Pee	Preprints NOT PEER-REVIEWE
1	Title Page
2	
3	Short title: Reinstatement of Pedicularis delavayi
4	
5	Reinstatement of Pedicularis delavayi (Orobanchaceae): a species
6	separating from <i>P. siphonantha</i> revealed by phylogenetic analyses
7	
8	Wen-Bin Yu ^{1*} , Hong Wang ^{2, 3*} , Min-Lu Liu ² , Alisa E. Grabovskaya ⁴ , and De-Zhu Li ^{2, 3*}
9	
10	1 Center for Integrative Conservation, and Southeast Asia Biodiversity Research Institute, Xishuangbanna
11	Tropical Botanical Garden, Chinese Academy of Sciences, Mengla 666303, China
12	2 Key Laboratory for Plant Diversity and Biogeography of East Asia, Kunming Institute of Botany, Chinese
13	Academy of Sciences, Kunming 650201, China
14	3 Plant Germplasm and Genomics Center, Germplasm Bank of Wild Species, Kunming Institute of Botany,
15	Chinese Academy of Sciences, Kunming 650201, China
16	4 Komarov Botanical Institute, Prof. Popov str., 2 197376, St. Petersburg, Russia
17	

18 *Correspondences: yuwenbin@xtbg.ac.cn, waghong@mail.kib.ac.cn, dzl@mail.kib.ac.cn

NOT PEER-REVIEWED

Peer Preprints

1 ABSTRACT

- Background. Morphological identification of *Pedicularis* depends on floral characters because
 interspecific variations these characters is very large. However, some important characters may be lost
 during the process of pressing the specimen. *Pedicularis delavayi* was described from Lijiang, northwestern
 Yunnan, and it was widely adopted as a variety of *P. siphonantha*. Unfortunately, the name "*P. siphonantha*var. *delavayi* incorrectly referred to *P. milliana/P. tenuituba* in some identified specimens and publications.
 Moreover, phylogenetic relationships between *P. delavayi* and *P. siphonantha* and its allies (*P. millana* and
- 8 *P. tenuituba*) were not fully resolved.
- 9 Methods. In total, we sampled 75 individuals representing 56 taxa. Of them, 10 taxa were from P.
- 10 siphonantha group, and 11 individuals of P. delavayi represented 9 populations. One nuclear gene (nrITS)
- 11 and four chloroplast genes/regions (matK, rbcL, trnH-psbA, and trnL-F) were sequenced for phylogenetic
- 12 analyses. The nrITS and plastid datasets were combined to analyze. Bayesian Inference (BI) and Maximum
- 13 Likelihood (ML) methods were used to reconstruct phylogenetic trees. Species identification, morphological
- comparison and line drawing were based on field observation, dried herbarium specimens and FFA-fixedsamples.
- 16 **Results.** Phylogenetic analyses showed that the *P. siphonantha* group was polyphyletic: monophyletic *P.*
- 17 *delavayi*, sister to *P. obliquigaleata*, fell into clade A; and the remaining species of *P. siphonantha* group
- 18 was in clade B as monophyletic (i.e. *P. siphonantha* lineage). In the *P. siphonantha* lineage, *P. milliana*, *P.*
- 19 *siphonantha*, and *P. tenuituba* (= *P. siphonantha* var. *stictochila*) were well supported as monophyletic, and
- 20 P. dolichosiphon (= P. siphonantha var. dolichosiphon) was sister to P. leptosiphon. Morphologically, P.
- 21 *delavayi* differs from species of the *P. siphonantha* lineage in having a long petiole (~ 50 mm) and pedicel
- 22 (\sim 40 mm), a ridged corolla tube, and a folded lower-lip of the corolla.
- Discussion. Phylogenetic analyses showed evolution of *P. delavayi* was independent from the *P. siphonantha* lineage. The long-tubular corollas in *P. delavayi* and *P. siphonantha* lineage might be a
 convergent character. Both morphological and phylogenetic evidence strongly supported to reinstate *P. delavayi* as an independent species. In addition, *P. neolatituba* was proposed to reduce as a new synonymy of *P. delavayi*.
- 28
- 29 Keywords: Pedicularis, Pedicularis delavayi, Pedicularis milliana, Pedicularis siphonantha, species
- 30 delimitation, the Hengduan Mountains

1 INTRODUCTION

Flowers of *Pedicularis* L. (Orobanchaceae) show striking interspecific variations (Li 1951; Yu et al. 2008), 2 so morphological identification of these species depends much on floral characters (Liu, Yu & Wang 2013). 3 4 Generally, one species is easily distinguished from another morphologically similar species using fresh flowers in the field. However, floral shape and structure may be changed during the process of pressing and 5 drying the specimen and, in practice, herbarium specimens of closely related species are very difficult to 6 7 discriminate. DNA barcodes have been widely applied to assist species identification (Hebert et al. 2003; 8 Kress et al. 2005; China Plant BOL Group 2011), particularly when morphological identification is 9 uncertain. In *Pedicularis*, the nuclear ribosomal internal transcribed spacer (nrITS) or nrITS+rbcL can discriminate at least 78% of species in the genus (Yu et al. 2011; Liu, Yu & Wang 2013). However, there is 10 little consensus between the phylogenetic tree and traditional classification in *Pedicularis* (Ree 2005; Yang 11 & Wang 2007; Tkach et al. 2014; Robart et al. 2015; Yu et al. 2015), and morphologically similar species 12 may not be sister to each other in phylogenetic analyses. Therefore, DNA sequences are very useful to 13 14 delimit species and to confirm phylogenetic relationship among species.

15 Pedicularis delavayi was firstly named by Franchet after J. M. Delavay, who collected the type material (Fig. S1) at Yulong Mountain in Lijiang, northwestern Yunnan, China in 1886, while it was validly 16 published by Maximowicz (1888). This species was treated as an independent species (Prain 1890; Bonati 17 1910; Limpricht 1924; Li 1949) until Tsoong (1963), who downgraded it to a variety in P. siphonantha D. 18 Don. According to illustrations in Chinese Floras (Tsoong 1963; Yang, Holmgren & Mill 1998), the lower 19 20 lip of *P. siphonantha* var. *delavavi* (Franch. ex Maxim.) P. C. Tsoong should be similar to *P. siphonantha* D. 21 Don var. siphonantha as spreading (see Fig. 1A). From illustrated publications, the name "P. siphonantha var. delavayi was used for a "long-tubed and purple-red species" (Fig. 1B), which is a common species in 22 23 alpine meadow at altitudes from 3000 m to 4000 m a.s.l. (above sea level, hereafter) in northwestern Yunnan (e.g., Guan 1998; Yu, Zhang & Wang 2008; Xu, Li & Sun 2013). In addition, some publications 24 used the name "P. siphonantha" for this "long-tubed and purple-red species" (e.g., Yang et al. 2002; Yang, 25 26 Sun & Guo 2005; Huang & Fenster 2007; Yu et al. 2013a; Huang, Wang & Sun 2016).

27 During field expeditions for *Pedicularis* in the Hengduan Mountains region from 2006 to 2010, we 28 collected an unknown long-tubed species with a purple-red corolla in Shangri-La, northwestern Yunnan, and Jiulong and Kangding, Sichuan, at altitudes around 4000 m (Figs. 1C-F). This species differs from 29 infraspecfic taxa on P. siphonantha and other long-tubed and purple-red species in series Longiflorae Prain 30 31 by having an incurved middle lobe of the lower lip and a ridged corolla tube (Figs. 1G, H). DNA barcoding 32 showed that samples of this species were separated from *P. siphonantha* (Yu et al. 2011). We considered 33 that this species may be new until we checked the type materials of *P. delavayi* conserved at the herbaria of 34 the V. L. Komarov Botanical Institute in St. Petersburg (LE) (Fig. S1) and the Muséum National d'Histoire Naturelle in Paris (P). Based on morphological comparisons of specimens, we found that our specimens 35 were very similar to the type of P. delavayi. In order to clarify the taxonomical confusion, we examined 36 herbarium specimens of *P. siphonantha* collected from Yulong Mountain (type location of *P. delvavi*) 37 38 conserved at the herbaria of CAS Kunming Institute of Botany (KUN) and CAS Institute of Botany (PE).

