

# AliTV – interactive visualization of whole genome comparisons

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## **ABSTRACT**

Whole genome alignments and comparative analysis are key methods in the quest of unraveling the dynamics of genome evolution. Interactive visualization and exploration of the generated alignments, annotations, and phylogenetic data are important steps in the interpretation of the initial results. Limitations of existing software inspired us to develop our new tool AliTV, which provides interactive visualization of whole genome alignments. AliTV reads multiple whole genome alignments or automatically generates alignments from the provided data. Optional feature annotations and phylogenetic information are supported. The user-friendly, web-browser based and highly customizable interface allows rapid exploration and manipulation of the visualized data as well as the export of publication-ready high-quality figures. AliTV is freely available at https://github.com/AliTVTeam/AliTV.

Keywords: Comparative genomics, alignment, visualization

## INTRODUCTION

Advances in short- and long-read sequencing and assembly over the last decade (Salzberg et al., 2011; Chin et al., 2013; Hackl et al., 2014) have made whole genome sequencing a routine task for biologists in various fields. Public sequence databases already contain several thousand of draft and finished genomes (Benson et al., 2013), with many more on the way (Pagani et al., 2012). In particular, high throughput sequencing projects of pathogen strains related to recent outbreaks (Rasko et al., 2011), and large scale ecological studies targeting microbial communities and pan genomes of populations using metagenome and single cell sequencing approaches contribute in this process (Turnbaugh et al., 2007; Kashtan et al., 2014). These rich data sets can be explored for large-scale evolutionary processes using comparative genomics and whole genome alignments, revealing genomic recombinations (Didelot et al., 2012; Namouchi et al., 2012; Yahara et al., 2014), islands and horizontal gene transfer (Avrani et al., 2011; Coleman et al., 2006; Langille et al., 2008) as well as the often related dynamics of mobile or endogenous viral elements (Fischer, 2015; Touchon and Rocha, 2007). Other applications of whole genome comparisons include the analysis of paleopolyploidization events (Vanneste et al., 2014) and quantitative measurements of intra-tumour heterogeneity (Schwarz et al., 2015).

However, to facilitate proper interpretation of the obtained whole genome comparisons, visualization is key. One of the first tools to provide an interactive graphical representation of aligned genomes is the multiple whole genome alignment program Mauve (Darling et al., 2004). Mauve represents genomes in a co-linear layout with homologous syntenic blocks indicated by colors and connecting lines. The interactive stand-alone viewer ACT (Carver et al., 2008), in addition to alignment blocks, supports the representation of genomic annotations, such as genes. The R library genoPlotR (Guy et al., 2010) and the Python based application EasyFig (Sullivan et al., 2011), both also based on a co-linear layout and supporting feature annotations, lack interactive analysis features as they are designed to generate static figures.

In addition to co-linear layouts, tools using circular representations of genomes have been developed. BLASTatlas (Hallin et al., 2008) and BRIG (Alikhan et al., 2011) use multiple concentric rings to represent data of individual genomes, with BRIG also providing an interactive

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graphical interface. GenomeRing (Herbig et al., 2012) uses a circular representation as well, however, places all genomes on the same ring and syntenic blocks are connected with arcs extending into the center of the ring.

The web-based comparative genomics software Sybil (Riley et al., 2012) provides interactive co-linear visualization of multiple whole genome alignments with feature annotations and also supports a phylogenetic tree alongside the alignments. The software builds on a relational Chado database schema and, therefore, requires upload and import of custom data sets prior to analysis.

During our analysis of existing software, we found that interactive tools are useful for data exploration, but offer limited support for the figure export and at low qualities. Scripting-based tools provide higher levels of customization and figure quality, however, require familiarity with the respective language, thus often rendering the generation of figures time-consuming. For web- and database-based suites, such as <code>Sybil</code>, the upload and import procedure complicate utilization and limit applicability.

