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# Creating functional groups of marine fish from categorical traits

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**Background.** Functional groups serve two important functions in ecology, they allow for simplification of ecosystem models and can aid in understanding diversity. Despite their important applications, there has not been a universally accepted method of how to define them. A common approach is to cluster species on a set of traits, validated through visual confirmation of resulting groups based primarily on expert opinion. The goal of this research is to determine a suitable procedure for creating and evaluating functional groups that arise from clustering nominal traits.

**Methods.** To do so we produced a species by trait matrix of 22 traits from 116 fish species from Tasman Bay and Golden Bay, New Zealand. Data collected from photographs and published literature were predominantly nominal, and a small number of continuous traits were discretized. Some data were missing, so the benefit of imputing data was assessed using four approaches on data with known missing values. Hierarchical clustering is utilised to search for underlying data structure in the data that may represent functional groups. Within this clustering paradigm there are a number of distance matrices and linkage methods available, several combinations of which we test. The resulting clusters are evaluated using internal metrics developed specifically for nominal clustering. This revealed the choice of number of clusters, distance matrix and linkage method greatly affected the overall within- and between- cluster variability. We visualise the clustering in two dimensions and the stability of clusters is assessed through bootstrapping.

**Results.** Missing data imputation showed up to 90% accuracy using polytomous imputation, so was used to impute the real missing data. A division of the species information into three functional groups was the most separated, compact and stable result. Increasing the number of clusters increased the inconsistency of group membership, and selection of the appropriate distance matrix and linkage method improved the fit.

**Discussion.** We show that the commonly used methodologies used for the creation of functional groups are fraught with subjectivity, ultimately causing significant variation in the composition of resulting groups. Depending on the research goal dictates the appropriate strategy for selecting number of groups, distance matrix and clustering algorithm combination.

# 1 Creating functional groups of marine fish 2 from categorical traits

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## 12 ABSTRACT

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14 ecosystem models and can aid in understanding diversity. Despite their important applications, there  
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38 matrix and clustering algorithm combination.

## 39 INTRODUCTION

40 Marine ecosystems are large and complex, requiring simplification of their components in order to be  
41 studied and understood. One such simplification is the construction of functional species groups, which  
42 involves creating distinct sets of species according to a selection of their functional traits (Tilman, 2001).  
43 The groups are defined by the niche requirements of the species, rather than by their taxonomy (Root,  
44 1967), or their economic importance. In other words, a functional group comprises species with similar  
45 life history that respond to environmental fluctuations in a similar way within a given habitat. Defining  
46 functional groups “allows a context-specific simplification of the real world...” (pg. 5; Gitay and Noble,  
47 1997). This provides a basis from which food web analysis and relationships with other components of

48 the ecosystem can be derived (Gravel et al., 2016). There are two primary uses of functional groups:  
49 to simplify the numerous species contained in an ecosystem for modelling; and to assess the diversity  
50 of an ecosystem. It is a particularly important step in ecosystems modelling as it identifies the basic  
51 structures that become the inputs of the model, thus making the outputs more interpretable (Fulton et al.,  
52 2003). If functional groups are used in assessing the diversity of an ecosystem (in addition to or instead of  
53 species richness), the problem of functional redundancy can be avoided (Stuart-Smith et al., 2013), and  
54 the variation in the productivity of a given ecosystem can be more clearly observed (Tilman et al., 1997).

55 Functional groups for ecosystem models typical have been established using expert knowledge of the  
56 system and its inhabitants (Baretta et al., 1995; Olivier and Planque, 2017), while groups representing  
57 functional diversity have been created using trait or diet data and statistical classification methods  
58 (Petchey and Gaston, 2002). Diet data are commonly used to create functional groups of fishes in marine  
59 ecosystems, because diet can demonstrate resource partitioning between species, which is a key indicator  
60 of interspecific competition (Colloca et al., 2010; Sala and Ballesteros, 1997). However, diet data are time  
61 consuming and expensive to collect, and this type of analysis only takes into account part of the species  
62 role in the ecosystem. Therefore diet data should be complemented with morphological traits (Reecht  
63 et al., 2013; Albouy et al., 2011), known habitat associations and/or other life history traits (Stewart et al.,  
64 2006; Gravel et al., 2016) to derive functional groups. The usefulness of these groupings depends on the  
65 ecosystem of interest and the intended use of the groupings. Such intentions could be used to identify  
66 specialists (Dehling et al., 2016; Clavel et al., 2013), habitat use (Franco et al., 2008; Elliott et al., 2007)  
67 or predict prey selection (Spitz et al., 2014).

68 With such a wide array of applications there are inevitably many variations in approaches to deriving  
69 the groups. One approach is to record traits that reflect how species use the environment and its resources,  
70 and use those to cluster groups based on their similarities (Mindel et al., 2016). Selecting functional traits  
71 for classification is a crucial step in the grouping process as these ultimately determine how species group  
72 together (Bremner et al., 2006a). Functional groups can be defined by continuous traits, for example  
73 Albouy et al. (2011); Córdova-Tapia and Zambrano (2016) used continuous morphological measurements  
74 to infer a given species food source and its acquisition which were used to derive functional groups.  
75 These traits are time consuming and expensive to collect and measuring many traits for all members  
76 of species-rich ecosystems is impractical (Madin et al., 2016). The traits that will be most valuable in  
77 practice will be those available for most species (Costello et al., 2015). To create functional groups of  
78 benthic invertebrate communities categorical (nominal) traits are utilised to classify species (Bremner  
79 et al., 2006a). Using categorical rather than numerical features allows the data to be measured without units,  
80 and as traits are rarely measured with a common methodology this may lead to more reliable, complete  
81 and comparable data (McGill et al., 2006).

82 Functional groups defined from clustering using continuous data collected from species measurements  
83 (Albouy et al., 2011; Sibbing and Nagelkerke, 2000) can utilise traditional approaches to cluster analysis  
84 (e.g. Euclidean distances with Ward's minimum variance clustering (Dumay et al., 2004)). Clustering  
85 categorical (nominal) traits can use the same hierarchical approach, but cannot make use of most  
86 distance matrix algorithms. Instead, several alternative approaches to clustering nominal data have been  
87 suggested by Boriah et al. (2008) and Šulc and Řezanková (2014) that can then be used with traditional  
88 linkage methods. An important consideration that should be made and is often missed in these type of  
89 analyses is that choice of both distance matrix and linkage method will ultimately alter the composition of  
90 the clusters (Clifford et al., 2011).

91 While unsupervised learning (such as clustering) is a potentially powerful solution for finding func-  
92 tional groups, as yet there is no agreed method for assessing validity. Functional groups that arise from  
93 clustering are often evaluated visually and with expert understanding of the underlying ecology. The  
94 major concern with this approach is the inevitable influence of researcher bias on selecting an appropriate  
95 result (Handl et al., 2005). Even with dozens of cluster evaluation metrics available, they are rarely  
96 utilised as there is no single cluster evaluation index that can outperform others (Arbelaitz et al., 2013;  
97 Milligan and Cooper, 1985). There are two possible ways of evaluating the distribution of variables to  
98 clusters - internal and external (Šulc, 2016). External indices are used to evaluate supervised learning  
99 problems where the model solution is evaluated against the known solution. This allows the use of well  
100 known and understood metrics of clustering reliability such as accuracy, sensitivity and precision. Despite  
101 these methods requiring a prior known outcome, they are often used in the evaluation of new unsupervised  
102 learning approaches, even when by definition these methods have no known outcome. Instead internal

103 evaluation methods may be used to evaluate an unsupervised learning outcome (Arbelaitz et al., 2013).  
104 Internal indices have been developed to calculate the within- and between- cluster variability and select  
105 the optimal number of clusters (Borjesson et al., 2008). The number of functional groups (clusters) selected  
106 will affect ecosystem model outcomes and running time (Fulton et al., 2003), and choosing too few will  
107 mean that the functionality is not well represented, while choosing too many will over-fit the problem  
108 (Mason et al., 2003).