NOT PEER-REVIEWED

We found that specimens labeled as "P. siphonantha var. delavayi" included two taxa: one is similar to the 1 2 type of *P. delavayi*, and another is the "long-tubed and purple-red species" (Fig. 1B), a common species in 3 northwestern Yunnan. Indeed, it is very difficult to discriminate the herbarium specimens as two taxa. Based on the field investigations, we found that P. delavayi differed from the "long-tubed and purple-red 4 species" by having a long petiole and pedicel, inflated calyx tube in the middle upper parts, and folded 5 6 lower lip of the corolla, as well as occurring at altitudes over 3600 m a.s.l. During recent field expeditions, specimens of *P. delavavi* were collected from the Yulong Mountain at over 4000 m a.s.l (Fig. 1F), and those 7 of the "long-tubed and purple-red species" between 3600 m and 4000 m a.s.l. (Fig. 1B). Therefore, we 8 confirmed that P. delavayi and the "long-tubed and purple-red species" were two separated species, and the 9 "long-tubed and purple-red species" should be an undescribed species. To reduce descriptive confusion, we 10 11 proposed a name "P. millina W. B. Yu & H. Wang" for this taxon in following statements and discussion.

12 Recently, a comprehensive phylogeny of Chinese Pedicularis shows that P. siphonantha is a polyphyletic group, var. delavavi ($\equiv P. delavavi$) and other varieties of P. siphonantha falling into two 13 subclades in clade 3 (Yu et al. 2015). Pedicularis delavayi was sister to P. obliquigeleata in subclude A. In 14 the subclade 3B, var. siphonantha and var. stictochila H. Wang & W.B. Yu (= P. tenuituba H.L. Li), and 15 16 five species from series *Longiflorae* with purple/red/pink corollas with twisted beaks formed a strongly supported lineage, i.e. the *P. siphonantha* lineage (that is molecular delimitation). In the present study, we 17 made extensive sampling of P. delavayi from 9 populations (10 new samples), and two additional samples 18 19 of P. millina from Yulong Mountains in Lijiang, one additional new samples of P. tenuituba, and all 20 recognized taxa in clade 3 and the sister P. axillaris Franch. ex Maxim. (Yu et al. 2015). DNA sequences from nrITS and four chloroplast regions (*matK*, *rbcL*, *trnH-psbA* and *trnL-F*) were generated and analyzed. 21 Our main goal was to evaluate the monophyly of P. delavayi, and its phylogenetic relationship to the P. 22 siphonantha lineage, in particular to P. milliana and P. tenuituba. If monophyletic P. delavavi was excluded 23 from the *P. siphonantha* lineage, *P. delavayi* should be reinstated as an independent species. 24

25

26 **MATERIAL AND METHODS**

27 Plant samplings. In total, we sampled 75 individuals representing 56 taxa, including all taxa were identified in clade 3 and its sister P. axillaris (Yu et al. 2015). Pedicularis siphonantha group, based on 28 morphological delimitation, consisted of 11 species (Table S1). Of them, four species P. delavayi, P. 29 30 milliana (new species), P. siphonantha, and P. tenuituba (= P. siphonantha var. stictochila) have a wide 31 distribution range. The remaining seven species only collected once or few gatherings around the type 32 locality: 1) P. sigmoidea Franch. ex Maxim. were found in around Eryuan and Lijiang, northwestern Yunnan; 2) P. dolichosiphon (Hand.-Mazz.) H.L.Li) (≡ P. siphonantha var. dolichosiphon Hand.-Mazz.), P. 33 dolichantha Bonati, P. leptosiphon H. L. Li, P. variegata H. L. Li and P. humilis Bonati were recollected 34 from the type locality; and 3) P. fastigiata Franch. only had the type, which did not include in this study. In 35 36 this study, we chose 11 samples of P. delavayi (three samples from the type locality Yulong Mountain, 37 Lijiang), and three samples of *P. milliana* and *P. tenuituba* (Table S2). Natural population of *P. humilis* was 38

just rediscovered in 2015 (Li et al. 2016). It is the first time to include this species for phylogenetic

1 analyses.

Fresh leaf tissues were collected in the field and preserved in silica gel. All DNA samples and voucher
specimens are stored at the Germplasm Bank of Wild Species and the herbarium of CAS Kunming Institute
of Botany (KUN), respectively. There are 284 sequences from 64 individuals which have been published in
other studies (China Plant BOL Group 2011; Yu et al. 2011; Liu & Yu 2015; Yu et al. 2015). In this study,
we generated 62 new sequences from 23 individuals (with 11 new individuals). A conspectus of voucher
information is presented in Table S2.

8

9 Specimen examination and identification. Fresh specimens were observed in the field. Fresh flowers
10 were collected and fixed in FAA solution. Herbarium specimens from the herbaria CDBI, KUN, LE, and
11 PE were examined and identified, and digital images of types from the herbaria E, K and P were accessed
12 online. Flower and fruits characters in the line drawings of *P. delavayi* were based on field photos and
13 FAA-preserved flowers.

14

Molecular methods. For the 11 new samples, total genomic DNA was extracted from silica gel-dried tissue using a modified 2× CTAB method. Five DNA loci, one nuclear gene (nrITS) and four chloroplast genes/regions (*matK*, *rbcL*, *trnH-psbA*, and *trnL-F*), were sequenced in this study. Primer information for the five loci were presented in previous studies (Yu et al. 2011; Yu et al. 2013b). Protocols for polymerase chain reaction (PCR) amplification and sequencing followed the study of (Yu et al. 2011).

20

Sequence assembly and alignment. The newly obtained raw sequences were assembled and edited using
Geneious version 7.1 (Kearse et al. 2012). The nrITS is a multiple copy region. These copies showed
evolutionary consistent in the sequenced 85 samples, only three samples had one ambiguous basecall (i.e.
multiple superimposed peaks in chromatograms). Two samples, LID1199 and HW10244, belonging to *P. tenuituba*, and sample HW10163 is *P. milliana*. The ambiguous site was assigned using IUPAC ambiguity
characters.

Preliminary alignments were automatically aligned using MAFFT version 7.2 (Katoh & Toh 2010),
then adjusted manually in Geneious. The aligned matrix was concatenated to a combined matrix using
SequenceMatrix version 1.73 (Vaidya, Lohman & Meier 2011). Sequence characteristics were calculated
using MEGA version 6.0 (Tamura et al. 2013).

31

Phylogenetic analyses. Bayesian Inference (BI) and Maximum Likelihood (ML) methods were used to reconstruct phylogenetic trees. The nrITS and plastid datasets were combined to analyze. No nucleotide positions were excluded from analyses. Partitioned BI analyses were performed using MrBayes (Ronquist & Huelsenbeck 2003), with DNA substitution models selected for each gene partition by the Bayesian information criterion (BIC) using jModeltest (Guindon & Gascuel 2003; Darriba et al. 2012). Markov Chain Monte Carlo (MCMC) analyses were run in MrBayes for 10,000,000 generations for each dataset, with two simultaneous runs, and each run comprising four incrementally heated chains. The BI analyses were started

5/14

NOT PEER-REVIEWED

with a random tree and sampled every 1000 generations. Number of generations for the three datasets were 1 2 sufficient, because the average standard deviation of split frequencies for the datasets was lower than 0.005 3 (0.002900), and Potential Scale Reduction Factor of Convergence Diagnostic (Gelman & Rubin 1992) for the datasets was 1.00. The first 25% of the trees was discarded as burn-in, and the remaining trees were used 4 5 to generate a majority-rule consensus tree. Posterior probability values (PP) ≥ 0.95 were considered as well 6 supported (Alfaro, Zoller & Lutzoni 2003; Erixon et al. 2003; Kolaczkowski & Thornton 2007). The ML 7 tree searches and bootstrap estimation of clade support were conducted with RAxML (Stamatakis, Hoover & Rougemont 2008). These analyses used the GTR substitution model with gamma-distributed rate 8 heterogeneity among sites and the proportion of invariable sites estimated from the data. The dataset was 9 partitioned by genes. Support values for the node and clade were estimated from 1000 bootstrap replicates. 10 11 Bootstrap support (BS) \geq 70 are considered well supported (Hillis & Bull 1993). Both BI and ML analyses, 12 as well as jModeltest, were performed at the CIPRES Science Gateway (http://www.phylo.org).

