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1 **Title page**

2

3 **Article title:** Effect of stratification and gibberellic acid on epicotyl dormancy release
4 in seeds of *Yunnanopilia longistaminea* (Opiliaceae)

5 **Types of contribution:** Notes and Comments

6 **Running title:** Seed germination of *Y. longistaminea*

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10 **Keywords:** *Yunnanopilia longistaminea*, epicotyl dormancy, radicle emergence,
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30 **Abstract:**

31 *Yunnanopilia longistaminea* is an endangered monotypic species belonging to
32 Opiliaceae. This edible plant is an important germplasm source with a high economic
33 value in China. The seed dormancy and germination of Opiliaceae species have been
34 rarely investigated. This study examined the effects of scarification, soaking in
35 gibberellic acid, and dehydration on the seed germination of *Y. longistaminea*. Results
36 indicated that the seed germination of this species involves two stages: radicle
37 emergence and epicotyls (shoot) emergence. During radicle emergence, the optimum
38 temperatures were 28 °C and 28 °C/20 °C. Seed moisture content and viability
39 decreased as dehydration occurred. Thus, the seeds may be recalcitrant. The optimum
40 GA₃ solution for the seeds undergoing shoot emergence was 100 mg·L⁻¹. The
41 percentages of shoot emergence in 7 and 14 days of stratification at 5 °C were slightly
42 higher than those in other groups. This study is the first to describe epicotyl dormancy
43 in *Y. longistaminea* seeds. From the seed grow to the seedling of *Y. longistaminea*
44 subjected to a autumn→winter→spring temperature process in nature conditions.
45 Warm and cold stratification can alleviate radicle and epicotyl dormancy, respectively.
46 The duration of cold stratification also significantly affects the epicotyl dormancy
47 release of *Y. longistaminea*. The researches on the seeds breaking methods:
48 warm(28°C/20°C)→cold(5°C)→GA₃(100mg·L⁻¹)→warm(28°C/20°C).

49 **Key words:** *Yunnanopilia longistaminea*, epicotyl dormancy, radicle emergence,
50 endangered plant, stratification

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59 Introduction

60 Seed dormancy is a survival mechanism through which germination time and
61 distribution area are adjusted on the basis of different environmental conditions
62 (Copete et al., 2011). Temperate plant seeds are typically dormant in the seed
63 formation period; as such, these seeds require an appropriate temperature to germinate
64 and for the seedlings to grow (Baskin and Baskin, 2004; Baskin et al., 2000).
65 Dormancy ensures seed germination at appropriate temperature or rainfall conditions;
66 this phenomenon also results in a high probability of successful seedling
67 establishment (Bewley et al., 2006).

68 *Yunnanopilia longistaminea* (W. Z. Li) C. Y. Wu et D. Z. Li, a monotypic species
69 belonging to Opiliaceae, is a vulnerable plant endemic to Red River Valley, Yunnan
70 Province, Southwest China. As the sole member of a geographically isolated genus, *Y.*
71 *longistaminea* plays a significant part in the phylogeny and evolution of Opiliaceae.
72 Furthermore, *Y. longistaminea* is an important resource plant consumed as a delicious
73 wild woody vegetable. However, this species is endangered because of excess
74 utilization and habitat destruction. As a key component of germplasm conservation
75 and access to wild plant resource, artificial propagation, including seed germination
76 and seedling establishment, is an essential step in conservation procedure plans (Shen
77 et al., 2015; Khanna et al., 2013). However, the germination characteristics of *Y.*
78 *longistaminea* have yet to be investigated.

