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# Effects of ultraviolet radiation on metabolic rate and fitness of *Aedes albopictus* and *Culex pipiens* mosquitoes

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Environmental changes will alter many environmental factors in the coming years including temperature, precipitation, humidity, and the amount of solar radiation reaching the earth's surface, which in turn will have an impact on living organisms like invertebrates. In this study, we assessed the effect of UV-B radiation upon the metabolic rate and upon three fitness parameters (survival, development time, and body size) of the mosquitoes *Aedes albopictus* and *Culex pipiens*, and upon the production of microbial resources on which mosquito larvae feed in aquatic microcosms. We set up three UV-B radiation treatments mimicking levels typically measured in full-sun (FS) and shade (S) conditions, as well as a control group with no UV-B radiation (NUV). The metabolic rate expressed as heat production ( $\mu\text{watts/ml}$ ) for larvae and microbial community was measured at days 1, 8, and 15. Our results indicated that UV-B radiation affected the metabolic rate of both *Cx. pipiens* and *Ae. albopictus* larvae; metabolic rates were significantly higher in full-sun (FS) compared to shade (S) and no-UV condition (NUV), at days 8 and 15 compared to day 1 (Figures 1A and 1B). *Culex pipiens* metabolic rates were significantly higher than *Ae. albopictus* at day 15 compared to days 1 and 8 (Figure 1B). Metabolic rates were significantly lower in microbial communities from vials with *Ae. albopictus* larvae, *Cx. pipiens* larvae, and no larvae in FS conditions compared to vials from S and NUV conditions, especially at day 8 (Figure 2A and 2B). There was a major effect of UV-B conditions only on the survival of *Ae. albopictus* and *Cx. pipiens* mosquitoes, with significantly lower survival in FS compared to S and NUV conditions. UV-B radiation at levels found in aquatic environments in open fields showed a negative impact on the metabolic rate of *Ae. albopictus* and *Cx. pipiens* larvae and on the microbial communities on which they feed. These negative impacts could have important implications for the distribution and abundance of these mosquitoes and for the transmission rate of illness caused by the pathogens that these two broadly distributed mosquitoes transmit.

1 **Effects of ultraviolet radiation on metabolic rate and fitness of *Aedes albopictus***  
2 **and *Culex pipiens* mosquitoes**

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39 **Abstract**

40

41 Environmental changes will alter many environmental factors in the coming years  
42 including temperature, precipitation, humidity, and the amount of solar radiation reaching the  
43 earth's surface, which in turn will have an impact on living organisms like invertebrates. In this  
44 study, we assessed the effect of UV-B radiation upon the metabolic rate and upon three fitness  
45 parameters (survival, development time, and body size) of the mosquitoes *Aedes albopictus* and  
46 *Culex pipiens*, and upon the production of microbial resources on which mosquito larvae feed in  
47 aquatic microcosms. We set up three UV-B radiation treatments mimicking levels typically  
48 measured in full-sun (FS) and shade (S) conditions, as well as a control group with no UV-B  
49 radiation (NUV). The metabolic rate expressed as heat production ( $\mu\text{watts/ml}$ ) for larvae and  
50 microbial community was measured at days 1, 8, and 15. Our results indicated that UV-B radiation  
51 affected the metabolic rate of both *Cx. pipiens* and *Ae. albopictus* larvae; metabolic rates were  
52 significantly higher in full-sun (FS) compared to shade (S) and no-UV condition (NUV), at days  
53 8 and 15 compared to day 1 (Figures 1A and 1B). *Culex pipiens* metabolic rates were significantly  
54 higher than *Ae. albopictus* at day 15 compared to days 1 and 8 (Figure 1B). Metabolic rates were  
55 significantly lower in microbial communities from vials with *Ae. albopictus* larvae, *Cx. pipiens*  
56 larvae, and no larvae in FS conditions compared to vials from S and NUV conditions, especially  
57 at day 8 (Figure 2A and 2B). There was a major effect of UV-B conditions only on the survival of  
58 *Ae. albopictus* and *Cx. pipiens* mosquitoes, with significantly lower survival in FS compared to S  
59 and NUV conditions. UV-B radiation at levels found in aquatic environments in open fields  
60 showed a negative impact on the metabolic rate of *Ae. albopictus* and *Cx. pipiens* larvae and on  
61 the microbial communities on which they feed. These negative impacts could have important

62 implications for the distribution and abundance of these mosquitoes and for the transmission rate  
63 of illness caused by the pathogens that these two broadly distributed mosquitoes transmit.

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65 **Key words:** Ultraviolet radiation, metabolic rate, *Ae. albopictus*, *Cx. pipiens*, microbial  
66 communities

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**87 Introduction**

88

89 Environmental changes (e.g., global warming, climate change) will trigger major changes  
90 in environmental factors (e.g., temperature, solar radiation) in coming years. These changes are  
91 likely to have profound impacts on insect ecology and physiology, including survival,  
92 development time and underlying metabolic processes (Helbling and Zagarese, 2003).  
93 Environmental changes will differ with geographical regions, and the impacts on the ecology and  
94 physiology of insects will vary with the extent of temperature increase, amounts and patterns of  
95 precipitation and humidity, and changes in incoming solar radiation, especially UV-B radiation.  
96 Despite many uncertainties, there is consensus that environmental changes have had and will have  
97 impacts upon insect metabolic processes, fitness variables, geographical ranges, and abundance;  
98 upon species extinction; upon activity and abundance of natural enemies; and upon the  
99 transmission of vector-borne diseases (Shuman, 2011; Gray, 2013).

