

Towards a Barrier Height Benchmark Set for Biologically Relevant Systems

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ABSTRACT

We have collected computed barrier heights and reaction energies (and associated model structures) for five enzymes from studies published by Himo and co-workers. Using this data, obtained at the B3LYP/6-311+G(2d,2p)[LANL2DZ]/B3LYP/6-31G(d,p) level of theory, we then benchmark PM6, PM7, PM7-TS, and DFTB3 and discuss the influence of system size, bulk solvation, and geometry re-optimization on the error. The mean absolute differences (MADs) observed for these five enzyme model systems are similar to those observed for PM6 and PM7 for smaller systems (10-15 kcal/mol), while DFTB results in a MAD that is significantly lower (6 kcal/mol). The MADs for PMx and DFTB3 are each dominated by large errors for a single system and if the system is disregarded the MADs fall to 4-5 kcal/mol. Overall, results for the condensed phase are neither more or less accurate relative to B3LYP than those in the gas phase. With the exception of PM7-TS, the MAD for small and large structural models are very similar, with a maximum deviation of 3 kcal/mol for PM6. Geometry optimization with PM6 shows that for one system this method predicts a different mechanism compared to B3LYP/6-31G(d,p). For the remaining systems geometry optimization of the large structural model increases the MAD relative to single points, by 2.5 and 1.8 kcal/mol for barriers and reaction energies. For the small structural model the corresponding MADs decrease by 0.4 and 1.2 kcal/mol, respectively. However, despite these small changes, significant changes in the structures are observed for some systems, such as proton transfer and hydrogen bonding rearrangements. The paper represents the first step in the process of creating a benchmark set of barriers computed for systems that are relatively large and representative of enzymatic reactions, a considerable challenge for any one research group but possible through a concerted effort by the community. We end by outlining steps needed to expand and improve the data set and how other researchers can contribute to the process.

Keywords: enzyme mechanism, semiempirical methods, benchmarks

1 INTRODUCTION

Semiempirical electronic structure methods are increasingly parameterized and benchmarked against data obtained by DFT or wavefunction-based calculations rather than experimental data (Stewart, 2007; Scholten, 2003; Gaus et al., 2013). Using calculated data has the advantage that it represents the precise value (usually the electronic energy) that is being parameterized, with little random noise and even coverage of chemical space, including molecules that are difficult to synthesize or perform measurements on. Carefully curated benchmark sets, such as GMTKN30 (Goerigk and Grimme, 2011), are therefore an invaluable resource to the scientific community and heavily used.

For example, Korth and Thiel (2011) used the GMTKN24-hcno dataset (21 subsets of the GMTKN24 data set (Goerigk and Grimme, 2010), an earlier version of GMTKN30) to show that modern semiempirical methods are approaching the accuracy of PBE/TZVP and B3LYP/TZVP calculations. While this is encouraging one concern is whether the results obtained for the small systems that make up these data sets are representative of those one would obtain for large systems. For example, Yilmazer and Korth (2013) performed a benchmark study of hundreds of protein-ligand complexes that included protein atoms within up to 10 Å from the ligand and showed, for example, that the mean absolute difference (MAD) between interaction energies computed using PM6-DH+ and BP86-D2/TZVP was 14 kcal/mol.

In comparison the MADs for the S22 interaction energy subset of GMTKN24 are <2 kcal/mol for both dispersion corrected PM6 and DFT/TZVP calculations (Korth and Thiel, 2011). One likely explanation is that the systems in the S22 subset are too small to exhibit many-body polarization contributions to the binding energy that semi-empirical methods fail to capture. Another, or additional, possibility is that the S22 subset does not include ionic groups, which are quite common in proteins and ligands.

The Yilmazer and Korth (2013) study raises a similar question about whether benchmark results for semiempirical barrier height-predictions on small systems, such as the BH76 and BHPERI subsets of GMTKN24/30, are transferable to barrier height predictions for enzymes. The first step towards answering this question is to create a benchmark set of barriers computed for systems that are relatively large and representative of enzymatic reactions. This is a considerable challenge because, unlike for ligand-protein complexes, there is no large database of corresponding transition state (TS) structures (or even substrate-enzyme structures) to start from. Thus, TS structures must be computed which is time-consuming and hard to automate. There are a significant number of such structures in the literature but many are not computed at a high enough level of theory to serve as benchmarks. Furthermore, TS structures are known to dependent significantly on the level of theory used and it is therefore important that the benchmark set is computed using identical or very similar levels of theory. Creating such a benchmark set is this a considerable challenge for any one research group but can be addressed by a concerted effort by the community. This paper represents the first step in this process.

We have collected barrier heights and reaction energies (and associated structures) for five enzymes from studies published by Himo and co-workers (Chen et al., 2007; Georgieva and Himo, 2010; Hopmann and Himo, 2008; Liao et al., 2011; Sevastik and Himo, 2007), on a GitHub repository (github.com/jensengroup/db-enzymes). Using this data, obtained at the same level of theory, we then benchmark PM6, PM7, PM7-TS, and DFTB3 and discuss the influence of system size, bulk solvation, and geometry re-optimization on the error. We end by outlining steps needed to expand and improve the data set and how other researchers can contribute to the process.

2 COMPUTATIONAL METHODOLOGY

Five systems are investigated: L-aspartate α -decarboxylase (AspDC), 4-oxalocrotonate tautomerase (4-OT), phosphotriesterase (PTE), histone lysine methyltransferase (HKMT), and haloalcohol dehalogenase (HheC). The reaction mechanisms that are investigated are shown schematically in Figure 1. The B3LYP/6-311+G(2d,2p)//B3LYP/6-31G(d,p) (LANL2DZ is used for Zn in PTE) barrier heights and reaction energies are taken from the literature: AspDC (Liao et al., 2011), 4-OT (Sevastik and Himo, 2007), PTE (Chen et al., 2007), HKMT (Georgieva and Himo, 2010), and HheC (Hopmann and Himo, 2008) and the corresponding atomic coordinates are taken from the supplementary information or supplied by Fahmi Himo. 4-OT and PTE have two-step mechanisms resulting in two barrier heights and reaction energies. All energies are taken relative to the reactant state which results in a negative barrier for the second step in the 4-OT mechanism (4-OT-2 in Table 1). The largest model system for each study is used unless noted otherwise and PCM results are for a dielectric constant of 80.