109 The aim of this research is to find a clustering method suitable for identifying functional groups of  
110 fish from nominal data. In this paper, we evaluate the utility of using hierarchical cluster analysis to  
111 find functional groups of fish from nominal traits. A good clustering result would find groups that are  
112 compact, well-separated, connected, and stable while still being ecologically relevant (Brock et al., 2008).  
113 Therefore, our focus is largely on evaluating results with internal cluster evaluation metrics, bootstrapping  
114 and visualisation.

## 115 MATERIALS & METHODS

116 Here we describe a step-by-step approach to derive functional groups from nominal traits by first creating  
117 a species by trait matrix (Part 1; Fonseca and Ganade, 2001) and classifying the groups via hierarchical  
118 cluster analysis (Part 2; Petchey and Gaston, 2002). Importantly, during the analysis stage we provide  
119 strategies for dealing with missing data, and selection of distance matrix and clustering algorithm. This is  
120 done by evaluating the compactness, separation and stability of group fits.

### 121 Part 1: Creation of the trait matrix

122 In this section we describe the steps used for creating a species · trait matrix as described by Fonseca and  
123 Ganade (2001). This methodology can be used for making a trait matrix for any group of species. Tasman  
124 Bay and Golden Bay (TBGB; co-ordinates:  $-41^{\circ}\text{E}$ ,  $-173^{\circ}\text{N}$ ) located on the north of New Zealand's South  
125 Island is used as a case study and we focus on fish (*Actinopterygii* and *Chondrichthyes*). TBGB is one of  
126 many areas used for commercial fish catches in the New Zealand Exclusive Economic Zone (EEZ). This  
127 region is characterised by its relatively shallow water habitat that has large ocean currents that enter this  
128 system from the Tasman sea bring nutrient rich cold water that makes the area highly productive. Large  
129 sheltered areas mean that this area is home to a diverse range of species, from small reef bound species to  
130 large migrating pelagic species.

#### 131 (i) Select the functional group to be defined

132 The type of functional group defined will be dependent on the ecosystem that is being modelled. Different  
133 ecosystems require different functions in order for their production to be exploited by its inhabitants  
134 (Fonseca and Ganade, 2001). For example, coral reef fishes need strong, sharp teeth in order to exploit  
135 polyps, while large pelagic species need to be fast moving in order to capture prey. Functional groups of  
136 species should be defined by how the species use their environment and its resources as ecosystem models  
137 attempt to model the entire process of an ecosystem spatially and temporally (Fulton et al., 2004). As we  
138 are modelling an open ecosystem, where species can enter and leave, it is important to try and capture  
139 some of the diversity of how species use an ecosystem daily, seasonally and yearly. The final groupings  
140 of species should exhibit similar responses to environmental conditions and have similar effects on the  
141 ecosystem processes (Fonseca and Ganade, 2001), though a good way to test these characteristics is yet to  
142 be found.

#### 143 (ii) Select species to include

144 The species selected to include should represent the taxonomy, time and space that the functional groups  
145 are trying to capture (Fonseca and Ganade, 2001). That is, species that rarely occupy the area of interest,  
146 or species with greatly differing biomasses should be included in the analysis. This is because including  
147 many species in functional groups better explains changes in the biodiversity of a given system (Naeem and  
148 Li, 1997). For this study, a comprehensive list of species of fishes from TBGB was made from the latest  
149 published account of trawl data (Stevenson and MacGibbon, 2015) and from published accounts of species  
150 known to inhabit TBGB (Roberts et al., 2015). While there are obvious functional differences between  
151 adults and juveniles of many species, that should be addressed and incorporated, such a delineation was  
152 beyond the scope of this project.

153 **(iii) Select functions of interest**

154 To avoid functional redundancy more functions can be selected to increase the chances of species having  
155 unique roles within the ecosystem, while ensuring that species who display the same traits across a number  
156 of functions truly belong to the same functional group. We selected four different functions to represent  
157 how the species of interest utilise their environment: diet, morphology, habitat use and life history traits  
158 (Villéger et al., 2017b; Gravel et al., 2016; Costello et al., 2015). Diet determines a species influence  
159 on other organisms in the environment and its position in the food web (Costello et al., 2015). Habitat  
160 preference allow us to understand how the different species might aggregate in the environment and  
161 can provide information about the likely lifestyle of the species (Chan, 2001; Vadas Jr and Orth, 1997).  
162 Morphology traits are important in defining the range of food sources, behaviour, adaptation and habitat  
163 use available to a certain species (Sibbing and Nagelkerke, 2000). Life history primarily reflected the  
164 reproductive strategies of the species which may be indicative of their abundance and resilience in the  
165 environment (Villéger et al., 2017b).

166 **(iv) Trait selection**

167 Traits should reflect the functions of interest. A literature review was conducted that identified 94 potential  
168 traits that could be recorded from fish species. As cost and time are often significant motivators for  
169 conducting research, it was a goal of this study to record functional trait information only from published  
170 resources or from photographs, rather than collecting and measuring specimens. We identified 40 traits  
171 that could be recorded without measuring species directly (Table S1). For some cases, variables that  
172 previously required a specimen to be measured were able to be categorised into nominal variables. For  
173 example, caudal peduncle aspect ratio was recorded as caudal fin shape. Where information differed  
174 ontogenetically within species, the information for adult females was recorded. The final list of recorded  
175 traits is provided in Table 1.

176 Morphology traits describe how species move around their environment and can potentially be used  
177 as an indicator of prey preferences (Albouy et al., 2011). Most of the traits recorded for morphology  
178 were determined from pictures of the species. Descriptions of the species fins were recorded either as  
179 their position on the body (pelvic), the shape of the fin (caudal) or the fin composition (soft ray or spines  
180 - dorsal). The shape of the caudal fin is important in determining the ability of a species to transition  
181 between vertical habitats (Bridge et al., 2016). The swimming mode of the species was recorded as either  
182 body caudal fin (BCF) locomotion or median paired fin (MPF) locomotion that is an indicator of the  
183 evasiveness of the food types targeted (Sfakiotakis et al., 1999; Webb, 1984b). The body form of the  
184 species was recorded as either fusiform, flat, cylindrical or compressed which is an indicator of how  
185 species acquire their food (Webb, 1984a). Eye position indicates the likely location of the species in  
186 the water column (Mindel et al., 2016). The spiny dorsal fin type may be an indicator of protection (i.e.  
187 from the number of spines - another recorded variable) but can also indicate the manoeuvrability of the  
188 species. The soft dorsal fin can help a fish to remain stable while swimming but is also able to generate  
189 thrust (Lauder and Drucker, 2004). Oral gape position can indicate feeding position in the water column  
190 (Albouy et al., 2011) and prey types that may be acquired (Zhao et al., 2014). Teeth shape indicate the  
191 type of prey consumed and the substrate on which a species may be feeding (Bellwood et al., 2014). Body  
192 length is an indicator of potential prey available and it correlates with size at maturity, fecundity, growth  
193 rate and longevity (Sibbing and Nagelkerke, 2000; Mindel et al., 2016). Physical protection was recorded  
194 as present or absent as an indicator of how difficult the species would be to use as prey (Reecht et al.,  
195 2013).

196 The life history traits selected primarily reflect the reproductive strategies for each of the species.  
197 Parental care (care, no care) was included as it can indicate where a species chooses to breed as well as  
198 the size and amount of the offspring (Franco et al., 2008). The spawning season and location (river, bay,  
199 ocean) were also recorded as it indicates when species would be expected to be found together and their  
200 potential seasonal movements. Gregariousness or schooling type was defined as solitary, facultative or  
201 obligative which help to explain how species aggregate and how often. Fish that are obligative schoolers  
202 (highly gregarious) tend to be preferred prey of large and fast predators (Spitz et al., 2014). Mortality  
203 and maximum age are indicators of population turnover rates and longevity and may also be an indicator  
204 of population size (Palomares and Pauly, 1998). Age or length at maturity affects the resilience of a  
205 population, as species that mature younger are more resilient (Froese and Binohlan, 2000). Number of  
206 eggs or brood size is an indicator of fecundity (Clavel et al., 2013). Spawning frequency was recorded  
207 as singular (semelparous), batch or serial spawning and annual which can indicate stability of stocks

**Table 1.** Diet, habitat and morphology traits included in the analysis along with trait type, function, categories, percent missing and references.

