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Quantitative assessment of *Fgfr1* expression in Neurons and Glia

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Fibroblast growth factors (FGFs) and their receptors (FGFRs) have numerous functions in the developing and adult CNS. For example, the FGFR1 receptor is important for proliferation of radial glial cells in the cortex and hippocampus, oligodendrocyte proliferation and regeneration, midline glia morphology and soma translocation, Bergmann glia morphology, and cerebellar morphogenesis. In addition, FGFR1 signaling in astrocytes is required for postnatal maturation of interneurons expressing parvalbumin (PV). FGFR1 is implicated in synapse formation in the hippocampus, and alterations in the expression of *Fgfr1* and its ligand, *Fgf2* accompany major depression. Understanding which cell types express *Fgfr1* during development may elucidate its roles in normal development of the brain as well as illuminate possible causes of certain neuropsychiatric disorders. Here, we used a BAC transgenic reporter line to trace *Fgfr1* expression in the developing murine CNS. The specific transgenic line employed was created by the GENSAT project, *tgFGFR1-EGFP-GP338Gsat*, and includes a gene encoding enhanced green fluorescent protein (*EGFP*) under the regulation of the *Fgfr1* promoter, to trace *Fgfr1* expression in the developing CNS. This model reveals that *Fgfr1* is primarily expressed in glial cells, in both astrocytes and oligodendrocytes, along with some neurons. Dual labeling experiments indicate that the proportion of GFP+ (*Fgfr1+*) cells that are also GFAP+ increases from postnatal day 7 (P7) to 1 month, illuminating dynamic changes in *Fgfr1* expression during postnatal development of the cortex. In postnatal neurogenic areas, GFP expression was also observed in SOX2, doublecortin (DCX), and brain lipid-binding protein (BLBP) expressing cells. *Fgfr1* is also highly expressed in DCX positive cells of the dentate gyrus, but not in the rostral migratory stream. *Fgfr1* driven GFP was also observed in tanycytes and GFAP+ cells of the hypothalamus, as well as in Bergmann glia and astrocytes of the cerebellum. Understanding which cell types express *Fgfr1* may elucidate its role in neuropsychiatric disorders and brain development.
Quantitative assessment of *Fgfr1* expression in Neurons and Glia

Short title: *Fgfr1* expression in Neurons and Glia

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**Running head:** *Fgfr1* is expressed in neurons and glia

**Key words:** Fibroblast Growth Factor, astrocyte,
oligodendrocyte, doublecortin, SOX2, cortex, hippocampus, SVZ
Abstract

**Background.** Fibroblast growth factors (FGFs) and their receptors (FGFRs) have numerous functions in the developing and adult CNS. For example, the FGFR1 receptor is important for proliferation of radial glial cells in the cortex and hippocampus, oligodendrocyte proliferation and regeneration, midline glia morphology and soma translocation, Bergmann glia morphology, and cerebellar morphogenesis. In addition, FGFR1 signaling in astrocytes is required for postnatal maturation of interneurons expressing parvalbumin (PV). FGFR1 is implicated in synapse formation in the hippocampus, and alterations in the expression of *Fgfr1* and its ligand, *Fgf2* accompany major depression. Understanding which cell types express *Fgfr1* during development may elucidate its roles in normal development of the brain as well as illuminate possible causes of certain neuropsychiatric disorders.

**Methods.** Here, we used a BAC transgenic reporter line to trace *Fgfr1* expression in the developing postnatal murine CNS. The specific transgenic line employed was created by the GENSAT project, *tgFGFR1-EGFP* *Gsat*, and includes a gene encoding enhanced green fluorescent protein (*EGFP*) under the regulation of the *Fgfr1* promoter, to trace *Fgfr1* expression in the developing CNS. Unbiased stereological counts were performed for several cell types in the cortex and hippocampus.

**Results.** This model reveals that *Fgfr1* is primarily expressed in glial cells, in both astrocytes and oligodendrocytes, along with some neurons. Dual labeling experiments indicate that the proportion of GFP+ (*Fgfr1+*) cells that are also GFAP+ increases from postnatal day 7 (P7) to 1 month, illuminating dynamic changes in *Fgfr1* expression during postnatal development of the cortex. In postnatal neurogenic areas, GFP expression was also observed in SOX2, doublecortin (DCX), and brain lipid-binding protein (BLBP) expressing cells. *Fgfr1* is also highly expressed...
in DCX positive cells of the dentate gyrus, but not in the rostral migratory stream. *Fgfr1* driven GFP was also observed in tanycytes and GFAP+ cells of the hypothalamus, as well as in Bergmann glia and astrocytes of the cerebellum.

**Conclusions.** The *tgFGFR1-EGFPGP338Gs* mouse model expresses GFP that is congruent with known functions of FGFR1, including hippocampal development, glial cell development, and stem cell proliferation. Understanding which cell types express *Fgfr1* may elucidate its role in neuropsychiatric disorders and brain development.

**Introduction**

Differential binding of Fgfs to Fgf receptor isoforms may confer a high degree of selectivity leading to signaling events that lead to a multitude of specific cellular responses (Fortin et al., 2005). Multiple *Fgf* ligands and three of the *Fgfrs* (*Fgfr1*, *Fgfr2*, and *Fgfr3*) are expressed in the CNS (el-Husseini et al., 1994, Belluardo et al., 1997, Bansal et al., 2003). This complex system of Fgfs and Fgfrs plays a pivotal role in the normal development, maturation, and function of the central nervous system (CNS) (Iwata and Hevner, 2009, Stevens et al., 2010b, Hebert, 2011). FGF signaling is implicated in patterning of the CNS, in determining neuronal and glial cell fate, in influencing cerebral cortex size through maintenance of radial glial stem cells, in cerebellar development, and in regional patterning of the neocortex and midbrain-hindbrain boundaries (Vaccarino et al., 1999, Fukuchi-Shimogori and Grove, 2001, Korada et al., 2002, Storm et al., 2003, Shin et al., 2004, Gutin et al., 2006, Smith et al., 2006,
Storm et al., 2006, Cholfin and Rubenstein, 2007, Muller Smith et al., 2008, Kang et al., 2009, Stevens et al., 2010a, Rash et al., 2011, Muller Smith et al., 2012, Rash et al., 2013, Kang et al., 2014, Smith et al., 2014).

