# A peer-reviewed version of this preprint was published in PeerJ on 19 July 2016.

<u>View the peer-reviewed version</u> (peerj.com/articles/2128), which is the preferred citable publication unless you specifically need to cite this preprint.

Jaskuła R, Rewicz T, Płóciennik M, Grabowski M. 2016. Pleistocene phylogeography and cryptic diversity of a tiger beetle, *Calomera littoralis*, in North-Eastern Mediterranean and Pontic regions inferred from mitochondrial COI gene sequences. PeerJ 4:e2128 <a href="https://doi.org/10.7717/peerj.2128">https://doi.org/10.7717/peerj.2128</a>



# Pleistocene phylogeography of tiger beetle, *Calomera littoralis*, in North-Eastern Mediterranean and Pontic regions inferred from mitochondrial COI gene sequence

Radomir Jaskuła, Tomasz Rewicz, Mateusz Płóciennik, Michał Grabowski

**Background.** Calomera littoralis is a Palearctic species, widely distributed in Europe; inhabiting predominantly its Atlantic, Mediterranean and Black Sea coastlines.

**Methods.** Its phylogeography on the Balkan Peninsula and on the north-western Black Sea coast was inferred using a 697 bp long portion of the mitochondrial COI gene, amplified from 169 individuals collected on 43 localities.

**Results.** The results revealed two genetically divergent groups/lineages, the southern one inhabiting both the Balkan Peninsula and the Pontic Region and the northern one found exclusively in the Pontic Region. Species delimitation based on DNA barcoding gap suggested an interspecific level of divergence between these groups. Multivariate analysis of eight male and female morphometric traits detected no difference between the groups, implying they may represent cryptic species. The Bayesian time-calibrated reconstruction of phylogeny suggested that the lineages diverged ca. 2.3 Ma, in early Pleistocene.

**Discussion.** This might result from contemporary isolation of the Pontic basin from the Mediterranean that broke the continuous strip of coastal habitats inhabited by *C. littoralis*. Demographic analyses indicated that both lineages have been in demographic and spatial expansion since ca. 0.15 Ma. It coincides with the terminal stage of MIS-6, i.e. Wartanian/Saalian glaciation, and beginning of MIS-5e, i.e. Eemian interglacial, during which, due to eustatic sea level rise, a wide connection between Mediterranean and the Pontic basin was re-established. This, along with re-appearance of coastal habitats could initiate north-east expansion of the southern lineage and its secondary contact with the northern one. Isolation of Pontic basin from Mediterranean during the Weichselian glaciation did not have probably any effect on their phylogeography.



### NOT PEER-REVIEWED

Pleistocene phylogeography of tiger beetle, Calomera littoralis, in North-Eastern 2 Mediterranean and Pontic regions inferred from mitochondrial COI gene sequences 3 Radomir Jaskuła<sup>1\*</sup>, Tomasz Rewicz<sup>2</sup>, Mateusz Płóciennik<sup>1</sup>, Michał Grabowski<sup>1</sup> <sup>1</sup>Department of Invertebrate Zoology and Hydrobiology, University of Lodz, Banacha 12/16, 4 5 90-237 Lodz, Poland 6 <sup>2</sup>Laboratory of Microscopic Imaging and Specialized Biological Techniques, University of 7 Lodz, Banacha 12/16, Łódź 90-237, Poland 8 9 10 Corresponding author: 11 Radomir Jaskuła<sup>1</sup> 12 Ul. Banacha 12/16, 90-237 Łódź, Łódź, Poland Email address: radekj@biol.uni.lodz.pl 13 14 15

17 Abstract

18 **Background.** Calomera littoralis is a Palearctic species, widely distributed in Europe; 19 inhabiting predominantly its Atlantic, Mediterranean and Black Sea coastlines. 20 Methods. Its phylogeography on the Balkan Peninsula and on the north-western Black Sea 21 coast was inferred using a 697 bp long portion of the mitochondrial COI gene, amplified from 22 169 individuals collected on 43 localities. 23 **Results.** The results revealed two genetically divergent groups/lineages, the southern one 24 inhabiting both the Balkan Peninsula and the Pontic Region and the northern one found 25 exclusively in the Pontic Region. Species delimitation based on DNA barcoding gap 26 suggested an interspecific level of divergence between these groups. Multivariate analysis of 27 eight male and female morphometric traits detected no difference between the groups, 28 implying they may represent cryptic species. The Bayesian time-calibrated reconstruction of 29 phylogeny suggested that the lineages diverged ca. 2.3 Ma, in early Pleistocene. 30 **Discussion.** This might result from contemporary isolation of the Pontic basin from the 31 Mediterranean that broke the continuous strip of coastal habitats inhabited by C. littoralis. 32 Demographic analyses indicated that both lineages have been in demographic and spatial 33 expansion since ca. 0.15 Ma. It coincides with the terminal stage of MIS-6, i.e. 34 Wartanian/Saalian glaciation, and beginning of MIS-5e, i.e. Eemian interglacial, during 35 which, due to eustatic sea level rise, a wide connection between Mediterranean and the Pontic 36 basin was re-established. This, along with re-appearance of coastal habitats could initiate 37 north-east expansion of the southern lineage and its secondary contact with the northern one. 38 Isolation of Pontic basin from Mediterranean during the Weichselian glaciation did not have 39 probably any effect on their phylogeography. 40



- 41 **Key words:** Coleoptera Cicindelidae ice age interglacial sea level changes population
- 42 expansion southern Europe