13

14 **RESULTS**

Information of DNA sequences. Sequence characteristics of five DNA regions and the concatenated datasets are summarized in Table 1. In the total dataset, the numbers of variable and parsimony informative sites were highest for nrITS, followed by *trnH-psbA*, *trnL-F*, *matK* and *rbcL*. For three selected groups, three spacers (nrITS, *trnH-psbA* and *trnL-F*) were more variable and informative than two coding genes (*matK* and *rbcL*), then *matK* was more than *rbcL*. One exception for *P. delavayi*, alignment of *matK* had only one variable site in the 11 individuals, whereas alignment of *rbcL* had three variable and two informative sites, respectively.

22

23 Phylogenetic analyses. The BI tree using the total dataset was presented in Figure 2. The topology was 24 similar to that in previous study (Yu et al. 2015). Two major clades were recovered, named as A and B 25 following Yu et al. (2015). Pedicularis delavayi fell into clade A, and P. siphonantha lineage was in clade B. 26 Both P. delavavi (BS/PP = 100/1.00) and P. siphonantha lineage (BS/PP = 96/1.00) were strongly as 27 monophyletic, respectively. In the clade of *P. delavayi*, three Lijiang samples (type locality) formed a group 28 (BS/PP = 98/1.00), which was weakly supported sister to the remaining eight samples (PP = 0.62); three Sichuan samples were strongly supported as monophyletic (BS/PP = 92/1.00), and two Yunnan samples 29 (HW10130 and HW10172) as sister. The P. siphonantha lineage split in two groups. One group included 30 clade P. dolichosiphon + P. leptosiphon (BS/PP = 100/1.00), and monophyletic P. siphonantha (BS/PP = 31 32 100/1.00) and P. tenuituba (= P. siphonantha var. stictochila) (BS/PP = 100/1.00). Another group comprised 33 of the remaining five sampled species (including P. humilis) and sample LIDZ1518. Three samples of P. *millina* from Lijiang were monophyletic by moderate supporting (BS/PP = 55/0.88), then the Lijiang 34 sample (LIDZ1584) of *P. sigmoidea* was resolved as sister (BS/PP = 99/1.00), followed by the Ervuan 35 sample (YWB2015059) of *P. sigmoidea* (BS/PP = 100/1.00). *Peducularis humilis* nested with sample 36 LIDZ1518 (BS/PP = 84/0.79), with long branch length, and P. variegata was sister to them (BS/PP = 37 38 90/1.00).

NOT PEER-REVIEWED

Morphologically, P. delavayi is similar to P. siphonantha by in having Morphological comparisons. 1 large and bi-lobed middle lobe of lower-lip, and semi-circle and crestless beak. However, the middle lobe 2 3 of P. delavayi was significantly incurved (Figs. 1C-G; vs. spreading in P. siphonantha, Fig. 1A), which was crushed in herbarium specimens (e.g. Fig. S1). Based on comparisons of flowering specimens, we found 4 5 that P. delavayi also differed from P. siphonantha by having a long petiole (~ 50 mm) and pedicel (~ 40mm), a furfuraceous surface on the abaxial leaf blade, a ridged corolla tube, a folded lower-lip of the 6 7 corolla, and four pubescent filaments. In addition, we found that the type of *P. neolatituba* was very close 8 to specimens of *P. delavayi* and should be reduced to a synonym of *P. delavayi*.

Pedicularis milliana (Fig. 1C) is very similar to P. siphonantha by having smooth corolla tube, 9 spreading corolla lower-lip, large and bi-lobed middle lobe, and semi-circle and crestless beak. Because the 10 11 distribution of P. delavayi overlaps with P. milliana in northwestern Yunnan, Tsoong (1963) might have 12 misidentified P. milliana as P. delavayi. Therefore, he downgraded P. delavayi to a variety under P. siphonantha. Clearly, P. milliana was separated from P. siphonantha, which is strongly supported by 13 phylogenetic analyses. In addition, sample LIDZ1518 was similar to P. milliana in having spreading corolla 14 lower-lip, large and bi-lobed middle lobe, and semi-circle beak, and to P. sigmoidea in having spreading 15 16 corolla lower-lip, large and bi-lobed middle lobe, and crested beak. However, phylogenetic analyses 17 indicating it was a separated lineage, close P. humilis and P. variegata.

18

19 DISCUSSION

Phylogenetic delimitation of P. siphonantha group. Pedicularis siphonantha was firstly described from 20 Nepal (Don, Hamilton & Wallich 1825), which has been recognized as endemic to the Himalayan region 21 22 (Pennell 1935; Li 1949). According to current taxonomic treatments (Tsoong 1963; Yang, Holmgren & Mill 1998), P. siphonantha var. delavay and var. stictochila occur in the Hengduan Mountains region, i.e. 23 northwestern Yunnan, western Sichuan, and southeastern Qinghai. Pedicularis siphonantha var. 24 dolichosiphon was discovered in Muli region of southern Sichuan (Handel-Mazzetti 1923), then upgraded 25 to an independent species by Li (1949). In the Chinese edition of Flora Reipublicae Popularis Sinicae, 26 Tsoong (1963) did not mention P. dolichosiphon, or he might have overlooked this species. According to 27 28 current phylogenetic analyses, P. siphonantha was polyphyletc, delimitation of P. siphonantha group needed to revise. Firstly, P. siphonantha var. delavayi was close to P. obliquigaleata in clade A, whereas the 29 30 other taxa of P. siphonantha were included in the P. siphonantha lineage. Therefore, P. siphonantha var. 31 *delavavi* should be reinstated as an independent species. Then, the remaining three infraspecific taxa of P. siphonantha (var. dolichosiphon, var. siphonantha and var. stictochila) and P. leptosiphon formed a clade, 32 33 and var. dolichosiphon was strongly supported as sister to P. leptosiphon. Of them, P. siphonantha var. 34 siphonantha has a semicircle beak, and the other three taxa have S-shaped beak. Integrating geographical 35 distribution, we agree with the treatment by Li (1949) to adopt var. dolichosiphon and var. stictochila as independent species as P. dolichosiphon and P. tenuituba, respectively. 36 Infraspecific delimitation of *P. siphonantha* was not fully resolved. In a taxonomical revision, Prain 37

38 (1890) included *P. hookeriana* Wall. ex Benth. as a synonym of *P. siphonantha* var. siphonantha, and *P.*

NOT PEER-REVIEWED

elephas Boiss. and P. punctata Decne. as synonyms of var. brevituba Prain. Nevertheless, some 1 taxonomists treated *P. hookeriana* and *P. punctata* as independent species (Pennell 1943; Yamazaki 1988; 2 3 Husain, Garg & Agnihotri 2010), and have placed P. elephas close to P. rhinanathoides Schrenk (Maximowicz 1888; Pennell 1943). A comprehensive phylogeny of *Pedicularis* showed that both *P*. 4 hookeriana and P. punctata fell into the clade of P. siphonantha from the Himalayas (R. Ree, Personal 5 6 Communication). In the early of 1900s, Bonati added two varieties under *P. siphonantha*, var. prostrata 7 Bonati (1904) from Sikkim and var. birmanica Bonati (1921) from upper Burma. In a revision of Pedicularis from Bhutan, Mill (2001) pointed out that P. siphonantha var. prostrata was easily confused 8 with P. hookeriana, whereas this variety had broader and ovate leaves and shorter corolla tubes. For P. 9 siphonantha var. birmanica, we found its type materials were close to that of P. humilis. 10