The PM6 (Stewart, 2007), PM7 and PM7-TS (Stewart, 2012) single point calculations are performed using MOPAC2012 while the DFTB3 (Gaus et al., 2011) single point calculations are performed using DFTB+ version 1.2.2 (Aradi et al., 2007) and version 3ob-3-1 of the 3OB parameter set (Gaus et al., 2013, 2014; Lu et al., 2015; Kubillus et al., 2015). PM7-TS calculations are only performed for barrier heights. The PMx/COSMO (Klamt and Schüürmann, 1993) are performed using a dielectric constant of 80. The PM6 geometry optimizations are done using Gaussian09 (Frisch et al., 2014). Figures 2-6 are made with Avogadro (Hanwell et al., 2012).

The B3LYP results include zero-point energy (ZPE) corrections and are therefore directly comparable to the relative enthalpy values predicted by PM6 and PM7. However, ZPE corrections are not included for the DFTB3 calculations and we note that the ZPE can contribute to the difference observed between the DFTB3 and B3LYP results.

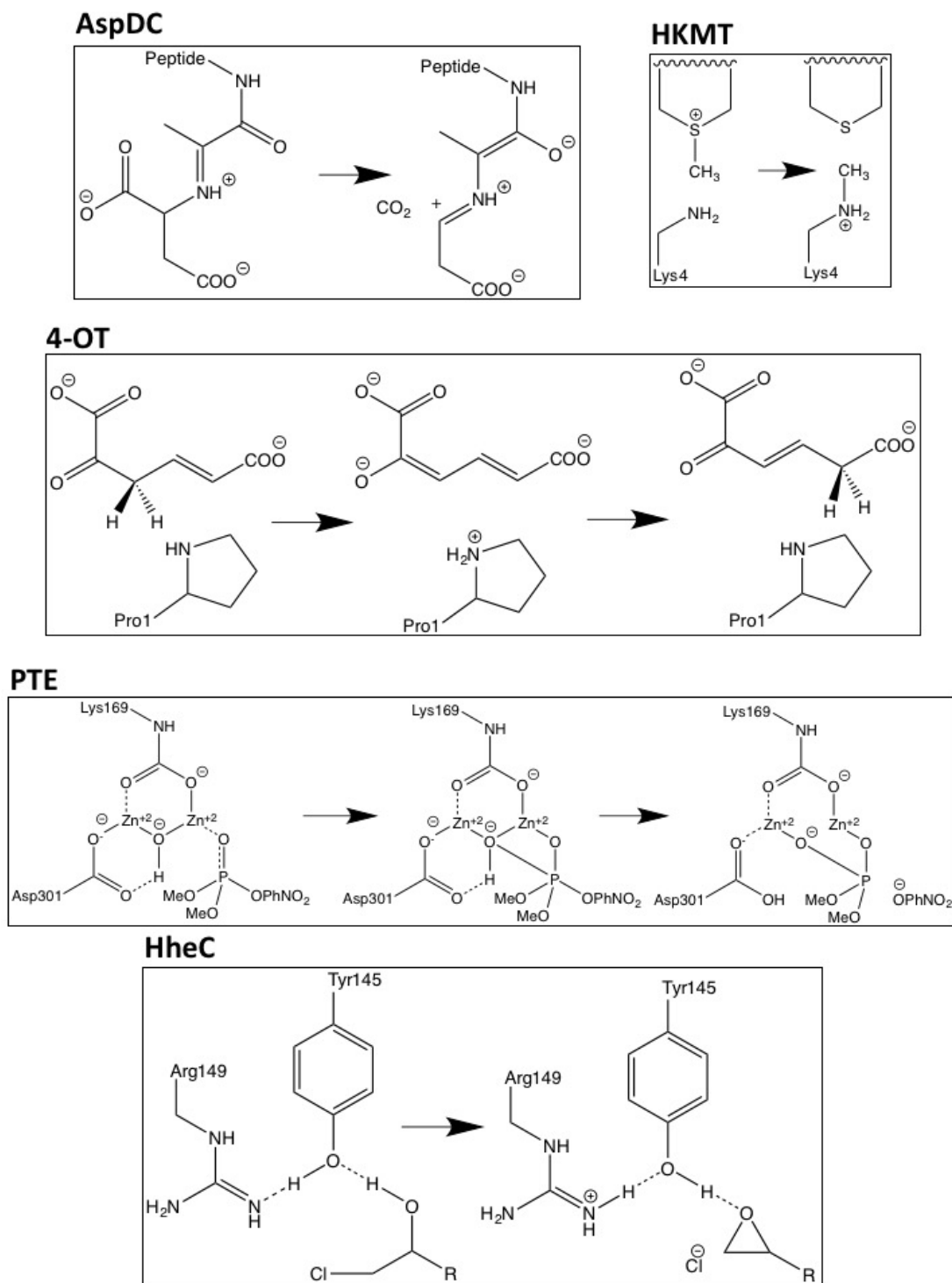


Figure 1. Schematic representations of the reactions mechanisms for the five enzymes studied. In the case of PTE Lys169 is carboxylated and two histidine ligands to each each Zn ion are omitted for clarity.