100 Temperature is the most well-studied environmental factor that affects insect biology and  
101 ecology. Most insects are ectothermic, meaning that their bodily heat source is primarily sourced  
102 from the environment; through thermoregulation, they regulate their body temperature to optimally  
103 support survival and reproduction (Klowden, 2007; Terblanche et al. 2005, 2009; Klowden, 2007).  
104 Although in the context of environmental changes much of the focus has been on changing  
105 temperatures, precipitation and humidity also have important impacts on insects. Terrestrial insects  
106 lose water through their cuticle, and aquatic insects require water for habitat. Water availability  
107 could affect insect activity, distribution patterns, and species richness, especially for those insects  
108 that inhabit ephemeral habitats (e.g., mosquitoes). Other environmental variables may also have  
109 subtle yet important effects upon insects; perhaps the most interesting of these is UV radiation.

110           Ultraviolet radiation (UVR) is part of the electromagnetic spectrum emitted by the sun,  
111 with a wavelength range between 400 and 100 nm (Andrady et al., 1998); it is subdivided into  
112 three subtypes: UV-A (400-315 nm), UV-B (315-280 nm) and UV-C (280-100 nm). Of these three  
113 subtypes UV-C, the most harmful, does not reach the earth's surface (Dyer, 2001; Caldwell et al.,  
114 2003). Of the UVR that reaches the earth's surface, around five percent corresponds to UV-B  
115 radiation and ninety five percent to UV-A radiation. Of these two, UV-B radiation is more harmful  
116 to biotic and abiotic environments because of its shorter wavelength, which means higher energy  
117 levels (Andrady et al.,1998). Variation in exposure to radiation throughout the landscape, because  
118 of varying shade conditions, can moderate the direct and indirect effects of UVR upon insects.  
119 Relatively few studies have examined the effects of UVR, and the few that have been published  
120 have focused on insect control through the use of UVR traps (Shimoda and Honda, 2013; Sliney  
121 et al., 2016) or, the use of UVR to affect insect physiology (e.g., flight behavior, orientation, visual  
122 ecology) in greenhouse facilities (Johansen et al., 2011).

123           Mosquitoes are blood-feeding insects of the order Diptera. They are medically important  
124 because they transmit vector-borne diseases. *Aedes albopictus* and *Cx. pipiens* are two common  
125 mosquitoes in urban areas of the eastern United States (Joy et al., 2003; Joy 2004; Costanzo et al.,  
126 2005). *Aedes albopictus* is an important vector for the transmission of many viral pathogens,  
127 including yellow fever, dengue, and Chikungunya (Lambrechts et al., 2010). *Aedes albopictus* is  
128 also capable of hosting the Zika virus and it is therefore considered a potential vector for Zika  
129 virus (Wong et al., 2013). *Culex pipiens* is an important vector for the transmission of West Nile  
130 virus, Japanese encephalitis, and meningitis (Gerhardt et al., 2001; Kim et al., 2005; Molaei et al.,  
131 2006). *Aedes albopictus* and *Cx. pipiens* are also capable of transmitting the dog heartworm

132 (*Dirofilaria immitis*), which not only affects dogs but also cats, foxes, coyotes, and other animals  
133 (Cancrini, 2007).

134 Mosquitoes have a complex life cycle; they lay eggs in aquatic environments where the  
135 larvae and pupae develop in several weeks until adults emerge into the terrestrial environment  
136 where they can freely move (Juliano, 2009). Mosquito larvae feed on microbial communities  
137 (Juliano, 2009). Environmental effects on larval stages have important consequences for some  
138 adult traits (Terblanche and Chown, 2007). Larval ecology affects distribution and abundance of  
139 adults, by modulating survival as well as adult fitness parameters, such as body size, that can affect  
140 adult survival, biting rate, and ultimately the ability to vector and transmit pathogens. There is little  
141 information on the effect of UV-B radiation on mosquito metabolic rate and survival and on the  
142 microbial community on which mosquitoes feed. One of the few studies that has assessed the  
143 effects of UVR on mosquitoes dates back to the 1930's (MacGregor, 1932); the author,  
144 demonstrated clear negative effects on larvae and pupae of *Ae. aegypti* and *Cx. pipiens* mosquitoes.  
145 However, a significant limitation of the study is that the UVR levels used were not comparable to  
146 field conditions.

147 Other studies have demonstrated that UV-B radiation has effects on microbial communities  
148 (Pancotto et al., 2003), but none have examined how these effects may impact mosquito  
149 populations. Future variations in UV-B, resulting from climate change and anthropogenic activities  
150 (e.g., change in land use, pollution), may have more important consequences for microbial  
151 communities and for decomposition of dead plant and animal material than the changes in UV-B  
152 caused by ozone depletion, thereby affecting the food chains that depend on microbial  
153 communities (Ballare et al., 2011). The goal of this study is to test the effect of field-relevant UV-B  
154 radiation on the metabolic rate, larval survival, development time, and adult body size of *Ae.*



155 *albopictus* and *Cx. pipiens* mosquitoes, and on the production of the microbial communities on  
156 which the larvae feed.

## 157 **Materials and methods**

### 158 *Collection and maintenance of mosquitoes*

159  
160 *Aedes albopictus* and *Cx. pipiens* larvae were collected from multiple locations in College  
161 Park, Baltimore, and Towson, Maryland. Neither *Ae. albopictus* or *Cx. pipiens* are endangered,  
162 and collection sites were either on publically accessible lands or on private lands where consent  
163 for collections was granted at the time of collection; thus, no field permits were required to collect  
164 them. Field collected *Ae. albopictus* and *Cx. pipiens* larvae were reared to adulthood at 25°C at  
165 16:8 (L:D) h photoperiod, and then released into 1-m<sup>2</sup> single-species cages. Adults were kept in  
166 an insectary at 25°C and >85% RH, 16:8 (L:D) h photoperiod. Both colonies were supplied 20%  
167 sugar solution. Females from both colonies were fed horse or rooster blood once a week via an  
168 artificial feeder (Hemotek, Accrington, UK) to ensure egg production and experimental larvae.  
169 *Aedes albopictus* females oviposited on seed paper in 500 ml black cups covered filled with 200  
170 ml of deionized (DI) water. Eggs were collected over multiple weeks and stored at >80% RH and  
171 16:8 hours (L:D) photoperiod until hatching for the experiment. *Culex pipiens* oviposited egg rafts  
172 into a 500 ml black bowl filled with 400 ml of DI water. *Culex pipiens* eggs cannot be held without  
173 hatching; thus, egg rafts were collected within 24 h of oviposition, hatched in a lactobumina: yeast  
174 solution, and larvae were transferred into the experiment after being rinsed. *Aedes albopictus* eggs  
175 that had been stored were also hatched in a lactobumina: yeast solution and transferred into the  
176 experiment after being rinsed and within 24 h of hatching. Experimental larvae of both species  
177 were F<sub>1-3</sub> generation.