Here we present our stand-alone application <code>AliTV</code> (Alignment Toolbox and visualization) designed for interactive visualization of multiple whole genome alignments. <code>AliTV</code> aims to enable researches to either directly read or automatically generate new whole genome alignments, rapidly explore the results, manipulate and customize the visualization and, at the end of the day, export appealing, publication-grade figures. <code>AliTV</code> reads sequence and annotation or alignment data in common formats (FASTA, GenBank, GFF, MAF, Newick, ...), and internally computes alignments using <code>lastz</code> (Harris, 2007). The user-friendly interface is built on the state-of-the-art <code>D3.js</code> JavaScript framework and can be utilized in a platform independent manner with common web browsers. Genomes are represented in a highly customizable co-linear layout including annotations and an optional phylogenetic tree. The tree is not computed by <code>AliTV</code> but has to be provided during data generation. Also the order of genomes is not automatically optimized to minimize rearrangements. Customizations to the figure by the user can be saved, reloaded, and exported to high quality <code>SVG</code> files.

# **METHODS**

Our tool AliTV is divided into two parts. The first non-interactive part is required for the generation of the input files for our interactive viewer. The second part represents that interactive viewer in the form of a SVG file embedded in a HTML5 website. The latest version of our code can be obtained from GitHub (https://github.com/AliTVTeam/AliTV). It is planned to adjust AliTV in order to integrate it into the BioJS registry (https://biojsnet.herokuapp.com/, Corpas et al. (2014)). The general design of AliTV assures, that AliTV runs on different hard- and software platforms, e.g. Linux, MacOSX, and Windows. The following sections describe those parts in more detail.

## **Data Preparation**

The data preparation is performed by a single Perl script named alitv.pl. This script uses a set of different Perl modules to import incoming data and generate valid JSON input data for our visualization engine described in the next paragraph. One of our aims is to support as many different input formats for sequence and annotation information as possible. Therefore, we used the well tested and broadly accepted BioPerl as basis for our modules (Stajich et al., 2002).

The script alitv.pl uses a YAML file to specify the different input files. Moreover, an easy-to-use-mode is available which requires only a couple of input files and generates the required YAML file on the fly. This generated YAML settings file might be used to reproduce AliTV results or can be used as starting point to alter configuration parameters.

During the preparation step, AliTV requires all-vs-all alignments of the complete sequence set. Those alignments are generated or user provided. The current version of alitv.pl requires lastz to generate all alignments in MAF format (Harris, 2007). Nevertheless, BioPerl supports a broad range of alignment formats. Therefore, other programs can easily be added to the list of supported alignment programs. Moreover, the ability to use existing alignments allows a huge time benefit, when AliTV parameters are changed to optimize the visualization via YAML settings file in a non-interactive manner. Thus future versions of alitv.pl will support caching of alignments based on checksums to avoid unnecessary recalculations.

The final result of our ality.pl is a JSON file, which can be load into our interactive visualization page.



**Table 1.** Chloroplast genomes of the parasitic and non-parasitic plants used in the case study.

Species	Accession	Life-style	Reference
Olea europaea	NC_013707	non-parasitic	Messina (2010)
Lindenbergia philippensis	NC_022859	non-parasitic	Wicke et al. (2013)
Cistanche phelypaea	NC_025642	holo-parasitic	Wicke et al. (2013)
Epifagus virginiana	NC_001568	holo-parasitic	Wolfe et al. (1992)
Orobanche gracilis	NC_023464	holo-parasitic	Wicke et al. (2013)
Schwalbea americana	NC_023115	hemi-parasitic	Wicke et al. (2013)
Nicotiana tabacum	NC_001879	non-parasitic	Kunnimalaiyaan and Nielsen (1997)

#### Interactive Visualization

AliTV is implemented in JavaScript. Our code is documented using JSDoc 3 (version 3.4.0 http://usejsdoc.org/, 02.06.2016). AliTV generates a SVG which is presented within a browser using HTML5. A tutorial is available at https://alitv.readthedocs.io/en/latest/index.html.