3 RESULTS AND DISCUSSION

3.1 Gas Phase

Table 1 lists barrier heights and reaction energies computed using PM6, PM7, PM7-TS, and DFTB3 single point energies on B3LYP/6-31G(d,p) geometries. For barrier heights the mean absolute differences (MADs) are 13, 15, 20, and 6 kcal/mol for PM6, PM7, PM7-TS and DFTB3. The 13 kcal/mol MAD for PM6 is comparable to the 10-15 kcal/mol MADs computed for PM6 by Korth and Thiel (2011) and Dral et al. (2016) for various small molecule benchmark sets for barrier heights. For PM6 the accuracy is best for AspDC and 4-OT, for which the models only consist of atoms in the first and second row of the periodic table. For PM6 and PM7 the MADs are dominated by the PTE system (the only system containing a transition metal, Zn) where the errors range from 30 to 41 kcal/mol. Removing these two entries reduces the MADs to 5 and 7 kcal/mol, respectively for PM6 and PM7, which is 2-3 times lower than the MADs computed for PM6 by Korth and Thiel (2011). As we will show below, PM6 does not predict the same mechanism for PTE as B3LYP/6-31G(d,p). For DFTB3 the MAD is dominated by AspDC with an errors of 18 kcal/mol, while the first and second barrier for PTE is reproduced reasonably and very well, respectively. If the AspDC system is neglected the MADs for barrier heights decreases to 3.7 kcal/mol.

The errors computed for reaction energies have MADs of 9, 14, and 6 kcal/mol for PM6, PM7, and DFTB3. The lower MAD for PM6 compared to barrier heights is primarily due to the fact that the 10 error in the second step of the PTE mechanism (i.e. the difference between the product and the reactant) is considerably smaller than the 33 kcal/mol error in the corresponding barrier height. In the all cases there is generally a correlation between errors in the reaction energies and errors in the corresponding barrier heights. Just as for the barriers, the MAD is reduced significantly for PM6 and PM7 (to 4.0 and 7.2 kcal/mol, respectively, if PTE is disregarded). Similarly, the MAD for DFTB3 is reduced to 4.4 kcal/mol if AspDC is disregarded.

In summary, the MADs observed for these five enzyme model systems are similar to those observed for PM6 and PM7 for smaller systems (10-15 kcal/mol), while DFTB results in a MAD that is significantly lower (6 kcal/mol). The MADs for PMx and DFTB3 are dominated by large errors for one system (PTE and AspDC, respectively) and if the system is disregarded the MADs fall to 4-5 kcal/mol.

3.2 Effect of Solvation

The inclusion of bulk solvation effects leads to very modest (≤ 0.5 kcal/mol) decreases in the MADs (Table 1). Some errors decrease, by as much as 3.0 kcal/mol for the reaction energy of AspDC, while others increase, by as much as 3.5 kcal/mol for the second reaction energy of PTE. This is in part due to the fact that the effect of bulk solvation on the B3LYP results is at most 4 kcal/mol for all methods, including B3LYP. The one exception is the second step in the PTE mechanism where solvation increases decreases the reaction energy by 10.7, 12.1, and 14.2 kcal/mol at the B3LYP, PM6, and PM7 level of theory. Thus, overall, results for the condensed phase are neither more or less accurate than those in the gas phase.

3.3 Effect of System Size

In four of the five systems Himo and co-workers computed barrier heights and reaction energies for between two and five systems of different size. In Table 2 the columns marked "Model 0" lists the barrier heights and reaction energies for the smallest system, while the remaining "Model" columns lists the change in energetics on going to the next larger system. For example, at the B3LYP level the barrier height for AspDC is 0.5 kcal/mol larger when computed using Model 2 compared to Model 1. In the case of 4-OT "Model 0-1" and "Model 0-2" refer to the first and second barrier height computed using Model 0 in the case of barrier heights and, in the case of reaction energies, the energy of the intermediate and product relative to the reactant. The last column in Table 2 list the MAD of the energy changes (i.e. excluding Model 0) relative to B3LYP.

The data in Table 2 is summarized in Table 3. Columns two and three list the MAD relative to B3LYP for barrier heights computed using Model 0 and the largest model (excluding PTE), while columns four

Table 1. Barrier heights and reaction energies calculated with a list of semi-empirical methods and compared to B3LYP/6-311+G(2d,2p)[LANL2DZ]//B3LYP/6-31G(d,p) values taken from the literature. For barriers "-1" and "-2" refer to the first and second transition state in the mechanism, while for reaction energies they refer to the intermediate and product, respectively. MAD* values are computed without PTE. All values are in kcal/mol.

	AspDC	4-OT-1	4-OT-2	PTE-1	PTE-2	HKMT	HheC	MAD	MAD*
Gas Phase Barrier Heights									
B3LYP	13.5	6.9	-1.6	11.7	13.3	18.9	18.2		
PM6	11.9	1.4	-1.5	-18.1	-19.4	27.8	28.0	12.6	5.2
PM7	5.6	8.1	5.8	-18.4	-27.3	24.0	31.7	15.1	7.0
PM7-TS	23.1	-10.5	-18.5	-20.4	-18.5	30.5	34.4	19.4	14.3
DFTB3	-4.9	9.2	3.3	18.7	14.1	18.3	24.6	5.8	6.5
Gas Phase Reaction Energies									
B3LYP	9.0	-5.7	-2.9	11.9	-4.6	-9.2	5.5		
PM6	2.0	-8.4	-1.7	-24.2	-14.2	-10.0	13.6	9.4	4.0
PM7	-2.3	-2.3	2.7	-30.1	-26.5	-11.8	18.8	14.3	7.2
DFTB3	-8.9	1.7	-3.0	21.7	-4.9	-7.6	12.7	6.3	6.8
Solution Phase Barrier Heights									
B3LYP	13.3	6.7	-0.8	10.5	11.1	19.1	17.0		
PM6	14.9	0.2	-1.3	-18.8	-21.3	27.7	25.1	12.4	5.1
PM7	8.1	6.7	6.0	-20.0	-32.2	24.2	29.3	14.7	5.9
PM7-TS	24.8	-13.3	-19.2	-19.9	-18.9	29.3	34.0	19.6	15.4
Solution Phase Reaction Energies									
B3LYP	10.0	-5.4	-2.6	8.9	-15.3	-12.4	4.2		
PM6	6.0	-9.6	-2.0	-27.0	-26.3	-12.6	10.3	8.9	3.0
PM7	1.2	-3.3	2.2	-34.9	-40.7	-13.7	16.0	14.0	5.7

and five lists the corresponding values for reaction energies. The last two columns list the average MAD for the changes in barrier heights and reaction energies, respectively, due to increasing system size.

