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6	A simple method for measuring long-term integrated
7	testosterone levels in men
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9	Devorah Matas and Lee Koren [#]
10	The Mina and Everard Goodman Faculty of Life Sciences
11	Bar-Ilan University
12	Ramat Gan, 5290002
13	Israel
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16	Corresponding author:
17	[#] Lee Koren, Ph.D.
18	The Mina and Everard Goodman Faculty of Life Sciences,
19	Bar Ilan University, Ramat Gan, Israel;
20	Phone: +972-3-7384371
21	Fax: +972-3-7384372
22	E-mail: Lee.Koren@biu.ac.il
23	

24 Abstract

25	Steroids play multiple roles in the regulation of development, physiology, reproduction, and
26	behavior. Measuring circulating steroids is especially challenging since concentrations are
27	extremely labile, responding to stressors within minutes. Matrices that integrate long-term
28	steroid levels are therefore valuable as biomarkers of baseline, as well as chronic steroid
29	exposures. Here we report on a simple method to extract and measure accumulated
30	testosterone from human fingernails using commercial EIA kits. Further, we demonstrate
31	known human testosterone sex and age trends. Thus, this method is a potential tool for
32	biomonitoring endogenous as well as exogenous steroid exposure.
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54	Key words: biomonitoring, endogenous steroid levels, integrated matrices, non-invasive,
35	fingernails.
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37	Introduction
38	Steroid hormones influence and are influenced by development, physiology, and behavior.
39	Circulating and integrated steroid levels can provide information on growth and reproduction,
40	and offer prospective biomarkers for well-being and survival (Koren et al. 2012). Measuring
41	steroids in integrated matrices, such as hair, feathers, and nails, is a progressively developing
42	field. Unlike traditional blood sampling, hair and nail collection is relatively non-invasive,
43	pain and infection-free. Hair and nails may be cut and collected quickly and efficiently,
44	without the need for patient privacy or researcher health precautions. They are also easy to
45	transport and store, without needing electricity and cooling. Moreover, while circulating

- 46 steroid levels represent the momentary total (i.e., protein-bound and free) state, hair and nails
- 47 provide a longer time-frame, presumably integrating free steroids over the time of their
- 48 growth (i.e., weeks or months).

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49	In mammals, nails develop by keratinocytes cell divisions in the proximal germinal
50	regions (Baran 1981; De Berker et al. 2007). Nail growth is attributed primarily to the nail
51	matrix and to a lesser extent to the nail bed. The nail matrix is mainly composed of specific
52	keratin proteins, cross-linked by keratin-associated proteins. Diverse factors influence nail
53	growth, including numerous physiological and pathological conditions (reviewed in Zaiac &
54	Walker (Zaiac & Walker 2013)). On average, human fingernails grow at a rate of 3 mm per
55	month, while human toenails grow at a rate of 1 mm per month. During nail growth, multiple
56	materials are transferred from the capillary blood vessels nourishing the nail to the growing
57	nail (Palmeri et al. 2000). These are embedded in the nails, making this abundant matrix an
58	attractive candidate for biomonitoring multiple blood borne substances. Indeed, poisonous
59	materials (Barbosa et al. 2005; Button et al. 2009; Mehra & Juneja 2005; Suzuki et al. 1989),
60	drugs (Shu et al. 2015) and endocrine-disruptors (Li et al. 2013) have been forensically
61	detected in human nails. In addition, endogenous steroids have been analyzed in nail extracts.
62	For example, infants that experienced in utero stress had higher DHEA
63	(dehydroepiandrosterone) compared to infants from non-stressed mothers (Tegethoff et al.
64	2011). In another study, nail cortisol-to-DHEA ratio was found to be related to perceived
65	exam stress in students (Warnock et al. 2010). Exogenous testosterone and its derivatives
66	were measured in nails using LC-MS (Brown & Perrett 2011). However, the authors
67	concluded that detection of these substances was not sensitive enough. GC-MS was used in a
68	different study, yet this method requires derivatization (Choi et al. 2001). Here, we aimed to
69	develop a simple and repeatable quantitative method for measuring endogenous testosterone
70	in nails, and biologically validate it using known age-related trends.

