

A peer-reviewed version of this preprint was published in PeerJ on 21 April 2016.

[View the peer-reviewed version](https://peerj.com/articles/1897) (peerj.com/articles/1897), which is the preferred citable publication unless you specifically need to cite this preprint.

Simeone MC, Grimm GW, Papini A, Vessella F, Cardoni S, Tordoni E, Piredda R, Franc A, Denk T. 2016. Plastome data reveal multiple geographic origins of *Quercus* Group Ilex. PeerJ 4:e1897
<https://doi.org/10.7717/peerj.1897>

Plastome data reveal multiple geographic origins of *Quercus* Group Ilex

Marco Cosimo Simeone, Guido W Grimm, Alessio Papini, Federico Vessella, Simone Cardoni, Enrico Tordoni, Roberta Piredda, Alain Franc, Thomas Denk

Nucleotide sequences from the plastome are currently the main source for assessing taxonomic and phylogenetic relationships in flowering plants and their historical biogeography at all hierarchical levels. One exception is the large and economically important genus *Quercus* (oaks). Whereas differentiation patterns of the nuclear genome are in agreement with morphology and the fossil record, diversity patterns in the plastome are at odds with established taxonomic and phylogenetic relationships. However, the extent and evolutionary implications of this incongruence has yet to be fully uncovered. The DNA sequence divergence of four Euro-Mediterranean Group Ilex oak species (*Quercus ilex*, *Q. coccifera*, *Q. aucheri*, *Q. alnifolia*) was explored at three chloroplast markers (*rbcL*, *trnK-matK*, *trnH-psbA*). Phylogenetic relationships were reconstructed including worldwide members of additional 55 species representing all *Quercus* subgeneric groups. Family and order sequence data were harvested from gene banks to better frame the observed divergence in larger taxonomic contexts. We found a strong geographic sorting in the focal group and the genus in general that is entirely decoupled from species boundaries. Main plastid haplotypes shared by distinct oak lineages from the same geographic region and high plastid diversity in members of Group Ilex are indicative for a polyphyletic origin of their plastomes. The results suggest that incomplete lineage sorting and repeated phases of unidirectional introgression among ancestral lineages of Group Ilex and two other main Groups of Eurasian oaks (Cyclobalanopsis and Cerris) caused this complex pattern. Comparison with the current phylogenetic synthesis also suggests an initial high- versus mid-latitude biogeographic split within *Quercus*. High plastome plasticity of Group Ilex reflects geographic area disruptions, possibly linked with high tectonic activity of past and modern distribution ranges, that did not leave imprints in the nuclear genome of modern species and infrageneric lineages.

1 **Title: Plastome data reveal multiple geographic origins of *Quercus* Group Ilex**

2

3 **Authors**

4 Marco Cosimo Simeone ^{1,*}, Guido W. Grimm ², Alessio Papini ³, Federico Vessella ¹, Simone
5 Cardoni ¹, Enrico Tordoni ⁴, Roberta Piredda ⁵, Alain Franc ^{6,7}, Thomas Denk ⁸

6

7 **Affiliations**

8 ¹ = Department of Agricultural and Forestry Science (DAFNE), Università degli Studi della
9 Tuscia, via S. Camillo de Lellis, 01100 Viterbo, Italy

10 ² = Department of Palaeontology, University of Wien, Althanstrasse 14 (UZA II), 1090 Wien,
11 Austria

12 ³ =Dipartimento di Biologia Vegetale, Università degli studi di Firenze, via La Pira 4, 50121
13 Firenze, Italy

14 ⁴ =Department of Life Science, Università degli studi di Trieste, via L. Giorgieri 10, 34127
15 Trieste, Italy

16 ⁵ =Stazione Zoologica Anton Dohrn, Villa Comunale, 8012 Napoli, Italy

17 ⁶ = INRA, UMR BIOGECO-1202, 69 route d'Arcachon, F-33612 Cestas, France

18 ⁷ = Université Bordeaux, UMR BIOGECO-1202, Avenue des Facultés, F-33405 Talence, France

19 ⁸ = Department of Palaeobiology, Swedish Museum of Natural History, Box 50007, 10405
20 Stockholm, Sweden

21

22 **Corresponding Author**

23 Dr. Marco Cosimo Simeone

- 24 Department of Agricultural and Forestry Science (DAFNE), Università degli Studi della Tuscia,
25 via S. Camillo de Lellis, 01100 Viterbo, Italy
26 E-Mail address: mcsimeone@unitus.it

27 **Abstract**

28 Nucleotide sequences from the plastome are currently the main source for assessing taxonomic
29 and phylogenetic relationships in flowering plants and their historical biogeography at all
30 hierarchical levels. One exception is the large and economically important genus *Quercus* (oaks).
31 Whereas differentiation patterns of the nuclear genome are in agreement with morphology and
32 the fossil record, diversity patterns in the plastome are at odds with established taxonomic and
33 phylogenetic relationships. However, the extent and evolutionary implications of this
34 incongruence has yet to be fully uncovered. The DNA sequence divergence of four Euro-
35 Mediterranean Group Ilex oak species (*Quercus ilex* L., *Q. coccifera* L., *Q. aucheri* Jaub. &
36 Spach., *Q. alnifolia* Poech.) was explored at three chloroplast markers (rbcL, trnK-matK, trnH-
37 psbA). Phylogenetic relationships were reconstructed including worldwide members of
38 additional 55 species representing all *Quercus* subgeneric groups. Family and order sequence
39 data were harvested from gene banks to better frame the observed divergence in larger
40 taxonomic contexts. We found a strong geographic sorting in the focal group and the genus in
41 general that is entirely decoupled from species boundaries. Main plastid haplotypes shared by
42 distinct oak lineages from the same geographic region and high plastid diversity in members of
43 Group Ilex are indicative for a polyphyletic origin of their plastomes. The results suggest that
44 incomplete lineage sorting and repeated phases of unidirectional introgression among ancestral
45 lineages of Group Ilex and two other main Groups of Eurasian oaks (Cyclobalanopsis and Cerris)
46 caused this complex pattern. Comparison with the current phylogenetic synthesis also suggests
47 an initial high- versus mid-latitude biogeographic split within *Quercus*. High plastome plasticity
48 of Group Ilex reflects geographic area disruptions, possibly linked with high tectonic activity of

49 past and modern distribution ranges, that did not leave imprints in the nuclear genome of modern
50 species and infrageneric lineages.

51

52 **Keywords:** Fagaceae, Mediterranean, Plastome polyphyly, Ancient introgression, Incomplete
53 lineage sorting, Decoupled phylogenies

54 **Introduction**

55 *Quercus* L. (oaks) is among the most ecologically diverse and economically important
56 extratropical tree genera in the northern hemisphere (Govaerts and Frodin, 1998). *Quercus* is the
57 largest genus in the order Fagales, comprising ca. 400–500 species. Oaks are concentrated in the
58 Americas (Groups *Quercus*, *Lobatae* and *Protobalanus*; Flora of North America Editorial
59 Committee, 1997) and Southeast Asia and southern China (Group *Cyclobalanopsis*; Flora of
60 China Editorial Committee, 1999). In contrast, a relatively lower number of species can be found
61 in western Eurasia and the Mediterranean (Groups *Ilex* and *Cerris*; Kubitzki, 1993; Menitsky,
62 2005). The six major infrageneric lineages of *Quercus* occur from the tropics to the high
63 mountains of the temperate zone and to the boreal continental, cold temperate regions (Denk and
64 Grimm, 2010). The northern limit of oaks in North America and Eurasia coincides with the
65 border of *Dfb* to *Dfc* and *Dwb* to *Dwc* climates, snow climates with warm versus cool summers
66 (Köppen, 1936; Kottek et al., 2006; Peel et al., 2007).

67 Recent molecular phylogenetic studies at and below the genus level focussed on the nucleome of
68 oaks (Oh and Manos, 2008; Denk and Grimm, 2010; Hipp et al., 2014; Hubert et al., 2014).

69 These studies consistently recovered two main lineages, the ‘New World Clade’ comprising the
70 white oaks (Group *Quercus*), red oaks (Group *Lobatae*) and golden-cup oaks (Group
71 *Protobalanus*), and the ‘Old World Clade’ consisting of the cycle-cup oaks (Group

72 Cyclobalanopsis), the Ilex oaks (Group Ilex) and the Cerris oaks (Group Cerris). Evidence from
73 nuclear markers and the fossil record suggests that the initial split in the ‘New World Clade’ was
74 pre-Oligocene between the lineages leading to Group Lobatae and Group Protobalanus/Quercus
75 (Bouchal et al., 2014; Hubert et al., 2014; Grímsson et al., 2015). This early radiation of the
76 Quercus/Protobalanus lineage left its imprints in the molecular signatures of the few modern
77 species of Group Protobalanus and two narrow endemic white oak species, *Quercus pontica*
78 (north-eastern Turkey, south-western Georgia; Denk and Grimm, 2010) and *Q. sadleriana*
79 (California; Hubert et al., 2014). Within the ‘Old World Clade’, the major split was established
80 between the evergreen Groups Cyclobalanopsis and Ilex during the Eocene/Oligocene, whereas
81 the chiefly temperate Group Cerris is suggested to have evolved (‘budded’) from a Group Ilex
82 stock, possibly in Europe, not before the earliest Miocene (Denk and Grimm, 2009; Kmenta,
83 2011; Hubert et al., 2014; Velitzelos et al., 2014).

84 Nuclear amplicon data sets have also contributed to resolve the circumscription of these six
85 groups and to delineate some intergroup and interspecies relationships (López de Heredia et al.,
86 2007; Pearse and Hipp, 2009; Denk and Grimm, 2010; Hubert et al., 2014); well-resolved
87 within-lineage relationships were recently obtained from phylogenomic data in the genetically
88 least-diverged, but species-rich Group Quercus (Hipp et al., 2014). Nucleome-based studies,
89 therefore, clearly indicate a strong correlation between morphology/speciation and nuclear
90 differentiation in oaks. In contrast, oak plastid haplotypes are extensively shared between groups
91 of species (Whittemore and Schaal, 1991; Belahbib et al., 2001; Manos and Stanford, 2001; Petit
92 et al. 2002; Kanno et al., 2004; López de Heredia et al., 2007; Okaura et al., 2007; Neophytou et
93 al., 2010; Gugger and Cavender-Bares 2013). Notably, this was also observed in other genera of
94 Fagaceae such as *Fagus* (Fujii et al. 2002; Lei et al. 2012; Zhang et al. 2013b) and *Lithocarpus*

95 (Cannon and Manos, 2003), and other Fagales such as the northern hemispheric *Carya*
96 (Juglandaceae; Zhang et al., 2013a) and the South American *Nothofagus* (Nothofagaceae; Acosta
97 and Premoli, 2010; Premoli et al., 2012). Plastomes of this large group of long-lived woody
98 plants appear to retain molecular signatures of evolutionary events that cannot be investigated
99 when considering the nuclear DNA alone (e.g., Cavender-Bares et al., 2011; Premoli et al.,
100 2012). As such, they can provide additional information to complement hypotheses on
101 diversification and speciation processes. However, the extent and evolutionary implications of
102 nuclear-plastome incongruence in *Quercus* have yet to be fully uncovered.

103 Testing the potential of DNA barcoding in western Eurasian oaks, Simeone et al. (2013) recently
104 found puzzling diversity in the plastid haplotypes of samples belonging to Group Ilex. In
105 addition to interspecific haplotype sharing, paraphyly to Groups Cerris and *Quercus* and an
106 underlying geographic partitioning was suggested. In the present study, we increased the
107 geographic coverage and taxon sampling to explore the complex patterns of plastome evolution
108 in *Quercus* Group Ilex. This species group is today confined to extra-tropical regions of Eurasia,
109 spanning from arid Mediterranean maquis to high mountain and sub-alpine Himalayan forests
110 and thickets, and to subtropical forests of SE Asia. Group Ilex includes some 35 evergreen,
111 mostly sclerophyllous taxa, whose taxonomy is still controversial (see Table 1) and
112 biogeographic history is not yet well understood (Menitsky, 2005; Denk and Grimm, 2010). In
113 this work, we compiled plastid sequence data for 81 accessions of 20 oak taxa of Group Ilex. The
114 main sampling effort was put into the four species currently occurring in the Mediterranean and
115 adjacent regions in North Africa (Atlas Mountains) and northern Turkey (Black Sea region): the
116 widespread *Quercus ilex* L. and *Q. coccifera* L., and the two East Mediterranean narrow
117 endemics *Q. aucheri* Jaub. & Spach. and *Q. alnifolia* Poech. Data for additional 56 individuals of

118 ca. 40 species were also produced to integrate all subgeneric *Quercus* groups and their
119 worldwide geographic distribution. Additionally, Fagales data sets were harvested from gene
120 banks to allow interpretation of the observed divergence in the plastid markers within a larger
121 taxonomic frame. Our objectives were: (1) to assess the extent of plastome diversity in the Euro-
122 Mediterranean focal group; (2) to outline key phylogeographic patterns within *Quercus* Group
123 Ilex; (3) to establish major evolutionary steps for the differentiation of the ‘Old World Clade’.

124

125 **Material and methods**

126

127 *Plant Material, DNA amplification and analyses*

128 Our analysis included 59 individuals of the four Mediterranean *Quercus* Group Ilex species
129 (Table S1) covering their entire range in North Africa and western Eurasia. Additionally, 22
130 individuals of 16 Asian species of Group Ilex were analysed. The final dataset also included all
131 species of the western North American Group Protobalanus (five species, 10 individuals), 16
132 species of Group Quercus (20 individuals, from North America and Eurasia), five species of the
133 East Asian Group Cyclobalanopsis (11 individuals), seven species of the American Group
134 Lobatae (eight individuals), and six species of Group Cerris (seven individuals). The outgroup
135 set was represented by one sample each of the monotypic genera *Notholithocarpus* and
136 *Chrysolepis* (western North America) and one species each of *Castanea* and *Castanopsis* [(NCBI
137 GenBank accessions HQ336406 (complete plastid genome of *C. mollissima*), JN044213,
138 JF941179, FJ185053). Based on their genetic (plastid) signatures these genera are the closest
139 relatives of *Quercus* within the Fagaceae (Manos et al., 2008). For voucher information and
140 accession numbers see Table S1. The molecular analyses included three plastid DNA regions: a

141 part of the *rbcL* gene, the *trnH-psbA* intergenic spacer and a portion of the *trnK/matK* region (3'
142 intron and partial gene). These markers were chosen based the variability displayed in previous
143 works (e.g. Manos et al., 2001; Okaura et al., 2007; Simeone et al., 2013) and on the of the high
144 number of their sequences available on GenBank. DNA extractions, primers and PCR protocols
145 were the same as in (Piredda et al., 2011) and (Simeone et al., 2013). Sequencing was performed
146 at Macrogen (<http://www.macrogen.com>); electropherograms were edited with CHROMAS 2.3
147 (<http://www.technelysium.com.au>) and checked visually.

148

149 *Statistical tools*

150 The diversity of the investigated regions was evaluated with MEGA 5.2 (Tamura et al., 2011) and
151 DNASP 5.1 (Librado and Rozas, 2009). For comparisons of divergence patterns across all
152 Fagales, available data in gene banks were processed using GBK2FAS (Göker et al., 2009);
153 multiple sequence alignments were done with MAFFT v.7 (Kato and Standley, 2013) using
154 default setting and checked by eye to remove inconsistencies and erroneous sequences (taxa and
155 sequence numbers in Supporting Information). To minimise the effect of alignment gaps, and
156 since we were primarily interested in assessing intra- and intergeneric divergence, alignments
157 included only subsets of the Fagales: 1) Nothofagaceae (data covering all four genera); 2)
158 Fagaceae (10 genera including *Quercus*); 3) Betulaceae-*Ticodendron*-Casuarinaceae (11 genera);
159 4) Juglandaceae (9 genera); 5) Myricaceae (4 genera). Pairwise distance matrices (uncorrected *p*-
160 distance, K2P, HKY, GTR+ Γ) for each marker were calculated with PAUP* 4.0 (Swofford,
161 2002). Minimum intra-specific and minimum/maximum inter-specific distances (calculated with
162 G2CEF; Göker and Grimm, 2008) within and between genera, subgenera in the case of *Fagus*, and
163 infrageneric groups in case of *Quercus*, are listed in Table S2.