11

12 Parallel evolution of long-tubular corollas in *Pedicularis*. During revision of *Pedicularis*, Li (1948, 1949, 1951) and Tsoong (1955, 1963) hypothesized that long-tubular corollas were independently evolved 13 14 at least six and ten times, respectively. Phylogenetic inferences supported their hypotheses that long-tubular corollas were independently derived from short-tubular corollas at least eight times (Ree 2005), or up to 21 15 16 times (Yu et al. 2015). Long-tubular species occurred in seven of 13 clades, plus two unresolved species P. 17 batangensis Franch. & Bur. and P. flexuosa Hook. f. (Yu et al. 2015). Series Longiflorae Prain included more than 20 long-tubular species from the Himalaya-Hengduan Mountains region (Li 1949; Tsoong 1963; 18 Yamazaki 1988; Mill 2001). Species of series Longiflorae fell into clade 3, however, this series was 19 20 supported as monophyletic (Yu et al. 2015; this study). Phylogenetic analyses tended to split series Longiflorae into four groups: a) P. siphonantha lineage, b) P. delavayi, c) P. longiflora, and 4) P. armata – P. 21 22 cranolopha group (including a short-tubular species, P. fletcherii). From morphological similarity and geographical distribution, P. delavayi and P. longiflora were close to P. siphonantha lineage and P. armata 23 -P. cranolopha group, respectively. However, phylogenetic evidence indicated that the four groups may 24 evolve independently. 25

Evolution of long-tubular corollas in *Pedicularis* were hypothesized to adopt long-tongued pollinators 26 (Li 1951). However, pollination observations showed that long-tubular species were exclusively pollinated 27 28 by bumblebees (Macior & Sood 1991; Macior & Tang 1997; Macior, Tang & Zhang 2001; Wang & Li 2005; Huang & Fenster 2007; Huang, Wang & Sun 2016). Long-tubular corollas are associated with beaked galea, 29 30 and beaked species rewards pollinators for pollen alone (Macior & Sood 1991; Yu et al. 2008). Due to 31 anthers are tightly enclosed by the beaked galea, long-tongued Lepidoptera are impossible to dislodge pollen from the tightly enclosed anthers. Only bumblebees can open concealed anthers from the beaked 32 galea using forelegs, and release pollen by vibrating wings in high speed, i.e. buzz-pollination (Yu, Li & 33 34 Wang 2012). When long-tongued pollinators driving evolution of long-tubular corollas was rejected, an 35 alternative hypothesis for enhancing pollination attractiveness was proposed (Macior & Sood 1991; Macior, Tang & Zhang 2001). However, pollinator attraction hypothesis was not supported by experiments on P. 36 siphonantha (corrected as P. milliana herein) and P. tricolor Hand.-Mazz. (Huang, Wang & Sun 2016). 37 38 Pollination treatments indicated that elongation of corolla tube (and pistil length) may put more selective

NOT PEER-REVIEWED

pressure for male-to-male competition during the pollen germination (Yang & Wang 2015). Moreover, 1 plants growing in more fertilized conditions can produce longer corolla tube (Huang, Wang & Sun 2016). 2 3 We suggested that evolution of long-tubular corollas may have some advantages in high altitudes, because most of long-tubular species occur in alpine meadow over 3000 m a.s.l. in the Himalya-Hengduan 4 Mountains region (Tsoong 1956, 1961). Such ecological factors may independently drive elongation of 5 6 corolla tube in different lineages. Subsequent diversification of lineage may be mainly induced by 7 geographical isolation. Pedicularis siphonantha lineage is one good example to illustrate geographical 8 isolation facilitating species divergence in the Himalya-Hengduan Mountains region (Yu et al. 2015).

9

10 Reinstatement of *Pedicularis delavayi*. Phylogenetic analyses strongly support P. delavayi as a 11 separated species, which is sister to P. obliquigaleata in clade A, not included in P. siphonantha lineage in 12 clade B. From floral color and beak shape, P. delavayi was easy to misplace into the P. siphonantha group. In the revision of Chinese Pedicularis, Li (1949) cited dozens of specimens for P. delavavi; however, some 13 14 Sichuan specimens were P. tenuituba, and some Yunnan specimens were P. milliana. Subsequently, Tsoong (1963) might be failed to check diagnostic characters of *P. delvayi*, or might misidentify *P. milliana* or *P.* 15 16 tenuituba as P. delavavi, thus he downgraded P. delavavi as a variety in P. siphonantha. Unfortunately, 17 Tsoong's incorrect treatment has been widely adopted by current Chinese Floras (Yang, Holmgren & Mill 1998; Wang 2006), checklists (Wang & Wu 1994; Guan 1998; Xu, Li & Sun 2013) and other publications 18 (Cai et al. 2004; Yu, Zhang & Wang 2008). Moreover, illustrations and/or voucher specimens of "P. 19 20 siphonantha var. delvavi" from northwestern Yunnan were P. milliana, or mixed with P. milliana (e.g. Wang & Wu 1994; Guan 1998; Cai et al. 2004; Yu, Zhang & Wang 2008; Xu, Li & Sun 2013). Some herbarium 21 22 specimens of *P. tenuituba* from Sichuan were misidentified as "*P. siphonantha* var. delvavi". Phylogenetic analyses supported P. tenuituba sister to P. siphonantha, and P. milliana sister to P. sigmoidea. To further 23 clarify phylogenetic relationship between P. milliana and P. sigmoidea needs to extensively sample more 24 populations of them in northwestern Yunnan. According to morphological and phylogenetic evidence, we 25 propose to reinstate P. delavavi as an independent species. Full description and line drawing (Fig. 3) were 26 27 provided.

28 Pedicularis neolatituba P. C. Tsoong was described from Songpan, northern Sichuan, which had short plant (less than 10 cm), long pedicel (up to 4 cm) and basal circinate-incurved galea (Tsoong 1963). In 29 protologue, Tsoong proposed this species similar to three long-pedicelled species, P. franchetiana, P. 30 31 mussotii, and P. mychophila, then established series Neolatitubae P. C. Tsoong. After checking the type specimen of *P. neolatituba* (Fig. S2), we found that it was difficult to distinguished from specimens of *P.* 32 33 delavayi. Pedicularis delavayi also has long pedicel, anterior cleft and mid-upper part inflated calyx, basal 34 twisted galea, semi-circle beak, ciliate corolla lobes and pubescent filaments. The plant height is variable in 35 different specimens. Therefore, we proposed to reduce P. neolatituba as a new synonymy of P. delavayi.

36

37 Taxonomic treatments

38 Pedicularis delavayi Franch. ex Maxim., Bull. Acad. Imp. Sci. Saint-Pétersbourg 32: 531, pl. 1, fig. 7. 1888

1

= Pedicularis siphonantha var. delavayi (Franch. ex Maxim.) P. C. Tsoong, Fl. Reipubl. Popularis Sin.

- 68: 374. 1963 Type: CHINA. Yunnan: Lijiang (Li-kiang), Yulong Snow Mountain (Suee Shan), alt.
 4,000m, 14 Aug. 1886, *J. M. Delavay s.n.* (holotype: LE!, barcode 01010308; isotypes: K!, barcode
 000708729, MPU!, barcode 020765, P!, barcode 02987194).
- 5 Synonymy: Pedicularis neolatituba P. C. Tsoong, in Fl. Reipubl. Popularis Sin. 68: 418–419, pl. 72, f. 1–3.
- 6 1963. *Syn. nov.* Type: CHINA. Sichuan: Songpan (Dongrergo), alt. 4,700m, 9 Aug 1922, *H. Smith* 7 3162 (holotype: PE!, barcode 00033070; isotype: PE!, barcode 00119661).

