

1 **The transcription factors and its implications in adaptive evolution**

2 Víctor A. Zapata Trejo¹. *

3 ¹Department of Marine Biology, Autonomous University of Yucatan, Campus of Biological and Agricultural
4 Sciences, highway Merida-Xmatkuil Km. 15.5, Merida 97000, Mexico.

5 *Correspondence: A16018093@alumnos.uady.mx

6 Telephone number: +52 999 207 3228

7 ORCID: 0000-0002-8930-4195

8 **Abstract** The random to explain the emergence of variations in sequence of the alleles is the current scientific
9 paradigm of evolutionary biology. Here is argued that external stimuli responsible of manifestation of an instinct
10 together with transcription factors (TFs) involved are the main cause of emergence of such variations. Advances in
11 epigenomics show that this molecular function plays an important role in the regulation of gene expression of all cells,
12 both prokaryotes and eukaryotes, and indicates which specific genes should be transcribed and which should be
13 translated. Under this context, the present work pretends evaluate the current evidence on the increase in mutation rate
14 caused by transcription-associated mutational pressure in primordial germ cells due to presence of TFs also present in
15 somatic cells involved in an instinct. In conclusion is established that adaptive evolution can understood as biological
16 superposition of 4 functional states. This work that is added to the evolutionary theoretical framework contributes
17 with an alternative causal understanding of adaptive evolution.

18 **Keywords** adaptive system, gametogenesis, stimuli, stochastic, robustness.

19

20

21

22

23

24

25 New data, theoretical findings and approaches on niche construction, developmental plasticity and organism-
26 environment feedback, suggest that is needed an alternative causal understanding of adaptive evolution¹⁻⁵.
27 Evolutionary biology currently accepts that DNA sequences transmitted in eukaryotic gametes are not affected by the
28 experiences of individuals, as well as that variations in sequence of the alleles arise at random and not in response to
29 any need of the organisms⁶.

30 This paper evaluates the implications of transcription factors (TFs) in mutation rate of somatic cells and primordial
31 germ cells and complement the hypothesis, as far as I am aware, initially proposed by Bernard⁷ with premises that
32 explains how gene expression in somatic cells caused by an instinct increase mutation rate of same genes in primordial
33 germ cells before and during the gametogenesis⁸. Here is established that random does not exist and instead is offered
34 a causal explanation based in the superposition of 4 functional states.

35 **Transcription-associated mutational pressure**

36 If we knew the number of molecules of the mRNAs that integrate the transcriptome of a certain cell type in any
37 moment and the number of nucleotides of the genes that to the mRNAs correspond, would be possible calculate the
38 stochastic minimum (1) for possible mutations considering the following: the probability of that a gene after finished
39 the transcription has undergone at least 1 mutation during the interaction will always be 1/2.

40 equation (1)

41 $\text{stochastic minimum} = (1/s)(t)(1/2)$

42 Where s is the number of nucleotides of a gene and t the number of molecules of mRNAs that to the gene correspond.
43 The equation describes molecular behavior caused by transcription-associated mutational pressure and is offered as
44 formal theoretical definition.

45 **4 functional states for adaptive evolution**

46 Keeping the above in mind, is argued that emergence of a point mutation in sequence of an allele is result of an increase
47 in mutation rate due mainly by transcription-associated mutational pressure in genetic networks in demand^{9, 10}. To
48 understand how this molecular behavior affect adaptive evolution is necessary draw upon to the well-known principle
49 of superposition: a state whose function can understood as sum of the independent functions of each state that

50 composes it. To this principle, for a biological system, is needed add that superposition is a reconfigurable event of
 51 dynamic and robust¹¹ nature (2), that is, a likely event of dynamic configuration in space.