Loss of FGFR1 function by hGFAP-Cre-induced deletion of Fgfr1^floxfloxfloxflox alleles in the dorsal telencephalon of mice results in decreased hippocampal size and volume, without altering hippocampal patterning, and also results in a reduction in the number of dividing progenitor cells of the ventricular zone and dentate gyrus (Ohkubo et al., 2004). The hGFAP-Cre transgene targets the radial glial of the dorsal telencephalon during mid neurogenesis (Ohkubo et al., 2004). Fgfr1 mutants also exhibit a disruption in corpus callosum and hippocampal commissure due to abnormal midline glia development (Smith et al., 2006, Tole et al., 2006). The midline glial cells fail to undergo soma translocation and formation of the indusium griseum leading to midline commissural axon guidance defects (Smith et al., 2006). Furthermore, these mice exhibit postnatal loss of maturation in GABAergic interneurons expressing parvalbumin (PV) and exhibit behavioral hyperactivity (Muller Smith et al., 2008, Smith et al., 2014). Hyperactivity and a decrease in number of interneurons in the cortex co-occur in patients with schizophrenia, bipolar disorder, and Tourette syndrome (Benes et al., 2000, Volk et al., 2000, Kalanithi et al., 2005, Akbarian and Huang, 2006, Hashimoto et al., 2008, Kataoka et al., 2010, Gonzalez-Burgos et al., 2011, Volk and Lewis, 2013). Interestingly, Fgfr1 expression was found to be increased in the prefrontal cortex of individuals with schizophrenia (Volk et al., 2016). Dual inactivation of floxed alleles of Fgfr1 and Fgfr2 with the hGFAP-Cre transgene results in abnormal cerebellar morphogenesis including reduced size of the cerebellum due to a defect in proliferation of both cerebellar glia and granule cell precursors, abnormal orientation and morphology of Bergmann glia, and loss of laminar architecture (Muller Smith et al., 2012). This phenotype is similar to that
observed in Fgf9 mutants (Lin et al., 2009). FGFRs are implicated in maintaining astrocytes in a non-reactive state, and in impeding glial scar formation (Kang et al., 2014). Fgfr1 deletions targeted to oligodendrocyte lineages do not disrupt oligodendrocyte birth, but affect myelin sheath thickness, as well as remyelination and recovery in chronic demyelination models (Furusho et al., 2012, Furusho et al., 2015). Therefore, FGFR1 has important roles in glial function and development including stem cell proliferation, migration, morphology, and support of neural maturation.

FGF signaling is implicated as a contributing factor in affective disorders, including depression. Patients with major depressive disorder (MDD) and bipolar disorder have altered gene expression of FGFs and FGFRs (Evans et al., 2004, Gaughran et al., 2006). In situ hybridization revealed that mRNA for Fgfr1, and its ligand Fgf2, were both down regulated in the hippocampus of rats that had undergone the social defeat paradigm (Turner et al., 2008). Microinjections of FGF2 into the lateral ventricles of rats resulted in an increase in Fgfr1 mRNA in the dentate gyrus within 24 hours post FGF2 injections and was accompanied by acute antidepressant-like effects in the force swim test (Elsayed et al., 2012). Intracerebroventricular infusions of FGF2 into mice were sufficient to block the anhedonia-like behavior caused by chronic unpredictable stress (CUS). Furthermore, intracerebroventricular infusions of FGF2 blocked CUS-induced inhibition of proliferating glia in the prefrontal cortex, and produced antidepressant actions in the novelty suppressed feeding test and forced swim test (Elsayed et al., 2012). Furthermore, increased anxiety, dysregulation of the hypothalamic pituitary axis and decreased hippocampal glucocorticoid receptor expression is observed in FGF2 knockout mice. These effects are reversible by administration of FGF2 (Salmaso et al., 2016). FGF22 and FGF7 are presynaptic organizing molecules that promote differentiation of excitatory and inhibitory
presynaptic terminals in the hippocampal CA3 region through combinatorial signaling of sets of FGFRs (Umemori et al., 2004, Terauchi et al., 2010, Dabrowski et al., 2015). Given the data that FGF2/FGFR1 signaling is important for the regulation of mood and affect, and that FGFR1 signaling may participate in synaptogenesis, a better understanding of the cell types expressing Fgfr1 is important to improving our understanding of its actions.

Previous estimates of Fgfr1 expression have been derived from *in situ* hybridization studies and from immunochemistry using antibodies against FGFR1 (Gonzalez et al., 1995, Belluardo et al., 1997, Bansal et al., 2003, Ohkubo et al., 2004, Blak et al., 2005). In embryonic mice, Fgfr1 is strongly expressed in the hippocampal hem, choroid plexus, cortical ventricular zone, and cortical midline (el-Husseini et al., 1994, Bansal et al., 2003, Ohkubo et al., 2004, Smith et al., 2006). During brain development, expression of Fgfrs change within the oligodendrocyte lineage in a spatial and temporal fashion (Bansal et al., 2003). Fgfr1 is expressed in all three stages: proliferation, migration, and differentiation of oligodendrocytes (OL) (Fortin et al., 2005). An *in situ* hybridization study showed that Fgfr1 is not expressed where Olig2-positive and Pdgfra-positive OL progenitors arise in the ventral ventricular zone (Bansal et al., 2003). In adult mice, the strongest Fgfr1 expression is observed in the hippocampus (Ohkubo et al., 2004). Based on a literature review, Turner and colleagues surmised that neuronal populations in the adult hippocampus and cortex mostly express Fgfr1, in contrast to other FGF receptors that are considered to be expressed primarily in glia (Turner et al., 2012a).

Despite clear advances in our understanding of FGF signaling derived from *in situ* hybridization, it suffers from poor cell-type resolution. Likewise, although immunocytochemistry using antibodies raised to FGFR1 have proven to be important, cross-
reactivity to other FGFRs remains a concern. To circumvent these issues, we investigated Fgfr1 expression in PV+ interneurons, employing a transgenic reporter line, tgFGFR1-EGFPGP338Gsat bacterial artificial chromosome (BAC), that was obtained from GENSAT, http://www.gensat.org (Smith et al., 2014). In this transgenic line, the gene encoding enhanced green fluorescent protein (EGFP) is regulated under the same promoter as Fgfr1. Thus, GFP fluorescence should indicate expression of genes encoding Fgfr1. We previously showed that in tgFGFR1-EGFPGP338Gsat mice, PV+ interneurons did not colocalize with GFP+ cells. Thus, the decrease in PV+ interneurons due to inactivation of Fgfr1 occurs non-cell-autonomously (Smith et al., 2014). We also observed that a large number of glia appeared to express Fgfr1 in adult mice. In the present study, we extend our studies and present a quantitative analysis of the relative expression of Fgfr1 in neurons versus glial cells during postnatal development of the telencephalon. We show that Fgfr1 is differentially expressed, primarily in GFAP+ astrocytes and OLIG2+ cells, with a minority of cells colocalizing with NeuN+ neurons. Furthermore, SOX2+ cells, BLBP+ cells and DCX+ cells in the cortex, hippocampus, subventricular zone (SVZ), and hypothalamus of mice are colocalized with the GFP signal, indicating that these cells also express Fgfr1.

