

Biological soil crusts inhibit seed germination in a temperate pine barren ecosystem Jessica A. Gilbert^{1,2} and Jeffrey D. Corbin^{1,3} ¹Department of Biological Sciences, Union College, Schenectady, NY 12308 ²Orchid ID: 0000-0002-5506-666X ³Corresponding Author: <u>corbinj@union.edu</u>; 518-388-6097; Orchid ID: 0000-0002-3377-5916 **Acknowledgements** This research was made possible by funding from Union College in support of JAG. The Albany Pine Bush Preserve allowed us to collect crust samples and generously provided seeds for our experiment. We highlight the Preserve staff's dedicated stewardship on behalf of this unique ecosystem. Rachel Thiet made helpful suggestions on a previous version of this paper and also identified lichens found at APB. Lorinda Leonardi identified bryophytes. Jacqueline Sharry and Samantha Williams assisted with field collection.



Λ	hst	tra	ct
А	115	111	

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

Biological soil crusts (BSCs) are known to affect plants' germination and seedling establishment in arid ecosystems, but their ecological role in more mesic climates is not so well-known. We tested the effects of moss-crusted versus uncrusted soils on seed germination dynamics in a temperate pine barren ecosystem. We conducted a 35-day laboratory assay of seed germination on moss-crusted soils versus uncrusted soils from the Albany (NY) Pine Bush Preserve. We tested total seed germination and the number of days to 50% of total germination (T50) of two herbaceous perennial forb species in each soil type. Three and five times more seeds germinated on uncrusted soil than on crusted soil for bush clover (Lespedeza capitata) and wild lupine (Lupinus perennis), respectively. Seeds of both species also germinated approximately 10 days earlier on uncrusted soil than on crusted soil. This study, and others in similar habitats, show that BSCs in mesic climates can influence germination and other early life-history stages of plants. We hope that further study of the interactions between BSCs and vascular plants in mesic climates will contribute to our understanding of the ecology of BSCs outside the arid and semiarid climates where they are more extensively studied.

39

40

Keywords: Albany Pine Bush Preserve; Crypotogamic crust; Bryophytes; Inland sand plain;

41 Lespedeza capitata ; Lupinus perennis

42



Introduction

A variety of biotic and abiotic factors can strongly influence the rates of seed germination and other early plant life stages that, in turn, influence plant population structure and community composition (Crawley 1990; Eriksson and Ehrlén 1992; Harper 1977). Understanding such factors, and how they vary in space and time, are critical if we are to understand ecosystem processes and resilience to disturbances (Hoffmann 1999; Platt et al. 1988; Prevéy and Seastedt 2015).

The combined effects of abiotic and biotic influences on plant establishment have been demonstrated in habitats supporting biological soil crusts (BSCs) worldwide. BSCs are aggregations of cyanobacteria, algae, lichens, and bryophytes that form thin layers on soil surfaces (Belnap 2003; Belnap and Lange 2001; Weber et al. 2016). They can play an integral role in soil stability (Belnap and Gillette 1997; Jimenez Aguilar et al. 2009), soil moisture retention (Bu et al. 2015; Chamizo et al. 2016) and the nitrogen cycle (Belnap 2002; Castillo-Monroy et al. 2010). They can also influence the vascular plant community through their impacts on seed germination and seedling survival (Zhang et al. 2016). The direction and magnitude of BSCs' impacts on plants, however, has been shown to vary widely by plant species and crust community composition. For this reason, biocrust-plant interactions should be tested empirically rather than inferred from other studies.

BSCs have been most-often studied in arid to semi-arid ecosystems, but they can also occur in more mesic climates including the temperate and subtropical US (e.g. Hawkes and Flechtner 2002; Neher et al. 2003; Sedia and Ehrenfeld 2003; Smith et al. 2004; Thiet et al. 2014). BSCs in these habitats can be found in open barrens, pine barrens, pavement barrens, sand plains, and dunes, and include a diverse assemblage of algae, cyanobacteria,



lichens, and bryophytes (Corbin and Thiet, *unpublished data*). Many of these habitats are recognized as systems of high conservation value for their unique ecology and the presence of rare and threatened plants and animals (e.g. Edinger 2014; Gawler and Cutko 2010; Motzkin and Foster 2002; Sperduto and Kimball 2011).

