

1 Resolving relationships in an exceedingly young orchid lineage using
2 Genotyping-by-sequencing data

3 Oscar Alejandro Pérez-Escobar ^{a,b,c*}; Diego Bogarín ^d; Rowan Schley ^{a,k}; Richard M. Bateman ^a;
4 Günter Gerlach ^e; Dörte Harpke ^f; Jonathan Brassac ^f; Mario Fernández-Mazuecos ^g; Steven
5 Dodsworth ^{h*}; Eric Hagsater ⁱ; Marc Gottschling ^j; Frank R. Blattner ^{f*}

6
7 ^a Royal Botanic Gardens Kew, Richmond TW9 3AE, UK

8 ^b Department of Biological and Environmental Sciences, University of Gothenburg, 405 30
9 Gothenburg, Sweden

10 ^c Gothenburg Global Biodiversity Centre, Gothenburg, Sweden

11 ^d Jardín Botánico Lankester, Universidad de Costa Rica, P.O. Box 302-7050, Cartago, Costa Rica

12 ^e Botanischer Garten München-Nymphenburg, Menzinger Str. 61, D-86 638 Munich, Germany

13 ^f Institute of Plant Genetics and Crop Plant Research (IPK), D-06 466 Gatersleben, Germany

14 ^g Department of Plant Sciences, University of Cambridge, Cambridge CB2 3AE, UK

15 ^h School of Life Sciences, University of Bedfordshire, Luton LU1 3JU, UK

16 ⁱ Herbarium AMO, Montañas Calizas 490, Lomas Barrilaco, Mexico

17 ^j Department of Biology, Systematic Botany and Mycology, GeoBio-Center, Ludwig-
18 Maximillians University, Menzinger Str. 67, D-80 638 Munich, Germany

19 ^k Department of Life Sciences, Imperial College London, Silwood Park, Ascot, Berkshire, SL5
20 7PY, UK

21
22 * Corresponding author.

23 E-mail address: o.perez-escobar@kew.org; steven.dodsworth@beds.ac.uk; [blattner@ipk-](mailto:blattner@ipk-gatersleben.de)
24 [gatersleben.de](mailto:blattner@ipk-gatersleben.de)

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26 **Abstract [max 300 words]:**

27 Poor morphological and molecular differentiation in recently diversified lineages is a
28 widespread phenomenon in plants. Phylogenetic relationships within such species complexes are
29 often difficult to resolve because of the low variability in traditional molecular loci, as well as
30 various other biological phenomena responsible for topological incongruence such as ILS and
31 hybridization. In this study, we employ a Genotyping-by-sequencing (GBS) approach to
32 disentangle evolutionary relationships within a species complex belonging to the Neotropical
33 orchid genus *Cycnoches*. The complex includes seven taxa distributed in Central America and
34 the adjacent Chocó biogeographic region, nested within a clade estimated to have first diversified
35 in the early Quaternary. Previous phylogenies inferred from a handful of loci failed to provide
36 support for internal relationships within the complex. Our Neighbor-net and coalescent-based
37 analyses inferred from *ca.* 13,000 GBS loci obtained from 31 individuals belonging to six of the
38 seven traditionally accepted *Cycnoches* species provided a robustly supported network. The
39 resulting three main clades are corroborated by morphological traits and geographical

40 distributions. Similarly, Maximum Likelihood (ML) inferences of concatenated GBS-loci
41 produced results comparable with those derived from coalescence and network-based methods,
42 albeit always with poor statistical support. The low support evident in the ML phylogeny might
43 be attributed to the abundance of uninformative GBS loci, which can account for up to 50% of
44 the total number of loci recovered. The phylogenomic framework provided here, as well as
45 morphological evidence and geographical patterns, suggest that the six entities previously
46 thought to be different species might actually represent only three distinct segregates. Our study
47 is the first to demonstrate the utility of GBS data in phylogenomic research of a very young
48 Neotropical plant clade (~2 Ma), and it paves the way for the study of the many other species
49 complexes that populate the species-rich orchid family.

50 *Keywords:* American Tropics, high-throughput-sequencing, Orchidaceae, phylogenetic
51 incongruence, rapid diversification.

52 **1. Introduction**

53 Species complexes are aggregates of putative species that exhibit little morphological
54 differentiation or genetic divergence. They present a challenge to systematists and molecular
55 biologists because their phylogenetic relationships are particularly difficult to resolve (Després et
56 al., 2003; Escudero et al., 2014). This is particularly true when attempts to disentangle
57 genetically the evolutionary history within such lineages are based on only a small number of
58 loci (Taberlet et al., 2007), because the number of informative positions they provide is very
59 limited. Moreover, past and/or ongoing biological phenomena that are known to produce
60 discordance among gene trees (e.g. hybridization and incomplete lineage sorting [ILS]: (Fehrer
61 et al., 2007; Pérez-Escobar et al., 2016a)) further exacerbate the difficulty of species-tree
62 inference for species complexes (Fernández-Mazuecos et al., 2018; Salichos et al., 2014).

63 The recent advent of several high-throughput sequencing methods has facilitated the
64 generation of millions of base pairs of DNA sequence data (Dodsworth, 2015). These approaches
65 have enabled researchers to sequence hundreds or thousands of loci in parallel for multiple
66 individuals (Hipp et al., 2014), potentially leading to a great increase in the number of
67 informative sites with which to infer evolutionary relationships inside lineages with little
68 divergence (e.g. Escudero et al., 2014). Among the plethora of high-throughput sequencing
69 techniques, Genotyping-by-sequencing approaches (GBS, also referred as to Restriction-site
70 Associated DNA sequencing, RADseq; Andrews et al. 2016) reduce the complexity of a genome
71 by means of restriction enzymes (Andrews et al., 2016; Elshire et al., 2011). Enzymes specific to
72 restriction cut sites digest a genome at specific lengths, thus enabling the sequencing of
73 fragments adjoining enzyme cut sites and the generation of huge yet tractable numbers of genetic
74 markers per individual (Hipp et al., 2014). For the particular case of GBS, methylation-sensitive
75 restriction enzymes are employed for genome digestion, thus enabling more efficient access to
76 low-copy regions (Elshire et al., 2011).

77 Several studies have demonstrated the utility of reduced-representation genomic data for
78 resolving the phylogenetic relationships of plant and animal lineages derived from both ancient
79 (e.g. oaks: Hipp et al., 2014) and very recent rapid diversifications (e.g. cichlid fishes: Wagner et
80 al., 2013, toadflaxes (*Linaria*): Fernández-Mazuecos et al., 2018; *Diospyros*: Paun et al., 2016).
81 RADseq has also been successfully employed to tease apart the inter- and intraspecific
82 relationships of a recently diversified clade of *Heliconius* butterflies (Nadeau et al., 2013).
83 However, the extent to which such data are useful for inferring phylogenetic relationships in
84 plant species complexes has been little studied (Anderson et al., 2017).