With the exception of PM7-TS, the MAD for the small and large systems are very similar, with a maximum deviation of 3 kcal/mol for PM6. This indicates that the error observed in Table 1 stem primarily from the part of the system where bonds are being broken and formed. This is corroborated by the fact that the MAD for the change in barrier heights and reaction energies are all ≤ 3 kcal/mol (again with the exception of PM7-TS). It is not at all clear why the errors in barrier heights computed using PM7-TS differ significantly more from B3LYP for small structural models, compared to large.

3.4 Effect of Geometry Optimization

Table 4 compares the barrier heights and reaction energies computed using B3LYP and PM6 single points (from Table 1) to the corresponding values computed using PM6 optimized geometries using the largest and smallest structural models. The PTE system is excluded for reasons described below. The data for the large structural models indicate that with the exception of the 4-OT system the effect of optimization on the energetics is relatively minor (< 4 kcal/mol) and does not necessarily improve the agreement with B3LYP.

In the case of 4-OT the agreement with B3LYP is improved considerably for the first barrier, reducing the error from 5.5 to 0.4 kcal/mol, while the agreement is worsened considerably for the second barrier (error increased from 0.1 to 18.9 kcal/mol) and the intermediate (error increased from 2.7 to 11.3 kcal/mol). Corresponding calculations using the smaller structural model of 4-OTA (Sevastik and Himo, 2007) leads to < 2 kcal/mol changes in the barrier heights due to geometry optimization, with the exception of the intermediate, whose stability is increased by 5.9 kcal/mol due to a proton transfer from Arg11 to the

Table 2. Barrier heights and reaction energies calculated with a list of semi-empirical methods and compared to B3LYP/6-311+G(2d,2p)[LANL2DZ]//B3LYP/6-31G(d,p) values taken from the literature. The column labeled "Model 0" is the energetics computed using the smallest structural model, while the remaining columns represent the change n going to the next-largest model. For ApsDC 4.1 and 4.2 refer to two structural models of roughly equal size and the change of going to model 5 is computed relative to Model 4.2. For barriers of 4-TA "-1" and "-2" refer to the first and second transition state in the mechanism, while for reaction energies they refer to the intermediate and product, respectively. All values are in kcal/mol.

	AspDC Barriers							MAD
	Model 0	Model 1	Model 2	Model 3	Model 4.1	Model 4.2	Model 5	
B3LYP	0.1	8.2	0.5	0.2	4.9	4	0.5	
PM6	-1.5	5.2	1.9	1.1	2	2.5	2.7	2.0
PM7	-4.5	4.1	3.2	-1.2	8.4	5.7	-1.7	2.6
PM7-TS	4.5	17.6	3.4	-2.6	6	-2.9	3.1	4.3
DFTB3	-6.8	-4.8	4.7	1.4	-1.3	1.6	-1.0	4.7
AspDC Reaction energies								
B3LYP	-9.5	9.8	-0.9	-1.4	9.1	3.4	4.8	
PM6	-15.5	9.2	-0.5	1.6	6.4	1.8	5.4	1.5
PM7	-15.9	10.8	2.1	-3.8	11.7	3.2	1.3	2.1
DFTB3	-18.8	1.0	0.3	5.0	2.6	1.2	2.4	4.6
4-OT Barriers								
	Model 0-1	Model 0-2	Model 1-1	Model 1-2				
B3LYP	12.8	7	-5.9	-8.6				
PM6	5.4	0.9	-4	-2.4				
PM7	10.4	8.6	-2.3	-2.8				
PM7-TS	-16.8	-19.9	6.3	1.4				
DFTB3	18.3	12.6	-9.1	-9.3				
4-OT Reaction Energies								
B3LYP	9.8	-3.7	-15.5	0.8				
PM6	1.3	-3.6	-9.7	1.9				
PM7	5.4	-3.1	-7.7	5.8				
DFTB3	17.1	-5.5	-15.3	2.5				
HKMT Barriers								
	Model 0	Model 1	Model 2	Model 3				
B3LYP	18.8	2.9	-6.3	3.5				
PM6	29.2	1.8	-5.2	2				
PM7	28.8	0.9	-7.2	1.5				
PM7-TS	39.6	-0.4	-10.3	1.6				
DFTB3	16.3	2.7	-4.7	4.0				
HKMT Reaction Energies								
B3LYP	-2.9	3.4	-17.2	7.5				
PM6	-5.5	5.5	-14.1	4.1				
PM7	-2.2	4.5	-17.3	3.2				
DFTB3	-1.2	5.5	-15.2	3.3				
HheC Barriers								
	Model 0	Model 1	Model 2					
B3LYP	23.0	-5.1	0.3					
PM6	37.4	-6.5	-2.9					
PM7	42.8	-8.2	-2.9					
PM7-TS	49.6	-6.2	-9.0					
DFTB3	30.6	-7.3	1.3					
HheC Reaction Energies								
B3LYP	17.5	-3.4	-8.6					
PM6	30.8	-3.7	-13.5					
PM7	34.3	-6.6	-8.9					
DFTB3	24.8	-5.4	-6.7					

Table 3. "MAD TS Small" refers to the MAD from B3LYP in barrier heights computed using the small structural models listed in Table 2 and similarly for the reaction energies ("MAD RxnE Small"). The values labeled "Big" are the corresponding MADs computed for the large systems taken from Table 1. The columns marked Δ and the MADs for the changes in barrier heights and reaction energies listed in Table 2. All values are in kcal/mol.

