

A peer-reviewed version of this preprint was published in PeerJ on 13 February 2014.

[View the peer-reviewed version](https://doi.org/10.7717/peerj.255) (peerj.com/articles/255), which is the preferred citable publication unless you specifically need to cite this preprint.

Proietti MC, Reisser J, Marins LF, Marcovaldi MA, Soares LS, Monteiro DS, Wijeratne S, Pattiaratchi C, Secchi ER. 2014. Hawksbill × loggerhead sea turtle hybrids at Bahia, Brazil: where do their offspring go? PeerJ 2:e255 <https://doi.org/10.7717/peerj.255>

Hawksbill x loggerhead sea turtle hybrids at Bahia, Brazil: where do their offspring go?

Maira C. Proietti¹, Julia Reisser^{2,3}, Luis F. Marins⁴, Maria A. Marcovaldi⁵, Luciano S. Soares⁶, Danielle S. Monteiro⁷, Sarath Wijeratne², Charitha Pattiaratchi² and Eduardo R. Secchi¹

¹Instituto de Oceanografia, Universidade Federal do Rio Grande, Rio Grande, Brazil

²School of Environmental Systems Engineering & Oceans Institute, University of Western Australia, Perth, Australia

³CSIRO Wealth from Oceans Flagship, Perth, Australia

⁴Instituto de Ciências Biológicas, Universidade Federal do Rio Grande, Rio Grande, Brazil

⁵Fundação Pró-Tamar, Praia do Forte, Brazil

⁶Archie Carr Center for Sea Turtle Research & Department of Biology, University of Florida, Gainesville, USA

⁷Núcleo de Educação e Monitoramento Ambiental, Rio Grande, Brazil

Correspondence to:

Maira C. Proietti

Instituto de Oceanografia, Universidade Federal do Rio Grande

Av. Itália KM 08, Rio Grande RS, 96201-900, Brazil

phone +55 53 81249203

email mairaproietti@gmail.com

Abstract

Hybridization between hawksbill (*Eretmochelys imbricata*) and loggerhead (*Caretta caretta*) breeding groups is unusually common in Bahia state, Brazil. Such hybridization is possible because hawksbill and loggerhead nesting activities overlap temporally and spatially along the coast of this state. Nevertheless, the destinations of their offspring are not yet known. This study is the first to identify immature hawksbill x loggerhead hybrids (n = 4) from this rookery by analyzing the mitochondrial DNA (mtDNA) of 157 immature turtles morphologically identified as hawksbills. We also compare for the first time modeled dispersal patterns of hawksbill, loggerhead, and hybrid offspring considering hatching season and oceanic phase duration of turtles. Particle movements varied according to season, with a

34 higher proportion of particles dispersing southwards throughout loggerhead and hybrid
35 hatching seasons, and northwards during hawksbill season. Hybrids from Bahia were not
36 present in important hawksbill feeding grounds of Brazil, being detected only at areas more
37 common for loggerheads. The genetic and oceanographic findings of this work indicate that
38 these immature hybrids, which are morphologically similar to hawksbills, could be adopting
39 behavioral traits typical of loggerheads, such as feeding in temperate waters of the western
40 South Atlantic. Understanding the distribution, ecology, and migrations of these hybrids is
41 essential for the development of adequate conservation and management plans.

43 Introduction

44 Interspecific hybridization occurs naturally or as a result of anthropogenic actions such
45 as habitat modification and fragmentation, species introduction, and population declines
46 (Rhymer and Simberloff, 1996; Allendorf et al., 2001). It is estimated that 25% of plant and
47 10% of animal species undergo hybridization (Mallet, 2005). This process can contribute to
48 the evolution of many taxa (Barton, 2001), but may also lead to lower fitness and fertility, and
49 even genetic extinction of species (Rhymer and Simberloff, 1996). In the marine environment
50 hybridization has been described for a range of organisms including corals (Willis et al.,
51 2006), fish (Hubbs, 2013), dolphins (Yazdi, 2002), seals (Kovacs, 1997), whales (Glover et
52 al., 2013) and sea turtles (Karl et al., 1995). Natural hybridization between Cheloniid sea
53 turtle species has been reported for green *Chelonia mydas* x hawksbill *Eretmochelys*
54 *imbricata*, loggerhead *Caretta caretta* x hawksbill, green x loggerhead, loggerhead x olive
55 ridley *Lepidochelys olivacea*, and olive ridley x hawksbill turtles (Wood et al., 1983;
56 Conceição et al., 1990; Karl et al., 1995; Seminoff et al., 2003; James et al., 2004; Lara-Ruiz
57 et al., 2006; Reis et al., 2010; Vilaça et al., 2012). Possible sterility and lower fitness of these
58 hybrids is concerning since all sea turtle species are currently threatened (IUCN 2012);
59 however, the exact causes and consequences of these hybridizations are not yet understood.