Variable	Function	Data type	Categories	Missing	Reference/s
Diet	Diet	Nominal	Omnivore; Invert feeder, Piscivore, Herbivore, Gelatinous inverts	0%	Villegier et al. (2017)
Trophic level	Diet	Continuous/Discretized	Low (0-3); Medium (3-3.5); High (3.5-4); Very high (4+)	0%	FishBase (2016)
Common maximum depth (m)	Habitat	Continuous/Discretized	Reef (0-20.1); Shallow (20.2-54.6); Ocean (54.7-148.4); Deep (148.5+)	0%	New
Maximum depth (m)	Habitat	Continuous/Discretized	Reef (0-20.1); Shallow (20.2-54.6); Ocean (54.7-148.4); Deep (148.5-403.4); Bathy (403.4+)	0%	New
Temperature preference	Habitat	Nominal	Deep, Temperate, Subtropical, Tropical	0%	FishBase (2016)
Vertical habitat	Habitat	Nominal	Reef, Pelagic, Demersal, Benthopelagic, Bathypelagic, Bathydemersal	0%	FishBase (2016)
Horizontal habitat	Habitat	Nominal	Coast, Neritic, Ocean	0%	FishBase (2016)
Caudal fin shape	Morphology	Nominal	Forked, Rounded, Truncated, Emarginate, Heterocercal, Continuous, Lanceolate	0%	Fishes of New Zealand
Swimming mode	Morphology	Nominal	Body caudal fin (BCF), Median paired fin (MPF)	0%	Villegier et al. (2017); Sfakiotakis et al. (1999)
Body form	Morphology	Nominal	Compressed, Cylindrical, Eel, Flat, Fusiform	0%	Villegier et al. (2017); Sfakiotakis et al. (1999)
Eye position	Morphology	Nominal	Mid, Side, Top	0%	Mindel et al., (2016)
Oral gape position	Morphology	Nominal	Subterminal, Terminal, Hyper-protusable, Inferior, Snout projecting, Lower jaw projecting, Tubular	0%	Gravel et al. (2016); Sibbing & Nagelkerke (2001)
Maximum length (cm)	Morphology	Continuous/Discretized	Small (0-20.1); Medium (20.2-54.6); Large (54.7-148.4); Very large (148.5+)	0%	Gravel et al. (2016); Sibbing & Nagelkerke (2001)
Reproductive strategy	Life history	Nominal	Oviviparous, Ovoviparous, Viviparous	1.7%	Franco et al. (2008); Bremner et al. (2006)
Sexual differentiation	Life history	Nominal	Gonochoristic, Hermaphrodite	1.7%	Bremner et al. (2006)
Migration	Life history	Nominal	Anadromous, Catadromous, Oceanic, None	12.1%	Spitz et al. (2014)
Parental care	Life history	Nominal	None, Paternal, Resource defence polygeny (RDP), Sheltered	2.6%	Gravel et al. (2016); Franco et al. (2008)
Egg attachment	Life history	Nominal	Pelagic, Benthic, Adhesive, None	7.8%	Gravel et al. (2016); Franco et al. (2008)
Reproduction location	Life history	Nominal	Bay, Ocean, River	23.3%	Franco et al. (2008)
Gregariousness/Schooling type	Life history	Nominal	Facultative, Obligatory, Solitary	18.1%	Spitz et al. (2014)
Population doubling	Life history	Nominal	High, Medium, Low, Very low	12.1%	FishBase (2016)

208 between years, where species that spawn more often tend to have more stable populations (Longhurst,  
 209 2002). Fish that provide parental care or give birth to live young (viviparous) tend to give birth to fewer,  
 210 larger offspring, often in more sheltered habitats such as estuaries.

211 Habitat traits are important in defining how a species uses their environment. As we focused on a  
 212 small ecosystem the habitat variables of a given species must match the available habitat of that ecosystem.  
 213 We included the minimum and maximum known depth of the species as TBGB is a relatively shallow bay  
 214 (max depth 200m). Knowing the vertical space that the species occupy informs us of potential intraspecific  
 215 competition (Munday et al., 2001). We included the preferred temperature gradient (tropical, subtropical,  
 216 temperate or deep) as temperature is an important indicator of how species use the ecosystem (Malavasi  
 217 et al., 2004). Horizontal habitat (coastal, neritic or ocean) was used as another indicator of how species  
 218 may group together in similar habitats.

219 Diet traits allow us to understand a species position within a food web. Diet can be recorded in a  
 220 number of ways, but for our purposes we sought a simple classification of diet. Therefore we have two  
 221 diet variables only; diet category (omnivore; invertivore, piscivore, herbivore and gelatinous invertebrate  
 222 feeders) and trophic level (obtained from FishBase for consistency).

### 223 **Data sources**

224 Functional traits of species were sourced primarily from FishBase - a global information system on fishes  
 225 (Froese and Pauly, 2017) and from 'The fishes of New Zealand' - a comprehensive text with citations of  
 226 all known fish species in New Zealand (Roberts et al., 2015). Additional trait data were obtained from a  
 227 combination of published research and reports. When data was obtained from sources other than FishBase  
 228 or Roberts et al. (2015) the source is referenced. To obtain traits from FishBase we utilised the R package

229 rfishbase (Boettiger et al., 2012).

## 230 **Part 2: Statistical analysis**

231 In this section we describe the steps taken to analyse and group the data. Our approach differs to traditional  
232 functional group analyses as we use categorical (nominal) data. In order to use nominal data we must  
233 ensure we have a complete dataset (no missing values) and our continuous variables must be discretized.  
234 These two steps are detailed in our data preparation stage, followed by a description of the distance  
235 matrices available for nominal data. We then describe some linkage options and finally detail the data  
236 evaluation stage. Our approach utilises the R package *nomclust* which is designed exclusively for  
237 clustering observations with nominal variables (Šulc and Řezanková, 2015; Team, 2018).

### 238 **(v) Data preparation**

239 Only 22 of the 40 recorded traits had less than 25% missing data and were retained for analysis. 25% was  
240 selected as the cutoff as the accuracy of imputed datasets is seriously degraded above 20-25% for small  
241 datasets (Clavel et al., 2014). Distance matrix calculations require complete information, therefore we  
242 choose to impute the missing data in these 22 variables. Numerous methods exist for imputing data, and  
243 many of these have been examined for their precision in imputing continuous variables (Penone et al.,  
244 2014; Clavel et al., 2014). What is unknown is how well these packages perform for nominal variables.  
245 To find the most accurate imputation method for nominal data we used three different approaches (all  
246 implemented in R packages): random forests implemented in *missForest* (Stekhoven and Bühlmann,  
247 2012), multiple correspondence analysis (MCA) implemented in *missMDA* (Josse and Husson, 2016)  
248 and polytomous logistic regression implemented in *MICE* (van Buuren and Groothuis-Oudshoorn, 2011)  
249 (described in Data S1). We also selected a simple imputation method using the mode value for each  
250 variable to serve as a baseline. In the mode replacement method, all missing values are replaced with the  
251 same value that is most frequently occurring. This method was used to compare against other imputation  
252 methods that use more information to inform the imputation (Taugourdeau et al., 2014). To test the  
253 accuracy of the different imputation methods we first selected all 13 variables from the database with  
254 complete information (Table 1). For each method, we ran a simulation in which data were randomly  
255 deleted and imputed 100 times. The probability of the method correctly imputing values were tested  
256 over a range of proportions of missing data ranging from 0.05 to 0.45, increasing in steps of 0.05. The  
257 final accuracy was calculated as the number of incorrect imputations divided by the number of possible  
258 imputations.

259 Four of the 22 trait variables were continuous and were discretized to turn them into categorical  
260 variables. It was a goal of the discretization process to maintain the underlying distribution of the data  
261 while creating similar number of categories in each variable (Teletchea et al., 2009). Each continuous  
262 variable was plotted on a histogram and bins were selected such that the distribution of the variable was  
263 maintained using four or five bins (Figure S1). The final categories for each continuous variable and their  
264 values are reported in Table 1. The final trait matrix consisted of  $m = 22$  traits and  $n = 116$  fish species.