164

165 *Phylogenetic analyses*

166 Multiple sequence alignments for the focal group were obtained with CLUSTALW 1.81

167 (Thompson et al., 1994) and checked by eye. The matrices were concatenated with the Python

168 programme COMBINEX2_0.PY (PYTHON v. 2.6.4; BIOPYTHON 1.57).

169 Maximum likelihood trees were inferred with GARLI (Zwickl, 2006; run on the CIPRES portal,

170 http://www.phylo.org/sub_sections/portal/) using four data partitions (*rbcL* and *matK* codons,171 *trnK* intron and *trnH-psbA* spacer). MRMODELTEST 2.0 (Nylander, 2004) and the Akaike

172 Information Criterion (AIC; Akaike, 1974) were used to decide on the best-fitting substitution

173 model for each partition.

174 MRMODELTEST2.0 results were also used for setting up Bayesian inference, performed with

175 MRBAYES 3.4b4 (Ronquist and Huelsenbeck 2003; Ronquist et al. 2012). RAXML v. 7.0.4

176 (Stamatakis et al., 2008) was used for calculating maximum likelihood bootstrap support (1000

177 replicates). Trees were edited with FIGTREE 1.3.1 (Rambaut, 2014) and MESQUITE v. 2.75

178 (Maddison and Maddison, 2011). Median-joining (MJ) haplotype networks were inferred with

179 NETWORK 4.6.1.1 (<http://www.fluxus-engineering.com/>) for each gene region (*rbcL*, *trnK/matK*,180 *trnH-psbA*), treating gaps either as missing or 5th state. MJ algorithm was invoked with default

181 parameters (equal weight of transversion/transition), in order to handle large datasets and

182 multistate characters.

183 Primary data, analysis and supplementary files (S1–S3) are provided for anonymous download in

184 an online supporting archive hosted at www.palaeogrimm.org/data/Smn15_OSA.zip

185

186 **Results**

187

188 *Levels of intra- and interspecies plastome divergence in Quercus*

189 The entire dataset included 423 plastid DNA sequences (141 samples, three markers each). Table
190 2 shows that *trnH-psbA* was the most variable marker region (a 34-bp inversion occurring in
191 approximately 50% of the samples was not considered). The least variable region was *rbcL*, as
192 expected. No indels were found in the *rbcL* and *matK* coding regions. The combined cpDNA
193 dataset (*trnH-psbA*, *trnK/matK*, *rbcL*) resulted in an alignment of 2082 characters (sites), of
194 which 122 were variable (thereof 72 parsimony-informative; gaps not considered). The alignment
195 had a nucleotide diversity of 0.0056 and included 74 different haplotypes of which 50 were
196 unique (restricted to a single accession). As a result, the overall haplotype diversity was high (Hd
197 = 0.978 ± 0.005). With gaps considered, the number of haplotypes increased to 110, of which 89
198 were unique ($Hd = 0.9939$).

199 In general, the infrageneric divergence calculated in *Quercus* is comparable to that found in other
200 genera of the Fagaceae and Betulaceae, and higher than in Juglandaceae (Table 3). All three gene
201 regions allow distinguishing the generic affinity of an oak individual; the same haplotype may be
202 shared by several or many oak species (usually within the same infrageneric group; Table 3), but
203 not with other genera of the Fagaceae.

204 At the infrageneric level in *Quercus*, minimal inter-species distances can be zero for all three
205 markers and within all infrageneric groups. Notably, maximal inter-species distances within
206 infrageneric groups of *Quercus* can reach or even exceed the level of inter-generic differentiation
207 in Fagaceae (e.g. between *Notholithocarpus*, *Lithocarpus*, *Castanopsis*, *Castanea*, *Chrysolepis*),
208 Juglandaceae and Myricaceae. The maximum intra-specific distance found in Mediterranean

209 individuals of *Quercus* Group Ilex equals the maximum inter-specific divergence found within
210 this group.

211

212 *Phylogenetic placement of Mediterranean Quercus Group Ilex plastid haplotypes*

213 Individuals of the Mediterranean species of *Quercus* Group Ilex cluster in three well supported
214 distinct clades (Fig. 1). The first clade ('Euro-Med') accommodates most accessions of *Q. ilex*
215 and *Q. coccifera*. In the second clade ('Cerris-Ilex'), accessions of *Q. ilex*, *Q. coccifera*, and one
216 of the five samples of *Q. aucherii* group together with all representatives of *Quercus* Group
217 Cerris and two Himalayan-East Asian species of Group Ilex. Sister to this clade are the three
218 representatives of the single Japanese species of Group Ilex (*Q. phillyraeoides*). In the third clade
219 (West Asia-Himalaya-East Asia; 'WAHEA') the remaining specimens of *Q. aucherii* form a
220 subclade along with the Cypriote endemic *Q. alnifolia*, and several Eastern Mediterranean *Q.*
221 *coccifera*. The second, more divergent and poorly supported subclade comprises two western
222 Himalayan species (*Q. baloot*, *Q. floribunda*), two individuals of Himalayan-East Asian species
223 of *Quercus* Group Ilex, and one Central China accession of a Cyclobalanopsis member (*Q.*
224 *oxyodon*) sympatric with many group Ilex oaks, including *Q. semecarpifolia*, *Q.*
225 *leucotrichophora*, *Q. floribunda* (Menitsky, 2005). In contrast to Group Ilex, all other
226 infrageneric groups show relatively high chlorotypic coherence, usually forming clades or
227 grouped within the same subtree. The actual root of the tree is obscured; representatives of
228 *Castanea*, *Castanopsis*, and *Notholithocarpus/Chrysolepis* that could be used as putative
229 outgroups are placed in different subtrees.

230

231 *Evolutionary significance of plastid haplotypes in western Mediterranean oaks of Quercus*
232 *Group Ilex*

233 The MJ network for the plastid region with the highest overall variability (*trnH-psbA*, only
234 length-homogenous parts considered; Fig. 2) highlights the evolutionary significance of the three
235 main haplotypes, ‘Euro-Med’, ‘Cerris-Ilex’, and ‘WAHEA’. Three main clusters differ by a
236 minimum of two conserved mutations: 1) Group *Quercus*, *Protobalanus* and *Lobatae* (‘New
237 World Oaks’); 2) individuals with ‘Euro-Med’ haplotypes; 3) individuals with ‘Cerris-Ilex’ and
238 ‘WAHEA’ haplotypes, representatives of Group *Cerris* and East Asian species of Group *Ilex* and
239 Group *Cyclobalanopsis* (‘Old World Oaks’). In general, haplotypes (File S3 includes MJ-
240 networks for the other three regions, *rbcL* gene, *matK* gene, 3’ *trnK* intron,) found in the western
241 Eurasian members of Group *Ilex* represent unique or ancestral variants. Unique haplotypes of
242 Group *Cerris* are directly derived from the Group *Ilex* or shared ‘Cerris-Ilex’ haplotypes.
243 Haplotypes of Group *Cyclobalanopsis* are identical to or can be derived from East Asian
244 members of Group *Ilex*. The graphs further highlight a close relationship of haplotypes of
245 *Chrysolepis* and *Notholithocarpus* with those of the ‘New World’ oaks; those of *Castanea* and
246 *Castanopsis* can be derived from the ‘Old World’ oaks basic type.

247 Figures 1 and 2 clearly illustrate that differentiation in the plastid sequences of *Quercus* (and
248 related *Fagaceae*) is independent from the formation or at least the genetic homogenization
249 (lineage sorting) of the modern clades.

250

251 *Phylogeographic structure in Quercus Group Ilex*

252 Haplotypes forming the ‘Euro-Med’, ‘Cerris-Ilex’ and ‘WAHEA’ lineages are geographically
253 sorted. The phylogenetically isolated ‘Euro-Med’ haplotypes are encountered in the western

254 Mediterranean populations of *Q. ilex* and *Q. coccifera* (North Africa, Iberia, Southern France,
255 Italy), along the Adriatic coast and into Central Greece (Fig. 3). Also included here are isolated
256 populations of *Q. ilex* from Crete and the southern Black Sea coast. ‘Cerris-Ilex’ and ‘WAHEA’
257 haplotypes are confined to the eastern Mediterranean region. ‘Cerris-Ilex’ haplotypes are found
258 in the Aegean region (*Q. ilex*, *Q. coccifera* and *Q. aucheri* individuals) and replaced by
259 ‘WAHEA’ haplotypes (*Q. coccifera*, *Q. aucheri*, *Q. alnifolia*) in south-western Turkey and
260 extending to the east (Levant; Fig. 3). The ‘Cerris-Ilex’ type is also found in the *Q. coccifera*
261 individual from northern Turkey, representing the north-easternmost population of this species.

262

263 Discussion

264

265 Despite resolution issues due to weak signals regarding intergeneric relationships, all data on
266 Fagaceae show a deep incongruence between nuclear and plastid data. Nuclear phylogenies
267 unambiguously point towards an inclusive common origin of *all* oaks, i.e. a monophyletic (s.
268 str.) genus *Quercus* (Oh and Manos, 2008; Denk and Grimm, 2010; Hubert et al., 2014). At the
269 same time plastid data repeatedly failed to resolve all oaks as one clade (Manos et al., 2008; this
270 study). Instead, a split emerges (with varying support) between the North American
271 *Notholithocarpus* and North American/northern temperate clade of oaks, the 'New World Oaks',
272 and the Eurasian *Castanea*, *Castanopsis* and oak lineages, the 'Old World oak' clade; an
273 observation that holds independent from the exact placement of the root in a plastid tree. If we
274 accept the monophyly of the genus *Quercus*, which is backed also by morphology and evidence
275 from the fossil record, haplotypes of *Castanea/Castanopsis* and *Notholithocarpus* that group
276 with the 'New World' and 'Old World' oaks, respectively, can hence only be the result of

277 incomplete lineage sorting during the formation of the modern genera. In addition, the plastid
278 genepool of the earliest oaks must have shown a genetic gradient that was to some extent caused
279 by biogeographic patterns. Although it is impossible to pinpoint the place of origin of oaks, it is
280 clear that the ancient oaks must have been widespread, allowing them to pick up and propagate
281 geographic signatures inherited from their common ancestors with *Notholithocarpus*, *Castanea*
282 and *Castanopsis*. Geographic signatures in two Mediterranean species of Group Ilex, *Q. ilex* and
283 *Q. coccifera*, are discussed in the following.

284

285 *Major trends of plastome differentiation*

286 The overall low genetic intra- and intertaxonomic (intrageneric lineages, genera) distances
287 suggest low evolutionary rates for the chloroplast genomes of Fagales, at least at the examined
288 loci. However, the data coverage is far from sufficient for most genera and families to precisely
289 evaluate the plastome potential variation within this plant group. In Fagaceae, a comparison with
290 the (genetically) more diverse Nothofagaceae and Betulaceae families reveals that haplotype
291 variation at the *trnH-psbA* locus can be sufficiently high to allow inferences at the
292 phylogeographic and systematic level (see Premoli et al., 2012; Grimm and Renner, 2013). In
293 analogy, haplotypes of intrageneric lineages of *Quercus* differ in this marker. Furthermore, a
294 geographic pattern is evident for the most widely sampled groups. Groups Ilex, Lobatae and
295 *Quercus* appeared the most variable, whereas Group Cerris exhibited the lowest differentiation
296 rates. Interestingly, interclade differentiation among all *Quercus* groups equalled or exceeded the
297 values scored by the four outgroup genera (*Castanea*, *Castanopsis*, *Notholithocarpus* and
298 *Chrysolepis*). As a consequence, the outgroup taxa appear scattered across the tree, rather than
299 being culled in a distinct subtree, rendering the plastome of *Quercus* ‘non-monophyletic’.

300 Outgroup selection as a potential source of topological ambiguity was already pointed out by
301 Hubert et al. (2014; 108 oak taxa, eight nuclear markers). Ambiguous relationships among this
302 group of genera independently of the strength of the obtained phylogenetic signal were also
303 suggested by a recent study on Fagales (based on molecular, fossil and reproductive syndromes
304 analyses), which resolved the majority of inter-generic relationships in each family except in the
305 Quercoideae group making *Castanopsis* and *Quercus* non-monophyletic (Xiang et al., 2014).

306

307 *Plastid phylogeny does not conform to the current synopsis of oak evolution*

308 Figure 4 highlights the incongruence of the plastid genealogy tree with the current understanding
309 of the evolution of Fagaceae and oaks based on molecular sequence data from the non-coding
310 nuclear gene regions (Manos et al., 2008; Denk and Grimm, 2010), a recent time-calibrated
311 nuclear phylogeny of oaks (Hubert et al. 2014), and the fossil record of modern lineages as
312 documented by pollen investigated under the scanning-electron microscope (Grímsson et al.,
313 2015; see also Denk and Grimm, 2009). Two evolutionary mechanisms (incomplete lineage
314 sorting, reticulation) may account for the observed, highly complex pattern.

315 Firstly, speciation processes in *Quercus* do not immediately leave imprints on the plastome (e.g.
316 Neophytou et al., 2010; Cavender-Bares et al., 2011) as also well documented for *Nothofagus*
317 (Acosta and Premoli, 2010; Premoli et al., 2012). Low mutation rate and long generation time
318 can contribute to slow evolutionary rates and incomplete lineage sorting of organellar genomes
319 (Cavender-Bares et al., 2015; Besnard et al., 2007). In addition, reiterated extinctions and re-
320 colonisations involving bottlenecks, genetic drift, and founder effects may cause random fixation
321 of haplotypes, increasing the probability for retaining ancestral traits. Oaks in general, and
322 especially the Mediterranean taxa, are also characterised by a marked resprouting ability in

323 response to disturbances of varying frequencies and intensities, including the action of man,
324 herds, and wild fires (Barbero et al., 1990). This could also have contributed to clonally preserve
325 and transmit ancestral plastid lineages (maternally inherited) during multiple and reiterated
326 unfavourable conditions since the origin of the Mediterranean region (Blondel and Aronson,
327 1999). At the same time, different environmental selection and adaptation, large population sizes,
328 and long distance pollen dispersal, might have homogenised the nuclear genomes in local
329 populations of a species but not their organelle genomes.

