

The influence of rater training on inter-rater reliability when using the rat grimace scale

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Background. Rodent grimace scales facilitate evaluation of the affective component of pain and can identify a range of acute pain levels. Reported rater training in the use of these scales varies considerably and may contribute to observed variability in inter-rater reliability. This study evaluated the effect of training on inter-rater reliability with the Rat Grimace Scale (RGS). Methods. Two training sets, of 42 and 150 images, were prepared from several acute pain models. Four trainee raters, with no previous experience with the RGS, progressed through 2 rounds of training, first scoring 42 images (S1) followed by 150 images (S2a). After each round, trainees reviewed the RGS and any problematic images with an experienced rater. The 150 images were re-scored in a final round (S2b). Interrater reliability was evaluated using the intra-class correlation coefficient (ICC) and ICCs compared with a Feldt test. Results. Inter-rater reliability increased from moderate (ICC 0.58 [95%CI: 0.43-0.72]) to very good (ICC 0.85 [0.81-0.88]) between S1 and S2b (p < 0.01) with a significant increase also observed between S2a and S2b (p < 0.01). The ICCs for individual action units orbital tightening, ears and nose/cheek also improved from S1 to S2b (p < 0.01). The action units with the highest and lowest ICCs at S2b were orbital tightening (0.84 [0.80-0.87]) and whiskers (0.63 [0.57-0.70]), respectively. In comparison to an experienced rater the ICCs for all trainees improved, ranging from 0.88 to 0.91 at S2b. Discussion. Training improves inter-rater reliability between trainees, with an associated reduction in 95%CI. Additionally, training resulted in improved inter-rater reliability alongside an experienced rater. Training improves the scoring of individual action units though scoring of whiskers is more difficult that other sites. Conclusion. The beneficial effects of training potentially reduce data variability and improve experimental animal welfare.

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Abstract

Background. Rodent grimace scales facilitate evaluation of the affective 18 component of pain and can identify a range of acute pain levels. Reported 19 20 rater training in the use of these scales varies considerably and may contrib-21 ute to observed variability in inter-rater reliability. This study evaluated the effect of training on inter-rater reliability with the Rat Grimace Scale (RGS). 22 **Methods.** Two training sets, of 42 and 150 images, were prepared from sev-23 eral acute pain models. Four trainee raters, with no previous experience with 24 the RGS, progressed through 2 rounds of training, first scoring 42 images (S1) 25 followed by 150 images (S2a). After each round, trainees reviewed the RGS 26 and any problematic images with an experienced rater. The 150 images were 27 re-scored in a final round (S2b). Inter-rater reliability was evaluated using the 28 intra-class correlation coefficient (ICC) and ICCs compared with a Feldt test. 29 30 **Results.** Inter-rater reliability increased from moderate (ICC 0.58 [95%CI: 0.43-0.72) to very good (ICC 0.85 [0.81-0.88]) between S1 and S2b (p < 31 0.01) with a significant increase also observed between S2a and S2b (p < 32 0.01). The ICCs for individual action units orbital tightening, ears and 33 nose/cheek also improved from S1 to S2b (p < 0.01). The action units with 34 the highest and lowest ICCs at S2b were orbital tightening (0.84 [0.80-0.87]) 35 and whiskers (0.63 [0.57-0.70]), respectively. In comparison to an experi-36 enced rater the ICCs for all trainees improved, ranging from 0.88 to 0.91 at 37 S2b. 38



- 39 **Discussion.** Training improves inter-rater reliability between trainees, with
- 40 an associated reduction in 95%CI. Additionally, training resulted in improved
- 41 inter-rater reliability alongside an experienced rater. Training improves the
- 42 scoring of individual action units though scoring of whiskers is more difficult
- 43 that other sites.
- 44 **Conclusion.** The beneficial effects of training potentially reduce data variab-
- 45 ility and improve experimental animal welfare.



