A peer-reviewed version of this preprint was published in PeerJ on 28 March 2019.

View the peer-reviewed version (peerj.com/articles/6600), which is the preferred citable publication unless you specifically need to cite this preprint.

https://doi.org/10.7717/peerj.6600
Evaluation of a toxoid fusion protein vaccine produced in plants to protect poultry against necrotic enteritis

Joseph G Hunter 1,2, Shyra Wilde 1, Amanda M Tafoya 1, Jamie Horsman 1, Miranda Yousif 1, Andrew G Diamos 1,2, Kenneth L Roland 1, Hugh S Mason Corresp. 1,2

1 Center for Immunotherapy, Vaccines and Virotherapy, The Biodesign Institute, Arizona State University, Tempe, AZ, United States
2 School of Life Sciences, Arizona State University, Tempe, AZ, United States

Corresponding Author: Hugh S Mason
Email address: hugh.mason@asu.edu

Background. Necrotic enteritis (NE) is caused by type A strains of the bacterium Clostridium perfringens. Total global economic losses to the poultry industry due to NE is estimated to be over 2 billion dollars annually. Traditionally, NE has been effectively controlled by inclusion of antibiotics in the diet of poultry. However, recent concerns regarding the impact of this practice on increasing antibiotic resistance in human pathogens have led us to consider alternative approaches, such as vaccination, for controlling this disease. NE strains of C. perfringens produce two major toxins, a-toxin and NetB. Immune responses against either toxin can provide partial protection against NE.

Methods. We have developed a fusion protein combining a non-toxic carboxyl-terminal domain of a-toxin (PlcC) and an attenuated, mutant form of NetB (NetB-W262A) for use as a vaccine antigen to immunize poultry against NE. We utilized a DNA sequence that was codon-optimized for Nicotiana benthamiana to enable high levels of expression. The 6-His tagged PlcC-NetB fusion protein was synthesized in N. benthamiana using a geminiviral replicon transient expression system, purified by metal affinity chromatography, and used to immunize broiler birds.

Results. Immunized birds produced a strong serum IgY response against both the plant produced PlcC-NetB protein and against bacterially produced His-PlcC and His-NetB. Immunized birds were significantly protected against a subsequent in-feed challenge with virulent C. perfringens when treated with the fusion protein. These results indicate that a plant-produced PlcC-NetB toxoid is a promising vaccine candidate for controlling NE in poultry.
Evaluation of a toxoid fusion protein vaccine produced in plants to protect poultry against necrotic enteritis

Joseph G. L. Hunter\textsuperscript{1,2A}, Shyra Wilde\textsuperscript{1AB}, Amanda M. Tafoya\textsuperscript{1}, Jamie Horsman\textsuperscript{A}, Miranda Yousif\textsuperscript{1}, Andrew G. Diamos\textsuperscript{1,2}, Kenneth L. Roland\textsuperscript{1} and Hugh S. Mason\textsuperscript{1,2} *

\textsuperscript{1}Center for Immunotherapy, Vaccines and Virotherapy, The Biodesign Institute, Arizona State University, Tempe, Arizona, USA

\textsuperscript{2}School of Life Sciences, Arizona State University, Tempe, Arizona, USA

\textsuperscript{A}These authors contributed equally to this work

\textsuperscript{B}Current address: Department of Microbiology and Molecular Genetics, Emory University

*Corresponding Author

Email addresses:

Joseph Hunter - jghunte1@asu.edu

Shyra Wilde - shyra.lynn6@gmail.com

Amanda M. Tafoya - Amanda.Gonzales@asu.edu

Jamie Horsman - jhorsma1@asu.edu
23  Miranda Yousif - mlyousif@asu.edu
24  Andrew Diamos - adiamos@asu.edu
25  Kenneth L. Roland – Kenneth.roland@asu.edu
26  Hugh Mason - Hugh.Mason@asu.edu
Abstract

Background. Necrotic enteritis (NE) is caused by type A strains of the bacterium *Clostridium perfringens*. Total global economic losses to the poultry industry due to NE is estimated to be over 2 billion dollars annually. Traditionally, NE has been effectively controlled by inclusion of antibiotics in the diet of poultry. However, recent concerns regarding the impact of this practice on increasing antibiotic resistance in human pathogens have led us to consider alternative approaches, such as vaccination, for controlling this disease. NE strains of *C. perfringens* produce two major toxins, \(\alpha\)-toxin and NetB. Immune responses against either toxin can provide partial protection against NE.

Methods. We have developed a fusion protein combining a non-toxic carboxyl-terminal domain of \(\alpha\)-toxin (PlcC) and an attenuated, mutant form of NetB (NetB-W262A) for use as a vaccine antigen to immunize poultry against NE. We utilized a DNA sequence that was codon-optimized for *Nicotiana benthamiana* to enable high levels of expression. The 6-His tagged PlcC-NetB fusion protein was synthesized in *N. benthamiana* using a geminiviral replicon transient expression system, purified by metal affinity chromatography, and used to immunize broiler birds.