To gain advanced application possibilities we use different libraries. The JavaScript library D3.js 3.5.17 (http://d3js.org/, 06.06.2016) provides a wide range of pre-built functions for calculating and drawing the interactive figure. In addition, AliTV employes JQuery 2.2.4 (https://jquery.com/, 06.06.2016) to ease access to several parts of the figure. This is helpful for hiding selected sequences, genes or links. JQueryUI 1.11.4 (https://jqueryui.com/, 06.06.2016) gives us the possibilities to add user-friendly interactions to AliTV. With sliders the user has the chance to specify values for link length and link identity. Context menus offer direct and native interactions with the figure.

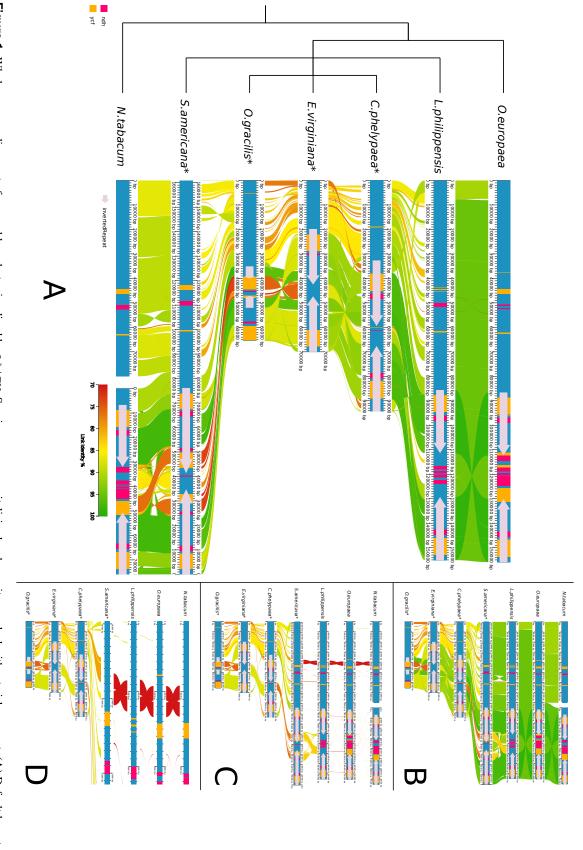
To guarantee correct code functionality we engineer AliTV according to the Test Driven Development. First we write an automated test case that defines a new function. Then we add the minimum amount of code to make the test pass. Finally we refactor the code to accepted standards. We use Jasmine 2.3 (http://jasmine.github.io/, 06.06.2016), as framework for testing our JavaScript code. The tests can run either via the SpecRunner or the command line using the taskrunner grunt 1.0.0 (http://gruntjs.com/, 06.06.2016).

# **RESULTS AND DISCUSSION**

To demonstrate the capabilities of AliTV we describe a short case study using seven published chloroplast genomes (table 1). Four of the chloroplasts belong to parasitic plant species and three to non-parasitic ones. Parasitic plants rely much less or not at all on photosynthetic activity, a trait that should be reflected in the genomic structure of their chloroplast genomes. To assess this hypothesis the chloroplast genomes were downloaded from NCBI and processed with ality.pl. For demonstration purposes, the chloroplast genome of Nicotiana tabacum was split in two pieces to represent an unfinished genome with more than one contig, and the genome sequence of Schwalbea americana was reverse-complemented (flipped). The pair-wise whole genome alignments are visualized by AliTV (figure 1A). The left-hand side of the display panel shows the phylogenetic tree for the seven species with species names as tip labels (parasitic plants are highlighted with an asterisk). The tree has been created provided in accordance to NCBI taxonomy (Sayers et al., 2009). Next to the tip labels, each genome is drawn as a scaled and annotated horizontal bar. The orientation of the S. americana genome was swapped back to match the orientation of the other genomes, indicated by the tick coordinates in reverse order (0 on the right side). N. tabacum is represented by two bars as the sequence has been split into two parts. On those bars features (e.g. genes or inverted repeats (IRs)) are shown as either rectangles or arrows. Alignments between adjacent genomes are represented as colored ribbons. The bottom legend shows the default color scale from red to green corresponding to low and high identity respectively.