71

72 Materials and methods

73 Nails collection

- 74 The study was approved by Bar Ilan University's Research on Human Subjects' Ethics
- 75 Committee. In order to collect nail samples, we sent an electronic mail to all graduate

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76	students, staff, and faculty members in the Faculty of Life Sciences at Bar Ilan University,
77	requesting fingernail donations from men, women and children. Samples were placed in an
78	empty paper envelope marking only the sex and age of the donator on it. Envelopes were
79	deposited anonymously in our laboratory mailbox. Between November 2014 and May 2015
80	we collected 52 samples from men (ages 18.5-74), 6 samples from boys (ages 0.4-2), 11
81	samples from women (ages 18-53) and 11 samples from girls (ages 3-9).
82	
83	Steroid extraction
84	We extracted steroids from nails using our published protocol for hair and claw-testing
85	(Koren & Geffen 2009; Koren et al. 2006; Koren et al. 2008; Koren et al. 2002; Koren et al.
86	2012; Matas et al. 2016), with several modifications. Nails were weighed to the nearest 0.01
87	mg in safe-lock polypropylene tubes (Sarstedt, Germany). Samples were ground in mixer mill
88	MM 400 (Retsch, Germany) for 10 minutes at 25 Hz. Methanol (UPLC grade, Sigma, Israel)
89	was added, and the minced samples were sonicated for 30 minutes and then incubated
90	overnight at 50°C, with gentle shaking. Next day, the supernatant was centrifuged at 17,000 g,
91	transferred into a glass tube (Corning Inc., USA) and evaporated under a stream of nitrogen.
92	Samples were reconstituted and testosterone was quantified in the assay buffer that was
93	provided with the commercial enzyme-linked immunosorbent assays (ELISA) according to
94	manufacturer's recommendations.
95	
96	Quantitation of steroids
97	High sensitivity salivary ELISA kits (Salimetrics; item no.1-2402; Ann Arbor, USA) were
98	validated for nails by examining parallelism with kit standards and linearity. Serial dilutions
99	of separate pools for men, women and children all showed parallelism with the provided kit

- standards (univariate analysis of variance in SPSS; P=0.5; P=0.46 and P=0.48 for men,
- 101 women, and children respectively). Linearity was demonstrated for men between 1-20 mg of
- 102 nail extract, corresponding to 6-230 pg/mL of kit standard. For women, linearity was

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- 107 to the manufacturer, antibody cross-reactivity was reported as 36.4% with
- dihydrotestosterone, 21.02% with 19-nortestosterone, 1.9% with 11-hydroxytestosterone, 108
- 109 1.157% with androstenedione and less than 0.489% for all other steroids. By measuring six
- 110 samples of the pool on the same ELISA plate we determined intra-assay variability to be

111 2.7%. By quantifying four samples of the pool on four different days we calculated inter-

- 112 assay variability to be 11.78%. Recovery was calculated to be 86% by the addition of a
- 113 known amount of testosterone to the nail extract. The presence of testosterone in human nails
- 114 was further validated utilizing a second antibody, by the DRG testosterone ELISA kit (DRG
- 115 International item no. EIA-1559; NJ, USA) following the manufacturer's protocol. DRG
- 116 reports that the testosterone antibody cross-reacts with 11β -Hydroxyestosterone (3.3%), 19-
- 117 Nortestosterone (3.3%) and other steroids $(\leq 0.9\%)$.
- 118

119 **Results and Discussion**

120 We developed a simple protocol for human fingernail testosterone extraction and quantitation. 121 According to our validations, whereas 5 mg of nails were needed to quantify testosterone in 122 men, ten-fold was necessary to quantify testosterone in women and girls (i.e., 50 mg). This is comparable to the documented association between circulating testosterone in men vs. women 123 (Longcope, 1986). Since a large amount of nail matrix was needed for the female analysis, we 124 125 created a pool from all donated female nails. Figure 1 shows nail testosterone in adult men (mean for N=52; 5.04 pg/mg nails), women (mean for N=11; pool; 2.3 pg/mg nails), boys 126 (mean for N=6; 1.8 pg/mg nails), and girls (mean for N=11; pool; 1.07 pg/mg nails). In order 127 to perform statistical comparisons, testosterone levels were transformed using the Johnson Su 128 129 transformation for normal distribution. All analysis was performed in JMP 12 (SAS Institute 130 Inc.). Analysis of variance showed significant differences in testosterone levels between sex