To date, only a few studies have examined how BSCs in these more mesic climates influence plant dynamics, and, as has been observed for BSC-plant dynamics in general, the results are system- and species-specific. BSCs in New Jersey's Pine Barrens inhibited germination of three perennial species (Sedia and Ehrenfeld 2003), but in a Florida sand scrub, BSCs promoted germination of three short-lived perennial herbaceous species and an annual/occasional perennial (Hawkes 2004). Meanwhile, BSCs in a German sand ecosystem inhibited perennial species' germination but had either a positive effect or no effect on annual species' germination (Langhans et al. 2009). Thiet et al. (2014) found that algal and lichen-moss crusts from a coastal sand dune increased seedling survivorship and growth of a perennial grass and a woody shrub; but moss-only crusts decreased seedling performance. The range of results demonstrates that a general relationship between crusts and plant demography in such habitats is not possible.

This study contributes to the growing but still small body of research investigating the role of BSCs on early-stage plant population dynamics in mesic climates. We assessed seed germination on moss-crusted versus uncrusted (sand) substrate in an inland pine barren ecosystem in New York, USA. This is the first known examination of BSCs in such a habitat, and adds needed understanding to the ecology of BSCs in temperate ecosystems.



Materials and Methods

Study Area

The Albany Pine Bush (APB) Preserve (42° 42′ N; 73° 52′ W; Figure 1) is an inland pine barren that is formed of deep, sandy, glacial outwash soils (Barnes 2003). Though its cold temperate climate supports nearby eastern deciduous forest, vegetation cover at the APB is relatively sparse due to the edaphically xeric conditions and frequent fires. Native vegetation is dominated by scrub oak (*Quercus ilicifolia*), blueberry (*Vaccinium* spp.), and scattered pitch pine (*Pinus rigida*). Areas of APB were invaded by the non-native black locust tree (*Robinia pseudoacacia*) in the latter 20th Century and subsequently restored with mechanical removal of locust stems and roots (Rice et al. 2004). Both native pine-oak scrub and areas from which black locust trees have been removed are managed using prescribed fire (Bried et al. 2015).

Biological soil crusts are abundant in areas of APB where fires or other recent disturbances maintain an open canopy. Vascular plants, BSCs, and uncrusted sand at APB and other pine barren ecosystems form a spatial mosaic at the scale of 0.5-10 m² per patch (Sedia and Ehrenfeld 2003; personal observation). Crust composition ranges from incipient algal mats to moss-dominated crusts that also contain lichens and cyanobacteria (Corbin and Thiet, *unpublished data*). Common species include the mosses *Ceratodon purpureus* and *Polytrichum piliferum* and the lichens *Cladonia cristatella* and *C. gracilis* ssp. *turbinata*, and *C. ochrochlora*.



Sample collection

In June 2017, we collected the substrate for our germination trials from a single site that had been cleared of the black locust tree in 2008 and subsequently replanted with a native species mix including little bluestem (*Schizachyrium scoparium* (Michx.) Nash), wild perennial lupine (*Lupinus perennis* L.) and round-headed bush clover (*Lespedeza capitata* Michx.). We collected mature moss-dominated BSCs by inserting 40, 3.5-cm diameter plastic Petri dishes into the soil to a depth of 2 cm. We also collected 40 dishes of bare sand, hereafter referred to as "uncrusted" samples, from the same site and within meters of our crusted samples. Collections were approved by the Albany Pine Bush Preserve

Commission. Crusted samples were dominated by the mosses *C. purpureus* and *P. commune*. Lichens made up a minor component, though in several dishes they increased in cover during experimental watering. Though the uncrusted samples were free of visible mosses and lichens and had a loose-sand texture, some turned green during the experiment, indicating that algae and perhaps cyanobacteria were present.

All dishes were stored in a refrigerator (4°C) for three weeks until the experiment began.