### 265 **(vi) Distance matrices**

266 Hierarchical clustering methods utilise distance matrices to make groups. A distance matrix in this context  
267 is a measure of pairwise similarities or dissimilarities between species (rows) based on their trait values  
268 (columns). There are a wide range of distance matrices and clustering methods available to cluster nominal  
269 data, and the combinations selected will influence the resulting groups. Having nominal data prevents  
270 us from using some measures, such as Euclidean distances, as they assume an inherent ordering within  
271 variables. For binary data treating data (0 or 1) as continuous is a valid measure of difference, but for  
272 variables with more than two categories the various distances between values do not represent meaningful  
273 differences. Boriah et al. (2008) evaluate 14 alternative measures of calculating distance matrices for  
274 nominal data and here we evaluate five: simple matching (SM - as in Gower's distance), Eskin, Lin,  
275 inverse frequency of occurrence (IOF), and Goodall's, available in the R package *nomclust* which are  
276 described in Data S2. The other measures available are derivatives of these measures and were not shown  
277 to improve performance in preliminary analyses. Briefly, the five distance matrices are described. The SM  
278 distance, which is the simplest approach to creating a distance matrix, awarding 1 to observations that are  
279 the same and 0 if not. This is the approach used for Gower's similarity measure of nominal data (Gower,  
280 1967). Eskin's distance, which uses a SM criteria that gives more weight to mismatches on variables  
281 that have more categories (Eskin et al., 2002). The inverse occurrence frequency (IOF) distance has



282 the same approach as Eskin but gives less weight to mismatches on variables that have more categories  
 283 (Sparck Jones, 1972). This uses the absolute frequencies of observed categories. Goodall's distance,  
 284 which when comparing two observations of a given variable, takes into account relative frequencies of  
 285 categories (Goodall, 1966). A similarity value is assigned based on the normalised similarity between  
 286 the two observations, where the similarity value is higher if a category occurs infrequently. This method  
 287 takes into account that individuals attributes occur stochastically and independently in a population. Lin's  
 288 distance is an information theoretic definition of similarity based on relative frequencies (Lin, 1998).  
 289 Matches are given higher weightings when they occur infrequently, and conversely mismatches are given  
 290 higher weightings when they occur infrequently.

### 291 (vii) Clustering methods

As we do not know the number of functional groups in the ecosystem a priori, we used hierarchical clustering to visualise group association given our chosen distance metric. Hierarchical clustering first places all  $n$  objects in  $n$  separate single member clusters, and larger clusters are formed by sequentially joining first individual observations and then groups of observations until at last all observations are in a single group. The closeness of pairs of observations or groups of observations to another are determined by a measure of distance calculated in the preceding step. In linkage, all pairwise inter-cluster dissimilarities are calculated. The pair of clusters that are least dissimilar (that is, most similar) is identified and these two clusters are fused. Once observations or clusters are joined to a group they remain as a part of that cluster for the remainder of the analysis. There are a number of linkage methods that can be used for this type of data and here we explore three methods available in the R package `nomclust` (Blashfield, 1976). To describe the linkage methods we use the following notation:  $D(A, B)$  is the distance between clusters  $A$  and  $B$ , which have sizes  $n_A$  and  $n_B$  respectively. In single linkage (minimising inter-cluster dissimilarity), the dissimilarity between two clusters is the smallest of all pairwise distances between the observations in the two clusters:

$$D(A, B) = \min[d(x, y) : x \in A, y \in B] \quad (1)$$

In complete linkage (maximises inter-cluster dissimilarity), the dissimilarity between two clusters is the largest of all pairwise distances between the observations in the two clusters:

$$D(A, B) = \max[d(x, y) : x \in A, y \in B] \quad (2)$$

In average linkage, the dissimilarity between two clusters is the average of all pairwise distances between observations in the two clusters:

$$D(A, B) = \frac{1}{n_A n_B} \sum_{x \in A} \sum_{y \in B} d(x, y) \quad (3)$$

### 292 (viii) Selection of distance matrices, clustering methods and number of clusters

Evaluating clustering outputs can occur in two ways; external, where the resulting clusters are compared against known groupings (as in supervised learning), or internal evaluation, where some metric (there are many) is used to evaluate cluster separation and compactness. Since in our case the true groupings are unknown only internal evaluation is considered. To select the best distance matrix and clustering method for our data we utilised internal evaluation measures available from `nomclust` (Šulc and Řezanková, 2015). The within-cluster entropy coefficient (WCE) is a measure of compactness which evaluates the variability of each cluster by calculating a measure of normalised entropy (the number of variables that have the same categories from each of the variables evaluated) (Šulc, 2016). WCE is measured from 0 to 1, where a lower value indicates intra-cluster homogeneity. Due to the way that these values are calculated they will generally always improve by adding clusters to the solution because the within cluster variability decreases:

$$WCE(k) = \sum_{g=1}^k \frac{n_g}{n \times m} \sum_{c=1}^m \left( - \sum_{u=1}^{K_c} \left( \frac{n_{gcu}}{n_g} \ln \frac{n_{gcu}}{n_g} \right) \right) \quad (4)$$

293 Where  $n$  is the total number of objects (species),  $m$  is the number of variables (traits),  $n_g$  is the number  
 294 of objects in the  $g^{th}$  cluster ( $g = 1, \dots, k$ ) and  $n_{gcu}$  is the number of objects in the  $g^{th}$  cluster by the  $c^{th}$   
 295 variable with the  $u^{th}$  category ( $u = 1, \dots, K_c$ ).

To select the number of groups we use the pseudo F coefficient based on the entropy (PSFE), a measure of separation (Šulc, 2016). The PSFE is a measure of entropy of the between- and within-cluster variability adjusted for the number of clusters and number of objects in the cluster where a higher value indicates a better grouping:

$$PSFE(k) = \frac{(n-k)[nWCE(1) - nWCE(k)]}{(k-1)nWCE(k)} \quad (5)$$

where  $n$  is the number of observations and  $k$  is the number of clusters,  $nWCE(1)$  is the variability in the whole dataset, and  $nWCE(k)$  the within-cluster variability in the  $k$ -cluster solution.

Therefore, a more informative measure of performance is the degree of improvement with increasing number of clusters. Results from these measures are therefore presented as the difference between the  $k_{th}$  cluster and the  $k_{th-1}$ . Equivalent measures of all the aforementioned evaluation techniques are available in *nomclust* using the Gini coefficient instead of entropy and are provided in Figure S2 as a reliability measure of our results.

We use t-Distributed Stochastic Neighbour Embedding (t-SNE) (Van Der Maaten, 2014) to construct a two-dimensional scatter plot in which each point represents a species. t-SNE minimises the distance between two distributions, one that was derived from a similarity matrix, and one that is derived from embedding the same matrix. To do so, a principal components analysis (PCA) is constructed from a dissimilarity matrix which allows species with similar trait profiles to be mapped in two-dimensions. These graphs provide a visual demonstration of similar species by the closeness of their points, and we use these graphs to evaluate our final group clustering. After the number of clusters was selected, we evaluated a cluster-wise measure of cluster stability through a bootstrapping procedure available in the R package *fpc* (Hennig, 2013). The `clusterboot` function draws a sample of size  $N$  from the original data set, computes the clustering using partitioning around medoids (PAM), then calculates the maximum Jaccard coefficient between the most similar cluster in the bootstrapped data sets (Hennig, 2007). PAM is an agglomerative clustering approach that moves a pre-defined number of centres, here 3 and 9, around a group of data to find the total minimum distance between the centres and the observations (Brock et al., 2008). This is repeated 100 times and an average Jaccard coefficient is found for each of the clusters which is representative of cluster stability. The more stability in clusters the less deviation evident in the Jaccard coefficient and as such the results are plotted as error bars. Finally, the adjusted Rand index (Hubert and Arabie, 1985) is used to compare the partitioning of groups for the different combinations of distance matrix and cluster algorithm for 3, 5, 7 and 9 cluster groups. The index can take a minimum of 0 (groups are completely random, may be negative if the index is less than the expected index), or 1 (each group contains the same observations).

The results of these analyses are discussed in terms of connectedness, compactness, separation and stability. Compact groups are those which minimise the spread of observations within a cluster and are assessed with the WCE and visually with the t-SNE graphs. Separation refers to the between cluster distances, which ideally should be maximised and are assessed by the PSFE, and visually with the t-SNE graphs. A well connected cluster is one where an observations nearest neighbour is from its own cluster and is assessed solely by visualisation. Stability is assessed via the results of the bootstrapping procedure.