330 Secondly, Fagaceae lineages are susceptible to hybridisation and introgression (Arnold, 2006).
331 This may lead to the formation of morphologically unambiguous individuals of a species with
332 plastid signatures of another (Whittemore and Schaal, 1991; Petit et al., 2004). There is
333 increasing evidence for local introgression in oak communities with morphologically distinct
334 species in the case of European white oaks (Group Quercus; *Q. robur*, *Q. petraea*, *Q. pyrenaica*,
335 *Q. pubescens*, *Q. frainetto*; Curtu et al., 2007; Valbuena-Carabaña et al., 2007; Lepais et al.,
336 2009), as well as in members of *Quercus* subsection *Virentes*, a subgroup of Group Quercus, in
337 North America (Cavender-Bares et al., 2015), and across a wide range of Group Lobatae (Dodd
338 and Afzal-Rafii, 2004; Peñaloza-Ramírez et al., 2010; Moran et al., 2012; Valencia-Cuevas et al.,
339 2015). In our focal group, hybrids and different levels of genetic introgression among
340 morphologically pure individuals were molecularly documented in *Q. ilex*/*Q. coccifera* (Ortego
341 and Bonal, 2010) and, to a lesser extent, in *Q. coccifera*/*Q. alnifolia* (Neophytou et al., 2011).
342 Also, the potential for inter-group hybridisation was experimentally demonstrated for *Q. ilex* and
343 *Q. robur* (Group Quercus; Schnitzler et al., 2004), and natural introgression in *Q. ilex*/*Q. suber*
344 was identified in Southern France (Mir et al., 2009) and Iberia (Burgarella et al., 2009).
345 Therefore, it is possible that ancient hybridization and introgression, favoured by the well-known

346 sexual promiscuity between closely related taxa and their ability to disperse pollen over long
347 distances, obscure the true evolutionary origin of an oak species or entire lineage.
348 In the Mediterranean, the dramatic geological and ecological changes during the Neogene
349 (Blondel and Aronson, 1999) likely caused extinction, re-colonisation, range fragmentation and
350 hybridisation linked to secondary contact, especially when species were still young and
351 reproductive barriers likely weaker than today. Taken together, incomplete sorting of ancestral
352 traits and introgression of haplotypes thus appear highly likely mechanisms to decrease inter-
353 species plastid differentiation while at the same time increasing intra-species variation. This
354 provides an explanation why the widespread, morphologically and genetically (at the nuclear
355 level) distinct species *Q. coccifera* and *Q. ilex* have accumulated three distinct main plastid
356 haplotypes, which we discuss in the following.

357

358 *Polyphyletic clues in Quercus Group Ilex*

359 The most striking finding of this study is the plastid polyphyly and a clear geographic pattern
360 displayed by a group of Mediterranean Ilex oaks (*Q. ilex*, *Q. coccifera* and *Q. aucheri*).
361 Phylogeographic patterns reflecting distant vicariant events and a complex history of range
362 expansions and contractions have been previously inferred for other Mediterranean woody
363 species (Besnard et al., 2007; Desamore et al., 2011; Migliore et al., 2012; Chen et al., 2014).
364 Mediterranean *Laurus* (Rodriguez-Sanchez et al., 2009), for instance, also comprises three
365 plastid haplotype lineages roughly corresponding to biogeographic patterns as seen in the
366 Mediterranean Ilex oaks: (1) an eastern lineage in Turkey and the Near East, (2) a second one in
367 the Aegean region, and (3) a probably ancestral lineage of central and western Mediterranean
368 populations. The importance of the Mediterranean basin in shaping the intraspecific divergence

369 of Tertiary plant species is therefore well-acknowledged (Nieto Feliner, 2014). Unfortunately,
370 none of the above studies investigated multiple species, leaving the question whether the
371 observed divergence is restricted to a single species or shared by several species as in the case of
372 *Quercus* Group Ilex.

373 Extensive sampling is more likely to document hidden polyphyly (Wiens and Servedio, 2000);
374 besides a weak phylogenetic signal resulting in poorly resolved and potentially inaccurate gene
375 trees (countered here by using median-joining and bipartition networks), incomplete sorting of
376 ancestral lineages and introgression represent very general sources of polyphyly. However,
377 distinguishing the separate effects of these mechanisms can be very difficult in the absence of
378 nuclear markers and (palaeo-)geography as complementing information (Funk and Omland,
379 2003). As a general rule, the phylogenetically basal position of polyphyly usually hints at
380 retained ancestral polymorphism, while recently introgressed haplotypes may assume a highly
381 derived position in a gene tree. At the same time, incomplete sorting is not predicted to promote
382 the geographic proximity of interspecifically shared haplotypes that may be seen under local
383 introgression (Hare and Avise, 1998, Masta et al., 2002).

384 In our reconstruction, Group Ilex oaks appear to conform to both instances. Nuclear data
385 covering the entire range of *Q. ilex* and *Q. coccifera* in the Mediterranean region unambiguously
386 resolved the two species as close, but mutually monophyletic sister taxa (Denk and Grimm,
387 2010). In the absence of nucleome data for all here included individuals, it is impossible to infer
388 to which degree introgression and incomplete lineage sorting contributed to the plastid gene pool
389 of the Mediterranean species of Group Ilex. Nevertheless, the most straightforward explanation
390 for the observed scenario would be a combined effect of the two mechanisms: asymmetrical
391 introgression of ancestral haplotypes resulting in local genetic clusters decoupled from

392 taxonomic boundaries, in which plastome accessions of species or species complexes may form
393 grades or multiple clades in phylogenetic trees, thus appearing polyphyletic (e.g. Rieseberg and
394 Soltis, 1991; Whittmore and Schaal, 1991).

395 As modelled by Excoffier et al. (2009), interspecific interactions during historical range
396 fluctuations can profoundly affect the observed phylogeographic patterns, and manifest as
397 paraphyly or reticulation. In fact, most range expansions do not occur in completely uninhabited
398 areas, and interbreeding between local and an expanding (invasive) species with subsequent
399 asymmetrical introgression can develop also in absence of selection (Lepais et al., 2009). Plastid
400 haplotypes referring to the original ('lost') species are indeed likely to persist over long
401 evolutionary periods, and may still be found in the invading species. Noteworthy, environmental
402 changes and disturbance of local communities have been shown to increase hybridisation rates
403 (Lagache et al., 2013), hence, the potential for widespread, imbalanced introgression. In Group
404 *Ilex* oaks, the interspecific capture of plastids among sexually incompletely isolated species
405 likely occurred on the geological timescale, concealing the species relationships at various stages
406 in the history of the genus. In a comprehensive study of the genus *Ilex* (Manen et al., 2010), the
407 high incongruence between a taxonomically compatible nuclear gene tree and a geographically
408 structured plastid tree was explained with extensive extinctions between the Cretaceous and
409 Miocene and multiple hybridization and introgression events between distantly related lineages.
410 This has been documented also for *Platanus* (Grimm and Denk, 2010) and more recently
411 suggested for the evergreen white oaks of *Quercus* subsection *Virentes* (Eaton et al., 2015).
412 Similar ancient lateral transfers have been also inferred to explain the paraphyly of the
413 maternally inherited mtDNA of *Picea* (Bouillè et al., 2011) and *Pinus* (Tsutsui et al., 2009). As
414 noted above, introgression has been widely demonstrated across a wide range of oaks (Dodd and

415 Afzal-Rafii, 2004; Curtu et al., 2007; Valbuena-Carabaña et al., 2007; Burgarella et al., 2009;
416 Lepais et al., 2009; Mir et al., 2009; Ortego and Bonal, 2010; Peñaloza-Ramírez et al., 2010;
417 Neophytou et al., 2011; Moran et al., 2012; Valencia-Cuevas et al., 2015) and suggested as
418 explanation for the wide haplotype sharing revealed by *Q. suber* and *Q. cerris* in the Italian
419 peninsula (Magri et al., 2007).

420

421 *Temporal and spatial framework of plastome evolution*

422 The three distinct plastid haplotypes observed in modern Mediterranean members of *Quercus*
423 Group Ilex may reflect three radiation phases (range extensions), followed by range disruptions
424 and isolation of plastome lineages within the ‘Old World Clade’ of *Quercus*. Considering the
425 high diversity of haplotypes in Group Ilex as compared to other major oak lineages (or other
426 genera in the Fagales; see Table 3; Table S2) it can be assumed that the geographical disruptions
427 in the plastome of the ancestors of Group Ilex and interacting lineages predate the manifestation
428 of modern taxa (species and infrageneric groups; Fig. 1). Haplotypes shared between members of
429 Group Ilex and its sister lineages Group Cerris and Group Cyclobalanopsis may indicate
430 common (geographic) origin or may be the result of secondary contact and unidirectional
431 introgression.

432 Independent from the position of the plastid root, the divergence of the ‘Euro-Med’ haplotype
433 must have coincided with the initial differentiation in *Quercus* (Fig. 1). Oaks had achieved a
434 wide northern hemispheric range by the Eocene. Unequivocal fossils are known from high
435 latitudes (North America, Greenland, North Europe; Crepet and Nixon, 1989; Manchester, 1994;
436 Grímsson et al., 2015) and mid latitudes (Central Europe, South East Asia; Kvaček and Walther,
437 1989; Hofmann, 2010). All major lineages of oaks were established by the end of the Eocene, ca.

438 35 Ma, as evidenced by the fossil record and molecular dating using eight nuclear gene regions
439 (Bouchal et al., 2014: fig. 14; Hubert et al., 2014; Grímsson et al., 2015). During this time, one
440 fraction of oaks must have been geographically and reproductively isolated which would have
441 caused a major split in the plastid gene pool (Fig. 1). Today, the ‘Euro-Med’ haplotype is the
442 only one exclusively shared by just two, but widespread Mediterranean species of *Quercus*
443 Group Ilex, *Q. ilex* and *Q. coccifera*. Two evolutionary scenarios can explain the establishment
444 of this haplotype in *Q. ilex-Q. coccifera* (Fig. 5): (i) The ‘Euro-Med’ haplotype is the remnant of
445 an extinct oak lineage that was intrograded (invaded) and consumed by members of Group Ilex;
446 under this scenario Group Ilex would have migrated into Europe at some point prior to the
447 Miocene where it came into contact with this extinct oak lineage. (ii) The ‘Euro-Med’ haplotype
448 represents the original plastome of Group Ilex; under this scenario, the first split within the
449 modern ‘Old World clade’ would have been between a western Group Ilex and an eastern Group
450 Cyclobalanopsis (Fig. 5). Fossil evidence and available phylogenies (discussed in the following)
451 lend high credibility to scenario (i) as the most plausible explanation.

452 The ‘Cerris-Ilex’ haplotype is shared between all species of *Quercus* Group Cerris (western
453 Eurasian and East Asian), East Mediterranean (Aegean) individuals and two East Asian species
454 of Group Ilex. This is in agreement with Denk and Grimm (2009) who suggested that *Quercus*
455 Group Cerris evolved from Group Ilex by budding (a hypothesis further confirmed by the 8-
456 nuclear gene data set used by Hubert et al., 2014), and the low support for a Group Ilex clade in
457 an all-Fagaceae (excluding *Fagus*) tree based on over 1000 nuclear ITS sequences (Denk and
458 Grimm, 2010). Hubert et al. (2014) inferred a Miocene age for this budding event, which
459 corresponds to the earliest unequivocal fossil of *Quercus* Group Cerris (Kmenta, 2011) and is
460 younger than the earliest definite fossil record of *Quercus* Group Ilex in Europe (early

461 Oligocene, Cospuden; Denk et al., 2012). Also, dispersed pollen from the Paleogene
462 Changchang Formation, Hainan (Hofmann, 2010), resembles both *Quercus* Group Ilex and
463 Group Cyclobalanopsis; the age of this formation is considered late early to early late Eocene
464 (Lei et al., 1992). The most closely related haplotype to the ‘Cerris-Ilex’ haplotype is
465 encountered in the widespread East Asian *Q. phillyraeoides*, the only species of Group Ilex
466 extending to Japan (the East Asian members of Group Cerris have a much wider range in north-
467 eastern Asia; Menitsky, 2005). Regarding its phylogenetic position, the emergence of the
468 ‘Cerris-Ilex’ haplotype appears linked with a major taxonomic sorting event in Eurasian
469 Fagaceae, resulting in distinct haplotypes restricted to genera and intrageneric groups of *Quercus*
470 (Fig. 1). Based on the palaeobotanical record, these lineages (*Castanopsis*, *Castanea*, *Quercus*
471 Group Ilex, *Quercus* Group Cyclobalanopsis) were well established at least by the Eocene (Table
472 4, Fig. 5; Grímsson et al., 2015); a deep divergence is reflected by their distinctly different
473 nuclear genomes (Oh and Manos, 2008; Denk and Grimm, 2010; Hubert et al., 2014). Two
474 evolutionary scenarios can explain the occurrence of the ‘Cerris-Ilex’ haplotype in Aegean
475 individuals of *Q. ilex* and *Q. coccifera* and the westernmost *Q. aucheri*: (i) Group Cerris evolved
476 in western Eurasia/Himalaya from an (extinct) subtropical to temperate sublineage of Group Ilex,
477 which left its imprint in the Aegean members of Group Ilex, and *Q. spinosa*, *Q. engleriana* and
478 *Q. phillyraeoides*; (ii) Group Cerris shares a common ancestry with the north-east Asian *Q.*
479 *phillyraeoides*. Under this scenario, the budding event of the group took place in north-eastern
480 Asia, from where it migrated into western Eurasia and the Aegean region; in relatively recent
481 times, Group Cerris came into contact with the Mediterranean members of Group Ilex and were
482 locally intrograded.

483 The high similarity of ‘Cerris-Ilex’ haplotypes lends some credibility to the second scenario.
484 Furthermore, there is evidence for current introgression and occasional hybridization of *Q. suber*
485 (Group Cerris) and *Q. ilex* in the western Mediterranean (Burgarella et al., 2009; Mir et al.,
486 2009). However, it is difficult to explain why *Q. ilex-coccifera* should only intrograde into
487 populations of Cerris oaks at a large scale in the Aegean region. Today, Group Cerris is more
488 diverse than Group Ilex in the East Mediterranean (*Q. brantii*, *Q. cerris*, *Q. ithaburensis*, *Q.*
489 *macrolepis*, *Q. libani*, *Q. trojana*) with some species adapted to distinctly continental climates
490 (Browicz and Zieliński, 1982; Menitsky, 2005), outside the range of Group Ilex. This diversity
491 and the vast distribution of only two species of Group Cerris in East Asia may point towards a
492 young radiation of the group. During the early and middle Miocene, Cerris did not play an
493 important role in western Eurasia. In contrast, two distinct species complexes of *Quercus* Group
494 Ilex were prominently represented in eastern Mediterranean and Paratethyan plant assemblages,
495 *Q. drymeja* Unger and *Q. mediterranea* Unger (e.g. Velitzelos et al., 2014). Intriguingly, the *Q.*
496 *drymeja* complex includes morphotypes found today in *Q. ilex* as part of its intraspecific
497 variation, and of a range of East Asian species including *Q. engleriana*. *Quercus mediterranea* is
498 the morphological equivalent of *Q. ilex*, *Q. coccifera* and a range of East Asian species including
499 *Q. spinosa*. Hence, the fossil record clearly favours a western Eurasian-Himalayan origin of
500 Group Cerris (scenario i).

501 The West Asian-Himalayan-East Asian (WAHEA) haplotype represents Eastern Mediterranean
502 members of *Quercus* Group Ilex and is sister to a clade comprising several Asian species of
503 Group Ilex (Himalayas to the mountains of Southeast Asia). It reflects the second radiation
504 within the Old World Clade and allies after the isolation of the ‘Euro-Med’ original lineage and
505 prior to the radiation and subsequent sorting within the clade comprising the Cerris-Ilex

506 haplotype (Fig. 1). The modern distribution of species with the WAHEA haplotype follows the
507 Himalayan corridor (Kitamura, 1955; Güner and Denk, 2012). The Himalayan corridor is a
508 narrow band along the southern flanks of the Himalaya with a subtropical to temperate climate
509 (*Cwa*, *Cwb*; Peel et al. 2007) providing a refuge for plants that were more widespread before the
510 Himalayan uplift. In addition to *Quercus* Group Ilex (Zhou, 1992; Velitzelos et al., 2014),
511 prominent relic taxa include species of *Acer*, *Aesculus*, *Cedrus*, *Cotinus*, *Juglans*, *Platanus*, and
512 *Rhododendron* among others. The ‘WAHEA’ haplotype represents the western counterpart to the
513 haplotype lineage found in East Asian species of Group Ilex and *Cyclobalanopsis*. The relic *Q.*
514 *alnifolia*, today preserved only in the mid-montane region of Cyprus (Mt. Troodos), would be a
515 witness of this expansion (Menitsky, 2005).

