manghera and Douville, 2013) linking HERV-K activity with the developmental program. The burst of sex hormones during maturation leads to inducing of HERV-K transcription activity. However, age-associated chronic inflammation is not developed immediately due to sex hormones which suppress the NF- κ B signaling pathway (McKay and Cidlowski, 1998). The equilibrium between NF- κ B and sex hormones prevents the deleterious effect of HERV-K because endogenous retroviruses lack lytic ability. Endogenous retroviruses are able to harm organism mostly through activation of innate immune response and then promoting chronic inflammation. It is demonstrated by the dynamics of the transcriptional levels of HERVs during the lifespan (Balestrieri et al., 2015). Median transcription level of HERV-K in childhood is negligible but dramatically elevates during puberty. The HERV-K transcription activity slightly decreases in young adults, but then constantly rises with age. The HERV-W activity drops in young adults and drastically surges in the middle-aged and slightly decreases in the elderly people (Balestrieri et al., 2015). Age-associated changes in the activity of HERVs explains why individual development is linked to aging

processes because growth retardation and late maturation lead to increased lifespan prompting an idea of the developmental origin of aging (de Magalhães, 2012). The rising activity of endogenous retroviruses cause chronic inflammation and suppresses sex hormones production through inflammation of hypothalamus (Zhang et al., 2013).

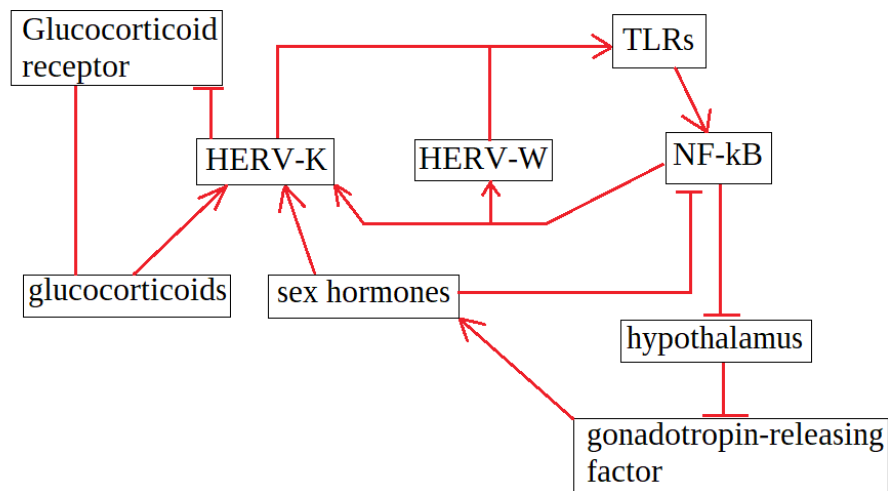


Figure 4. Mutual antagonism of pro-inflammatory pathways and sex hormones provides the delayed deleterious effect of endogenous retroviruses.

Conclusion

The detrimental program of aging is likely evolved during the transposon-driven evolution. Viral particles of endogenous retroviruses induce the innate immune response. This innate immune response is not accompanied by the adaptive immune response due to the ability of endogenous retroviruses to immune response evasion. The signaling pathways involved in innate immune response provide a clue how chronic inflammation induces senescence and age-associated diseases. Most aging-associated effects like DNA mutations and increased ROS production may be considered as attempts of adaptive systems of the organism to overcome intrinsic stress stimulus. However, the organism cannot overcome this stress stimulus because endogenous retroviruses are already part of the genome. Activation of endogenous retroviruses occurs during maturation but their deleterious effect is displayed only after some period of time when endogenous retroviruses are able to suppress sex hormone production through an impact on the endocrine system. Sex hormones suppress chronic inflammation in young adults and alterations in the endocrine system drive aging process. However, this program is very stable due to multiple copies of active endogenous retroviruses and cannot be completely broken without gene editing. The most active class of human endogenous retroviruses is HERV-K that has binding sites for NF- κ B and sex hormones and HERV-K is considered to be the most probable culprit of aging.

