

BioMine-DB: A database for metazoan biomineralization proteins

Biomineralization is the process by which living organisms construct hard skeletons creating complex structures that range from specialized tissues such as bone or teeth to ecosystems such as coral reefs. Biominerals are composed of both inorganic minerals and proteins, which give them extra hardness and special attributes. Biomineralization proteins are also known to be associated with multiple bone disorders and are therefore of biomedical importance. Herein we describe BioMine-DB, a biomineralization centric protein database. Availability and implementation: BioMine-DB can be accessed at http://biomine.net, SQL dump, FASTA files and source code are available for download as well at https://github.com/bishoyh/biomineDB "



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11	Introduction
12	Biomineralization is a process in which minerals form inside or outside the cells of a
13	variety of organisms (Lowenstam & Weiner 1989; Simkiss & Wilbur 1989). In animals,
14	these minerals are primarily calcium carbonates and calcium phosphates (Knoll 2003).
15	The majority of biominerals formed in bones, shells, skeletons and spicules are composed
16	of mineral crystals, however all biominerals contain various amounts of other proteins
17	that give these minerals extraordinary properties. The cell orchestrates the mineral
18	formation process through the expression and translocation of proteins that nucleate the
19	crystals either intracellularly or extracellularly. More importantly, the cell has to inhibit
20	mineral formation and crystal growth in unwanted sites (Kawasaki et al. 2009; Marin et
21	al. 1996). Both nucleation and inhibition can be achieved through multiple cellular
22	mechanisms. For example, the cell will produce enzymes that modify proteins by
23	breaking them into smaller peptides (Qin et al. 2004) thus changing their function. The
24	cell is able to tightly regulate the biomineralization process by molecular modification
25	(e.g. adding sugars or other moieties) and regulation of ion transport across membranes
26	(Qin et al. 2004; Saavedra 1994; Sarashina et al. 2006). Such modifications to
27	biomineralization-associated proteins will determine how they interact with other
28	proteins, other cells, and with the biomineral in general. Biominerals are essential to the
29	survival of a broad range of animal taxa because they deliver protection against
30	predation, act as energy storage, provide support and unique optical properties (Addadi et
31	al. 2006). In particular, biomineralization plays a pivotal role in multiple human diseases



32 and other pathological phenomena such as coronary artery calcification (Atlan et al. 33 1997; Collette et al. 2010; Fisher et al. 2001; Lopez et al. 1992; Rousselle & Heymann 34 2002; Salih et al. 1996; Wallin et al. 2001; Westbroek & Marin 1998; Yang et al. 2002). 35 A growing interest in bio-inspired materials has generated a large body of work that uses 36 proteins and other biological scaffolds for in vitro mineralization and synthetic materials 37 (Chiu et al. 2012; Perry et al. 2009). 38 39 The process of biomineralization is ubiquitous throughout the animal tree. Such 40 distribution has generated speculation about the origin of metazoan biomineralization and 41 its evolutionary history. Biomineralization is a complex process that relies on multiple 42 cellular pathways (Knoll 2003; Marin et al. 1996). Many of the studied biomineralization 43 proteins are part of other important processes such as cell adhesion, extracellular matrix organization and immune functions (Bryden et al. 1999; Clendenon et al. 2009). This 44 45 evidence favors the idea that biomineralization independently evolved in multiple phyla 46 using pre-existing pathways in the eumetazoan ancestor. It can also be argued that 47 biomineralization was present in the early eumetazoan ancestor yet various parts of the 48 pathway were lost in several animal lineages. According to fossil evidence and when 49 mapped onto a phylogeny, carbonate skeletons seem to have evolved at least 20 different 50 times in metazoans (Knoll 2003). If biomineralization evolved multiple times, it is 51 relevant to understand which components of the process exactly underwent innovations. 52 Since biomineralization is an active process, it requires 1) targeted localization of 53 calcium and carbonate, 2) an organic matrix as a template for the mineral nucleation, 3) 54 growth, and 3) efficient inhibitors in order to stop undesired calcification or even 55 formation of the mineral (Jackson et al. 2010; Marin et al. 1996). When all these different requirements are taken into account, it seems unlikely that such diverse biochemical 56 57 processes involved in metazoan biomineralization evolved independently more than 20 58 times. A process such as transport is quite conserved across animal lineages and it shows 59 a clear history of gene duplication events (Dean et al. 2001; Saier et al. 2009). Such 60 complexity presents us with a conundrum. While the biomineralization process, with 61 different minerals and methods of calcification and clear evolutionary novelties, is found 62 across multiple animal phyla (Jackson et al. 2007a; Jackson et al. 2006; Jackson et al.