79 Limited information on seed dormancy and germination in Opiliaceae has been
80 published. Willis et al. (2014) presented the phylogeny of seed plant families with
81 current dormancy classes and proposed that Opiliaceae seeds exhibit
82 morphophysiological dormancy (MPD). A common type of this condition is deep
83 simple epicotyl MPD, generally referred to as epicotyl dormancy. Epicotyl dormancy
84 occurs in many species growing in various vegetation types. In this case, various
85 temperature conditions, in addition to the epicotyl, can break radicle dormancy; warm
86 and cold temperatures independently alleviate radicle and epicotyl dormancy (Baskin
87 and Baskin, 1998; Barton, 1933; Nikolaeva, 1977). We hypothesized that *Y.*
88 *longistaminea* seeds may undergo epicotyl dormancy. This study generally aimed to

89 characterize the germination requirements for radicle and epicotyls (shoot) emergence
90 of *Y. longistaminea*. The desiccation tolerance of *Y. longistaminea* seeds was also
91 examined because large seeds are sensitive to dehydration.

92

93 **Materials and methods**

94 *Seed material and seed collection*

95 *Y. longistaminea* is mainly distributed in the Red River Valley in Yunnan,
96 Southwest China. All individual plants were distributed vertically from 1,500 m to
97 3,000 m in elevation. The annual mean temperature was 18 °C with a minimum of
98 5.2 °C in January and a maximum of 35.8 °C in July. The annual average rainfall was
99 approximately 2000 mm, which is mostly recorded between June and September. The
100 region is characterized by a typical high humidity geographical environment with an
101 annual mean relative humidity of more than 80%. There were two stages in *Y.*
102 *longistaminea* of seeds seedlings, the first stages was radicle-emergence; the second
103 stages was shoot emergence (epicotyl-plumule emergence). Radicle-emergence
104 procedure includes four processes: embryo growth, testa rupture, endosperm rupture.
105 In the study, radicles grown to ≥ 2 mm, shoot grown to ≥ 5 mm were considered
106 as radicle-emergence and epicotyl-plumule emergence. All experiments were finished
107 when the surplus seeds (no germinated seeds) were for a consecutive one month until
108 eventually rotted out.

109 In the nature, *Y. longistaminea* drupes reach maturity between June to July, and
110 seedlings emerge in spring next year. In this study, mature *Y. longistaminea* fruits
111 were manually harvested in germination tests started one week after collection. 2015.
112 *Y. longistaminea* drupes were full grow, it got a yellow tone on the skin. The drupes
113 yellow were selected within the sarcocarp to softening. The seeds were manually
114 separated from the drupes and divided into two portions. One portion was reserved in
115 paper bags for germination tests; the other portion was immediately placed under
116 moisture-controlled condition for desiccation tolerance tests (Kunming, Yunnan
117 Province).

118 *Seed morphology, weight, and moisture content*

119 The dimensions (breadth and length) of the seeds were measured by using a
120 vernier caliper in triplicates and the 1000-seed-weight was determined. Moisture
121 content was examined gravimetrically at $103 \text{ }^\circ\text{C} \pm 2 \text{ }^\circ\text{C}$ for 17 h in accordance with
122 standard procedures. The final moisture content was expressed as dry weight basis (%)
123 (Shen et al., 2010). Four replicates were used to determine moisture content expressed
124 as dry mass basis.

125 *Effect of stratification and desiccation on radicle-emergence*

126 *Stratification*

127 Seeds were surface-sterilized with 0.85% sodium hypochlorite for 1 min and
128 washed with distilled water. Fresh seeds were placed on moist autoclaved fine sand in
129 a plastic box ($25 \times 10 \times 8$ cm length, width, height). The seeds were incubated within
130 a 12 h/12 h photoperiod at daily alternating temperatures of $28 \text{ }^\circ\text{C}/15 \text{ }^\circ\text{C}$ and
131 $28 \text{ }^\circ\text{C}/20 \text{ }^\circ\text{C}$ and at constant temperatures of $5 \text{ }^\circ\text{C}$, $15 \text{ }^\circ\text{C}$, $20 \text{ }^\circ\text{C}$, $25 \text{ }^\circ\text{C}$, $28 \text{ }^\circ\text{C}$, and
132 $30 \text{ }^\circ\text{C}$. All temperatures were set in four replicates with 10 seeds in each replicate. The
133 seeds were then checked for radicle emergence every 5 days throughout each
134 treatment duration. Distilled water was added as needed to ensure that moisture was
135 non-limiting during germination.