44

#### Introduction

45 The Eastern Mediterranean including the Pontic area is recognised as one of the major 46 biodiversity and endemism hot spots on a global scale as well as a major glacial refugium in 47 Europe (eg. Myers et al., 2000; Kotlík, Bogutskaya & Ekmekçi, 2004; Blondel et al., 2010). 48 Among others, it is a consequence of complex geological history of the region that was an 49 archipelago and united with rest of the European continent only in Neogene (Pffifner, 2014). 50 On the other side, a shallow epicontinental sea, Paratethys, occupied vast areas of the continent 51 and regressed gradually leaving relics, such as Black, Azov and Caspian Sea (Nahavandi et al., 52 2013). Local isostatic and eustatic changes of sea level were among superior phenomena 53 shaping local landscapes. For example, there were at least twelve saline water intrusions from 54 the Mediterranean Sea, and eight intrusions from the Caspian Lake to the Black Sea during the 55 last 0.67 million years (Ma) (Badertscher et al., 2011). Inevitably, they played an important role 56 in modelling diversity and distribution patterns for numerous organisms, particularly those 57 inhabiting coastal ecosystems both in Mediterranean and in Pontic area. However, the evidence 58 comes mostly from aquatic, predominantly marine or brackish water, taxa (e.g. Audzijonyte, 59 Daneliya & Vainola, 2006; Neilson & Stepien, 2011). There is a deficiency of studies focusing 60 upon coastal species inhabiting terrestrial habitats in this region (Akin et al., 2010). 61 Tiger beetles, Cicindelidae Latreille, 1806, seem to be ideal model organisms to test such 62 assumptions. The family, with more than 2600 species, has a worldwide distribution with 63 exception of polar regions and some oceanic islands (Pearson & Cassola 2005). Most species, 64 both in larval and adult stage, prefer various types of sandy areas and are habitat specialists; often inhabiting coastal areas (Pearson & Vogler 2001). Several studies dealt with 65

### NOT PEER-REVIEWED

phylogeography of tiger beetles in various regions of the world (eg. Vogler et al., 1993; Cardoso & Vogler, 2005; Woodcock et al., 2007), yet so far only few focused on the role of sea level 67 oscillations in their evolutionary history (Vogler & DeSalle, 1993; Sota et al., 2011) or 68 69 compared the diversity patterns on both, the molecular and morphological, levels (Cardoso, 70 Serrano & Vogler, 2009; Tsuji et al., 2016). 71 The tiger beetle, Calomera littoralis Fabricius 1787, is widely distributed in Palaearctic, 72 from the Iberian Peninsula and Morocco in the west to the Middle Asia and Russian Far East 73 in the east (Putchkov & Matalin, 2003; Serrano, 2013; Jaskuła, 2011, 2015). Generally, it is 74 recognised as euryoecious (Jaskuła, 2011, 2013, 2015). However, in Europe it occupies predominantly the very narrow stretch of Atlantic, Mediterranean and Black Sea coastal 75 76 habitats (Cassola & Jaskuła, 2004; Franzen, 2006; Jaskuła, 2007a-b; Jaskuła, Peśić & Pavicević, 77 2005; Serrano, 2013). 78 Our study aims at (1) exploring and comparing spatial patterns of molecular and 79 morphological diversity of this coastal species in the Mediterranean and Pontic region, (2) interpreting the observed patterns in the context of local paleogeography. 80

81

82

84

85

86

87

88

#### Material and methods

### 83 Sample collection and identification

- In total, 169 imagines of *Calomera littoralis* were collected with entomological hand net on 43 sites on the Mediterranean coasts of the Balkan Peninsula, Crete and Turkey as well as on the northern and western coast of the Black and Azov Seas, in the years 2009-2012 (Figure 1, Table 1). At a site the material was fixed in 96% ethanol for DNA preservation. Taxonomic identification of the collected material followed Mandl (1981).
- 89 DNA extraction, amplification and sequencing

90

91 from all the collected individuals. Air-dried DNA pellets were eluted in 100 µl of TE buffer, 92 pH 8.00, stored at 4°C until amplification, and subsequently at -20°C for long-term storage. 93 Fragments of mitochondrial cytochrome oxydase subunit I gene (COI), ca. 700 bp long, 94 were amplified using the Jerry and Pat pair of primers (Simon et al., 1994). Each PCR reaction 95 was conducted in a total volume of 10 µl and contained DreamTag Master Mix (1x) Polymerase (ThermoScientific), 200nM of each primer and 1 µl of DNA template. The thermal regime 96 97 consisted of initial denaturation at 94°C for 2min, followed by 34 cycles of denaturation at 94°C 98 for 30s, annealing at 44°C for 30s, and elongation at 72°C for 60s, completed by a final 99 extension at 72°C for 10min. The amplified products were visualized on 2.0% agarose gels 100 stained with MidoriGreen (Nippon Genetics) to verify the quality of the PCR reactions. Then, 101 the PCR products were chemically cleaned up of dNTPs and primer residues by adding 1U of 102 Exonuclease I (Thermo Scientific) and 2U of FastAP Alkaline Phosphatase (Thermo Scientific) 103 per sample. Sequencing was performed using BigDye sequencing protocol (Applied 104 Biosystems 3730xl) by Macrogen Inc., Korea. 105 Molecular data analysis 106 First, all the obtained sequences were positively verified as Calomera DNA using 107 GenBankBLASTn searches (Altschul et al., 1990). Then they were edited and assembled with 108 CLUSTALW algorithm (Chenna et al., 2003) using BIOEDIT© 7.2.5. The resulting alignment was 109 697 bp long with no gaps, and composed of 169 COI sequences. The sequence data and trace 110 files were uploaded to BOLD and subsequently also to GenBank (accession nos KU905171-111 KU905339). 112 Pairwise Kimura 2-parameter (K2p) distances between sequences were estimated using 113 MEGA 6.2 (Tamura et al., 2013). Haplotypes were retrieved using DNASP v5 (Librado & Rozas, 114 2009). Phylogenetic relationships between the haplotypes were visualised with phylogenetic

Following Hillis et al. (1996) the standard phenol-chloroform method was used to extract DNA



network computed using the neighbour-net algorithm and uncorrected p-distances in SplitsTree ver. 4.13.1 (Huson & Bryant 2006).

To test for presence of distinct operational taxonomic units (OTUs) that may represent potential cryptic species/subspecies in the sequenced pool of individuals we used the Automatic Barcode Gap Discovery (ABGD) procedure (Puillandre et al., 2012). The default value of 0.001 was used as the minimum allowed intraspecific distance. The maximum allowed intraspecific distance was set to P max = 0.03 and 0.06, as both threshold values have been already used in literature to delimit insect species (Hebert et al., 2003; Hebert, Ratnasingham & de Waard, 2003). We applied the K2P model sequence correction, which is a standard for barcode analyses (Hebert et al., 2003). We used primary partitions as a principal for group definition for they are usually stable over a wider range of prior values, minimise the number of false positive (over split species) and are usually close to the number of groups described by taxonomists (Puillandre et al., 2012).