178 ***Experiment set up***

179

180           The experimental design was a split plot-randomized complete block design (RCBD) with  
181 UV-B radiation condition as the main plot, mosquito larvae cohorts (*Ae. albopictus*, *Cx. pipiens*,  
182 or no larvae) as the sub-plots, and three replicate blocks. Individuals of *Ae. albopictus* and *Cx.*  
183 *pipiens* were sorted into single species cohorts of 10 newly hatched individuals and added to 20  
184 ml vials with 17 ml of DI water that were inoculated with 1 ml of water containing a microbial  
185 community that was collected from discarded tires. A total of 45 vials were prepared. Ten newly  
186 hatched *Ae. albopictus* or ten newly hatched *Cx. pipiens* were randomly added to 15 vials each. 15  
187 vials only contained microbial community; no larvae were placed on these vials. Fifteen of the  
188 total vials (5 of each treatment) were randomly allotted to one of three Percival reach-in  
189 environmental chambers, model I-36 VL, located in the Aqua Engineering laboratory in the  
190 Environmental Science and Technology Program (ENST). Each chamber was kept at 25° C, 16:8  
191 (L: D), and 80- 90 % of humidity, to mimic typical summer conditions in the northeastern U.S.  
192 (Day et al., 1993; Li et al., 2006). At the end, each environmental chamber had 5 vials containing  
193 10 first instar larvae of *Ae. albopictus*, 5 vials containing 10 first instar larvae of *Cx. pipiens*, and  
194 5 vials containing no larvae, only the microbial community. Vials represent sub-samples and each  
195 experimental unit was a group of 5 vials in one of the three environmental chambers. Each vial  
196 was checked daily to collect pupae and place them in individual vials with water from that vial  
197 until adults emerged. I recorded the following information for each adult: date of emergence, sex,  
198 species, and replicate (notebook for the experiment). On the day of emergence, the adults were  
199 killed by placing them on a drying oven for further analysis such as wing length measurements.  
200 The Experiment was run until all larvae had died or eclosed.

201 One of three UV-B radiation conditions mimicking full-sun (10.82  $\mu\text{mol}/\text{m}^2/\text{s}$ ), shade (6.1  
202  $\mu\text{mol}/\text{m}^2/\text{s}$ ), and a no-UV control group (0  $\mu\text{mol}/\text{m}^2/\text{s}$ ), were applied to each chamber. To achieve  
203 the required UV-B levels in the full-sun and shade treatment chambers, cellulose diacetate filters  
204 were applied on four UV-B-313 lamps (Q Panel Lab Products, Cleveland, OH) in each chamber,  
205 and vials were placed 5 cm and 20 cm from the bulbs, respectively. For the control group, I used  
206 regular Phillips 32 watts bulbs, model 205047, which simulate a visible range of sunlight (400 nm  
207 - 700 nm). To assure uniform exposure to UV radiation, vials were rotated daily. I ran the  
208 experiment three times (blocks) and applied a different UV-B treatment to each incubator each  
209 time to minimize incubator-treatment confounding effect.

#### 210 *Measurement of metabolic rate*

211  
212 Metabolic rate was measured as the rate of heat production ( $\mu\text{watts}/\text{ml}$ ) by a heat  
213 conduction, multicell differential scanning calorimeter (MC-DSC model 4100, Calorimetry  
214 Sciences Corp.). The multicell differential calorimeter was set up in isothermal mode, at a  
215 temperature of 25 °C, which allows concurrent measurements of two samples using two 1  $\text{cm}^3$   
216 ampoules. For larvae, five were selected randomly from each sub-sample of each of the three  
217 treatments. Before being placed in the ampoule, they were washed in sterilized water and placed  
218 inside the ampoule with 1 ml of deionized water. The heat production was monitored for 60  
219 minutes to allow for temporal equilibration and consistency of final readings. Previous to this step,  
220 I ran a blank sample (just deionized water) for 60 minutes. After obtaining the reading in  $\mu\text{watts}$   
221 ( $\mu\text{W}$ ) I subtracted the blank reading from the sample reading, and the result was the final metabolic  
222 rate value in  $\mu\text{watts}/\text{ml}$  (Zhang et. al., 2009). Metabolic rates were measured when larvae were  
223 first instars (within 2 to 24 hours of hatching); the second measurement was when larvae were 8  
224 days old; and the final measurement was made when larvae were 15 days old. For mosquito larvae

225 metabolic rate measurements, from each species, five larvae were collected randomly from each  
226 vial, together with 5 ml of water from the same vial and placed in a sterile bottle (5 ml) and  
227 transported to the laboratory for metabolic rate measurements. Vials in chambers were refilled  
228 with deionized water as needed.

229 To measure the metabolic rate of the microbial community I followed the same procedure  
230 described for mosquito larvae metabolic rate measurements. The only difference was that I placed  
231 1 ml of water sampled from the vials in the ampoules. We measured the microbial metabolic rate  
232 24 hours after inoculation of vials with a 1 ml of microbial inoculum from tires (day 1). After  
233 completing microbial metabolic measurements for day 1, we placed the first instar larvae in the  
234 corresponding vials as described in previous paragraph. The metabolic rate of the microbial  
235 community was also measured on days 8 and 15. Before placing samples in the ampoules, they  
236 were washed in sterilized water and sterilized with ethanol before each run (Zhang et al., 2009).  
237 To place larvae and water microbial samples into the ampoules, we used sterile pipettes, tips, and  
238 forceps to avoid any kind of sample contamination. To collect microbial community samples to  
239 measure metabolic rate, we mixed the liquid content in each vial with a manual stirrer and took a  
240 2 ml sample and placed it in a sterile bottle (5 ml) for its transportation to the laboratory for  
241 metabolic rate measurements.