85 Orchidaceae (the orchid family) is a hyper-diverse lineage of flowering plants distributed
86 worldwide (Chase et al., 2015), which is known for its rich and often intricate morphological
87 diversity (Mondragón-Palomino, 2013; Gramzow and Theißen, 2010; Mondragón-Palomino and
88 Theißen, 2008). Species complexes are common across the orchid family (e.g. Bateman et al.,
89 2017; Gale et al., 2018; Johnson and Steiner, 1997). As such, orchids provide a good opportunity
90 to test the utility of GBS data for resolving phylogenetic relationships among taxa with little or
91 overlapping morphological variation. Notable among orchids with intricate reproductive
92 morphology (Fig. 1, inset) is the Neotropical genus *Cycnoches* (Darwin, 1877), which
93 encompasses 34 epiphytic species distributed from Southern Mexico to Bolivia and Central
94 Brazil (Pérez-Escobar, 2016a).

95 Previous studies using a combination of three nuclear and two plastid loci failed to
96 resolve the phylogenetic relationships of very young clades within *Cycnoches* (Batista et al.,
97 2014; Gerlach and Pérez-Escobar, 2014; Pérez-Escobar et al., 2017a; Pérez-Escobar et al.,
98 2017b). This statement is particularly true for the *C. egertonianum* species alliance, a complex
99 consisting of seven species and two subspecies restricted to Central America and the Chocó
100 region of South America. Their members are characterized by male flowers with narrow,
101 elongated tepals (which can be pale green to deep purple), and by a labellum that is divided into
102 oblong to capitate dactylar processes (Fig. 1) (Carr, 2012; Romero and Gerlach, n.d.). Ample
103 intraspecific morphological variability exists and is most evident in tepal coloration patterns and
104 morphology of the dactylar processes that extend radially from the labellar margin (Fig. 1).
105 Moreover, herbarium material of *Cycnoches* is very scarce; some putative species of the
106 *egertonianum* complex are known only from drawings of the type specimen (*C. pachydactylon*,
107 *C. amparoanum*, *C. pauciflorum*, *C. powellii*, *C. stenodactylon*), thus effectively precluding
108 species delimitation by means of morphology.

109 The available molecular phylogenies of *Cycnoches* recovered the *C. egertonianum*
110 complex as a monophyletic group composed of two clades. Further comparison between nuclear
111 and plastid phylogenies revealed strongly supported incongruences within the complex, thus
112 suggesting ILS and/or that primary hybridization or introgression events have taken place (Pérez-
113 Escobar et al., 2016b). In this study, we test the utility of GBS data for resolving the evolutionary
114 history of the *C. egertonianum* complex by investigating six members (four species and two

115 subspecies; Carr 2012) using phylogenomic analyses. We specifically ask: (i) do all species in
116 the *C. egertonianum* complex conform to monophyletic groups? (ii) are ML phylogenies
117 congruent with trees derived from coalescent-based methods? And (iii) how informative are the
118 loci produced by GBS reduced-representation genomic data for resolving phylogenetic
119 relationships in the ~2 Myr-old *C. egertonianum* complex? Answering these questions allows us
120 to draw more general conclusions regarding the applicability of GBS methods to recent and rapid
121 diversifications.

122 2. Material and Methods

123 2.1. Taxon sampling, DNA extraction and Genotyping-by-sequencing (GBS) library preparation

124 We analysed 29 accessions together representing four species and two subspecies of the
125 *C. egertonianum* complex, collectively sampled from four localities. Additionally, we included
126 as outgroup single accessions of *C. barthiorum* and *C. herrenhusanum*. The number of
127 morphotypes sampled per species and voucher information are provided in Table S1. Genomic
128 DNA was extracted from fresh leaf tissue (preserved in silica gel) with the NucleoSpin® plant
129 kit (Macherey-Nagel; Düren, Germany), following the manufacturer's protocol. Total genomic
130 DNA was analysed on a 1.5% Agarose gel, and concentration and fragment length distribution
131 were assessed relative to a DNA standard. Genomic library preparation for GBS was performed
132 at the Institute for Plant Genetics and Crop Plant Research (IPK), Germany, following a protocol
133 of Elshire et al. (2011) as modified by Weltman (2016), using the rare-cutting enzyme *PstIHF*®
134 (recognition site: CTGCA'G) and the methylation-sensitive enzyme *MspI* (recognition site:
135 C'CGG). Genomic libraries were sequenced at the IPK on an Illumina HiSeq 2000, generating
136 single-end reads of 100 bp.

137 2.2. DNA sequence data analysis, SNP detection and phylogenomic inference

138 Illumina reads were trimmed, filtered and *de novo* assembled with the pipeline ipyRAD
139 v.3.0 (Eaton, 2014) using the default parameters recommended for single-end read data. We
140 produced GBS loci assemblies using a clustering value (*c*) of 0.95 and the statistical base-calling
141 parameters of (Li et al., 2008), specifically a minimum depth coverage of six and a maximum
142 number of five Ns in the consensus sequence. Paralogous loci were filtered by setting two alleles
143 and eight heterozygous positions per consensus sequence. To assess whether missing data had an
144 impact on tree topologies (Huang and Knowles, 2016), we employed three contrasting thresholds
145 of taxon coverage (i.e. minimum number of species per locus – *s*), specifically 15%, 50% and
146 100% of the total number of species, each in combination with *c* = 0.95. Thus, three DNA
147 matrices were set for phylogenomic inference: c95s15, c95s50 and c95s100.

148 We inferred Maximum Likelihood trees from the three DNA matrices using RAxML
149 v.8.0 (Stamatakis, 2014) with the GTR+G substitution model and 1000 bootstrap replicates,
150 operating via the CIPRES Science Gateway computing facility (Miller et al., 2015). We also

151 analysed the same DNA matrices in SplitsTree4 (Huson, 1998) to produce Neighbor-net
152 networks (i.e. split graphs) derived from uncorrected P-distances. Split graphs are considered
153 more suitable than phylograms or ultrametric trees to represent evolutionary histories that are
154 still subject to reticulation (Rutherford et al., 2018).

155 Maximum Likelihood analyses of concatenated loci might produce biased topologies
156 (Rokas et al., 2003), particularly in the presence of biological phenomena responsible for gene
157 tree incongruence such as hybridization and ILS (Fernández-Mazuecos et al., 2018). Thus, to
158 obtain a species tree from the complete multi-locus data sets while considering ILS, we
159 performed the coalescence-based analysis SVDquartets (Chifman and Kubatko, 2014). To test
160 the monophyly of the putative species and subspecies in *C. egertonianum* complex, we carried
161 out analyses assigning tips to delimitations indicated by concatenated and network analyses
162 produced in this study (i.e. with the ‘taxon partition’ option) and also without the ‘taxon
163 partition’ option. We executed SVDquartets in the software PAUP* v.4.0a (Swofford, 2001), and
164 for each dataset, we evaluated 100,000 random quartets and performed 100 bootstrap replicates
165 under the multispecies coalescent tree model.