To reveal the temporal framework for the divergence of the two phylogenetic lineages (potential cryptic species) revealed in *Calomera littoralis*, the time calibrated phylogeny was reconstructed in BEAST, version 1.8.1 (Drummond et al., 2012). A COI sequence of *Calomera lugens aphrodisia* Baudi di Selve 1864 from GenBank (acc. no. KC963733) was used as an outgroup. This analysis was performed on a reduced dataset, containing only the most distant haplotypes from each OTU. The HKY model of evolution and coalescent model were set for priors. The strict clock with rate 0.0115, widely used for phylogenetic studies upon insects, was applied for the analyses (Brower, 1994). Five runs of 20M iterations of Markov chain Monte Carlo (MCMC) sampled each 2000 iterations were performed. The runs were examined using Tracer v 1.6 and all sampled parameters achieve sufficient effective sample sizes (ESS>200). Tree files were combined using Log-Combiner 1.8.1 (Drummond et al., 2012), with removal of

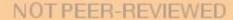
the non-stationary 20% burn-in phase. The maximum clade credibility tree was generated using
TreeAnnotator 1.8.1 (Drummond et al., 2012).

To provide insight into historical demography, i.e. the temporal changes of the effective population size of *Calomera littoralis* in the studied region, we performed Bayesian Skyline Plot (BSP) analysis (Drummond et al., 2005) in BEAST, version 1.8.1 (Drummond et al., 2012). Separate analysis was performed for each of the two phylogenetic lineages revealed in our study (see Results). The Northern Lineage was represented by 84 individuals from 22 localities, while the Southern Lineage was represented by 85 individuals from 32 localities. The HKY+I model of evolution was used as the best fitting model in case of the Eastern Lineage, while TN93+I was used in case of the Western Lineage. Two runs of MCMC, 20M iterations long sampled each 2000 iterations, were performed. In both cases the runs were examined using Tracer v 1.6 (Drummond et al., 2012) and all sampled parameters achieved sufficient effective sample sizes (ESS>200).

Two models of population expansion, demographic and spatial, were examined using mismatch distribution analysis (Slatkin & Hudson, 1991; Rogers & Harpending, 1992) and Tajima's D neutrality test (Tajima, 1989). Analyses were performed for the COI groups, using Arlequin 3.5.1.3 (Excoffier & Lischer, 2010) with 1000 replicates.

#### Morphometric data analysis

To test whether variation of morphometric traits reflects presence of two genetically divergent lineages (potential cryptic species), measurements of eight body parameters (Fig. 2) were taken from all the 69 males and 100 females used previously for the molecular analyses: 1 – right mandible length (RML), 2 – length of head (LH), 3 – width of head (WH), 4 – maximum pronotum width (MPW), 5 – pronotum length (PL), 6 – elytra length (EL), 7 – maximum elytra width (MEW), and 8 – total body length (TBL). The principal component analysis (PCA) was performed separately for each sex (Figure 3, Table 1). To test for significance (p<0.01) of



morphological differences (separately for males and females) between the two divergent lineages one-way ANOSIM Pairwise Test was performed. All the above statistical analyses were done with PRIMER 6 software (Clarke & Gorley, 2006).

#### Results

#### Molecular data

A total of 81 haplotypes were identified in the dataset composed of 169 individuals from 43 sites from the Mediterranean and the Pontic areas (Table 1). The phylogenetic network illustrating phylogenetic relationships among haplotypes (Fig.4) uncovered presence of two distinct haplotype groups (phylogenetic lineages). The first group, from now on defined as southern lineage, includes 36 haplotypes present all over the studied range including the Balkan Peninsula and the Pontic area. The other group, from now on defined as northern lineage, is composed of 45 haplotypes present exclusively along the north-western coast of the Black Sea. The mean K2p genetic distance between both groups of haplotypes is relatively high (0.039, SD 0.007). Both variants of the ABGD analysis resulted in partitioning of the dataset into two OTUs, that may represent distinct operational taxonomic units – potential cryptic species or subspecies within *Calomera littoralis* in the studied area (Fig. 3A).

The Bayesian time-calibrated reconstruction of phylogeny shows that the two lineages split at ca. 2Ma, i.e. in early Pleistocene (Fig. 5A). Results of the BSP analyses showing the temporal changes of the effective population size suggests that both lineages experienced rapid population growth that has started ca. 0.15Ma (Fig. 5B). In both cases, a small decline in effective population size may be observed in most recent times (<0.05 Ma). Results of the mismatch analysis shows that both lineages are currently in the stage of both demographic and spatial expansion (Fig 5C). Interestingly, geographical distribution of both lineages shows that the spatial expansion of southern lineage was efficient enough to spread eastwards into the

Black Sea and colonise effectively the north-western Black Sea coast. The northern lineage has

spread only in the Pontic region.

### Morphometric data

The results of PCA and ANOSIM revealed no differences in the analysed morphometric traits

between the southern and the northern lineages, neither in males nor in females (Fig. 3B). In

PCA (Fig. 3B), a very weak gradient (R=0.03) could be seen in case of female body length.

Females from the northern lineage clade seem to be slightly larger than those from the southern

one (body length; ANOSIM Pairwise Tests p=0.03).

#### **Discussion and conclusions**

### Cryptic diversity of Calomera littoralis

Known as very important hotspot of biodiversity, endemicity and cryptic diversity (eg. Myers et al., 2000; Kryštufek & Reed, 2004; Huemer & Timossi, 2014; Blondel et al., 2010; Previšić et al., 2014; Caković *et al.*, 2015), the southern Europe holds also most diverse tiger beetle fauna in the entire Palearctic realm (Jaskuła, 2011). Presence of cryptic diversity was already pointed out for *Cicindela hybrida* in the Mediterranean (Cardoso, Serrano & Vogler, 2009) as well as for several species of tiger beetles occurring in other part of the world (Vogler & Pearson, 1996; López-López, Hudson & Galián, 2012). Thus, existence of well-defined OTUs within *Calomera littoralis* is not surprising in the studied area. The level of divergence, 0.04 K2p distance, between the northern and the southern lineage is similar as those found between species of tiger beetles in other studies (eg. Cardoso & Vogler, 2005; López-López, Abdul Aziz & Galián, 2015). Interestingly, we could not detect any conclusive morphological differences between the two lineages based on the multivariate analysis of eight morphometric traits. It must be mentioned that three subspecies of *Calomera littoralis*, described on the basis of morphology, were reported from the studied area: *C. l. nemoralis* from all the studied Balkan