## 242 *Analyses*

243

244 All the data were analyzed using linear models using the SAS 9.4 software (SAS Institute  
245 Inc. 2013). The metabolic rate of *Ae. albopictus* and *Cx. pipiens* larvae and microbial community  
246 from containers with *Ae. albopictus* larvae, *Cx. pipiens* larvae and no larvae were analyzed as a  
247 three-way analysis of variance (ANOVA) containing one repeated factor (day of sampling) using  
248 the PROC MIXED procedure, with UV-B condition, species and days as fixed effects, day as the

249 repeated variable, and block as a random effect. Metabolic rate was measured on days 1, 8, and  
250 15. To account for assumptions of normality and homogeneity of variances, data were  $\log_{10}(y)$   
251 transformed.

252 For vials with larval mosquito cohorts, fitness parameters were calculated (proportion  
253 survival, development time, and wing length). To determine survival rate, the number of adults  
254 were compared with the initial number of larvae placed in the experimental units; to measure mean  
255 development time, we considered the days from hatching to adulthood; and to measure wing  
256 length, we used a dissecting microscope and the image analysis system called Image Pro Plus 6.0.  
257 These fitness parameters were analyzed as a two-way ANOVA using the PROC MIXED  
258 procedure; we considered UV conditions and species as fixed effects, and block as a random effect  
259 in the model. To account for assumptions of normality and homogeneity of variances, data were  
260  $\log_{10}(y+1)$  transformed. We did a pairwise mean comparison in the mixed procedures using the  
261 LSMEANS statement with tukey adjustment. For all analyses experiment-wise  $\alpha = 0.05$ ; marginal  
262 significance was defined  $\alpha = 0.05-0.10$ .

## 263 Results

### 264 *Metabolic rate of mosquito larvae*

265  
266 There was an interaction between UV-B condition and day of sampling (Table 1). There  
267 was also an interaction between species and day of sampling for larval metabolic rate, indicating  
268 differences in metabolic rate depending on the day of sampling for both *Ae. albopictus* and *Cx.*  
269 *pipiens* (Table 1, Figure 1B). At day 15, metabolic rate of *Cx. pipiens* was significantly higher  
270 compared to *Ae. albopictus* in FS conditions; this was not seen on days 1, and 8 (Figure 1B). Main  
271 effects of UV-B condition, species, and days were detected on larval metabolic rates (Table 1).  
272 Metabolic rates of both *Ae. albopictus* and *Cx. pipiens* were significantly higher under FS

273 condition compared to NUV condition (Figure 1A). Metabolic rates of both *Ae. albopictus* and *Cx.*  
274 *pipiens* were higher at days 8 and 15 compared to day 1, with *Cx. pipiens* metabolic rates being  
275 higher than *Ae. albopictus* metabolic rates (Figure 1B).

### 276 ***Metabolic rate of microbial community***

277

278 There was an interaction between UV-B condition and day of sampling for microbial  
279 metabolic rate (Table 2). Main effects of UV-B condition and day of sampling were detected on  
280 microbial metabolic rates (Table 2, Figures 2A and 2B). Metabolic rates of microbial communities  
281 from vials with *Ae. albopictus*, *Cx. pipiens* larvae and no larvae were significantly lower in FS  
282 condition compared to S and NUV conditions (Figure 2A). Metabolic rates of microbial  
283 communities from vials with *Ae. albopictus*, *Cx. pipiens* larvae and no larvae were significantly  
284 lower at day 8 compared to days 1 and 15 (Figure 2B).

### 285 ***Mosquito Fitness parameters***

286

287 There was no an interaction between UV-B condition and species for *Ae. albopictus* and  
288 *Cx. pipiens* fitness parameters: survival, development time, and body size (Table 3, Figure 3). UV-  
289 B condition affected the survival of *Ae. albopictus* and *Cx. pipiens* mosquitoes similarly, with  
290 significantly lower survival of both species under FS conditions compared to S and NUV  
291 conditions (Figure 3A and 3B). There was also a main effect of species on body size between *Ae.*  
292 *albopictus* and *Cx. pipiens*, with *Cx. pipiens* being the larger on average (Figure 3E and 3F).

### 293 **Discussion**

294

295 Ultraviolet radiation may have important effects on the development of mosquitoes, effects  
296 that could have important implications for the distribution and abundance of pathogen-transmitting  
297 species and their microbial food resources. This is the first study that has assessed the effect of

298 UV-B radiation comparable to that which reaches water bodies in open fields (full-sun), shaded  
299 areas (shade), and no-UV radiation conditions (control group) upon the fitness (survivorship,  
300 development time, and body size) and metabolic rates of two of the most broadly distributed  
301 mosquito species in the world: *Ae. albopictus* and *Cx. pipiens*; and upon the microbial communities  
302 on which they feed. In previously published field studies, resting metabolic rate increased in  
303 mosquito larvae between emergence and day 4 to 5 (Gray and Bradley, 2003). Our results showed  
304 that larval metabolic rate of both *Cx. pipiens* and *Ae. albopictus* mosquitoes significantly increased  
305 in FS conditions compared to NUV conditions (Figure 1A). In addition, larval metabolic rate is  
306 significantly higher at day 8 and day 15, indicating a direct effect of UV-B radiation on mosquito  
307 metabolism. Furthermore, at day 15, metabolic rate was significantly higher for *Cx. pipiens*  
308 compared to *Ae. albopictus* (Figure 1B).