The most striking observation is that three of the chloroplast genomes have drastically reduced sizes. All of those are parasitic (table 1). Interestingly the chloroplast genome size of *S. americana* is similar to that of the non-parasitic plants. This can be explained by the life style of *S. americana* which is hemi-parasitic in contrast to the other parasitic plants which are holo-parasites. The features shown are the IR regions as arrows, the hypothetical chloroplast open reading frames as orange and the genes of the ndh family as pink rectangles. First, it can be seen that there is a big variation in size of the inverted repeats. While the IR of *Orobanche gracilis* is the shortest with roughly 5000 bp, that of *S. americana* is the largest with roughly 35 000 bp. Second, there are less genes of the ndh

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plants at the bottom. (C) links filtered by identity (only those with 50 % to 90 % identity are drawn). (D) zoom in on a potential segmental duplication (red 'X'-shaped links) in the top phylogenetic tree on the left-hand side and genomes represented by co-linear horizontal bars on the right; genes and inverted repeats are displayed as rectangles and arrows, Figure 1. Whole genome alignment of seven chloroplasts visualized by AliTV. Species names were italicized and parasites marked with asterisks ex post. (A) Default layout with a four genomes respectively; colored ribbons connect corresponding regions in the alignment. (B-D) Customized layouts: (B) reordered genomes, non-parasitic plants at the top and holo-parasitic



family on Cistanche phelypaea, Epifagus virginiana, O. gracilis, and S. americana. Members of the ndh gene family encode subunits of the NADH dehydrogenase-like complex, which is involved in chlororespiration (Martín and Sabater, 2010). However they are not required for plant growth under optimal conditions (Burrows, 1998). The absence of ndh genes in chloroplasts of parasitic plants has been studied in detail in Wicke et al. (2013). Loss of ndh genes has also been reported for photosynthetic plants such as some conifers and orchids (Wakasugi et al., 1994; Kim et al., 2015). Looking at the pairwise similarities of adjacent genomes, it is apparent that the non-parasitic plants (e.g. Olea europaea and Lindenbergia philippensis) have high overall sequence identity. In contrast, the sequence similarity within parasitic plants is lower. This observation can help framing a hypothesis about the evolutionary pressure on chloroplasts of parasitic plants. Another interesting observation is the distribution of missing regions of C. phelypaea in comparison to L. philippensis. Missing regions are distributed all over the genome and the order of the remaining parts remains stable. Wicke et al. (2013) describe an inversion in the large single copy region of S. americana compared to non-parasitic plants which is clearly visible by the link to N. tabacum around the 115 kbp position. All these observations can be made by simply looking at the raw figure created by ality.pl and visualized by AliTV. However the figure can be analyzed interactively in more detail. One shortcoming of the linear representation of whole genome alignments is the limited comparability of non-adjacent sequences. Therefore, AliTV provides a way for the user to re-order the genomes on the figure (figure 1B). If reordering causes inconsistencies with the phylogenetic tree, the tree is hidden and a warning message is displayed. Furthermore, the links can be filtered by their alignment identity. The default setting is to display only links with minimal identity of 70%. But sometimes it might be interesting to look at regions with less similarity. To see these regions it is also important to hide large regions with high similarity. This can be achieved by changing the identity via a slider (figure 1C). After setting the identity range to 50 % to 90 % red 'X'-shaped links between N. tabacum, O. europaea, L. philippensis, and S. americana become apparent. For detailed inspection of regions of interest, AliTV provides a zoom function (figure 1D). This way the exact location of the alignments can be traced to the locations of psaA and psaB. Moreover AliTV provides functions like alignment length filtering, selective hiding of sequences, links and features, change of orientation (reverse complement) and rotation of circular chromosomes. Finally, it is possible to tweak many graphical parameters, such as colors, labels or spacing, directly via the interface to produce a publication quality figure which can be saved in SVG format. Furthermore, the current state can be saved in JSON format in order to share it with collaborators or continue the work with AliTV at a later time.