166 2.3. Assessing per-locus phylogenetic informativeness

167 We assessed the performance of every GBS locus to resolve phylogenetic relationships in
168 *C. egertonianum* complex by estimating the net and per-site phylogenetic informativeness (PI)
169 across an arbitrary time scale (tips assigned to time 0 and root to 1) following the method of
170 Townsend (2007) and using the web service PhyDesign (Townsend, 2007) (available at
171 <http://phydesign.townsend.yale.edu/>). This package requires locus alignments with complete
172 taxon sampling and an ultrametric tree. Thus, we sampled sequences of 29 individuals (one
173 outgroup and 28 ingroup samples) from 2297 locus alignments derived from the c95s100 dataset.
174 We concatenated the sampled alignments to form a new super matrix, and inferred an ML tree
175 with the same settings as specified above; the tree was later converted into a chronogram with
176 PATHd8, a program for phylogenetic dating without a molecular clock
177 (<https://www2.math.su.se/PATHd8/>) (Britton et al., 2007; Schoch et al., 2009). The PI profiles
178 were estimated with the HyPhy substitution rates algorithm for DNA sequences (Kosakovsky-
179 Pond et al., 2005). We identified the sites with unusually high substitution rates that may cause
180 phylogenetic noise with the R script and filtering method described by Fragoso-Martínez et al.
181 (2016) using a cut-off value of five. The identified sites were removed manually from the
182 alignments using the software Geneious v8.1 (Biomatters Ltd.; Kearsse et al., 2012) and these
183 corrected matrices were uploaded again to PhyDesign as described above.

184 The net and per-site PI of the GBS loci was compared with the performance of an
185 Internal Transcribed Spacer (ITS) alignment for the same set of species. Here, ITS sequences for
186 each individual were obtained by mapping trimmed reads (GBS sequence data) for each species
187 against an nrITS reference sequence of *C. herrenhusanum* (GenBank acc. MF285490) and

188 producing a majority rule consensus sequence in Geneious v8.1. The ITS sequences were then
 189 aligned in MAFFT (Katoh et al., 2017) using the Q-INS-I strategy, which is optimized to
 190 consider secondary structures during the alignment process. A ML tree from the ITS alignment
 191 was inferred and then converted to an ultrametric tree, after which it was analysed in PhyDesign
 192 using the aforementioned settings.

193 To understand the extent to which particular GBS loci resolve phylogenetic relationships
 194 at any node with confidence, we computed (i) the number of nodes receiving null to maximum
 195 Likelihood Bootstrap Support (LBS) values, and (ii) the distribution of LBS across node depth.
 196 Here, we obtained ML trees for every locus using RAxML v.8.0 (Stamatakis, 2014) with the
 197 GTR+G substitution model and 100 bootstrap replicates. We classified LBS values at nodes into
 198 the following intervals: [0–10], [11–20], [21–30], [31–40], [41–50], [51–60], [61–70], [71–80],
 199 [81–100]. Node depth values were obtained for every locus tree by converting the ML phylogeny
 200 into an ultrametric tree with a root age of 1 using the penalized likelihood method implemented
 201 in the function *chronos* from the R package APE (Paradis et al., 2004) which is faster and easier
 202 to parallelize than PATHd8 for analyses of large numbers of trees. Here, we set a root age of 1,
 203 following the approach of Lee et al. (2018), and used a lambda value of 0 for rate smoothing.
 204 Node number vs LBS intervals and node depth vs LBS values were plotted using the R package
 205 GGPLOT2 (Wickham, 2009).

206 3. Results

207 3.1. Genotyping-by-sequencing data recovery

208 Illumina sequencing produced *ca.* 66 million raw reads, bases with a quality score of Q30
 209 for 96.3% reads and a GC content of 45.7%. The number of loci recovered under different taxon
 210 coverages and the proportion of missing data are summarised in Table 1. As expected, the
 211 highest total number of loci recovered (13,960) was achieved under the smallest taxon coverage
 212 (i.e. $s = 15$) whereas the smallest number of sequenced loci (2,297) was obtained under the
 213 highest taxon coverage, $s = 100$. These loci have almost the same length (median=102 bp,
 214 mean=100.8, SD=11.5).

Data set	Minimum taxon coverage	Number of loci	Concatenated length	Missing data (%)
c95s15	15%	13,960	1,382,653	35.2%
s95s50	50%	9,626	949,726	17.15%
c95s100	100%	2,297	231,238	0%

216

217 **Table 1**

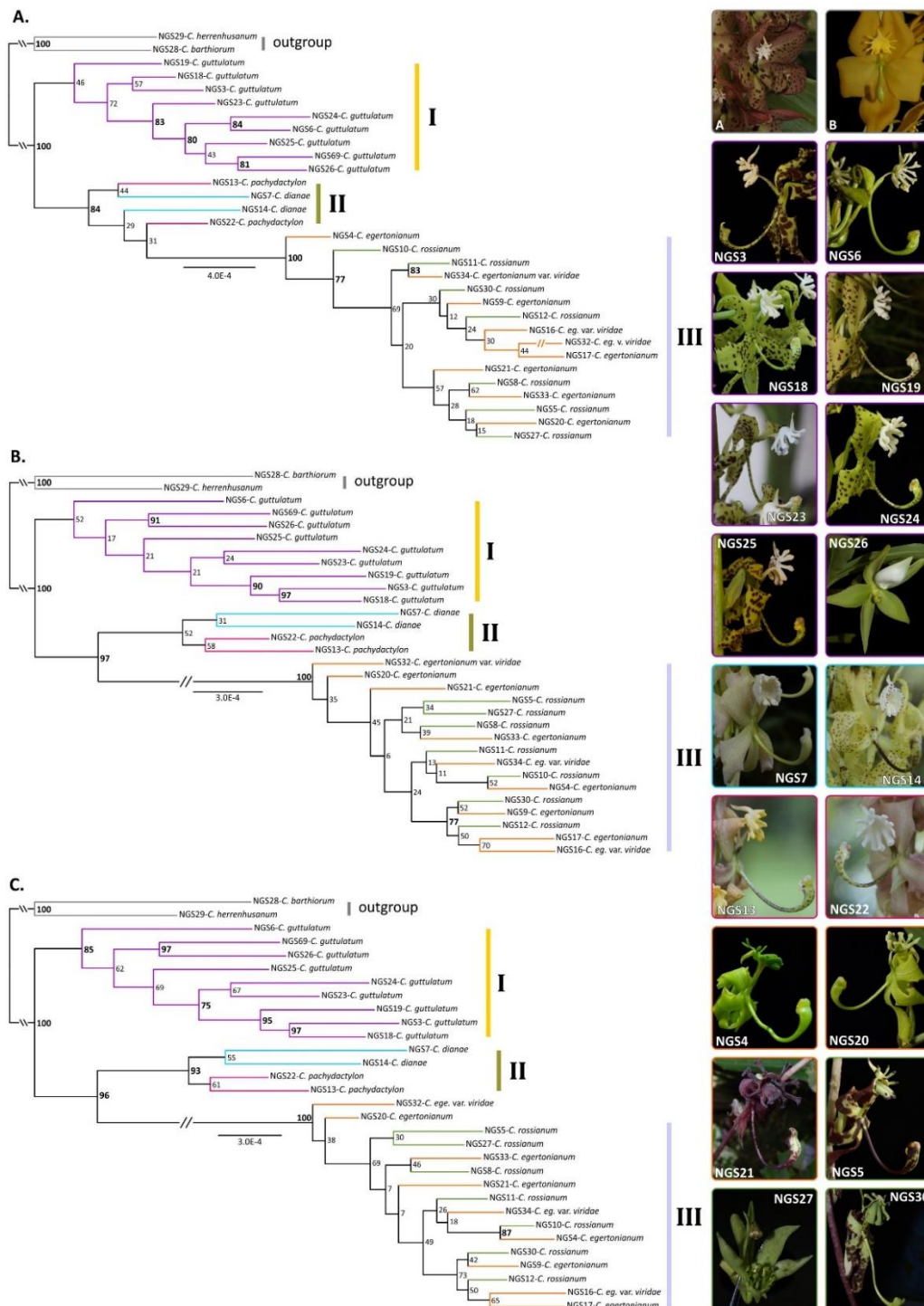
218 Characteristics of the three assembled GBS datasets under ipyRAD.