Results. Immunized birds produced a strong serum IgY response against both the plant produced PlcC-NetB protein and against bacterially produced His-PlcC and His-NetB. Immunized birds were significantly protected against a subsequent in-feed challenge with virulent *C. perfringens* when treated with the fusion protein. These results indicate that a plant-produced PlcC-NetB toxoid is a promising vaccine candidate for controlling NE in poultry.
INTRODUCTION

Clostridium perfringens (C. perfringens), a Gram-positive anaerobic spore-forming bacterium, is the causative agent of necrotic enteritis (NE) in poultry and has become an economically significant issue for the broiler industry [1]. The pathogenesis of NE is complex and results from the concerted actions of a number of Clostridial genes combined with reduced gut health generated by environmental factors, feed, mycotoxins or the presence of other pathogens [2, 3]. NE typically occurs in broiler chicks at 2 to 6 weeks of age, but may occur in older birds as well [4, 5]. The onset of NE at around 2 weeks is believed to be due to waning levels of anti-Clostridium maternal antibodies [5]. The acute form of the disease leads to increased mortality in broiler flocks, which can account for high losses of up to 1% per day, reaching mortality rates up to 10-40% [6]. In the subclinical form, damage to the intestinal mucosa caused by C. perfringens leads to poor productivity (reduced growth, reduced feed efficiency) without mortality. The estimated cost of subclinical NE is around $0.05 per chicken with a total global loss of nearly $2 billion per year [7, 8].

C. perfringens is ubiquitous; readily found in soil, dust, feces, feed, used poultry litter and is normally present in the intestines of healthy chickens. The enterotoxaemia that results in clinical disease most often occurs either following an alteration in the intestinal microflora or from a condition that results in damage to the intestinal mucosa (e.g. coccidiosis). High dietary levels of animal by-products such as fishmeal, or certain grains with
high levels of indigestible fiber including wheat, barley, oats, and rye predispose birds to NE.

Dietary fat source can also influence the *C. perfringens* population [9]. Current thinking is that predisposing factors such as a high protein diet (e.g. fishmeal) and/or *Eimeria* infection result in alterations of the chicken gut microbiota that allow incoming or resident *C. perfringens* strains to become established [10].

Historically, *C. perfringens* outbreaks in the broiler industry were avoided using growth-promoting antimicrobials in the diet. However, concerns regarding antibiotic resistance led to restrictions on the use of antibiotics [11]. When antibiotics were banned in European and Scandinavian countries, it led to increases in *C. perfringens* infections, including NE [12]. Thus, other methods for controlling *C. perfringens* infections are needed.

Toxins have traditionally been targeted as antigens of interest for controlling clostridial infections. The *C. perfringens* α-toxin, encoded by the *plc* gene, is the major virulence determinant for gas gangrene [13]. Antibodies to *C. perfringens* α-toxin prevent gas gangrene in mice [14]. The protein is divided into two domains; the amino-terminal domain and carboxy-terminal domain [15]. The amino-terminal domain encodes the catalytic site responsible for phospholipase activity [15-17], while the carboxy-terminal domain is involved in interactions with phospholipids, targeting the enzyme to host cell membranes [18]. The α-toxin carboxy-terminal fragment (aa 247 – 370) is non-toxic [19] and immunization with this fragment confers protection against α-toxin and *C. perfringens* in a gas gangrene mouse model [14]. In chickens, immune responses against the C-terminal domain (PlcC) can provide protection against intestinal lesions following challenge with *C. perfringens* [14, 20-22].

For many years, the chromosomally-encoded α-toxin was considered to be the major toxin associated with NE [13, 14, 23, 24]. The C-terminal domain of the α-toxin (PlcC) has been
studied extensively as a vaccine against *C. perfringens* infection, delivered as a purified protein [25-27] or by live attenuated bacteria [22, 28]. Currently the only commercially available vaccine for necrotic enteritis is composed of an α-toxoid derived from a *C. perfringens* type A strain [29].

More recent studies have identified a β-like toxin, designated NetB toxin, linked to necrotic enteritis. It was originally identified in a *C. perfringens* type A strain isolated from an Australian outbreak of NE [30]. Keyburn et al. showed that *C. perfringens* supernatants contain a factor that is toxic for chicken leghorn male hepatoma cells [30]. The factor was present in supernatant from a *plc* knockout mutant, but not a *netB* knockout mutant of *C. perfringens*, implicating NetB as the toxin. Further, the *C. perfringens netB* mutant was attenuated for virulence in chickens, whereas a *plc* mutant of the same parent strain retained full virulence [31].

Screens for *netB* in *C. perfringens* field isolates indicate that the presence of the *netB* gene is highly correlated with necrotic enteritis causing strains [30, 32, 33]. Thus, NetB has emerged as a critical virulence factor. NetB functions by binding to cholesterol in membranes, forming heptameric pores [34, 35]. There are a number of single amino acid substitutions in the rim loop region of the protein, believed to interact with the host membrane, that significantly reduce its toxicity. These include Y191A, R200A, W257A and W262A [34] and S254L, R230Q and W287R [35].

NetB is also a protective antigen, providing significant protection against *C. perfringens* challenge, particularly in combination with other immunogenic components [36-38]. In addition, immunization with non-toxic rim mutants, W262A [36] and S254L [38], have been shown to generate protective immune responses in chickens. A study that examined serum antibody levels against *C. perfringens* α-toxin and NetB toxin in commercial birds from field outbreaks of NE
showed that the levels of serum antibodies against both α-toxin and NetB toxin were significantly higher in apparently healthy chickens compared to birds with clinical signs of NE, suggesting that these antitoxin antibodies may play a role in protection [39]. These results indicate a correlation between the presence of antitoxin antibodies in the serum and protective immunity against NE.