516

517 *Towards an integrated biogeography of oaks*

518 Plant biogeographic studies at the genus level have commonly relied on few to many chloroplast
519 markers and a single or very few accessions per taxon. In the case of woody angiosperms with a
520 subtropical to temperate distribution such as for example Nothofagaceae (Svenson et al., 2001;
521 Knapp et al., 2005), *Rhus* (Yi et al., 2004), *Cornus* (Xiang et al., 2005), *Carpinus* (Yoo and Wen,
522 2007), *Castanea* (Lang et al., 2007), *Juglans* (Aradhya et al. 2007), and *Carya* (Zhang et al.
523 2013a), such an approach runs the risk of capturing but a limited aspect of the evolutionary
524 history of the focal group. Mere combination with e.g. nuclear ITS data can be problematic, too
525 (compare data shown here with data provided by Denk and Grimm, 2010, on western Eurasian
526 members of Group Ilex). The decoupled evolutionary signals in plastomes and the
527 nucleome/morphology as documented for *Nothofagus* (Acosta and Premoli, 2010; Premoli et al.,
528 2012) and *Quercus* Group Ilex (this study) suggests that the traditional placeholder sampling

529 strategy is not ideal. Signals from few-marker/many-samples data sets are likely to be complex
530 or even puzzling (Figs 1–4), but at the same time provide entirely new perspectives on plant
531 evolution worth exploring. For *Quercus* Group Ilex, our pilot study focussing on Mediterranean
532 species reveals a crucial aspect of oak evolution not seen in the combined nuclear,
533 morphological, and fossil data: large-scale introgression and incomplete lineage sorting among
534 ancestral lineages of modern major groups and species. The new data corroborate hypotheses
535 that Group Cerris evolved (‘budded’) relatively recent from Group Ilex (over 600 ITS and over
536 900 5S-IGS accessions covering all western Eurasian oak species, Denk and Grimm, 2010;
537 signal from six single-copy nuclear regions, Hubert et al., 2014). *Quercus* Group Cerris probably
538 evolved in western Eurasia and the Himalayas when the then chiefly subtropical low latitude
539 Group Ilex radiated into temperate niches. Within modern members of Group Cerris, a wide
540 spectrum of leaf traits is found from pseudo-evergreen in *Q. suber*, to semi-evergreen in *Q.*
541 *brantii*, *Q. ithaburensis*, *Q. trojana* (partly) and fully deciduous in *Q. acutissima*, *Q.*
542 *castaneifolia*, *Q. cerris*, *Q. libani* and *Q. variabilis*. The conspicuous plastid diversity in the
543 Mediterranean species of Group Ilex and the lineage in general (Figs 1, 2; Table 2) reflects the
544 highly complex geographical history of this group. The ‘Euro-Med’ haplotype evidences an
545 initial phase of west-east differentiation in low-latitude Eurasian oaks, the ‘Old World Clade’,
546 probably triggered by the complex topography within its potential range essentially since the
547 Eocene (Fig. 5). During its evolution, Group Ilex was continuously affected by range disruptions
548 caused by tectonic activity south of the Paratethys linked to the collision of Africa and the Indian
549 subcontinent with Eurasia (Fig. 6); progressive rarefaction of the original haplotypes and the
550 occurrence of (repeated) invasion and introgression events that left imprints in the plastome even
551 within the same species is highly likely.

552 Although decoupled from taxonomy, the plastid phylogeny provides important, independent
553 information on the geographic differentiation of *Quercus* prior to the formation of modern
554 species/species groups. The major split within oaks has traditionally been considered between
555 ‘New World’ and ‘Old World’ oaks (Manos and Stanford, 2001) because of the current
556 distribution of the major lineages of oaks. The plastid data presented here strongly suggest that
557 the early evolution of oaks instead was geographically bound to high latitude Arctic regions and
558 to low latitude subtropical regions (Fig. 5). The high latitude lineages remained genetically
559 homogeneous in the nucleome, but also in the plastome to some degree. Continuous circum-
560 polar distribution prevented pronounced genetic drift in the high latitude lineage, which became
561 the ‘New World Clade’, and explains low genetic differentiation in deciduous high and mid
562 latitude white oaks until today (Denk and Grimm, 2010). At the same time, the Atlantic, the
563 proto-Mediterranean, and the Paratethys isolated the Eurasian low latitude lineage.

564 Our data should only be viewed as a first step towards a more complete understanding of the
565 biogeography and evolution of oaks. The next step would be to map the plastid variation of
566 *Quercus* Group Ilex across its entire range by sampling multiple stands of the Himalayan and
567 East Asian species to characterise the geographic and taxonomic ranges of the various plastid
568 lineages.

569

570 **Conclusion**

571

572 Taken all evidence together, the first major split of oaks, consistently found in all molecular
573 phylogenies, would have been into two clades. A northern, high-latitude clade leading to white,
574 red and golden-cup oaks, which evolved and diversified in the tectonically quiet parts of the

575 Northern Hemisphere. Its counterpart was a southern, mid-latitude clade made up by Group Ilex
576 and Group Cyclobalanopsis (and later by Group Cerris), in the southern part of Eurasia, and
577 perhaps western North America. Both *Quercus* Group Cyclobalanopsis and Group Ilex were
578 present in southern Eurasia, close to the shores of the Tethys, and western North America by the
579 middle Eocene. The outlined history of further steps in the southern clades appear to consist of
580 two radiations: one lead to the Group Cerris clade (early Miocene) with migration westwards
581 along the southern slopes of the Himalayas, and eastwards over China and Japan. The highly
582 coherent ‘Cerris-Ilex’ haplotypes are key witnesses of this event indicating that westwards and
583 eastwards migration and radiation of the monophyletic Group Cerris may have been relatively
584 recent. A putative radiation centre is the East-Mediterranean Paratethys region, in which
585 members of Group Ilex and Cerris coexist *and* share highly similar to identical haplotypes. The
586 second radiation is likely to have occurred in the Miocene, too. From a Himalayan Group Ilex
587 stock, the clade with ‘WAHEA’ haplotype expanded towards the eastern Mediterranean basin.
588 Both the ancient western Eurasian clade, now extinct but evidenced by the ‘Euro-Med’
589 haplotype, and the originally Himalayan clade had been invaded by the late Neogene by the
590 direct ancestors of today’s *Q. coccifera* and *Q. ilex*. Modern forms of these two Mediterranean
591 oaks (and their two closest relatives, *Q. aucheri* and *Q. alnifolia*) would have preserved the
592 ancestral haplotypes, keeping strong geographic indications of those events along with the
593 formation of the Mediterranean region. Moreover, the Himalayan uplift coincided with the
594 development of the modern monsoon climate; hence adaptations or exaptation to phases of
595 drought may have played an important role at least since 15 million years ago (Wang and Wu
596 2015). Our reconstruction is still speculative, but consistent with (i) all fossil observations, (ii)
597 known molecular phylogenies of plastid haplotypes, and (iii) known processes shaping the

598 diversity of plastid haplotypes by speciation and introgression, linked with invasions. Further
599 detailed studies, especially in the Himalayan and Chinese regions, are needed to validate or
600 disprove this scenario.

601 Because of similar strong correlation between plastome differentiation and geographic
602 distribution at the population level and the species/genus level, similar or identical plastid
603 haplotypes typically shared between co-occurring and often distantly related taxa, polyphyletic
604 signals and reproductive biology, the same processes could have likely played a key role in the
605 evolutionary history of other Fagaceae (e.g., *Fagus*, *Castanea*, *Castanopsis*). Broadening the
606 sampling efforts of phylogenetic analyses of the plastome could help decipher the speciation
607 history of these genera. At the same time, extended nucleome investigations will be obviously
608 necessary to definitely assess a clear molecular phylogeny of Fagaceae.

609 **Acknowledgments**

610 We thank all the Directors and curators of the Botanic Gardens and Arboreta who provided the
611 investigated material: Anthony S. Aiello, Wolfgang Bopp, Anne Boscawen, Peter Brownless,
612 Béatrice Chassé, Laszlo Csiba, Dirk De Meyere, Holly Forbes, Anett Krämer, Isabel Larridon,
613 Gitte Petersen, David Scherberich, Patrick Thompson, Jef Van Meulder, Michael Wall, David
614 Zuckerman, and all the colleagues and friends who contributed to the sampling design: Jeannine
615 Cavender-Bares, Hanno Schaefer, Charalambos Neophytou, Martina Temunovic, Gianni Bedini,
616 Laura Genco, Enara Otaegi Veslin.

617

618

619

620 **References**

- 621 **Acosta MC, Premoli AC.** 2010. Evidence of chloroplast capture in South American *Nothofagus*
622 (subgenus *Nothofagus*, Nothofagaceae). *Molecular Phylogenetics and Evolution* **54**: 235–
623 242.
- 624 **Akaike H.** 1974. A new look at the statistical model identification. *IEEE Transactions on*
625 *Automatic Control* **19**: 716–723.
- 626 **Aradhya MK, Potter D, Gao F, Simon CJ.** 2007. Molecular phylogeny of *Juglans*
627 (Juglandaceae): a biogeographic perspective. *Tree Genetics and Genomes* **3**: 363–378.
- 628 **Arnold ML** (2006) *Evolution through genetic exchange*, vol 3. Oxford: Oxford University Press.
- 629 **Barbero M, Bonin G, Loisel R, Quezel P.** 1990. Changes and Disturbances of Forest
630 Ecosystems Caused by Human Activities in the Western Part of the Mediterranean Basin.
631 *Vegetatio* **87** : 151-173.
- 632 **Belahbib N, Pemonge MH, Ouassou A, Sbay H, Kremer A, Petit R.** 2001. Frequent
633 cytoplasmic exchanges between oak species that are not closely related: *Quercus suber*
634 and *Q. ilex* in Morocco. *Molecular Ecology* **10**: 2003-2012.
- 635 **Besnard G, Rubio de Casas R, Vargars P.** 2007. Plastid and nuclear DNA polymorphism
636 reveals historical processes of isolation and reticulation in the olive tree complex (*Olea*
637 *europaea*). *Journal of Biogeography* **34**: 736-752.
- 638 **Blondel J, Aronson J.** 1999. *Biology and wildlife of the Mediterranean region*. Oxford: Oxford
639 University Press.
- 640 **Bouchal J, Zetter R, Grímsson F, Denk T.** 2014. Evolutionary trends and ecological
641 differentiation in early Cenozoic Fagaceae of western North America. *American Journal*
642 *of Botany* **101**: 1–18.

- 643 **Bouillé M, Senneville S, Bousquet J.** 2011. Discordant mtDNA and cpDNA phylogenies
644 indicate geographic speciation and reticulation as driving factors for the diversification of
645 the genus *Picea*. *Tree Genetics and Genomes* 7: 469–484.
- 646 **Browicz K, Zieliński J.** 1982. *Chorology of trees and shrubs in South-West Asia and adjacent*
647 *regions*, Vol. 1. Warsaw: Polish Scientific Publishers
- 648 **Burgarella C, Lorenzo Z, Jabbour-Zahab R, Lumaret R, Guichoux E, Petit R, Soto A, Gil**
649 **L.** 2009. Detection of hybrids in nature: application to oaks (*Quercus suber* and *Q. ilex*).
650 *Heredity* 102: 442-452
- 651 **Cannon CH, Manos PS.** 2003. Phylogeography of the Southeast Asian stone oaks
652 (*Lithocarpus*). *Journal of Biogeography* 30: 211-226
- 653 **Cavender-Bares J, Gonzalez-Rodriguez A, Eaton DAR, Hipp AAL, Beulke A, Manos PS.**
654 2015. Phylogeny and biogeography of the American live oaks (*Quercus* subsection
655 *Virentes*): A genomic and population genetics approach. *Molecular Ecology* (in press)
- 656 **Cavender-Bares J, Gonzalez-Rodriguez A, Pahlich A, Koehler K, Deacon N.** 2011.
657 Phylogeography and climatic niche evolution in live oaks (*Quercus* series *Virentes*) from
658 the tropics to the temperate zone. *Journal of Biogeography* 38: 962–981
- 659 **Chen C, Qi ZC, Xu XH, Comes HP, Koch MA, Jin XJ, Fu CX, Qiu YX.** 2014.
660 Understanding the formation of Mediterranean-African-Asian disjunctions: evidence for
661 Miocene climate-driven vicariance and recent long-distance dispersal in the Tertiary
662 relict *Smilax aspera* (Smilacaceae). *New Phytologist* 204: 243-255.
- 663 **Crepet WL, Nixon KC.** 1989. Earliest megafossil evidence of Fagaceae: phylogenetic and
664 biogeographic implications. *American Journal of Botany* 76: 842–855

- 665 **Curtu AL, Gailing O, Finkeldey R.** 2007. Evidence for hybridization and introgression within
666 a species-rich oak (*Quercus* spp.) community. *Bmc Evolutionary Biology* **7**: 218
- 667 **Denk T, Grimm GW.** 2009. Significance of pollen characteristics for infrageneric classification
668 and phylogeny in *Quercus* (Fagaceae). *International Journal of Plant Sciences* **170**: 926–
669 940
- 670 **Denk T, Grimm GW.** 2010. The oaks of western Eurasia: traditional classifications and
671 evidence from two nuclear markers. *Taxon* **59**: 351-366
- 672 **Denk T, Grímsson F, Zetter R.** 2012. Fagaceae from the early Oligocene of Central Europe:
673 persisting New World and emerging Old World biogeographic links. *Review of*
674 *Palaeobotany and Palynology* **169**: 7–20
- 675 **Desamore A, Laenen B, Devos N, Popp M, Gonzalez-Mancebo JM, Carine MA,**
676 **Vanderpoorten A.** 2011. Out of Africa: north-westwards Pleistocene expansions of the
677 heather *Erica arborea*. *Journal of Biogeography* **38**:164-176
- 678 **Dodd RS, Afzal-Rafii Z.** 2004. Selection and dispersal in a multispecies oak hybrid zone.
679 *Evolution* **58**: 261-269
- 680 **Eaton DAR, Gonzalez-Rodriguez A, Hipp AL, Cavender-Bares J.** 2015. *Introgression*
681 *obscures and reveals historical relationships among the American live oaks*. bioRxiv
682 DOI: 10.1101/016238.
- 683 **Excoffier L, Foll M, Petit RJ.** 2009. Genetic Consequences of Range Expansions. *Annual*
684 *Review of Ecology, Evolution and Systematics* **40**: 481-501
- 685 **Flora of China Editorial Committee.** 1999. *Flora of China, Cycadaceae through Fagaceae,*
686 Vol. 4. St. Louis: Missouri Botanical Garden

- 687 **Flora of North America Editorial Committee.** 1997. *Flora of North America north of Mexico*,
688 Vol. 3. New York: Oxford University Press
- 689 **Funk D, Omland K.** 2003. Species-level paraphyly and polyphyly: frequency, causes and
690 consequences, with insights from animal mitochondrial DNA. *Annual Reviews of*
691 *Ecology, Evolution and Systematics* **34**: 397–423
- 692 **Fujii N, Tomaru N, Okuyama K, Koike T, Mikami T, Ueda K.** 2002. Chloroplast DNA
693 phylogeography of *Fagus crenata* (Fagaceae) in Japan. *Plant Systematics and Evolution*
694 **232**: 21-33
- 695 **Göker M, García-Blázquez G, Voglmayr H, Tellería MT, Martín MP.** 2009. Molecular
696 taxonomy of phytopathogenic fungi: a case study in *Peronospora*. *Plos One* **4**: e6319
- 697 **Göker M, Grimm GW.** 2008. General functions to transform associate data to host data, and
698 their use in phylogenetic inference from sequences with intra-individual variability. *Bmc*
699 *Evolutionay Biology* **8**: 86
- 700 **Govaerts R, Frodin DG.** 1998. *World checklist and bibliography of Fagales (Betulaceae,*
701 *Corylaceae, Fagaceae and Ticodendraceae)*. Royal Botanic Gardens, Kew
- 702 **Grimm GW, Denk T.** 2010. The reticulate origin of modern plane trees (*Platanus*, Platanaceae)
703 - a nuclear marker puzzle. *Taxon* **59**: 134-147.
- 704 **Grimm GW, Renner SS.** 2013. Harvesting Betulaceae sequences from GenBank to generate a
705 new chronogram for the family. *Botanical Journal of the Linnean Society* **172**: 465–477
- 706 **Grímsson F, Zetter R, Grimm GW, Krarup Pedersen G, Pedersen AK, Denk T.** 2015.
707 Fagaceae pollen from the early Cenozoic of West Greenland: revisiting Engler's and
708 Chaney's Arcto-Tertiary hypotheses. *Plant Systematics and Evolution* **301**: 809–832