219

220 3.2. Phylogenomic inferences

221 Maximum Likelihood (ML) analyses of the c95s15, c95s50 and c95s100 concatenated
 222 DNA matrices produced similar topologies, and provided strong statistical support for the C.

223 *egertonianum* complex (LBS = 100; Fig. 1). They recovered three weakly to moderately
 224 supported main groups (I–III; Fig. 1). Group I included all samples identified as *C. guttulatum*
 225 (LBS = 46–85) and was placed as sister to groups II plus III. Group II clustered all specimens
 226 assigned to *C. diana*e and *C. pachyactylon* (LBS = 52–93), except in the ML tree of c95s100,
 227 which rendered the group paraphyletic (Fig. 1A). However, the topologies of c95s15 and c95s50
 228 grouped *C. diana*e and *C. pachyactylon* samples as reciprocally monophyletic, albeit with weak
 229 statistical support (LBS = 31–55, and 58–61, respectively). Group III included samples of *C.*
 230 *rossianum* and the two subspecies of *C. egertonianum* (LBS = 100), all intermingled across the
 231 clade rather than clustering according to traditional taxonomy.



264 **Fig. 1.** Best scoring Maximum Likelihood trees inferred under contrasting proportions of missing data:
265 (A) c95s100 (i.e. no missing data); (B) c95s50 (minimum number of 50% of species per locus included);
266 (C) c95s15 (minimum number of 15% of species per locus included). Likelihood Bootstrap Support
267 (LBS) values > 75 are displayed in boldface. Branches are colour coded according to taxonomic
268 identities. Clades I–III are indicated by vertical bars. (Inset) Pictures of individuals sampled, colour coded
269 according to taxonomic identity. Left: *Cynoches barthiorum*; Right: *C. herrenhusanum*.

270
271

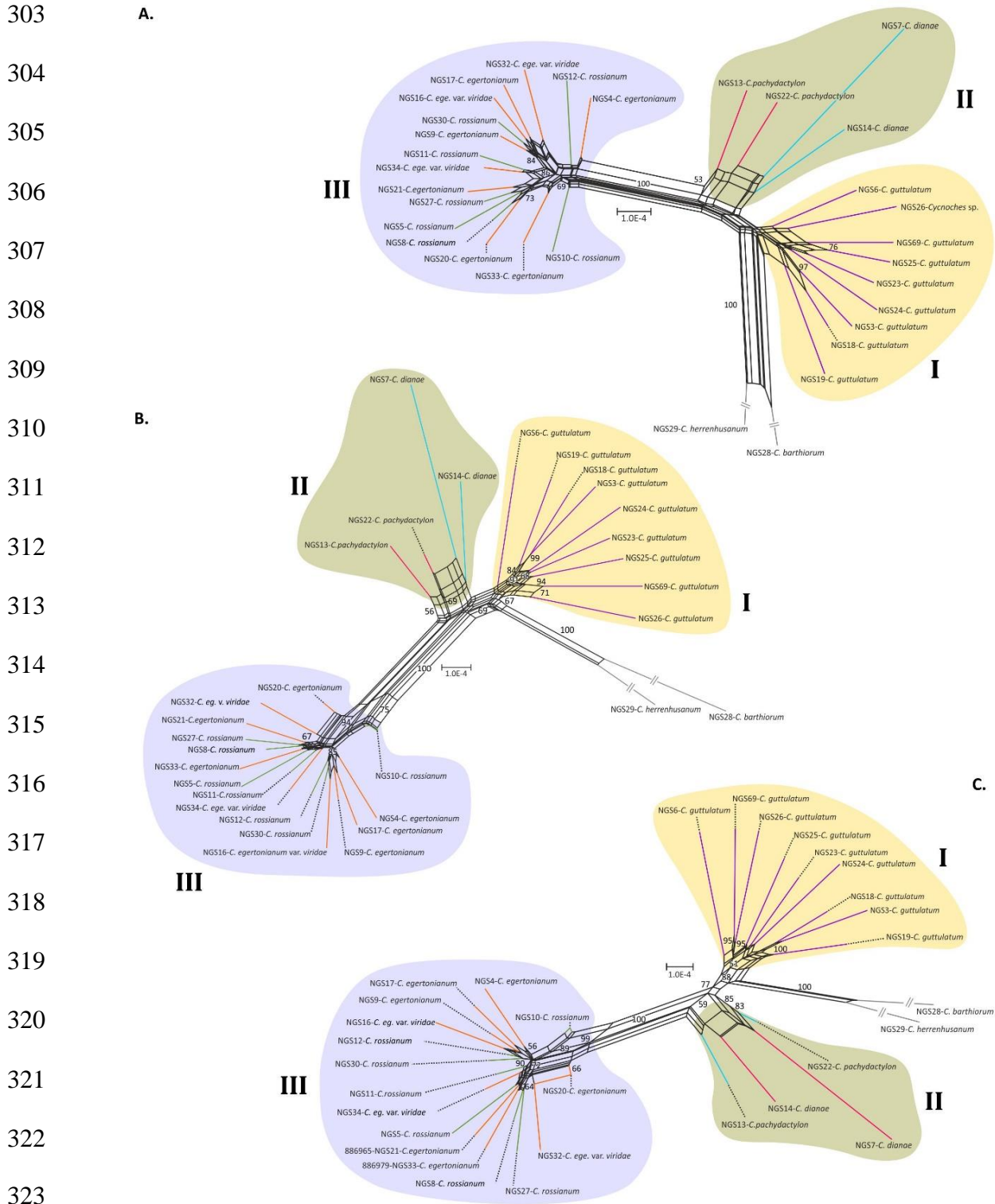
272 Uncorrected P-distance split networks of all DNA matrices recovered the same groups
273 (i.e. clades I–III; Fig. 2) produced by the ML phylogenies and revealed a clear distinction
274 between each cluster. Here, groups I and II were distanced from each other by very short splits,
275 whereas group III was placed more distantly from both groups I plus II and the outgroup, a
276 topology consistent with the ML trees. The bootstrap support (B) of splits was proportional to the
277 amount of missing data; the c95s100 network contained nine splits with support values
278 exceeding 50, compared with 18 such splits obtained in the network derived from c95s15. The
279 splits leading to group III from groups I plus II were in all instances strongly supported (B =
280 100), while splits joining groups I and II were only moderately supported (i.e. B = 85, 88) in the
281 network derived from c95s15 (Fig. 2C). None of the split networks produced clear clustering
282 patterns between samples of the same named taxon within either group II or group III.