References

- Afanas'ev, I. (2013). New Nucleophilic Mechanisms of Ros-Dependent Epigenetic Modifications: Comparison of Aging and Cancer. *Aging Dis* 5, 52–62.
- Ahmed, M., and French-Constant, C. (2016). Extracellular Matrix Regulation of Stem Cell Behavior. *Curr Stem Cell Rep* 2, 197–206.
- Angelovich, T.A., Hearps, A.C., and Jaworowski, A. (2015). Inflammation-induced foam cell formation in chronic inflammatory disease. *Immunol. Cell Biol.* 93, 683–693.
- Ariza, M.-E., and Williams, M.V. (2011). A human endogenous retrovirus K dUTPase triggers a TH1, TH17 cytokine response: does it have a role in psoriasis? *J. Invest. Dermatol.* 131, 2419–2427.
- Balestrieri, E., Pica, F., Matteucci, C., Zenobi, R., Sorrentino, R., Argaw-Denboba, A., Cipriani, C., Bucci, I., and Sinibaldi-Vallebona, P. (2015). Transcriptional Activity of Human Endogenous Retroviruses in Human Peripheral Blood Mononuclear Cells. *Biomed Res Int* 2015.
- Banerjee, D., Sinha, A., Saikia, S., Gogoi, B., Rathore, A.K., Das, A.S., Pal, D., Buragohain, A.K., and Dasgupta, S. (2018). Inflammation-induced mTORC2-Akt-mTORC1 signaling promotes macrophage foam cell formation. *Biochimie* 151, 139–149.
- Bano, G., Trevisan, C., Carraro, S., Solmi, M., Luchini, C., Stubbs, B., Manzato, E., Sergi, G., and Veronese, N. (2017). Inflammation and sarcopenia: A systematic review and meta-analysis. *Maturitas* 96, 10–15.
- Belancio, V.P., Roy-Engel, A.M., Pochampally, R.R., and Deininger, P. (2010). Somatic expression of LINE-1 elements in human tissues. *Nucleic Acids Res* 38, 3909–3922.
- Bernales, S., Schuck, S., and Walter, P. (2007). ER-phagy: selective autophagy of the endoplasmic reticulum. *Autophagy* 3, 285–287.
- Bitterman, K.J., Medvedik, O., and Sinclair, D.A. (2003). Longevity Regulation in *Saccharomyces cerevisiae*: Linking Metabolism, Genome Stability, and Heterochromatin. *Microbiol Mol Biol Rev* 67, 376–399.
- Bond, M., Chase, A.J., Baker, A.H., and Newby, A.C. (2001). Inhibition of transcription factor NF-kappaB reduces matrix metalloproteinase-1, -3 and -9 production by vascular smooth muscle cells. *Cardiovasc. Res.* 50, 556–565.
- Brunet, T.D.P., and Doolittle, W.F. (2015). Multilevel Selection Theory and the Evolutionary Functions of Transposable Elements. *Genome Biol Evol* 7, 2445–2457.
- Carson, J.A., and Baltgalvis, K.A. (2010). Interleukin-6 as a Key Regulator of Muscle Mass during Cachexia. *Exerc Sport Sci Rev* 38, 168–176.