	MAD TS Small	MAD TS Big	MAD RxnE Small	MAD Rxn Big	Δ Barrier	Δ RxnE
PM6	8.0	5.2	6.1	4.0	2.4	2.6
PM7	7.7	7.0	5.8	7.2	3.0	3.0
PM7-TS	21.7	14.3			5.9	
DFTB3	5.6	6.5	5.5	6.8	2.3	2.6

substrate (Figure 2). Thus, the change in energetics upon geometry optimization observed for 4-OT is most likely due to different interactions with ligands not immediately adjacent to the substrate.

Using the small structural models, the effect of optimization on the PM6 barrier heights is also relatively modest (< 3 kcal/mol), with the exception of HheC, where the barrier drops by 8.6 kcal/mol. The drop in barrier height is likely due to a shift in position of Arg149 upon PM6 optimization so that it is now hydrogen bonded to the Ser132 oxygen and the oxygen on the substrate, rather than the oxygen of Tyr145 as in the B3LYP optimized structure (Figure 3).

For reaction energies substantial decreases of 10.9 and 15.2 kcal/mol are observed for HKMT and HheC, respectively. In the case of HheC the change is due to a rather large structural rearrangement in which a proton from Arg149 is transferred to the Cl^- (Figure 4) while for HKMT the change appears to be due to a rather subtle change in the interaction between the S atom and the methyl group in the methylamine (Figure 5).

In the case of PTE the bonding in the reactant completely changes upon geometry optimization using PM6 (Figure 6a and b). The Zn-phosphate bond is broken and a Zn-Zn bond is formed instead. Furthermore, a minimum is found on the PM6 potential energy surface (Figure 6d) that is very similar to the second transition state found with B3LYP (Figure 6c) except that the proton on the Zn-bridging OH group has not been transferred to Asp301 and the P-O bond to the nitrophenyl group is shorter by 0.40 Å in the PM6 optimized structure. Thus, taking B3LYP/6-31G(d,p) as the standard, PM6 fails to predict the correct mechanism for PTE, which also explains the very large errors observed for the PM6 single point calculations in Table 1. The PTE mechanism has been studied using AM1 (Wong and Gao, 2007), PM3 (Zhang et al., 2009), and AM1/d-based QM/MM methods (López-Canut et al., 2012) and that these studies have suggested other reaction mechanisms. We emphasize that we not explored the mechanism of PTE using PM6 but simply compared the structures resulting from the PM6 geometry optimizations initiated from the B3LYP/6-31G(d,p) geometries obtained by Chen et al. (2007).

In summary, geometry optimization with PM6 shows that for PTE this method predicts a different mechanism compared to B3LYP/6-31G(d,p), consistent with the fact that PM6//B3LYP/6-31G(d,p) energetics differ very significantly from the reference values as discussed above. For the remaining systems geometry optimization of the large structural model increases the MAD relative to single points, by 2.5 and 1.8 kcal/mol for barriers and reaction energies. For the small structural model the corresponding MADs decrease by 0.4 and 1.2 kcal/mol, respectively. However, despite these small changes significant changes in the structure, such as proton transfer and hydrogen bonding rearrangements is observed for some systems.

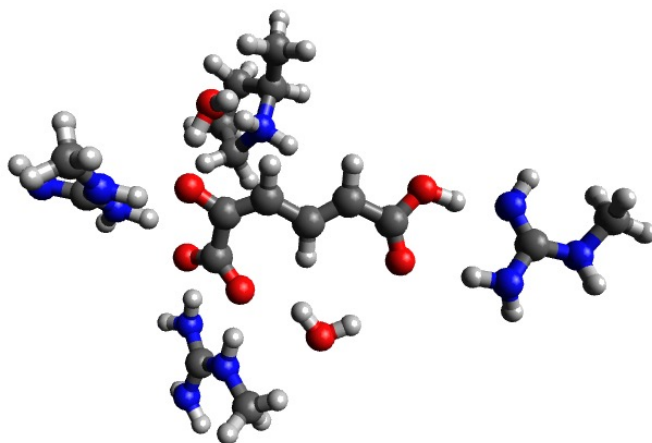


Figure 2. PM6 optimized small structural model of the intermediate in the 4-OT reaction mechanism

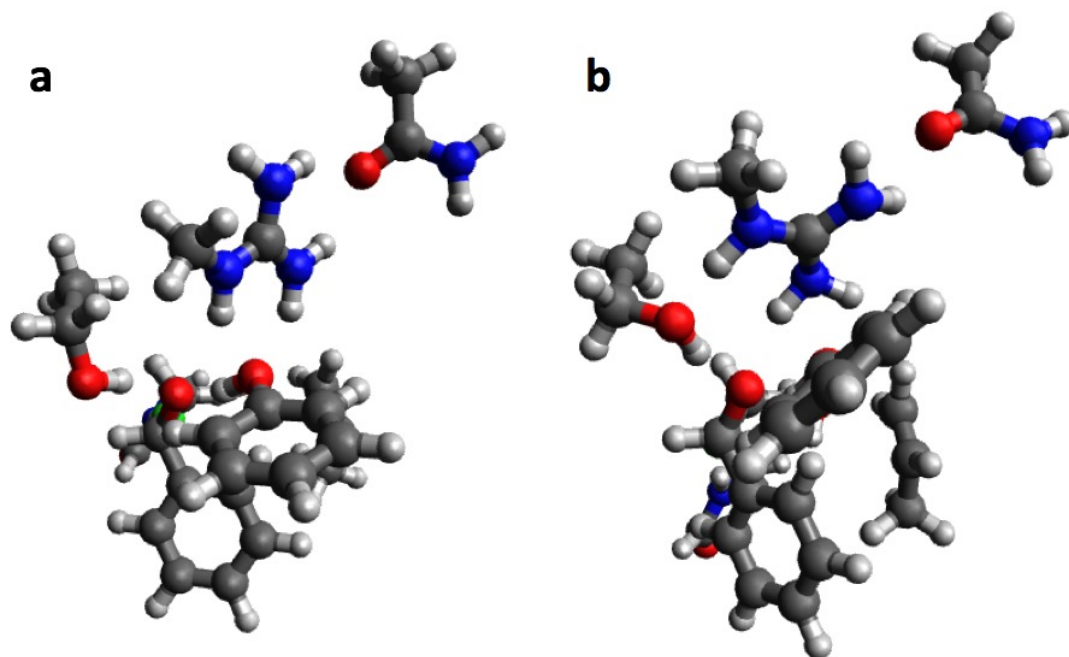


Figure 3. B3LYP/6-31G(d,p) (a) and PM6 (b) optimized small structural model of the transition state in the HheC reaction mechanism