# **CONCLUSION**

The case study demonstrates the suitability of AliTV as a tool for visualizing and analyzing whole genome comparisons. AliTV can be used to easily create a figure that show cases many genomic features at once. Furthermore the rich interactive features enable the exploratory analysis and discovery of previously unknown features. Thus novel hypotheses can be generated that can then be validated with experimental methods. So AliTV is a useful tool that will help scientists to find biologically meaningful information in the vast amount of genomic data.

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# REFERENCES

Alikhan, N.-F., Petty, N. K., Ben Zakour, N. L., and Beatson, S. A. (2011). Blast ring image generator (brig): simple prokaryote genome comparisons. *BMC Genomics*, 12:402.

Avrani, S., Wurtzel, O., Sharon, I., Sorek, R., and Lindell, D. (2011). Genomic island variability facilitates Prochlorococcus–virus coexistence. *Nature*, 474(7353):604–608.

Benson, D. A., Cavanaugh, M., Clark, K., Karsch-Mizrachi, I., Lipman, D. J., Ostell, J., and Sayers, E. W. (2013). Genbank. *Nucleic Acids Res*, 41(Database issue):D36–D42.

Burrows, P. A. (1998). Identification of a functional respiratory complex in chloroplasts through analysis of tobacco mutants containing disrupted plastid ndh genes. *The EMBO Journal*, 17(4):868–876.

Carver, T., Berriman, M., Tivey, A., Patel, C., Böhme, U., Barrell, B. G., Parkhill, J., and Rajandream, M.-A. (2008). Artemis and ACT: viewing, annotating and comparing sequences stored in a relational database. *Bioinformatics (Oxford, England)*, 24(23):2672–6.