283
284

3.3. Coalescence-based phylogenies

285 The SVDquartet analyses produced backbone topologies very similar to those derived
286 from ML analyses (Figs. S1, S2). Inferences with and without the taxon partition option
287 produced similar phylogenies in both topology and bootstrap support values across the different
288 taxon coverages (Fig. 3), and strongly supported monophyly of the *C. egertonianum* complex.
289 All analyses without the taxon partition recovered all accessions of *C. guttulatum* as a well-
290 supported monophyletic group (I in Figs. S1, S2) and placed it as sister to the rest of the alliance.
291 The samples of *C. diana* were revealed as a paraphyletic group, and one of them (NGS14)
292 formed a moderately supported clade (B = 81–88) together with the two accessions of *C.*
293 *pachydactylon* (group II). The accessions of *C. rossianum* and the two subspecies of *C.*
294 *egertonianum* were intermingled in a strongly supported clade (together constituting group III; B
295 = 100). No clustering pattern between samples of the same species was evident within this clade.

296 The trees produced using taxon partitions converged on the same topology derived from
297 analyses without taxon partitions, albeit with some differences in bootstrap support values (Fig.
298 3). Here, *C. guttulatum* was recovered as sister to rest of the species alliance (group I).
299 *Cynoches diana* and *C. pachydactylon* were placed as sister lineages in a moderate-to-strongly
300 supported clade (group II, B = 84–94). Group II was in turn recovered as sister to the *C.*
301 *egertonianum* + *C. rossianum* clade (group III) with high statistical support (B = 99–100). Group
302 III received similarly strong statistical support (B = 100).



324 **Fig. 2.** Uncorrected P-distance split networks under different proportions of missing data: (A) c95s100
 325 (i.e. no missing data); (B) c95s50; (C) c95s15. Bootstrap support values (B) > 50 at splits are shown.
 326 Main clusters are colour coded according to main clades (I–III) shown in Fig. 1.

327 3.4. Phylogenetic informativeness and node support of GBS loci

328 The net phylogenetic informativeness (PI) analyses of the GBS loci generated a total of
329 231,997 observations. The per-site and net PI values for all loci varied dramatically, ranging
330 from 0 to 0.13, and from 0 to 135.09, respectively. Most of the loci reached their highest PI
331 values between a time interval of 0.06–0.20, which includes the most recent common ancestors
332 (MRCAs) of *C. egertonianum* + *C. rossianum* accessions. 1,296 out of 2,297 loci (56.4%) were
333 completely uninformative (net and per-site PI = 0). Only 346 loci had net and per-site PI values
334 higher than the mean and peaked at an average time of 0.3, though the most informative peaked
335 closer to the present. Of these, two loci (locus679 and locus787) showed disproportionate
336 increases in net and per-site PI values towards the present (between times 0.08 and 0.15). Both
337 net and per-site PI values of the GBS loci yielded almost identical results. The mean net PI value
338 of nrITS was ~210, and it peaked at time 0.5, near the MRCAs of *C. pachydactylon* + *C. diana*
339 *grade* + *egertonianum* + *C. rossianum* and *C. guttulatum* (Fig. 4). This locus was the most
340 informative compared with all GBS loci for the net PI. In contrast, the maximum per-site PI
341 value of the nrITS (~0.028) was lower than 42 GBS loci but still much higher than the average of
342 the remaining GBS.