countries, Crete, Moldova, western Ukraine and western Turkey; *C. l. conjunctaepustulata* (Dokhtouroff, 1887) from the Azov Sea area; *C. l. winkleri* (Mandl, 1934) from Crete and the coastal zone of southern Turkey (Werner, 1991; Putchkov & Matalin, 2003; Avgın & Özdikmen, 2007). However, the morphological differences between the subspecies, such as body size, maculation of elytra and shape of aedeagus, are poorly defined and did not allow the identification of the studies material further than to the species level. Unfortunately, we had no opportunity to study the topotypical material – Provence, France, is *locus typicus* for *C. l. nemoralis*, Tibet for *C. l. conjunctaepustulata*, and Cyprus for *C. l. winkleri*. Thus, we cannot exclude a possibility that the two lineages we found in our material overlap with any of the above mentioned subspecies. However, only a further taxonomic revision combining more phenotypic traits, including e.g. cuticle ultrastructure, with several, mitochondrial and nuclear DNA data, could help to resolve this problem. Until such revision is done, we propose to use the tentative name "*Calomera littoralis* complex" for populations from the studied area.

### Phylogeography of Calomera littoralis

Occurrence of *C. littoralis* in Europe is restricted predominantly to marine shorelines with sandy beaches and salt marshes as main habitats (e.g. Franzen, 2006; Jaskuła, 2011; Serrano, 2013). In the eastern Mediterranean it is distributed continuously all along the Adriatic and Aegean coasts, Turkish Straits and the Black Sea coastline (Cassola & Jaskuła, 2004; Jaskuła Peśić & Pavicević 2005; Franzen, 2006; Jaskuła, 2007a; Jaskuła 2007b). However, pronounced genetic structure with two divergent operational taxonomic units (OTUs) implies prolonged spatial isolation in the evolutionary past of this species. The observed level of divergence indicates that this isolation initiated an allopatric speciation. Their present distribution i.e. sympatry in the Pontic region reveals secondary contact of the already divergent lineages in this area. The Bayesian time-calibrated reconstruction of phylogeny shows that split between these OTUs begun in early Pleistocene. This coincides with beginning of recurrent glaciations

239

240

241

242

243

244

245

246

247

248

249

250

251

252

253

254

255

256

257

258

259

260

261

262

263

resulting in eustatic sea level changes and climate aridisation that ever since dominated the global climate and landscape/habitat distribution (Fagan, 2009). In the Mediterranean and in the Pontic region such global effects overlaid and strengthen the local effects of tectonic plate collision leading to Alpine orogeny, i.e. local land uplift and subsidence resulting in isostatic sea level changes, salinity fluctuations from freshwater to fully marine and habitat mosaicism (Stanley & Blanpied, 1980). For example, during that time the connections of Pontic basin to Mediterranean Sea was lost and regained for more than a dozen times (Kerey et al., 2004; Badertscher et al., 2011). A profound impact of these events on the evolution and, hence, distribution of local both aquatic (Audzijonyte, Daneliya & Vainola, 2006; Nahavandi et al., 2013) and terrestrial taxa (eg. Böhme et al., 2007; Ferchaud et al., 2012). We can assume that in case of C. littoralis, a halophilic species bound to coastal habitats, sea level fluctuations would significantly affect its distribution. The 2 Ma divergence time for C. littoralis OTUs derived from our data coincides with one particular disconnection of the Mediterranean and Pontic basins. At that time, from ca. 2 to ca. 1.5 Ma, the Meothic Sea, one of several predecessors of the Black Sea, turned into the predominantly freshwater Pontos Sea/Lake (Grinevetsky et al., 2015). This surely broke the formerly continuous stretch of coastal habitats connecting the two basins and thus, could be an effective barrier leading to split of C. littoralis population into the allopatric southern and northern lineages. Their detailed history is impossible to unravel, yet results of BSP analyses reconstructing past changes in effective population size indicate that both lineages started their demographic expansions at ca. 0.15 Ma. This coincides with the terminal stage of MIS-6, i.e. Wartanian/Saalian glaciation, and beginning of MIS-5e, i.e. Eemian interglacial (Lisiecki & Raymo, 2005; Marks, 2011). The latter was characterized by warmer climate and sea level higher by 6-9 m in comparison to Holocene (Kopp et al., 2009; Dutton & Lambeck, 2012). In result, a wide connection between Mediterranean and the Pontic basin was re-established and the coastal habitats extended again,

264

265

266

267

268

269

270

271

272

273

274

275

276

277

278

279

280

281

282

283

284

285

enabling exchange of faunas. Due to deficiency of local studies, it is hard to compare our results to evolutionary history of any other terrestrial taxa in the area. However, a wealth of studies showing very similar spatiotemporal scenario in animal taxa comes from the coastal regions of the Gulf of Mexico and the adjacent Atlantic coast (summarised by Avise, 1992). During Pleistocene, Cuba was connected with a land bridge to the Florida Peninsula what lead to divergence of populations of several terrestrial and aquatic animals, including also a local tiger beetle species Cicindela dorsalis Say, 1817 (Vogler & DeSalle, 1993). Interestingly enough, however according to our results both lineages are until now in the stage of demographic and spatial expansion, only the southern one has crossed the present Turkish straits. This asymmetry is hard to explain. Another interesting fact is that the isolation of Pontic basin from Mediterranean during the following Weichselian glaciation did not have probably any effect on the demography and phylogeography of the species. Based on the mitochondrial DNA marker only we cannot also conclude, whether the secondary contact of the divergent lineages effected in hybridization and or introgression. Answering this question requires employment of nuclear marker, what leaves a space for the future studies – much wider in terms of geographic coverage and molecular markers used. Concluding, we have demonstrated that Pleistocene glaciations and associated sea level changes in the Mediterranean/Pontic region had a profound effect on the genetic diversity and distribution of widely distributed coastal insect species, generating some level of cryptic diversity. Our case study casts more light on the evolutionary relationships between populations of terrestrial animals inhabiting both the Mediterranean and Black Sea shorelines - a phenomenon that is still weakly explored in literature.

286

287

#### Acknowledgements

- 288 The first author would like to thank to Iwona Jaroszewska, Piotr Jóźwiak, Błażej Pawicki,
- 289 Maciej Podsiadło, Anna Stepień, and Bartosz Ukleja for their kind help during and material
- 290 collecting TB-Quest Expedition to the Balkans.