Introduction

4/	The effectiveness of a pain assessment scale lies in its validity (does a scale
48	measure what is intended) and reliability (measurement error). Rodent grim-
49	ace scales have renewed interest in measuring the affective component of
50	pain and have been promoted as a means of overcoming the shortfalls of
51	nociceptive threshold testing (Mogil & Crager, 2004; Langford, Bailey &
52	Chanda et al., 2010; Sotocinal, Sorge & Zaloum et al., 2011; Oliver, De
53	Rantere & Ritchie et al., 2014; De Rantere, Schuster & Reimer et al., 2016).
54	There is increasing evidence that grimace scales discriminate painful and
55	non-painful states in a range of acute pain models and interventions (Lang-
56	ford, Bailey & Chanda et al., 2010; Sotocinal, Sorge & Zaloum et al., 2011;
57	Oliver, De Rantere & Ritchie et al., 2014; De Rantere, Schuster & Reimer et
58	al., 2016; Leach, Klaus & Miller et al., 2012). However, there are conflicting
59	reports regarding reliability when multiple raters score images (Langford,
60	Bailey & Chanda et al., 2010; Sotocinal, Sorge & Zaloum et al., 2011; Oliver,
61	De Rantere & Ritchie et al., 2014; Faller, McAndrew & Schneider et al., 2015;
62	Mittal, Gupta & Lamarre et al., 2016). Factors contributing to this variability
63	may include a lack of structured training and variation in individual learning
64	curves (Campbell, Hecker & Biau et al. 2014; de Oliveira Filho, 2002; Roughar
65	& Flecknell, 2006).



It is unclear what level of training is required to attain proficiency in using 66 grimace scales. Most studies include minimal, non-specific descriptions of 67 training (Langford, Bailey & Chanda et al., 2010; Sotocinal, Sorge & Zaloum 68 et al., 2011; Oliver, De Rantere & Ritchie et al., 2014; Leach, Klaus & Miller et 69 70 al., 2012; Faller, McAndrew & Schneider et al., 2015; Mittal, Gupta & Lamarre et al., 2016) and few report any measure of reliability (Langford, Bailey & 71 Chanda et al., 2010; Sotocinal, Sorge & Zaloum et al., 2011; Oliver, De 72 73 Rantere & Ritchie et al., 2014; Mittal, Gupta & Lamarre et al., 2016). Trainees progress at different rates during training to achieve proficiency in a task 74 (Mittal, Gupta & Lamarre et al., 2016; Campbell, Hecker & Biau et al. 2014; 75 Roughan & Flecknell, 2006); therefore, in addition to training, some assess-76 77 ment of score reliability is necessary. The impact of training on scoring reliability with the Rat Grimace Scale (RGS) has not been formally evaluated. The 78 79 objective of this study was to assess the effect of training on inter-rater reliability when scoring was performed with single and multiple raters applying 80 the RGS. We hypothesized that training would improve inter-rater reliability. 81 **Materials and Methods** 82



Two sets of training images were created from images collected during an un-83 related project that had received institutional animal care and use committee 84 85 approval from the University of Calgary Health Sciences Animal Care Committee (protocol IDs: AC13-0161 and AC13-0124)(De Rantere, Schuster & Reimer 86 et al., 2016). This project used the following acute pain models: intraplantar 87 carrageenan or Complete Freund's adjuvant or plantar incision. Animals were 88 adult (> 10 weeks old) male Wistar (n = 34) rats, from a commercial source 89 90 (Charles River Laboratories, Canada). The methodology used to generate images was as previously described (So-91 92 tocinal, Sorge & Zaloum et al., 2011). Briefly, still images were captured from high-definition video-recordings and cropped so that only the face was vis-93 ible. Each image was presented on a single slide in presentation software 94 (Microsoft PowerPoint, version 14.0, Microsoft Corporation, Redmond, WA, 95 96 USA). Slide order was randomized and identifying information (animal ID, time point, model) removed. 97 98 Images were selected based on image quality alone, by an individual not involved with the study. Two unique sets of training images were created, of 42 99 100 (S1) and 150 (S2) images. Images were scored using the RGS (scale range 0-2 for each action unit) and the average score calculated from four action 101 units: orbital tightening, nose/cheek flattening, ear changes, and whisker 102 change. 103



- None of the 4 trainee raters recruited had previous experience with the RGS.
- All raters were female undergraduate and graduate students (age range 20-
- 25 years), studying veterinary medicine, biology (n = 2) and health sciences
- and recruited when joining the research group as project students.