Methods and Materials

Animals

Wild type Swiss Webster mice were crossed with mice expressing enhanced green fluorescent protein (EGFP) under the same promoter as Fgfr1 (tgFGRI-EGFPGP338Gsat). The transgenic line, tgFGRI-EGFPGP338Gsat was generated from the GENSAT project (GENSAT.org) by microinjecting bacterial artificial chromosome with Fgfr1 promoter driving
EGFP into the pronucleus of fertilized mouse eggs. GENSAT produces transgenic BAC-EGFP reporter and BAC-Cre recombinase driver mouse lines with the aim to map the expression of genes in the CNS of mice (Heintz, 2004). This line was obtained from the Mutant Mouse Resource Center (MMRRC.org) at UC Davis. This study was conducted in an ethical manner, utilizing the recommendations of The Guide for the Care and Use of Laboratory Animals of the National Institutes of Health. Animals were euthanized under the University of Louisiana at Lafayette IACUC committee APS numbers 2012-8717-046, 2013-8717-053, 2014-8717 040, 2015-8717-033. Tissue collected for this study was performed under ketamine/xylazine cocktail for P7 and older animals.

**Genotyping**

The mice were genotyped by polymerase chain reaction (PCR) for GFP and through GFP screening with goggles containing a GFP filter (BLS LTD). For PCR based genotyping, tails of mice were collected and DNA was extracted from the tail using 50mM sodium hydroxide (95°C for 30 min), followed by neutralization with 1M TRIS (pH 7.6). Master mix for 1 reaction of PCR for amplifying *GFP* was created using 2.5 µl of 10x PCR buffer, 0.5 µl of 10mM dNTP mix, 1 µl of eGFP forward and reverse primer mix (Forward:AAGTTCATCTGCACCACCG and Reverse: TGCTCAGGTAGTGGTGTCG ), 0.2 µl of 5 units/ µl Hot start Taq Polymerase and 18.8 µl of distilled water. Two microliters of DNA sample were added to 23 µl of Master mix per PCR tube and the samples were amplified in the Applied Biosystems 96 Well Thermocycler. In adult *tgFGR1-EGFPGP338Gs* positive mice, we found we could reliably genotype mice due to GFP expression in the eye, using GFP goggles (available from BLS).

**Immunostaining**
P7 mice were anesthetized with ketamine/xylazine (100 mg and 10 mg per kg of body weight respectively) and euthanized by cervical dislocation. Brains were dissected out and in 4% PFA overnight. Adult mice were euthanized by cardiac perfusion under deep anesthesia (ketamine/xylazine as above). Mice were perfused with cold 1x Phosphate Buffered Saline (PBS) followed by 4% PFA in 1x PBS. Brains were post fixed overnight and cryoprotected as described above. Brains collected from P7 and one-month ages were cryoprotected in 20% sucrose/1xPBS, and cryopreserved with exposure to dry ice and embedded in optimal cutting temperature (OCT) at the time of sectioning. The tissue was thick sectioned (50 µm, free floating) in a cryostat (Microm, HM 505 E) in a series of 10 vials containing 1xPBS. Samples were stored in PBS with 0.2% sodium azide at 4°C, and protected from light exposure.

Prior to antibody incubation, sections were blocked with 10% serum in PBS with 0.2% triton x (Sigma Aldrich) and 0.1% tween 20 (Sigma Aldrich), except for anti-Gad67 staining which did not include detergents. Primary antibodies (Table 1) were detected with Alexa conjugated secondary antibodies (Jackson labs and Abcam) or AMCA conjugated secondary antibodies (Vector) in 5% serum. VECTASHIELD DAPI used for double staining and VECTASHIELD without DAPI used for triple staining when mounting sectioned tissue onto polyprep slides.

**Cell Counting**

Unbiased estimated counts of astrocytes expressing glial fibrillary acidic protein (GFAP), neurons expressing NeuN, and oligodendrocytes and oligodendrocyte precursors expressing Olig2 were obtained using StereoInvestigator software (Microbrightfield) coupled with a AxioCam MRm on the Zeiss axioimager microscope equipped with an ApoTome.2. Tops of cells were counted in three-dimensional counting boxes, which were set to specific parameters.
For counts of the cortex and hippocampus, 50 µm sections were sampled every 20th (cortex) and every 10th section (Hippocampus). Fluorescence images were acquired through StereoInvestigator imaging software.

To determine the percentage of GFP+ cells that were neuronal stem cells expressing Sox2 and neuroblasts expressing DCX, we acquired z stack images of the anterior dentate gyrus hemisphere and a section 500 µm posterior to this first section. The z stack images obtained to count SOX2 markers were 19 µm thick with each slice 1 µm thick. The z stack images acquired to count DCX markers were 10 µm thick with each slice 1 µm thick.

**Statistical Analysis**

Data from the StereoInvestigator software were entered into Excel, imported to JMP Pro 11, and analyzed by student t-tests or ANOVA using SAS.

**Results**

*Fgfr1* is expressed in various cell types of the dentate gyrus, CA, and subventricular zone (SVZ) at P7