- 292 Literature
- 293 Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic Local Alignment Search
- 294 Tool. *Journal of Molecular Biology* 215: 403-410.
- Akin C, Bilgin CC, Beerli P, Westaway R, Ohst T, Litvinchuk SN, Uzzell T, Bilgin M, Hotz
- 296 H, Guex GD, Plötner J. 2010. Phylogeographic patterns of genetic diversity in eastern
- 297 Mediterranean water frogs were determined by geological processes and climate change in the
- 298 Late Cenozoic. Journal of Biogeography 37 (11): 2111-2124.
- 299 Audzijonyte A, Daneliya ME, Vainola R. 2006. Comparative phylogeography of Ponto-
- 300 Caspian mysid crustacean: isolation and exchange among dynamic inland sea basins.
- 301 *Molecular Ecology* 15: 2969–2984.
- Avgin S, Özdikmen H. 2007. Check-list of tiger beetles of Turkey with review of distribution
- and biogeography (Coleoptera: Cicindelidae). *Munis Entomology and Zoology* 2 (1): 87-102.
- 304 Avise JC. 1992. Molecular population structure and the biogeographic history of a regional
- fauna: a case history with lessons for conservation biology. *Oikos* 63: 62-76.
- 306 Badertscher S, Fleitmann D, Cheng H, Edwards RL, Göktürk OM., Zumbühl A, Leuenberger
- M, Tüysüz O. 2011. Pleistocene water intrusions from the Mediterranean and Caspian seas
- into the Black Sea. *Nature Geoscience* 4: 236–239.
- 309 Blondel J, Aronson J, Bodiou J-Y, Boeuf G. 2010. The Mediterranean Region. Biological
- 310 *Diversity in Space and Time*. 2<sup>nd</sup> Edition. Oxford: Oxford University Press.

- Böhme MU, Fritz U, Kotenko T, Džukić G, Ljubisavljević K, Tzankov N, Berendonk TU.
- 312 2007. Phylogeography and cryptic variation within the *Lacerta viridis* complex (Lacertidae,
- 313 Reptilia). Zoologica Scripta 36: 119–131.
- 314 Brower AVZ. 1994. Rapid morphological radiation and convergence among races of the
- 315 butterfly *Heliconius erato* inferred from patterns of mitochondrial DNA Evolution.
- 316 Proceedings of the National Academy of Sciences USA 91: 6491-6495.
- Caković D, Stešević D, Schönswetter P, Frajman B. 2015. How many taxa? Spatiotemporal
- 318 evolution and taxonomy of *Amphoricarpos* (Asteraceae, Carduoideae) on the Balkan
- Peninsula. Organisms Diversity & Evolution 15(3): 429-445.
- 320 Cardoso A, Vogler AP. 2005. DNA taxonomy, phylogeny and Pleistocene diversification of
- 321 the Cicindela hybrida species group (Coleoptera: Cicindelidae). Molecular Ecology 14: 3531-
- 322 3546.
- 323 Cardoso A, Serrano A, Vogler AP. 2009. Morphological and molecular variation in tiger
- beetles of the *Cicindela hybrida* complex: is an 'integrative' taxonomy possible? *Molecular*
- 325 Ecology 18: 648-664.
- 326 Cassola F, Jaskuła R. 2004. Material to the knowledge of the tiger beetles of Romania
- 327 (Coleoptera: Cicindelidae). *Polskie Pismo Entomologiczne* 73: 193–214.
- 328 Chenna R, Sugawara H, Koike T, Lopez R, Gibson TJ, Higgins DG, Thompson JD. 2003.
- 329 Multiple sequence alignment with the Clustal series of programs. *Nucleic Acids Research* 31:
- 330 3497–3500.
- Clarke KR, Gorley RN. 2006. *PRIMER v6: User Manual/Tutorial*. PRIMER-E, Plymouth.
- Drummond AJ, Rambaut A, Shapiro B, Pybus OG. 2005. Bayesian coalescent inference of
- past population dynamics from molecular sequences. *Molecular Biology and Evolution* 22:
- 334 1185-1192.

- Drummond AJ, Suchard MA, Xie D, Rambaut A. 2012. Bayesian Phylogenetics with BEAUti
- and the BEAST 1.7. Molecular Biology and Evolution 29: 1969–1973.
- Dutton A, Lambeck K. 2012. Ice volume and sea level during the last interglacial. *Science*
- 338 337 (6091): 216–219.
- Excoffier L, Lischer HEL. 2010. Arlequin suite ver 3.5: a new series of programs to perform
- population genetics analyses under Linux and Windows. *Molecular Ecology Resources* 10:
- 341 564-567.
- Fagan B (ed.). 2009. The complete Ice Age. How climate change shaped the world. London:
- 343 Thames & Hudson Ltd.
- Ferchaud A-L, Ursenbacher S, Cheylan M, Luiselli L, Jelić D, Halpern B, Major A, Kotenko
- T, Keyan N, Behrooz R, Crnobrnja-Isailović J, Tomović L, Ghira I, Ioannidis Y, Arnal V,
- Montgelard C. 2012. Phylogeography of the *Vipera ursinii* complex (Viperidae):
- 347 mitochondrial markers reveal an east–west disjunction in the Palaearctic region. *Journal of*
- 348 *Biogeography* 39: 1836–1847.
- Franzen M. 2006. Verbreitung und Lebensräume der Sandlaufkäfer der Peloponnes-Halbinsel,
- 350 Griechenland (Coleoptera, Cicindelidae). Nachrichtenblatt der Byerischen Entomologen 55
- 351 (3/4): 46–64.
- 352 Grinevetsky SR, Zonn IS, Zhiltsov SS, Kosarev AN, Kostianoy AG. 2015. The Black Sea
- 353 Encyclopedia. Berlin: Springer.
- Hebert PDN, Cywinska A, Ball SL, deWaard JR. 2003a. Biological identification through
- 355 DNA barcodes. to the fast publication and worldwide dissemination of high-quality research.
- 356 Proceedings of the Royal Society of London B: Biological Sciences 270: 313–321.
- 357 Hebert PDN, Ratnasingham S, deWaard JR. 2003b. Barcoding animal life: cytochrome c
- oxidase subunit 1 divergences among closely related species. *Proceedings of the Royal*
- 359 *Society of London B: Biological Sciences* Suppl. 27: 96–99.