8 Perennial herb, barely 10 cm tall, drying black or not. Roots fleshy, fusiform. Stems 1 to several, unbranched and erect or \pm ascending, 2–10 cm, with lines of hairs. Basal leaves numerous, mostly 9 membraneous and no leaf blade when beginning to flowering, blades development delayed; petiole up to 5 10 11 cm, winged, glabrescent; leaf blades lanceolate-oblong, 10–30 mm, sparely pubescent on both surfaces, abaxially furfuraceous, pinnatipartite; leaf segments 5-10 pairs, triangular-ovate to oblong-ovate, margin 12 dentate; leafe veins sparely pubescent. Cauline leaves alternate or pseudo-opposite; petiole 0.5–5 cm. 13 sparely pubescent; leaf blades and segments similar to basal ones. Flowers alternate and axillary, dense, 14 flowering \pm synchronous; pedicel 0.5–4 cm, sparely pubescent. Calvx tube 0.8–1 cm, 1/3-2/5 cleft 15 16 anteriorly, mid-upper part inflated in flowering, sparsely long-pubescent; calyx lobes 3 or 5, rarely 2, lateral 17 lobes leaflike, and posterior lobe \pm entire or absent. Corolla purple-red, base whitish, and white spots on the base of galea and the center of lower lip; corolla tube 3-6.5 cm, slender, glabrescent, ridged; galea strongly 18 twisted apically; beak slender, semicircular or slightly S-shaped, bent upward, to 1.2 cm; lower lip ciliate, 19 20 $1.5-2.0 \times 1.5-1.8$ cm, lobes emarginate, middle lobe smaller and involute; filaments attached near tube 21 throats, pubescent. Capsule obliquely oblong, apiculate, $1.4-1.7 \times 0.4-0.6$ cm; seed black, linear-ovate.

Distribution and Habitat. — Pedicularis delavayi is endemic to the Hengduan Mountains regions.
 After re-examination of the herbarium specimens and extensive field expeditions, we confirmed that this
 species occurs in Northwest Yunnan (Deqin, Lijiang, and Shangri-La counties), and West and North
 Sichuan (Daocheng, Jiulong, Kangding, Luding, Miangning, Muli, Songpan, and Xiangcheng counties).
 This species mainly grows in alpine meadows or at the margin of alpine shrub, at the altitude over 3600 m
 a.s.l.

Phenology. — According to field collection and herbarium records, flowering individuals were
 collected from early June to early August. Fruiting specimens conserved at the herbaria were difficult to
 identify. In August, 2007 and 2008, we collected fruiting individuals with mature seeds at Daxueshan
 Mountain of Shangri-Lain, Northwest Yunnan.

- 32 *Conservation status. Pedicularis delavayi* is not common, and it is restricted to alpine meadows.
 33 Its habitats may be threatened by human activities in pasture and tourism. This species can be considered
 34 Least Concern (LC) according to IUCN Red List criteria.
- Selected specimens examined. CHINA. Yunnan: Deqin, L.-M. Gao et al., 25794 (KUN); Lijiang,
 Yulong Mt. H.-D. Li & H. Tang LHD2014-01 (KUN), LHD201-20 (KUN), LHD201340 (KUN);
 Shangi-La, W.-B.Yu 015 (KUN), W.-B.Yu et al. HW10130 (KUN), HW10172 (KUN), HW10200 (KUN),
- 38 LIDZ1258 (KUN). Sichuan: Daocheng, Bowa Mt., Sichuan Vegetation Exped. 1923 (CDBI); Jiulong,

Qing-Quan Wang 20508 (CDBI), W.-B.Yu et al., LIDZ1116 (KUN), YWB201507224 (KUN),
 YWB201507260 (KUN); Kangding, Zheduo Mt., Ru Jiang & Cun-Li Jin 02086 (KUN, PE), W.B.Yu et al.,

- 3 HW10316 (KUN); Xiangcheng, Wuming Mt., *Fu-Sheng Yang Y0071* (PE).
- 4

5 ACKNOWLEDGMENTS

We thank the curators of K, MPU, P and PE for providing online access to specimens, and the curators of
CDBI, KUN, and PE for allowing study of the specimens. We are grateful to L.-M. Gao, H.-J. He, W. Jiang,
H.-D. Li, R. Li, Y. Luo, X.-C. Shi, H. Tang, D. Wu, J.-D. Ya, Q.-L. Yang and S.-D. Zhang for their kind
help in the field and/or providing samples for this study; to P.-H. Huang, J. Yang, and C.-X. Zeng for their
assistance in molecular experiments; to X.-L. Wu for line drawings; and to H.-D. Li and Z.-K. Wu for their
permissions in using photographs.

12

13 Funding

14 This study is supported by grants from the National Natural Science Foundation of China (grants 31470323,

and 31200185), the National Key Basic Research Program of China (2014CB954100), the Major

16 International Joint Research Project of National Natural Science Foundation of China (31320103919), the

17 West Light Foundation of the Chinese Academy of Sciences (Y2227111W1), Visiting Scholar Fellowship

18 of Chinese Academy of Sciences, and the research project of the Komarov Botanical Institute of the

19 Russian Academy of Sciences: "Collections of vascular plants in BIN RAS (history, conservation, study

- 20 and replenishment).
- 21

22 References

- Alfaro ME, Zoller S, and Lutzoni F. 2003. Bayes or bootstrap? A simulation study comparing the performance of
 Bayesian Markov chain Monte Carlo sampling and bootstrapping in assessing phylogenetic confidence.
 Molecular Biology and Evolution 20:255-266. DOI 10.1093/molbev/msg028
- Bonati G. 1904. The alpine and subalpine vegestation of south-east Sikkim. *Records of the Botanical Survey of India* 4:323-431.
- Bonati G. 1910. Contribution à l'étude du genre Pedicularis. Bulletin de la Société Botanique de France 57
 (18):1-35.

Bonati G. 1921. New species of the genera *Phtheirospermum* and *Pedicularis*. *Notes from the Royal Botanic Garden, Edinburgh* 13:103-141.

Cai J, Wang H, Gu Z-J, Mill RR, and Li D-Z. 2004. Karyotypes of thirteen species of Pedicularis
 (Orobanchaceae) from the Hengduan Mountains Region, NW Yunnan, China. *Caryologia* 57:337-347.