52 equation (2)

$$53 \quad f(a + b \dots \pm n) = f(a) + f(b) \dots \pm n$$

54 Where a y b are states and $\pm n$ represents the dynamic of states that can added or retired of the function. Thus, adaptive
 55 evolution can understand as biological superposition of 4 functional states (3), each one composed by a superposition
 56 of complex networks (4). The first is stochastic minimum, whose number is in function of gene expression (stochastic
 57 state). The following states can guide by the questions: what are mechanisms involved in mutagenesis? what
 58 mechanisms intervene in DNA repair, what are their signaling pathways and what TFs are involved? (cellular state);
 59 how does substitution of amino acids affect the structure and functionality of proteins and how influence in function
 60 of organism? (structural-functional state); how do organisms interact with ecosystem and what are states that define
 61 dynamic and robustness of interaction? (ecological state).

62 equation (3)

$$63 \quad f(st \in c \subseteq sf \subset e) = f(st) \in f(c) \subseteq f(sf) \subset f(e)$$

64 Where st , c , sf , and e are the states stochastic, cellular, structural-functional and ecological, respectively.

65 equation (4)

$$66 \quad f(r_1 + r_2 \dots + r_n) = f(r_1) + f(r_2) \dots + f(r_n)$$

67 Where r is a biological network whose function can understood as sum of the independent functions of each state that
 68 composes it (2).

69 **Robust interactions that respond to external stimuli**

70 The evolutionary theoretical framework explain that basic process of biological evolution is a process at the population
 71 level where adaptive evolution is due to shifting gene frequencies by natural selection, from an abundant pre-existing
 72 variation¹². If basic process of biological evolution is a process at the population level, instincts of the population
 73 should be determinant for adaptation of organisms. Therefore, instincts must have a role in evolutionary process.

74 However, instincts itself are phenotypes that responds to external stimuli of environment^{13, 14}, and according to theory
75 of niche construction, this process of dynamic and robust interaction between instincts of a population and its
76 environment can affect selection that acts on same population and on other species^{15, 16}.

77 Any function in prokaryotic and eukaryotic cells exerts a genetic demand in genome and there are stimuli that,
78 depending of cell type, activate transcription of required genetic product. The cell recognizes effectively each stimulus
79 through TFs whose evolutionary function is closely related to cell diversity¹⁷. Therefore, for appear a certain network
80 (cellular or structural-functional) associated to specific stimuli, is needed that cells locate in genome the operator sites
81 of DNA sequences that to TFs correspond¹⁸. To regulate gene expression, cells use diverse mechanisms that can, for
82 example, silence transcription or suppress translation of a genetic network even when stimuli associated to TFs are
83 present^{19, 20}. Some of these mechanisms are DNA methylation and MicroRNAs (miRNAs) that modulate gene
84 expression at the post-transcriptional level^{21, 22}. To interaction of these mechanisms with genome to control
85 transcription and translation of a specific genetic network is called epigenomics function²³. Therefore, epigenomics
86 function have an important role in gene expression of specific genetic networks that cells need to perform its functions
87 in presence of stimuli associated to TFs. All cells of a eukaryotic organism have the same genetic information, but
88 each cell type that integrates it have a different epigenomics function²⁴.

89 In eukaryotic organisms there is evidence that show how exposure to a specific stimulus can regulate gene expression
90 of cells involved²⁵⁻²⁷. For example, in zebra finch, the perception of singing is sexually dimorphic and implies that
91 during spermatogenesis, oogenesis, embryogenesis and development the genes involved has sex-biased expression
92 levels²⁸. In male, miR-2954 gene associated with habituation of song has higher levels of expression than in female
93 when both sexes are exposed to stimulus of new song although with frequent exposure of same song expression of
94 miR-2954 is gradually inhibited. Therefore, if a new song is frequently repeated, genes that intervene in positive
95 control of transcription associated to habituation of song will be susceptible to higher transcription rates²⁹.