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All raters followed the same scoring protocol: S1 images were scored independently by each individual, using the training manual provided by Sotocinal et al. (2011) (which contains prototypic images of each action unit at each score), with additional images from our laboratory (Supplementary Data S1). Raters were encouraged to record comments for any images they found difficult to score. Following S1 scoring, raters reviewed their scores as a group with an experienced rater (DP), discussing recorded comments and areas of inconsistency. Images with the most variation between raters were selected for review. The primary goal of the discussion was to improve standardisation of scoring images assigned a score of 0 or 2. Disagreement in scores was tolerated provided differences between raters did not exceed 1 point on the scale. The standard of scoring was set by the experienced rater, following establishment of the technique within the laboratory with the support of the Mogil laboratory (McGill University). Once review of S1 scoring was complete, S2 images were scored independently by each individual and comments recorded as before (S2a). The S2 image set was then scored independently a second time (S2b) after a facilitated group discussion with the experienced rater (as per the S1 image set discussion). Approximately 15-30 images were reviewed during group discussions, with 2-3 weeks between reviews.



Intraclass correlation coefficients (ICCs, MedCalc version 12.6.1.0, MedCalc 128 Software, Ostend, Belgium) were calculated to measure the reliability of RGS 129 130 scoring between raters for the individual action unit scores and average RGS 131 scores. An absolute model was used for the ICC calculation and single meas-132 ure reported. This was done for each dataset (S1, S2a, S2b). ICCs were also calculated for the comparison between individual rater scores and those of 133 the experienced rater (DP) to determine reliability of an individual rater. Cal-134 culated ICCs were compared with a Feldt test (critical F set at alpha = 0.01135 and differences considered significant if the observed F value was greater 136 than the critical F value) (Feldt, Woodruff & Salih, 1987; Kuzmic, 2015). Inter-137 pretation of the ICC followed the same divisions as used previously: "very 138 139 good" (0.81-1.0), "good" (0.61-0.80), "moderate" (0.41-0.60), "fair" (0.21-0.40), "poor" (< 0.20)(Oliver, De Rantere & Ritchie et al., 2014). During the 140 141 training process, raters were said to be proficient when calculated ICCs \pm 142 95%CI overlapped with those published in a study reporting inter-rater reliab-143 ility(Oliver, De Rantere & Ritchie et al., 2014). To assess the potential impact of scores memorised during group discussion between S2a and S2b introdu-144 cing bias in to the ICC calculation for S2b, images with the greatest scoring 145 variability at S2a (those with a difference of 2 points between any 2 raters 146 and therefore the most likely to have been discussed) were removed and the 147 ICCs for S2b recalculated. Data are presented as ICC (± 95%CI) and a correc-148 ted p value for multiple comparisons of ≤ 0.017 was considered significant. 149 Scoring accuracy was assessed by comparing scores for images collected at 150 baseline and 6-9 hours after treatment (when a peak in RGS scores could be 151



- expected for the models studied (De Rantere, Schuster & Reimer et al.,
- 2016); paired t test with alpha set at 0.05). The datasets generated from this
- study are available in the Harvard Dataverse repository (Pang, 2018).

Results

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- Four raters completed the study. All training images were scored by every
- rater, and all scores included in the subsequent analysis.
- 158 Training was associated with a progressive improvement in inter-rater reliabil-
- ity and narrowing 95%CI (Fig. 1). The first training round (S1) resulted in a
- moderate ICC for the average RGS scores, with wide 95%CI (0.58 [0.43-
- 161 0.72]). The increase in average RGS ICC between S1 and S2a (0.68 [0.58-
- 162 0.76]) was not statistically significant ($F_{0.01:149.41} = 1.88$, observed F = 1.31, p
- > 0.05). A significant improvement was observed at S2b (0.85 [0.81-0.88])
- 164 compared with S1 (observed F = 2.8) and S2a ($F_{0.01:149.149} = 1.47$, observed F
- = 2.13, p < 0.01 for both comparisons). The resultant S2b ICC was classified
- as very good and comparable with published values (Fig. 1)(Oliver, De
- 167 Rantere & Ritchie et al., 2014).