- Chermnykh, E., Kalabusheva, E., and Vorotelyak, E. (2018). Extracellular Matrix as a Regulator of Epidermal Stem Cell Fate. *Int J Mol Sci* 19.
- Covarrubias, A.J., Aksoylar, H.I., and Horng, T. (2015). Control of macrophage metabolism and activation by mTOR and Akt signaling. *Semin Immunol* 27, 286–296.
- Cruickshanks, H.A., McBryan, T., Nelson, D.M., VanderKraats, N.D., Shah, P.P., van Tuyn, J., Rai, T.S., Brock, C., Donahue, G., Dunican, D.S., et al. (2013). Senescent cells harbour features of the cancer epigenome. *Nat Cell Biol* 15, 1495–1506.
- De Cecco, M., Criscione, S.W., Peterson, A.L., Neretti, N., Sedivy, J.M., and Kreiling, J.A. (2013). Transposable elements become active and mobile in the genomes of aging mammalian somatic tissues. *Aging (Albany NY)* 5, 867–883.
- De Santa, F., Totaro, M.G., Prosperini, E., Notarbartolo, S., Testa, G., and Natoli, G. (2007). The histone H3 lysine-27 demethylase Jmjd3 links inflammation to inhibition of polycomb-mediated gene silencing. *Cell* 130, 1083–1094.
- Dewannieux, M., Esnault, C., and Heidmann, T. (2003). LINE-mediated retrotransposition of marked Alu sequences. *Nat. Genet.* 35, 41–48.
- Doolittle, W.F., and Sapienza, C. (1980). Selfish genes, the phenotype paradigm and genome evolution. *Nature* 284, 601–603.
- Doonan, R., McElwee, J.J., Matthijssens, F., Walker, G.A., Houthoofd, K., Back, P., Matscheski, A., Vanfleteren, J.R., and Gems, D. (2008). Against the oxidative damage theory of aging: superoxide dismutases protect against oxidative stress but have little or no effect on life span in *Caenorhabditis elegans*. *Genes Dev.* 22, 3236–3241.
- Evsikov, A.V., and Marín de Evsikova, C. (2016). Friend or Foe: Epigenetic Regulation of Retrotransposons in Mammalian Oogenesis and Early Development. *Yale J Biol Med* 89, 487–497.
- Failla, G. (1958). The aging process and cancerogenesis. *Ann. N. Y. Acad. Sci.* 71, 1124–1140.
- Fulop, T., McElhaney, J., Pawelec, G., Cohen, A.A., Morais, J.A., Dupuis, G., Baehl, S., Camous, X., Witkowski, J.M., and Larbi, A. (2015). Frailty, Inflammation and Immunosenescence. *Interdiscip Top Gerontol Geriatr* 41, 26–40.
- Gasior, S.L., Wakeman, T.P., Xu, B., and Deininger, P.L. (2006). The human LINE-1 retrotransposon creates DNA double-strand breaks. *J. Mol. Biol.* 357, 1383–1393.
- Gibcus, J.H., and Dekker, J. (2013). Connecting the genome: dynamics and stochasticity in a new hierarchy for chromosome conformation. *Mol Cell* 49, 773–782.
- Gudkov, A.V., Gurova, K.V., and Komarova, E.A. (2011). Inflammation and p53. *Genes Cancer* 2, 503–516.

- Guelen, L., Pagie, L., Brasset, E., Meuleman, W., Faza, M.B., Talhout, W., Eussen, B.H., de Klein, A., Wessels, L., de Laat, W., et al. (2008). Domain organization of human chromosomes revealed by mapping of nuclear lamina interactions. *Nature* 453, 948–951.
- Hannum, G., Guinney, J., Zhao, L., Zhang, L., Hughes, G., Sadda, S., Klotzle, B., Bibikova, M., Fan, J.-B., Gao, Y., et al. (2013). Genome-wide methylation profiles reveal quantitative views of human aging rates. *Mol. Cell* 49, 359–367.
- Harman, D. (1956). Aging: a theory based on free radical and radiation chemistry. *J Gerontol* 11, 298–300.
- Horvath, S. (2013). DNA methylation age of human tissues and cell types. *Genome Biology* 14, 3156.
- Horváth, V., Merenciano, M., and González, J. (2017). Revisiting the Relationship between Transposable Elements and the Eukaryotic Stress Response. *Trends in Genetics* 33, 832–841.
- Hotamisligil, G.S. (1999). Mechanisms of TNF-alpha-induced insulin resistance. *Exp. Clin. Endocrinol. Diabetes* 107, 119–125.
- Hurst, T.P., and Magiorkinis, G. (2015). Activation of the innate immune response by endogenous retroviruses. *Journal of General Virology* 96, 1207–1218.
- Hurst, T.P., and Magiorkinis, G. (2017). Epigenetic Control of Human Endogenous Retrovirus Expression: Focus on Regulation of Long-Terminal Repeats (LTRs). *Viruses* 9.
- Irvin, M.R., Aslibekyan, S., Do, A., Zhi, D., Hidalgo, B., Claas, S.A., Srinivasasainagendra, V., Horvath, S., Tiwari, H.K., Absher, D.M., et al. (2018). Metabolic and inflammatory biomarkers are associated with epigenetic aging acceleration estimates in the GOLDN study. *Clin Epigenetics* 10.
- Jintaridith, P., and Mutirangura, A. (2010). Distinctive patterns of age-dependent hypomethylation in interspersed repetitive sequences. *Physiol. Genomics* 41, 194–200.
- Jylhävä, J., Kananen, L., Raitanen, J., Marttila, S., Nevalainen, T., Hervonen, A., Jylhä, M., and Hurme, M. (2016). Methylomic predictors demonstrate the role of NF-κB in old-age mortality and are unrelated to the aging-associated epigenetic drift. *Oncotarget* 7, 19228–19241.
- Kawasaki, T., and Kawai, T. (2014). Toll-Like Receptor Signaling Pathways. *Front Immunol* 5.
- Kaya, A., Lobanov, A.V., and Gladyshev, V.N. (2015). Evidence that mutation accumulation does not cause aging in *Saccharomyces cerevisiae*. *Aging Cell* 14, 366–371.
- Koonin, E.V. (2016). Viruses and mobile elements as drivers of evolutionary transitions. *Philos Trans R Soc Lond B Biol Sci* 371.