96 In theory, operator sites in genes that respond to presence of their TFs are conserved in genetic information of all cells
97 that make up a eukaryotic organism. So, in primordial germ cells is likely that TFs also present in somatic cells act in
98 operator sites to which they are associated. Recently, has been proposed that type of regulator mechanism of gene
99 expression of an instinct can affect likelihood of trait plasticity evolving³⁰. Here is added to the above that external
100 stimuli responsible of manifestation of an instinct can influence in transcription rate of somatic cells implicated, and

101 by means of TFs, too in primordial germ cells. Consequently, external stimuli can increase mutation rate in genome,
102 molecular evolution rate and affect likelihood of trait plasticity evolving.

103 **Transcription-associated mutational pressure and GC-rich content**

104 Not long ago was suggested that gene of platelet phosphofructokinase in songbirds has enriched its guanine and
105 cytosine content due mainly by transcription-associated mutational pressure and it is important because the platelet
106 phosphofructokinase is a tissue enzyme that should not be transcribed during gametogenesis and yet it does partially
107 in one or several steps of this process³¹. The compartmentalized of guanine and cytosine is one of defining
108 characteristics to the eukaryotic genome³². This quality is associated with positive control of transcription. For
109 example, empirical evidence in studies on pre-implantation in both human and mouse indicated that expression levels
110 increased for those genes that were in regions with GC-rich content while than those that were in regions with GC-
111 poor content decreased expression levels³³. The GC-rich regions are also associated with negative control of
112 transcription. For example, in exons of some homologous copies of MET1 gene family of hexaploid genome of wheat
113 has been suggested that presence of GC-rich regions was caused by point mutations that induced pseudogenization
114 and DNA methylation³⁴.

115 From the above is concluded that exons located in GC-rich regions present evidence of evolutionary changes in
116 sequences associated with control both positive and negative of transcription. If is considered the epigenomics
117 function, is also concluded that transcription-associated point mutations had to occur in primordial germ cells³⁵, and
118 if in they transcription-associated mutational pressure responds to an increase in genetic demand as in somatic cells³⁶,
119 could deducted that currently exons located in areas with GC-rich content were under transcription-associated
120 mutational pressure³⁷, just what has been proposed for platelet phosphofructokinase in songbirds.

121 *In silico* genomic evidence already has confirmed the relation between expression levels, transcription-associated
122 mutational pressure and increase observed in mutation rate in yeast as well as the relation between increase observed
123 in mutation rate in germline cells and expression levels in humans³⁸. Therefore, scientific solidity of hypothesis
124 initially proposed by Bernard⁷ and complemented in this paper is demonstrated.

125 **Biological superposition as a principle to understand adaptive evolution**

126 The relation between transcription-associated mutational pressure and TFs is really superposition of two functional
127 states: the stochastic and cellular, respectively. The other two superpositions correspond to the instinct inside of
128 structural-functional state and to the niche construction inside of ecological state. This approach explains robustness
129 of an adaptive system with their environment where perturbation caused by point mutations is restricted by
130 superposition of complex networks inside ecological state and such mutations are optimal only if cause the upgrade
131 in flow of energy and information³⁹. This is where Dawkins's concepts on The Extended Phenotype⁴⁰ and The Selfish
132 Gene⁴¹ take on special importance.

133 The restriction by superposition of complex networks is also reason of why neutral mutations define the evolution of
134 genome, contrary to what happens at the organism level, where adaptive changes are strictly optimal and not neutral.
135 A neutral mutation can change composition of genome but not affect superposition of networks. As result, evolution
136 rate of genome is higher than observed at the organism level⁴².

137 **Three axioms to address the origin of life**

138 In a biological sense: the entropy, number of probable configurations in space for a certain state, decreases as
139 complexity of networks increase. Likewise, the superposition decreases as robustness of networks increase. One of
140 questions that forces us to consider this way of see natural world is: how does information behave in universe and
141 what are laws that govern its activity? What is possible add to theory about flow of information is that increases when,
142 in a determinate space, superposition of complex networks does it, and that information to be decode, encode and
143 recode in function of the stimuli perceived by organism must travel in packages constituted of elementary information
144 that would correspond to a universal information language.