Previous *in situ* studies indicated high levels of *Fgfr1* mRNA in the hippocampus, including dentate gyrus (DG) and cornu ammonis (CA regions). To investigate the cell types expressing *Fgfr1* in the hippocampus of P7 control and *tgfgfr1-EGFP*+ mice, samples were stained by immunocytochemistry for cell-type markers to identify each of the following: GFAP+ stem cells and astrocytes, NeuN+ neurons, BLBP+ stem cells, and DCX+ neuroblasts. The cell-type marker staining (red fluorescence) was imaged alongside GFP immunostaining (green fluorescence) to reveal which cell types express *Fgfr1* and with DAPI counterstaining (blue fluorescence) to image nuclei (*Fig. 1*). The colocalization of a cell type marker and Fgfr1
expression is indicated by yellow fluorescence caused by overlapping red and green fluorescence. GFAP and GFP immunostaining showed GFP+ cells in the GFAP+ stem cells of the subgranular zone (SGZ) of the DG of tfgfr1-EGFP+ mice (Fig. 1A-C, Video S1) with little to no green fluorescence in GFP- controls (Fig. 1D-F). In the CA region of the hippocampus, GFAP+/GFP+ cells were primarily observed surrounding the stratum pyramidale, in the stratum radiatum, stratum oriens, as well as the white matter above the CA region (Fig. 1 G-I) with little to no green fluorescence in GFP- littermate controls (Fig. 1 J-L). There was strong GFP fluorescence in the stratum pyramidale of the CA region. This GFP staining colocalized with NeuN+ cells (neurons) (Fig. 2A-C) with little to no green fluorescence in GFP- littermate controls (Fig. 2D-F). NeuN+/GFP+ cells were also observed in some, but not all, granule cell layer neurons of the developing DG of tfgfr1-EGFP+ mice (Fig. 2G-I) with little to no green fluorescence in GFP- controls (Fig. 2J-L). Stereological analysis of tfgfr1-EGFP expression in GFAP+ cells of the dentate gyrus showed that of GFP+ cells, 24% ± 8% are GFAP positive, and of GFAP+ cells, 50% ± 8% are GFP+. Of GFP+ cells in the CA, 50% ± 5% are GFAP positive and of GFAP+ cells in the CA, 43% ± 9% are GFP positive (Table 3). One-way ANOVA statistical analysis revealed that the total number of GFAP+ cells in the DG and CA of tfgfr1-EGFP+ are not significantly different from their littermate controls (Table 3). This analysis suggests that the insertion of tfgfr1-EGFP is not lethal to GFAP+ stem cells and astrocytes. We further explored Fgfr1 promoter driven expression of GFP in the postnatal neurogenic niches of the DG and SVZ of the lateral ventricles at P7. Immunostaining for DCX (neuroblasts) and GFP extensively colocalized in the SGZ and granule cell layer (GCL) of the DG (Fig. 3A and 3B, low and high magnification respectively) with nearly all DCX positive cells co-expressing GFP. Postnatally, the SVZ of the lateral ventricles gives rise to olfactory bulb
interneurons. GFP+ cells were observed in the SVZ, consistent with previous studies of postnatal
Fgfr1 expression (Fagel et al., 2009). However, DCX did not colocalize with GFP staining in the
SVZ of the lateral ventricles (Fig. 3C), in contrast to what was observed in the DG of the
hippocampus. Immunostaining with BLBP (stem cells) and GFP antibodies revealed strong
colocalization of Fgfr1 driven GFP in the subgranular zone (SGZ) of the dentate gyrus (DG)
(Fig. 3D and 3E, low and high magnification, respectively) as well as BLBP+ cells of the SVZ
(Fig. 3F).

Fgfr1 is expressed in cortical GFAP+ astrocytes and NeuN+ neurons at P7
To determine which cortical cells express Fgfr1, we stained the cortical tissue of P7
tgfgfr1-EGFP+ and control mice with GFAP and NeuN antibodies. Whereas GFP+ cells
colocalized with GFAP+ astrocytes throughout the cortex (Fig. 4A-C), GFP+ cells colocalized
with NeuN+ cells (neurons) mostly in layers 5 and 6 of the cortex at this age (Fig. 4D-F).
Stereological analysis of the cortex region revealed that of GFP+ cells in the cortex, 32% ± 4%
are GFAP positive. Of the cortical GFAP+ cells, 93% ± 2% are GFP positive. This analysis
indicates that most Gfap expressing astrocytes also express Fgfr1. The total number of cortical
GFAP+ cells of tgfgr1-EGFP+ mice was not significantly different from the number of cells in
littermate controls (Table 3), suggesting that the insertion of the transgene does not significantly
alter the number of astrocytes expressing Gfap.

We examined whether tgfgr1-EGFP+ was expressed in oligodendrocytes and their
precursors by staining for OLIG2 in the cortex. We found that OLIG2+ staining colocalized with
GFP in the cortex (Fig. 4G-I) and SVZ and subcortical white matter (Fig. 4J). At P7, radial glia
of the cortex are undergoing soma translocation to become astrocytes, and can be detected with
BLBP. BLBP and GFP immunostaining revealed strong colocalization throughout the cortex (Fig. 4K-M). At P7, GFP staining colocalized with BLBP+ stem cells and Bergmann Glia in the developing cerebellum (Fig. 4N). This finding is consistent with the participation of Fgfr1/Fgfr2 signaling in Bergmann Glia morphology and cerebellar development (Muller Smith et al., 2012). Furthermore, GFP+ cells colocalized with GFAP+ astrocytes of the cerebellum, but not within granule cell layer or molecular layer neurons that stain with NeuN. GFP+ cells likely colocalized with NeuN- Purkinje neurons based on the location and size of GFP+ cells in the Purkinje layer (Fig. S1 and Video S2).

**Fgfr1 is expressed in specific cell types of the hippocampus and SVZ at 1 month**

We next investigated Fgfr1 expression in 1-month *tgfgfr1-EGFP*+ mice. Immunostaining for SOX2+ cells (stem cells), DCX+ cells (neuroblasts), NeuN+ neurons, GFAP+ astrocytes and stem cells, and OLIG2+ oligodendrocytes was performed in combination with GFP immunostaining to determine which cell types express Fgfr1 in the dentate gyrus, CA, and SVZ. GFP+ cells colocalized with SOX2+ cells (Fig. 5A-E) and DCX+ cells (Fig. 5F-J) in the SGZ and granule cell layers. To determine the percentage of GFP+ cells that were SOX-2 positive and DCX positive, we obtained z stack images of an anterior dentate gyrus hemisphere and posterior dentate gyrus hemisphere, and performed counts from these images. Of the GFP+ cells counted in the z stack, 33% ± 2% are SOX2 positive. Conversely, the majority of SOX2+ cells, 71% ± 2%, are GFP positive (Fig. 5E). Of the GFP+ cells counted in the z stack, 64% ± 0.9% are DCX positive and of DCX+ cells, 86% ± 2% are GFP positive (Fig. 5J). Triple staining with GFP, GFAP and NeuN antibodies revealed that GFAP+ cells (stem cells) and NeuN+ cells (neurons) colocalized with GFP+ cells in the SGZ (Fig. 5K-N) and in the CA region (Fig. 5O-R),
respectively. GFP+ cells did not colocalize with OLIG2+ cells (oligodendrocytes) in the dentate gyrus (Fig. S2). Taken together, these results indicate that GFP (Fgfr1) was expressed in stem cells and neuroblasts of the DG. Stereological analysis of tgfgrf1-EGFP expression in NeuN+ cells of the dentate gyrus showed that of GFP+ cells, 16% ± 6% are NeuN positive and of NeuN+ cells, 5% ± 0.7% are GFP positive. Of GFP+ cells in the CA, 25% ± 7% are NeuN positive and of NeuN+ cells, 15% ± 6% are GFP positive. Stereological analysis of tgfgrf1-EGFP expression in GFAP+ cells of the dentate gyrus showed that of GFP+ cells, 51% ± 3% are GFAP positive and of GFAP+ cells, 85% ± 4% are GFP positive. Of GFP+ cells in the CA, 61% ± 9% are GFAP positive and of GFAP+ cells, 81% ± 3% are GFP positive. These findings indicate that most GFAP positive astrocytes express Fgfr1 at 1 month. The total number of SOX2+ cells, DCX+ cells, NeuN+ cells and GFAP+ cells of tgfgrf1-EGFP+ mice were not significantly different from their littermate controls in the DG and CA (Sox2: p=0.23, DCX: p=0.88 and Table 3).