- 709 **Gugger PF, Cavender-Bares J.** 2013. Molecular and morphological support for a Florida origin
710 of the Cuban oak. *Journal of Biogeography* **40**: 632-645
- 711 **Güner TH, Denk T.** 2012. The genus *Mahonia* in the Miocene of Turkey: taxonomy and
712 biogeographic implications. *Review of Palaeobotany and Palynology* **175**: 32–46
- 713 **Hare MP, Avise JC.** 1998. Population structure in the American oyster as inferred by nuclear
714 gene genealogies. *Molecular Biology and Evolution* **15**: 119–28
- 715 **Hipp AL, Eaton DAR, Cavender-Bares J, Fitzek E, Nipper R, Manos PS.** 2014. A
716 framework phylogeny of the American oak clade based on sequenced RAD data. *Plos*
717 *One* **9**: e93975
- 718 **Hofmann C-C.** 2010. Microstructure of Fagaceae pollen from Austria (Paleocene/Eocene
719 boundary) and Hainan Island (?middle Eocene). Paper presented at the 8th European
720 Palaeobotany-Palynology Conference, 6-19 July, Budapest, Hungary
- 721 **Hubert F, Grimm GW, Jouselin E, Berry V, Franc A, Kremer A.** 2014. Multiple nuclear
722 genes stabilize the phylogenetic backbone of the genus *Quercus*. *Systematics and*
723 *Biodiversity* **12**: 405–423.
- 724 **Huzioka K, Takahasi E.** 1970. *The Eocene flora of the Ube coal-field, southwest Honshu,*
725 *Japan.* Akita University, Mining College, Japan
- 726 **Kanno M, Yokoyama J, Suyama Y, Ohyama M, Itoh T, Suzuki M.** 2004. Geographical
727 distribution of two haplotypes of chloroplast DNA in four oak species (*Quercus*) in
728 Japan. *Journal of Plant Research* **117**: 311-317
- 729 **Katoh K, Standley DM.** 2013. MAFFT multiple sequence alignment software version 7:
730 improvements in performance and usability. *Molecular Biology and Evolution* **30**: 772–
731 780

- 732 **Kitamura S.** 1955. Flowering plants and ferns. In: Kihara H,ed. *Fauna and Flora of Nepal*
733 *Himalaya*. Kyoto: Fauna and Flora Research Society, 73–290
- 734 **Kmenta M.** 2011. *Die Mikroflora der untermiozänen Fundstelle Altmittweida, Deutschland*.
735 Wien: University of Wien, Austria
- 736 **Knapp M, Stöckler K, Havell D, Delsuc F, Sebastiani F, Lockhart PJ.** 2005. Relaxed
737 molecular clock provides evidence for long-distance dispersal of *Nothofagus* (Southern
738 Beech). *Plos Biology* **3**: e14
- 739 **Köppen W.** 1936. Das geographische System der Klimate. In: Köppen W, Geiger R eds.
740 *Handbuch der Klimatologie, Band 1*. Berlin: Teil C. Gebr. Borntraeger, 1–44
- 741 **Kottek M, Grieser J, Beck C, Rudolf B, Rubel F.** 2006. World map of the Köppen-Geiger
742 climate classification updated. *Meteorologische Zeitschrift* **15**: 259-263
- 743 **Kubitzki K.** 1993. Fagaceae. In: Kubitzki K, Rohwer JG, Bittrich V,eds. *The families and*
744 *genera of vascular plants*, vol 2. Berlin: Springer, 301–309
- 745 **Kvaček Z, Walther H.** 1989. Palaeobotanical studies in Fagaceae of the European Tertiary.
746 *Plant Systematics and Evolution* **162**: 213-229
- 747 **Lagache L, Klein EK, Guichoux E, Petit RJ.** 2013. Fine-scale environmental control of
748 hybridization in oaks. *Molecular Ecology* **22**: 423-436.
- 749 **Lang P, Dane F, Kubisiak TL, Huang H.** 2007. Molecular evidence for an Asian origin and a
750 unique westward migration of species in the genus *Castanea* via Europe to North
751 America. *Molecular Phylogenetics and Evolution* **43**: 49-59
- 752 **Lei M, Wang Q, Wu ZJ, Lopez-Pujol J, Li DZ, Zhang ZY.** 2012. Molecular phylogeography
753 of *Fagus engleriana* (Fagaceae) in subtropical China: limited admixture among multiple
754 refugia. *Tree Genetics and Genomes* **8**: 1203-1212.

- 755 **Lei YZ, Zhang QR, He W, Cao XP.** 1992. Tertiary. In: Wang XF,ed. *Geology of Hainan*
756 *Island. I. Stratigraphy and Palaeontology* [in Chinese]. Beijing: Geological Publishing
757 House, 218–266
- 758 **Lepais O, Petit R, Guichoux E, Lavabre J, Alberto F, Kremer A, Gerber S.** 2009. Species
759 relative abundance and direction of introgression in oaks. *Molecular Ecology* **18**: 2228-
760 2242
- 761 **Librado P, Rozas J** (2009) DnaSP v5: A software for comprehensive analysis of DNA
762 polymorphism data. *Bioinformatics* **25**: 1451–1452
- 763 **López de Heredia U, Jiménez P, Collada C, Simeone MC, Bellarosa R, Schirone B, Cervera**
764 **MT, Gil L.** 2007 Multi-marker phylogeny of three evergreen oaks reveals vicariant
765 patterns in the Western Mediterranean. *Taxon* **56**: 1209-1209
- 766 **Maddison WP, Maddison DR.** 2011. *Mesquite: a modular system for evolutionary analysis.*
767 *Version 2.75.* <http://mesquiteproject.org>.
- 768 **Magri D, Fineschi S, Bellarosa R, Buonamici A, Sebastiani F, Schirone B, Simeone MC,**
769 **Vendramin GG.** 2007. The distribution of *Quercus suber* chloroplast haplotypes
770 matches the palaeogeographical history of the western Mediterranean. *Molecular Ecology*
771 **16**: 5259-5266
- 772 **Manchester SR.** 1994. Fruits and seeds of the Middle Eocene nut beds flora, Clarno Formation,
773 Oregon. *Palaeontographica Americana* **58**: 1–205
- 774 **Manos PS, Cannon CH, Oh S-H.** 2008. Phylogenetic relationships and taxonomic status of the
775 paleoendemic Fagaceae of Western North America: recognition of a new genus,
776 *Notholithocarpus*. *Madroño* **55**: 181–190

- 777 **Manos PS, Stanford AM.** 2001. The historical biogeography of Fagaceae: tracking the tertiary
778 history of temperate and subtropical forests of the Northern Hemisphere. *International*
779 *Journal of Plant Sciences* **162**: S77-S93
- 780 **Masta SE, Sullivan B, Lamb T, Routman EJ.** 2002. Phylogeography, species boundaries, and
781 hybridization among toads of the *Bufo americanus* group. *Molecular Phylogenetics and*
782 *Evolution* **24**: 302–14
- 783 **McIntyre DJ.** 1991. Pollen and spore flora of an Eocene forest, eastern Axel Heiberg Island,
784 N.W.T. In: Christie RL, McMillan NJ, eds. *Tertiary Fossil Forest of the Geodetic Hills,*
785 *Axel Heiberg Island, Arctic Archipelago.* Geological Survey of Canada, Bulletin, vol 403,
786 83–97
- 787 **McIver EE, Basinger JF.** 1999. Early Tertiary floral evolution in the Canadian high arctic.
788 *Annals of the Missouri Botanical Garden* **86**: 523–545
- 789 **Menitsky YL.** 2005. *Oaks of Asia.* Science Publishers, Enfield, New Hampshire, USA
- 790 **Migliore J, Baumel A, Juin M, Medail F.** 2012. From Mediterranean shores to central Saharan
791 mountains: key phylogeographical insights from the genus *Myrtus*. *Journal of*
792 *Biogeography* **39**: 942-956
- 793 **Mir C, Jarne P, Sarda V, Bonin A, Lumaret R.** 2009. Contrasting nuclear and cytoplasmic
794 exchanges between phylogenetically distant oak species (*Quercus suber* L. and *Q. ilex*
795 L.) in Southern France: inferring crosses and dynamics. *Plant Biology* **11**: 213-226
- 796 **Moran EV, Willis J, Clark JS.** 2012. Genetic evidence for hybridization in red oaks (*Quercus*
797 sect. Lobatae, Fagaceae). *American Journal of Botany* **99**: 92-100

- 798 **Neophytou C, Aravanopoulos FA, Fink S, Dounavi A.** 2011. Interfertile oaks in an island
799 environment: II. Limited hybridization *Quercus alnifolia* Poech and *Q. coccifera* L. in a
800 mixed stand. *European Journal of Forest Research* **130**: 623–635
- 801 **Neophytou C, Dounavi A, Fink S, Aravanopoulos FA.** 2010. Interfertile oaks in an island
802 environment: I. High nuclear genetic differentiation and high degree of chloroplast DNA
803 sharing between *Q. alnifolia* and *Q. coccifera* in Cyprus. A multipopulation study.
804 *European Journal of Forest Research* **130**: 543–555
- 805 **Nieto Feliner G.** 2014. Patterns and processes in plant phylogeography in the Mediterranean
806 Basin. A review. *Perspectives in Plant Ecology, Evolution and Systematics* **16**: 265-278.
- 807 **Nylander JAA.** 2004. *MrModeltest. 2.1.* Program distributed by the author. Evolutionary
808 Biology Centre, Uppsala University, Uppsala, Sweden
- 809 **Oh S-H, Manos PS.** 2008. Molecular phylogenetics and cupule evolution in Fagaceae as
810 inferred from nuclear CRABS CLAW sequences. *Taxon* **57**: 434–451
- 811 **Okaura T, Quang ND, Ubukata M, Harada K.** 2007. Phylogeographic structure and late
812 Quaternary population history of the Japanese oak *Quercus mongolica* var. *crispula* and
813 related species revealed by chloroplast DNA variation. *Genes and Genetics Systems* **82**:
814 465-477
- 815 **Ortego J, Bonal R.** 2010. Natural hybridisation between kermes (*Quercus coccifera* L.) and
816 holm oaks (*Q. ilex* L.) revealed by microsatellite markers. *Plant Biology* **12**: 234-238
- 817 **Pearse IS, Hipp AL.** 2009. Phylogenetic and trait similarity to a native species predict herbivory
818 on non-native oaks. *Proceedings of the National Academy of Sciences of the USA* **106**:
819 18097-18102

- 820 **Peel MC, Finlayson BL, McMahon TA.** 2007. Updated world map of the Köppen-Geiger
821 climate classification. *Hydrology and Earth System Sciences* **11**: 1633-1644
- 822 **Peñaloza-Ramírez JM, González-Rodríguez A, Mendoza-Cuenca L, Caron H, Kremer A,**
823 **Oyama K.** 2010. Interspecific gene flow in a multispecies oak hybrid zone in the Sierra
824 Tarahumara of Mexico. *Annals of Botany* **105**: 389-399
- 825 **Petit RJ, Bodénès C, Ducouso A, Roussel G, Kremer A.** 2004. Hybridization as a mechanism
826 of invasion in oaks. *New Phytologist* **161**: 151-164
- 827 **Petit RJ, Csaikl UM, Bordács S, Burg K, Coart E, Cottrell J, van Dam B, Deans JD,**
828 **Dumolin-Lapègue S, Fineschi S.** 2002. Chloroplast DNA variation in European white
829 oaks: phylogeography and patterns of diversity based on data from over 2600
830 populations. *Forest Ecology and Management* **156**: 5-26
- 831 **Piredda R, Simeone MC, Attimonelli M, Bellarosa R, Schirone B.** 2011. Prospects of
832 barcoding the Italian wild dendroflora: oaks reveal severe limitations to tracking species
833 identity. *Molecular Ecology Resources* **11**: 72-83
- 834 **Premoli AC, Mathiasen P, Acosta MC, Ramos VA.** 2012. Phylogeographically concordant
835 chloroplast DNA divergence in sympatric *Nothofagus* s.s. How deep can it be? *New*
836 *Phytologist* **193**: 261–275
- 837 **Rambaut A.** 2014. FigTree, a graphical viewer of phylogenetic trees, v1.4.2.. Institute of
838 Evolutionary Biology University of Edinburgh. <http://tree.bio.ed.ac.uk/software/figtree/>.
- 839 **Rieseberg LH, Soltis DE.** 1991. Phylogenetic Consequences of Cytoplasmic Gene Flow in
840 Plants. *Evolutionary Trends in Plants* **5**: 65-84

- 841 **Rodriguez-Sanchez F, Guzman B, Valido A, Vargas P, Arroyo J.** 2009. Late Neogene history
842 of the laurel tree (*Laurus* L., Lauraceae) based on phylogeographical analyses of
843 Mediterranean and Macaronesian populations. *Journal of Biogeography* **36**: 1270-1281
- 844 **Ronquist F, Huelsenbeck JP.** 2003. MrBayes 3: Bayesian phylogenetic inference under mixed
845 models. *Bioinformatics* **19**: 1572-1574
- 846 **Ronquist F, Klopstein S, Vilhelmsen L, Schulmeister S, Murray DL, Rasnitsyn AP.** 2012.
847 A total-evidence approach to dating with fossils, applied to the early radiation of the
848 hymenoptera. *Systematics Biology* **61**: 973–999
- 849 **Schnitzler JP, Steinbrecher R, Zimmer I, Steigner D, Fladung M.** 2004. Hybridization of
850 European oaks (*Quercus ilex* × *Q. robur*) results in a mixed isoprenoid emitter type. *Plant*
851 *Cell and Environment* **27**: 585-593
- 852 **Simeone MC, Piredda R, Papini A, Vessella F, Schirone B.** 2013. Application of plastid and
853 nuclear markers to DNA barcoding of Euro–Mediterranean oaks (*Quercus*, Fagaceae):
854 problems, prospects and phylogenetic implications. *Botanical Journal of the Linnean*
855 *Society* **172**: 478–499
- 856 **Stamatakis A, Hoover P, Rougemont J.** 2008. A rapid bootstrap algorithm for the RAxML
857 web servers. *Systematics Biology* **57**: 758–771
- 858 **Svenson U, Backlund A, McLoughlin S, Hill RS.** 2001. *Nothofagus* biogeography revisited
859 with special emphasis on the enigmatic distribution of subgenus *Brassospora* in New
860 Caledonia. *Cladistics* **17**: 28–47
- 861 **Swofford DL.** 2002. PAUP*: *Phylogenetic Analysis Using Parsimony (and Other Methods)* 4.0
862 *Beta*. Sunderland, MA: Sinauer Associates