In a previous study, we used a live attenuated Salmonella vaccine to deliver PlcC, NetB, or both to chickens. We found that birds immunized against both toxins developed fewer severe lesions after C. perfringens challenge than birds immunized with either antigen alone [20]. Based on this finding, we have been developing approaches to combine the immunogenic components of each toxoid into a single protein. In this study, we describe a novel plcC-netB fusion gene modified to encode the rim mutation W262A and codon-optimized for production in Nicotiana benthamiana. The gene was introduced into N. benthamiana via Agrobacterium tumefaciens-mediated transformation of a geminiviral replicon transient expression system. The resulting fusion protein was purified and used to inject broiler chicks. Results indicate that the protein was highly immunogenic and elicited partial protection against challenge with a virulent strain of C. perfringens.

MATERIALS AND METHODS

2.1 Expression vector construction

A PlcC-NetB fusion protein gene was designed using codons preferred for N. benthamiana [40]. Amino acid sequences were based on GenBank accessions AAP15462.1 for PlcC and ABW71134.1 for NetB. For targeting of nascent protein to the endoplasmic reticulum (ER), the barley alpha amylase signal peptide, followed by a 6His tag, was added to the N-
terminus of PlcC. A linker peptide GGSGGSGGPSGGSGG was inserted between the PlcC and NetB sequences (Fig. 1). The fusion protein nucleotide sequences are in GenBank accession numbers MH883808 and MH883809. The gene was synthesized commercially with flanking XbaI and SacI sites. The 1402 bp XbaI-SacI fragment was ligated with geminiviral vector pBYR2eK2M [41] to make pBYR2eK2M-6HplcCnetB. Removal of the 417 bp BamHI fragment containing the PlcC and linker sequences yielded the ER-targeted 6His-NetB construct pBYR2eK2M-6HnetB. To make the ER-targeted 6His-PlcC vector, we PCR end-tailored the PlcC sequence to introduce a stop codon and SacI site at its 3’ end, using the primer PlcC-Sac-R (Table S1), inserting the resulting product to yield pBYR2eK2M-6HplcC. To produce the cytosol-targeted proteins, the signal peptide was removed by introducing an XbaI site and start codon 5’ of the 6His sequence by PCR with primer 6H-Xba-F and using the ER-targeting constructs above as templates. The resulting XbaI-SacI fragments were ligated with plasmid pBYR2e3K2Mc-GFP [42, 43] digested XbaI-SacI, thus yielding the vectors pBYR2e3K2Mc-c6HplcCnetB, pBYR2e3K2Mc-c6HplcC, and pBYR2e3K2Mc-c6HnetB.

2.2 Agroinfiltration of plants for protein production

Expression vectors were inserted into Agrobacterium tumefaciens EHA105 via electroporation. PCR and restriction digestion of the purified plasmids were used to confirm the resulting strains. Agrobacterium strains were grown overnight at 30°C in YENB (7.5 g Yeast Extract and 8 g Nutrient Broth for 1 liter) + 50 mg/L kanamycin and 2.5 mg/L rifampicin. Bacteria were pelleted for 10 minutes at 5,000g, then resuspended in infiltration buffer [10 mM 2-(N-morpholino) ethanesulfonic acid (MES), pH 5.5 and 10 mM MgSO₄] with a final OD600 = 0.2. Suspensions were infiltrated into N. benthamiana leaves at 5- to 6-weeks of age using a
syringe without needle via a small puncture [43]. Infiltrated leaves were harvested at 5 days post infiltration (DPI).

2.3 Protein extraction, acid precipitation clarification, and filtration

Total protein was extracted from agroinfiltrated leaf samples using a 1:3 (w:v) ratio of extraction buffer (50 mM sodium phosphate, 200 mM sodium chloride, pH 8.1) at 4°C. Leaf samples were homogenized for 5-10 min until an even consistency was obtained using a two speed Conair™ Waring™ Laboratory Blender (Fisher Scientific, Waltham, MA). Homogenized leaf tissue was stirred at high speed for 10 minutes at 4°C to improve protein solubility.

Acid precipitation was performed on crude extracts after this 10-minute stirring period. Note that no filtration, sedimentation, or centrifugation was performed before acid precipitation. 1 M phosphoric acid was added to the crude extract as it spun until a pH of 4.4-4.6 was reached, as measured by a VWR Sb70 Symphony Ph Meter (VWR, Radnor, PA). Extract was stirred for 10 min at 4°C. 2 M Tris base was then added to the extract until pH reached 7.5-7.7, and the mixture was stirred for another 10 min at 4°C before being filtered through Miracloth and centrifuged at 17,000 g, 4°C, for 30 min. The extract was then subjected to a second filtration through Miracloth resulting in a clarified extract, which was passed through a 0.45-micron filter for sterilization before metal affinity chromatography.