145 **Conclusion**

146 The superposition demonstrated among transcription-associated mutational pressure (stochastic state), TFs (cellular
147 state), instinct (structural-functional state) and the niche construction (ecological state) indicates that gene expression
148 of somatic cells caused by an instinct due to external stimuli can increase transcription-associated mutational pressure
149 in spermatogonias and oogonias by presence of TFs that act in operator sites of same genetic networks that is involved
150 in instinct. Consequently, this system of non-direct interaction between primordial germ cells and environment

151 mediated by TFs have repercussions in developmental plasticity, adaptive evolution, molecular evolution rate and
152 represent a contribution to the evolutionary theoretical framework.

153 **References**

154 1. Gilbert, S.F., Bosch, T.C.G. & Ledón-Rettig, C. Eco-Evo-Devo: developmental symbiosis and developmental
155 plasticity as evolutionary agents. *Nat. Rev. Genet.* **16**, 611-622 (2015). doi:10.1038/nrg3982

156 2. Laland, K.N. *et al.* The extended evolutionary synthesis: its structure, assumptions and predictions. *Proc. R. Soc.*
157 *London, Ser. B* **282**, 20151019 (2015). doi:10.1098/rspb.2015.1019

158 3. Noble, D. Evolution viewed from physics, physiology and medicine. *Interface Focus* **7**, 20160159 (2017).
159 doi:10.1098/rsfs.2016.0159

160 4. Saltz, J.B. & Nuzhdin, S.V. Genetic variation in niche construction: implications for development and evolutionary
161 genetics. *Trends Ecol. Evol.* **29**, 8-14 (2014). <https://doi.org/10.1016/j.tree.2013.09.011>

162 5. Svensson, E.I. On Reciprocal Causation in the Evolutionary Process. *Evolutionary Biology* **45**, 1-14 (2017).
163 <https://doi.org/10.1007/s11692-017-9431-x>

164 6. Futuyma, D.J. Evolutionary biology today and the call for an extended synthesis. *Interface Focus* **7**, 20160145
165 (2017). doi:10.1098/rsfs.2016.0145

166 7. Bernard, D.D. Transcriptional bias: a non-Lamarckian mechanism for substrate-induced mutations. *PNAS* **86**, 5005-
167 5009 (1989). <https://doi.org/10.1073/pnas.86.13.5005>

168 8. Sandip, P., Million-Weaver, S., Chattopadhyay, S., Sokurenko, E. & Merrikh, H. Accelerated gene evolution
169 through replication–transcription conflicts. *Nature* **495**, 512-515 (2013). <https://doi.org/10.1038/nature11989>

170 9. Chen, X. & Zhang, J. No Gene-Specific Optimization of Mutation Rate in *Escherichia coli*. *Mol. Biol. Evol.* **30**,
171 1559-1562 (2013). <https://doi.org/10.1093/molbev/mst060>

172 10. Cui, P., Lin, Q., Ding, F., Hu, S. & Yu, J. The Transcript-centric Mutations in Human Genomes. *GPB* **10**, 11-22
173 (2012). [https://doi.org/10.1016/S1672-0229\(11\)60029-6](https://doi.org/10.1016/S1672-0229(11)60029-6)

174 11. Kitano, H. Biological robustness. *Nat. Rev. Genet.* **5**, 826-837 (2004). <https://doi.org/10.1038/nrg1471>