Many GFAP+ cells also express GFP in the SVZ (Fig. 6A-H) of 1 month tgfgrf1-EGFP+ mice. Colocalized GFP positive cells and GFAP positive fibers were observed both within the SVZ and in the white matter above it (Fig. 6D, 6G, and 6H). There was also significant colocalization of GFP with SOX2+ cells (Fig. 6I-L) indicating that GFAP+ and SOX2+ stem cells of the SVZ express Fgfr1. NeuN+/GFP+ cells were not observed in the SVZ. Similar to what was found at P7, GFP+ cells did not colocalize with DCX+ cells (neuroblasts) in the SVZ (Fig. 6M-O) or in the DCX+ cells of rostral migratory stream (RMS) as it enters the olfactory bulb (Fig. 6P). In the rostral migratory stream, GFP staining surrounds the DCX+ cells as would be expected from astrocytes surrounding the migrating neuroblasts.
**Fgfr1** is expressed in cortical GFAP+ astrocytes, NeuN+ neurons and OLIG2+ cells at 1 month

To determine which cortical cells express **Fgfr1**, we examined cortical tissue of 1-month old *tgfgfr1-EGFP*+ and control mice immunostained with GFAP, NeuN or OLIG2 antibodies alongside GFP antibodies. Analysis of **Fgfr1** expression in the cortex revealed that GFP+ cells colocalized with GFAP+ and NeuN+ cells (Fig. 7A-E, compared to control D). Of note, GFP immunofluorescence appeared less brightly fluorescent in cells colocalizing with NeuN compared to those colocalizing with GFAP. GFP+ cells also colocalized with OLIG2+ cells (Fig. 7G-H) with little to no green fluorescence occurring in littermate controls (Fig. 7I).

Stereological analysis of the cortex revealed that of GFP+ cells, 57% ± 7% are GFAP positive. Of the GFAP+ astrocytes, 83% ± 6% are GFP positive. Of GFP+ cells, 25% ± 4% are NeuN positive and of NeuN+ cells, 12% ± 2% are GFP positive. Of GFP+ cells, 29% ± 0.9% are OLIG2 positive and of OLIG2+ cells, 58% ± 0.08% are GFP positive. The total number of cortical GFAP+, NeuN+, and OLIG2+ cells of *tgfgfr1-EGFP*+ mice were not significantly different from their littermate controls (Table 3).

**Fgfr1** is expressed in various cell types of the hypothalamus at P7 and 1 month

Comparing **Fgfr1** expression in different regions of the P7 and 1-month old mice, we observed GFP expression in cells of the hypothalamus and among cells lining the third ventricle, within the arcuate nucleus and medial eminence (Fig. 8, Fig. S3 and Video S3). In 1-month *tgfgfr1-EGFP*+ mice, GFP+ cells strongly colocalized with SOX2+ cells throughout the hypothalamus and third ventricle and arcuate nucleus (Fig. 8D-I). GFAP+ tanycytes, including those of the arcuate nucleus were among the hypothalamic cells observed to express **Fgfr1** both
at 1 month and at P7 (Fig. 8J-L). Tanycytes participate in neuroendocrine regulation and transport of molecules from the CSF to the hypothalamus, release of gonadotropin-releasing hormone, and production of triiodothyronine (T3) in the brain (Rodriguez et al., 2005). GFP expression was not observed in NeuN+ or OLIG2+ cells of the hypothalamus (Fig. 8M-O, Fig. S3 A-D). At P7, we also observed colocalization of BLBP and GFP in the hypothalamus near the third ventricle, but not many in the medial eminence (Fig. S3 E,F, Video S3).

Calretinin (CR) and somatostatin (SST) neurons express Fgfr1 in one-month old mice.

The NeuN+/GFP+ neurons observed at 1 month were not restricted to any specific cortical layer, and a minority population of GFP colocalized with NeuN+ neurons. We therefore sought to determine if inhibitory neurons express Fgfr1. Tissue from one-month old tgfgr1-EGFP+ mice and their control littermates was stained for GAD67. Some GAD67+ cells colocalized with GFP in the cortex (Fig. 9A, B), but not in the hippocampus (Fig. 9C) or SVZ (Fig. 9D). Our previous investigations determined that GFP was not colocalized with PV+ interneurons (Smith et al., 2014). This led to the experiments in which staining the one-month old tissue with calretinin (CR) and somatostatin (SST) was performed alongside GFP immunostaining. CR+ inhibitory neurons express GFP in the cortex (Fig. 9E, F), DG (Fig. 9G), and SVZ (Fig. 9H). Some SST+ inhibitory neurons express GFP in the anterior cingulate of the cortex (Fig. 9I, J), but none were observed in the DG (Fig. 9K).

Discussion

Through immunostaining for GFP in tgfgr1-EGFP+ mice of the tgfGR1-EGFPGP338Gsat line, we examined the cell specific expression of Fgfr1 in the cortex,
hippocampus, SVZ, hypothalamus, and cerebellum from P7 to one-month of age. In postnatal brains, Fgfr1 is highly expressed in the hippocampus, SVZ, hypothalamus, and in numerous cells of the cortex. In the hippocampus, Fgfr1 was expressed in DCX+ neuroblasts, in GFAP+ stem cells and astrocytes, and Sox2+ stem cells of the SGZ, and in neurons of the CA. Furthermore, Fgfr1 was expressed in astrocytes, neurons, and oligodendrocytes. Fgfr1 expression in cortical neurons was not restricted to a single layer and colocalized with CR+ and SST+ inhibitory neurons. Fgfr1 expression was also observed in Bergmann glia and GFAP+ astrocytes of the cerebellum.

The tgFgfr1-EGFP+ line has been shown to be a robust model in identifying which cell types express Fgfr1, corroborating the findings of previous in situ hybridization studies (el-Husseini et al., 1994, Gonzalez et al., 1995, Belluardo et al., 1997, Bansal et al., 2003, Ohkubo et al., 2004). This transgenic line gives the added benefit of allowing simple identification of the subtype of cells expressing Fgfr1 by immunofluorescence staining since GFP is regulated by the Fgfr1 promoter. Unlike the use of antibodies for FGFR1, which may have cross reactivity with similar FGFR family members, we can have greater confidence that we are identifying Fgfr1 expression accurately within a cell. Total neuronal and glial cell counts were not significantly different between the tgfgfr1-EGFP+ mice and their control littermates. Therefore, the insertion of this transgene into the host DNA does not cause toxicity amongst the neuronal and glial cells observed.