- 360 Hillis DM, Moritz C, Mable BK. 1996. *Molecular Systematics*, 2<sup>nd</sup> ed. Sunderland: Sinauer
- 361 Associates.
- Huemer P., Timossi G. 2014. Sattleria revisited: unexpected cryptic diversity on the Balkan
- Peninsula and in the south-eastern Alps (Lepidoptera: Gelechiidae). Zootaxa 3780 (2): 282-
- 364 296.
- 365 Huson DH, Bryant D. 2006. Application of Phylogenetic Networks in Evolutionary Studies,
- 366 *Molecular Biology and Evolution* 23(2): 254-267.
- Jaskuła R. 2007a. Remarks on distribution and diversity of the tiger beetle fauna (Coleoptera:
- 368 Cicindelidae) of Albania. Fragmenta Faunistica 50 (2): 127–138.
- Jaskuła R. 2007b. Furthern records of tiger beetles from Romania (Coleoptera: Cicindelidae).
- 370 *Cicindela* 39 (1–2): 27–34.
- Jaskuła R. 2011. How unique is the tiger beetle fauna (Coleoptera, Cicindelidae) of the
- 372 Balkan Peninsula? *ZooKeys* 100: 487-502.
- Jaskuła R. 2013. Unexpected vegetarian feeding behaviour of a predatory tiger beetle
- 374 Calomera littoralis nemoralis (Olivier, 1790) (Coleoptera: Cicindelidae). Journal of the
- 375 Entomological Research Society 15(1): 01–06.
- Jaskuła R. 2015. The Maghreb one more important biodiversity hot spot for tiger beetle
- fauna (Coleoptera, Carabidae, Cicindelinae) in the Mediterranean region. ZooKeys 482: 35-
- 378 53.
- Jaskuła R, Peśić V, Pavicević D. 2005. Remarks on distribution and diversity of the tiger
- beetle fauna of Montenegro (Coleoptera: Cicindelidae). Fragmenta Funistica 4 (1): 15–25.
- 381 Kerey IE, Meric E, Tunoglu C, Kelling G, Brenner RL, Dogan AU. 2004. Black Sea-
- Marmara Sea Quaternary connections: new data from the Bosphorus, Istanbul, Turkey.
- 383 Palaeogeography, Palaeoclimatology, Palaeoecology 204: 277–295.



- Kopp RE, Simons FJ, Mitrovica JX, Maloof AC, Oppenheimer M. 2009. Probabilistic
- assessment of sea level during the last interglacial stage. *Nature* 462 (7275): 863–867.
- Kotlík P, Bogutskaya NG Ekmekçi FG. 2004. Circum Black Sea phylogeography of *Barbus*
- freshwater fishes: divergence in the Pontic glacial refugium. *Molecular Ecology* 13: 87-95.
- 388 Kryštufek B, Reed M. 2004 Patterns and Process in Balkan Biodiversity an overview. In:
- Griffiths HI, Kryštufek B, Reed JM. (Eds) Balkan Biodiversity: Pattern and Process in the
- 390 European Hotspot. Dordrecht, Kluwer Academic Publishers, 203–217.
- 391 Librado P, Rozas J. 2009. Dna SPv5: a software for comprehensive analysis of DNA
- 392 polymorphism data. *Bioinformatics* 25: 1451–1452.
- 393 Lisiecki LE., Raymo ME. 2005. A Pliocene-Pleistocene stack of 57 globally distributed
- benthic  $\delta^{18}$ O records. *Paleoceanography* 20 (10): 1-17.
- 395 López-López A, Abdul Aziz A, Galián J. 2015. Molecular phylogeny and divergence time
- 396 estimation of Cosmodela (Coleoptera: Carabidae: Cicindelinae) tiger beetle species from
- 397 Southeast Asia. *Zoologica Scripta* 44: 437-445.
- 398 López-López A, Hudson P, Galián J. 2012. The blackburni/murchisona species complex in
- 399 Australian *Pseudotetracha* (Coleoptera: Carabidae: Cicindelinae: Megacephalini): evaluating
- 400 molecular and karyological evidence. Journal of Zoological Systematics and Evolutionary
- 401 Research 50 (3): 177-183.
- 402 Mandl K. 1981. Revision der unter Cicindela lunulata F. im Weltkatalog der Cicindelinae
- 403 zusammengefaßten Formen (Col., Cicindelidae). Entomologische Arbeiten aus dem Museum
- 404 *Gg. Frey* 29: 117-176.
- 405 Marks L. 2011. Quaternary glaciations in Poland. [in:]: EhlersJ., GibbardP.L., Hughes P.D.
- 406 (eds.) Quaternary Glaciations Extent and Chronology, a closer look. *Developments in*
- 407 *Quaternary Science* 15: 299-303.

