A peer-reviewed version of this preprint was published in PeerJ on 19 April 2016.

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O'Neill HL, Cassidy AP, Harris OB, Cassidy JW. 2016. BMP2/BMPR1A is linked to tumour progression in dedifferentiated liposarcomas. PeerJ 4:e1957 https://doi.org/10.7717/peerj.1957



- 1 BMPR1A signalling is linked to tumour progression in
- 2 dedifferentiated liposarcomas: lessons for rhBMP2 use in
- 3 spinal surgery

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Introduction

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The most commonly performed surgery to treat chronic lumbar back pain is vertebral fusion (arthrodesis) (Deyo, Nachemson, & Mirza, 2004). The procedure involves inter body fusion of spinal vertebrae; removing the degenerated disc and replacing it, most frequently with an autologous bone graft. Iliac crest harvest has been considered the 'gold standard' for successful arthrodesis of the lumbar spine, however, remains associated with multiple co-morbidities (Vaz et al., 2010). This has given rise to a number of alternatives: autologous bone from other donor sites, allogeneic bone, ceramics, and, in 2002, the FDA approved recombinant human bone morphogenic protein 2 (rhBMP2) as a bone graft substitute for single level anterior lumbar interbody fusion (Fu et al., 2013).

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BMP2, a member of the Transforming Growth Factor beta (TGF-β) superfamily of proteins, was first described for its essential role in the development of bone and cartilage (Chen, Zhao, & Mundy, 2004). Recent studies have highlighted its role as a multipurpose cytokine, involved in embryonic development (Munoz-Sanjuan & Brivanlou, 2002), migration (J. Song et al., 2014), angiogenesis (X. Song et al., 2011) and hedgehog signalling (Laforest et al., 1998). BMP2 signalling is essential for initiation of the endogenous bone repair response. Despite intact signalling cascades capable of directing osteogenesis, mice lacking BMP2 develop spontaneous fractures, which do not resolve without intervention (Tsuji et al., 2006). By combining osteoconductive hydrogel biomaterials with osteoinductive growth factors such as BMP2, bone repair can be dramatically improved (Ben-David et al., 2013). However, in recent years exogenous supply of high-dose recombinant human BMP2 (rhBMP2) has been linked to an increased incidence of new cancer events (Simmonds et al., 2013; Thawani et al., 2010), in addition to issues with ectopic bone formation. Thus, the North American Spine Society now stipulates that rhBMP2 be used solely in patients for whom other adjuncts are not appropriate or are not likely to lead to successful fusion (TCGA, 2013).



In order to initiate downstream SMAD signalling and the formation of the transcriptional complex, BMP2 binds type I and type II serine/threonine kinases at the cell surface. The current model suggests that type II receptors (BMPR2) can bind ligands in the absence of type I receptors but that type I receptors (BMPR1A and BMPR1B) are required for downstream signalling; likewise type I receptors require type II receptors for ligand binding (Bonor et al., 2012). Type I BMP receptors (BMPR1A and 1B) have well defined overlapping functions (Yoon et al., 2005) but may not be functionally redundant in all aspects of their biology. In the presence of ligand, a heterodimer forms between type I and type II receptors resulting in phosphorylation of BMPR1A/1B by BMPR2 and subsequent signal transduction through SMAD1/5/8 and SMAD4 (11).

Despite the clinical findings that rhBMP2 leads to an increase in new cancer incidents, there remains much debate over the role of BMP2 in tumour progression (Gitelis, Wilkins, & Yasko, 2008). Moreover despite the wide varieties of cancer types associated with rhBMP2 use, no systemic activity is expected due to its short *in vivo* half-life (FDA, 2002; Kisiel et al., 2013). Thus, we sought to profile the role of BMP2 tumour progression of the tissues surrounding the surgical site and therefor most likely to be affected by exogenous BMP2. Specifically we investigated the role of BMPR1A-biased BMP2 signalling in soft tissue sarcomas and found a significant association with reduced disease free survival. Through differential expression analysis and gene set enrichment, we show that this increase in disease recurrence is linked to increased extracellular remodelling in dedifferentiated liposarcomas. This study emphasises that increased research into the effects of locally administered rhBMP2 is needed, along with continued follow-up and screening of patients administered rhBMP2 during spinal surgeries.



Results

Differential Type 1 BMP Receptor expression in Normal Tissues and

Sarcomas

Type I BMP receptors (BMPR1A and 1B) have well defined overlapping functions (Yoon et al., 2005) but may not be functionally redundant in all aspects of their biology. To begin to test this hypothesis, we first profiled expression of the type I and type II BMP receptors across normal tissues (Supplementary Figure 1). Whilst BMPR1A and BMPR1B showed similar expression across some tissues (for example heart, lung, skin, colon etc.) we observed differential expression in numerous tissues. For example BMPR1B is highly expressed in all regions of the brain (cerebellum, hippocampus, cerebral cortex and lateral ventricle) whereas BMPR1A has only moderate expression in the cerebral cortex, suggesting that BMPR1B signalling predominates in the central nervous system.

We next profiled expression patterns across cancer cell lines (Supplementary Figure 2 and Figure 1A). In the three cell lines profiled by immunohistochemistry (IHC), BMPR1A exhibited both nuclear and cytoplasmic localisation, which matched BMPR2 expression patterns (Supplementary Figure 2). In contrast BMPR1B appeared predominantly localised to the nucleus, but not nucleolus. The osteosarcoma cell line U-2 OS showed highest expression of BMPR1A by IHC (Figure 1A), which was echoed in RNAseq data from three sarcoma cell lines (Figure 1B), where BMPR1A expression far exceeded that of BMPR1B. These data suggest that BMPR1A and BMPR1B may not be functionally redundant in all aspects of their biology.