136 *Desiccation*

137 The seeds were dried for 0, 6, 9, 12, 24, and 36 h in an airtight glass desiccator
138 overlaid with activated silica gel. The moisture content of the seeds was determined in
139 weight in each drying treatment. Afterward, four replicates of dried seeds were tested
140 for germination at $28 \text{ }^\circ\text{C}/20 \text{ }^\circ\text{C}$ in daylight for 12 h.

141

142 *Effect of GA_3 treatment and cold stratification on the epicotyl dormancy release of*
143 *radicle-emerged seeds*

144 GA_3

145 This experiment aimed to determine whether GA_3 eliminates the epicotyl
146 dormancy of radicle-emerged *Y. longistaminea* seeds. The seeds with the same radicle
147 lengths (about 5 cm) were independently soaked in 0, 50, 100, and $200 \text{ mg}\cdot\text{L}^{-1}$ GA_3
148 (95% purity, Sigma) for 10 h at the same temperature set to test the epicotyl plumule

149 emergence. The test conditions were the same as described above.

150 *Cold stratification*

151 The seeds with similar radicle lengths (about 5 cm) were stratified at 5 °C.
152 During stratification, Four replicates of 10 seeds were removed and monitored for
153 epicotyl-plumule emergence after 0, 7, 14, 21, and 35 days. The seeds with observed
154 epicotyl-plumule emergence were transferred into a temperature- and light-controlled
155 incubator (BSG-400 Illuminating Incubators, Shanghai, China) and incubated at an
156 alternating temperature of 28 °C/20 °C in the plastic boxes. Epicotyl dormancy was
157 considered broken when the cotyledons emerged partly out of the seed coat (Hidayati
158 et al., 2005).

159 *Statistical analysis*

160 The shoot emergence percentage (SP) and the final radicle emergence percentage
161 (REP) were determined as follows:

162

$$163 \quad \text{Radicle emergence percentage (\%)} = \left(\frac{\text{number of radicle emerged seeds}}{\text{number of seeds per sample}} \right) \times 100$$

$$164 \quad \text{Shoot emergence percentage (\%)} = \left(\frac{\text{number of shoot emerged seeds}}{\text{number of radicle has emerged}} \right) \times 100$$

165 Arcsine transformation was applied for REP and SP data before statistical
166 analysis was conducted to ensure variance homogeneity. Data given in figures were
167 not transformed. The factors influencing REP and SP were analyzed through ANOVA.
168 If a significant difference was detected by ANOVA, Fisher's least significant
169 difference (LSD) was applied to accomplish multiple comparison tests. Statistical
170 analyses were performed in SPSS 18.0.1. The critical level of significance was P =
171 0.05 in all of the tests.

172

173 **RESULTS**

174 *Characteristics of Y. longistaminea seeds*

175 *Y. longistaminea* seeds generally consist of a seed coat, an embryo, an endosperm,

176 and a cotyledon. The fresh seeds were large with 19.12 ± 1.22 mm (average \pm SE, $n =$
177 10) in length and 12.68 ± 0.63 mm (average \pm SE, $n = 10$) in width. The 1000-fresh
178 seed weight was 2580 ± 108 g. Indeed, *Y. longistaminea* seeds are large.

179 *Effects of stratification and desiccation on radicle growth*

180 *Stratification*

181 The radicle emergence of *Y. longistaminea* seeds is sensitive to temperature.
182 Radicles did not emerge from any of the newly matured seeds at 5 °C. The optimum
183 temperatures for radicle emergence were 28 °C and 28 °C/20 °C. High and low
184 temperatures significantly decreased the REP (Fig.1).