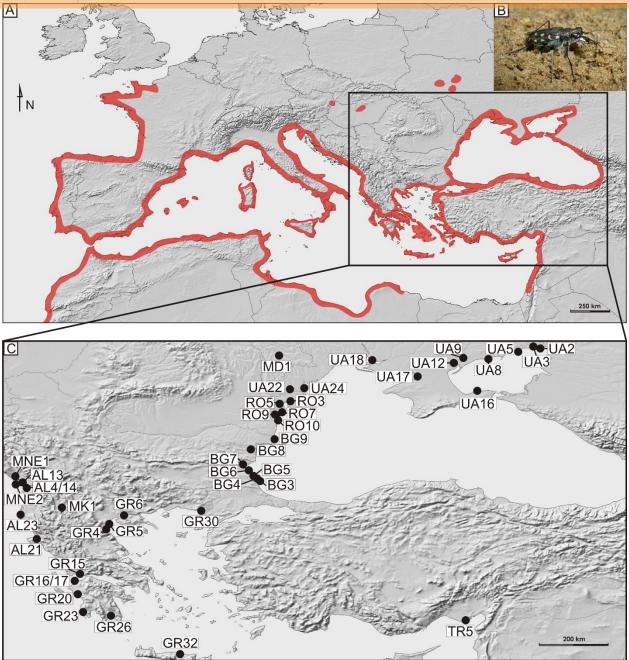


- Myers N, Mittermeier RA, Mittermeier CG, da Fonseca GAB, Jennifer K. 2000. Biodiversity
- 409 hotspots for conservation priorities. *Nature* 403: 853–858.
- Nahavandi N, Ketmaier V, Plath M, Tiedemann R. 2013. Diversification of Ponto-Caspian
- 411 aquatic fauna: Morphology and molecules retrieve congruent evolutionary relationships in
- 412 Pontogammarus maeoticus (Amphipoda: Pontogammaridae). Molecular Phylogenetics and
- 413 Evolution 69: 1063–1076.
- Neilson ME, Stepien CA. 2011. Historic speciation and recent colonization of Eurasian
- 415 monkey gobies (*Neogobius fluviatilis* and *N. pallasi*) revealed by DNA sequences,
- 416 microsatellites, and morphology. *Diversity and Distributions* 17: 688–702.
- Pearson DL, Cassola F. 2005. A quantitative analysis of species descriptions of tiger beetles
- 418 (Coleoptera: Cicindelidae), from 1758 to 2004, and notes about related developments in
- 419 biodiversity studies. *The Coleopterologist Bulletin* 59 (2): 184–193.
- 420 Pearson DL, Vogler AP. 2001. Tiger beetles: the evolution, ecology, and diversity of the
- 421 *cicindelids*. Ithaca and London: Cornell University Press.
- 422 Pfiffner OA. 2014. *Geology of the Alps*. New York: Wiley Blackwell.
- 423 Previšić A, Wolfram G, Vitecek S, Waringer J. 2014. Cryptic diversity of caddisflies in the
- 424 Balkans: The curious case of *Ecclisopteryx* species (Trichoptera: Limnephilidae). *Arthropod*
- 425 *Systematics and Phylogeny* 72(3): 309-329.
- 426 Puillandre N, Lambert A, Brouillet S, Achaz G. 2012. ABGD, Automatic Barcode Gap
- 427 Discovery for primary species delimitation. *Molecular Ecology* 21(8): 1864-77.
- 428 Putchkov AV, Matalin AV. 2003. Subfamily Cicindelinae Latreille, 1802. In: Löbl L,
- 429 Smetana A. (Eds) Catalogue of Palaearctic Coleoptera. V.1. Archeostemata Myxophaga –
- 430 Adephaga. Strenstrup: Apollo Books, 99–118.
- Rogers AR., Harpending H. 1992. Population growth makes waves in the distribution of
- pairwise genetic differences. *Molecular Biology and Evolution* 9: 552–569.

- 433 Serrano J. 2013. New catalogue of the family Carabidae of the Iberian Peninsula
- 434 (Coleoptera). Madrid: Editum Ediciones de la Universidad de Murcia.
- Simon C, Frati F, Beckenbach AT, Crespi B, Liu H, Flook P. 1994. Evolution, weighting, and
- 436 phylogenetic utility of mitochondrial gene sequences and a compilation of conserved
- polymerase chain reaction primers. *Annals of Entomological Society of America* 87: 51–701.
- 438 Slatkin M, Hudson RR. 1991. Pairwise comparisons of mitochondrial DNA sequences in
- stable and exponentially growing populations. *Genetics* 129: 555–562.
- Sota T, Liang H, Enokido Y, Hori M. 2011. Phylogeny and divergence time of island tiger
- beetles of the genus Cylindera (Coleoptera: Cicindelidae) in East Asia. Biological Journal of
- 442 *the Linnean Society* 102: 715-727.
- Stanley DJ, Blanpied C. 1980. Late Quaternary water exchange between the eastern
- Mediterranean and the Black Sea. *Nature* 285 (5766): 537-541.
- Tajima F. 1989. Statistical method for testing the neutral mutation hypothesis by DNA
- 446 polymorphism. Genetics 123: 585–595.
- Tamura K, Stecher G, Peterson D, Filipski A, Kumar S. 2013. MEGA6: Molecular
- Evolutionary Genetics Analysis version 6.0. *Molecular Biology* and Evolution 30: 2725–
- 449 2729.
- 450 Tsuji K, Hori M, Phyu MH, Liang H, Sota T. 2016. Colorful patterns indicate common
- ancestry in diverged tiger beetle taxa: molecular phylogeny, biogeography, and evolution of
- elytral coloration of the genus *Cicindela* subgenus *Sophiodela* and its allies. *Molecular*
- 453 *Phylogenetics and Evolution* 95: 1-10.
- Vogler AF, DeSalle R. 1993. Phyllogeographic patterns in coastal North American tiger
- beetles (Cicindela dorsalis Say) inferred from mitochondrial DNA sequences. Evolution 47
- 456 (4): 1192-1202.

### NOT PEER-REVIEWED

457	Vogler AP, Pearson DL. 1996. A molecular phylogeny of the tiger beetles (Cicindelidae):
458	congruence of mitochondrial and nuclear rDNA data sets. Molecular Phylogenetics and
459	Evolution 6 (3): 321–338.
460	Vogler AP, Knisley CB, Glueck SB, Hill JM, DeSalle R. 1993. Using molecular and
461	ecological data to diagnose endangered populations of the puritan tiger beetle Cicindela
462	puritana. Molecular Ecology 2: 375-383.
463	Werner K. 1991. Cicindelidae Regionis Palaearcticae. Megacephalini: Megacephala,
464	Cicindelini 1: Cicindela – Lophyridia. The Beetles of the World 13. Compiègne: Sciences
465	Nat
466	Woodcock MR, Kippenhan MG, Knisley CB, Foster JA. 2007. Molecular genetics of
467	Cicindela (Cylindera) terricola and elevation of Cicindela lunalonga to species level, with
468	comments on its conservation status. Conservation Genetics 8: 865-877.
469	



**Figure 1**. Distribution and sampling of *Calomera littoralis* in Europe. (A) General distribution of *Calomera littoralis* in Europe shown as red-shaded area. (B) Picture of *Calomera littoralis* beetle. (C) Sampling sites in Balkan Peninsula, Black Sea region and Turkey shown as black dots. Localities coded as in Table 1.