Since rhBMP2 has a short *in vivo* half-life and is expected to influence relatively few tissues after administration (FDA, 2002), we reasoned that the soft tissues surrounding the surgical site may be the most exposed to its actions. Hence, we elected to focus our analysis on BMP2 pathway activation in the soft tissue sarcoma TCGA dataset. The observations from Figure 1A and 1B (together with Supplementary Figure 1 and 2) that BMPR1A and



119	BMPR1B have differential expression patterns lead us to focus our analysis further on the
120	effects of BMPR1A versus BMPR1B biased BMP2 signalling within this group of patients.
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122	By stratifying the 263 samples represented in the TCGA sarcoma dataset based on the ratio
123	of BMPR1A to BMPR1B mRNA, we were able to define two clear groups (Figure 1C).
124	Although most patients expressed relatively similar levels of the two type I receptors there
125	were clear groups who favoured either BMPR1A (red) or BMPR1B (blue) but who did not
126	show differential expression of BMP2 (Figure 1D). Based on the observed patterns of
127	differential expression in normal tissues (Supplementary Figure 1) and cell lines
128	(Supplementary Figure 2), and the conclusion that BMPR1A and BMPR1B may not be
129	entirely functionally redundant, we reason that these groups may show distinct outcomes
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BMP2-BMPR1A signalling is associated with extracellular matrix remodelling

To test the hypothesis that patients favouring BMPR1A would have differential biological responses to high levels of endogenous or exogenous BMP2 to patients favouring BMPR1B we refined our patient groups from Figure 1C. Referred to as BMPR1A-biased or BMPR1B-biased signalling, we defined groups of patients in the upper quartile of the BMPR1A/BMPR1B ratio and in the upper quartile of BMP2 expression together with their reciprocals (high 1B/1A with high BMP2).

We first asked whether BMPR1A-biased signalling lead to differential gene expression to BMPR1B-biased signalling. In total we found 922 differentially expressed genes out of a possible 20,533 annotated transcripts, using a false discovery rate (FDR) of 0.05 (corrected p>0.05). Of these, 252 were up-regulated and 669 down-regulated. Figure 2A shows a volcano plot relating fold change (in patients with BMPR1A-biased versus BMPR1B-biased BMP2 signalling) to p value (multiple two-tailed t-tests with Bonferroni $post\ hoc$). The insert panel in Figure 2A highlights a selection of the most differentially expressed genes; the full list is available as Supplementary Table 2. Amongst the genes negatively associated with BMPR1A-biased signalling are WNT7B which has a role in osteoblast differentiation and mineralised nodule formation (Shao et al., 2016), whilst positively associated is SPON1 which inhibits excess bone formation (Palmer et al., 2014).

Although these data suggest that BMPR1A-biased signalling is associated with reduced bone formation, we sought to carry out a more unbiased analysis of our differential expression results in the form of a gene set enrichment analysis (GSEA). Figures 2B and C show two of the most highly enriched gene sets, each associated with the matrisome, a computationally predicted collection of extracellular matrix (ECM) constituents and their regulatory networks (Naba et al., 2012).



L79	Together, these data suggest that BMPR1A-biased BMP2 expression is associated with
180	reduced bone formation and an increased turnover of ECM components. The interaction of
181	tumour cells with ECM proteins can have profound effects on their biology, regulating gene
182	expression, signal transduction and migration/invasion (Cassidy, 2014; Lu, Weaver, &
183	Werb, 2012). Moreover, BMP2 signalling has previously been linked to increased ECM
184	production and degradation (Davidson et al., 2007). Thus, we next sought to determine
185	whether BMPR1A-biased signalling had any clinical phenotype.
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BMPR1A-biased BMP2 Signalling is Prognostic of Poor Clinical Outcome

Patients in the TCGA sarcoma dataset have a median disease free survival of 37 months (Figure 3A). Those patients with BMPR1B biased signalling (*n*=44) have no difference in their prognosis when compared to the dataset as a whole. However, those with BMPR1A-biased BMP2 signalling (*n*=32) have a significantly worse outcome when disease free survival is used as the primary endpoint (*p*=0.0087). To discount prognostic influences of BMPR1A, BMPR1B or BMP2 expression individually, we constructed Kaplan-Myer curves for the entire sarcoma dataset and compared patients with the highest and lowest 10% expression of BMPR1A, BMPR1B, BMPR2, BMP2 and BMP7. Despite several interesting trends, no significant associations were seen (Supplementary Figure 3). Thus, in addition to ECM remodelling and reduced bone formation, soft tissue sarcoma patients with BMPR1A-biased signalling are more likely to show tumour progression in the presence of high levels of endogenous BMP2.

Soft tissue sarcomas are classified based on their origin in the soft but not connective tissues of the body. They can be separated based on their cell of origin and are generally considered a heterogeneous mixture of diseases (Cormier & Pollock, 2004). Figure 3B shows the TCGA dataset broken down into subtypes of soft tissue sarcomas. The two best-represented subtypes are Leiomyosarcoma (LMS; smooth muscle sarcomas) and Dedifferentiated liposarcoma (DLS; adipocyte sarcomas), though the majority (60%) of the dataset is unclassified. Disease free survival is shown in Figure 3C; LMS tends to have a better prognosis than DLS though data for the three other subtypes is limited. To determine whether BMPR1A biased signalling was localised to a specific subtype, we profiled expression of BMPR1A, BMPR1B, BMPR2 and BMP2 across the three best-represented subtypes (UPS, LMS and DLS). No significant differences in terms of expression were noted, though UPS did trend toward lower overall expression of both ligand and receptors (Supplementary Figure 4). The two largest groups, LMS and DLS had indistinguishable expression values of BMP signalling nodules.



240	Interestingly, BMPR1A-biased BMP2 signalling conferred a worse prognosis on patients
241	than ether LMS or DLS. Thus, we next sought to determine whether the prognostic benefits
242	of BMPR1A-biased signalling were split evenly across these two major subtypes.
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BMPR1A prognostic signalling is localised to dedifferentiated

liposarcomas

We next profiled disease free survival in patients with LMS or DLS and investigated the effects of either BMPR1A-biased BMP2 signalling or high BMPR1A expression alone. BMPR1A and BMPR1A-biased signalling appeared to have little effect in terms of disease free survival in patients with LMS (Figure 4A), though high expression and biased signalling was highly detrimental in patients with DLS (Figure 4B).

To investigate the mechanism for this effect, we compared global gene expression in DLS patients with BMPR1A-biased signalling versus those with BMPR1B-biased signalling. Despite working with a much reduced dataset of 10 vs 10 patients, we found 1375 differentially expressed genes with a FDR of 0.05, the vast majority (1061) of which were down-regulated (Figure 4C). Such a high number of differentially expressed transcripts compared to our early analysis in the whole sarcoma dataset (Figure 2A) suggests patient groups which are more biologically robust. Amongst these differentially expressed genes, we noted a positive association between BMPR1A-biased signalling and GPR176, an orphan G-protein coupled receptor thought to be involved in atherosclerosis (Karagiannis, Weile, Bader, & Minta, 2013). We performed GSEA on these differentially expressed genes and found a positive enrichment for a number of extracellular matrix and matrisome gene sets (Figure 4D).