Germination Procedure

We tested germination rates of three herbaceous species – lupine, bush clover, and little bluestem. These three species are abundant in both mature pine barren habitat and also in the post-black locust removal sites such as where we collected our soil samples. Lupine and bush clover seeds were collected from wild populations at APB in 2016. Little bluestem was grown by Ernst Conservation Seeds, Inc. (Meadville, PA) using traditional



agricultural techniques and harvested in 2016. The source for the Pennsylvania-grown little bluestem seeds was a 2014 collection from APB. All seeds were stored in a freezer (-20 °C) between cleaning and use.

Our experiment was a full factorial experiment with two soil types – moss-crusted and uncrusted – and four plant species types – lupine, bush clover, little bluestem, and no seeds. There were 10 replicates of each soil type x plant species treatment combination. Each dish except the no seeds treatments received 20 seeds of a single species scattered onto the surface; seeds were not buried so as to best mimic natural wind-dispersal. The no seed treatments on each soil type were established as a control for seeds emerging from a potential buried seed bank; no seeds were added to this latter treatment.

Dishes were laid out randomly on a light table under 12-hour white light beginning July 2, 2017, and were rotated once a week to eliminate discrepancies in light conditions. All dishes were watered with 5 ml of water every day, which was the quantity that evenly moistened the soil or crust surface through the base of the dish. Germinating seeds, judged by the emergence of the radicle, were recorded every 1-2 days for 35 days. Once a seed germinated, it was removed. We recorded the total number of seeds that germinated in each dish and the number of days it took for each dish to reach 50% of total germination (T50). T50 was not calculated for any experimental dishes in which total germination was zero.

Data Analysis

We used two-way ANOVA (R version 3.4.1) to analyze the effect of soil type (moss-crusted versus uncrusted), plant species, and the interaction between soil type and plant



species on total germination and T50 (α = 0.05). Little bluestem germination was extremely low in both soil conditions, and thus was omitted from data analysis. Otherwise, the data were found to meet assumptions of normality and homogeneity of variance.

Results

There was a strong effect of soil type on the number of bush clover and lupine seeds that germinated, as three and five times more bush clover and lupine, respectively, germinated on uncrusted soil than on moss-crusted soil (Table 1; Figure 2A). Seeds on uncrusted soil reached T50 approximately 10 days earlier than seeds on crusted soils. Bush clover seeds' T50 was approximately 5 days earlier than lupine seeds' T50 on both crusted and uncrusted soils (Table 1; Figures 2B and 3). There was no interaction between soil type and plant species on T50 (Table 1). By the end of the experiment, total germination did not differ between the two species, nor was there a significant interaction between soil type and plant species on total germination (Table 1; Figures 2 and 3).

No seeds germinated in either our crusted or uncrusted control ("no seeds") dishes that would indicate the presence of a stable seed bank.

Discussion

Moss-covered BSCs at the Albany Pine Bush clearly inhibited seed germination in our experiment. Fewer seeds germinated, and they took longer to do so, on moss crusts than they did on uncrusted sand. Though some germination assays have found that BSCs can enhance rates of germination (Godínez-Alvarez et al. 2012; Hawkes 2004), our findings are consistent with other studies in mesic and arid climates in which BSCs inhibited



germination (Deines et al. 2007; Jeschke and Kiehl 2008; Langhans et al. 2009; Serpe et al. 2008; Thiet et al. 2014; Zaady et al. 1997).

Though we were only able to test the effects of BSCs on two species, both of which were herbaceous perennial forbs, they both responded in similar ways. We saw no interaction between plant species and the type of soil for either total germination nor the time it took for germination to reach 50% of total (T50). The lone grass we tested, little bluestem, did not germinate in any conditions. A number of other studies have reported mixed effects of crusts on germination: crusts had negative effects on germination of some species and no effect or positive effects on others (e.g. Godínez-Alvarez et al. 2012; Hawkes 2004; Langhans et al. 2009; Zaady et al. 1997; Zhang and Belnap 2015). However, until more species with a wider range of life history traits are assayed for their response to crusts in our system, we have no evidence that there are species-specific effects of BSCs on seed germination.