Three mL of Talon metal affinity resin (Clontech/Takara Bio, Mountain View, CA) was spun down at 10,000 g, 4°C, for 5 min and the supernatant removed from the bed. The resulting 1.5 mL resin bed was washed using 15 mL of extraction buffer (referred to below as wash buffer) and once again spun down at 10,000 g, 4°C, for 5 min. Supernatant was discarded and the resin resuspended in 15 mL wash buffer before being added to a 20 mL column. A final wash of
20 mL was performed before the filtered extract was passed through the column. The resin was 
washed with 60 mL of wash buffer, and elution of bound proteins achieved with PBS + 150 mM 
imidazole, pH 7.4, collected in five 1 mL samples. Elution samples were dialyzed overnight in 1 
liter of sterile PBS using either a 3 mL or 10 mL 3,500 MWCO Slide-A-Lyzer G2 dialysis 
cassette (Thermo Scientific, Waltham, MA), following manufacturer’s instructions. Elution 
concentrations were determined both before and after dialysis by an A$_{280}$ absorption using PBS 
and PBS + 150 mM imidazole, respectively, as blank solutions. Protein concentrations were 
confirmed by Coomassie staining of SDS-PAGE (below) analyzed with ImageJ [44].

2.4 SDS-PAGE and Western blot

Crude extracts, clarified extracts, and eluted samples were mixed with SDS sample buffer 
(50 mM Tris-HCl, pH 6.8, 2% SDS, 10% glycerol, 0.02 % bromophenol blue) under reducing 
conditions (0.5 M DTT) and samples boiled for 10 min and separated on 4-15% polyacrylamide 
gels (Bio-Rad, Hercules, CA). Gels were stained with Coomassie stain (Bio-Rad) or electroblot 
transferred to a PVDF membrane. PVDF membranes were blocked with 5% PBSTM (PBS with 
0.05% Tween-20 and 5% nonfat dry milk) overnight at 4°C. Membranes were then washed with 
PBST before incubation with a 1:10,000 dilution of rabbit anti-PlcC or rabbit anti-NetB 
antibodies [20] in 1% PBSTM for 1 hour at 37°C. After washing a second time the membranes 
were incubated with a 1:10,000 dilution of goat anti-rabbit horseradish peroxidase conjugate in 
1% PBSTM for 1 hour at 37°C. Bound antibody was detected with ECL reagent (Amersham 
Biosciences, San Francisco USA). Stripping of blots for reprobing was performed using a 
modified Abcam protocol. PVDF membrane was washed for 10 min in mild stripping buffer (for 
1 liter: 15 g glycine, 1 g SDS, 10 mL Tween20, pH 2.2) per Abcam’s recipe. PVDF membrane
was then rinsed twice in 20 mL of PBS for 10 minutes followed by two 5-min rinse steps in 20 mL PBST. 5% PBSTM was used to block the membrane before reprobing.

2.6 Production of recombinant proteins in *E. coli* for immune assays

His-tagged recombinant PlcC (rPlcC) and NetB (rNetB) were cloned by PCR using primer sets Plc_pqe_f1/Plc_pqe_r2 and netB_pqe_f3/netB_pqe_r4, respectively (Table S1), using pYA5130 as the template. The resulting PCR fragments were digested with BamHI and SalI and ligated to BamHI/SalI-digested plasmid pQE30. Plasmids carrying the desired inserts were moved into *E. coli* strain M15(pREP4) (Qiagen). Protein synthesis was induced by the addition of IPTG according to the manufacturer’s instructions. rPlcC and rNetB were purified by passage over a TALON affinity column (Clontech Laboratories, Inc, Mountain View, CA) following the manufacturer’s recommendations. The GST-NetB protein shown in Figures 2 and 3 was purified as previously described [20].

2.7 Chicken experiments

All animal experiments were conducted in compliance with the Arizona State University Institutional Animal Care and Use Committee (Protocol 16-1480R) and the Animal Welfare Act, in accordance with the National Institutes of Health guide for the care and use of laboratory animals (NIH Publication No. 8023, revised 1978).

Groups of one-day-old Cornish × Rock broiler chickens were purchased from the Murray McMurray Hatchery (Webster City, IA) and randomly sorted and placed in pens with pine shavings on the floor. Food and water was supplied *ad libitum*. For experiments 1 and 2, groups of birds were immunized subcutaneously with 50 µg of fusion protein (Exp. 1) or 100 µg of
fusion protein (Exp. 2) with 50 µg of Quil-A adjuvant. Birds were given three immunizations at weekly intervals beginning at 8 days of age, with the final immunization at 22 days of age. Mock vaccinated controls were injected with adjuvant only. At 27 days of age, blood was taken for serum analysis. Three days later, when the birds were 30 days of age, birds were given an in-feed challenge with virulent *C. perfringens* for 5 days. The birds were scored for intestinal lesions the next day, at 35 days of age. For experiment 3, birds were injected with 50 µg of PlcC-NetB at 7 days of age and again at 12 days of age. Blood and tissues were taken for immunological analysis ten days after the boost. Birds were challenged two weeks after the final boost, when the birds were 26 days of age.

2.8 ELISA

ELISAs were performed in triplicate as described to determine the IgY responses against his-tagged PlcC, his-tagged Fba, his-tagged NetB and S. Typhimurium lipopolysaccharide (LPS) in chicken sera and IgA, IgY and IgM responses in intestinal washes. Biotinylated anti-chicken IgA (Alpha Diagnostic Intl. Inc), IgY (Southern Biotechnology) or IgM (Bioss) antibodies diluted 1:10,000 were used to detect the various antibody isotypes.