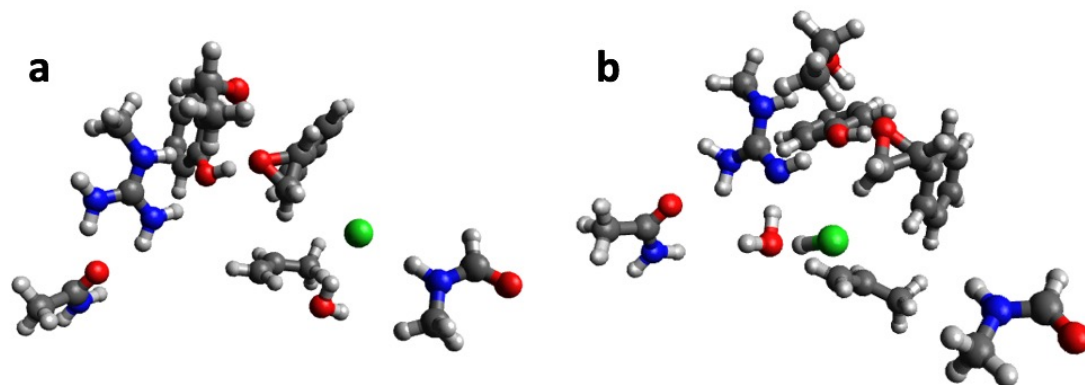


Figure 4. B3LYP/6-31G(d,p) (a) and PM6 (b) optimized small structural model of the product in the HheC reaction mechanism

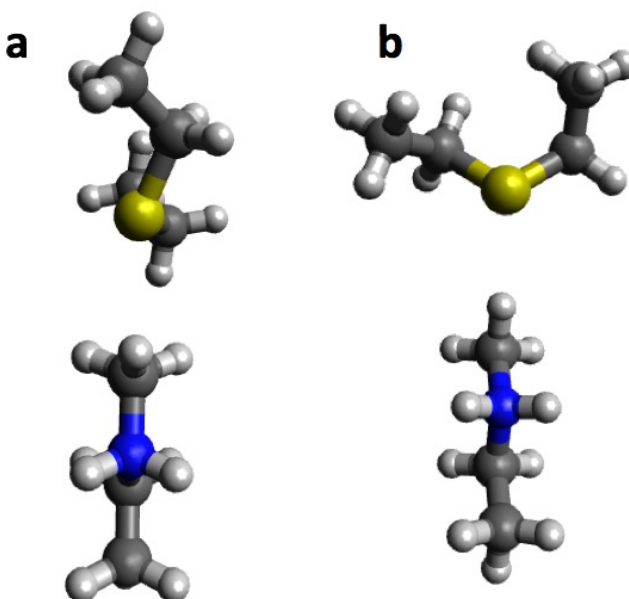


Figure 5. B3LYP/6-31G(d,p) (a) and PM6 (b) optimized small structural model of the product in the HKMT reaction mechanism

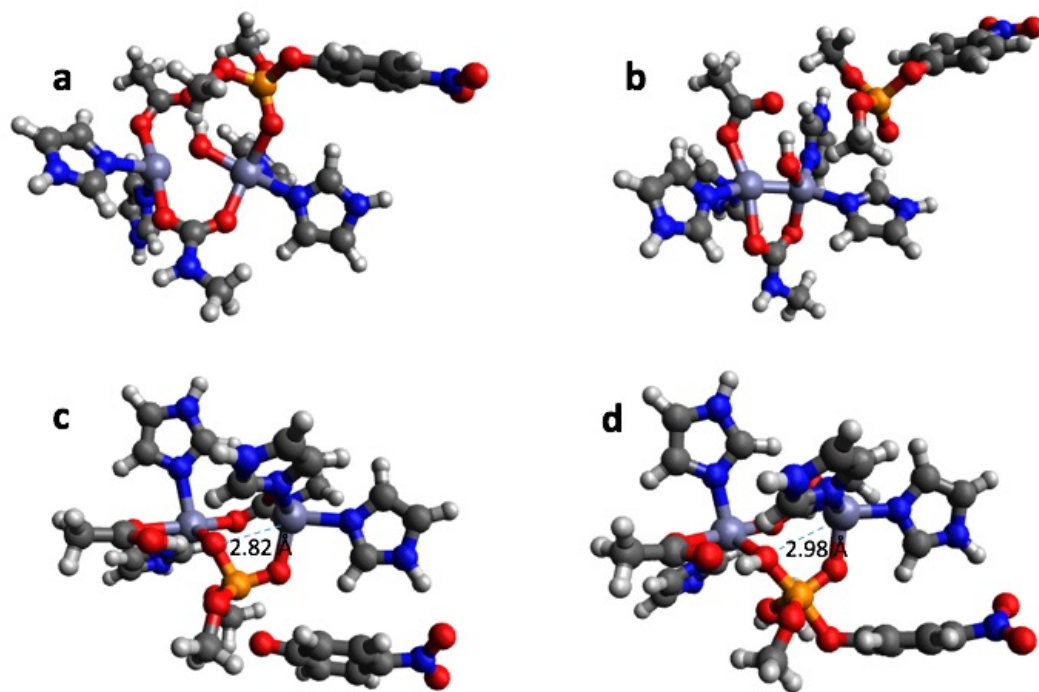


Figure 6. B3LYP/6-31G(d,p) and PM6 optimized structure of the reactant (a and b, respectively in the PTE mechanism. B3LYP/6-31G(d,p) structure of the second transition state (c) and PM6 optimized structure of a minimum that is very similar to this transition state (d).