BSCs' negative effects on seed germination have been explained, in part, by the physical barrier that they can create between seeds and soil (Jeschke and Kiehl 2008; Serpe et al. 2008; Song et al. 2017; Zhang and Belnap 2015). The mosses in our BSC treatments quickly absorbed the added water and thus seeds were more likely to dry out. Many of the seeds that did germinate on BSC treatments did so in cracks or gaps in the moss canopy, as has been observed in other studies (e.g. Langhans et al. 2009). By contrast, bare soil in the uncrusted treatments provided seeds with a larger area of soil contact and potentially more moisture as described by Song et al. (2017).

Our study did not consider the impact of BSCs on subsequent plant life stages, yet a variety of other studies have shown distinct effects on seedling growth and survival as well.



Mosses and other crust components can limit root penetration and therefore plants' access to moisture and nutrients (Deines et al. 2007; Serpe et al. 2008). This can be a major source of mortality for seedlings that germinate on crust surfaces (Jeschke and Kiehl 2008). However, if seedling roots can reach through the crust to the soil, they may find enhanced growing conditions. BSCs have been shown to positively affect seedling performance, perhaps through the higher nutrient and organic content of the soil under BSCs (Godínez-Alvarez et al. 2012; Langhans et al. 2009; Thiet et al. 2014).

BSCs' influence on the distribution and abundance of plant species may also have broader effects on the APB ecosystem. For example, lupines host the larvae of the federally endangered Karner blue butterfly (*Lycaeides melissa samuelis*) (Barnes 2003).

Furthermore, both lupines and bush clover form symbioses with nitrogen-fixing bacteria, and N inputs from such plants are known to influence the nitrogen status of other components of the ecosystem (Rascher et al. 2012). Finally, BSCs in temperate (Sedia and Ehrenfeld 2003; Thiet et al. 2005) and dryland (Belnap 2002) systems are known to, themselves, affect soil C, N, micronutrients, and moisture. BSCs' role in northeastern barren and dune ecosystems is worthy of further study.

Our study is evidence that intact BSC communities can influence the plant community of pine barren ecosystems by affecting the fate of seeds. This mechanism likely contributes to the mosaic of BSC patches interwoven with vegetated patches and is likely a significant source of habitat heterogeneity. Sedia and Ehrenfeld (2003) hypothesized that BSCs and vascular plants form alternative stable states in pine barren ecosystems, potentially mediated by seasonal weather conditions. Until recently, there were relatively few efforts to further explore the interactions between BSCs and vascular plants in mesic



climates, and how those int	eractions vary over space in time (but see Büdel et al. 2014;
Langhans et al. 2009; Thiet	et al. 2014). We hope that further awareness of crusts outside
of the arid and semiarid clin	mates where they are more extensively studied will encourage
more exploration.	



234	References
235	Barnes JK (2003) Natural history of the Albany Pine Bush. New York State Museum, Albany,
236	USA
237	Belnap J (2002) Nitrogen fixation in biological soil crusts from southeast Utah, USA. Biology
238	and Fertility of Soils 35:128-135. doi:10.1007/s00374-002-0452-x
239	Belnap J (2003) The world at your feet: desert biological soil crusts. Frontiers in Ecology
240	and the Environment 1:181-189. doi:doi:10.1890/1540-
241	9295(2003)001[0181:TWAYFD]2.0.CO;2
242	Belnap J, Gillette DA (1997) Disturbance of biological soil crusts: impacts on potential wind
243	erodibility of sandy desert soils in southeastern Utah. Land Degradation &
244	Development 8:355-362. doi:doi:10.1002/(SICI)1099-145X(199712)8:4<355::AID-
245	LDR266>3.0.CO;2-H
246	Belnap J, Lange OL (2001) Biological soil crusts: Structure, function, and management.
247	Ecological Studies 150. Springer, Berlin
248	Bried JT, Gifford NA, Robertson KM (2015) Predicted Crown Fire Risk Adds Incentive to
249	Restore Open-Canopy Pine Barrens at the Wildland-Urban Interface. Journal of
250	sustainable forestry 34:147-167
251	Bu C, Wu S, Han F, Yang Y, Meng J (2015) The Combined Effects of Moss-Dominated
252	Biocrusts and Vegetation on Erosion and Soil Moisture and Implications for
253	Disturbance on the Loess Plateau, China. PLOS ONE 10:e0127394.
254	doi:10.1371/journal.pone.0127394