2.9 Cellular proliferation assay

A proliferation assay was performed to evaluate cell-mediated immunity. Lymphocytes in the blood were harvested using the gentle swirl technique [1] and plated in quadruplicate, in a 96-well plate at 10^5 cells/well in RPMI-1640 without phenol red. Spleens were placed through a 70 µm cell strainer to obtain single cell suspensions. Red blood cells were lysed with Red Blood Cell Lysis solution (eBioscience). Splenocytes were then washed, suspended in RPMI and
plated at 10^6 cells/well. Each set of cells was incubated at 37°C, 5% CO₂ for 72 h with or without 4 µg/ml of either His-Fba, His-NetB, His-PlcC, or 1 µg/ml PMA. Cell proliferation was measured using the Vybrant®MTT Cell Proliferation Assay Kit (Molecular Probes). Mean absorbance value of antigen stimulated wells divided by mean absorbance of non-stimulated control wells was used to calculate stimulation index.

2.10 Challenge with C. perfringens

Chickens were fed an antibiotic-free starter feed containing 21% protein for 20 days, at which time the feed was switched to a high protein (28% protein), wheat-based feed containing 36% fish meal and zinc at 400 ppm (customized by Reedy Fork Farm, NC) to predispose the birds to necrotic enteritis [45, 46]. Birds were challenged with virulent C. perfringens strain CP4 [47] in feed for 5 days as described [46] with some modifications. Feed was withdrawn on the day prior to challenge for 15 h. The following day, chickens were inoculated with 1.0 mL of an overnight culture of C. perfringens CP4 grown in cooked meat medium (CMM; Difco) by oral gavage. Immediately after gavage, infected feed was provided thereafter for 5 consecutive days. To prepare infected feed, C. perfringens was grown in CMM medium for 24 h at 37°C, which then was inoculated into fluid thioglycollate medium (FTG; Difco) at a ratio of 0.3% (v/v) and incubated at 37°C for 15 h (morning challenge) or 23 h (evening challenge). The C. perfringens culture was mixed with feed at a ratio of 1:1 (v/w). Infected feed was prepared freshly twice daily. All birds were euthanized and necropsied the day following the final challenge.
Protection against *C. perfringens* challenge was assessed on the basis of gross intestinal lesion scores at necropsy. On day 33, chickens were euthanized with CO₂ and their small intestines (defined here as the section between the gizzard and Meckel’s diverticulum) were examined for visible gross lesions. Intestinal lesions were scored as follows: 0 = no gross lesions; 1 = thin or friable wall or diffuse superficial but removable fibrin; 2 = focal necrosis or ulceration, or non-removable fibrin deposit, 1 to 5 foci; 3 = focal necrosis or ulceration, or non-removable fibrin deposit, 6 to 15 foci; 4 = focal necrosis or ulceration, or non-removable fibrin deposit, 16 or more foci; 5 = patches of necrosis 2 to 3 cm long; 6 = diffuse necrosis typical of field cases [46]. Birds that died during challenge were assigned a score of 6.

2.12 Statistical analysis

All statistics were carried out using GraphPad Prism 6.0 (Graph-Pad Software, San Diego, CA). Antibody titers and lesion scores were analyzed using one-way ANOVA followed by Tukey’s posttest. The values were expressed as means ± SEM, and differences were considered significant at $P < 0.05$.

RESULTS

3.1 Construction of a PlcC-NetB fusion protein.

We designed a fusion protein comprising an N-terminal 6His tag, PlcC, linker GGSGGSGGPGSGGSGG, and NetB (Fig. 1). The long, flexible linker was used to facilitate correct folding of the PlcC and NetB domains, and to permit unencumbered access of B-cell receptors to the important immunogenic regions. PlcC contains the C-terminal receptor-binding...
domain of the alpha toxin [18]. The NetB sequence contains a mutation W262A in the rim domain [34], which substantially attenuates the toxicity of the protein in cell culture assays. We noted that PlcC and NetB contain a total of 5 eukaryotic Asn-linked glycosylation sites (N-X-S/T), which could result in the addition of glycans to the Asn residue when the protein is targeted to the ER in plant cells. We constructed expression vectors to target proteins either to the ER (pBYR2eK2M-6HplcCnetB, -6HplcC, and -6HnetB) or to the cytosol (pBYR2e3K2Mc-c6HplcCnetB -c6HplcC, and -c6HnetB). We expected the cytosol targeted proteins to be unglycosylated, while the ER-targeted proteins may be glycosylated at any combination of or all of the 5 consensus N-X-S/T sites.

3.2 Production of PlcC-NetB and PlcC-NetB-NG fusion proteins in plants.

Using the methods outlined herein two different fusion proteins were produced. One, targeted to the endoplasmic reticulum (ER), was glycosylated (PlcC-NetB). The other, targeted to the cytoplasm, was non-glycosylated (PlcC-NetB-NG). Both fusion proteins were probed with rabbit anti-PlcC and rabbit anti-NetB antibodies which yielded the same results on a western blot (Fig. 2). PlcC-NetB also has incrementally increased band sizes above the initial band which aligns with the singular PlcC-NetB-NG band at approximately 49 kDa, the expected relative size of the fusion proteins (Fig 2). Both NetB and PlcC expressed individually were also probed and appear at the expected sizes of 34 kDa and 15 kDa, respectively. While ER and cytoplasmic targeted NetB and PlcC were produced only representative results are shown in Figure 2.