Previous studies have shown that inactivation of FGFR1/FGFR2 and FGFR1/FGFR2/FGFR3 signaling in the developing dorsal VZ results in premature depletion of radial glial stem cells and a smaller cortex, whereas loss of Fgfr1 alone is sufficient to result in reduced hippocampal volume and reduced hippocampal stem cell proliferation during
embryogenesis and the early postnatal period (Ohkubo et al., 2004, Kang et al., 2009, Stevens et al., 2010b, Rash et al., 2011). Our studies of Fgfr1 expression in embryos and at P1 will be described elsewhere. At P7, Fgfr1 was expressed in BLBP positive cells. At this age, the BLBP+ radial glial cells are undergoing soma translocation and differentiating into astrocytes of the cortical parenchyma. BLBP is also expressed in Bergmann glia of the cerebellum, which are cells that have dual roles as stem cells and as a scaffold for granule cell neuron migration and alignment of Purkinje neuron dendrites. At P7, BLBP positive cells expressed Fgfr1 in stem cells and Bergmann Glia of the cerebellum. These findings are consistent with the demonstrated occurrence of cooperative signaling between FGFR1 and FGFR2 in the formation of the cerebellum, and specifically, in the morphology and pial attachment of Bergmann glia of the cerebellar anlage (Lin et al., 2009, Muller Smith et al., 2012).

At P7, and one-month, Fgfr1 was expressed in the dentate gyrus and CA regions of the hippocampus, as well as in the SVZ, hypothalamus, and all layers of the cortex. At P7, Fgfr1 was expressed in the granule cell layer of the dentate gyrus, whereas at 1 month, Fgfr1 was expressed primarily in the subgranular zone or in the granule zone adjacent to it. The Fgfr1 expressing cells in the granule cell layer of P7 mice sparsely colocalized with GFAP positive astrocytes and NeuN positive neurons, and the majority colocalized with DCX positive neuroblasts, of which there are many more at P7 than in the more mature brain of a 1-month old mouse. At one month, the Fgfr1 expressing cells in the subgranular zone colocalized with SOX2+ and GFAP+ stem cells, as well as in the DCX positive neuroblasts. The expression of Fgfr1 seems to be downregulated from DCX+ cells as they mature into NeuN expressing neurons. The Fgfr1 expressing cells in the CA region at P7 and 1 month colocalize with GFAP positive astrocytes and NeuN positive neurons in the stratum pyramidale.
GFAP positive astrocytes and BLBP stem cells in the SVZ at P7 are expressing Fgfr1, indicating that stem cells of the SVZ express Fgfr1. GFAP positive astrocytes at 1 month also express Fgfr1. At P7 and 1 month, Fgfr1 expressing cells in the SVZ do not colocalize with DCX positive neuroblasts and NeuN positive neurons. Thus, DCX positive cells express Fgfr1 in one of the postnatal neurogenic regions – dentate gyrus. The difference in Fgfr1 expression observed between DCX positive cells of the DG and SVZ implies that the staining in DCX positive cells of the hippocampus is not due to residual GFP protein that has not been degraded once a daughter cell born from a stem cell stops expressing Fgfr1. Therefore, we can further accurately identify Fgfr1 expression within cells using this transgenic model. Fgfr1 expressing cells were also colocalized with SOX2 positive cells at 1 month. In the SVZ, there may be additional cell types not investigated in the current study, including glial progenitor cells such as O4 or Nestin, which may colocalize with Fgfr1 expression.

FGFR signaling participates in development of hippocampal synaptic transmission and synaptogenesis (Cambon et al., 2004, Umemori et al., 2004, Terauchi et al., 2010). Activation of FGFR1 promotes synapse formation in hippocampal neurons (Cambon et al., 2004, Terauchi et al., 2010, Dabrowski et al., 2015). Changes in Fgfr1 expression, and FGF2 levels in the hippocampus are linked to major depression and anxiety, as well as to responses to antidepressants (Evans et al., 2004, Newton and Duman, 2004, Gaughran et al., 2006, Turner et al., 2008, Elsayed et al., 2012). It is hypothesized that Fgf2 and Fgfr1 are downregulated in depression and anxiety, and this downregulation can be partially reversed by antidepressant treatment (Turner et al., 2012b, a). The chronic unpredictable stress model causes anhedonia and is a model of depression that results in decreased (mRNA) Fgfr1 levels in the prefrontal cortex (Elsayed et al., 2012). Antidepressant treatment resulted in increased (mRNA) Fgf2 levels, and
the antidepressant effects could be blocked by an FGFR1 inhibitor. These authors further showed that FGF2 administration also had an antidepressant effect (Elsayed et al., 2012).

Most GFAP positive astrocytes in the cortex coexpressed Fgfr1, and the majority of the Fgfr1 expressing cells were GFAP+ astrocytes. A minority of Fgfr1 expressing cells were NeuN positive neurons at P7 and one month. While it was previously believed that Fgfr2 is expressed in glia and that Fgfr1 is mostly expressed by neuronal populations in the adult cortex, our findings support the view that most of the cortical cells expressing Fgfr1 in the adult brain are, in fact, astrocytes or oligodendrocytes (Turner et al., 2012a). In this study we see that most GFAP positive astrocytes express Fgfr1 and a small proportion of NeuN positive neurons express Fgfr1 in one month old mice. A higher percentage of Fgfr1 expressing cells are GFAP positive astrocytes than NeuN positive neurons. Thus, astrocytes constitute a majority of Fgfr1 expressing cells in the adult brain. This idea is consistent with previous mRNA profiling of astrocytes in which Fgfr1 was identified as a gene with enriched expression in astrocytes as compared to other cell types in the brain (Lovatt et al., 2007, Cahoy et al., 2008, Doyle et al., 2008).