255	Büdel B et al. (2014) Improved appreciation of the functioning and importance of biological
256	soil crusts in Europe: the Soil Crust International Project (SCIN). Biodiversity and
257	Conservation 23:1639-1658. doi:10.1007/s10531-014-0645-2
258	Castillo-Monroy AP, Maestre FT, Delgado-Baquerizo M, Gallardo A (2010) Biological soil
259	crusts modulate nitrogen availability in semi-arid ecosystems: insights from a
260	Mediterranean grassland. Plant and Soil 333:21-34. doi:10.1007/s11104-009-0276-
261	7
262	Chamizo S, Cantón Y, Rodríguez-Caballero E, Domingo F (2016) Biocrusts positively affect
263	the soil water balance in semiarid ecosystems. Ecohydrology 9:1208-1221.
264	doi:doi:10.1002/eco.1719
265	Crawley MJ (1990) The population dynamics of plants. Phil Trans R Soc Lond B 330:125-
266	140
267	Deines L, Rosentreter R, Eldridge DJ, Serpe MD (2007) Germination and seedling
268	establishment of two annual grasses on lichen-dominated biological soil crusts.
269	Plant and Soil 295:23-35. doi:10.1007/s11104-007-9256-y
270	Edinger GJ, D. J. Evans, S. Gebauer, T. G. Howard, D. M. Hunt, and A. M. Olivero (2014)
271	Ecological Communities of New York State. Second Edition. A revised and expanded
272	edition of Carol Reschke's Ecological Communities of New York State. Albany, NY
273	Eriksson O, Ehrlén J (1992) Seed and microsite limitation of recruitment in plant
274	populations. Oecologia 91:360-364
275	Gawler SC, Cutko A (2010) Natural landscapes of Maine: a guide to natural communities
276	and ecosystems. Maine Natural Areas Program, Department of Conservation,



277	Godínez-Alvarez H, Morín C, Rivera-Aguilar V (2012) Germination, survival and growth of
278	three vascular plants on biological soil crusts from a Mexican tropical desert. Plant
279	Biology 14:157-162. doi:doi:10.1111/j.1438-8677.2011.00495.x
280	Harper JL (1977) Population biology of plants. Academic Press, London
281	Hawkes CV (2004) Effects of biological soil crusts on seed germination of four endangered
282	herbs in a xeric Florida shrubland during drought. Plant Ecology 170:121-134.
283	doi:10.1023/b:Vege.0000019035.56245.91
284	Hawkes CV, Flechtner VR (2002) Biological soil crusts in a xeric Florida shrubland:
285	Composition, abundance, and spatial heterogeneity of crusts with different
286	disturbance histories. Microbial Ecology 43:1-12. doi:10.1007/s00248-001-1017-5
287	Hoffmann WA (1999) Fire and population dynamics of woody plants in a neotropical
288	savanna: Matrix model projections Ecology 80:1354-1369. doi:doi:10.1890/0012-
289	9658(1999)080[1354:FAPDOW]2.0.CO;2
290	Jeschke M, Kiehl K (2008) Effects of a dense moss layer on germination and establishment
291	of vascular plants in newly created calcareous grasslands. Flora - Morphology,
292	Distribution, Functional Ecology of Plants 203:557-566.
293	doi:https://doi.org/10.1016/j.flora.2007.09.006
294	Jimenez Aguilar A, Huber-Sannwald E, Belnap J, Smart DR, Arredondo Moreno JT (2009)
295	Biological soil crusts exhibit a dynamic response to seasonal rain and release from
296	grazing with implications for soil stability. Journal of Arid Environments 73:1158-
297	1169. doi: https://doi.org/10.1016/j.jaridenv.2009.05.009
298	Langhans TM, Storm C, Schwabe A (2009) Biological soil crusts and their
299	microenvironment: Impact on emergence, survival and establishment of seedlings.