- Korotchkina, L.G., Leontieva, O.V., Bukreeva, E.I., Demidenko, Z.N., Gudkov, A.V., and Blagosklonny, M.V. (2010). The choice between p53-induced senescence and quiescence is determined in part by the mTOR pathway. *Aging (Albany NY)* 2, 344–352.
- Lacativa, P.G.S., and Farias, M.L.F. de (2010). Osteoporosis and inflammation. *Arq Bras Endocrinol Metabol* 54, 123–132.
- Lamming, D.W., and Sabatini, D.M. (2013). A central role for mTOR in lipid homeostasis. *Cell Metab* 18.
- Levine, M.E., Lu, A.T., Quach, A., Chen, B.H., Assimes, T.L., Bandinelli, S., Hou, L., Baccarelli, A.A., Stewart, J.D., Li, Y., et al. (2018). An epigenetic biomarker of aging for lifespan and healthspan. *Aging (Albany NY)* 10, 573–591.
- Li, M., Radvanyi, L., Yin, B., Li, J., Chivukula, R., Lin, K., Lu, Y., Shen, J., Chang, D.Z., Li, D., et al. (2017). Down-regulation of human endogenous retrovirus type K (HERV-K) viral env RNA in pancreatic cancer cells decreases cell proliferation and tumor growth. *Clin Cancer Res* 23, 5892–5911.
- Lin, Q., and Wagner, W. (2015). Epigenetic Aging Signatures Are Coherently Modified in Cancer. *PLoS Genet* 11.
- Liu, G.-H., Qu, J., and Shen, X. (2008). NF- κ B/p65 antagonizes Nrf2-ARE pathway by depriving CBP from Nrf2 and facilitating recruitment of HDAC3 to MafK. *Biochimica et Biophysica Acta (BBA) - Molecular Cell Research* 1783, 713–727.
- Lleras, R.A., Adrien, L.R., Smith, R.V., Brown, B., Jivraj, N., Keller, C., Sarta, C., Schlecht, N.F., Harris, T.M., Childs, G., et al. (2011). Hypermethylation of a Cluster of Krüppel-Type Zinc Finger Protein Genes on Chromosome 19q13 in Oropharyngeal Squamous Cell Carcinoma. *The American Journal of Pathology* 178, 1965–1974.
- Local, A., Huang, H., Albuquerque, C.P., Singh, N., Lee, A.Y., Wang, W., Wang, C., Hsia, J.E., Shiau, A.K., Ge, K., et al. (2018). Identification of H3K4me1-Associated Proteins at Mammalian Enhancers. *Nat Genet* 50, 73–82.
- de Magalhães, J.P. (2012). Programmatic features of aging originating in development: aging mechanisms beyond molecular damage? *FASEB J.* 26, 4821–4826.
- Mameli, G., Astone, V., Khalili, K., Serra, C., Sawaya, B.E., and Dolei, A. (2007). Regulation of the syncytin-1 promoter in human astrocytes by multiple sclerosis-related cytokines. *Virology* 362, 120–130.
- Manghera, M., and Douville, R.N. (2013). Endogenous retrovirus-K promoter: a landing strip for inflammatory transcription factors? *Retrovirology* 10, 16.