- Chin, C.-S., Alexander, D. H., Marks, P., Klammer, A. a., Drake, J., Heiner, C., Clum, A., Copeland, A., Huddleston, J., Eichler, E. E., Turner, S. W., and Korlach, J. (2013). Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. *Nature Methods*, (may):1–9.
- Coleman, M. L., Sullivan, M. B., Martiny, A. C., Steglich, C., Barry, K., Delong, E. F., and Chisholm, S. W. (2006). Genomic islands and the ecology and evolution of Prochlorococcus. *Science (New York, N.Y.)*, 311(5768):1768–70.
- Corpas, M., Jimenez, R., Carbon, S. J., García, A., Garcia, L., Goldberg, T., Gomez, J., Kalderimis, A., Lewis, S. E., Mulvany, I., Pawlik, A., Rowland, F., Salazar, G., Schreiber, F., Sillitoe, I., Spooner, W. H., Thanki, A., Villaveces, J. M., Yachdav, G., and Hermjakob, H. (2014). BioJS: an open source standard for biological visualisation its status in 2014. F1000Research.
- Darling, A. C. E., Mau, B., Blattner, F. R., and Perna, N. T. (2004). Mauve: multiple alignment of conserved genomic sequence with rearrangements. *Genome Res*, 14(7):1394–1403.
- Didelot, X., Méric, G., and Falush, D. (2012). Impact of homologous and non-homologous recombination in the genomic evolution of Escherichia coli. *BMC*.
- Fischer, M. G. (2015). Virophages go nuclear in the marine alga ¡i¿Bigelowiella natans¡/i¿. *Proceedings of the National Academy of Sciences*, 112(38):11750–11751.
- Guy, L., Kultima, J. R., and Andersson, S. G. E. (2010). genoplotr: comparative gene and genome visualization in r. *Bioinformatics*, 26(18):2334–2335.
- Hackl, T., Hedrich, R., Schultz, J., and Förster, F. (2014). proovread: large-scale high-accuracy PacBio correction through iterative short read consensus. *Bioinformatics*, 30(21):3004–3011.
- Hallin, P. F., Binnewies, T. T., and Ussery, D. W. (2008). The genome BLASTatlas-a GeneWiz extension for visualization of whole-genome homology. *Molecular bioSystems*, 4(5):363–371.
- Harris, R. S. (2007). *Improved pairwise alignment of genomic DNA*. PhD thesis, Pennsylvania State University.
- Herbig, A., Jäger, G., Battke, F., and Nieselt, K. (2012). GenomeRing: alignment visualization based on SuperGenome coordinates. *Bioinformatics (Oxford, England)*, 28(12):i7–15.
- Kashtan, N., Roggensack, S. E., Rodrigue, S., Thompson, J. W., Biller, S. J., Coe, A., Ding, H., Marttinen, P., Malmstrom, R. R., Stocker, R., Follows, M. J., Stepanauskas, R., and Chisholm, S. W. (2014). Single-Cell Genomics Reveals Hundreds of Coexisting Subpopulations in Wild Prochlorococcus. *Science (New York, NY)*, 344(6182):416–420.
- Kim, H. T., Kim, J. S., Moore, M. J., Neubig, K. M., Williams, N. H., Whitten, W. M., and Kim, J.-H. (2015). Seven New Complete Plastome Sequences Reveal Rampant Independent Loss of the ndh Gene Family across Orchids and Associated Instability of the Inverted Repeat/Small Single-Copy Region Boundaries. *PLOS ONE*, 10(11):e0142215.
- Kunnimalaiyaan, M. and Nielsen, B. L. (1997). Fine mapping of replication origins (ori A and ori B) in Nicotiana tabacum chloroplast DNA. *Nucleic acids research*, 25(18):3681–3686.
- Langille, M., Hsiao, W., and Brinkman, F. (2008). Evaluation of genomic island predictors using a comparative genomics approach. *BMC bioinformatics*.
- Martín, M. and Sabater, B. (2010). Plastid ndh genes in plant evolution. *Plant Physiology and Biochemistry*, 48(8):636–645.
- Messina, R. (2010). Olea europaea chloroplast, complete genome. URL: http://www.ncbi.nlm.nih.gov/nuccore/NC\_013707.2. 30.06.2015.
- Namouchi, A., Didelot, X., Schöck, U., and Gicquel, B. (2012). After the bottleneck: Genome-wide diversification of the Mycobacterium tuberculosis complex by mutation, recombination, and natural selection. *Genome*.
- Pagani, I., Liolios, K., Jansson, J., Chen, I.-M. A., Smirnova, T., Nosrat, B., Markowitz, V. M., and Kyrpides, N. C. (2012). The Genomes OnLine Database (GOLD) v.4: status of genomic and metagenomic projects and their associated metadata. *Nucleic acids research*, 40(Database issue):D571–9.
- Rasko, D. A., Dale, W. R., Sahl, J. W., Bashir, A., Boisen, N., Scheutz, F., Paxinos, E. E., Sebra, R., Chin, C.-s., Iliopoulos, D., Klammer, A., Peluso, P., Lee, L., Kislyuk, A. O., Bullard, J., Kasarskis, A., Wang, S., Eid, J., Rank, D., Redman, J. C., Steyert, S. R., Frimodt-møller, J., Struve, C., Petersen, A. M., Krogfeld, K. A., Nataro, J. P., Schadt, E. E., and Waldor, M. K. (2011). Origins of the E. coli Strain Causing an Outbreak of Hemolytic–Uremic Syndrome in Germany. *The New England journal of medicine*, 365(8):709–717.
- Riley, D. R., Angiuoli, S. V., Crabtree, J., Dunning Hotopp, J. C., and Tettelin, H. (2012). Using sybil for interactive comparative genomics of microbes on the web. *Bioinformatics*, 28(2):160–166.
- Salzberg, S. L., Phillippy, A. M., Zimin, A. V., Puiu, D., Magoc, T., Koren, S., Treangen, T., Schatz, M. C., Delcher, A. L., Roberts, M., Marcais, G., Pop, M., and Yorke, J. A. (2011). GAGE: A