300	Flora - Morphology, Distribution, Functional Ecology of Plants 204:157-168.
301	doi:https://doi.org/10.1016/j.flora.2008.01.001
302	Motzkin G, Foster DR (2002) Grasslands, heathlands and shrublands in coastal New
303	England: historical interpretations and approaches to conservation. Journal of
304	Biogeography 29:1569-1590. doi:doi:10.1046/j.1365-2699.2002.00769.x
305	Neher DA et al. (2003) Biological soil crust and vascular plant communities in a sand
306	savanna of northwestern Ohio. The Journal of the Torrey Botanical Society 130:244-
307	252. doi:10.2307/3557543
308	Platt WJ, Evans GW, Rathbun SL (1988) The Population Dynamics of a Long-Lived Conifer
309	(<i>Pinus palustris</i>). The American Naturalist 131:491-525. doi:10.1086/284803
310	Prevéy JS, Seastedt TR (2015) Effects of precipitation change and neighboring plants on
311	population dynamics of <i>Bromus tectorum</i> Oecologia 179:765-775.
312	doi:10.1007/s00442-015-3398-z
313	Rascher KG, Hellmann C, Máguas C, Werner C (2012) Community scale $^{15}\mathrm{N}$ isoscapes:
314	tracing the spatial impact of an exotic N_2 -fixing invader. Ecology Letters 15:484-491
315	Sedia EG, Ehrenfeld JG (2003) Lichens and mosses promote alternate stable plant
316	communities in the New Jersey Pinelands. Oikos 100:447-458.
317	doi:doi:10.1034/j.1600-0706.2003.12058.x
318	Serpe MD, Zimmerman SJ, Deines L, Rosentreter R (2008) Seed water status and root tip
319	characteristics of two annual grasses on lichen-dominated biological soil crusts.
320	Plant and Soil 303:191-205. doi:10.1007/s11104-007-9498-8



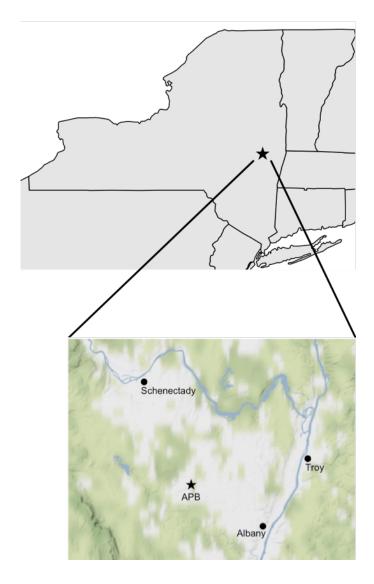
321	Smith SM, Abed RMM, Gercia-Pichel F (2004) Biological soil crusts of sand dunes in Cape
322	Cod National Seashore, Massachusetts, USA. Microbial Ecology 48:200-208.
323	doi:10.1007/s00248-004-0254-9
324	Song G, Li X, Hui R (2017) Effect of biological soil crusts on seed germination and growth of
325	an exotic and two native plant species in an arid ecosystem PLOS ONE 12:e0185839
326	doi:10.1371/journal.pone.0185839
327	Sperduto D, Kimball B (2011) The Nature of New Hampshire: Natural Communities of the
328	Granite State. University Press of New England, Hanover, NH
329	Thiet RK, Boerner REJ, Nagy M, Jardine R (2005) The effect of biological soil crusts on
330	throughput of rainwater and N into Lake Michigan sand dune soils. Plant and Soil
331	278:235-251. doi:10.1007/s11104-005-8550-9
332	Thiet RK, Doshas A, Smith SM (2014) Effects of biocrusts and lichen-moss mats on plant
333	productivity in a US sand dune ecosystem. Plant and Soil 377:235-244.
334	doi:10.1007/s11104-013-2002-8
335	Weber B, Büdel B, Belnap J (2016) Biological soil crusts: an organizing principle in
336	drylands. vol 226. Springer,
337	Zaady E, Gutterman Y, Boeken B (1997) The germination of mucilaginous seeds of <i>Plantago</i>
338	coronopus, Reboudia pinnata, and Carrichtera annua on cyanobacterial soil crust
339	from the Negev Desert. Plant and Soil 190:247-252. doi:10.1023/a:1004269031844
340	Zhang Y, Aradottir AL, Serpe M, Boeken B (2016) Interactions of biological soil crusts with
341	vascular plants. In: Biological soil crusts: an organizing principle in drylands.
342	Springer, pp 385-406