Finally, we sought to determine whether the detrimental effect of BMP2-BMPR1A signalling were a pan-cancer phenomenon. rhBMP2 use is associated with an increased incidence in a wide variety of solid tumour types (Supplementary Table 1). Moreover, several large studies performed by The Cancer Genome Atlas (TCGA) show copy number changes and mutational events in BMP2 interacting partners as defining features of that malignancy (Supplementary Figure 5). For example, two separate studies in Ovarian cancer show copy number gains as frequent events, which we later determined to be due to amplifications in the BMP2 and BMP7 loci (Supplementary Figure 6). Interestingly, mRNA



up regulation of both ligands and receptors were frequent events in each of the cancers profiled. Deep deletion of BMPR1A was a frequent event in prostate cancers, this has previously been shown to be related to the loss of the PTEN locus, which is associated with invasive forms of prostate cancer (Yoshimoto et al., 2012). In order to test whether our identified BMPR1A-biased signalling biomarker was useful across tumour types, we investigated disease free survival in Ovarian, Lung, Pancreatic and Prostate cancers in response to BMP2 biased signalling but found no significant alterations (Supplementary Figure 5). Thus, it appears that the detrimental effects of BMPR1A biased signalling are confined to the context of soft tissue sarcomas and in particular dedifferentiated liposarcomas. Clearly as rhBMP2 is associated with multiple new cancer events, it is probable that some other prognostic signalling network could be uncovered in these tumour types in the future.



Discussion

rhBMP2 has substantial utility in vertebral fusion surgery, however, recent concerns over its safety has lead to an FDA black box warning. Specifically, rhBMP2 is used as an adjuvant, along with a collagen type I carrier within the surgical site in order to promote osteoblast differentiation and osteogenesis around an interbody titanium cage (Fu et al., 2013). Following several concerns over its safety, a large meta-analysis confirmed a significant increase in the number of new cancer events in patients given high dose (40 mg) rhBMP2 (Carragee et al., 2013) (Supplementary Table 1). However, the *in vivo* half-life of rhBMP2 is short and consequently it is not expected to have systemic activity (FDA, 2002; Simmonds et al., 2013). Herein, we characterise the role of BMP2 signalling in the soft tissue sarcomas. Although this rare group of diseases were unlikely to be observed within the timeframe of rhBMP2 safety trials (Cormier & Pollock, 2004), we reason that their proximity to the surgical site increases their relevance and warrants further research.

Several studies have shown BMPR1A and BMPR1B have largely overlapping roles in BMP2 signal transduction, for example during chondrogenesis (Yoon et al., 2005). However recent work has highlighted opposing roles in certain situations. Sahni and colleagues found that BMPR1A, but not BMPR1B, knockout mice exhibited defective astrocytic hypertrophy and increased inflammatory infiltrate after spinal chord injury (Sahni et al., 2010). The authors go on to show opposing roles for BMPR1A and BMPR1B in posttranscriptional regulation of astrocytic microRNA-21 (Sahni et al., 2010). We find that BMPR1A and BMPR1B expression is highly heterogeneous throughout the body without any obvious correlation between expression patterns. Moreover we observe differences in subcellular localisation in three separate cell lines. Together, these observations suggest that type I BMP receptors may not be functionally redundant, indeed differential expression of BMPR1A and BMPR1B may lead to differential outcomes of BMP ligand signalling in different tissue types. We reasoned that as BMPR1A expression is enriched relative to BMPR1B in sarcoma cell lines, BMP2-BMPR1A signalling may be important in progression of this tumour type.



Loss of BMPR1A has previously been associated with the formation of juvenile polyposis, a predisposing factor for gastrointestinal and colorectal cancers (Calva-Cerqueira et al., 2010). Reduced expression is also associated with a poor prognosis in pancreatic cancer (Voorneveld et al., 2013). Exemplifying the dichotomy of this signalling pathway in cancer, BMPR1A antagonists reduce growth and induce cell death in lung cancer cell lines (Langenfeld, Hong, Lanke, & Langenfeld, 2013). Similarly, we find that sarcoma patients with BMPR1A biased signalling, despite having unchanged levels of BMP2, have significantly reduced disease free survival. Interestingly, we find that the prognostic value of BMPR1A biased signalling in sarcomas is localised to dedifferentiated liposarcomas and not leiomyosarcomas. Together these subtypes represent over 35% of the sarcoma dataset, with 60% undefined. As the trend holds true for the dataset as a whole, it is possible that other underrepresented subtypes (myxofibrosarcoma, giant cell sarcomas, undifferentiated pleomorphic sarcomas etc.) would show the same pattern if dataset annotation was complete or if the dataset were larger.

Further to linking BMPR1A-biased BMP2 signalling to reduced disease free survival, we report a detailed differential expression analysis in both the sarcoma dataset as a whole and dedifferentiated liposarcomas. Amongst the genes negatively associated with BMPR1A-biased signalling are *WNT7B* which has a role in osteoblast differentiation and mineralised nodule formation (Shao et al., 2016), whilst positively associated is *SPON1* which inhibits excess bone formation (Palmer et al., 2014). The ECM is far from an inert passenger to tumour progression. Rather, cells exist in a dynamic milieu of proteins, cytokines and growth factors which can have profound influence on the propensity to malignancy (Cassidy, 2014; Cassidy et al., 2014) and tumour heterogeneity (Cassidy, Caldas, & Bruna, 2015). Indeed we find that patients with BMPR1A-biased signalling also exhibit increased expression of genes typically linked with the matrisome and ECM remodelling (Naba et al., 2012). Metalloproteinase expression has previously been linked to prognosis in soft tissue sarcomas (Benassi et al., 2001), and BMPR1A is necessary for extracellular matrix deposition by osteoblasts (Mishina et al., 2004). It is tempting to speculate that increased turnover of extracellular matrix in sarcomas is at least in part mediated by BMPR1A and



this contributes to the observed poor prognosis of patients with BMPR1A-biased signalling. Together these data suggest a shift from an osteogenic phenotype to one of increased extracellular matrix remodelling. In summary, we present here evidence that BMP2-BMPR1A signalling leads to extracellular matrix remodelling and tumour progression in dedifferentiated liposarcomas. Although this trend is present in the sarcoma dataset as a whole, we did not observe prognostic ability of BMPR1A signalling in four other solid tumours. rhBMP2 is used extensively as an adjuvant therapy for spinal fusion surgery and it is possible that such treatment could fuel progression of pre-existing sarcomas in these patients. Sarcomas are rare cancers and as such are underdiagnosed in the early stages, but this research underscores the need for continued study of the BMP2-BMPR1A signalling axis in cancer. Moreover, patients administered rhBMP2 should have substantial clinical follow-up to minimise the risk posed by any new cancer events.