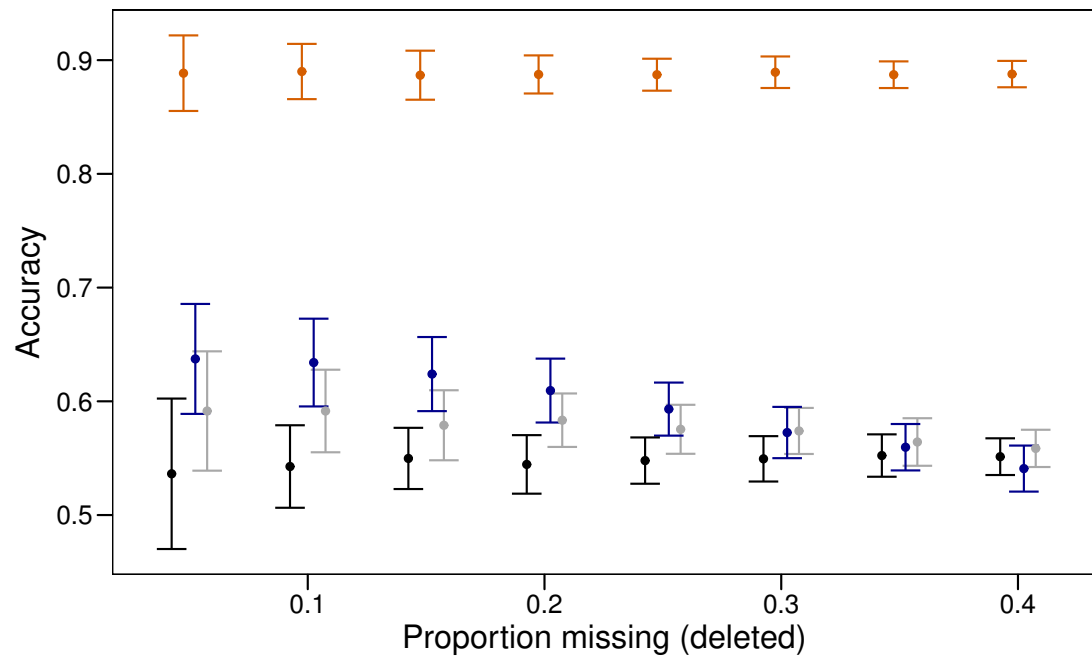
## RESULTS

### Imputing missing data

Three imputation methods were compared with a baseline method of using the mode to replace the randomly deleted missing value. The polytomous regression from the package *MICE* clearly outperformed all other methods, imputing data correctly between 85 and 92% of the time (Figure 1). Both *missForest* and *missMDA* performed better than imputing the data from the mode, but across the range of proportions of missing data none were significantly better than the baseline. Over the range of missing data proportions as the amount of missing data increased, the variability in imputed accuracy decreased.

### Cluster evaluation

The values of PSFE changed depending on the combination of number of clusters, distance matrix and linkage method selected (Figure 2). The single linkage method was clearly the poorest performer with all but one PSFE score below that of the other linkage methods. The PSFE values for the complete

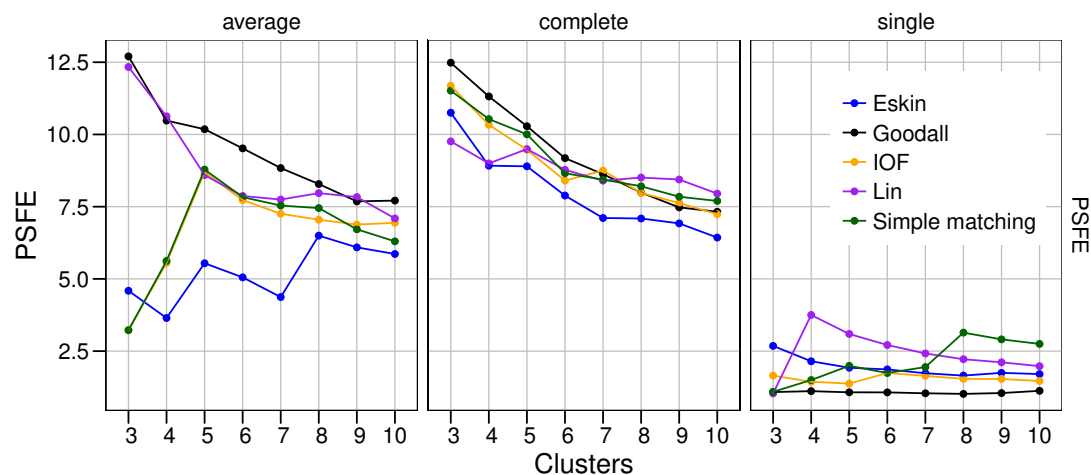


**Figure 1.** Proportion of values imputed correctly (accuracy) and 95% confidence interval for different imputation methods across varying amounts of missing data. The four imputation methods displayed are; MICE (orange), missForest (blue), missMDA (grey) and mode (black). Bars are jittered for clarity.

341 linkage method tended to decrease with an increasing number of clusters, with all distance matrices  
 342 showing that three is the optimal cluster number. In contrast, distance matrices under the average linkage  
 343 method showed a variety of patterns (Figure 2). Only two distance matrices selected three as the optimal  
 344 cluster size, two selected five and one (Eskin) showed eight clusters was optimal. The Goodall and Lin  
 345 distance matrices appeared to have the highest PSFE scores across the range of cluster sizes using the  
 346 average linkage method, and the values decreased as the number of clusters increased. Distance matrices  
 347 calculated using Eskin produced the lowest PSFE scores for average and complete linkage methods.  
 348 While there is no combination of distance matrix and linkage method that is uniformly superior to the  
 349 others, three clusters seem to fit best under a variety of conditions, as shown by high PSFE scores. In  
 350 addition, Goodall and Lin distance matrices appear to perform slightly better, particularly for the average  
 351 linkage method.

352 Similar to PSFE, the combination of distance matrix and linkage method selected impacted the overall  
 353 WCE score (Figure 3). A lower value of WCE indicates more intra-cluster homogeneity. Using this metric,  
 354 the within cluster variability continues to decrease across all numbers of clusters for all combinations  
 355 of linkage method and distance matrix. Under average linkage, the Goodall and Lin distance matrices  
 356 demonstrated the lowest scores across cluster numbers, indicating the lowest within cluster variance. For  
 357 complete and single linkage there was no clear distance matrix that performed better. The WCE score will  
 358 always decrease as the number of clusters increase, so is not robust to clustering complexity. We therefore  
 359 examined the magnitude of improvement of the WCE score with increasing the numbers of cluster.

360 Figure 4 presents the WCE and PSFE scores simultaneously. This figure attempts to extract some of  
 361 the more complex relationships underlying the clustering results, but must be interpreted with the previous  
 362 two figures (Figure 2 and 3). The height of the bars represents the difference of the WCE between the  
 363 labelled cluster ( $x$ -axis) and the previous clustering. A higher value in this figure represents a higher  
 364 decrease in within-cluster variability. The largest differences in WCE correspond to the highest PSFE  
 365 score (Figure 4; red bars). Distance matrices calculated using Goodall and Lin show that a lower number  
 366 of clusters is generally a better solution. Eskin now prefers a large number of clusters (8-10), while using  
 367 the IOF and SM distance matrices it was found that 5-8 clusters is a better solution under the average  
 368 linkage method. All distance matrices have the highest values for the three cluster solution under complete  
 369 linkage.



**Figure 2.** Evaluation of the optimal number of clusters using the pseudo F coefficient based on the entropy (PSFE). All five distance matrices (coloured lines) tested are displayed across three clustering algorithms (facets).

370 Hierarchical clustering is usually presented as a dendrogram, but due to the large number of species in  
 371 the dataset we take advantage of dimensionality reduction techniques to plot the clusters in two dimensions.  
 372 Well formed clusters are those that distinct from other clusters and are compact. Here we present the  
 373 resulting clusters for a subset of distance matrices that have low WCE values for the cluster numbers with  
 374 high PSFE scores. Only the average linkage method is shown here as complete returned similar results,  
 375 and SM did not perform well on any metric.