- China Plant BOL Group. 2011. Comparative analysis of a large dataset indicates that internal transcribed spacer
 (ITS) should be incorporated into the core barcode for seed plants. *Proceedings of the National Academy of Sciences of the United States of America* 108:19641-19646. 10.1073/pnas.1104551108
- 37 Darriba D, Taboada GL, Doallo R, and Posada D. 2012. jModelTest 2: more models, new heuristics and parallel
 38 computing. *Nature Methods* 9:772.
- 39 Don D, Hamilton F, and Wallich N. 1825. Prodromus florae Nepalensis : sive Enumeratio vegetabilium quae in
 40 itinere per Nepaliam proprie dictam et regiones conterminas, ann. 1802-1803. Londini :: J. Gale.
- 41 Erixon P, Svennblad B, Britton T, and Oxelman B. 2003. Reliability of Bayesian posterior probabilities and 11/14

Pee	Preprints NOT PEER-REVIEWED
1	bootstrap frequencies in phylogenetics. Systematic Biology 52:665-673.
2	Gelman A, and Rubin DB. 1992. Inference from iterative simulation using multiple sequences.457-472.
3	10.1214/ss/1177011136
4	Guan KY. 1998. Highland flowers of Yunnan. Kunming: Yunnan Science and Techology Press.
5	Guindon S, and Gascuel O. 2003. A simple, fast, and accurate algorithm to estimate large phylogenies by
6	maximum likelihood. Systematic Biology 52:696-704. Doi 10.1080/10635150390235520
7	Handel-Mazzetti HREv. 1923. Plantae novae sinenses. Akademie der Wissenschaften in Wien
8	Mathematisch-Naturwissenschaftliche Klasse 60:114-118.
9	Hebert PDN, Cywinska A, Ball SL, and deWaard JR. 2003. Biological identifications through DNA barcodes.
10	Proceedings of the Royal Society of London Series B-Biological Sciences 270:313-321. DOI
11	10.1098/rspb.2002.2218
12	Hillis DM, and Bull JJ. 1993. An empirical test of bootstrapping as a method for assessing confidence in
13	phylogenetic analysis. <i>Systematic Biology</i> 42:182-192. 10.1093/sysbio/42.2.182
14	Huang S-Q, and Fenster CB. 2007. Absence of long-proboscid pollinators for long-corolla-tubed Himalayan
15	Pedicularis species: implications for the evolution of corolla length. International Journal of Plant
16	Sciences 168:325-331.
17	Huang S-Q, Wang X-P, and Sun S-G. 2016. Are long corolla tubes in Pedicularis driven by pollinator selection?
18	Journal of Integrative Plant Biology 58:698-700. 10.1111/jupb.12460
19	Husain T, Garg A, and Agnihotri P. 2010. Genus Pedicularis L. (Scrophulariaceae) in India: a revisionary Study:
20	Bishen Singh Mahendra Pal Singh.
21	Katon K, and Ton H. 2010. Parallelization of the MAFFT multiple sequence alignment program. <i>Bioinformatics</i>
22	26:1899-1900. 10.1093/bioinformatics/btq224
23	Kearse M, Molr R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran
24	C, Interer I, Ashton B, Meintjes P, and Drummond A. 2012. Genetous Basic: An integrated and
25	extendable desktop software platform for the organization and analysis of sequence data. <i>Bioinformatics</i>
20	28.1047-1049. DOI 10.1095/01011101111atics/018199
27	probabilities for phylogenetic hypotheses Molecular Biology and Evolution 24:2108 2118 DOI
20	10 1003/molbey/msm141
30	Kress WI Wurdack KI Zimmer EA Weigt IA and Janzen DH 2005 Use of DNA barcodes to identify
31	flowering plants. Proceedings of the National Academy of Sciences of the United States of America
32	102:8369-8374 DOI 10 1073/pnas 0503123102
33	Li H-L 1948 A revision of the genus <i>Pedicularis</i> in China part I <i>Proceedings of the Academy of Natural</i>
34	Sciences of Philadelphia 100:205-378
35	Li H-L 1949 A revision of the genus <i>Pedicularis</i> in China part II <i>Proceedings of the Academy of Natural</i>
36	Sciences of Philadelphia 101:1-214
37	Li H-L 1951 Evolution in the flowers of <i>Pedicularis Evolution</i> 5:158-164
38	Li R Shi X Yu W-B Feng S and Sun W 2016 Rediscovery of the supposedly extinct <i>Pedicularis humilis</i> in the
39	eastern Himalayas. <i>Oryx</i> 50:204-204. doi:10.1017/S0030605316000016
40	Limpricht W. 1924. Studien über die Gattung Pedicularis. Repertorium Specierum Novarum Regni Vegetabilis
41	20:161-265.
42	Liu M-L, and Yu W-B. 2015. Pedicularis wanghongiae (Orobanchaceae). a new species from Yunnan.
43	southwestern China. <i>Phytotaxa</i> 217:53-62.
44	Liu M-L, Yu W-B, and Wang H. 2013. Rapid identification of plant species and iflora: application of DNA
	17 / 14

Pe	erJ Preprints NOT PEER-REVIEWER
1	barcoding in a large temperate genus Pedicularis (Orobanchaceae). Plant Diversity and Resources
2	35:707-714.
3	Macior LW, and Sood SK. 1991. Pollination ecology of Pedicularis megalantha D. Don (Scrophulariaceae) in
4	the Himachal Himalaya. Plant Species Biology 6:75-81.
5	Macior LW, and Tang Y. 1997. A preliminary study of the pollination ecology of Pedicularis in the Chinese
6	Himalaya. Plant Species Biology 12:1-7.
7	Macior LW, Tang Y, and Zhang J-C. 2001. Reproductive biology of Pedicularis (Scrophulariaceae) in the
8	Sichuan Himalaya. Plant Species Biology 16:83-89.
9	Maximowicz CJ. 1888. Diagnoses plantarum novarum asiaticarum. VII. Bulletin de l'Academie Imperiale des
10	Sciences de St-Petersbourg, sér 3 32:477-629.
11	Mill RR. 2001. Pedicularis L. (Scrophulariaceae). In: Grierson AJC, and Long DG, eds. Flora of Bhutan.
12	Edinburgh: Royal Botanic Garden Edinburgh, 1156-1234.
13	Pennell FW. 1935. The Scrophulariaceae of eastern temperate North America. Philadelphia: The Academy of
14	Natural Sciences of Philadelphia.
15	Pennell FW. 1943. The Scrophulariaceae of the Western Himalayas. The Academy of Natural Sciences of
16	Philadelphia Monographs 5:1-163.
17	Prain D. 1890. The species of Pedicularis of the Indian Empaire and its frontiers. Annals of the Royal Botanical
18	<i>Garden</i> 3:1-196.
19	Ree RH. 2005. Phylogeny and the evolution of floral diversity in Pedicularis (Orobanchaceae). International
20	Journal of Plant Sciences 166:595-613. doi:10.1086/430191
21	Robart BW, Gladys C, Frank T, and Kilpatrick S. 2015. Phylogeny and biogeography of North American and
22	Asian Pedicularis (Orobanchaceae). Systematic Botany 40:229-258. 10.1600/036364415X686549
23	Ronquist F, and Huelsenbeck JP. 2003. MrBayes 3: Bayesian phylogenetic inference under mixed models.
24	Bioinformatics 19:1572-1574. DOI 10.1093/bioinformatics/btg180
25	Stamatakis A, Hoover P, and Rougemont J. 2008. A rapid bootstrap algorithm for the RAxML web servers.
26	Systematic Biology 57:758-771. Doi 10.1080/10635150802429642
27	Tamura K, Stecher G, Peterson D, Filipski A, and Kumar S. 2013. MEGA6: Molecular Evolutionary Genetics
28	Analysis Version 6.0. Molecular Biology and Evolution 30:2725-2729. 10.1093/molbev/mst197
29	Tkach N, Ree RH, Kuss P, Roser M, and Hoffmann MH. 2014. High mountain origin, phylogenetics, evolution,
30	and niche conservatism of arctic lineages in the hemiparasitic genus Pedicularis (Orobanchaceae).
31	Molecular Phylogenetics and Evolution 76:75-92. DOI 10.1016/j.ympev.2014.03.004
32	Tsoong P-C. 1955. A new system for the genus Pedicularis. Acta Phytotaxonomica Sinica 4:71-147.
33	Tsoong P-C. 1956. A new system for the genus Pedicularis (continued II). Acta Phytotaxonomica Sinica
34	5:205-278.
35	Tsoong P-C. 1961. A new system for the genus Pedicularis (continued III). Acta Botanica Sinica 9:230-274.
36	Tsoong P-C. 1963. Scrophulariaceae (Pars II). In: Chien S-S, and Chun W-Y, eds. Flora Reipublicae Popularis
37	Sinacae. Beijing: Science Press, 1-378.
38	Vaidya G, Lohman DJ, and Meier R. 2011. SequenceMatrix: concatenation software for the fast assembly of
39	multi-gene datasets with character set and codon information. Cladistics 27:171-180. DOI
40	10.1111/j.1096-0031.2010.00329.x
41	Wang H. 2006. Pedicularis L. In: Chen S-K, and Wang H, eds. Flora Yunnanica, Vol 16. Beijing: Science Press,
42	468-611.
43	Wang H, and Li D-Z. 2005. Pollination biology of four Pedicularis species (Scrophulariaceae) in northwestern
44	Yunnan, China. Annals of the Missouri Botanical Garden 92:127-138.