The process of purification for the PlcC-NetB protein is outlined in Figure 3. A Coomassie-stained gel (Fig. 3A) shows a wild-type, uninfiltrated (WT) control followed by crude extract which had already been acid precipitated. Note that the crude extract has highly
reduced plant protein contaminants, due to the acid precipitation that caused contaminating proteins to become insoluble while the fusion proteins remain soluble. The flow-through fraction contained bands of a similar profile to the crude extract on the Coomassie, while both washes showed no detectable protein. Three elution fractions contained bands at approximately 49 kDa. Anti-NetB antibodies (Fig 3B) reacted primarily with the standard and elution 2, while elutions 1 and 3 had visible but less prominent bands. When probed with anti-PlcC antibodies (Fig 3C), similar bands were present in every lane except the blank and NetB standard lanes. Elution fraction bands were as strong as those seen in the crude extract lane. Flow-through and wash fractions had lighter but detectable bands when probed with anti-PlcC. Bands that reacted with anti-PlcC antibodies were also seen at approximately 49 kDa.

3.3 Serum antibody responses to fusion protein antigens.

We evaluated the serum antibody responses in birds immunized with PlcC-NetB and in control birds against the glycosylated fusion protein and against non-glycosylated recombinant rPlcC and rNetB produced in bacteria. In experiments 1 and 2, the anti-PlcC-NetB titers were significantly higher than controls (Fig. 4). The titers against rPlcC and rNetB were also significantly elevated over controls. In experiment 3, where birds were immunized twice, only the titers against the fusion protein were significantly higher than controls, although the serum titers against rNetB were elevated.

3.4 Cellular responses against PlcC-NetB in Experiment 3.

Immunization with the fusion protein led to variable cellular responses. The rPlcC-NetB fusion protein elicited weak, but significant, cellular responses against rNetB and rPlcC-NetB in
spleenocytes (Fig. 5). No antigen-specific stimulation was detected in lymphocytes (data not shown).

3.5 Protection against C. perfringens challenge.

Protection against challenge was assessed by scoring intestinal lesions using a 6 point scoring system [46]. Immunization with the rPlcC-NetB fusion protein resulted in a significant reduction in lesion scores in all three experiments (Fig. 5). Note that in Experiment 3, birds were challenged a week earlier than in experiments 1 and 2. The younger age of the birds may have contributed to the higher lesion scores for some birds. Two of the non-immunized birds died during challenge. This was the only experiment in which any birds died following challenge.

DISCUSSION

There is a growing need for methods to control NE that do not rely on antibiotics. Vaccination is a proven approach for controlling many poultry diseases and immunization with toxoid antigens is known to provide some protection against necrotic enteritis [37, 48]. Immunizing with both α-toxoid and NetB toxoid provides better protection than when either antigen is used alone [49]. Based on these observations, we designed a novel fusion protein combining toxoids of both toxins. The use of a plant expression system to produce this antigen has many advantages, including high yields, ease of stockpiling, rapid and easy scaling, quick response to emerging pathogens, and lack of any animal components in the final vaccine product.

The production of a PlcC-NetB fusion was readily accomplished in N. benthamiana using a geminiviral replicon transient expression system. A previously described Salmonella expression system was used to attempt production a fusion protein between PlcC and GST-NetB.
However, production levels were low and the protein was rapidly degraded within the *Salmonella* cells [20]. In contrast, the plant expression system was shown to produce the PlcC-NetB fusion protein at a high capacity without degradation. High levels of protein production were due to our use of the geminiviral replicon transient expression system which has been recently optimized and has undergone improvement since the production of the PlcC-NetB fusion protein [41, 42, 50]. Acid precipitation (a process by which the PI of proteins, and their repulsive electromagnetic forces, equal zero and results in protein aggregation) proved to be an effective clarification step, over other purification methods, as both the PlcC-NetB and PlcC-NetB-NG fusion proteins proved to be acid stable [51]. This allowed for easy purification using metal affinity chromatography because many of the plant protein contaminants were removed from solution while the fusion proteins remained soluble. Metal affinity chromatography resulted in substantial purification although contaminants could be seen upon concentration of elutions (data not shown). However, contaminants were deemed not to be an issue for vaccination as they were in such low quantities.