A similar pattern of improvement was observed in the scores of individual ac-168 tion units (Table 1). Significant increases in ICCs were observed between S1 169 and S2b for orbital tightening (observed F = 1.94), ear changes (observed F170 = 2.14) and nose/cheek flattening (observed F = 2.21, p < 0.01 all comparis-171 172 ons), but not whisker changes (observed F = 1.65, p > 0.05). And between S2a and S2b: orbital tightening (observed F = 1.81), ear changes (observed F173 = 1.96) and nose/cheek flattening (observed F = 1.72, p < 0.01 all comparis-174 ons), but not whisker changes (observed F = 1.35, p > 0.05). At all stages, or-175 bital tightening had the highest ICC, improving from 0.69 to 0.84. Following 176 177 training, ICCs for individual action units fell within the good or very good range (Table 1). 178 179 Comparing individual rater performance against the experienced rater showed considerable variation following the first training round with ICCs ran-180 181 ging from fair to good. All trainee raters showed improvement with training (Table 2). 182 183 There were 28 images (19%) with score differences between raters of 2 points at S2a. Removing these scores had a minimal effect on the recalcu-184 lated ICCs for S2b; the 95%CI of the ICCs overlapped with those for the full 185 data set (Supplementary Data S2). 186 There was a significant increase in RGS scores between baseline (n = 41, 187 0.45 ± 0.07) and 6-9 hours after treatment (n = 29, 0.92 ± 0.08, p < 0.001, 188 95%CI of mean difference 0.27 to 0.68, Supplementary Data S3). 189



Discussion

l 9 1	Little is known regarding the need for, or role of, rater training in the use of
192	rodent grimace scales. Where training has been described, it ranges from re-
193	viewing the grimace scale training manuals (Leach, Klaus & Miller et al.,
194	2012; Faller, McAndrew & Schneider et al., 2015) to a single training session
195	of variable length (Langford, Bailey & Chanda et al., 2010; Sotocinal, Sorge &
196	Zaloum et al., 2011; Oliver, De Rantere & Ritchie et al., 2014; De Rantere,
197	Schuster & Reimer et al., 2016) or multiple training sessions (Mittal, Gupta &
198	Lamarre et al., 2016). Few studies describe an assessment of reliability
199	(Langford, Bailey & Chanda et al., 2010; Sotocinal, Sorge & Zaloum et al.,
200	2011; Oliver, De Rantere & Ritchie et al., 2014; Mittal, Gupta & Lamarre et
201	al., 2016). The results of this study show that an assessment of reliability is
202	necessary to confirm that training will lead to proficiency.
203	Our results suggest that reliability is limited when training is limited to re-
204	viewing the training manual, improving when feedback and discussion with
205	an experienced rater are included.



The rate at which individuals achieve proficiency in a task is highly variable 206 and, as such, it is erroneous to assume that training guarantees proficiency. 207 208 Neither a single training session nor repeated attempts at a task ensure proficiency (Campbell, Hecker & Biau et al. 2014; de Oliveira Filho, 2002; Roughan 209 210 & Flecknell, 2006). A simple method of evaluating rater proficiency is to assess inter-rater reliability (Streiner & Norman, 2008). This provides assurance 211 that variability in scoring is at an acceptable level and enables roque raters 212 to be identified (Mittal, Gupta & Lamarre et al., 2016; Brondani, Mama & 213 Luna; 2013). Identification of such raters during training allows for further 214 testing and assessment or removal from participation in scoring (Mittal, 215 Gupta & Lamarre et al., 2016). Ensuring reliability will reduce data variability 216 217 and consequently, animal use. An alternative approach is to use a single rater; however, it is still useful to compare the performance of a single rater 218 against that of an experienced rater, or a standard set of scores, to confirm 219 reliability and consistency over time (Oliver, De Rantere & Ritchie et al., 220 221 2014). The presence of systematic bias may negatively affect data interpretation and pain management (Faller, McAndrew & Schneider et al., 2015). 222 Orbital tightening had the highest associated ICC following the initial round of 223 scoring, which was maintained throughout training. In contrast, the reliability 224 of whisker scoring remained relatively low throughout training. These results 225 support previous findings that assessing the whisker change action unit is 226 more difficult for raters than orbital tightening (Oliver, De Rantere & Ritchie 227 et al., 2014). 228



A limitation of this study was re-scoring the 150 image set in the final training round, with the potential for memorised scores assigned during the group discussion following the second training round being applied rather than a rater scoring independently. We feel this is unlikely due to the large number of images scored, the similar appearance of rodent faces from similar strains, the time elapsed between review rounds, the small number of images reviewed during group discussion and the nature of the group discussion, where disagreement between raters was acceptable. The minimal difference in ICCs after removal of the 28 image scores supports this assertion.

Images for training were selected on the basis of quality rather than to allow comparison between treatment groups. This limits any assessment of construct validity but the comparison of baseline and predicted peak pain periods indicates that accuracy was preserved.