- critical evaluation of genome assemblies and assembly algorithms. *Genome Research*, pages gr.131383.111-.
- Sayers, E. W., Barrett, T., Benson, D. A., Bryant, S. H., Canese, K., Chetvernin, V., Church, D. M.,
  DiCuccio, M., Edgar, R., Federhen, S., Feolo, M., Geer, L. Y., Helmberg, W., Kapustin, Y.,
  Landsman, D., Lipman, D. J., Madden, T. L., Maglott, D. R., Miller, V., Mizrachi, I., Ostell, J.,
  Pruitt, K. D., Schuler, G. D., Sequeira, E., Sherry, S. T., Shumway, M., Sirotkin, K., Souvorov, A.,
  Starchenko, G., Tatusova, T. A., Wagner, L., Yaschenko, E., and Ye, J. (2009). Database resources
  of the National Center for Biotechnology Information. *Nucleic Acids Research*, 37(Database issue):D5–15.
- Schwarz, R. F., Ng, C. K. Y., Cooke, S. L., Newman, S., Temple, J., Piskorz, A. M., Gale, D., Sayal, K., Murtaza, M., Baldwin, P. J., Rosenfeld, N., Earl, H. M., Sala, E., Jimenez-Linan, M., Parkinson, C. A., Markowetz, F., and Brenton, J. D. (2015). Spatial and Temporal Heterogeneity in High-Grade Serous Ovarian Cancer: A Phylogenetic Analysis. *PLOS Med*, 12(2):e1001789.
- Stajich, J. E., Block, D., Boulez, K., Brenner, S. E., Chervitz, S. A., Dagdigian, C., Fuellen, G., Gilbert, J. G. R., Korf, I., Lapp, H., Lehväslaiho, H., Matsalla, C., Mungall, C. J., Osborne, B. I., Pocock, M. R., Schattner, P., Senger, M., Stein, L. D., Stupka, E., Wilkinson, M. D., and Birney, E. (2002). The bioperl toolkit: Perl modules for the life sciences. *Genome Res*, 12(10):1611–1618.
- Sullivan, M. J., Petty, N. K., and Beatson, S. A. (2011). Easyfig: a genome comparison visualizer. *Bioinformatics*, 27(7):1009–1010.
- Touchon, M. and Rocha, E. (2007). Causes of insertion sequences abundance in prokaryotic genomes. *Molecular biology and evolution*.
- Turnbaugh, P. J., Ley, R. E., Hamady, M., Fraser-Liggett, C. M., Knight, R., and Gordon, J. I. (2007). The human microbiome project. *Nature*, 449(7164):804–10.
- Vanneste, K., Baele, G., Maere, S., and Peer, Y. V. d. (2014). Analysis of 41 plant genomes supports a wave of successful genome duplications in association with the Cretaceous–Paleogene boundary. *Genome Research*, 24(8):1334–1347.
- Wakasugi, T., Tsudzuki, J., Ito, S., Nakashima, K., Tsudzuki, T., and Sugiura, M. (1994). Loss of all ndh genes as determined by sequencing the entire chloroplast genome of the black pine Pinus thunbergii. *Proceedings of the National Academy of Sciences*, 91(21):9794–9798.
- Wicke, S., Müller, K. F., de Pamphilis, C. W., Quandt, D., Wickett, N. J., Zhang, Y., Renner, S. S., and Schneeweiss, G. M. (2013). Mechanisms of functional and physical genome reduction in photosynthetic and nonphotosynthetic parasitic plants of the broomrape f amily. *The Plant cell*, 25(10):3711–25.
- Wolfe, K. H., Morden, C. W., and Palmer, J. D. (1992). Function and evolution of a minimal plastid genome from a nonphotosynthetic parasitic plant. *Proceedings of the National Academy of Sciences of the United States of America*, 89(22):10648–10652.
- Yahara, K., Didelot, X., Ansari, M., and Sheppard, S. (2014). Efficient inference of recombination hot regions in bacterial genomes. *Molecular biology and*.