- 175 12. Stoltzfus, A. Why we don't want another "Synthesis". *BMC Biology Direct* **12**, 23 (2017).
176 <https://doi.org/10.1186/s13062-017-0194-1>
- 177 13. Tinbergen, N. *The Study of Instinct* (Oxford University Press, 1989).
- 178 14. Yan, H. *et al.* Eusocial insects as emerging models for behavioural epigenetics. *Nat. Rev. Genet.* **15**, 677-688
179 (2014). <https://doi.org/10.1038/nrg3787>
- 180 15. Constant, A., Ramstead, M.J.D., Veissière, S.P.L., Campbell, J.O. & Friston, K.J. A variational approach to niche
181 construction. *J. R. Soc. Interface* **15**, 20170685 (2018). doi:10.1098/rsif.2017.0685
- 182 16. Jänes, H., Herkül, K. & Kotta, J. Environmental niche separation between native and non-native benthic
183 invertebrate species: Case study of the northern Baltic Sea. *Mar. Environ. Res.* **131**, 123-133 (2017).
184 <https://doi.org/10.1016/j.marenvres.2017.08.001>
- 185 17. Konstantinides, N. *et al.* Phenotypic Convergence: Distinct Transcription Factors Regulate Common Terminal
186 Features. *Cell* **174**, 622-635 (2018). <https://doi.org/10.1016/j.cell.2018.05.021>
- 187 18. Getz, L.J. & Thomas, N.A. The Transcriptional Regulator HlyU Positively Regulates Expression of *exsA*, Leading
188 to Type III Secretion System 1 Activation in *Vibrio parahaemolyticus*. *J. Bacteriol.* doi:10.1128/JB.00653-17 (2018).
- 189 19. Liew, Y.J., *et al.* Epigenome-associated phenotypic acclimatization to ocean acidification in a reef-building coral.
190 *Sci. Adv.* **4**, eaar8028 (2018). doi:10.1126/sciadv.aar8028
- 191 20. Nusrin, S. *et al.* Regulation of CYP11B1 and CYP11B2 steroidogenic genes by hypoxia-inducible miR-10b in
192 H295R cells. *Mar. Pollut. Bull.* **85**, 344-351 (2014). <https://doi.org/10.1016/j.marpolbul.2014.04.002>
- 193 21. Ni, P. *et al.* Methylation divergence of invasive *Ciona* ascidians: Significant population structure and local
194 environmental influence. *Ecol. Evol.* <https://doi.org/10.1002/ece3.4504> (2018).
- 195 22. Zhang, X. *et al.* Identification and characterization of microRNAs involved in ascidian larval metamorphosis.
196 *BMC Genomics* **19**, 168 (2018). <https://doi.org/10.1186/s12864-018-4566-4>
- 197 23. Carlberg, C. & Molnár, F. *Human Epigenomics* (Springer, 2018). <https://doi.org/10.1007/978-981-10-7614-5>