185 *Desiccation treatments*

186 The moisture content of *Y. longistaminea* seeds steadily decreased for 24 h of
187 drying, and $19.23 \pm 1.45\%$ moisture was reached after 24 h of dehydration. The REP
188 of the seeds decreased during dehydration. The REP decreased from $38.89 \pm 4.81\%$ to
189 $0 \pm 0\%$ when the seed moisture content decreased from $25.6 \pm 1.55\%$ to $19.23 \pm$
190 1.45% (Fig. 2). This finding indicated that REP is sensitive to dehydration.

191 *Effect of GA₃ treatment and stratification on the epicotyl dormancy release of* 192 *radicle-emerged seeds*

193 *GA₃*

194 The optimum GA₃ solution for the shoot emergence of radicle-emerged seeds
195 was $100 \text{ mg}\cdot\text{L}^{-1}$. Although 50, 200, and $0 \text{ mg}\cdot\text{L}^{-1}$ (control) did not significantly differ,
196 the SP of the seeds with emerged radicles was improved by the GA₃ treatment (Fig. 3).
197 The mean germination time decreased rapidly.

198 *Cold stratification*

199 After 7, 14, and 21 days of cold stratification at 5 °C, the shoot emergence was
200 significantly increased to $86.67 \pm 5.77\%$, $80 \pm 10\%$, and $66.67 \pm 5.77\%$, respectively.
201 By contrast, SP declined when the duration of cold stratification was prolonged (Fig.
202 4).

203

204 **Discussion**

205 The seed germination of *Y. longistaminea* involves two stages: radicle and

206 epicotyl emergence. Warm stratification can break radicle emergence and cold
207 stratification can stimulate the epicotyl emergence of radicle-emerged seeds. GA₃
208 treatments can also improve both radicle and epicotyl emergence. According to seed
209 dormancy classification criteria, *Y. longistaminea* seeds undergo epicotyl dormancy.
210 Numerous species in the Araceae, Araliaceae, Aristolochiaceae, Berberidaceae,
211 Gentianaceae, Liliaceae, Papaveraceae, and Ranunculaceae families also experience
212 epicotyl dormancy (Baskin et al., 2004; Baskin and Baskin, 2005; Adams et al., 2003;
213 Baskin et al., 1993; Baskin and Baskin, 2004; Kondo et al., 2004; Karlsson et al.,
214 2003; Walck et al., 2000). However, this study is the first to report an Opiliacea
215 species with epicotyl dormancy.

216 *Y. longistaminea* seeds that dispersed in June-July germinated in the field in the
217 following spring. The drupes reach maturity between June to July, and seedlings
218 emerge in spring next year in the nature conditions, the process time is about 10
219 months. The time of *Y. longistaminea* seeds germination was shorter after
220 stratification and GA₃ treatment. In this case, they were exposed to a sufficient warm
221 stratification period after they were dispersed for radicle emergence. They also
222 received cold stratification in winter for epicotyl emergence. This finding is similar to
223 those observed in other species with epicotyl dormancy (Hidayati et al., 2005).
224 However, the embryonic growth in *Y. longistaminea* seeds during radicle and epicotyl
225 emergence should be further investigated.

226 Dehydration-sensitive seeds typically grow in large, broadleaf evergreen tropical
227 or subtropical zones and then mature in the rainy season (Pammenter and Berjak,
228 2000; Shen et al., 2015). In our study, *Y. longistaminea* seeds matured in the rainy
229 season (June) at >25 °C in their natural habitat. In the dehydration test, the seed
230 moisture content of *Y. longistaminea* decreased gradually during desiccation. Seed
231 viability also declined as the duration of dehydration was prolonged. Thus, *Y.*
232 *longistaminea* seeds are recalcitrant.

233 In summary, this study is the first to demonstrate that *Y. longistaminea* seeds
234 undergo epicotyl dormancy and exhibit recalcitrance. Considering the high frequency
235 of human disturbances in the natural habitat of this endangered species, we speculate

236 that the prolonged duration of *Y. longistaminea* seed germination may affect its natural
237 regeneration. Thus, we recommend the artificial promotion of dormancy release and
238 germination for its conservation and management. The studies should be proceed with
239 GA₃ and temperature for embryo becoming coarser in the future.