309         The increase of metabolic rate in *Cx. pipiens* larvae from day 8 to day 15 under FS  
310 conditions compared to *Ae. albopictus* presumably reflects that UV-B radiation had a greater  
311 negative effect on *Cx. pipiens* larvae. It has been shown in many insect species that metabolic rate  
312 is strongly related to the physical and biological factors that influence metabolism (Gray and  
313 Bradley, 2003). The *Culex pipiens* mosquito larvae probably increases its metabolic rate under FS  
314 conditions, especially at day 15, in response to negative effects of UV-B radiation. These effects  
315 could cause larvae to increase energy expenditure in order to perform physical and biological  
316 activities such as getting food, growing, or competing with conspecifics, which is reflected in  
317 greater metabolic rates. It has been shown previously that UV-B radiation has a greater negative  
318 effect on the larval stage than the pupae stage, as the pupae stage is more resistant to damage by  
319 UV radiation (MacGregor, 1932). In that study, larvae of *Cx. pipiens* exposed to UV-B radiation  
320 were affected when exposed for long periods of time (more than 48 hours). Larvae lost movement

321 coordination and increased swimming rates after 24 hours of exposure but still 60 % of larvae were  
322 able to pupate. However, none were able to become adults. Histological analyses showed that  
323 larvae suffered damage in the cuticle, there was disintegration of the abdominal segments, that the  
324 peristaltic-wave no longer travelled between the 7th and 8th segment, and that the pulse rate was  
325 lowered (MacGregor, 1932).

326         Natural and artificial container aquatic habitats (e.g., puddles, tree holes, tires) are  
327 inhabited by a specialized community of macroinvertebrates (e.g., mosquitoes) that feed on  
328 microbial communities associated with decaying organic matter from insect carcasses and leaf  
329 litter (Walker et al., 1988; Walker et al., 1991). Microbial abundance and diversity could be  
330 affected by environmental stressors (e.g., contaminants, UV radiation). In this study, we assessed  
331 how microbial communities from microcosms that contain *Ae. albopictus* larvae, *Cx. pipiens*  
332 larvae, or no larvae (just microbial community), were affected by UV-B radiation. We used  
333 microbial community metabolic rate expressed as heat production ( $\mu$ watts/ml) as an indicator of  
334 the quantity of microbial community. Microbial community metabolic rate in the three different  
335 microcosms was significantly lower in FS compared to S and NUV conditions, especially for  
336 microbial communities from microcosms that do not contain larvae; metabolic rate decreased in  
337 35.73 % from NUV conditions and in 33.93 % from S conditions compared to FS conditions. This  
338 showed that UV radiation levels reaching water bodies on open fields had a negative effect in the  
339 metabolic rate of microbial community compared with water bodies in shade areas. This could be  
340 a mechanism leading to low reproduction and even dying of bacterial community, which would  
341 indirectly affect larvae that feed on them. At day 8, microbial community metabolic rate in the  
342 three different microcosms was significantly lower compared to days 1 and 15. Increase of  
343 metabolic rate from day 8 to day 15 is probably due to the input of dead carcasses from larvae that



344 were dying in the vials due to the negative effect of UV-B radiation; this material probably served  
345 as a nutrient source that increased microbial community size and metabolic rate.

346 In regard to the effect of UV radiation on microbial and algae communities that serve as  
347 food resources for mosquito larvae (Pelz-Stelinski et al., 2011), there are different points of view.  
348 Some authors indicate that UV-B radiation has negative effects on microorganisms and algae (Wu  
349 et al., 2009; Hader et al., 2007). Gao et al (2008) reported that UV-B radiation damages the DNA,  
350 proteins, membranes, and photochemical efficiency of photosynthetic prokaryote organisms like  
351 *Arthrospira platensis* (cyanobacteria), affecting photosynthesis and biomass production. The  
352 spiral structure of *A. platensis* is broken, and there is inhibition of photosynthetic activity with  
353 exposure to UV-B radiation in a temperature range of 18 to 20 °C, and this results in low biomass  
354 densities. The damage to these cells is temperature and density dependent (Gao et al., 2008). Wu  
355 et al. (2005) also found that exposure to 6 hours of UV-B radiation breaks the spiral filaments of  
356 *A. platensis* into small pieces and it also affects photosynthesis activity. UV-B radiation  
357 specifically affects the photosynthetic electron transport and pigment-protein complexes of *A.*  
358 *platensis* (Wu et al., 2005). Hader et al. (2007) showed that UV radiation affects negatively algae  
359 and microbial communities in aquatic ecosystems on which mosquito larvae feed. UV radiation  
360 penetrates significant depths in aquatic systems, depending on water transparency, with effects  
361 ranging from effects on major biomass producers such as phytoplankton to effects on consumers  
362 in the food web such as mosquito larvae. Davidson and Belbin (2002) found that marine  
363 phytoplankton and protozoan community assemblages exposed to UV radiation at less than 2  
364 meters depth for more than a day suffer a reduction in biomass and concentration per cubic meter  
365 of water, which would represent less availability of food for mosquito larvae that feed on this  
366 microbial assemblage. On the other hand, other authors have suggested that UV radiation could be

367 beneficial to microbial communities because of increased availability of dissolved organic carbon;  
368 this would promote bacterial growth and bacterial abundance, leading to the increase of food  
369 resources for mosquito larvae (De Lange et al., 2003).