63 2009; Jackson et al. 2007b; Marin et al. 2000; Marin & Luquet 2004; Marin et al. 2008; 64 Marin et al. 1996), it also remains that many parts of biomineralization pathways must be 65 conserved. 66 67 As a first step in tackling such questions in the evolution of animal biomineralization, we 68 have created a database to accumulate, annotate and curate biomineralization proteins and 69 protein-coding sequences. The database aims to serve the community by bridging the gap 70 between the few identified biomineralization proteins, and the unannotated plethora of 71 Expressed Sequence Tags (ESTs), draft genome gene models and next-generation 72 sequencing datasets. We employed various bioinformatics techniques using domain-73 based searches to collect and identify novel biomineralization proteins in metazoans. We 74 hope that due to the increasing surge of sequence information along with broad phylogenetic representation in the public domain, a clearer picture of the evolutionary 75 76 history of biomineralization proteins will emerge, rendering BioMine-DB as a dynamic 77 platform to answer not only fundamental questions in animal evolution but also about the 78 process of biomineralization in particular lineages. 79 80 Methods 81 **Biomineralization proteins list** 82 We carried out a wide primary literature and database survey in order to compile a list of 83 proteins that are functionally implicated biomineralization in animals. Specifically, we 84 included data from scleractinian corals, calcareous sponges, gastropod and bivalve 85 molluses, crustaceans, echinoderms, and vertebrates. Additional sequences were collected 86 from the AMIGO Gene Ontology database (Carbon et al. 2009). The complete 87 biomineralization gene list is accessible through the BioMine-DB web application 88 (http://biomine.net/). These already annotated sequences were used as a seed to search for 89 related biomineralization proteins in undocumented taxa or new sequence databases, and 90 were stored in a pre-computed BLAST search database. After building the list of 91 candidate proteins, each protein could be traced back to an original publication where it 92 was described. 93



94	Pfam domain search and protein homolog identification
95	To further improve the search strategy, using the Pfam database, we scanned for
96	conserved protein domains in the proteins we gathered from primary literature (Finn et al.
97	2008). The identified domains in the already known biomineralization proteins were
98	scanned against 6-frame translations of ESTs and protein sequences from dbEST and nr
99	databases (NCBI) of the taxa Cnidaria, Mollusca, Echinodermata and Vertebrata using
100	the HMMER 3 package (Eddy 2008; Eddy 2011). The tool used for the translation was
101	sixpack from the EMBOSS package (Rice et al. 2000).
102	
103	BLAST Searches
104	In addition to the domain searches, we conducted BLASTp searches for all the proteins in
105	the seed database against nr and against the 6-frame translations of the dbEST for the
106	selected taxa. For filtering the results, we only considered hits that match e-value <
107	0.000001 and bitscore > 150 to be significant.
108	
109	BioMine-DB construction
110	In order, to organize all the data in a searchable platform, we constructed a web
111	application that enables us to search the results and to submit new sequences into the
112	database. BioMine-DB is written in PHP and Perl, and relies on MySQL for relational
113	information Source code for BioMine-DB is under GPL v3 at
114	http://github.com/bishoyh/biomineDB. The MySQL database contains the results of all
115	the HMMER results in addition to the BLAST results and FASTA files of all sequences
116	can be downloaded from the website.
117	
118	Results
119	After assembling the initial list of biomineralization-related proteins, we identified
120	putative homologs of given candidate genes in calcifying lineages (molluscs, cnidarians,
121	arthropods and echinoderms). A protein domain search was initiated on our candidate list
122	of 472 proteins, based on Pfam models using HMMER 3 (Eddy 2008). In the Pfam
123	search 198 domain families were found to be linked to biomineralization. The search



124	results were stored in a relational database linking the detected domains with the
125	taxonomic information, in addition detected orthologs for every particular protein.
126 127	Below we describe two potential scenarios for the use of BioMine-DB by the scientific community.
128	Use case 1:
129	1) A user prepares a list of proteins from a newly sequenced organism.
130	2) The user submits the protein list to BioMine-DB through the web interface.
131	3) BioMine-DB generates potential matching biomineralization proteins in the
132	submitted dataset, together with the publications in which these similar proteins
133	have been described.
134	Use case 2:
135	1) A user is already working with a known biomineralization protein and is doing
136	functional work, i.e. the user is doing whole mount in situ gene expression
137	research in a given organism and finds it hard to explain the observed expression
138	pattern. Thus, the user thinks there could be other proteins involved.
139	2) The user submits his protein to BioMine-DB.
140	3) The user gets back a list of potential biomineralization from
141	
142	Discussion and Conclusion
143	By combining thorough literature scrutiny with similarity searches we were able to
144	construct a large dataset of biomineralization-related proteins. BioMine-DB proved
145	useful in annotating sequence data from non-model organisms involved in the particular
146	process of biomineralization. The ability to always link back to the primary literature
147	provides a unique opportunity to the investigator to directly examine the experimental
148	evidence that deemed a particular protein as biomineralization-associated. We believe
149	this should fast-forward research in non-model systems by knowledge transfer from
150	model species in biomineralization research. By providing a BLAST interface and
151	downloadable versions, we are certain that biomineralization researchers can benefit from
152	BioMine-DB, as there are no other resources that catalog and curate biomineralization
153	proteins.
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