Table 4. Gas phase barrier heights and reaction energies computed using B3LYP/6-311+G(2d,2p)//B3LYP/6-31G(d,p), PM6//B3LYP/6-31G(d,p), and PM6//PM6 ("PM6 opt") using the largest and smallest structural models

	AspDC	4-OT-1	4-OT-2	HKMT	HheC	MAD
Large Structural Models						
Barrier Heights						
B3LYP	13.5	6.9	-1.6	18.9	18.2	
PM6	11.9	1.4	-1.5	27.8	28.0	5.2
PM6 opt	15.8	7.3	17.3	27.5	26.4	7.7
Reaction Energies						
B3LYP	9.0	-5.7	-2.9	-9.2	5.5	
PM6	2.0	-8.4	-1.7	-10.0	13.6	4.0
PM6 opt	3.5	5.6	-3.0	-12.7	14.2	5.8
Small Structural Models						
Barrier Heights						
B3LYP	0.1	12.8	7.0	18.8	23.0	
PM6	-1.5	5.4	0.9	29.2	37.4	8.0
PM6 opt	1.6	3.7	-1.1	32.6	28.8	7.6
Reaction Energies						
B3LYP	-9.5	9.8	-3.7	-2.9	17.5	
PM6	-15.5	1.3	-3.6	-5.5	30.8	6.1
PM6 opt	-10.7	-4.6	-3.2	5.4	15.6	5.3

4 OUTLOOK

While our study identifies cases where semiempirical methods give results that differ significantly from the DFT and may require further attention, it is clear that five systems is not sufficient for a general and statistically significant assessment of the accuracy of a computational method. We plan to add more systems from the literature to the data set and invite other researchers to do the same. This can easily be done by making a free account on github.com and contributing to the project at github.com/jensengroup/enzyme-db.

While B3LYP/6-311+G(2d,2p)[LANL2DZ]//B3LYP/6-31G(d,p) may be an adequate level of theory to identify deficiencies in semiempirical methods it is unlikely to be accurate enough to parameterize against. In the case of intermolecular interactions the "gold standard" is CCSD(T)/CBS//MP2/TZVP computed using extrapolation (Jurecka et al., 2006; Řezáč and Hobza, 2013). This level of theory may be impractical for these size systems for the foreseeable future, but could be approximated by extrapolating from smaller systems using an ONIOM-like approach (Chung et al., 2015) or approaches like DLPNO-CCSD(T) (Liakos et al., 2015). Again we encourage researchers interested in developing or testing such methods to use the coordinates in the data set and deposit the barriers and reaction energies.

Acknowledgments

We thank Fahmi Himo for providing coordinates for the HKMT and HheC structural models and for providing useful comments on the manuscript.

REFERENCES

- Aradi, B., Hourahine, B., and Frauenheim, T. (2007). DFTB+ a Sparse Matrix-Based Implementation of the DFTB Method †. *J. Phys. Chem. A*, 111(26):5678–5684.
- Chen, S.-L., Fang, W.-H., and Himo, F. (2007). Theoretical Study of the Phosphotriesterase Reaction Mechanism. *The Journal of Physical Chemistry B*, 111(6):1253–1255.
- Chung, L. W., Sameera, W. M. C., Ranzani, R., Page, A. J., Hatanaka, M., Petrova, G. P., Harris, T. V., Li, X., Ke, Z., Liu, F., Li, H.-B., Ding, L., and Morokuma, K. (2015). The ONIOM Method and Its Applications. *Chemical Reviews*, 115(12):5678–5796.
- Dral, P. O., Wu, X., Spörkel, L., Kosłowski, A., and Thiel, W. (2016). Semiempirical quantum-chemical orthogonalization-corrected methods: Benchmarks for ground-state properties. *J. Chem. Theory Comput.*
- Frisch, M. J., Trucks, G. W., Schlegel, H. B., Scuseria, G. E., Robb, M. A., Cheeseman, J. R., Scalmani, G., Barone, V., Mennucci, B., Petersson, G. A., Nakatsuji, H., Caricato, M., Li, X., Hratchian, H. P., Izmaylov, A. F., Bloino, J., Zheng, G., Sonnenberg, J. L., Hada, M., Ehara, M., Toyota, K., Fukuda, R., Hasegawa, J., Ishida, M., Nakajima, T., Honda, Y., Kitao, O., Nakai, H., Vreven, T., Montgomery, Jr., J. A., Peralta, J. E., Ogliaro, F., Bearpark, M., Heyd, J. J., Brothers, E., Kudin, K. N., Staroverov, V. N., Kobayashi, R., Normand, J., Raghavachari, K., Rendell, A., Burant, J. C., Iyengar, S. S., Tomasi, J., Cossi, M., Rega, N., Millam, J. M., Klene, M., Knox, J. E., Cross, J. B., Bakken, V., Adamo, C., Jaramillo, J., Gomperts, R., Stratmann, R. E., Yazyev, O., Austin, A. J., Cammi, R., Pomelli, C., Ochterski, J. W., Martin, R. L., Morokuma, K., Zakrzewski, V. G., Voth, G. A., Salvador, P., Dannenberg, J. J., Dapprich, S., Daniels, A. D., Farkas, Ö., Foresman, J. B., Ortiz, J. V., Cioslowski, J., and Fox, D. J. (2014). Gaussian~09 Revision D.01. Gaussian Inc. Wallingford CT 2009.
- Gaus, M., Cui, Q., and Elstner, M. (2011). DFTB3: Extension of the Self-Consistent-Charge Density-Functional Tight-Binding Method (SCC-DFTB). *J. Chem. Theory Comput.*, 7(4):931–948.
- Gaus, M., Goez, A., and Elstner, M. (2013). Parametrization and Benchmark of DFTB3 for Organic Molecules. *J. Chem. Theory Comput.*, 9(1):338–354.
- Gaus, M., Lu, X., Elstner, M., and Cui, Q. (2014). Parameterization of DFTB3/3OB for Sulfur and Phosphorus for Chemical and Biological Applications. *J. Chem. Theory Comput.*, 10(4):1518–1537.
- Georgieva, P. and Himo, F. (2010). Quantum chemical modeling of enzymatic reactions: The case of histone lysine methyltransferase. *J. Comput. Chem.*
- Goerigk, L. and Grimme, S. (2010). A General Database for Main Group Thermochemistry Kinetics, and Noncovalent Interactions - Assessment of Common and Reparameterized (meta-)GGA Density Functionals. *J. Chem. Theory Comput.*, 6(1):107–126.