- 863 **Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S** (2011) MEGA5: molecular
864 evolutionary genetics analysis using maximum likelihood, evolutionary distance, and
865 maximum parsimony methods. *Molecular Biology and Evolution* **28**: 2731–2739
- 866 **Thompson JD, Higgins DG, Gibson TJ.** 1994. CLUSTAL W: improving the sensitivity of
867 progressive multiple sequence alignment through sequence weighting, position specific
868 gap penalties and weight matrix choice. *Nucleic Acids Research* **22**: 4673-4680
- 869 **Tsutsui K, Suwa A, Sawada K, Kato T, Ohsawa TA, Watano Y.** 2009. Incongruence among
870 mitochondrial, chloroplast and nuclear gene trees in *Pinus* subgenus *Strobus* (Pinaceae).
871 *Journal of Plant Research* **122**: 509–521
- 872 **Valbuena-Carabaña M, González-Martínez SC, Hardy OJ, Gil .L** 2007. Fine-scale spatial
873 genetic structure in mixed oak stands with different levels of hybridization. *Molecular*
874 *Ecology* **16**: 1207-1219
- 875 **Valencia-Cuevas L, Mussali-Galante P, Piñero D, Castillo-Mendoza E, Rangel-Altamirano**
876 **G, Tovar-Sánchez E.** 2015. Hybridization of *Quercus castanea* (Fagaceae) across a red
877 oak species gradient in Mexico. *Plant Systematics and Evolution* **301**:1085-1097
- 878 **Velitzelos D, Bouchal JM, Denk T.** 2014. Review of the Cenozoic floras and vegetation of
879 Greece. *Review of Palaeobotany and Palynology* **204**: 56–117.
- 880 **Wang N, Wu FX.** 2015. New Oligocene cyprinid in the central Tibetan Plateau documents the
881 pre-uplift tropical lowlands. *Ichthyological Research* **62**: 274-285
- 882 **Weitschat W, Wichard W.** 2003. *Atlas of Plants and Animals in Baltic Amber*. Munchen: Pfeil
883 Verlag
- 884 **Whittemore AT, Schaal BA** (1991) Interspecific gene flow in sympatric oaks. *Proceedings of*
885 *the National Academy of Sciences of the USA* **88**: 2540-2544

- 886 **Wiens JJ, Servedio MR.** 2000. Species delimitation in systematics: inferring diagnostic
887 differences between species. *Proceedings of the Royal Society of London Series B* **267**:
888 631–36
- 889 **Xiang Q-Y, Manchester SR, Thomas DT, Zhang W, Fan C.** 2005. Phylogeny, biogeography,
890 and molecular dating of Cornelian cherries (*Cornus*, Cornaceae): Tracking Tertiary plant
891 migration. *Evolution* **59**: 1685-1700
- 892 **Xiang X-G, Wang W, Li R-Q, Lin L, Liu Y, Zhou Z-K, Li Z-Y, Chen Z-D.** 2014. Large-
893 scale phylogenetic analyses reveal fagalean diversification promoted by the interplay of
894 diaspores and environments in the Paleogene. *Perspectives in Plant Ecology, Evolution*
895 *and Systematics* **16**: 101–110
- 896 **Yi T, Miller AJ, Wen J.** 2004. Phylogenetic and biogeographic diversification of *Rhus*
897 (*Anacardiaceae*) in the Northern Hemisphere. *Molecular Phylogenetics and Evolution* **33**:
898 861-879
- 899 **Yoo K-O, Wen J.** 2007. Phylogeny of *Carpinus* and subfamily Coryloideae (*Betulaceae*) based
900 on chloroplast and nuclear ribosomal sequence data. *Plant Systematics and Evolution*
901 **267**: 25–35
- 902 **Zhang J-B, Li R-Q, Xiang X-G, Manchester SR, Lin L, Wang W, Wen J, Chen Z-D.** 2013a.
903 Integrated fossil and molecular data reveal the biogeographic diversification of the
904 eastern Asian-eastern North American disjunct hickory genus (*Carya* Nutt.). *Plos One* **8**:
905 e70449
- 906 **Zhang ZY, Wu R, Wang Q, Zhang ZR, Lopez-Pujol J, Fan DM, Li DZ.** 2013b. Comparative
907 phylogeography of two sympatric beeches in subtropical China: Species-specific
908 geographic mosaic of lineages. *Ecology and Evolution* **3**: 4461-4472.

- 909 **Zhou Z.** 1992. The fossil history of *Quercus*. *Acta Botanica Yunnanica* **15**: 21-33
- 910 **Zwickl DJ.** 2006. *Genetic algorithm approaches for the phylogenetic analysis of large*
- 911 *biological sequence datasets under the Maximum Likelihood criterion*. Ph.D. thesis,
- 912 University of Texas, Austin, USA
- 913

914 **Figure captions**

915

916 **Figure 1** ML tree of plastid accessions; tentatively rooted with the *Notholithocarpus-Chrysolepis*
917 subtree. Stars indicate subtrees comprising accessions of Mediterranean members of *Quercus*
918 Group Ilex. Number at branches indicate non-parametric bootstrap support under maximum
919 likelihood using two different implementations and posterior probabilities calculated using
920 Bayesian inference

921

922 **Figure 2** Haplotype network based on length-conserved portions of the *trnH-psbA* spacer.
923 Colouration refers to the taxonomic affiliation of specimens

924

925 **Figure 3** Geographic pattern of plastid haplotype variation in Mediterranean members of
926 *Quercus* Group Ilex. (A) Map showing the taxonomic identity of sampled specimens. (B) Map
927 showing the plastid haplotypes of sampled specimens

928

929 **Figure 4** Mapping of chloroplast evolution in oaks (using the same rooting scenario as in Fig. 1)
930 on current evolutionary synopsis (based on nuclear sequence data, morphology, and the fossil
931 record; modified after (Grímsson et al., 2015, fig. 16). Colouring of the plastid lineages refers to
932 branches/subclades in Fig. 1: bluish, common (ancestral) and ‘New World’ oak/castanoids
933 plastid haplotype lineages; green, lineages of the unique ‘Euro-Med’ plastid haplotype found
934 only in Mediterranean members of Group Ilex; reddish, lineages of ‘Old World’ oaks and
935 Eurasian castanoids. Note that members of Group Ilex keep plastid haplotypes of five different

936 evolutionary sources/systematic affinities. Abbreviations: C = Cretaceous, Pa = Paleocene, E =
937 Eocene, O = Oligocene, M = Miocene, Pl = Plio-/Pleistocene

938

939 **Figure 5** Eocene set-up and the origin of the ‘Euro-Med’ haplotype (palaeogeographic map ©
940 Ron Blakey, Colorado Plateau Geosystems). **A** Unequivocal fossil record of oaks in the Eocene
941 mapped on a palaeotopographic map (© Ron Blakey, Colorado Plateau Geosystems)
942 highlighting a primary split into a high-latitude and mid-latitude lineage that likely correspond to
943 the deep phylogenetic split seen in nuclear and plastid sequence data of modern oaks between the
944 ‘New World Clade’ (Groups *Protobalanus*, *Quercus* and *Lobatae*) and the ‘Old World Clade’
945 (Groups *Cyclobalanopsis*, *Ilex*, *Cerris*). **B-C** Scenarios that can explain the occurrence of the
946 unique ‘Euro-Med’ haplotype in westernmost members of *Quercus* Group *Ilex*. **B** The ‘Euro-
947 Med’ haplotype belonged to an extinct oak lineage geographically/biologically separated from
948 both the ancestors of the New World and Old World Clade. Westward expansion of Himalayan
949 members of Group *Ilex* and subsequent large-scale introgression/hybridisation homogenised the
950 western members of Group *Ilex* and the extinct oak lineage, retaining and evolving the original
951 haplotype in the Mediterranean region. **C** The ‘Euro-Med’ haplotype reflects geographic
952 fragmentation within the Paleogene range of the Old World Clade that was overprinted to some
953 degree after later radiation phases of Group *Ilex*

954

955 **Figure 6** Tectonic activity during the Eocene (palaeogeographic map © Ron Blakey, Colorado
956 Plateau Geosystems) and past and modern distribution of the New World (white) and Old World
957 (yellow) groups within *Quercus*. Black lines indicate major subduction zones, red lines major
958 orogenies. Note that the high latitude lineage of oaks (*Quercus* Group *Lobatae*, Group *Quercus*,

959 Group Protobalanus) evolved in tectonically stable regions, whereas the low latitude lineage
960 (*Quercus* Group Ilex, Group Cyclobalanopsis, Group Cerris) evolved in tectonically unstable
961 regions. Uppercase and lowercase letters refer to extant and extinct distribution areas of major
962 oak lineages: P,p = Group Protobalanus; Q,q = Group Quercus; L,l = Group Lobatae; I,i = Group
963 Ilex; C,c = Group Cerris; Y,y = Group Cyclobalanopsis
964
965

966 **Supporting Information**

967 **Table S1** List of specimens and accessions included in this study

968 **Table S2** Intra- and intertaxonomic minimum and maximum pairwise genetic distances

969 **File S3** MJ networks based on different plastid gene regions

970

971 **Data Archival Location**

972 All sequence data generated as part of this study are available on GenBank

973 (<http://www.ncbi.nlm.nih.gov/genbank/>) under accession numbers LM652969-LM653098; other

974 relevant data are within the paper and its Supplementary files. Primary data and analyses are

975 provided for anonymous download at www.palaeogrimm.org/data/Smn15_OSA.zip

976

Figure 1(on next page)

ML tree of the investigated oak accessions

ML tree of plastid accessions; tentatively rooted with the Notholithocarpus-Chrysolepis subtree. Stars indicate subtrees comprising accessions of Mediterranean members of Quercus Group Ilex. Number at branches indicate non-parametric bootstrap support under maximum likelihood using two different implementations and posterior probabilities calculated using Bayesian inference

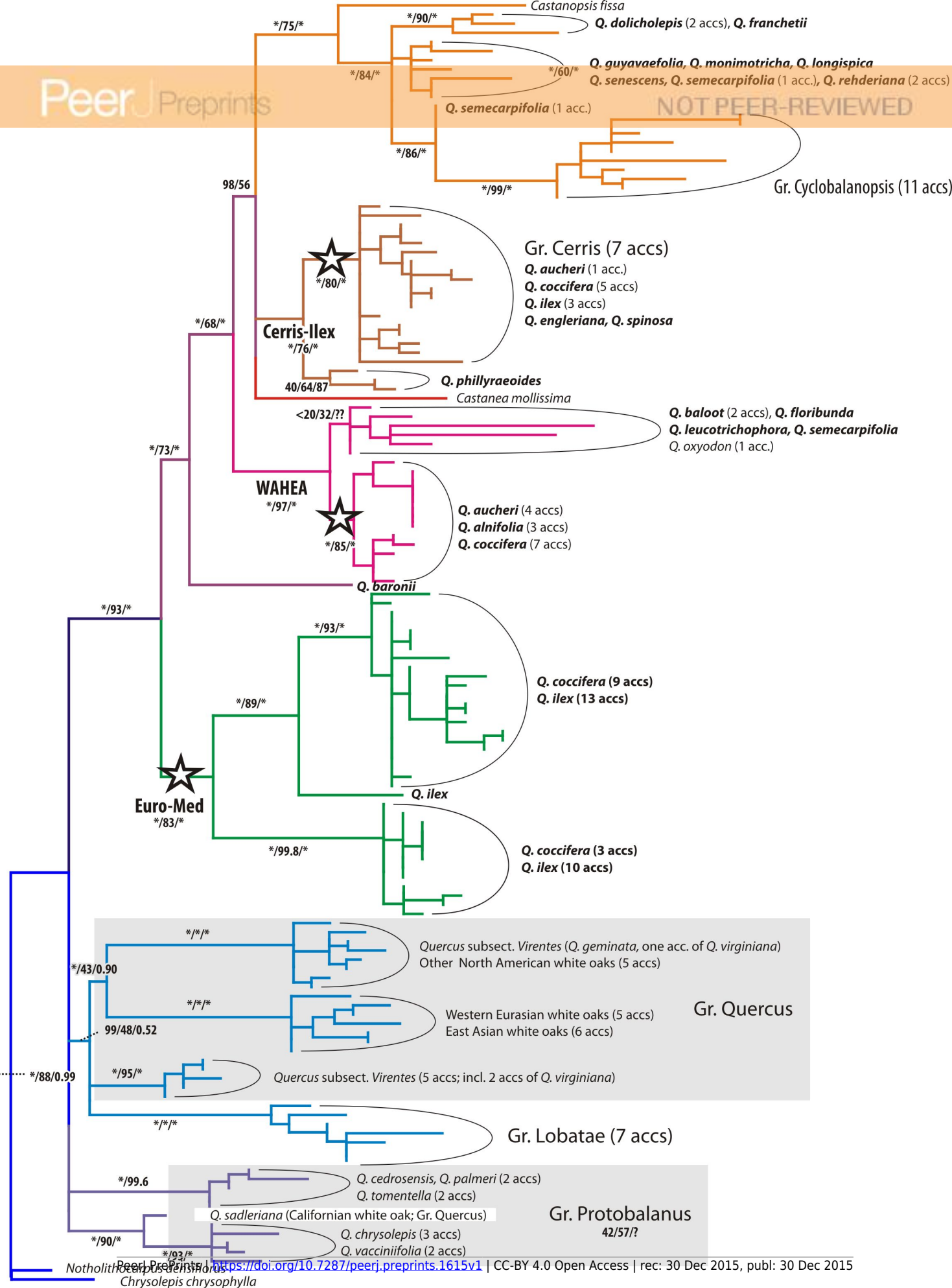


Figure 2 (on next page)

Haplotype network based on the trnH-psbA spacer

Haplotype network based on length-conserved portions of the trnH-psbA spacer. Colouration refers to the taxonomic affiliation of specimens