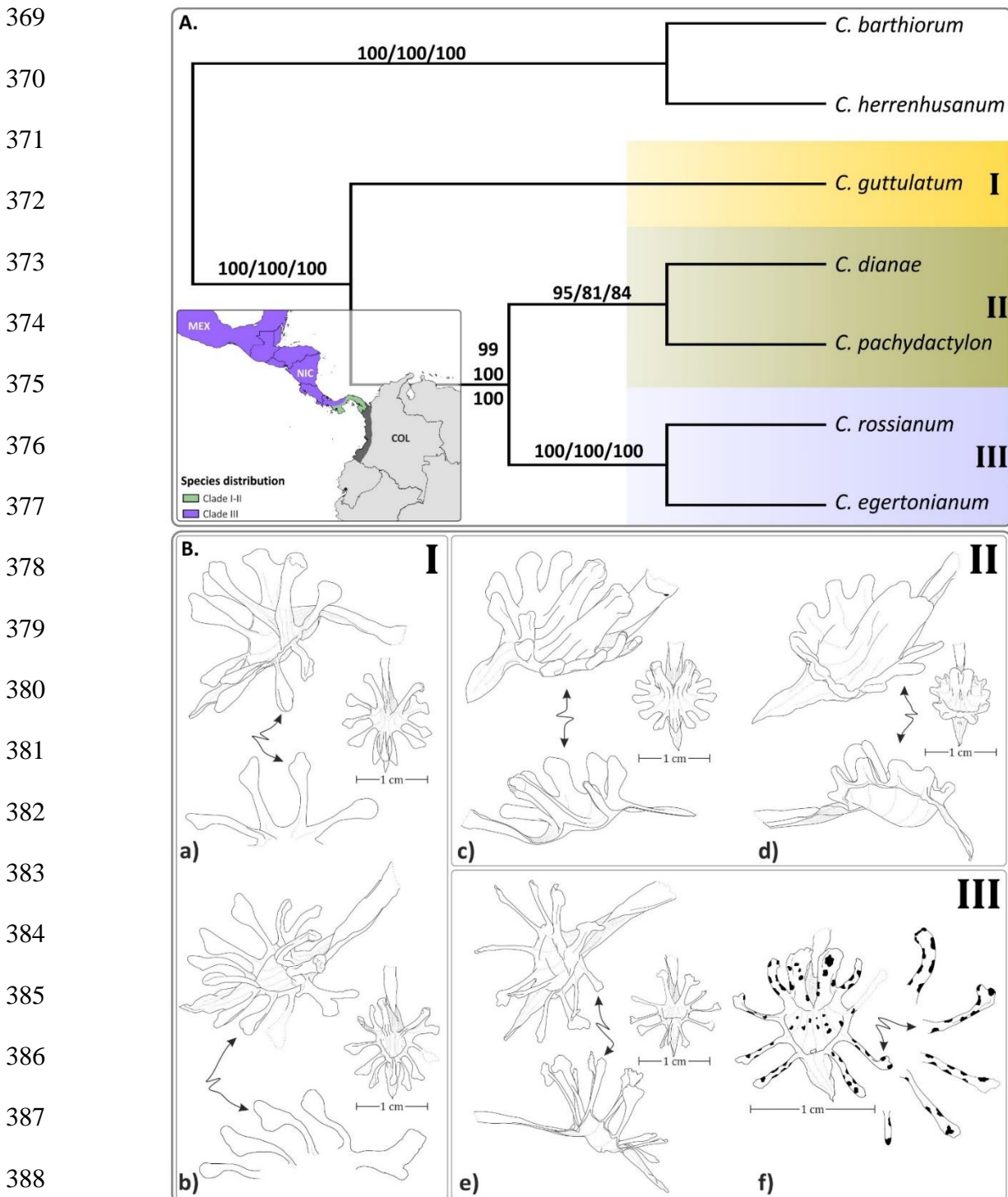
343 The distribution pattern of LBS values at nodes was virtually identical across the
344 c95s100, c95s50 and c95s15 datasets, most of the node support values being strongly skewed
345 towards the 0–10 LBS interval (Figs. S3–S5). Very few nodes were moderately to strongly
346 supported across gene trees (LBS interval 81–100), and they accounted for only ~1% of the total
347 number of nodes across gene trees in each dataset. Likewise, the distribution of LBS values
348 across node depth was strikingly similar across the three DNA datasets (Figs. S6–S8). Here, we
349 could detect no clear distribution pattern of LBS support at a given node depth.

350 4. Discussion

351 4.1. GBS-loci and ITS phylogenetic informativeness in the *C. egertonianum* complex

352 Several studies have convincingly demonstrated the utility of SNP discovery methods
353 coupled with high-throughput sequencing to resolve rapid plant diversifications as recent as ~5
354 Myr old (Fernández-Mazuecos et al., 2018; Hou et al., 2015). More recently, Lee et al. (2018)
355 showed that ddRADseq is subject to a dramatic decrease in the number of parsimony-
356 informative SNPs towards both shallow and deeper phylogenetic levels (i.e. suffers from locus
357 dropout). Our comprehensive analysis of LBS values across nodes in gene trees and species
358 trees, and the phylogenetic informativeness of GBS data, at least partially follows the locus
359 dropout behaviour previously observed by Lee et al. (2018). However, our results differ in that
360 the lack of support and informativeness is not confined to very shallow nodes but rather appears
361 to occur at all phylogenetic depths. Perhaps more importantly, their comparable average PI
362 values indicate that a widely used marker such as nrITS might be almost as informative as the
363 majority of the GBS loci recovered here. However, some shorter GBS loci may be good

364 candidates to improve resolution as they have higher per-site PI values than nrITS and peaked
 365 closer to the present (locus679 and locus787). The higher net PI values of nrITS are associated
 366 with the length of this locus (612 bp), which is six times the average of the GBS loci length.
 367 Thus, the concatenation of all GBS loci with positive PI values might bring considerably more
 368 phylogenetic signal to a particular dataset than would nrITS alone.



389 **Fig. 3.** (A) The 50% majority rule consensus tree derived from SVDQuartets analysis for all DNA
390 matrices (c95s100, c95s50 and c95s15) using taxon partition. Bootstrap support values > 75 are displayed
391 in boldface. Bootstrap values are listed in the following order: c95s100/c95s50/c95s15. Identical B values
392 produced from both datasets are shown only once. Clades I–III are indicated with vertical bars. (Inset)
393 Geographical distribution of terminals nested in clades I, II, and III. (B) Detailed illustrations of the
394 labellum (lateral and ventral view, including details of the dactylar processes indicated by the black
395 arrows) of preserved specimens of selected members of the *Cycnoches egertonianum* complex. (a) *C.*
396 *guttulatum* (drawn from the Isotype Powell 14 [AMES]); (b) *C. guttulatum* (Powell 20 [AMES]); (c) *C.*
397 *pachydactylon* (Gerlach 00/3415 [M]); (d) *C. diana*e (Powell 186 [AMES]); (e) *C. egertonianum* var.
398 *viridae* (Hamer 87 [AMES]); (f) *C. rossianum* (Tuerckheim 7777 [AMES]). Illustrations by O. Pérez-
399 Escobar extrapolated from rehydrated herbarium material or flowers preserved in spirits.

400 A striking result of our study is the large number of GBS loci with null PI values, which
401 can be more than 50% of the total GBS loci produced by ipyRAD. A similar lack of resolution
402 and phylogenetic structure have been reported within each of the nine very young clades that
403 together constitute the temperate terrestrial orchid genus *Ophrys* (Bateman et al., 2018). Here,
404 phylogenetic relationships within the young (< 1 Myr old) *O. sphegodes* and *O. fuciflora*
405 complexes were rendered poorly supported in ML analysis of concatenated RAD loci. The same
406 pattern was also recovered in a split network analysis, which revealed large phylogenetic
407 uncertainty within these clades and some residual uncertainty between them (Bateman et al.,
408 2018). Broadly similar results have been obtained from species complexes within the terrestrial
409 orchid genera *Dactylorhiza*, *Gymnadenia* and *Epipactis*.

410 Low statistical support is often attributed to the presence of phylogenetic incongruence
411 among gene trees (Aberer et al., 2013; Wilkinson, 1996). Alternatively, the exclusion of loci
412 with missing data might also negatively affect the phylogenetic accuracy and the probability of
413 recovering taxa as monophyletic (Huang and Lacey Knowles, 2016). The distribution of LBS
414 values across nodes in trees derived from the three different matrices with different degrees of
415 missing data (i.e. c95s100, c95s50, c95s15) is virtually identical; the majority of gene-tree nodes
416 have very low LBS values, and low LBS values occur at all phylogenetic depths. Likewise, the
417 phylogenies produced from the three datasets resulted in similar topologies, despite the fact that
418 each incurred poor statistical support (Fig. 1). Here, the only notable difference between the
419 datasets is the monophyly of group II, which is recovered as monophyletic (with weak to
420 moderate support) in c95s50 and c95s15 (i.e. datasets with greater proportions of missing data).
421 Thus, in the case of *Cycnoches*, the proportion of missing data did not have a notable impact on
422 the node support of any of our phylogenetic estimations, though it did affect to some extent the
423 topology. Additionally, given the overall low LBS support across loci trees, it is difficult to
424 assess whether there are statistically supported phylogenetic conflicts among our loci trees.