Methods

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Data Acquisition

- 423 Protein expression data was downloaded from the Human Protein Atlas (Uhlen et al., 2010;
- 424 Uhlén et al., 2015) (Cancer Atlas and the Cell Line Atlas; proteinatlas.org). Pan-cancer
- analysis was carried out using cBioPortal (cbioportal.org) (Cerami et al., 2012; Gao et al.,
- 426 2013). With pathway data obtained from Cytoscape 3.2.1 (cytoscape.org) (Lopes et al.,
- 427 2010). RNASeqV2 normalised gene results and clinical metadata were downloaded form
- 428 the TCGA Data Portal (tcga-data.nci.nih.gov), for the following cancer types: Sarcoma
- 429 (provisional), Prostate (provisional), Breast (TCGA, 2012), Ovarian (TCGA, 2011), Lung
- 430 (TCGA, 2014) and Head and Neck (TCGA, 2015).

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Data Analysis

- 433 Normalised RNASeqV2 expression values and corresponding clinical metadata were
- 434 manipulated in R Studio Desktop (Mac) 0.99.484 (rstudio.com). Combined data were
- 435 analysed in Microsoft Excel (Mac 14.4.3) and R Studio with results plotted in GrpahPad
- Prism 6 (Mac). Comparisons between two groups were made using two-tailed t-tests or
- 437 Mann-Whitney U-tests and more than two groups by one-way ANOVA or Kruskal-Wallis,
- 438 multiple comparisons were corrected for by Bonferroni post hoc tests. Kaplan-Meier
- disease free survival graphs were constructed in GraphPad Prism 6 and presented as whole
- 440 population versus experimental population. Comparisons between curves were made by
- Log-rank (Mantel-Cox) tests with the Mantel-Cox *p* value reported.

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Differential Expression Analysis

- 444 For patient groups with observed differences in phenotype (pathway activation/disease
- free survival) differential expression analysis was performed on normalised RNASeqV2
- gene counts. Date were combined in R studio and means across rows (genes) calculated.
- 447 Multiple two tailed t-tests with Bonferroni correction were used to compare means. Using a
- false discovery rate of 0.05 differentially expressed genes were ranked based on log₁₀ fold-
- change. Ranked genes were analysed through the Gene Set Enrichment pre-ranked plugin



450	(GSEAPreranked at genepattern.broadinstitute.org) with default parameters and 1000
451	permutations (GenePattern version 3.9.4 (Hubble et al., 2009)). Gene sets
452	c2.all.v5.0.symbols.gmt [Curated] and c6.all.v5.0symbols.gmt [Oncogenic signatures] were
453	used for comparison. Gene sets in the top 10 from the dataset based on enrichments score
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This study was completed without external funding and relies entirely on publicly available data. Without the open access policies adopted by journals and Research Councils over recent years, this study would not have been possible. Moreover, open source initiatives such as the R Project, TCGA, cBioPortal, Protein Atlas and GenePattern/GSEAPreranked this study would have been impossible. The authors are grateful to all those involved in the aforementioned initiatives and all patients who elected to share their data with the community.

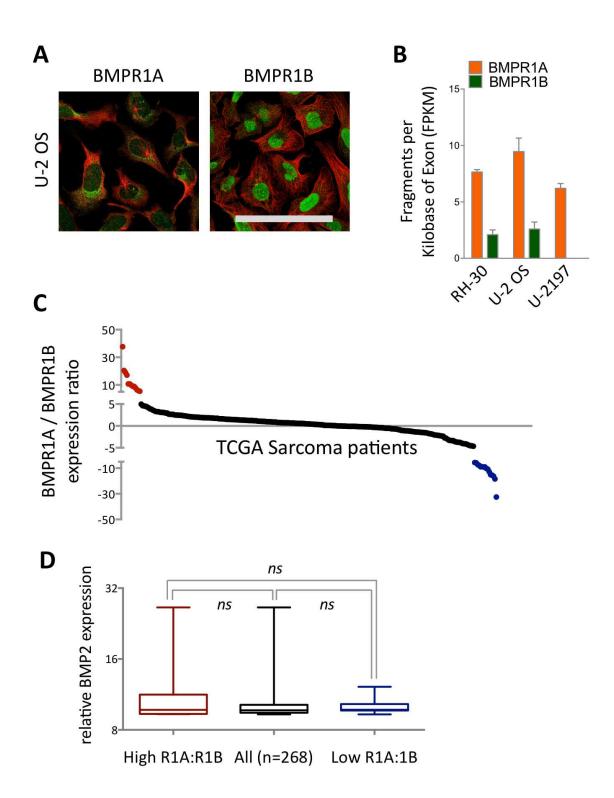
Competing Interests

The authors declare no competing financial interests.