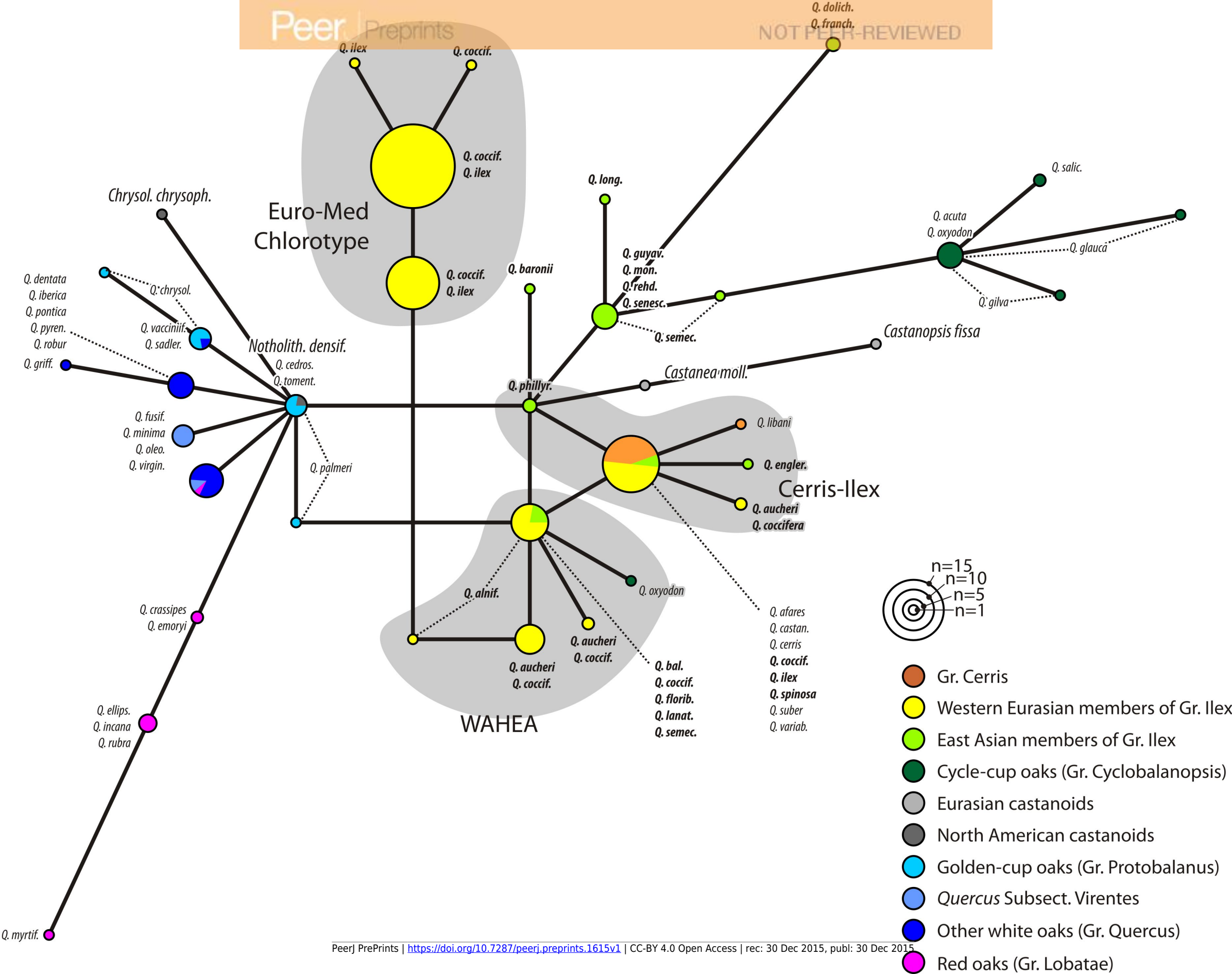
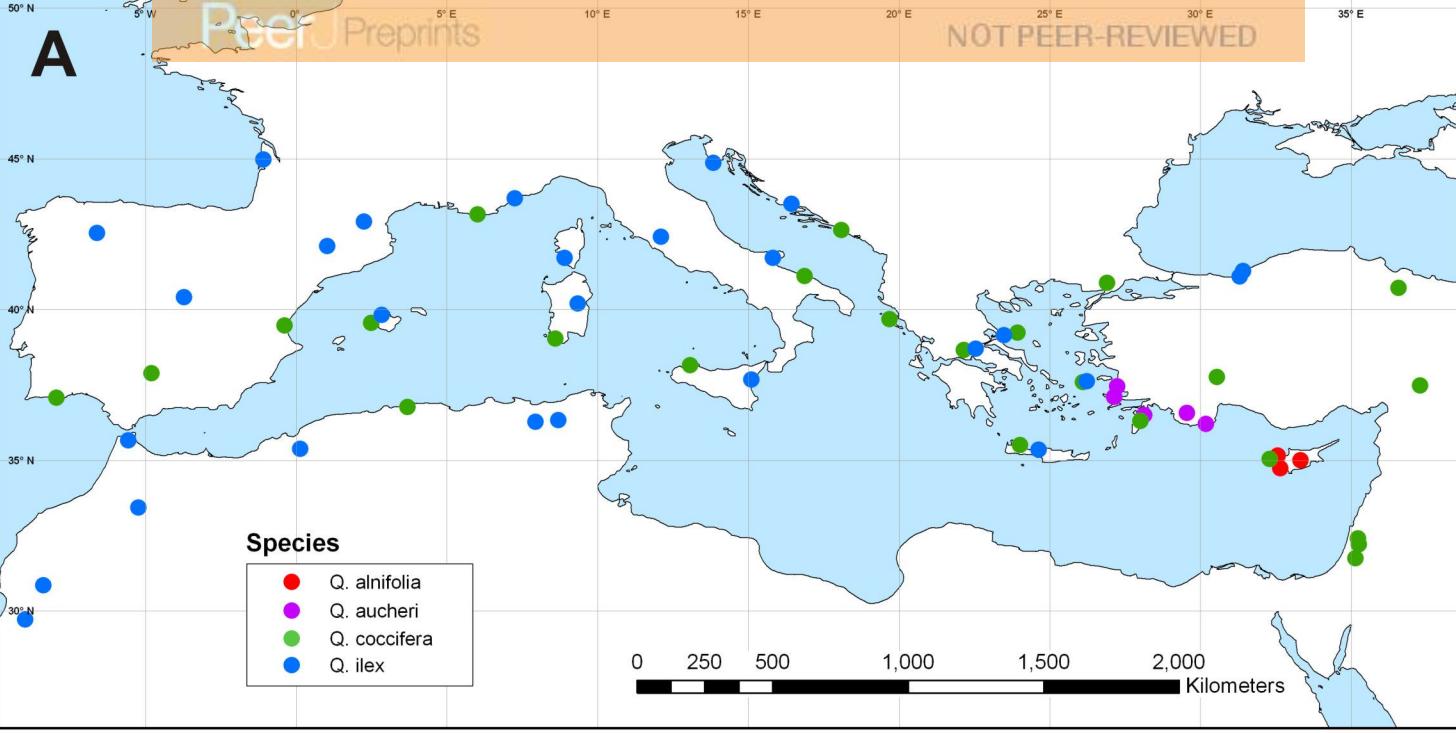


Figure 3(on next page)

Plastid haplotype variation in Mediterranean members of *Quercus* Group Ilex

Geographic pattern of plastid haplotype variation in Mediterranean members of *Quercus* Group Ilex. (A) Map showing the taxonomic identity of sampled specimens. (B) Map showing the plastid haplotypes of sampled specimens

A



B

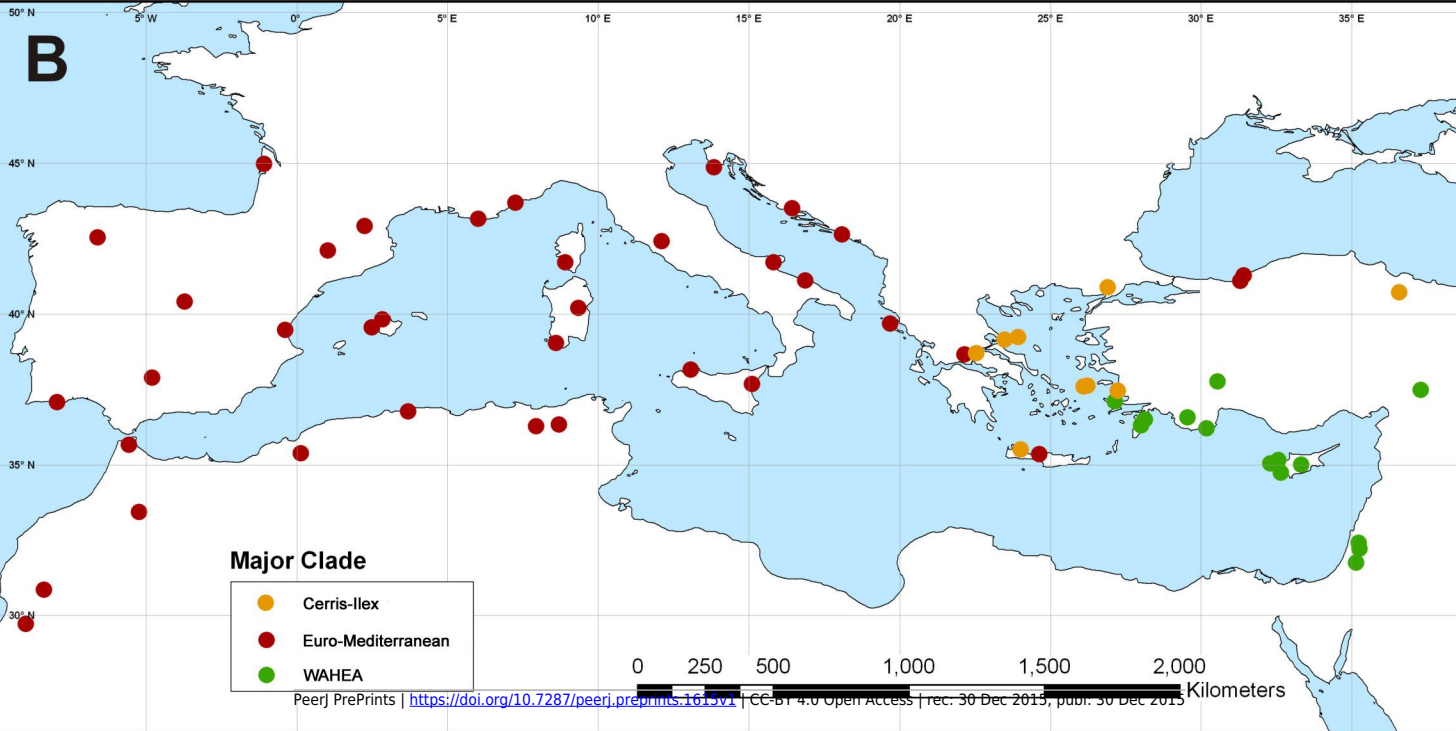
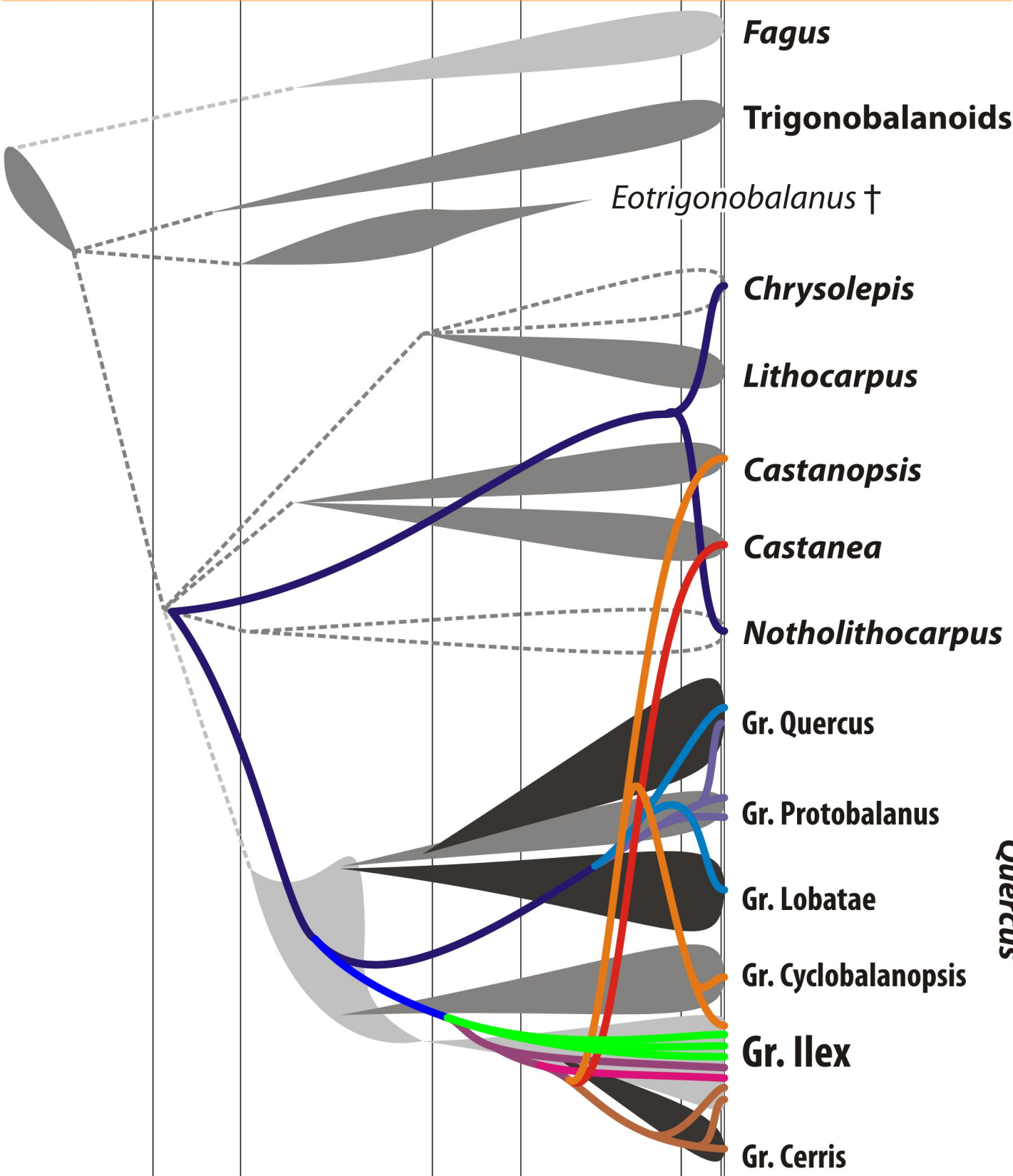


Figure 4(on next page)

Map of chloroplast evolution in oaks

Mapping of chloroplast evolution in oaks (using the same rooting scenario as in Fig. 1) on current evolutionary synopsis (based on nuclear sequence data, morphology, and the fossil record; modified after (Grímsson et al., 2015, fig. 16). Colouring of the plastid lineages refers to branches/subclades in Fig. 1: bluish, common (ancestral) and 'New World' oak/castanoids plastid haplotype lineages; green, lineages of the unique 'Euro-Med' plastid haplotype found only in Mediterranean members of Group Ilex; reddish, lineages of 'Old World' oaks and Eurasian castanoids. Note that members of Group Ilex keep plastid haplotypes of five different evolutionary sources/systematic affinities. Abbreviations: C = Cretaceous, Pa = Paleocene, E = Eocene, O = Oligocene, M = Miocene, Pl = Plio-/Pleistocene



Quercus

Figure 5(on next page)

Origin of the 'Euro-Med' haplotype

Eocene set-up and the origin of the 'Euro-Med' haplotype (palaeogeographic map © Ron Blakey, Colorado Plateau Geosystems). A Unequivocal fossil record of oaks in the Eocene mapped on a palaeotopographic map (© Ron Blakey, Colorado Plateau Geosystems) highlighting a primary split into a high-latitude and mid-latitude lineage that likely correspond to the deep phylogenetic split seen in nuclear and plastid sequence data of modern oaks between the 'New World Clade' (Groups Protobalanus, Quercus and Lobatae) and the 'Old World Clade' (Groups Cyclobalanopsis, Ilex, Cerris). B-C Scenarios that can explain the occurrence of the unique 'Euro-Med' haplotype in westernmost members of *Quercus* Group Ilex. B The 'Euro-Med' haplotype belonged to an extinct oak lineage geographically/biologically separated from both the ancestors of the New World and Old World Clade. Westward expansion of Himalayan members of Group Ilex and subsequent large-scale introgression/hybridisation homogenised the western members of Group Ilex and the extinct oak lineage, retaining and evolving the original haplotype in the Mediterranean region. C The 'Euro-Med' haplotype reflects geographic fragmentation within the Paleogene range of the Old World Clade that was overprinted to some degree after later radiation phases of Group Ilex

A

NOT PEER-REVIEWED

High-latitude lineage:
including ancestors of New World Clade

Affinity with East Asia
(*Quercus* Gr. *Cyclobalanopsis*, *Lithocarpus*)

Modern distribution
of Euro-Med chlorotype

- ★ Oak of unknown affinity/
extinct lineage
- ★ New World Clade
- ★ Group *Quercus*
- ★ Group *Lobatae*
- ★ Old World Clade