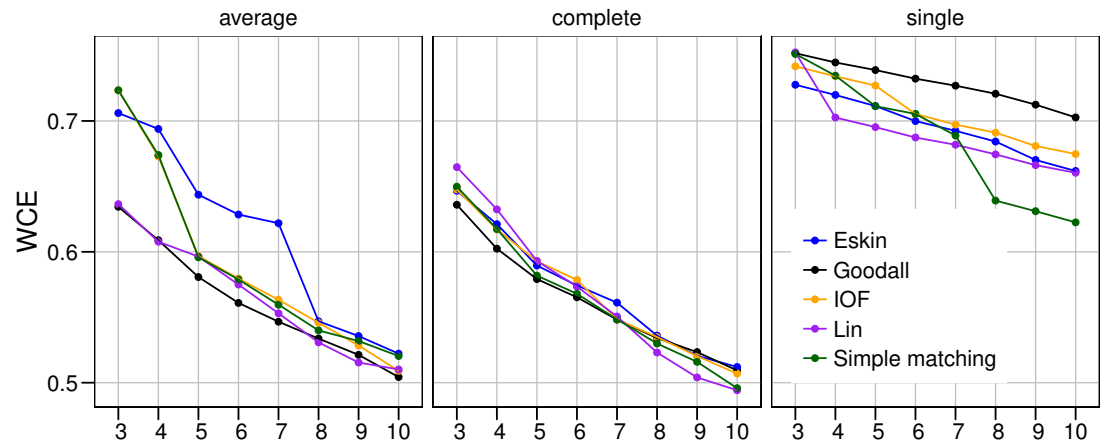
376 For clarity, we use t-SNE to plot the resulting groups in two dimensions (Figure 5). The most  
 377 connected groups are apparent when using a Lin or Goodall distance matrix with average linkage method  
 378 and three clusters. These clusters are relatively stable (Figure 6), but only one group (sharks - blue points)  
 379 is well separated and compact (Figure 5: Lin-3, Goodall-3). Increasing from three clusters does not seem  
 380 to increase the separation or compactness of groups. Instead, more small groups appear, scattered through  
 381 other groups, suggesting a loss of connectedness. In comparison, Eskin does a good job compacting  
 382 similar observations, and these groups appear more cohesive as the number of clusters increase. Similarly,  
 383 IOF creates more connected groups as the number of clusters increase, but again, only one group is  
 384 separated and compact. This is supported by the stability analysis which show that IOF and Eskin have  
 385 more stable clusters when a larger number is selected than Lin or Goodall (Figure 6).

386 Using the Rand index we compared distance matrix and linkage method combinations for three and  
 387 nine clusters. This confirmed that Goodall and Lin were consistently producing similar results for three  
 388 clusters with adjusted Rand index values of between 0.52 and 0.77. And IOF and Eskin produced similar  
 389 results with nine clusters with values between 0.52 and 0.89 (with the exception of IOF and complete  
 390 linkage).

391 Generally, within this dataset three main groups form which correspond to: reef and demersal fish  
 392 (including skates and rays), large pelagic and deep-sea fish, and sharks (Figure 5). The most obvious  
 393 distinction in these graphs is the group of 10-15 observations that always separate from the other clusters,  
 394 which correspond to the sharks. The four shark species that tend to not associate with the rest of the  
 395 cartilaginous fishes are the skates and stingrays, which cluster closely to the flatfish. As the number  
 396 of groups increases smaller groups tend to form, but these groups are highly unstable and are highly  
 397 dependent on the distance matrix selected.

## 398 DISCUSSION

399 Clustering species based on their traits theoretically allows functional groups to form. This is particularly  
 400 difficult to test, as it is unknown how many functional groups exist within a given ecosystem, nor which  
 401 traits are needed to find the functional groups (Bremner et al., 2006b). Cluster analysis allows for the  
 402 exploration of underlying data patterns when its presence and/or structure are unknown, but it lacks

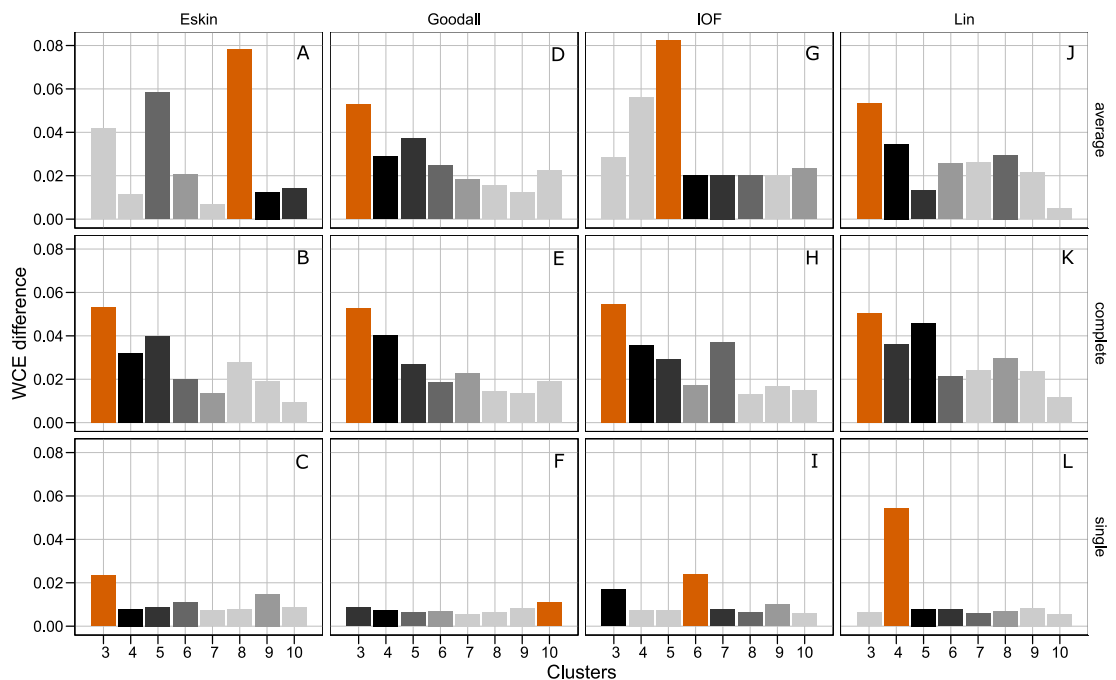


**Figure 3.** Evaluation of the optimal number of clusters using the within-cluster entropy coefficient (WCE). All five distance matrices (coloured lines) tested displayed across three clustering algorithms (facets).

403 an agreed method of evaluation. Using nominal trait data, we explored how changing distance matrix,  
 404 linkage method and number of clusters impacts the formation of functional groups of marine fish. We  
 405 utilise internal evaluation metrics available in the package `nomclust` to assess connectedness, separation  
 406 and compactness of the resulting groups (Handl et al., 2005), and we bootstrap the data to evaluate its  
 407 stability (Hennig, 2013). Our methodology demonstrates that the separation, compactness, and stability  
 408 of functional groups are dependent on the choice of distance metric, linkage method, and number of  
 409 clusters. While this may have been an intuitive conclusion (Gitay and Noble, 1997), our analysis provides  
 410 an indication of the level of variation that arises from these choices. This variation can be assessed by  
 411 comparing the final clustering results to one-another, revealing that the most similar clustering achieved a  
 412 Rand Index value of up to 0.89 (with 1 being a perfect match) and the most dissimilar clustering methods  
 413 scoring negative values.

414 Using 22 nominal traits representing diet, habitat, morphology and life history we explored the  
 415 combinations of distance matrix and linkage method that would best capture the structure in our dataset.  
 416 This combination revealed that there are probably three major groups of fish that exist in Tasman Bay  
 417 and Golden Bay. While this may appear to be a very simplistic summary of a complex ecosystem, using  
 418 these groupings in ecosystem models would help to increase efficiency in modelling time and output  
 419 interpretation. However, if more detailed analysis is warranted, because there is good evidence that a  
 420 greater number of functional groups exists in the system, or it is necessary to represent more diversity in  
 421 an ecosystem, then a different combination of distance matrix and linkage method would be required. For  
 422 this dataset a larger number of clusters is more accurately represented by Eskin distance metrics and the  
 423 average linkage method.