508 Figures

Figure 1

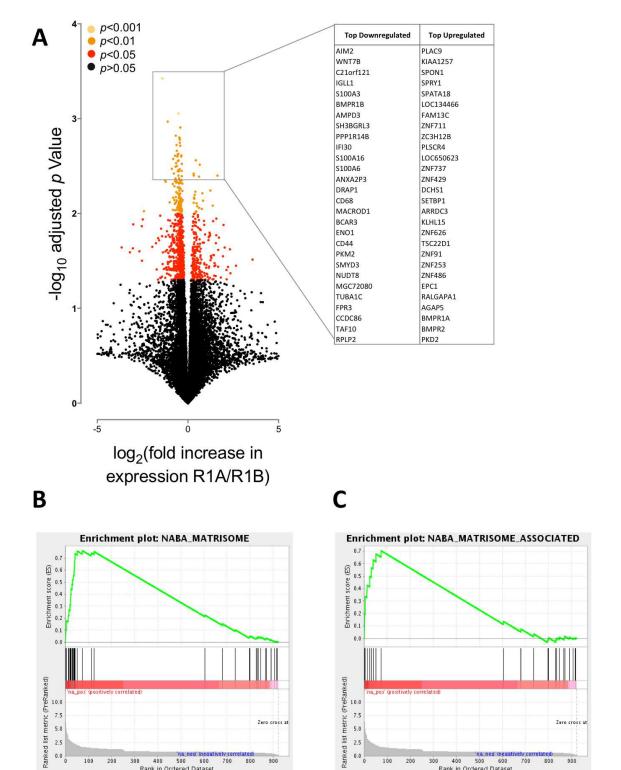




510	Figure 1 legend
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512	A: Immunocytochemistry of BMPR1A and BMPR1B in U-2 OS osteosarcoma cell line. Green
513	is receptor (Antibody CAB019398 for BMPR1A and HPA046821 for BMPR1B) red is
514	microtubules. Scale bar is 100 μm.
515	B: RNAseq data from three sarcoma cell lines, RH-30 is a Rhabdomyosarcoma and U-2197
516	is a Malignant fibrous histiocytoma.
517	C: Ratio of BMPR1A to BMPR1B expression for each patient in the TCGA sarcoma dataset
518	(n =263). Patients with high BMPR1A:BMPR1B expression (top 10%) are colour coded red
519	and patients with high BMPR1B:BMPR1A expression (top 10%) are colour coded blue.
520	D: BMP2 expression in patient sets defined in panel A. Boxplot represents median with
521	upper and lower quartiles with whiskers showing 95% confidence intervals, scale is log_2 .
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Figure 2 541



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Rank in Ordered Dataset

Ranking metric scores

Enrichment profile — Hits

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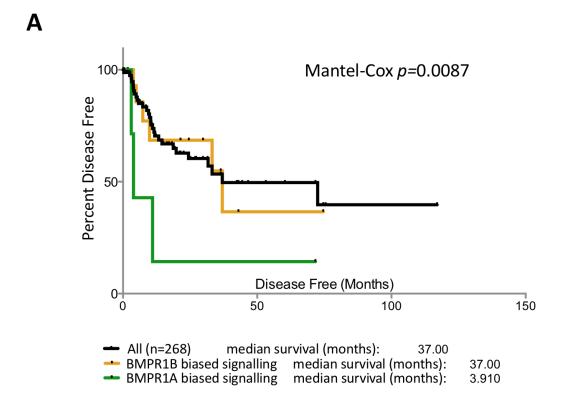
Ranking metric scores

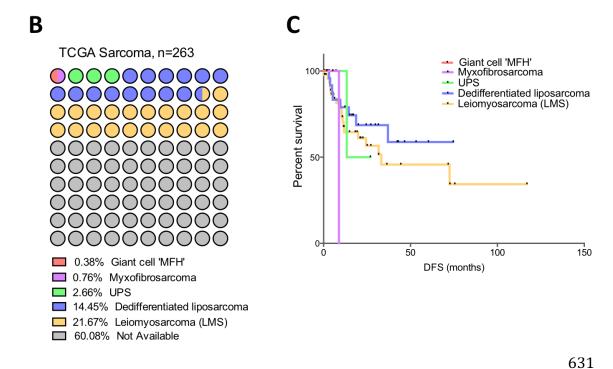
Enrichment profile — Hits



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573	Figure 2 shows the results of differential expression analysis comparing patients with high
574	BMPR1A-biased and high BMPR1B-biased BMP2 signalling i.e. upper quartile BMP2
575	expression and upper quartile R1A:B or R1B:A ratio.
576	A: volcano plot showing all differentially expressed transcripts colour-coded based or
577	adjusted p -value (multiple two-tailed t-tests with Bonferroni correction $post\ hoc$). Insert
578	shows top up- and down-regulated genes from this analysis.
579	B: GSEA enrichment plot for NABA Matrisome gene set; defined in silico based on lung and
580	colon tumour extracellular matrix components (Naba et al., 2012). False discovery rate is
581	p=0.05.
582	C: GSEA enrichment plot for NABA Matrisome associated gene set. False discovery rate is
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Figure 3



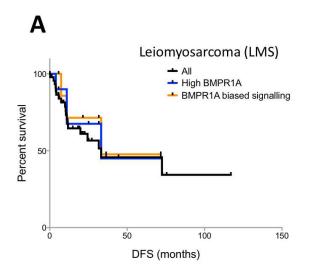


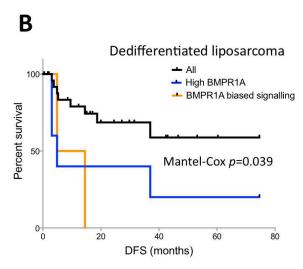


632	Figure 3 legend
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634	A: Kaplan-Myer survival curves (disease free survival) for patients with BMPR1A or
635	BMPR1B biased signalling. Biased signalling is defined as high BMP2 expression with either
636	high BMPR1A (R1A-biased) or high BMPR1B (R1B-biased). Median survival is shown.
637	B: Annotation of the TCGA sarcoma dataset with each dot colour-coded based on subtype
638	and corresponding to 2.6 patients. The majority of the dataset was unclassified but can be
639	expected to have similar proportions.
640	C: shows a Kaplan-Myer disease free survival curve for the different sarcoma subtypes,
641	colours correspond to <i>B</i> .
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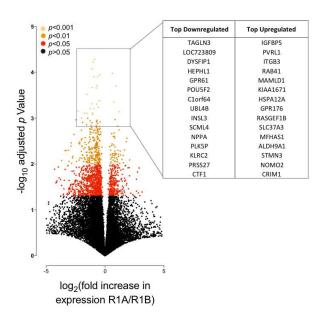