Since a plant expression system was used, native plant glycosylation of the fusion protein PlcC-NetB occurred. An illustration of this can be seen in both Figure 2 and Figure 3 as there are progressively higher bands above the primary fusion protein band at 49 kDa only in the ER targeted variant. Glycosylation was a concern as an immune response could potentially be directed towards the glycans rather than the fusion protein itself [52, 53]. This is the reason the PlcC-NetB-NG fusion was pursued. PlcC-NetB-NG proved to be troublesome as it precipitated after dialysis, a phenomenon that has been observed for rubisco, the most abundant plant protein contaminant. Insolubility of PlcC-NetB-NG may also be caused by stripping of metal ions from both PlcC and NetB that are normally present. According to the crystal structure of PlcC it likely
binds calcium ions, however, due to difficulty in forming a crystal structure calcium was substituted for cadmium (15). The crystal structure of native NetB toxin has been shown to bind magnesium (35). These metal ions could be removed during the final dialysis step, which might affect protein structure and solubility. This may indicate that the glycoforms play a role in protein stability and solubility as PlcC-NetB did not have solubility issues under the same buffer, concentration, and purification conditions. Results indicate that, even with plant native glycosylation, the PlcC-NetB fusion promoted an anti-toxoid response. We speculate that the glycoforms could have possibly had an adjuvant effect as antibody titers for the fusion protein were greater than *E. coli* produced PlcC and NetB delivered simultaneously [54]. This may also be due to the fusion of both proteins as well. In any case, production of the PlcC-NetB fusion protein was found to be efficient on a small scale which, due to the nature of plant expression systems, is readily scalable. Moreover, the fusion protein vaccine was found to be effective when injected into broiler birds as is discussed below.

In Experiments 1 and 2, three weekly injections with the fusion protein generated strong serum IgY responses against both toxin components ([Fig. 4](#fig4)). Vaccinated birds had geometric mean titers of around 2,000 against recombinant NetB and PlcC proteins purified from *E. coli* ([Fig. 4](#fig4)). Titers against the PlcC-NetB fusion protein produced in plants were around 8,000 in immunized birds. The difference in titers between single proteins and the fusion protein is speculated to be due to enhanced immune response to the fused protein, since PlcC and NetB are in close proximity during infection, or responses against the carbohydrate moieties on the glycosylated protein leading to an enhanced immune response.

In Experiment 3 birds were immunized at 7 and 12 days of age which lead to a reduced serum response. The chicken adaptive immune system does not fully mature until approximately
two weeks post-hatch, although humoral responses can be detected after immunization with a protein antigen at 12 days of age [55]. In this regard, an adjuvanted NetB toxoid was shown to be ineffective when injected into day-of-hatch chicks [56]. Thus, it is possible that immunizing the birds at 7 days of age did not serve as a priming dose, such that the immunization at 12 days of age was effectively the prime. Previous work showed that immunization at 12 days of age, but not 7 days of age, can be boosted by injection at 29 days of age [55]. Although we did not perform a vaccine boost, we infer that the birds were adequately primed since, despite the low serum antibody responses, they were protected against challenge (Fig. 5C).

CONCLUSIONS

We have demonstrated that this novel toxoid antigen produced in plants is immunogenic and protective in chickens. However, we acknowledge that a more practical immunization approach is needed in order to have a positive impact on the broiler industry. We envision several options. Immunization of parent stock is likely to induce maternal antibodies to protect chicks during the first few weeks of life. This approach has been demonstrated to be effective with an α-toxoid [29]. Alternatively, protein antigens can be successfully delivered via in ovo injection with an adequate adjuvant. Delivery of the fusion protein via cornmeal could also be a cheap and effective mass delivery method for vaccine produced in corn seeds. The effectiveness of these approaches will be assessed in future studies.
ACKNOWLEDGEMENTS

This project was supported by Agriculture and Food Research Initiative Competitive Grant no. 2016-67016-24947 from the USDA National Institute of Food and Agriculture and startup funds from ASU to Ken Roland, and by support from the Biodesign Institute at ASU for Hugh Mason. The funding agencies played no role in study design, data collection and analysis or preparation of the manuscript. The authors wish to thank Luis Armando Vazquez, Donovan Leigh, Penelope Roach, Melody Yeh, Jack Crawford, Matt Contursi, Mary Pardhe, Sara Aman, and Artemio Chaves for their technical assistance. We thank Dustin McAndrew, Randall Dalbey, Jacquelyn Kilbourne, Larisa Gilley and the rest of the ASU animal care staff for expert care of our research animals and procedural assistance.

FIGURE LEGENDS

Figure 1. Schematic of the his tagged PlcC-NetB fusion protein. Potential Asn-glycosylation sites are indicated by a short line (-). The site of the W262A mutation is indicated.

Figure 2. Western blot comparing different fusion constructs. (A) Blot probed with rabbit anti-NetB. (B) Blot used in (A) after stripping, probed with rabbit anti-PlcC. 6H-PlcC-NetB, ER targeted fusion protein expressed in N. benthamiana leaf; PlcC-NetB-NG, cytosolically targeted fusion protein expressed in N. benthamiana leaf; NetB-NG, cytosolically targeted NetB expressed in N. benthamiana leaf; GST-NetB, bacterially expressed standard protein; PlcC-NG, cytosolically targeted PlcC expressed in N. benthamiana leaf.
Figure 3. Purification of glycosylated PlcC-NetB fusion. (A) Coomassie-stained gel showing all purification steps. (B) Western blot of all purification steps probed with rabbit anti-NetB. (C) Western blot of all purification steps probed with rabbit anti-PlcC. Note: NetB standard is a GST-NetB fusion with a mass of 59.9 kDa [20].

Figure 4. IgY serum titers from immunized and non-immunized birds, assayed against NetB, PlcC, or NetB-PlcC fusion. All P values are compared to adjuvant only controls. Fusion refers to his-tagged, glycosylated PlcC-NetB. (A) Experiment 1, * P ≤ 0.009 (B) Experiment 2, ** P ≤ 0.0025. (C) Experiment 3, & P < 0.0002.