Mid-latitude lineage
including ancestors of
Old World Clade

B

Turgai Street closure

Extinct oak lineage

Westward expansion of Group *Ilex*

C

Turgai Street closure

Euro-Med chlorotype

Asian chlorotypes

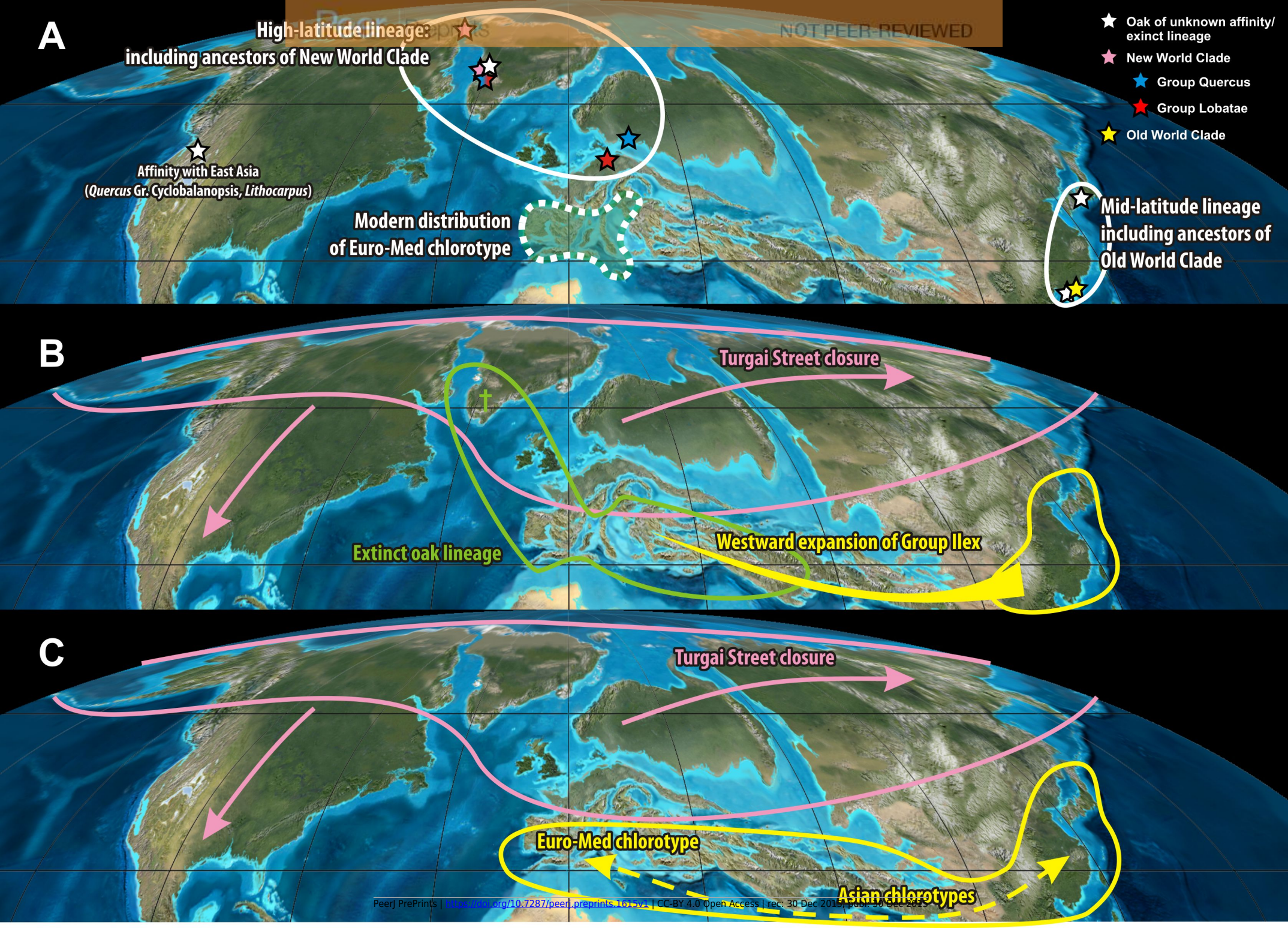


Figure 6(on next page)

Tectonic activity during the Eocene

Tectonic activity during the Eocene (palaeogeographic map © Ron Blakey, Colorado Plateau Geosystems) and past and modern distribution of the New World (white) and Old World (yellow) groups within *Quercus*. Black lines indicate major subduction zones, red lines major orogenies. Note that the high latitude lineage of oaks (*Quercus* Group Lobatae, Group Quercus, Group Protobalanus) evolved in tectonically stable regions, whereas the low latitude lineage (*Quercus* Group Ilex, Group Cyclobalanopsis, Group Cerris) evolved in tectonically unstable regions. Uppercase and lowercase letters refer to extant and extinct distribution areas of major oak lineages: P,p = Group Protobalanus; Q,q = Group Quercus; L,l = Group Lobatae; I,i = Group Ilex; C,c = Group Cerris; Y,y = Group Cyclobalanopsis

'New World Clade'

'Old World Clade'



Table 1 (on next page)

Species list

Species included in *Quercus* Group Ilex according to Denk and Grimm (2010); nomenclature followed Govaerts and Frodin (1998); species investigated in the present study are bolded.

Taxonomic remarks and species distributions according to Govaerts and Frodin (1998),

*Menitsky (2005) and **Flora of China Editorial Committee (1999).

1
2
3
4
5

Table 1 – Species included in *Quercus* Group Ilex according to Denk and Grimm (Denk and Grimm 2010); nomenclature followed Govaerts and Frodin (1998); species investigated in the present study are bolded. Taxonomic remarks and species distributions according to Govaerts and Frodin (1998), *Menitsky (2005) and **Flora of China Editorial Committee (1999).

Species	Taxonomic remarks	Distribution
<i>Q. acrodonta</i> Seemen	Includes <i>Q. handeliana</i> A. Camus*/**	C, E and S China
<i>Q. alnifolia</i> Poech		Cyprus
<i>Q. aquifolioides</i> Rehder & E.H. Wilson	Includes <i>Q. semecarpifolia</i> subsp. <i>glabra</i> *	Tibet, C and SW China to Myanmar
<i>Q. aucheri</i> Jaub. & Spach		SW Anatolia
<i>Q. baloot</i> Griff.		Pakistan, Afghanistan
<i>Q. baronii</i> Skan	Numerous morpho-ecological traits in common with members of Group Cerris*	NC and SW China
<i>Q. bawanglingensis</i> C.C. Huang, Ze X. Li & F.W. Xing	Poorly known; uncertain status, related to <i>Q. phillyreoides</i> **	SE China
<i>Q. coccifera</i> L.	Includes <i>Q. calliprinos</i> Webb*	Mediterranean
<i>Q. cocciferoides</i> Hand.-Mazz.	Includes <i>Q. taliensis</i> A. Camus**	CS China
<i>Q. floribunda</i> Lindl. ex A. Camus	Basionym: <i>Q. dilatata</i> Lindl. ex A.DC. <i>nom. illegit.</i>	Pakistan, Afghanistan, Nepal
<i>Q. dolicholepis</i> A. Camus	Includes <i>Q. fimbriata</i> **	CW to SW China
<i>Q. engleriana</i> Seemen		Tibet to E China, Myanmar
<i>Q. fimbriata</i> Y.C. Hsu & H. Wei Jen	Included in <i>Q. semecarpifolia</i> * or <i>Q. dolicholepis</i> **	C China
<i>Q. franchetii</i> Skan		C China to N Vietnam
<i>Q. gilliana</i> Rehder & E.H. Wilson	Included in <i>Q. spinosa</i> **	Tibet, C and S China
<i>Q. guyavifolia</i> H. Lév.	Included in <i>Q. semecarpifolia</i> subsp. <i>glabra</i> *; <i>Q.</i>	C and S China

	<i>pannosa</i> or <i>Q. aquifolioides</i> var. <i>rufescens</i> **	
<i>Q. ilex</i> L.		Mediterranean
<i>Q. kingiana</i> Craib		C China to N Thailand
<i>Q. lanata</i> Sm.	Included in <i>Q. leucotrichophora</i> **	Buthan to Vietnam
<i>Q. leucotrichophora</i> A. Camus	Basionym: <i>Q. incana</i> Roxb. <i>nom. illegit.</i>	N Pakistan, N India, Nepal to N Vietnam
<i>Q. lodicosa</i> O.E. Warb. & E.F. Warb.		SE Tibet to Myanmar
<i>Q. longispica</i> (Hand.-Mazz.) A. Camus	Includes <i>Q. semecarpifolia</i> subsp. <i>glabra</i> *; <i>Q. rehderiana</i> **	C and S China
<i>Q. marlipoensis</i> Hu & Cheng	Poorly known*; very close to <i>Q. engleriana</i> **	C China
<i>Q. monimotricha</i> Hand.-Mazz.		C and S China
<i>Q. oxyphylla</i> Hand.-Mazz.	Includes <i>Q. spathulata</i> Seemen; included in <i>Q. dolicholepis</i> **	C and S China
<i>Q. pannosa</i> Hand.-Mazz.	Possibly conspecific with <i>Q. semecarpifolia</i> *	C and S China
<i>Q. phillyreoides</i> A. Gray	Includes <i>Q. utilis</i> *	C China to Japan
<i>Q. pseudosemecarpifolia</i> A. Camus	Includes <i>Q. semecarpifolia</i> subsp. <i>glabra</i> *, <i>Q. rehderiana</i> **	Tibet to CS China
<i>Q. rehderiana</i> Hand.-Mazz.	Included in <i>Q. semecarpifolia</i> subsp. <i>glabra</i> *; includes <i>Q. longispica</i> and <i>Q. pseudosemecarpifolia</i> **	Tibet to C and S China
<i>Q. semecarpifolia</i> Sm.	Includes: <i>Q. fimbriata</i> , <i>Q. gujavifolia</i> , <i>Q. aquifolioides</i> , <i>Q. rehderiana</i> , <i>Q. longispica</i> , <i>Q. pseudosemecarpifolia</i> *	Afghanistan to Myanmar
<i>Q. senescens</i> Hand.-Mazz.		E Himalaya, Tibet, C and S China
<i>Q. setulosa</i> Hickel & A. Camus		C China to Vietnam
<i>Q. spinosa</i> David		NC and SW China to Taiwan
<i>Q. tarokoensis</i> Hayata		E Taiwan
<i>Q. utilis</i> Hu & Cheng	Included in <i>Q. phillyreoides</i> subsp. <i>fokiensis</i> *	C China

Table 2 (on next page)

Diversity values of the DNA fragments used for the analyses

1

2 **Table 2.** Diversity values and models of DNA evolution of the fragments used for the analyses in 59 *Quercus* species (137
3 individuals) and 4 outgroup taxa. AL: Aligned length (bp); P: uncorrected p-distance (min. – max.); N hap: Number of identified
4 haplotypes, brackets: with gaps considered; Hd: Haplotype diversity; S: Number of polymorphic sites; θ : Nucleotide polymorphism;
5 π : Nucleotide diversity; PIC: Number of Parsimony Informative Characters; ME: Model of evolution.

6

Markers	AL	P	Nhap	Hd	S	PICs	θ_w	π	ME
<i>rbcL</i>	743	0.00 – 0.008	28	0.846±0.027	26	18	0.0063	0.0027	HKY+I
<i>trnH-psbA</i>	634	0.00 – 0.035	37 (84)	0.944±0.008	38	23	0.0159	0.009	GTR+G
<i>trnK/matK</i>	705	0.00 – 0.022	49 (51)	0.952±0.008	59	31	0.0156	0.0064	n.d.
<i>trnK</i> (intron)	401	n.d.	32 (34)	0.821±0.028	36	16	0.0169	0.0048	GTR+G
<i>matK</i> (codons)	304	n.d.	25	0.925±0.008	23	15	0.0146	0.0086	HKY
<i>rbcL+trnK/matK+trnH-psbA</i>	2082	0.00 – 0.014	74 (110)	0.978±0.005	122	72	0.0119	0.0056	Combined
<i>rbcL + matK</i>	1047	n.d.	49	0.965±0.006	49	33	0.0085	0.0044	n.d.
<i>trnH-psbA + trnK</i>	1035	n.d.	57 (103)	0.954±0.008	69	34	0.0161	0.0067	n.d.
<i>trnH-psbA + trnK-matK</i>	1339	n.d.	65 (110)	0.970±0.006	92	49	0.0155	0.0072	n.d.

7

Table 3 (on next page)

Divergence patterns in other Fagaceae and Fagales

Divergence patterns in *Quercus* compared to other Fagaceae and Fagales based inter-species pair wise uncorrected p-distances of sequences retrieved from GenBank and produced in this study; Nt = number of taxa; Ns = number of sequences.

- 1 **Table 3.** Divergence patterns in *Quercus* compared to other Fagaceae and Fagales based inter-species pair wise uncorrected *p*-
 2 distances of sequences retrieved from GenBank and produced in this study; Nt = number of taxa; Ns = number of sequences.

Taxon	Intragenomic divergence						Mean intergeneric divergence at family level‡							
			<i>rbcL</i>		<i>matK</i>		<i>trnH-psbA</i>		<i>rbcL</i>		<i>matK</i>		<i>trnH-psbA</i>	
	Nt	Ns	Min.	Max.	Min.	Max.	Min.	Max.	Min.	Max.	Min.	Max.	Min.	Max.
<i>Quercus</i>	87/87/86	219/255/382	0.000	0.010	0.000	0.023	0.000	0.042	0.003	0.011	0.006	0.015	0.006	0.042
<i>Fagus</i>	8/9/6	33/30/19	0.000	0.014	0.000	0.007	0.000	0.042	0.024	0.036	0.091	0.098	0.120	0.147
Other Fagaceae	37/21/28	102/32/86	0.000	0.020	0.000	0.021	0.000	0.020	0.003	0.013	0.000	0.019	0.011	0.032
Nothofagaceae*	23/†/14	35/†/53	0.000	0.027	–†	–†	0.000	0.017	0.012	0.023	–†	–†	0.017	0.041
Betulaceae	55/19/77	131/34/247	0.000	0.011	0.000	0.006	0.000	0.069	0.006	0.024	0.014	0.033	0.011	0.079
Juglandaceae	18/†/21	23/†/28	0.000	0.005	–†	–†	0.000	0.007	0.000	0.021	–†	–†	0.006	0.034

* Values for *rbcL* may be over-estimated (data usually older than 15 years; sequences show features characteristic for sequencing and editing artifacts)

† Insufficient data.

‡ Values for *Quercus* and other Fagaceae not including *Fagus* (see *Fagus* for max. inter-generic divergence in Fagaceae)

3

Table 4 (on next page)

Eocene fossils of Quercus

1 **Table 4.** Eocene fossil record of *Quercus*.

Locality, site, age	Reference	Taxon, organ	Affinity
Clarno Fm., Oregon, western U.S.A.; ~48 Ma	Manchester 1994	“ <i>Quercus</i> ” <i>paleocarpa</i> Manchester; cupules and acorns	Group Cyclobalanopsis (? <i>Lithocarpus</i>)
Axel-Heiberg Island, Canadian Arctic; ~45 Ma	McIntyre 1991	Pollen and leaves	New World Clade;
Hareøen, western Greenland; ~42 Ma	McIver and Basinger 1999		<i>Quercus</i> Group <i>Quercus</i> /Lobatae
	Grimsson et al. 2015	<i>Quercus</i> sp. 4–5; pollen	Extinct/ancestral type
		<i>Quercus</i> sp. 6–7; pollen	New World Clade (aff. Group Protobalanus)
		<i>Quercus</i> sp. 1–3; pollen	<i>Quercus</i> Group <i>Quercus</i> and/or Lobatae
Baltic amber, northern Europe; ~45 Ma	Crepet and Nixon 1989; Weitschat and Wichard 2003	Flower and <i>in situ</i> pollen	<i>Quercus</i> Group <i>Quercus</i>
Königsau, near Aschersleben, Germany; middle Eocene (48–38 Ma)	Kvaček and Walther 1989	<i>Quercus subhercynica</i> H. Walther & Kvaček; leaf	<i>Quercus</i> Group Lobatae
Ube coal-field, southwestern Honshu, Japan; middle Eocene (48–38 Ma)	Huzioka and Takahasi 1970	<i>Cyclobalanopsis nathoi</i> Huzioka & Takahashi; leaf, acorn (?), cupule (?)	<i>Quercus</i> , affinity unclear
Changchang, Hainan; middle (?) Eocene (50–35 Ma)	Hofmann 2010	<i>Quercus</i> pollen types 2-8, 10; pollen	<i>Quercus</i> , affinity unclear (extinct, Group <i>Quercus</i> /Lobatae?, Group Protobalanus?)
		<i>Quercus</i> pollen type 1; pollen	<i>Quercus</i> Group <i>Ilex</i>
		<i>Quercus</i> pollen type 9; pollen	<i>Quercus</i> Group Cyclobalanopsis

2