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and age groups ($F_{59,3}$ =9.665; P<0.0001). Tukey-Kramer post-hoc statistics (q=2.648; α =0.05) 131 showed significant differences in nail testosterone levels between men and boys (difference 132 1.758; P<0.0001). These results support previous findings reported in the literature where 133 134 testosterone concentrations are higher in men and women than in boys and girls (Andersson et 135 al. 1997; Mouritsen et al. 2014). In this analysis, we found a two-fold difference between men and women's testosterone levels. Established normal ranges of circulating testosterone 136 137 concentrations indicate a five-fold difference between low levels in men (usually 300 ng/dL) 138 and high levels in women (usually 60 ng/dL). This discrepancy may be attributed to sampling 139 size, age and seasonal fluctuations, as well as specific matrix effect. Whereas blood samples 140 reflect a momentary concentration, nails show accumulated long-term levels, which integrate 141 concentrations over time.

142 We found a significant association between age and testosterone in male nails 143 $(R^2=0.405; N=58; P<0.0001; Figure 2)$. As documented in the literature (Vermeulen et al. 144 1972), testosterone levels reached a maximum in men in their 20s and than gradually 145 decreased. Although the variation in circulating testosterone is high, the pattern is profound. 146 In this study, we use commercial EIA kits to quantify testosterone. Immunoassays are widely 147 used to quantify steroids due to their high sensitivity and procedural simplicity. Although 148 mass spectrometry is superior in terms of specificity and flexibility to measure multiple 149 compounds simultaneously, a large amount of tissue is initially needed, due to loss during 150 sample prep and cleanup. For example, Choi et al. (2001) used 100 mg of nail matrix to find differences between testosterone in men and women using GC-MS(Brown & Perrett 2011; 151 Choi et al. 2001), and Brown and Perrett (2011) did not find differences between males and 152 females or exogenous testosterone users and non-users (Brown & Perrett 2011). Here we 153 present a simple and reliable antibody-based method for testosterone quantitation using a 154 155 minimal amount of nail matrix, without the need of derivatisation or costly equipment. Testosterone has been implicated in various diseases, fertility issues, anxiety and stress-156 157 related disorders, as well as risk-taking, sexual, and aggressive behaviors. Nails may serve as 158 an ideal matrix for long-term measurements of baseline or chronic testosterone levels related

Peer Preprints to stable physiological or psychological states, behavioral syndromes, and personality traits, 159

160 as well as follow-up of exogenous testosterone exposure. Nail collection is relatively non-161 invasive, and samples are easy to store and safe to handle. Other advantages include their bountiful quantity and slow growth, reflecting long-term steroid concentrations. Our method 162 163 offers the possibility for researchers to quantify steroids in nails in an easy, applicable way, and document stable trends. Utilizing nails for steroid analysis also enables periodic 164 quantitation by repeated cutting, opening an exciting array of questions and comparative 165 studies biomonitoring individual differences in endogenous steroid levels, as well as exposure 166 to exogenous steroids, endocrine disruptors and other environmental pollutants that may 167 168 interfere with the hypothalamus-pituitary-gonadal pathways. 169 170 Conclusions 171 Measuring integrated steroid levels that reflect long-term baseline is highly relevant, since 172 steroid concentrations are labile. Here we present a protocol for extraction and quantitation of

173 testosterone in human fingernails. We biologically validated our methodology by

174 demonstrating known changes in testosterone levels throughout men's lives, and expected sex

- 175 differences. These allow the usage of this protocol for further comparative research and
- 176 biomonitoring.

177

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243	

244 Figure legend

246	Figure 1: Nail testosterone levels in men, women and children. Testosterone was extracted
247	from 5 mg of nail clippings of individual men (N=52) and boys (N=6). Error bars were
248	constructed using 1 standard error from the mean. For women (N=11) and girls (N=11)
249	testosterone was extracted from 50mg of pooled nails. Significant differences were seen
250	between sex and age classes (ANOVA: F _{59,3} =9.665; P<0.0001).
251	
252	Figure 2: Transformed nail testosterone as a function of male age. Transformed testosterone =
253	-0.151 + 0.0245*age - 0.0016*(age-28.06) ² . Johnson Su transformation was used to
254	normalize the distribution of testosterone. Polynomial second degree fit $R^2=0.405$;

255 N=58; P<0.0001

256 Figure 1

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Figure 2

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