424 The separation of clusters in this analysis was evaluated by PSFE. Separation is a measure of distance  
 425 between clusters, therefore can be used to select the number of clusters. PSFE indicated that when using  
 426 the complete linkage method for all distance matrices investigated that three was the optimal number of  
 427 clusters, while under the average linkage method there was more variability in the number of clusters  
 428 selected, ranging from three to eight (Figure 2). The single linkage method tended to produce very low  
 429 values of PSFE, indicating less separation and overall a poor fit. Compactness was assessed by the WCE  
 430 which indicates the within-cluster homogeneity. Raw values of the WCE under the average linkage  
 431 method revealed that Goodall and Lin tended to have the lowest values across different cluster sizes, thus  
 432 had the most compact groups. Using the complete linkage method it was unclear which distance matrix  
 433 was performing best. The single linkage produced higher WCE scores (more variance) across the range of  
 434 distance measures, and in combination with the low PSFE scores was deemed a poor performer, therefore  
 435 was not considered for further analysis. As expected, the WCE decreased (indicating lower variance  
 436 within clusters) as the number of clusters increased, therefore this is not a good indicator of fit. Instead,

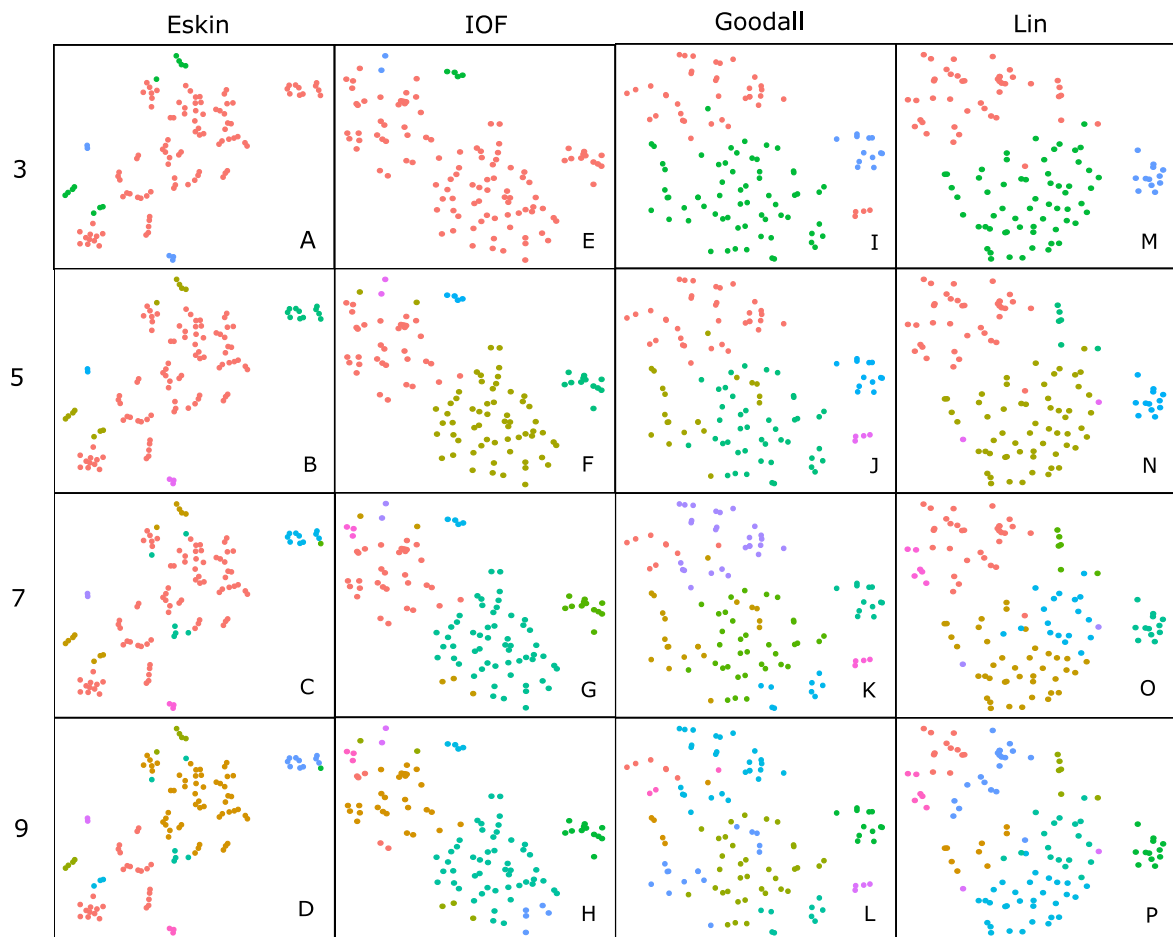


**Figure 4.** Evaluation of the optimal number of clusters using the difference between the  $k_{th}$  cluster and  $k_{th-1}$  of the WCE scores. The red bar corresponds to the highest PSFE score for that combination of distance matrix and linkage method indicating the optimal number of clusters. The black bar is the second highest score and colour gradient lightens as the PSFE scores lower (indicating a poor fit). A-C: WCE difference results for Eskin distance matrix with average, complete and single linkage; D-E: WCE difference results for Goodall distance matrix with average, complete and single linkage; G-I: WCE difference results for IOF distance matrix with average, complete and single linkage and J-L: WCE difference results for Lin distance matrix with average, complete and single linkage.

437 we explored the the difference in WCE score across number of clusters (Figure 4). This revealed that in  
 438 most cases the largest decrease in WCE (between cluster sizes) corresponded to the highest PSFE score.  
 439 This approach allowed us to see more clearly which combination of distance matrix and linkage method  
 440 was fitting our nominal data set best.

441 We used bootstrapping to assess cluster stabilities, where observations were re-sampled with replace-  
 442 ment and clustered repeatedly, with the Jaccard coefficient extracted after each clustering (Hennig, 2007).  
 443 A stable cluster is more likely to remain unchanged in composition (contain the same observations after  
 444 each bootstrap) during re-sampling. There was no clear pattern in stability between number of cluster or  
 445 distance matrices, but generally three clusters were the most stable, and had the lowest variation (Figure  
 446 6). This was expected as some species had more traits in common than others, making it more likely for  
 447 them to always be placed in the same group (less likely to change groups during re-sampling).

448 A good indication that true structure has been found in a dataset is when methods align in agreement  
 449 of cluster assignment (Handl et al., 2005). Here Goodall and Lin agree across a number of measures for a  
 450 low number of clusters (three), while Eskin and IOF demonstrate agreement when the number of clusters  
 451 selected increases (more than 5). This indicates that the different distance matrices are able to identify  
 452 different underlying data structures. The IOF or Eskin are better choices when increasing the number of  
 453 clusters, as the groups created are more connected than for other distance matrices, while Goodall and  
 454 Lin are better with fewer clusters as the connectivity and separation are better (Figure 4). Two different  
 455 measures matching may be a good indication of fit, however, if the methodologies are developed from the  
 456 same theory, then it would be expected that they would find the same result (Handl et al., 2005). With this  
 457 in mind, our findings are again supported as the matching methods have different fundamental approaches  
 458 (Data S2). Goodall and Eskin aim to weight values higher that match infrequently, while IOF and Lin  
 459 give greater weight to values that match frequently, and lower weight to infrequent matches (Boriah et al.,  
 460 2008). All of these measures have been shown previously to perform well on different datasets in different