240

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246

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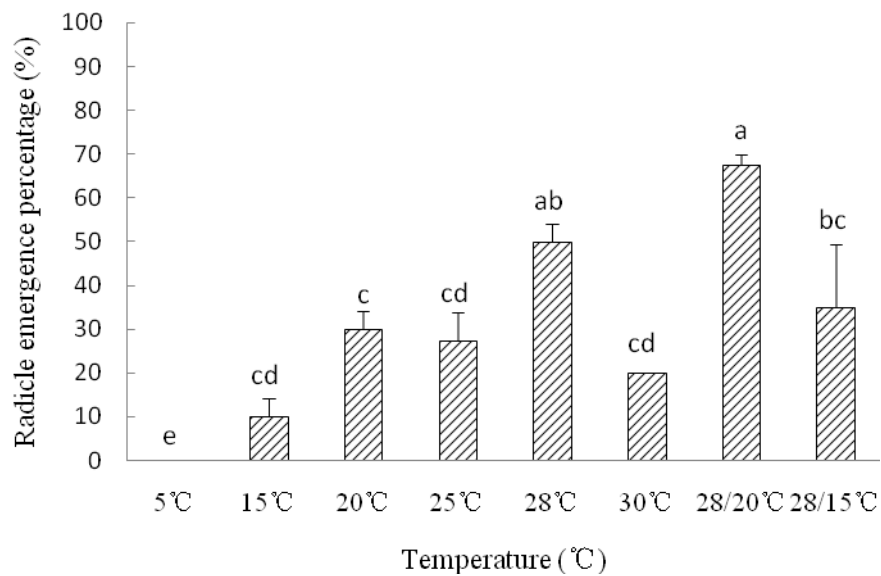
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305 **Figure legends**

306

307 **Fig. 1.** Seed radicle emergence of *Y. longistaminea* at different temperatures and a photoperiod of
308 12/12 h day/night. Values are mean \pm standard error (n = 3). Different letters indicate significant
309 differences between the temperatures (P < 0.05).

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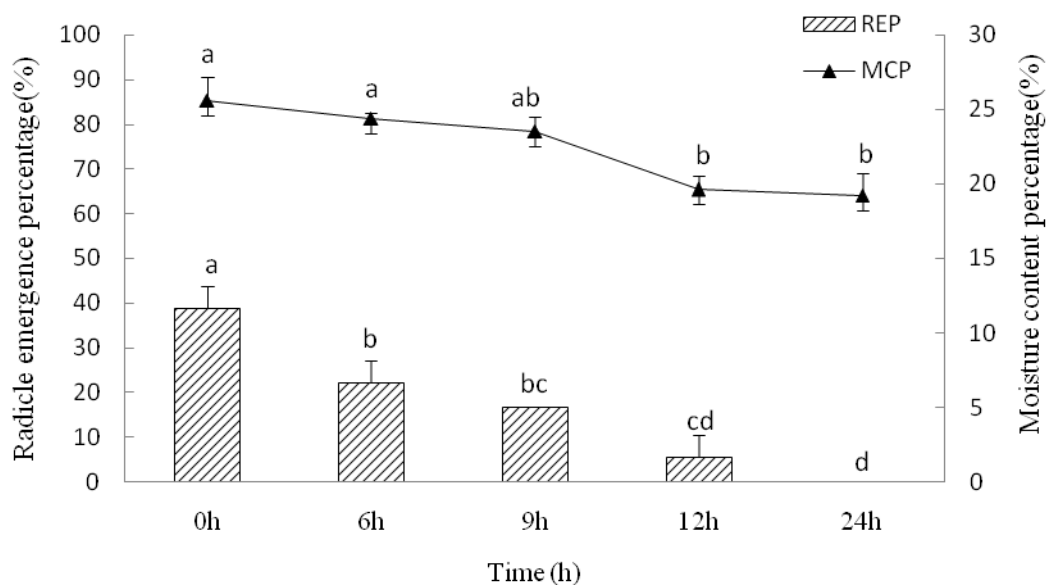
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319 **Fig. 2.** Seed radicle emergence of *Y. longistaminea* at different moisture content and dehydration
320 time. Values are mean \pm standard error (n = 3). Different letters indicate significant differences
321 between the dehydration time (P < 0.05).