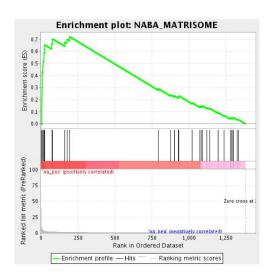
662 **Figure 4**





C Dedifferentiated liposarcoma





D



663	Figure 4 legend
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665	Survival and differential expression analysis between patients with BMPR1A and BMPR1B
666	biased signalling in dedifferentiated liposarcomas.
667	A: disease free Kaplan-Myer survival analysis in Leiomyosarcomas for both high BMPR1A
668	expression (top 10%) and BMPR1A-biased BMP2 signalling (defined as before). UPS is
669	Undifferentiated Pleomorphic Sarcoma.
670	B: disease free Kaplan-Myer survival in Dedifferentiated Liposarcomas for both high
671	BMPR1A expression (top 10%) and BMPR1A-biased BMP2 signalling (defined as before).
672	C: Volcano plot comparing gene expression in Dedifferentiated Liposarcoma patients with
673	BMPR1A-biased versus BMPR1B-biased BMP2 signalling. Top up- and down-regulated
674	genes are shown in the insert.
675	D: Gene set enrichment from differentially expressed genes based on a false discovery rate
676	of 0.05. NABA Matrisome was the most significantly associated gene set. Defined in silico
677	based on lung and colon tumour extracellular matrix components by Naba and collegues
678	(Naba et al., 2012).
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694	Supp	lementary	Figure	Legends
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Supplementary Figure 1

- Tissue specific protein expression of BMPR1A and BMPR1B based on IHC. Data from the
- 698 Protein Atlas (Uhlen et al., 2010).

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Supplementary Figure 2

- 701 Immunostaining of BMP receptors in three cancer cell lines showing intracellular
- localisation. U-2 OS is an osteosarcoma, A-431 is an epidermoid carcinoma and U-251/MG
- is a glioblastoma astrocytoma line. Green is receptor (Antibody CAB019398 for BMPR1A,
- HPA046821 for BMPR1B and HPA049014 for BMPR2) red is microtubules. Scale bar is 100
- 705 μm.

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Supplementary Figure 3

- 708 Kaplan-Myer disease free survival curves in the TCGA sarcoma dataset (n=263) based on
- 709 mRNA expression (top 10% and bottom 10% expressors). From top left; BMP2, BMP7,
- 710 BMPR1A, BMPR1B and BMPR2.

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Supplementary Figure 4

- 713 Median (± interquartile range) expression of BMPR1A, BMPR1B, BMPR2 and BMP2 in
- 714 different sarcoma subsets. UPS is Undifferentiated Pleomorphic Sarcoma and LMS is
- 715 Leiomyosarcoma.

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Supplementary Figure 5

- 718 A: Pan-cancer analysis of BMP2 in several large scale genomics projects. Data from
- 719 cBioPortal (cbioportal.org) (Cerami et al., 2012; Gao et al., 2013).
- 720 B: BMP2 interacting partners, defined in Cytoscape 3.2.1 (cytoscape.org) (Lopes et al.,
- 721 2010).



723	Supplementary Figure 6
724	Detailed waterfall plots for BMP pathway members deregulated (mutation, copy-number
725	up/down-regulation) in various cancer types. Vertical bars represent individual patients
726	from the following cancer types: Sarcoma (provisional), Prostate (provisional), Breast (T. C
727	G A. R. Network, 2012), Ovarian (T. C. G. A. R. Network, 2011), Lung (T. C. G. A. R. Network
728	2014) and Head and Neck (T. C. G. A. Network, 2015). Visualisation was cBioPorta
729	(cbioportal.org) (Cerami et al., 2012; Gao et al., 2013).
730	
731	Supplementary Figure 7
732	Kaplan-Myer disease free survival curves for BMPR1A-biased, BMPR1B-biased and BMPR2
733	signalling by high levels of endogenous BMP2. From top left, Prostate (provisional)
734	Pancreatic (provisional), Lung (T. C. G. A. R. Network, 2014) and Ovarian (T. C. G. A. R.
735	Network, 2011) cancers.
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753	Supplementary Table Legends
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755	Supplementary Table 1
756	Incidents of new cancers from a pivotal, multicentre, randomised controlled trial of
757	patients with degenerative lumbar spine conditions who underwent a single-level
758	instrumented posterolateral arthrodesis with either high-dose rhBMP-2 in a compression-
759	resistant matrix (CRM) (rhBMP-2/CRM; n = 239) or autogenous bone graft (control group;
760	n = 224).
761	
762	Supplementary Table 2
763	Differential expression analysis from BMPR1A-biased and BMPR1B-biased BMP2 signalling
764	in the TCGA sarcoma dataset.
765	
766	Supplementary Table 3
767	Differential expression analysis from BMPR1A-biased and BMPR1B-biased BMP2 signalling
768	in dedifferentiated liposarcomas.
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782	References
783	Ben-David, D., Srouji, S., Shapira-Schweitzer, K., Kossover, O., Ivanir, E., Kuhn, G., Livne, E.
784	(2013). Low dose BMP-2 treatment for bone repair using a PEGylated fibrinogen
785	hydrogel matrix. <i>Biomaterials</i> , 34(12), 2902–2910.
786	doi:10.1016/j.biomaterials.2013.01.035
787	Benassi, M. S., Gamberi, G., Magagnoli, G., Molendini, L., Ragazzini, P., Merli, M., Picci, P.
788	(2001). Metalloproteinase expression and prognosis in soft tissue sarcomas. Annals of
789	Oncology: Official Journal of the European Society for Medical Oncology / ESMO, 12(1),
790	75-80. doi:10.1023/A:1008318614461
791	Bonor, J., Adams, E. L., Bragdon, B., Moseychuk, O., Czymmek, K. J., & Nohe, A. (2012).
792	Initiation of BMP2 signaling in domains on the plasma membrane. Journal of Cellular
793	Physiology, 227(7), 2880-2888. doi:10.1002/jcp.23032
794	Calva-Cerqueira, D., Dahdaleh, F. S., Woodfield, G., Chinnathambi, S., Nagy, P. L., Larsen-
795	Haidle, J., Howe, J. R. (2010). Discovery of the BMPR1A promoter and germline
796	mutations that cause juvenile polyposis. Human Molecular Genetics, 19(23), 4654-
797	4662. doi:10.1093/hmg/ddq396
798	Carragee, E. J., Chu, G., Rohatgi, R., Hurwitz, E. L., Weiner, B. K., Yoon, S. T., Kopjar, B.
799	(2013). Cancer Risk After Use of Recombinant Bone Morphogenetic Protein-2 for
800	Spinal Arthrodesis. <i>The Journal of Bone & Done & Surgery</i> , 95(17), 1537–1545.
801	Cassidy, J. W. (2014). Nanotechnology in the Regeneration of Complex Tissues. Bone and
802	Tissue Regeneration Insights, 25. doi:10.4137/BTRI.S12331
803	Cassidy, J. W., Caldas, C., & Bruna, A. (2015). Maintaining Heterogeneity in Patient Derived
804	Tumour Xenografts. Cancer Research, 75(15), 1–6. doi:10.1158/0008-5472.CAN-15-
805	0727
806	Cassidy, J. W., Roberts, J. N., Smith, CA., Robertson, M., White, K., Biggs, M. J., Dalby, M. J.
807	(2014). Osteogenic lineage restriction by osteoprogenitors cultured on nanometric
808	grooved surfaces: the role of focal adhesion maturation. Acta Biomaterialia, 10(2),
809	651-60. doi:10.1016/j.actbio.2013.11.008