 Xu B, Li Z-M, and Sun H. 2013. Seed plants of the alpine subnival belt from the Hengduan mountians <i>China</i>. Beijing: Science Press. Yamazaki T. 1988. A revision of the genus <i>Pedicularis</i> in Nepal. In: Ohba H, and Malla SB, eds. <i>The Himal</i> <i>Plants</i>. Tokyo: University Museum, University of Tokyo, 91-161. Yang C-F, Sun S-G, and Guo Y-H. 2005. Resource limitation and pollen source (self and outcross) affecting production in two louseworts, <i>Pedicularis siphonantha</i> and <i>P. longiflora</i> (Orobanchaceae). <i>Bota</i> <i>Journal of the Linnean Society</i> 147:83-89. Yang CF, Guo YH, Gituru RW, and Sun SG. 2002. Variation in stigma morphology - How does it contribut pollination adaptation in <i>Pedicularis</i> (Orobanchaceae)? <i>Plant Systematics and Evolution</i> 236:89 DOI 10.1007/s00606-002-0223-z Yang CF, and Wang QF. 2015. Nectarless flowers with deep corolla tubes in <i>Pedicularis</i>: does long pistil le provide an arena for male competition? <i>Botanical Journal of the Linnean Society</i> 179:526- 10.1111/boj.12331 Yang F-S, and Wang X-Q. 2007. Extensive length variation in the cpDNA <i>trnT-trnF</i> region of hemipara <i>Pedicularis</i> and its phylogenetic implications. <i>Plant Systematics and Evolution</i> 264:251-264. 	
 <i>China</i>. Beijing: Science Press. Yamazaki T. 1988. A revision of the genus <i>Pedicularis</i> in Nepal. In: Ohba H, and Malla SB, eds. <i>The Himal</i> <i>Plants</i>. Tokyo: University Museum, University of Tokyo, 91-161. Yang C-F, Sun S-G, and Guo Y-H. 2005. Resource limitation and pollen source (self and outcross) affecting production in two louseworts, <i>Pedicularis siphonantha</i> and <i>P. longiflora</i> (Orobanchaceae). <i>Bota</i>. <i>Journal of the Linnean Society</i> 147:83-89. Yang CF, Guo YH, Gituru RW, and Sun SG. 2002. Variation in stigma morphology - How does it contribu pollination adaptation in <i>Pedicularis</i> (Orobanchaceae)? <i>Plant Systematics and Evolution</i> 236:89 DOI 10.1007/s00606-002-0223-z Yang CF, and Wang QF. 2015. Nectarless flowers with deep corolla tubes in <i>Pedicularis</i>: does long pistil le provide an arena for male competition? <i>Botanical Journal of the Linnean Society</i> 179:526- 10.1111/boj.12331 Yang F-S, and Wang X-Q. 2007. Extensive length variation in the cpDNA <i>trnT-trnF</i> region of hemipara <i>Pedicularis</i> and its phylogenetic implications. <i>Plant Systematics and Evolution</i> 264:251-264. 	SW
 Yamazaki T. 1988. A revision of the genus <i>Pedicularis</i> in Nepal. In: Ohba H, and Malla SB, eds. <i>The Himal</i> <i>Plants</i>. Tokyo: University Museum, University of Tokyo, 91-161. Yang C-F, Sun S-G, and Guo Y-H. 2005. Resource limitation and pollen source (self and outcross) affecting production in two louseworts, <i>Pedicularis siphonantha</i> and <i>P. longiflora</i> (Orobanchaceae). <i>Botal</i> <i>Journal of the Linnean Society</i> 147:83-89. Yang CF, Guo YH, Gituru RW, and Sun SG. 2002. Variation in stigma morphology - How does it contribut pollination adaptation in <i>Pedicularis</i> (Orobanchaceae)? <i>Plant Systematics and Evolution</i> 236:89 DOI 10.1007/s00606-002-0223-z Yang CF, and Wang QF. 2015. Nectarless flowers with deep corolla tubes in <i>Pedicularis</i>: does long pistil le provide an arena for male competition? <i>Botanical Journal of the Linnean Society</i> 179:526- 10.1111/boj.12331 Yang F-S, and Wang X-Q. 2007. Extensive length variation in the cpDNA <i>trnT-trnF</i> region of hemipara <i>Pedicularis</i> and its phylogenetic implications. <i>Plant Systematics and Evolution</i> 264:251-264. 	
 <i>Plants.</i> Tokyo: University Museum, University of Tokyo, 91-161. Yang C-F, Sun S-G, and Guo Y-H. 2005. Resource limitation and pollen source (self and outcross) affecting production in two louseworts, <i>Pedicularis siphonantha</i> and <i>P. longiflora</i> (Orobanchaceae). <i>Bota</i> <i>Journal of the Linnean Society</i> 147:83-89. Yang CF, Guo YH, Gituru RW, and Sun SG. 2002. Variation in stigma morphology - How does it contribut pollination adaptation in <i>Pedicularis</i> (Orobanchaceae)? <i>Plant Systematics and Evolution</i> 236:89 DOI 10.1007/s00606-002-0223-z Yang CF, and Wang QF. 2015. Nectarless flowers with deep corolla tubes in <i>Pedicularis</i>: does long pistil lee provide an arena for male competition? <i>Botanical Journal of the Linnean Society</i> 179:526- 10.1111/boj.12331 Yang F-S, and Wang X-Q. 2007. Extensive length variation in the cpDNA <i>trnT-trnF</i> region of hemipara <i>Pedicularis</i> and its phylogenetic implications. <i>Plant Systematics and Evolution</i> 264:251-264. 	iyan
 Yang C-F, Sun S-G, and Guo Y-H. 2005. Resource limitation and pollen source (self and outcross) affecting production in two louseworts, <i>Pedicularis siphonantha</i> and <i>P. longiflora</i> (Orobanchaceae). <i>Bota</i> <i>Journal of the Linnean Society</i> 147:83-89. Yang CF, Guo YH, Gituru RW, and Sun SG. 2002. Variation in stigma morphology - How does it contribut pollination adaptation in <i>Pedicularis</i> (Orobanchaceae)? <i>Plant Systematics and Evolution</i> 236:89 DOI 10.1007/s00606-002-0223-z Yang CF, and Wang QF. 2015. Nectarless flowers with deep corolla tubes in <i>Pedicularis</i>: does long pistil le provide an arena for male competition? <i>Botanical Journal of the Linnean Society</i> 179:526- 10.1111/boj.12331 Yang F-S, and Wang X-Q. 2007. Extensive length variation in the cpDNA <i>trnT-trnF</i> region of hemipara <i>Pedicularis</i> and its phylogenetic implications. <i>Plant Systematics and Evolution</i> 264:251-264. 	
 production in two louseworts, <i>Pedicularis siphonanth</i>a and <i>P. longiflora</i> (Orobanchaceae). <i>Bota</i> <i>Journal of the Linnean Society</i> 147:83-89. Yang CF, Guo YH, Gituru RW, and Sun SG. 2002. Variation in stigma morphology - How does it contribut pollination adaptation in <i>Pedicularis</i> (Orobanchaceae)? <i>Plant Systematics and Evolution</i> 236:89 DOI 10.1007/s00606-002-0223-z Yang CF, and Wang QF. 2015. Nectarless flowers with deep corolla tubes in <i>Pedicularis</i>: does long pistil le provide an arena for male competition? <i>Botanical Journal of the Linnean Society</i> 179:526-10.1111/boj.12331 Yang F-S, and Wang X-Q. 2007. Extensive length variation in the cpDNA <i>trnT-trnF</i> region of hemipara <i>Pedicularis</i> and its phylogenetic implications. <i>Plant Systematics and Evolution</i> 264:251-264. 	seed