- 198 24. Kenchanmane, R.S.K. & Niederhuth, C.E. Epigenetic Diversity and Application to Breeding. In: Christophe, M.
199 (ed) *Advances in Botanical Research* (Academic Press, 2018). <https://doi.org/10.1016/bs.abr.2018.08.001>
- 200 25. Gunaratne, P.H. *et al.* Song exposure regulates known and novel microRNAs in the zebra finch auditory forebrain.
201 *BMC Genomics* **12**, 277 (2011). <https://doi.org/10.1186/1471-2164-12-277>
- 202 26. McQuaid, J.B. *et al.* Carbonate-sensitive phytoferritin controls high-affinity iron uptake in diatoms. *Nature*
203 **555**, 534-537 (2018). <https://doi.org/10.1038/nature25982>
- 204 27. Nilsson, B., Jepsen, P.M., Bucklin, A. & Hansen, B.W. Environmental Stress Responses and Experimental
205 Handling Artifacts of a Model Organism, the Copepod *Acartia tonsa* (Dana). *Front. Mar. Sci.*
206 <https://doi.org/10.3389/fmars.2018.00156> (2018).
- 207 28. Luo, G.Z. *et al.* Genome-wide annotation and analysis of zebra finch microRNA repertoire reveal sex-biased
208 expression. *BMC Genomics* **13**, 727 (2012). <https://doi.org/10.1186/1471-2164-13-727>
- 209 29. Lin, Y.C., Balakrishnan, C.N. & Clayton, D.F. Functional genomic analysis and neuroanatomical localization of
210 miR-2954, a song-responsive sex-linked microRNA in the zebra finch. *Front. Neurosci.* **8**, 409 (2014).
211 <https://doi.org/10.3389/fnins.2014.00409>
- 212 30. Rittschof, C.C. & Hughes, K.A. Advancing behavioural genomics by considering timescale. *Nat. Commun.* **9**, 489
213 (2018). doi:10.1038/s41467-018-02971-0
- 214 31. Khrustalev, V.V., Barkovsky, E.V., Khrustaleva, T.A. & Lelevich, S.V. Intragenic isochores (intrachores) in the
215 platelet phosphofructokinase gene of Passeriform birds. *Gene* **546**, 16-24 (2014).
216 <https://doi.org/10.1016/j.gene.2014.05.045>
- 217 32. Costantini, M., Alvarez-Valin, F., Costantini, S., Cammarano, R. & Bernardi, G. Compositional patterns in the
218 genomes of unicellular eukaryotes. *BMC Genomics* **14**, 755 (2013). <https://doi.org/10.1186/1471-2164-14-755>
- 219 33. Barton, C., Iliopoulos, C.S., Pissis, S.P. & Arhondakis, S. Transcriptome activity of isochores during
220 preimplantation process in human and mouse. *FEBS Lett.* **590**, 2297-2306 (2016). <https://doi.org/10.1002/1873->
221 3468.12245

- 222 34. Thomas, M. *et al.* Evolutionary history of Methyltransferase 1 genes in hexaploid wheat. *BMC Genomics* **15**, 922
223 (2014). <https://doi.org/10.1186/1471-2164-15-922>
- 224 35. Percharde, M., Wong, P. & Ramalho-Santos, M. Global Hypertranscription in the Mouse Embryonic Germline.
225 *Cell Reports* **19**, 1987-1996 (2017). <https://doi.org/10.1016/j.celrep.2017.05.036>
- 226 36. Bergman, J., Betancourt, A.J. & Vogl, C. Transcription-Associated Compositional Skews in Drosophila Genes.
227 *Genome Biol. Evol.* **10**, 269-275 (2018). <https://doi.org/10.1093/gbe/evx200>
- 228 37. Touchon, M., Nicolay, S., Arneodo, A., d'Aubenton-Carafa, Y. & Thermes, C. Transcription-coupled TA and GC
229 strand asymmetries in the human genome. *FEBS Lett.* **555**, 579-582 (2003). <https://doi.org/10.1016/S0014->
230 [5793\(03\)01306-1](https://doi.org/10.1016/S0014-5793(03)01306-1)
- 231 38. Park, C., Qian, W. & Zhang, J. Genomic evidence for elevated mutation rates in highly expressed genes. *EMBO*
232 *rep.* **13**, 1123-1129 (2012). doi:10.1038/embor.2012.165
- 233 39. Chen, B.S. & Wu, W.S. Underlying Principles of Natural Selection in Network Evolution: Systems Biology
234 Approach. *Evol. Bioinf.* **3**, 245-262 (2007). <https://doi.org/10.1177/117693430700300010>
- 235 40. Dawkins, R. *The Extended Phenotype* (Oxford University Press, 2016).
- 236 41. Dawkins, R. *The Selfish Gene* (Oxford University Press, 2016).
- 237 42. Zhang, J. Neutral Theory and Phenotypic Evolution. *Mol. Biol. Evol.* **35**, 1327–1331 (2018).
238 <https://doi.org/10.1093/molbev/msy065>

239 **Acknowledgements**

240 The author is grateful with the friends, colleagues and academics who contributed to the gradual improvement of this
241 paper.

242 To my mother, the woman who makes my education possible.

243 **Competing interests**

244 The author(s) declare no competing interests.