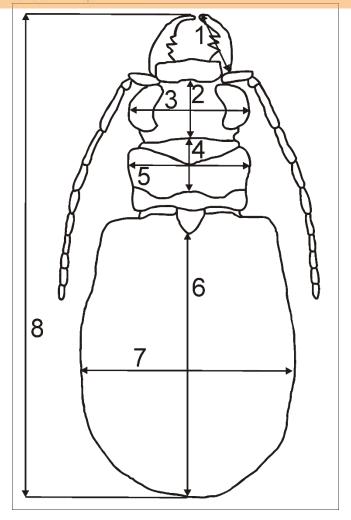
475

474

470

471

472



477

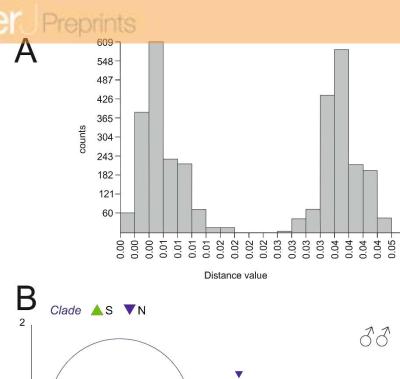
479

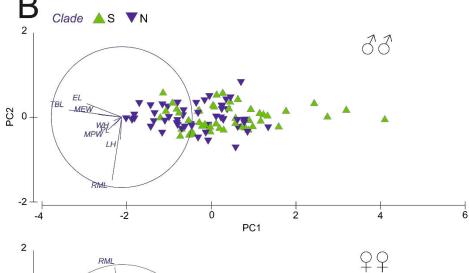
 $\textbf{Figure 2.} \ \ \text{Body parameters measured in } \textit{Calomera littoralis.} \ \ 1-\text{RLM}-\text{right mandible length,}$ 

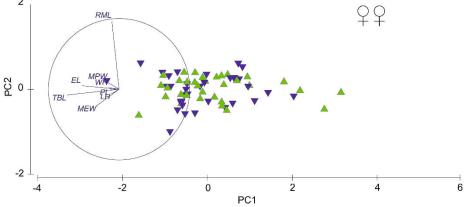
478 2 – WH – width of head, 3 – LH – length of head, 4 – MPW – maximum pronotum width, 5 –

PL – pronotum length, 6 – EL – elytra length, 7 – MEW – maximum elytra width, 8 – TBL –

480 total body length.







**Figure 3.** (A) Automatic Barcode Gap Discovery (ABGD) analysis of *Calomera littoralis* and (B) Results of Principal Component Analysis performed for investigated specimens on main body dimensions. S – southern lineage, N – northern lineage, RLM – right mandible length, WH – width of head, LH – length of head, MPW – maximum pronotum width, PL – pronotum length, EL – elytra length, MEW – maximum elytra width, TBL – total body length. Both in ABGD and PCA analyses 169 specimens from 43 sites from the Mediterranean and the Pontic areas were used.

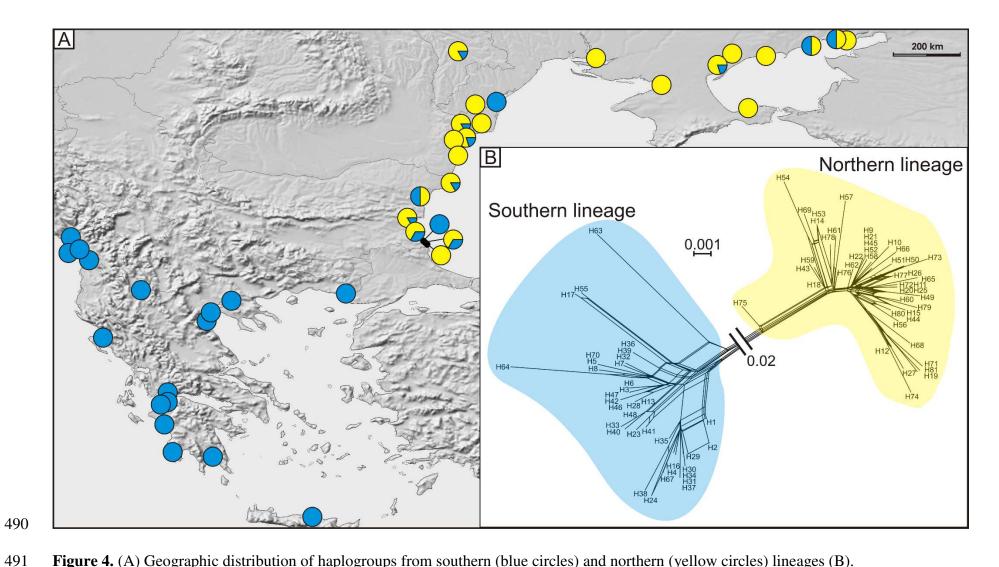
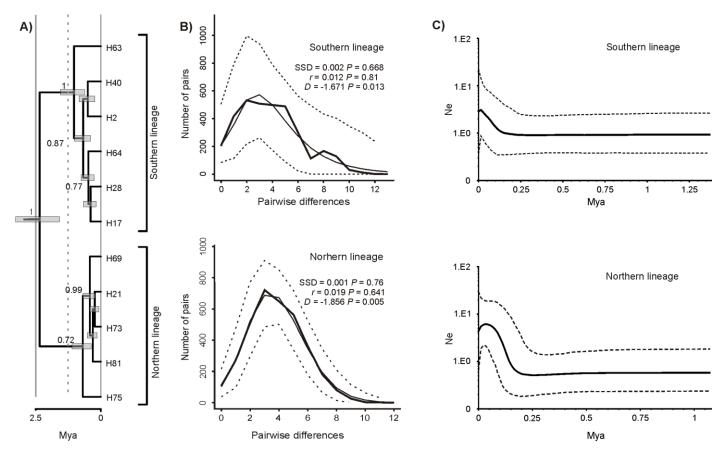


Figure 4. (A) Geographic distribution of haplogroups from southern (blue circles) and northern (yellow circles) lineages (B).



**Figure 5.** Phylogeny and demography of *Calomera littoralis*. (A) Maximum clade credibility chronogram with a strict molecular clock model inferred from COI sequences. The numbers next to the respective node indicate Bayesian posterior probabilities higher than 0.5. (B) Mismatch plots for southern and northern lineage. Thin solid lines indicate expected frequency under model of population demographic expansion, thick solid lines represent observed frequency, and dashed lines indicate 95% confidence intervals for the observed mismatch. SSD, sum of squared deviation;

r, Harpending's raggedness index; D, Tajima's D. (C) Bayesian skyline plots for southern and northern lineages of *Calomera littoralis*. Solid lines indicate the median posterior effective population size through time; dashed lines indicate the 95% highest posterior density interval for each estimate.