- Marttila, S., Kananen, L., Häyrynen, S., Jylhävä, J., Nevalainen, T., Hervonen, A., Jylhä, M., Nykter, M., and Hurme, M. (2015). Ageing-associated changes in the human DNA methylome: genomic locations and effects on gene expression. *BMC Genomics* 16.
- McClintock, B. (1984). The significance of responses of the genome to challenge. *Science* 226, 792–801.
- McKay, L.I., and Cidlowski, J.A. (1998). Cross-talk between nuclear factor-kappa B and the steroid hormone receptors: mechanisms of mutual antagonism. *Mol. Endocrinol.* 12, 45–56.
- Mitteldorf, J. (2010). Aging is not a process of wear and tear. *Rejuvenation Res* 13, 322–326.
- Morozov, V.A., Dao Thi, V.L., and Denner, J. (2013). The Transmembrane Protein of the Human Endogenous Retrovirus - K (HERV-K) Modulates Cytokine Release and Gene Expression. *PLoS One* 8.
- Moskalev, A.A., Shaposhnikov, M.V., Plyusnina, E.N., Zhavoronkov, A., Budovsky, A., Yanai, H., and Fraifeld, V.E. (2013). The role of DNA damage and repair in aging through the prism of Koch-like criteria. *Ageing Res. Rev.* 12, 661–684.
- Nexø, B.A., Villesen, P., Nissen, K.K., Lindegaard, H.M., Rossing, P., Petersen, T., Tarnow, L., Hansen, B., Lorenzen, T., Hørslev-Petersen, K., et al. (2016). Are human endogenous retroviruses triggers of autoimmune diseases? Unveiling associations of three diseases and viral loci. *Immunol Res* 64, 55–63.
- Niu, G., Briggs, J., Deng, J., Ma, Y., Lee, H., Kortylewski, M., Kujawski, M., Kay, H., Cress, W.D., Jove, R., et al. (2008). Signal transducer and activator of transcription 3 is required for hypoxia-inducible factor-1alpha RNA expression in both tumor cells and tumor-associated myeloid cells. *Mol. Cancer Res.* 6, 1099–1105.
- Orgel, L.E., and Crick, F.H. (1980). Selfish DNA: the ultimate parasite. *Nature* 284, 604–607.
- Osorio, F.G., Bárcena, C., Soria-Valles, C., Ramsay, A.J., de Carlos, F., Cobo, J., Fueyo, A., Freije, J.M.P., and López-Otín, C. (2012). Nuclear lamina defects cause ATM-dependent NF-κB activation and link accelerated aging to a systemic inflammatory response. *Genes Dev.* 26, 2311–2324.
- Ozes, O.N., Akca, H., Mayo, L.D., Gustin, J.A., Maehama, T., Dixon, J.E., and Donner, D.B. (2001). A phosphatidylinositol 3-kinase/Akt/mTOR pathway mediates and PTEN antagonizes tumor necrosis factor inhibition of insulin signaling through insulin receptor substrate-1. *Proc Natl Acad Sci U S A* 98, 4640–4645.
- Pérez, R.F., Tejedor, J.R., Bayón, G.F., Fernández, A.F., and Fraga, M.F. (2018). Distinct chromatin signatures of DNA hypomethylation in aging and cancer. *Aging Cell* 17, e12744.