Conclusion

These data show that reliance on access to the available manuals for rater training may be insufficient. Formal training improves inter-rater reliability and is likely to reduce data variability if rater proficiency is assessed before embarking on data collection. Collaborative training between research groups would ensure similar levels of rater proficiency and improve the reproducibility of research. Inclusion of clear descriptions of rater training and assessment would help in evaluating study results.



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- 255 versity of Calgary) for statistical advice.



256 **References**

- 257 Brondani JT, Mama KR, Luna SP, Wright BD, Niyom S, Ambrosio J, Vogel PR,
- Padovani CR. 2013. Validation of the English version of the UNESP-Botucatu
- 259 multidimensional composite pain scale for assessing postoperative pain in
- 260 cats. BMC Vet Res 9:143. DOI:10.1186/1746-6148-9-143.
- 261 Campbell RD, Hecker KG, Biau DJ, Pang DS. 2014. Student attainment of pro-
- 262 ficiency in a clinical skill: the assessment of individual learning curves. *PLoS*
- 263 One 9:e88526. DOI: 10.1371/journal.pone.0088526.
- 264 de Oliveira Filho GR. 2002. The construction of learning curves for basic skills
- in anesthetic procedures: an application for the cumulative sum method.
- 266 Anesth Analg 95:411-416.
- De Rantere, D, Schuster CJ, Reimer JN, Pang DS. 2016. The relationship be-
- 268 tween the Rat Grimace Scale and mechanical hypersensitivity testing in three
- 269 experimental pain models. Eur J Pain 20:417-426. DOI: 10.1002/ejp.742.
- Faller KM, McAndrew DJ, Schneider JE, Lygate CA. 2015. Refinement of analge-
- 271 sia following thoracotomy and experimental myocardial infarction using the
- 272 Mouse Grimace Scale. *Exp Physiol* 100:164–172. DOI:
- 273 **10.1113/expphysiol.2014.083139**.
- 274 Feldt LS, Woodruff DJ and Salih FA. 1987. Statistical inference for coefficient
- alpha. *Appl Psychol Meas* 11:93-103.
- 276 Kuzmic P. 2015. Critical values of F-statistics. Available at http://www.biokin.-
- 277 com/tools/f-critical.html (accessed 26 February 2018).



- 278 Langford DJ, Bailey AL, Chanda ML, Clarke SE, Drummond TE, Echols S, Glick
- 279 S, Ingrao J, Klassen-Ross T, Lacroix-Fralish ML, Matsumiya L, Sorge RE, Sotoci-
- 280 nal SG, Tabaka JM, Wong D, van den Maagdenberg AM, Ferrari MD, Craig KD,
- 281 Mogil JS. 2010. Coding of facial expressions of pain in the laboratory mouse.
- 282 Nat Methods 7:447-449. DOI: 10.1038/nmeth.1445.
- 283 Leach MC, Klaus K, Miller AL, Scotto di Perrotolo M, Sotocinal SG, Flecknell PA.
- 284 2012. The assessment of post-vasectomy pain in mice using behaviour and
- the Mouse Grimace Scale. *PLoS One* 7:e35656. DOI:
- 286 10.1371/journal.pone.0035656.
- 287 Mittal A, Gupta M, Lamarre Y, Jahagirdar B, Gupta K. 2016. Quantification of
- 288 pain in sickle mice using facial expressions and body measurements. *Blood*
- 289 Cells Mol Dis 57:58-66. DOI: 10.1016/j.bcmd.2015.12.006.
- 290 Mogil JS and Crager SE. 2004. What should we be measuring in behavioral
- 291 studies of chronic pain in animals? J Pain 112:12-15. DOI:
- 292 10.1016/j.pain.2004.09.028.
- 293 Oliver V, De Rantere D, Ritchie R, Chisholm J, Kecker KG, Pang DS. 2014. Psy-
- 294 chometric assessment of the Rat Grimace Scale and development of an anal-
- 295 gesic intervention score. *PLoS One* 9:e97882. DOI:
- 296 10.1371.journal.pone.0097882.
- Pang DS. 2018. Rat Grimace Scale rater training data 1.0. Available at
- 298 https://doi.org/10.7910/DVN/57K7PE (accessed 26 February 2018).