An imbalance of excitatory and inhibitory neurons, along with hyperactivity has been documented in certain neurological disorders (Benes et al., 2000, Volk et al., 2000, Kalanithi et al., 2005, Akbarian and Huang, 2006, Hashimoto et al., 2008, Kataoka et al., 2010, Gonzalez-Burgos et al., 2011, Volk and Lewis, 2013). Inactivation of Fgfr1 leads to a decrease in the abundance of parvalbumin interneurons and is associated with an increase in hyperactivity (Muller Smith et al., 2008). A subsequent study demonstrated that in these individuals, a decrease in interneurons occurs postnatally and that Fgfr1 expression does not occur in parvalbumin expressing interneurons (Smith et al., 2014). Thus the decrease in Fgfr1 expression
and decrease in parvalbumin interneurons occurs in a non-cell autonomous manner. Here, we tested other interneuron markers, including GAD67, calretinin, and somatostatin. At one month, GAD67, a marker for inhibitory neurons, colocalizes with Fgfr1 expressing cells in the cortex and not in the dentate gyrus. Calretinin interneurons express Fgfr1 throughout the cortex, and in the dentate gyrus, whereas somatostatin interneurons express Fgfr1 at the anterior cingulate of the cortex. Brandt et al., 2003 demonstrated newborn calretinin positive cells in the dentate gyrus do not express GABA (Brandt et al., 2003), offering a probable explanation as to why GAD67 is not observed colocalizing with Fgfr1 expressing cells in the DG.

The hypothalamus, responsible for controlling hormonal production, stress regulation, and feeding behaviors, has been found to contain a neural stem/progenitor cell niche (Robins et al., 2013). Our results show that GFAP positive astrocytes are Fgfr1 expressing cells. The morphology and location of the Fgfr1 expressing cells are consistent with the cells being tanycytes of the hypothalamus (Robins et al., 2013). The medial eminence of the hypothalamus contains Fgfr1 expressing cells that also express SOX2. Fgfr1 expressing cells that were not identified are similar in morphology and location to the neurons that release gonadotropin-releasing hormone (GNRH1). These results are in congruence with a study, which demonstrated FGFR1’s role in targeting of GNRH1 axons to the medial eminence (Gill and Tsai, 2006, Ojeda et al., 2008). Therefore, understanding FGFR signaling can give further insight to the hypothalamic regulation of homeostasis.

Conclusion

FGFR1 has been implicated as having multiple functions in CNS development, homeostasis, and behavior, but defining the cellular basis of its functions depends on having a
clear understanding of which cell types the *Fgfr1* gene is expressed in, and when. The GENSAT project was envisioned as a resource that would provide the tools for such detailed studies (Heintz, 2004). Here, we have extended the previously published *in situ* based studies and online resources with a detailed examination of the *tgFGR1-EGFP* line. Our data are congruent with *in situ* studies, but with the added feature of double immunofluorescence with glial and neuronal markers, and a quantification of the relative expression in glial versus neuronal cells in the young adult brain. We here find *Fgfr1* promoter driven GFP expression in a variety of stem cells of the CNS including the young adult SVZ, young adult SGZ, cerebellar Bergmann Glia, and SOX2+ cells of the hypothalamus. An additional study of *Fgfr1* expression in embryonic and perinatal stem cells will be described elsewhere. We also find that *Fgfr1* is expressed predominantly in glia in the young adult brain, although significant expression in DCX+ positive neuroblasts of the hippocampus was also observed. This was in stark contrast to DCX+ neuroblasts of the RMS. Our findings may shed light on the participation of FGF2/FGFR1 signaling in determining anxiety and mood, where a neuronal role for FGFR1 has been hypothesized (Turner et al., 2012a). Future studies are needed to determine whether *Fgfr1* expression colocalizes with other markers, such as S100β, O4, and NG2 at embryonic and postnatal time points. Since *Gfap* is not expressed in all astrocytes, S100β (glial specific marker) primarily expressed in astrocytes, but also in some ependymal cells) would be a good marker to further explore *Fgfr1* expression. The *tgFgfr1-EGFP* model can also be used to study additional stages in development, or *Fgfr1* expression after environmental manipulations previously shown to alter *Fgfr1* expression, including animal models for induced depression such as social defeat stress.
Acknowledgements

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References


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Table 2. Parameters used for Stereological Cell Counts

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<td>250 x 250</td>
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Table 3. Stereology Results for Hippocampal DG, CA, and Cortex

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<th>P-value</th>
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<td>DG</td>
<td>GFAP</td>
<td>5614 ± 467</td>
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<td>15974 ± 7447</td>
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<td>50% ± 8%</td>
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<td>GFAP</td>
<td>51258 ± 1741</td>
<td>0.42</td>
<td>46413 ± 14851</td>
<td>50% ± 5%</td>
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<td>GFAP</td>
<td>412147 ± 22494</td>
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<td>1220518 ± 53849</td>
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<td>93% ± 2%</td>
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<td>DG</td>
<td>NeuN</td>
<td>755542 ± 228698</td>
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<td>243542 ± 12945</td>
<td>16% ± 6%</td>
<td>5% ± 0.7%</td>
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<td>GFAP</td>
<td>149161 ± 22540</td>
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<td>243542 ± 12945</td>
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<td>85% ± 4%</td>
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<td>379768 ± 27634</td>
<td>25% ± 7%</td>
<td>15% ± 6%</td>
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<td>Olig2</td>
<td>80075 ± 137359</td>
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<td>1574962 ± 195563</td>
<td>29% ± 0.9%</td>
<td>58% ± 0.08%</td>
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Table 4. Qualitative comparison of *Fgfr1* expression