Peric-Hupkes, D., Meuleman, W., Pagie, L., Bruggeman, S.W.M., Solovei, I., Brugman, W., Gräf, S., Flicek, P., Kerkhoven, R.M., van Lohuizen, M., et al. (2010). Molecular maps of the reorganization of genome – nuclear lamina interactions during differentiation. *Mol Cell* 38, 603–613.

Pialoux, V., Mounier, R., Brown, A.D., Steinback, C.D., Rawling, J.M., and Poulin, M.J. (2009). Relationship between oxidative stress and HIF-1 alpha mRNA during sustained hypoxia in humans. *Free Radic. Biol. Med.* 46, 321–326.

Rakyan, V.K., Down, T.A., Maslau, S., Andrew, T., Yang, T.-P., Beyan, H., Whittaker, P., McCann, O.T., Finer, S., Valdes, A.M., et al. (2010). Human aging-associated DNA hypermethylation occurs preferentially at bivalent chromatin domains. *Genome Res* 20, 434–439.

Ren, J.-T., Wang, M.-X., Su, Y., Tang, L.-Y., and Ren, Z.-F. (2018). Decelerated DNA methylation age predicts poor prognosis of breast cancer. *BMC Cancer* 18.

Rolland, A., Jouvin-Marche, E., Viret, C., Faure, M., Perron, H., and Marche, P.N. (2006). The envelope protein of a human endogenous retrovirus-W family activates innate immunity through CD14/TLR4 and promotes Th1-like responses. *J. Immunol.* 176, 7636–7644.

Romero, Y., Bueno, M., Ramirez, R., Álvarez, D., Sembrat, J.C., Goncharova, E.A., Rojas, M., Selman, M., Mora, A.L., and Pardo, A. (2016). mTORC1 activation decreases autophagy in aging and idiopathic pulmonary fibrosis and contributes to apoptosis resistance in IPF fibroblasts. *Aging Cell* 15, 1103–1112.

Rosengardten, Y., McKenna, T., Grochová, D., and Eriksson, M. (2011). Stem cell depletion in Hutchinson-Gilford progeria syndrome. *Aging Cell* 10, 1011–1020.

Saito, T., Miyagawa, K., Chen, S.-Y., Tamosiuniene, R., Wang, L., Sharpe, O., Samayoa, E., Harada, D., Moonen, J.-R.A.J., Cao, A., et al. (2017). Upregulation of Human Endogenous Retrovirus-K Is Linked to Immunity and Inflammation in Pulmonary Arterial Hypertension. *Circulation* 136, 1920–1935.

Saleiro, D., and Platanias, L.C. (2015). Intersection of mTOR and STAT signaling in immunity. *Trends Immunol* 36, 21–29.

Salminen, A., Huuskonen, J., Ojala, J., Kauppinen, A., Kaarniranta, K., and Suuronen, T. (2008). Activation of innate immunity system during aging: NF-κB signaling is the molecular culprit of inflamm-aging. *Ageing Res. Rev.* 7, 83–105.

Sarparanta, J., García-Macia, M., and Singh, R. (2017). Autophagy and Mitochondria in Obesity and Type 2 Diabetes. *Curr Diabetes Rev* 13, 352–369.

Scaffidi, P., and Misteli, T. (2006). Lamin A-Dependent Nuclear Defects in Human Aging. *Science* 312, 1059–1063.