370         Of the fitness parameters we assessed, only survival was significantly negatively affected  
371 by UV-B radiation, in FS conditions in both species (Figure 3A, 3B). Lower survival rates in FS  
372 conditions were probably a consequence of the direct effect of UV-B radiation, but also of the  
373 indirect effect of lower amounts of food resources in microcosms in FS conditions compared to  
374 microcosms from S and NUV conditions. UV-B exposure probably stressed larvae, which was  
375 reflected in greater metabolic rates. Greater metabolic rates were probably due to greater  
376 expenditures of energy to keep with biological processes (e.g., growth) and searching for food  
377 resources, which provide the calories needed for biological processes, considering that food  
378 resources decline in habitats exposed to high UV-B radiation. Stress in insects had been detected  
379 through the release of stress hormones such as cortisol, epinephrine, octopamine (Peric-Mataruga  
380 et al., 2006; Farooqui, 2012). Furthermore, Development time showed a trend toward greater  
381 development time under FS conditions compared to NUV conditions in both species. Also, for  
382 body size of *Ae. albopictus* and *Cx. pipiens* there was a trend toward smaller body size in FS  
383 condition compared to NUV conditions. These results are similar to those of Sang et al. (2017),  
384 who found a negative effect of UV-B on survival, development time and reduced size in *tribolium*  
385 *castaneum*, which is not a mosquito, but it has a similar life cycle. The study of Hori et al. (2014)  
386 showed that not only could UV radiation have a negative impact on mosquito survival, but also  
387 that wavelengths in the violet and blue range could cause pupae mortality as high as 60 percent.  
388 Understanding the effects of ultraviolet radiation on larval metabolic rate and fitness parameters  
389 of mosquitoes could lead to the development of new ways to control mosquitoes, to predict future

390 geographic distribution due to changes in solar radiation, and to prevent outbreaks of illness caused  
391 by viruses transmitted by these mosquitoes.

392         In Summary, we observed that larval metabolic rate of *Ae. albopictus* and *Cx. pipiens* were  
393 significantly higher in full-sun conditions compared to no-UV conditions, especially at day 15,  
394 and that negative effects upon these mosquitoes were expressed in lower survival rates, greater  
395 development time, and smaller sizes of both species under FS conditions compared to NUV  
396 conditions. Also, we observed that the bacterial communities of container aquatic habitats  
397 demonstrated lower metabolic rates in response to disturbance by UV-B radiation in FS conditions.  
398 These findings enhance the understanding of how changes in UV-B radiation could affect  
399 mosquito fitness and the microbial communities on which mosquitoes feed; and they suggest  
400 impacts upon some key ecological processes such as decomposition, nutrient cycling, and  
401 microbial diversity, processes that should be evaluated in future studies.

## 402 **Conclusions**

403         *Aedes albopictus* and *Culex pipiens* are two mosquito species that are highly present in  
404 urban areas of the United States (Lounibos, 2002). *Aedes albopictus* is currently present in 33  
405 states and *Cx. pipiens* is present in 38 states (Evans et al., 2017). *Aedes albopictus* and *Cx. pipiens*  
406 mosquito populations overlap in their geographic distribution in 27 states where they coexist in  
407 spite of the fact that *Cx. pipiens* is an inferior competitor compared to *Ae. albopictus* (Carrieri et  
408 al., 2003; Costanzo et al., 2005; Costanzo et al., 2011). In the area where these mosquitoes overlap  
409 in their distribution, they could represent a threat to public health because they could promote  
410 increased human incidence of WNV, considering that *Cx. pipiens* is the main vector of WNV and  
411 that *Ae. albopictus* could act as a bridge vector for WNV (Brustolin et al., 2016).

412           Ultraviolet radiation may have important effects on *Ae. albopictus* and *Cx. pipiens*  
413 mosquitoes, especially in peridomestic areas. Understanding the factors affecting heterospecific  
414 competition of the immature stages of mosquitoes is important to the understanding of their  
415 distribution and of measures to control of adult populations. The overall goal of this dissertation,  
416 was to test the effects of UV-B radiation on the larval ecologies of *Ae. albopictus* and *Cx. pipiens*.

417           My results showed that both *Ae. albopictus* and *Cx. pipiens* had higher metabolic rates and  
418 lower survival in full-sun conditions compared to no-UV conditions, probably because they were  
419 under greater physiological stress. Stress could be due to direct UV-B exposure that demand  
420 greater partitioning of energy to maintaining bodily processes (e.g., feeding, growth, and  
421 reproduction), which would be expressed in lower survival rates and greater development time  
422 (MacGregor, 1932). Water samples exposed to full-sun conditions also showed lower microbial  
423 activity than samples exposed to no-UV and shade conditions, suggesting that full-sun exposure  
424 appears to decrease available microbial food resources for mosquito larvae. Thus, another form of  
425 stress could be via reduced food availability that could limit energy available to maintenance, or  
426 encourage larvae to forage for food for longer and incur injuries from increasing swimming, both  
427 of which could lead to reduced survivorship.

428           Overall, my study suggests that UV-B radiation can have strong effects on the larval  
429 ecology of both *Ae. albopictus* and *Cx. pipiens*, both through direct negative effects on metabolic  
430 processes and resultant decreases in survival, and indirectly through the decrease of food  
431 availability. Therefore, the effects of UV-B on larval ecology is likely to be especially important  
432 in dictating the distribution and abundance of both *Ae. albopictus* and *Cx. pipiens* mosquitoes in  
433 different mosquito habitats (e.g. tires) and in structuring their communities.

434           The effects of UV-B radiation are likely to be complex and may be manifest in both the  
435 immature (larval) and adult life stages, as well as in the microbial communities on which mosquito  
436 larvae feed. Additional research needs to examine the effects of UV-B radiation on other  
437 community processes, such as predation, parasitism, and on vector competence across other  
438 disease systems such as chikungunya and Zika virus.

439

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**Table 1** (on next page)

Analysis of variance of the effects of UV-B radiation conditions and species on mosquito larvae metabolic rates.

Three-way ANOVA of the effects of UV-B conditions (FS, S, and NUV) and species (*Ae. albopictus* and *Cx. pipiens*) at three different times (days 1,8, and 15) on the larvae metabolic rate of *Ae. albopictus* and *Cx. pipiens* mosquitoes.