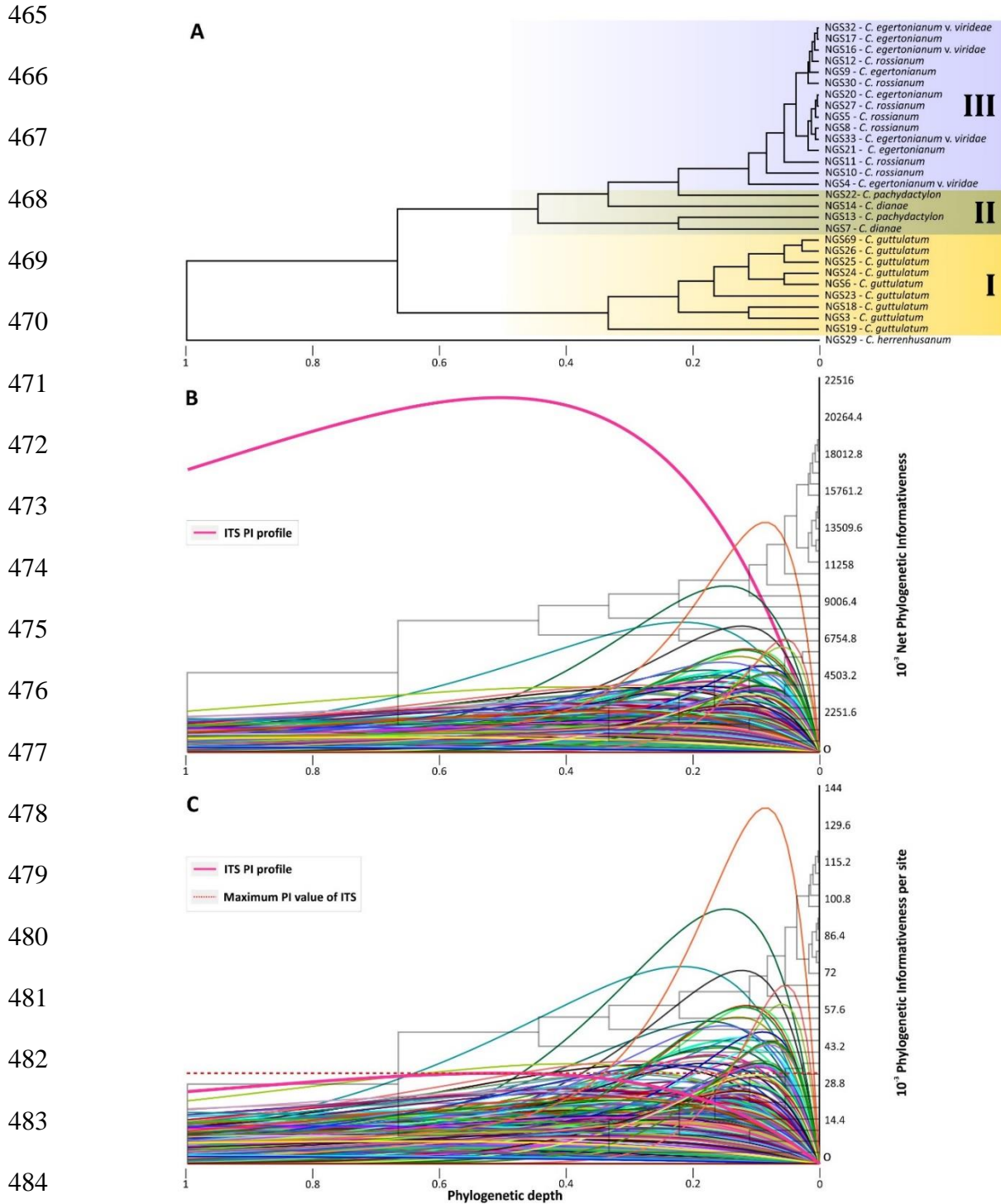
425 Lack of phylogenetic signal in the loci data, or rapid diversification events, are further
426 thought to produce low statistical support (Bogarín et al., 2018). Low PI profiles are derived
427 from sites evolving at rates that are either extremely fast or extremely slow (Townsend 2007).

428 Even though we do not disregard the influence of the biological phenomena responsible for
429 among-loci tree incongruence previously reported for *Cycnoches* by Pérez-Escobar et al. (2016b,
430 2016c), we speculate that poor statistical support in our ML phylogenies is indeed derived from
431 the overall low PI of the GBS loci here sequenced. The low PI values and poor bootstrap support
432 in our data indicate that perhaps more variable regions are needed to disentangle species
433 relationships resulted from rapid radiations (López-Giráldez et al., 2013; López-Giráldez and
434 Townsend, 2011; Townsend, 2007).

435 In this way, alternative reduced-representation approaches such as anchored hybrid
436 enrichment (AHE) have recently been employed to resolve phylogenetic relationships at very
437 shallow phylogenetic levels in both plant and animal lineages (Fragoso-Martínez et al., 2016;
438 Granados Mendoza et al., 2013; Lemmon and Lemmon, 2012; Wanke et al., 2017). In the case of
439 orchids, AHE did not yield loci alignments with PI values equal to 0. Therefore, they were more
440 informative than the GBS loci sequenced here (Bogarín et al., 2018). Moreover, the AHE loci
441 resolved intricate phylogenetic relationships with confidence in a taxonomically difficult
442 complex of a recent lineage of *Lepanthes* orchids. Thus, we suggest that alternative reduced-
443 representation methods such as targeted enrichment might also be also suitable to disentangle
444 phylogenetic relationships in very recent orchid diversifications. Here, lineage-specific sets of
445 targeted sequencing probes can retrieve hundreds (~500) of orthologous known-exons from all
446 plant genomic compartments plus their partial, contiguous intronic regions (Bogarín et al., 2018;
447 Johnson et al., 2018). Such intronic regions certainly enhance the phylogenetic informativeness
448 of gene alignments (~1000 bp) at shallow phylogenetic levels. The advantage of targeted
449 enrichment methods and the well-defined regions that they target stands in stark contrast with the
450 greatest major drawback of GBS approaches; GBS samples unknown genomic regions (Andrews
451 et al., 2016), some of which might be under contrasting selective pressures.