- Schmitt, K., Heyne, K., Roemer, K., Meese, E., and Mayer, J. (2015). HERV-K(HML-2) rec and np9 transcripts not restricted to disease but present in many normal human tissues. *Mob DNA* 6, 4.
- Schmitz, F., Heit, A., Dreher, S., Eisenächer, K., Mages, J., Haas, T., Krug, A., Janssen, K.-P., Kirschning, C.J., and Wagner, H. (2008). Mammalian target of rapamycin (mTOR) orchestrates the defense program of innate immune cells. *Eur. J. Immunol.* 38, 2981–2992.
- Serrato-Capuchina, A., and Matute, D.R. (2018). The Role of Transposable Elements in Speciation. *Genes (Basel)* 9.
- Singh, R., and Cuervo, A.M. (2012). Lipophagy: connecting autophagy and lipid metabolism. *Int J Cell Biol* 2012, 282041.
- Singh, R., Kaushik, S., Wang, Y., Xiang, Y., Novak, I., Komatsu, M., Tanaka, K., Cuervo, A.M., and Czaja, M.J. (2009). Autophagy regulates lipid metabolism. *Nature* 458, 1131–1135.
- Stamenkovic, I. (2003). Extracellular matrix remodelling: the role of matrix metalloproteinases. *J. Pathol.* 200, 448–464.
- Stuart, J.A., Maddalena, L.A., Merilovich, M., and Robb, E.L. (2014). A midlife crisis for the mitochondrial free radical theory of aging. *Longev Healthspan* 3, 4.
- Sung, J.Y., Lee, K.Y., Kim, J.-R., and Choi, H.C. (2018). Interaction between mTOR pathway inhibition and autophagy induction attenuates adriamycin-induced vascular smooth muscle cell senescence through decreased expressions of p53/p21/p16. *Exp. Gerontol.* 109, 51–58.
- Szilard, L. (1959). ON THE NATURE OF THE AGING PROCESS. *Proc. Natl. Acad. Sci. U.S.A.* 45, 30–45.
- Tang, D., Kang, R., Coyne, C.B., Zeh, H.J., and Lotze, M.T. (2012). PAMPs and DAMPs: signal 0s that spur autophagy and immunity. *Immunol. Rev.* 249, 158–175.
- Teschendorff, A.E., Menon, U., Gentry-Maharaj, A., Ramus, S.J., Weisenberger, D.J., Shen, H., Campan, M., Noushmehr, H., Bell, C.G., Maxwell, A.P., et al. (2010). Age-dependent DNA methylation of genes that are suppressed in stem cells is a hallmark of cancer. *Genome Res.* 20, 440–446.
- Tilstra, J.S., Robinson, A.R., Wang, J., Gregg, S.Q., Clauson, C.L., Reay, D.P., Nasto, L.A., St Croix, C.M., Usas, A., Vo, N., et al. (2012). NF-κB inhibition delays DNA damage-induced senescence and aging in mice. *J. Clin. Invest.* 122, 2601–2612.
- Valinluck, V., and Sowers, L.C. (2007). Endogenous cytosine damage products alter the site selectivity of human DNA maintenance methyltransferase DNMT1. *Cancer Res.* 67, 946–950.

Widschwendter, M., Fiegl, H., Egle, D., Mueller-Holzner, E., Spizzo, G., Marth, C., Weisenberger, D.J., Campan, M., Young, J., Jacobs, I., et al. (2007). Epigenetic stem cell signature in cancer. *Nat. Genet.* 39, 157–158.

Wilson, D., Jackson, T., Sapey, E., and Lord, J.M. (2017). Frailty and sarcopenia: The potential role of an aged immune system. *Ageing Research Reviews* 36, 1–10.

Wood, J.G., and Helfand, S.L. (2013). Chromatin structure and transposable elements in organismal aging. *Front Genet* 4.

Zhang, G., Li, J., Purkayastha, S., Tang, Y., Zhang, H., Yin, Y., Li, B., Liu, G., and Cai, D. (2013). Hypothalamic Programming of Systemic Aging Involving IKK β /NF- κ B and GnRH. *Nature* 497, 211–216.

Zhao, H., Sifakis, E.G., Sumida, N., Millán-Ariño, L., Scholz, B.A., Svensson, J.P., Chen, X., Ronnegren, A.L., Mallet de Lima, C.D., Varnoosfaderani, F.S., et al. (2015). PARP1- and CTCF-Mediated Interactions between Active and Repressed Chromatin at the Lamina Promote Oscillating Transcription. *Mol. Cell* 59, 984–997.

Zhou, F., Li, M., Wei, Y., Lin, K., Lu, Y., Shen, J., Johanning, G.L., and Wang-Johanning, F. (2016). Activation of HERV-K Env protein is essential for tumorigenesis and metastasis of breast cancer cells. *Oncotarget* 7, 84093–84117.