- Goerigk, L. and Grimme, S. (2011). Efficient and Accurate Double-Hybrid-Meta-GGA Density Functionals—Evaluation with the Extended GMTKN30 Database for General Main Group Thermochemistry Kinetics, and Noncovalent Interactions. *J. Chem. Theory Comput.*, 7(2):291–309.
- Hanwell, M. D., Curtis, D. E., Lonie, D. C., Vandermeersch, T., Zurek, E., and Hutchison, G. R. (2012). Avogadro: An advanced semantic chemical editor, visualization, and analysis platform. *J. Cheminformatics*, 4(1):17.
- Hopmann, K. H. and Himo, F. (2008). Quantum Chemical Modeling of the Dehalogenation Reaction of Haloalcohol Dehalogenase. *J. Chem. Theory Comput.*, 4(7):1129–1137.
- Jurecka, P., Spöner, J., Cerny, J., and Hobza, P. (2006). Benchmark database of accurate (MP2 and CCSD(T) complete basis set limit) interaction energies of small model complexes DNA base pairs, and amino acid pairs. *Phys. Chem. Chem. Phys.*, 8(17):1985.
- Klamt, A. and Schüürmann, G. (1993). COSMO: a new approach to dielectric screening in solvents with explicit expressions for the screening energy and its gradient. *J. Chem. Soc. Perkin Trans.*, 2(5):799.
- Korth, M. and Thiel, W. (2011). Benchmarking Semiempirical Methods for Thermochemistry Kinetics, and Noncovalent Interactions: OMx Methods Are Almost As Accurate and Robust As DFT-GGA Methods for Organic Molecules. *J. Chem. Theory Comput.*, 7(9):2929–2936.
- Kubillus, M., Kubař, T., Gaus, M., Řezáč, J., and Elstner, M. (2015). Parameterization of the DFTB3 Method for Br, Ca, Cl, F, I, K, and Na in Organic and Biological Systems. *J. Chem. Theory Comput.*, 11(1):332–342.
- Liakos, D. G., Sparta, M., Kesharwani, M. K., Martin, J. M. L., and Neese, F. (2015). Exploring the Accuracy Limits of Local Pair Natural Orbital Coupled-Cluster Theory. *J. Chem. Theory Comput.*, 11(4):1525–1539.
- Liao, R.-Z., Yu, J.-G., and Himo, F. (2011). Quantum Chemical Modeling of Enzymatic Reactions: The Case of Decarboxylation. *J. Chem. Theory Comput.*, 7(5):1494–1501.
- López-Canut, V., Ruiz-Pernía, J. J., Castillo, R., Moliner, V., and Tuñón, I. (2012). Hydrolysis of phosphotriesters: A theoretical analysis of the enzymatic and solution mechanisms. *Chemistry - A European Journal*, 18(31):9612–9621.
- Lu, X., Gaus, M., Elstner, M., and Cui, Q. (2015). Parametrization of DFTB3/3OB for Magnesium and Zinc for Chemical and Biological Applications. *The Journal of Physical Chemistry B*, 119(3):1062–1082.
- Řezáč, J. and Hobza, P. (2013). Describing Noncovalent Interactions beyond the Common Approximations: How Accurate Is the “Gold Standard,” CCSD(T) at the Complete Basis Set Limit? *J. Chem. Theory Comput.*, 9(5):2151–2155.
- Scholten, M. (2003). *Semiempirische Verfahren mit Orthogonalisierungskorrekturen: Die OM3 Methode*. PhD thesis, PhD thesis, Heinrich-Heine-Universität, Düsseldorf.
- Sevastik, R. and Himo, F. (2007). Quantum chemical modeling of enzymatic reactions: The case of 4-oxalocrotonate tautomerase. *Bioorganic Chemistry*, 35(6):444–457.
- Stewart, J. J. P. (2007). Optimization of parameters for semiempirical methods V: Modification of NDDO approximations and application to 70 elements. *Journal of Molecular Modeling*, 13(12):1173–1213.
- Stewart, J. J. P. (2012). Optimization of parameters for semiempirical methods VI: more modifications to the NDDO approximations and re-optimization of parameters. *Journal of Molecular Modeling*, 19(1):1–32.
- Wong, K.-Y. and Gao, J. (2007). The reaction mechanism of paraoxon hydrolysis by phosphotriesterase from combined QM/MM simulations †. *Biochemistry*, 46(46):13352–13369.
- Yilmazer, N. D. and Korth, M. (2013). Comparison of Molecular Mechanics Semi-Empirical Quantum Mechanical, and Density Functional Theory Methods for Scoring Protein–Ligand Interactions. *The Journal of Physical Chemistry B*, 117(27):8075–8084.
- Zhang, X., Wu, R., Song, L., Lin, Y., Lin, M., Cao, Z., Wu, W., and Mo, Y. (2009). Molecular dynamics simulations of the detoxification of paraoxon catalyzed by phosphotriesterase. *Journal of computational chemistry*, 30(15):2388–2401.