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<tr>
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<td>DG</td>
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<tr>
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<td>Hypothalamus</td>
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NA = not analyzed
Figure 1. *Fgfr1* expression in GFAP+ cells of the hippocampus at P7. GFAP (A,D) GFP (B,E) immunostaining of the DG in P7 tgfgrfr1-EGFP+ mice (A-C, n=3) and tgfgrfr1-EGFP-littermate controls (D-F, n=3) demonstrated strong GFAP/GFP colocalization in cells of the SGZ (arrowheads) and their radial fibers into the GCL (small arrows). GFAP and GFP immunostaining of the CA region in tgfgrfr1-EGFP+ mice (G-I, n=3) and tgfgrfr1-EGFP-controls (J-L, n=3). GFP+ staining was observed in stratum pyramidale (SP) as well as in cells above (stratum oriens, SO) and below (stratum radiatum, SR) this layer (H). GFAP+/GFP+ colabeling (arrowheads) was observed in primarily in the SO and SR within the CA region, and in the white matter dorsal to the hippocampus. Scale bar is 25 μm. SGZ = subgranular zone, GCL = granule cell layer, SP=stratum pyramidale.
Figure 2. *Fgfr1* expression in NeuN+ cells of the hippocampus at P7. NeuN (A,D) and GFP (B,E) immunostaining of the CA region in *tgfgr1*-EGFP+ mice (A-C, n=3) and *tgfgr1*-EGFP- controls (D-F, n=3). NeuN+/GFP+ staining was observed in stratum pyramidale (SP) of the CA region. NeuN (G,J) and GFP (H,K) immunostaining of the DG in *tgfgr1*-EGFP+ mice (G-I, n=3) and *tgfgr1*-EGFP- controls (J-L, n=3). There was some colabeling in the granule cell layer between GFP+ and NeuN+ cells. Scale bar is 25 μm in A-D and 50 μm in G-L. SP=stratum pyramidale.
Figure 3. Fgfr1 expression in neuroblasts of the hippocampus, and stem cells of the hippocampus and SVZ at P7. DCX and GFP colocalize in neuroblasts of the P7 mice in the granule cell layer of the DG in the hippocampus (A, low magnification, and B, high magnification), but not in the SVZ of the lateral ventricles (C). Many of the BLBP+ cells colocalize with GFP in the stem cells of the SGZ of the DG (D, low magnification, and E, high magnification), as well as stem cell in the SVZ (F). Scale bar is 50µm in A, C, D, F and 25µm in B and E.
Figure 3
Figure 4. Fgfr1 expression in cortex of P7 mice. Cortical GFAP, NeuN, BLBP and GFP immunostaining in P7 tgfgr1-EGFP+ mice (A, B, D, E, G-J, n=3) and tgfgr1-EGF- controls (C, F, n=3). GFP+ cells colocalize with GFAP+ cells (A, B). GFP+ cells weakly colocalize with NeuN+ at layers 5 and 6 of the cortex (D, E). GFP+ cells colocalize with Olig2+ cells in the cortex (G-I) and the subcortical white matter and SVZ (J). GFP+ cells colocalize with BLBP+ cells throughout the layers of the cortex (K-M) as well as the BLBP+ cells and Bergmann glia of the cerebellum (N). Scale bar is 50µm. 1, 2, 3, 4, 5, 6 = layers of the cortex.
Figure 5. Fgfr1 expression in the dentate gyrus and CA of 1-month mice. A majority of SOX2+ cells (A, C) in the DG of tgfgr1-EGFP+ mice colocalize with GFP (B, C) compared to GFP- controls (D). Counts of SOX2+ and GFP+ cells were made directly from Z stack images (19μm thick, 1 μm steps, from Z stack images using StereoInvestigator software from MBF). The relative proportions of single, and double-labeled cells in the DG were quantified (E), with 71% of SOX2 positive cells expressing GFP. A majority of DCX+ (F, H) cells in the DG also express GFP (G, H) compared to GFP- controls (I). Counts for DCX+ and GFP+ were performed on 10μm thick, 1μM step size, Z stack images using Image J (n=3) of the dentate gyrus. The relative proportions of single, and double-labeled cells in the DG were quantified (J) with 85% of DCX+ cells expressing GFP. GFAP (Red), NeuN (Blue), and GFP triple immunolabeling was perfomed on 1-month old mice. We observed GFAP and GFP colabeling in both the SGZ of the DG (K-N), as well as in the CA region (O-R) where most GFAP+ cells are present in the stratum oriens or stratum radiatum. NeuN+/GFP+ double positive cells were primarily observed in the stratum pyramidale (SP) of the CA region in the hippocampus. These cells were quantified by unbiased stereology with the StereoInvestigator (Table 3). Scale bar in I is 25 μm for images A-D and F-I, Scale bar in R is 50 μm for images K-R.
Figure 6. *Fgfr1* expression in the SVZ and rostral migratory stream of 1-month mice. GFP (A,E), GFAP (B,F), NeuN (C), immunostaining of the SVZ in 1-month *tgfgfr1-EGFP*+ mice (A-H, n=3). GFP+ cells of the SVZ colocalized with GFAP+ cells (D, high magnification in G and H. Arrowheads in E-H=GFAP/GFP+ cells) and GFAP+ fibers. GFP (I) and SOX2 (J) staining demonstrated that many, but not all SOX2+ cells also colabel with GFP+ (K and L, DAPI staining included in L). In contrast, DCX+ neuroblasts in the SVZ and rostral migratory stream did not colabel with GFP (L-O). Scale bar is 50µm in A-D and M-P and 25µm in E-L.
**Figure 7. Fgfr1 expression in the cortex of 1-month mice.** Cortical GFAP (Red), NeuN (Blue), and GFP immunostaining in 1-month *tgfgfr1-EGFP*+ mice (A-E, n=3) and *tgfgfr1-EGF*- mice (F, n=3). GFP+ cells colocalized with GFAP+ (arrows) (A, B, merged in D, low magnification in E), and some NeuN+ (double arrows) (B, C, merged in D, lower magnification in E). Immunostaining for Olig2+ in *tgfgfr1-EGFP*+ mice (G, H) compared to *tgfgfr1-EGF*- mice (I) demonstrated that many Olig2+ cells colocalized with GFP. Scale bars are 50µm.
Figure 8. *Fgfr1* expression in the hypothalamus. Sox2+ positive cells were observed along the third ventricle of *tgfgfr1-EGF*- control mice (A-C) and *tgfgfr1-EGF+* mice (D-I). These SOX2+ cells also express GFP under the Fgfr1 promoter (D-I). GFAP+ tanacytes along the third ventricle and arcuate nucleus also colocalize with GFP, as do GFAP+ cells within the hypothalamus (J-L). NeuN+ cells in the hypothalamus do not colocalize with GFP (M-O). A-L, 1 month; M-O, P7. Scale bar is 50µm.
Figure 8

A  SOX2  B  GFP  C

Control

D  SOX2  E  GFP  F

TGFr1-EGFP+

G  SOX2  H  GFP  I

J  GFAP  K  GFP  L

M  NeuN  N  GFP  O
Figure 9. *Fgfr1* expression in the 1-month interneurons. GAD67, CR, SST, and GFP immunostaining of the cortex, DG, and SVZ in one-month old *tgfgfr1-EGFP*- controls (A, E, I, n=2 SST, n=3 GAD67, CR) and *tgfgfr1-EGFP*+ mice (B-D, F-H, J, K, n=2 SST, n=3 GAD67, CR). GFP+ cells colocalized with GAD67+ cells in the cortex (B), but not in the DG (C) and SVZ (D). Some CR+ cells express GFP in the cortex (F), DG (G) and SVZ (H). Some SST+ cells express GFP in the anterior cingulate of the cortex (J), but not in the DG (K). Scale bars are 50µm. AC = anterior cingulate.
Figure 9

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<tbody>
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<td>A</td>
<td>B</td>
</tr>
<tr>
<td>E</td>
<td>F</td>
</tr>
<tr>
<td>I</td>
<td>J</td>
</tr>
</tbody>
</table>

**GFP/GAD67/DAPI**

**GFP/CR/DAPI**

**GFP/SST/DAPI**

**AC**