Figure 5. Splenocyte responses to PlcC-NetB, PlcC and NetB after immunization with PlcC-NetB in Figure 4, Experiment 3.

Figure 6. Intestinal lesions scores in (A) Experiment 1, (B) Experiment 2 and (C) Experiment 3. Each dot represents the score of an individual bird. Differences between immunized groups and controls are indicated. * P < 0.001; ** P = 0.004; &P = 0.0006; †P < 0.0001.

Table S1. Primers used in this study*

<table>
<thead>
<tr>
<th>Primer</th>
<th>DNA sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>PlcC-Sac-R</td>
<td>CCCGAGCTCCTATTTGATGGTTGAAGTGGAGTTTCC</td>
</tr>
<tr>
<td>6H-Xba-F</td>
<td>GACTCTAGAACAGTGGCTACCACATGCCATCATCAC</td>
</tr>
<tr>
<td>Plc_pqe_f1</td>
<td>ACATGGATCCGACCACCGTCCGATGGGCAACAC</td>
</tr>
<tr>
<td>Plc_pqe_r2</td>
<td>ACATGGATCCGACCACCGTCCGATGGGCAACAC</td>
</tr>
<tr>
<td>netB_pqe_f3</td>
<td>GGGATCCAGCGAAGCTGAACGAC</td>
</tr>
<tr>
<td>netB_Pqe_r4</td>
<td>GTCGACCTTTTACAGATAATATTC</td>
</tr>
</tbody>
</table>

*Underlined sequences indicate engineered restriction sites used for cloning.
REFERENCES


Figure 1

Schematic of the his tagged PlcC-NetB fusion protein

Potential Asn-glycosylation sites are indicated by a short line (-). The site of the W262A mutation is indicated.

**Figure 1.** Schematic of the His-tagged PlcC-NetB Fusion Protein. Potential Asn-glycosylation sites are indicated by a short line (●). The site of the W262A mutation is indicated.
Figure 2

Western blot comparing different fusion constructs

(A) Blot probed with rabbit anti-NetB. (B) Blot used in (A) after stripping, probed with rabbit anti-PlcC. 6H-PlcC-NetB, ER targeted fusion protein expressed in \textit{N. benthamiana} leaf; PlcC-NetB-NG, cytosolically targeted fusion protein expressed in \textit{N. benthamiana} leaf; NetB-NG, cytosolically targeted NetB expressed in \textit{N. benthamiana} leaf; GST-NetB, bacterially expressed standard protein; PlcC-NG, cytosolically targeted PlcC expressed in \textit{N. benthamiana} leaf.
Figure 3

Purification of glycosylated PlcC-NetB fusion

(A) Coomassie-stained gel showing all purification steps. (B) Western blot of all purification steps probed with rabbit anti-NetB. (C) Western blot of all purification steps probed with rabbit anti-PlcC. Note: NetB standard is a GST-NetB fusion with a mass of 59.9 kDa [20].

Figure 3. Purification of Glycosylated 6H-PlcC-NetB fusion. (A) Coomassie stain of all purification steps. (B) Western blot of all purification steps, rabbit anti-NetB probe. (C) Western blot of all purification steps, rabbit anti-PlcC probe. Note: NetB standard is a GST-NetB fusion.
Figure 4

IgY serum titers from immunized and non-immunized birds, assayed against NetB, PlcC, or NetB-PlcC fusion

All $P$ values are compared to adjuvant only controls. Fusion refers to his-tagged, glycosylated PlcC-NetB. (A) Experiment 1, * $P \leq 0.009$ (B) Experiment 2, ** $P \leq 0.0025$. (C) Experiment 3, $^\& P < 0.0002$.

---

Figure 4. IgY serum titers from immunized and non-immunized birds, assayed against NetB, PlcC, or NetB-PlcC fusion. All $P$ values are compared to adjuvant only controls. Fusion refers to his-tagged, glycosylated PlcC-NetB. (A) Experiment 1, * $P \leq 0.009$ (B) Experiment 2, ** $P \leq 0.0025$. (C) Experiment 3, & $P < 0.0002$. 

Coating Antigens

---

Log$_2$ Titer Serum IgY

NetB, PlcC, Fusion
Figure 5

Splenocyte responses

Splenocyte responses to PlcC-NetB, PlcC and NetB after immunization with PlcC-NetB in Figure 4, Experiment 3.
Figure 5. Splenocyte responses to PlcC-NetB, PlcC and NetB after immunization with PlcC-NetB in Figure 4, Experiment 3.
Figure 6

Intestinal lesions scores

Intestinal lesions scores in (A) Experiment 1, (B) Experiment 2 and (C) Experiment 3. Each dot represents the score of an individual bird. Differences between immunized groups and controls are indicated. * $P < 0.001$; ** $P = 0.004$; & $P = 0.0006$; †$P < 0.0001$

A. Expt 1

B. Expt 2

C. Expt 3

Figure 6. Intestinal lesions scores in Experiment 1 (A) Experiment 2 (B) and Experiment 3 (C). Each dot represents the score of an individual bird. Differences between immunized groups and controls are indicated. * $P < 0.001$; ** $P = 0.004$; & $P = 0.0006$; †$P < 0.0001$