310 311 312	The cBio Cancer Genomics Portal: An open platform for exploring multidimensional cancer genomics data. <i>Cancer Discovery</i> , 2(5), 401–404. doi:10.1158/2159-8290.CD-
313	12-0095
314 315	Chen, D., Zhao, M., & Mundy, G. R. (2004). Bone morphogenetic proteins. <i>Growth Factors</i> (Chur, Switzerland), 22(4), 233–241. doi:10.1080/08977190412331279890
316 317	Cormier, J. N., & Pollock, R. E. (2004). Soft Tissue Sarcomas. <i>CA: A Cancer Journal for Clinicians</i> , <i>54</i> (2), 94–109. doi:10.3322/canjclin.54.2.94
318 319	Data, A. M. I. (2013). Annals of Internal Medicine Review Safety and Effectiveness of Recombinant Human Bone Morphogenetic Protein-2 for Spinal Fusion.
320 321 322 323 324	Davidson, E. N. B., Vitters, E. L., van Lent, P. L. E. M., van de Loo, F. A. J., van den Berg, W. B., & van der Kraan, P. M. (2007). Elevated extracellular matrix production and degradation upon bone morphogenetic protein-2 (BMP-2) stimulation point toward a role for BMP-2 in cartilage repair and remodeling. <i>Arthritis Research & Therapy</i> , 9(5), R102–R102. doi:10.1186/ar2305
325 326 327	Deyo, R. A., Nachemson, A., & Mirza, S. K. (2004). Spinal-Fusion Surgery — The Case for Restraint. <i>New England Journal of Medicine</i> , <i>350</i> (7), 722–726. doi:10.1056/NEJMsb031771
328 329	FDA. (2002). Summary of Safety and Effectiveness Data for InFUSE Bone Graft for Spinal Fusion Procedures (Vol. P000058).
330 331 332 333	Fu, R., Selph, S., McDonagh, M., Peterson, K., Tiwari, A., Chou, R., & Helfand, M. (2013). Effectiveness and Harms of Recombinant Human Bone Morphogenetic Protein-2 in Spine FusionA Systematic Review and Meta-analysis. <i>Annals of Internal Medicine</i> , 158(12), 890–902.
334 335 336	Gao, J., Aksoy, B. A., Dogrusoz, U., Dresdner, G., Gross, B., Sumer, S. O., Schultz, N. (2013). Integrative Analysis of Complex Cancer Genomics and Clinical Profiles Using the cBioPortal. <i>Science Signaling</i> , 6(269), pl1–pl1. doi:10.1126/scisignal.2004088
337	Gitelis, B. S., Wilkins, R. M., & Yasko, A. W. (2008). BMPs and cancer: Is the risk real? AAOS



838	Now, 1, 2-5.
839 840 841	Hubble, J., Demeter, J., Jin, H., Mao, M., Nitzberg, M., Reddy, T. B. K., Ball, C. a. (2009). Implementation of GenePattern within the Stanford Microarray Database. <i>Nucleic Acids Research</i> , <i>37</i> (SUPPL. 1), 898–901. doi:10.1093/nar/gkn786
842 843 844	Karagiannis, G. S., Weile, J., Bader, G. D., & Minta, J. (2013). Integrative pathway dissection of molecular mechanisms of moxLDL-induced vascular smooth muscle phenotype transformation. <i>BMC Cardiovascular Disorders</i> , 13, 4. doi:10.1186/1471-2261-13-4
845 846 847	Kisiel, M., Klar, A. S., Ventura, M., Buijs, J., Mafina, M. K., Cool, S. M., & Hilborn, J. (2013). Complexation and Sequestration of BMP-2 from an ECM Mimetic Hyaluronan Gel for Improved Bone Formation. <i>PLoS ONE</i> , 8(10), 1–13. doi:10.1371/journal.pone.0078551
848 849 850	Laforest, L., Brown, C. W., Poleo, G., Géraudie, J., Tada, M., Ekker, M., & Akimenko, M. a. (1998). Involvement of the sonic hedgehog, patched 1 and bmp2 genes in patterning of the zebrafish dermal fin rays. <i>Development (Cambridge, England)</i> , 125(21), 4175–4184.
851 852 853	Langenfeld, E., Hong, C. C., Lanke, G., & Langenfeld, J. (2013). Bone Morphogenetic Protein Type I Receptor Antagonists Decrease Growth and Induce Cell Death of Lung Cancer Cell Lines. <i>PLoS ONE</i> , 8(4), e61256.
854 855 856	Lopes, C. T., Franz, M., Kazi, F., Donaldson, S. L., Morris, Q., & Bader, G. D. (2010). Cytoscape Web: an interactive web-based network browser. <i>Bioinformatics</i> , <i>26</i> (18), 2347–2348. doi:10.1093/bioinformatics/btq430
857 858	Lu, P., Weaver, V. M., & Werb, Z. (2012). The extracellular matrix: a dynamic niche in cancer progression. <i>The Journal of Cell Biology</i> , 196(4), 395–406. doi:10.1083/jcb.201102147
859 860 861 862	Mishina, Y., Starbuck, M. W., Gentile, M. a., Fukuda, T., Kasparcova, V., Seedor, J. G., Behringer, R. R. (2004). Bone morphogenetic protein type IA receptor signaling regulates postnatal osteoblast function and bone remodeling. <i>Journal of Biological Chemistry</i> , 279(26), 27560–27566. doi:10.1074/jbc.M404222200
863 864	Munoz-Sanjuan, I., & Brivanlou, A. H. (2002). Neural induction, the default model and embryonic stem cells. <i>Nat Rev Neurosci</i> , <i>3</i> (4), 271–280.
865	Naba, a., Clauser, K. R., Hoersch, S., Liu, H., Carr, S. a., & Hynes, R. O. (2012). The Matrisome:



866	In Silico Definition and In Vivo Characterization by Proteomics of Normal and Tumor
867868	Extracellular Matrices. <i>Molecular & Cellular Proteomics</i> , 11(4), M111.014647–M111.014647. doi:10.1074/mcp.M111.014647
869 870	Network, T. C. G. A. (2015). Comprehensive genomic characterization of head and neck squamous cell carcinomas. <i>Nature</i> , <i>517</i> (7536), 576–582.
871 872	Network, T. C. G. A. R. (2011). Integrated genomic analyses of ovarian carcinoma. <i>Nature</i> , 474(7353), 609–615.
873 874	Network, T. C. G. A. R. (2012). Comprehensive molecular portraits of human breast tumours. <i>Nature</i> , 490(7418), 61–70.
875 876	Network, T. C. G. A. R. (2014). Comprehensive molecular profiling of lung adenocarcinoma. <i>Nature</i> , <i>511</i> (7511), 543–550.
877 878 879	Palmer, G. D., Attur, M. G., Yang, Q., Liu, J., Moon, P., Beier, F., & Abramson, S. B. (2014). F-Spondin Deficient Mice Have a High Bone Mass Phenotype. <i>PLoS ONE</i> , <i>9</i> (5), e98388. doi:10.1371/journal.pone.0098388
880 881 882	Sahni, V., Mukhopadhyay, A., Tysseling, V., Hebert, A., Birch, D., Mcguire, T. L., Kessler, J. A (2010). BMPR1a and BMPR1b Signaling Exert Opposing Effects on Gliosis after Spinal Cord Injury, <i>30</i> (5), 1839–1855. doi:10.1523/JNEUROSCI.4459-09.2010
883 884 885	Shao, X., Lyon, K., Fong-Yee, C., Jia, W., Dunstan, C. R., Chen, D., Zhou, H. (2016). Wnt7b plays a unique and essential role in osteoblast differentiation. <i>Bone</i> , <i>47</i> , S370. doi:10.1016/j.bone.2010.09.116
886 887 888 889	Simmonds, M. C., Brown, J. V. E., Heirs, M. K., Higgins, J. P. T., Mannion, R. J., Rodgers, M. A., & Stewart, L. A. (2013). Safety and Effectiveness of Recombinant Human Bone Morphogenetic Protein-2 for Spinal FusionA Meta-analysis of Individual-Participant Data. <i>Annals of Internal Medicine</i> , 158(12), 877–889.
890 891 892	Song, J., McColl, J., Camp, E., Kennerley, N., Mok, G. F., McCormick, D., Münsterberg, A. E. (2014). Smad1 transcription factor integrates BMP2 and Wnt3a signals in migrating cardiac progenitor cells. <i>Proceedings of the National Academy of Sciences of the United</i>
893	States of America, 111(20), 7337-42. doi:10.1073/pnas.1321764111



394 395 396	angiogenesis but retard terminal differentiation of osteoblasts in bone regeneration by up-regulating Id1. <i>Acta Biochimica et Biophysica Sinica</i> , 43 (10), 796–804.
397 398 399	Thawani, J. P., Wang, A. C., Than, K. D., Lin, CY., La Marca, F., & Park, P. (2010). Bone morphogenetic proteins and cancer: review of the literature. <i>Neurosurgery</i> , 66(2), 233–246; discussion 246. doi:10.1227/01.NEU.0000363722.42097.C2
900 901 902 903	Tsuji, K., Bandyopadhyay, A., Harfe, B. D., Cox, K., Kakar, S., Gerstenfeld, L., Rosen, V. (2006). BMP2 activity, although dispensable for bone formation, is required for the initiation of fracture healing. <i>Nature Genetics</i> , <i>38</i> (12), 1424–1429. doi:10.1038/ng1916
904 905 906	Uhlén, M., Fagerberg, L., Hallström, B. M., Lindskog, C., Oksvold, P., Mardinoglu, A., Pontén F. (2015). Tissue-based map of the human proteome. <i>Science</i> , 347 (6220). doi:10.1126/science.1260419
907 908 909	Uhlen, M., Oksvold, P., Fagerberg, L., Lundberg, E., Jonasson, K., Forsberg, M., Ponten, F. (2010). Towards a knowledge-based Human Protein Atlas. <i>Nat Biotech</i> , <i>28</i> (12), 1248–1250.
910 911 912	Vaz, K., Verma, K., Protopsaltis, T., Schwab, F., Lonner, B., & Errico, T. (2010). Bone grafting options for lumbar spine surgery: a review examining clinical efficacy and complications. <i>SAS Journal</i> , <i>4</i> (3), 75–86. doi:10.1016/j.esas.2010.01.004
913 914 915	Voorneveld, P. W., Stache, V., Jacobs, R. J., Smolders, E., Sitters, A. I., Liesker, A., Hardwick, J. C. H. (2013). Reduced expression of bone morphogenetic protein receptor IA in pancreatic cancer is associated with a poor prognosis. <i>Br J Cancer</i> , <i>109</i> (7), 1805–1812.
916 917 918 919	Yoon, B. S., Ovchinnikov, D. A., Yoshii, I., Mishina, Y., Behringer, R. R., & Lyons, K. M. (2005). Bmpr1a and Bmpr1b have overlapping functions and are essential for chondrogenesis in vivo. <i>Proceedings of the National Academy of Sciences of the United States of America</i> , 102(14), 5062–5067. doi:10.1073/pnas.0500031102
920 921 922	Yoshimoto, M., Ludkovski, O., DeGrace, D., Williams, J. L., Evans, A., Sircar, K., Squire, J. A. (2012). PTEN genomic deletions that characterize aggressive prostate cancer originate close to segmental duplications. <i>Genes, Chromosomes and Cancer</i> , <i>51</i> (2), 149–160.