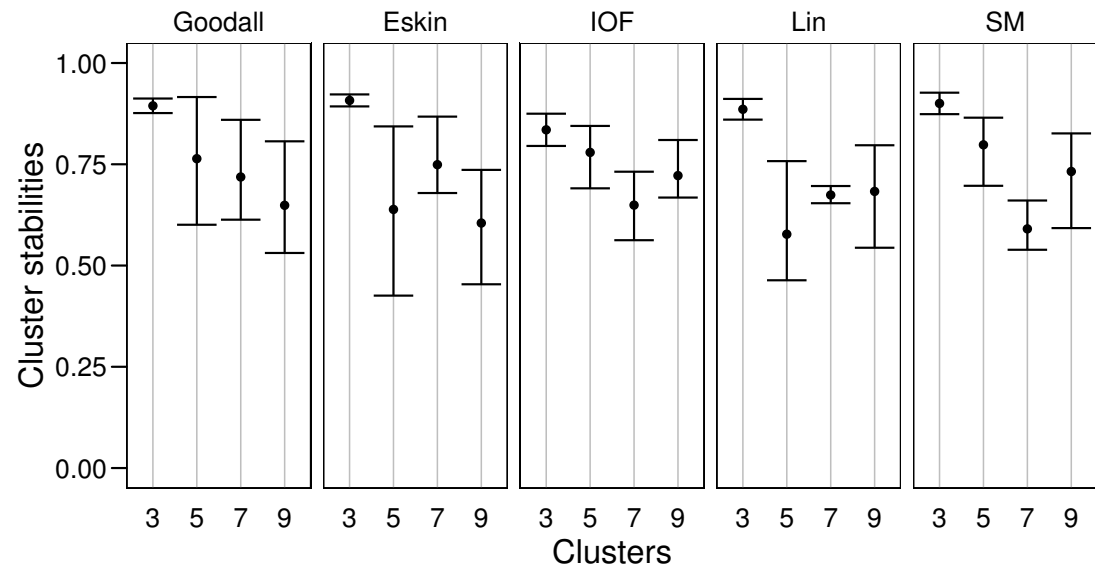


**Figure 5.** Clustering results using the average linkage method for four distance matrices (columns) for four different numbers of clusters (rows) displayed in two dimensions as the result of t-SNE. Colours represent the different groups found with hierarchical clustering. A-D: t-SNE clustering for Eskin linkage method with 3, 5, 7 and 9 clusters; E-H: t-SNE clustering for IOF linkage method with 3, 5, 7 and 9 clusters; I-L: t-SNE clustering for Goodall linkage method with 3, 5, 7 and 9 clusters and M-P: t-SNE clustering for Lin linkage method with 3, 5, 7 and 9 clusters

461 conditions (Šulc, 2016), emphasising the need to test a range of methodologies when clustering ecological  
 462 data.

463 The three groups that emerged from the analysis were reef and demersal fish (including skates and  
 464 rays), large pelagic and deep-sea fish and sharks. This finding contrasts with previous investigations where  
 465 a greater number of functional groups were found from fewer species (Córdova-Tapia and Zambrano,  
 466 2016; Reecht et al., 2013). Increasing the number of clusters may highlight different functional groups,  
 467 but as discovered by Córdova-Tapia and Zambrano (2016) this tends to result in groups occupied by a few  
 468 or single species. In a complex ecosystem such as this, we might expect to see much more separation  
 469 between the species, particularly if the traits selected truly represented different functions. Instead, we  
 470 find that across a range of distance matrices and linkage methods that just three groups continue to emerge.  
 471 The first distinct group is the sharks. These separate out first, and remain separated as the numbers of  
 472 clusters increase. The next two groups that commonly form roughly correspond to deep-sea and pelagic  
 473 fish, then reef and demersal fish (with skates and rays). The lack of separation of the groups may suggest  
 474 that we have not collected enough information about the species to robustly separate further groups. To  
 475 get a true representation of the functioning of ecosystems it is important to collect large numbers of traits  
 476 to predict functional groupings (Sibbing and Nagelkerke, 2000; Bremner et al., 2006b).

477 Mason et al. (2003) highlights the method used to classify functional groups as one of the three major  
 478 challenges of creating functional groups. While we attempted to resolve a number of issues with finding  
 479 functional groups from traits, there are some significant limitations that require investigation. To test our



**Figure 6.** Mean and standard deviation of the bootstrapped (n=100) Jaccard distance measure from PAM clustering for five nominal distance matrices across four cluster sizes.

480 methodology, we created a species by trait matrix with nominal trait data that could be extracted from the  
 481 literature or photographs. While obtaining trait information from published sources and distinctive features  
 482 from photographs can provide some information of how species use their environment, this strategy  
 483 cannot compensate for the rich data that can be collected from measuring species directly (Sibbing and  
 484 Nagelkerke, 2000). Inevitably, traits of both continuous and nominal types will be required and strategies  
 485 for how to analyse them. The problem of how to handle mixed data is yet to be resolved, particularly  
 486 as in many distance matrices nominal variables tend to have a higher influence on the similarity matrix  
 487 than continuous variable because they produce higher contrasts (Mirkin, 2012). Future analyses should  
 488 investigate using mixed (continuous and nominal) data to cluster functional groups.

489 As yet, there is no agreement on the set of functional traits to use that will provide meaningful  
 490 functional groups for fish, though various suggestions have been made (Sibbing and Nagelkerke, 2000;  
 491 Villéger et al., 2017a; Gravel et al., 2016). While an exhaustive list of functional traits can be provided  
 492 to assess their importance, a clustering model will try to include all of the variables provided, whether  
 493 they are important or not and there is going to be missing data. Here, we have used a combination  
 494 of dropping variables and imputing missing data. There is much support for imputing ecological data  
 495 (Nakagawa and Freckleton, 2011), but usually only for small amounts (Clavel et al., 2014). An important  
 496 aspect of functional group analysis that needs to be explored further is the impact of removing traits from  
 497 the analysis. Ideally, sensitivity analyses would be conducted to investigate their overall impact. We  
 498 treated our traits as nominal in order to equally weight all variables equally (Mason et al., 2003). This  
 499 approach will inevitably cause the loss of some information, as some traits were ordinal and some traits  
 500 contain more information than others. Using nominal data may limit the explanatory value of the trait by  
 501 excluding detailed information that continuous data can provide (Schleuter et al., 2010), as we have done  
 502 by discretizing some traits. Moving forward, it is likely that more traits are needed, and an assessment of  
 503 their importance to predicting group associations. One solution may be to use bi-clustering that is able to  
 504 perform dimensionality reduction by clustering traits, while simultaneously clustering species (Fernández  
 505 and Pledger, 2016).

## 506 CONCLUSIONS

507 Our results demonstrate that the best clustering solution for our data is three clusters using the Goodall  
 508 or Lin distance matrix with the average linkage method. If a larger number of clusters is the preferred  
 509 outcome, then the Eskin distance matrix with average linkage method should be used. While this result is  
 510 appropriate for this particular dataset, the results may of course change for different data. It is not correct



511 to assume that any combination of distance matrix and linkage method will be informative, nor that the  
512 combination used by a previous study is a good fit for your data. Instead, data exploration and evaluation  
513 analyses, such as those explored in this paper, must be employed. Not exploring the available options may  
514 lead to not finding a data structure when there is one, or randomly finding a structure among the noise  
515 when no clusters truly exist (Handl et al., 2005). This is because clustering algorithms are biased towards  
516 the properties on which they are built. Robust detection of genuine underlying structure requires that  
517 multiple algorithms find the same solution.

518 Deriving functional groups is an important process in developing our understanding of ecosystems. The  
519 goal of creating functional groups is to classify the species found in a given ecosystem into representative  
520 groups each of which contains species which have a similar way of responding to changes in their  
521 environment (Gitay and Noble, 1997). Functional group composition will affect the overall model  
522 outcomes and predictions of ecosystem models (Fulton et al., 2001), while the number of groups derived  
523 help us to understand functional diversity (Petchey and Gaston, 2002). Functional groups can be derived  
524 from expert knowledge, or from diet or trait based analyses, however these approaches incur significant  
525 costs, consume a lot of time and require invasive sampling of specimens (Albouy et al., 2011; Sibbing  
526 and Nagelkerke, 2000). We explored how individual species might cluster together based on information  
527 gathered about their diet, life history, morphology or habitat use, collected from published literature  
528 or observed from photographs of specimens. It was our aim to understand if meaningful groupings of  
529 teleost fish species can be made from known or easy to gather information. During this process, it quickly  
530 became apparent that there is no straightforward answer to how a functional group should be identified,  
531 and that there was not one most appropriate distance matrix or linkage method that could be applied to all  
532 situations. We therefore encourage future investigations to explore different distance matrices and linkage  
533 methods as they are easy to implement in statistical packages such as R (Ihaka and Gentleman, 1996).

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536 to making the trait matrix.

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