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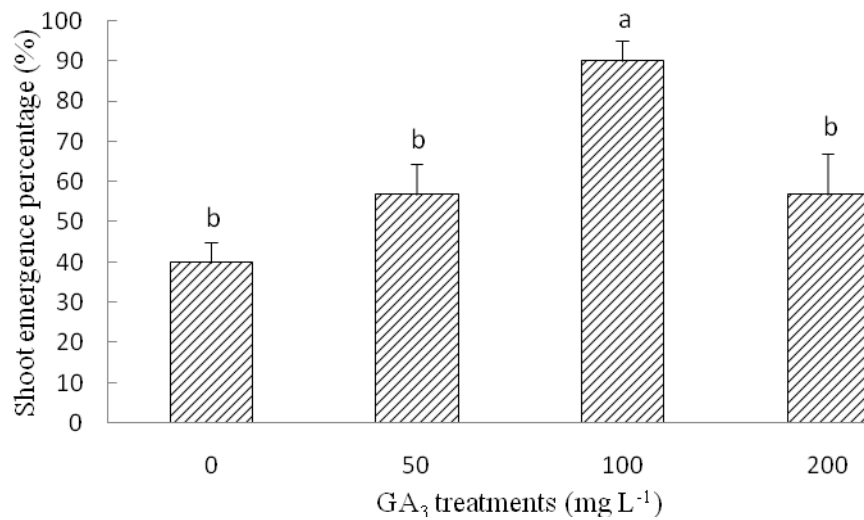
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330 **Fig. 3.** Effect of gibberellic acid (GA₃) treatment with shoot growth of *Y. longistaminea* under a
331 12/12 h photoperiod and 28/20 °C. Values are mean ± standard error (n = 3). Different letters
332 indicate significant differences between the different GA₃ solutions (P < 0.05).

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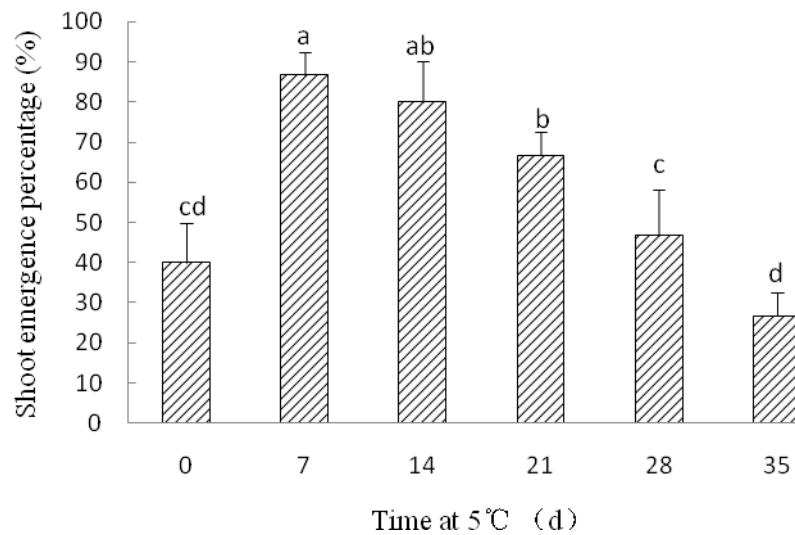
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347 **Fig. 4.** Effect of shoot growth in germinated seeds of *Y. longistaminea* following 0-35 days of cold
348 treatment at 5°C under a 12/12 h photoperiod and 28/20 °C. Values are mean \pm standard error (n =
349 3). Different letters indicate significant differences between the different cold treatment (P < 0.05).

350

351