343	Zhang Y, Belnap J (2015) Growth responses of five desert plants as influenced by biological
344	soil crusts from a temperate desert, China. Ecological Research 30:1037-1045.
345	doi:10.1007/s11284-015-1305-z
346	

Table 1: Analysis of variance comparing the effects of soil type (crusted versus uncrusted) and plant species (bush clover versus lupine), and their interaction, on total germination and time to 50% of total germination (T50). Significant p-values are indicated by bold text.

	Total Germination				T50		
Variable	d.f.	F	p-value	-	d.f.	F	p-value
Soil type	1, 36	54.4	<0.0001	-	1, 30	42.2	<0.0001
Plant species	1, 36	0.2	0.6		1, 30	13.0	0.002
Soil type x Plant species	1, 36	2.4	0.13		1, 30	0.4	0.6



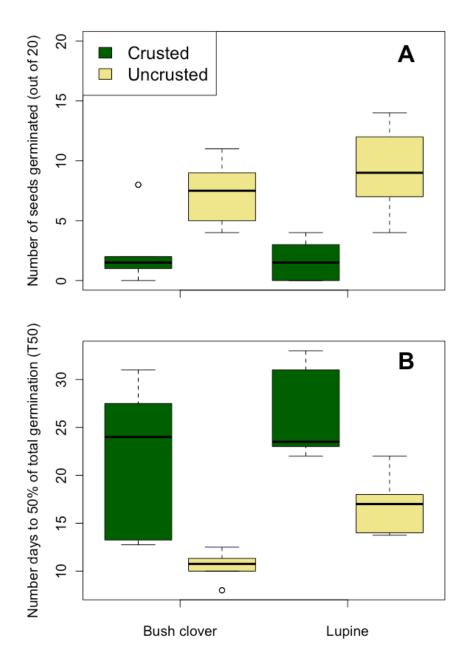
355

356

357

Figure 1 – Study area at Albany Pine Bush Preserve in New York State, indicated by the

358 stars.



361

362

Figure 2 – Boxplots of (A) total seed germination and (B) number of days to 50% of total germination (T50) for each soil type x plant species combination. The box in each box- and-whiskers plot represents the 25th and 75th percentiles, while the line in the middle of the



box is the median (50th percentile). The top and bottom whiskers extend to the most
extreme data points that are no more than 1.5 times the interquartile range from the box.
The ppen circle is an outliers beyond the whiskers. Total seed germination was lower, and
T50 was longer, on crusted soils compared to uncrusted soils. A similar number of seeds
germinated for each plant species, but bush clover seeds germinated significantly faster
than lupine seeds.

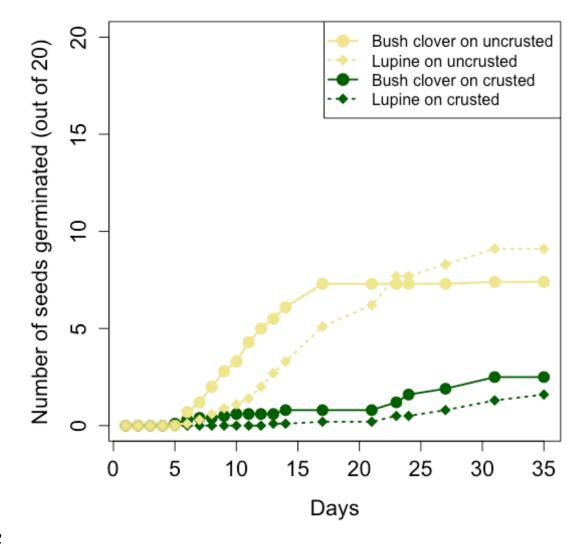


Figure 3 – Cumulative number of germinated seeds for bush clover and lupine on each soil type during the 35-day experiment.