1  
2

Variable	Larval metabolic rate		
	dfs	F	P
UV conditions	2,10	5.50	<b>0.0245</b>
Species	2,10	6.08	<b>0.0333</b>
UV conditions x Species	2,10	0.58	0.5799
Days	2,24	350.85	<b>&lt;0.0001</b>
UV conditions x Days	2,24	13.96	<b>&lt;0.0001</b>
Species x Days	2,24	14.79	<b>&lt;0.0001</b>
UV conditions x Species x Days	2,24	0.87	0.4975

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**Table 2** (on next page)

Analysis of variance of the effects of UV-B conditions, species inhabiting vials, and time on the metabolic rates of microbial communities.

Three-way ANOVA of the effects of UV-B conditions (FS, S, and NUV) and the species that inhabit the vials where microbial samples come from (*Ae. albopictus*, *Cx. pipiens*, and no larvae) at three different times (days 1, 8, and 15) in the metabolic rate of microbial community.

1

Variable	Microbial metabolic rate		
	dfs	F	P
UV conditions	2,16	10.74	<b>0.0011</b>
Species	2,16	1.13	0.3483
UV conditions x Species	4,16	0.79	0.5502
Days	2,36	5.69	<b>0.0071</b>
UV conditions x Days	4,36	3.65	<b>0.0135</b>
Species x Days	4,36	0.47	0.7562
UV conditions x Species x Days	8,36	0.54	0.8203

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**Table 3** (on next page)

Analysis of variance of the effects of UV-B conditions and specie on the fitness parameters of mosquitoes.

Two-way ANOVA of the effects of UV-B conditions (FS, S, and NUV) and specie (*Ae. albopictus* and *Cx. pipiens*) on the fitness parameters (survival, developmental time, and body size) of *Ae. albopictus* and *Cx. pipiens* mosquitoes.

1

Variable	Survival			Development time			Body size-wing length		
	dfs	F	P	dfs	F	P	dfs	F	P
UV conditions	2,11	7.11	<b>0.0104</b>	2,11	0.80	0.4773	2,11	1.05	0.3857
Species	1,11	0.01	0.9963	1,11	1.50	0.2491	1,11	16.36	<b>0.0023</b>
UV conditions x species	2,11	0.59	0.5717	2,11	0.30	0.7456	2,11	0.05	0.9518

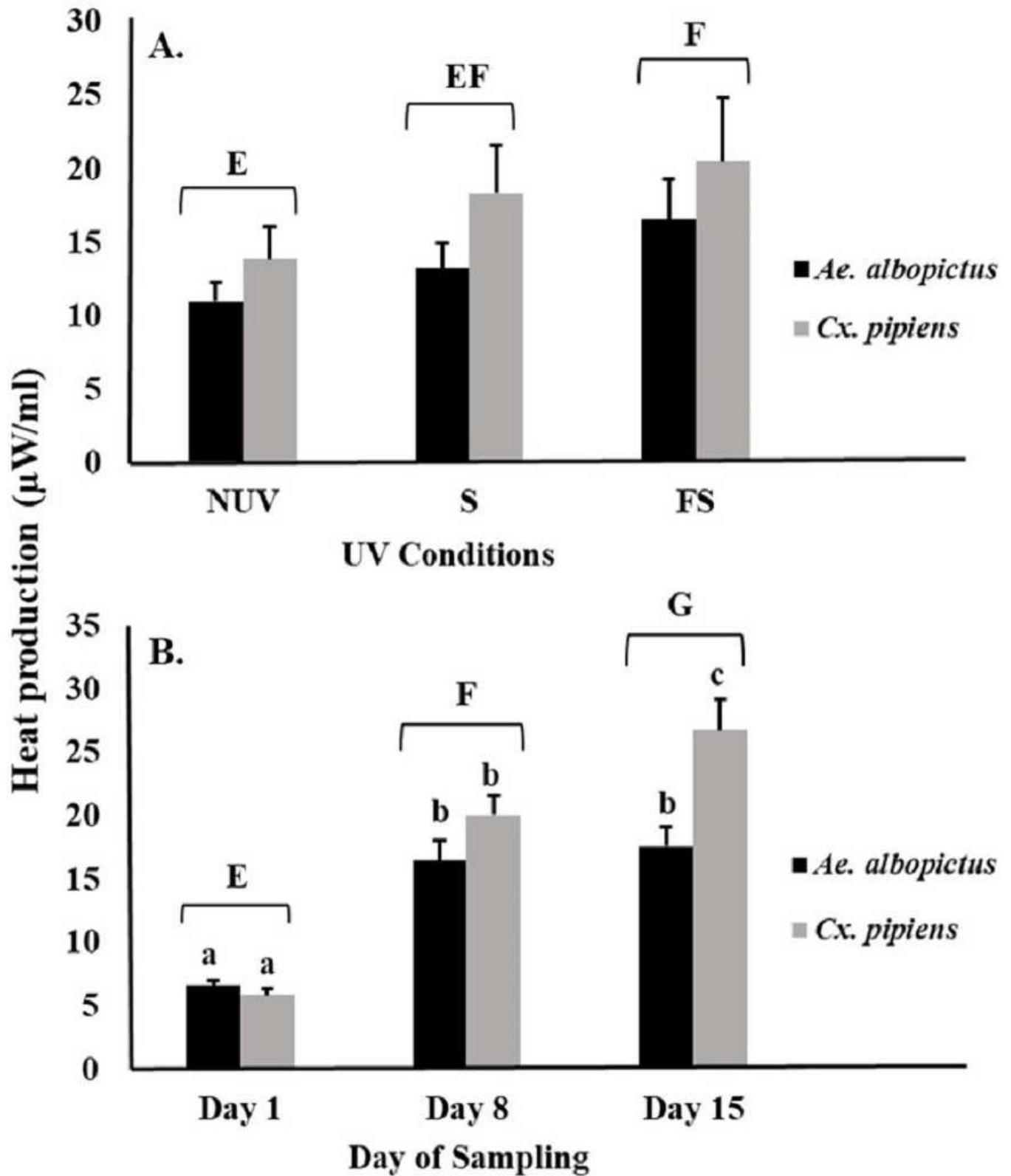
2

# Figure 1

Metabolic rate of mosquitoes in response to UV-B radiation conditions and time.