 <i>Journal of the Linnean Society</i> 147:83-89. Yang CF, Guo YH, Gituru RW, and Sun SG. 2002. Variation in stigma morphology - How does it contribut pollination adaptation in <i>Pedicularis</i> (Orobanchaceae)? <i>Plant Systematics and Evolution</i> 236:89 DOI 10.1007/s00606-002-0223-z Yang CF, and Wang QF. 2015. Nectarless flowers with deep corolla tubes in <i>Pedicularis</i>: does long pistil le provide an arena for male competition? <i>Botanical Journal of the Linnean Society</i> 179:526-10.1111/boj.12331 Yang F-S, and Wang X-Q. 2007. Extensive length variation in the cpDNA <i>trnT-trnF</i> region of hemipara <i>Pedicularis</i> and its phylogenetic implications. <i>Plant Systematics and Evolution</i> 264:251-264. 	ical
 Yang CF, Guo YH, Gituru RW, and Sun SG. 2002. Variation in stigma morphology - How does it contribut pollination adaptation in <i>Pedicularis</i> (Orobanchaceae)? <i>Plant Systematics and Evolution</i> 236:89 DOI 10.1007/s00606-002-0223-z Yang CF, and Wang QF. 2015. Nectarless flowers with deep corolla tubes in <i>Pedicularis</i>: does long pistil le provide an arena for male competition? <i>Botanical Journal of the Linnean Society</i> 179:526-10.1111/boj.12331 Yang F-S, and Wang X-Q. 2007. Extensive length variation in the cpDNA <i>trnT-trnF</i> region of hemipara <i>Pedicularis</i> and its phylogenetic implications. <i>Plant Systematics and Evolution</i> 264:251-264. 	
 pollination adaptation in <i>Pedicularis</i> (Orobanchaceae)? <i>Plant Systematics and Evolution</i> 236:89 DOI 10.1007/s00606-002-0223-z Yang CF, and Wang QF. 2015. Nectarless flowers with deep corolla tubes in <i>Pedicularis</i>: does long pistil le provide an arena for male competition? <i>Botanical Journal of the Linnean Society</i> 179:526- 10.1111/boj.12331 Yang F-S, and Wang X-Q. 2007. Extensive length variation in the cpDNA <i>trnT-trnF</i> region of hemipara <i>Pedicularis</i> and its phylogenetic implications. <i>Plant Systematics and Evolution</i> 264:251-264. 	te to
 DOI 10.1007/s00606-002-0223-z Yang CF, and Wang QF. 2015. Nectarless flowers with deep corolla tubes in <i>Pedicularis</i>: does long pistil le provide an arena for male competition? <i>Botanical Journal of the Linnean Society</i> 179:526- 10.1111/boj.12331 Yang F-S, and Wang X-Q. 2007. Extensive length variation in the cpDNA <i>trnT-trnF</i> region of hemipara <i>Pedicularis</i> and its phylogenetic implications. <i>Plant Systematics and Evolution</i> 264:251-264. 	-98.
 Yang CF, and Wang QF. 2015. Nectarless flowers with deep corolla tubes in <i>Pedicularis</i>: does long pistil le provide an arena for male competition? <i>Botanical Journal of the Linnean Society</i> 179:526-14 10.1111/boj.12331 Yang F-S, and Wang X-Q. 2007. Extensive length variation in the cpDNA <i>trnT-trnF</i> region of hemipara <i>Pedicularis</i> and its phylogenetic implications. <i>Plant Systematics and Evolution</i> 264:251-264. 	
 provide an arena for male competition? <i>Botanical Journal of the Linnean Society</i> 179:526-14 10.1111/boj.12331 Yang F-S, and Wang X-Q. 2007. Extensive length variation in the cpDNA <i>trnT-trnF</i> region of hemipara <i>Pedicularis</i> and its phylogenetic implications. <i>Plant Systematics and Evolution</i> 264:251-264. 	ngth
 14 10.1111/boj.12331 15 Yang F-S, and Wang X-Q. 2007. Extensive length variation in the cpDNA <i>trnT-trnF</i> region of hemipara 16 <i>Pedicularis</i> and its phylogenetic implications. <i>Plant Systematics and Evolution</i> 264:251-264. 17 10.1007/00000000000000000000000000000000	532.
 Yang F-S, and Wang X-Q. 2007. Extensive length variation in the cpDNA <i>trnT-trnF</i> region of hemipara <i>Pedicularis</i> and its phylogenetic implications. <i>Plant Systematics and Evolution</i> 264:251-264. 	
16 <i>Pedicularis</i> and its phylogenetic implications. <i>Plant Systematics and Evolution</i> 264:251-264.	sitic
	DOI
1/ 10.100//s00606-006-0510-1	
18 Yang H-B, Holmgren NH, and Mill RR. 1998. <i>Pedicularis</i> Linn. In: Wu Z-Y, and Raven P-H, eds. <i>Flow</i>	a of
19 <i>China</i> . St. Louis, Beijing: Missouri Botanical Garden Press & Science Press, 97-209.	
20 Yu W-B, Cai J, Li D-Z, Mill RR, and Wang H. 2013a. Floral ontogeny of <i>Pedicularis</i> (Orobanchaceae), with	h an
21 emphasis on the corolla upper lip. <i>Journal of Systematics and Evolution</i> 51:435-450.	Doi
22 10.1111/Jse.12018	
23 Yu W-B, Cai J, Wang H, and Chen J-Q. 2008. Advances in floral divergence and reproductive adaptation	n in
24 <i>Pedicularis</i> L. (Orobanchaceae). <i>Chinese Bulletin of Botany</i> 25:392-400.	
25 Yu W-B, Huang P-H, Li D-Z, and Wang H. 2013b. Incongruence between nuclear and chloroplast 1	NA
 26 phytogenies in <i>Pearcularis</i> section Cyathophora (Orobanchaceae). <i>PLOS One</i> 8:e/4828. 27 Vu W D, Huang D H, Bao BH, Liu M L, Li D Z, and Wang H. 2011. DNA homoding of <i>Badiaularia</i> 1 	inn
27 Yu w-B, Huang P-H, Ree KH, Liu M-L, Li D-Z, and wang H. 2011. DNA barcoding of <i>Peatcularis</i> 1	inn.
20 Iournal of Systematics and Evolution 40:425 427 10 1111/j 1750 6821 2011 00154 x	nus.
20 Vu W B Li D 7 and Wang H 2012 Highly efficient pollination by humblehees ensures seed productiv	n in
31 Pedicularis lachnoglossa (Orobanchaceae) an early-flowering Himalayan plant Journal of System	u m
<i>and Evolution</i> 50:218-226 10 1111/i 1759-6831 2012 00180 x	nies
33 Yu W-B Liu M-L Wang H Mill RR Ree RH Yang J-B and Li D-Z 2015 Towards a comprehe	sive
34 phylogeny of the large temperate genus Pedicularis (Orobanchaceae), with an emphasis on species	rom
35 the Himalaya-Hengduan Mountains. <i>Bmc Plant Biology</i> 15:176. 10.1186/s12870-015-0547-9	
36 Yu W-B, Zhang S-D, and Wang H. 2008. New taxa of <i>Pedicularis</i> (Scrophulariaceae) from the Heng	luan
37 Mountains, Southwestern China. <i>Novon</i> 18:125-129. 10.3417/2006032	
38	
39	
40	