- 299 Roughan JV and Flecknell PA. 2016. Training in behaviour-based post-opera-
- 300 tive pain scoring in rats an evaluation based on improved recognition of
- analgesic requirements. Appl Anim Behav Sci 96:327-342. DOI: 10.1016/j.ap-
- 302 planim.2005.06.012.
- 303 Sotocinal SG, Sorge RE, Zaloum A, Tuttle AH, Martin LJ, Wieskopf JS, Mapple-
- beck JC, Wei P, Zhan S, Zhang S, McDougall JJ, King OD, Mogil JS. 2011. The
- Rat Grimace Scale: a partially automated method for quantifying pain in the
- laboratory rat via facial expressions. *Mol Pain* 7:55. DOI: 10.1186/1744-8069-
- 307 **7-55**.
- 308 Streiner DL and Norman GR. 2008. Reliability. In: Streiner DL and Normal GR,
- 309 eds. Health Measurement Scales. New York: Oxford University Press, 167-
- 310 **210**.

311 Legends

- Figure 1. Average group ICCs for each of the three datasets (with 95%CI) with
- reference values (Oliver, De Rantere & Ritchie et al., 2014).
- 314 Supplementary Data_S3. Bar graph (mean ± SEM) showing RGS scores at
- baseline (n = 41 images) and 6-9 hours after treatment (n = 29 images: in-
- 316 traplantar Complete Freund's Adjuvant; n = 19 images, plantar incision; n =
- 317 10 images). Broken horizontal line indicates derived analgesia intervention
- threshold (Oliver, De Rantere & Ritchie et al., 2014).



Table 1(on next page)

Group Intra-class Correlation Coefficients (ICC) for each of the datasets.

S1, S2a and S2b are the first, second and third training round, respectively. Data are ICCsingle [95%CI]. Within a row, identical superscript letters indicate significant differences between the different training rounds, p < 0.01. Reference values are from Oliver, De Rantere, Ritchie et al. (2014).



- 1 Table 1. Group Intra-class Correlation Coefficients (ICC) for each of the
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- 3 spectively. Data are ICCsingle [95%CI]. Within a row, identical superscript let-
- 4 ters indicate significant differences between the different training rounds, p
- 5 < 0.01. Reference values are from Oliver, De Rantere, Ritchie et al. (2014).

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Action Unit	S1	S2a	S2b	Reference values
Orbital tighten-	0.69 [0.56-	0.71 [0.63-	0.84 [0.80-	0.92 [0.89-
ing	0.80]ª	0.78] ^b	0.87] ^{a,b}	0.95]
Ear changes	0.40 [0.25-	0.45 [0.35-	0.72 [0.66-	0.62 [0.51-
	0.56] ^a	0.54] ^b	0.77] ^{a,b}	0.72]
Nose/Cheek flat-	0.36 [0.21-	0.50 [0.41-	0.71 [0.65-	0.62 [0.51-
tening	0.52] ^a	0.58] ^b	0.76] ^{a,b}	0.72]
Whisker change	0.39 [0.26-	0.50 [0.42-	0.63 [0.57-	0.52 [0.39-
	0.55]	0.58]	0.70]	0.63]



Table 2(on next page)

Agreement of individual raters when compared to an experienced rater (DP).

Data are ICCsingle [95%CI]. Within a column, matching superscript letters indicate significant differences (p < 0.01).



Table 2: Agreement of individual raters when compared to an experienced rater (DP). Data are ICCsingle [95%CI]. Within a column, matching superscript letters indicate significant differences (p < 0.01).

Image set	Rater 1 vs DP	Rater 2 vs DP	Rater 3 vs DP	Rater 4 vs DP
S 1	0.41 [0.06-	0.70 [0.50-	0.62 [0.36-	0.42 [0.13-
	0.66] ^{a,b}	0.83] ^a	0.79] ^a	0.64] ^a
S2a	0.84 [0.79-	0.75 [0.68-	0.68 [0.25-	0.65 [0.38-
	0.88] ^a	0.82] ^b	0.84] ^b	0.79] ^b
S2b	0.89 [0.85-	0.88 [0.84-	0.91 [0.88-	0.90 [0.87-
	0.92] ^b	0.91] ^{a,b}	0.94] ^{a,b}	0.93] ^{a,b}



Figure 1

Average group ICCs for each of the three datasets (with 95%CI) with reference values (Oliver, De Rantere & Ritchie et al., 2014).