452 4.2. Paraphyly in the *C. egertonianum* complex

453 Previous maximum Likelihood and Bayesian analyses of *Cycnoches* inferred from three
454 nuclear and two plastid DNA loci have revealed two clades within the *C. egertonianum* complex,
455 and strongly supported the previously demonstrated monophyly of the alliance. However, they
456 failed to disentangle the internal relationships of these clades (Batista et al., 2014; Gerlach and
457 Pérez-Escobar, 2014; Pérez-Escobar et al., 2017). The copious amount of GBS loci obtained
458 from our multiple-accessions-per-species approach, together with comparative phylogenomic
459 analyses, do not support the monophyly of the three species (i.e. *C. diana*, *C. pachydactylon*
460 and *C. rossianum*) as well as the two subspecies of *C. egertonianum*. Only the monophyly of *C.*
461 *guttulatum* is (weakly to moderately) supported by all analyses. More importantly, they also
462 consistently point to the monophyly of two species aggregates, namely *C. pachydactylon* + *C.*
463 *diana* (clade II) and *C. egertonianum* + *C. rossianum* (clade III), implying that clades II and III
464 may in fact represent two *bona fide* species.



485 **Fig. 4.** (A) Ultrametric tree derived from a ML phylogeny computed from the concatenated c95s100
 486 dataset. (B) Net phylogenetic informativeness (PI) profile per locus of the filtered c95s100 GBS loci (for
 487 details of filtering strategy see Section 2.2). (C) Per-site PI profiles of filtered c95s100 GBS loci. The
 488 dashed red line indicates the maximum net PI of the nrITS for the same phylogeny and phylogenetic
 489 depth. The maximum PI value obtained for the filtered loci alignments is indicated. Note the lower
 490 maximum PI value of the nrITS.

491 The three clades confidently supported and recovered by all ML inferences and by
492 coalescence-based analyses with taxon partitions also exhibit a strong morphological and
493 geographical structure. Here, *C. guttulatum* and *C. pachydactylon* + *C. diana* are restricted to
494 the lowland South-eastern wet forest of Panama. *Cycnoches powellii*, another member of the *C.*
495 *egertonianum* complex (only known from the type specimen, therefore not available for
496 sampling in this study), greatly resembles *C. diana* and occurs within the distribution range of
497 this species. We therefore hypothesise that *C. powellii* belongs to the *C. diana* + *C.*
498 *pachydactylon* clade. *Cycnoches guttulatum* can easily be distinguished from *C. diana* + *C.*
499 *pachydactylon* by the elongated, capitate to oblong dactylar processes of the labellum (Figs. 1,
500 3A, B) and the green to pale yellow sepals and lateral petals bearing conspicuous macules (Carr,
501 2012). Members of clade II in turn have vestigial to short, rectangular dactylar processes, and the
502 sepals and lateral petals are either pink or pale green, usually lacking macules (when present,
503 these are inconspicuous; Figs. 1, 3C, D). The *C. egertonianum* + *C. rossianum* aggregate (clade
504 III) is distributed from the Cordillera de Talamanca in Panama northwards to Southern Mexico.
505 The morphology of the dactylar processes in this clade broadly resembles that of *C. guttulatum*,
506 though in this aggregate the sepals and lateral petals are usually deep purple, plain green or green
507 with conspicuous purple markings (Figs. 1, 3E, F).

508 Because phylogenetic incongruence is a pervasive phenomenon across the plant tree of
509 life (Eiserhardt et al., 2018; Rokas et al., 2003), species delimitation based on multilocus
510 phylogenies should always be accompanied by contrasting lines of evidence, preferably
511 including detailed morphological analyses (Bogarín et al., 2018). Thus, whether the three clades
512 here recovered represent three genuinely distinct entities merits further study. Even though our
513 coalescence-based phylogenies with taxon partition reject the monophyly of almost all of the
514 species previously recognised in the *C. egertonianum* complex, and both morphology and
515 geographical distribution clearly delimit clades I–III, further evidence is required to circumscribe
516 with sufficient confidence these three entities as distinct species. Our phylogenomic framework
517 does pave the way for further integrative studies combining morphological and ecological
518 information (e.g. plant-pollinator interactions) as well as presaging extended sampling to further
519 investigate the genome divergence among populations of members of the *C. egertonianum*
520 complex.

521

522 5. Conclusions

523 Our well-founded phylogenomic framework for the *C. egertonianum* complex suggests
524 that six of the seven sampled taxa recognised by traditional taxonomy more likely constitute only
525 three genuine species. Further testing through genomic, morphological and ecological studies is
526 desirable to confirm their circumscription and most appropriate rank. Genotyping-by-sequencing
527 reduced-representation genomic data coupled with coalescence-based inferences of relationships
528 is a useful tool to shed light on the phylogenetic relationships in recently diversified orchid

529 species complexes. However, given the lack of PI in a large proportion of the GBS loci
530 sequenced, alternative methods such as targeted enrichment – known to have effectively resolved
531 intricate phylogenetic relationships in equally young orchid lineages – might prove also suitable
532 for disentangling relationships within a species complex. In this study, exclusion of loci with
533 missing data did not notably affect node support, but rather clade recovery in concatenated ML
534 and coalescence-based inferences. Nevertheless, our results suggest that coalescence methods
535 with taxon partitioning produce better supported phylogenies compared with ML analyses, likely
536 due to the fact that coalescence methods can better account for topological incongruence among
537 loci, such as that resulting from ILS.

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549

550 **Appendix A**

551 **Table S1**

552 Species names and voucher information, including herbarium of voucher deposition for material
553 used in this study.

554 **Fig. S1.** The 50% majority rule consensus tree derived from SVDQuartets analysis for the DNA
555 matrix c95s100 (i.e. no missing data) without taxon partition. Bootstrap support values (B) are
556 provided at branches. Clades I–III are indicated with vertical bars.

557 **Fig. S2.** The 50% majority rule consensus tree derived from SVDQuartets analysis for the DNA
558 matrix c95s50 and c95s15 without taxon partition. Bootstrap support values > 75 are displayed in
559 boldface; values are shown in the order c95s50/c95s15. Clades I–III are indicated with vertical
560 bars.

561 **Fig. S3.** Distribution of LBS values in six different intervals at nodes for the c95s100 dataset.

562 **Fig. S4.** Distribution of LBS values in six different intervals at nodes for the c95s50 dataset.

563 **Fig. S5.** Distribution of LBS values in six different intervals at nodes for the c95s15 dataset.

564 **Fig. S6.** Distribution of LBS values in relation to phylogenetic depth for the c95s100 dataset.

565 **Fig. S7.** Distribution of LBS values in relation to phylogenetic depth for the c95s50 dataset.

566 **Fig. S8.** Distribution of LBS values in relation to phylogenetic depth for the c95s15 dataset.

567

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