Least squares means ( $\pm$  SE) for metabolic rate expressed as heat production ( $\mu$ W/ml) of larvae of *Ae. albopictus* and *Cx. pipiens* in response to (a) UV-B conditions (NUV, S, and FS) and (b) day of metabolic rate measurement (days 1, 8, and 15). Data were statistically tested using ANOVA. Significant pairwise comparisons among treatment levels for main effects of (a) UV-B conditions and (b) day of sampling are indicated by capitalized letters, and interaction effects of UV-B conditions and day of sampling are indicated by lower case letters.

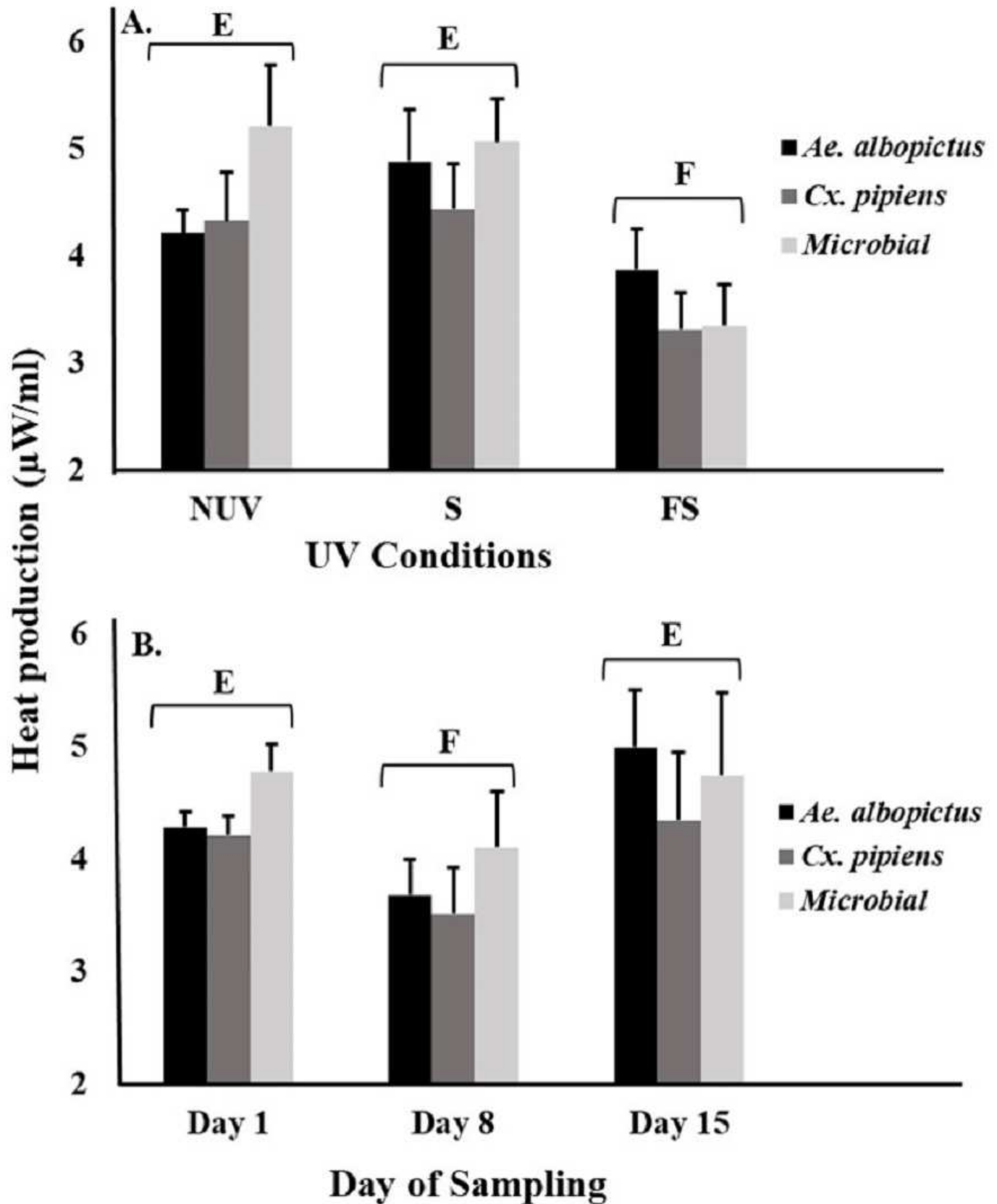




## Figure 2

Metabolic rate of microbial communities in response to container inhabitants, UV-B radiation conditions, and time.

Least squares means ( $\pm$  SE) for metabolic rate expressed as heat production ( $\mu$ W/ml) of microbial community from vials that contain *Ae. albopictus* larvae, *Cx. pipiens* larvae, and no larvae (just microbial community) in response to (a) UV conditions (NUV, S, and FS) and (b) day of metabolic rate measurement (days 1, 8, and 15). Data were statistically tested using ANOVA. Significant pairwise comparisons among treatment levels for main effects of (a) UV-B conditions and (b) day of sampling are indicated by capitalized letters.



## Figure 3

Mosquito fitness parameters in response to UV-B radiation conditions.

Least squares means ( $\pm$  SE) for fitness parameters of *Ae. albopictus* and *Cx. pipiens* mosquitoes. (a) survival percentage of *Ae. albopictus* (b) survival percentage of *Cx. pipiens* (c) development time of *Ae. albopictus* (d) development time of *Cx. pipiens* (e) body size of *Ae. albopictus*, and (f) body size of *Cx. pipiens* in response to UV-B conditions (FS, S, and NUV). Data were statistically tested using ANOVA. Significant pairwise comparison among treatment levels for main effects are indicated by different letters above bars.