60 In Brazil, hawksbill and loggerhead breeding groups present exceptionally high
61 hybridization rates (Lara-Ruiz et al., 2006). The largest rookeries of both species overlap
62 along the coast of Bahia state, where approximately 420 hawksbills and 1240 loggerheads lay
63 their eggs each season (Marcovaldi and Chaloupka, 2007; Marcovaldi et al., 2007). They also
64 overlap temporally, with hawksbills nesting from November to March, and loggerheads from
65 September to February (Marcovaldi and Chaloupka, 2007; Marcovaldi et al., 2007). Studies
66 have shown that 42% of nesting females with hawksbill morphology were actually hybridized

with loggerheads, presenting the typical loggerhead mitochondrial DNA (mtDNA) haplotypes BR3 and BR4 (Lara-Ruiz et al., 2006). Since mtDNA is maternally inherited, the first generation (F1) of these hybrids is a cross between female loggerheads and male hawksbills; this could indicate a gender bias since to date no hybrids have presented hawksbill mtDNA (Vilaça and Santos, 2013). This bias has been attributed to the larger loggerhead population and the temporal overlap in nesting at the area. Since the hawksbill season begins around the loggerhead nesting peak (November – December), hawksbill males encounter an abundance of both hawksbill and loggerhead females for mating; meanwhile, by the time a large number of hawksbill females arrive, loggerhead males have already mated and left the area (Vilaça et al., 2012). Interestingly, the hawksbill x loggerhead hybrids are reproductively viable and produce hatchlings, possibly due to an ongoing introgression process (Lara-Ruiz et al., 2006; Vilaça et al., 2012).

After hatching, hawksbill turtles undergo an epipelagic dispersal stage followed by recruitment to tropical coastal areas (Bolten, 2003), usually coral or rocky reefs, where they feed preferably upon incrusting benthic organisms such as sponges and zoanthids (León and Bjørndal, 2002; Proietti, Reisser, and Secchi, 2012). Loggerheads also undergo an initial dispersal phase but are adapted to a broader latitudinal distribution range, recruiting to coastal or oceanic areas from tropical to temperate zones, where they feed mainly upon crustaceans, mollusks and fish (Davenport, 1997; Witzell, 2002). Immature loggerhead distribution in Brazil is not well known, but recognized high-use areas include the temperate waters along the southern continental shelf and the Rio Grande rise, a seamount located ca. 800 km off of the coast (Bugoni et al., 2003; Monteiro et al., 2006; Sales et al., 2008). High-occurrence hawksbill feeding areas include the oceanic islands of Rocas Atoll, Fernando de Noronha and São Pedro and São Paulo, and the coastal islands of the Abrolhos National Marine Park (Marcovaldi et al., 1998; Proietti, Reisser, and Secchi, 2012). The genetic characterization of hawksbills at these feeding grounds has until now been limited to Rocas Atoll and Fernando de Noronha, and one hybrid individual, representing a hawksbill x loggerhead hybrid backcrossed with a hawksbill (>F1 generation), was found. However it most likely originated from West Africa since it presented an mtDNA haplotype typical of hawksbills from São Tomé and Príncipe (Monzón-Argüello et al., 2011). Therefore, despite the elevated hybridization between these species in Bahia, how hybrid offspring disperse and where they recruit to is still a mystery. This is likely due to the relatively short timespan of this

phenomenon (~40 years, Lara-Ruiz et al., 2006) and limited surveys at hawksbill and loggerhead feeding grounds.

Understanding how hybridization affects the distribution and ecology of these animals is a complex task that is nevertheless fundamental when defining conservation strategies. In this work, we analyzed mtDNA of 157 immature turtles morphologically identified as hawksbills at high and occasional occurrence areas along the coast of Brazil, and modeled the dispersal patterns of turtles hatched at the Bahia rookery. We report for the first time immature hawksbill x loggerhead hybrids in Brazilian waters and show how temporal variability in hatching period leads to differences between the dispersal patterns of loggerhead, hawksbill, and hybrid offspring from Bahia. Finally, we consider the ecological and conservation implications of this exceptionally frequent phenomenon in Brazil.

Methods

Ethics statement: according to Normative Instruction 154/March 2007, all capture, tagging, sampling and transport of biological samples of wild animals for scientific purposes must have approval from Instituto Chico Mendes de Conservação da Biodiversidade (ICMBio) SISBIO committees. This study was approved by the Instituto Chico Mendes de Conservação da Biodiversidade, and conducted under SISBIO licenses #225043, #14122, and #159622. All animal handling was performed by trained personnel, following widely accepted and ethical protocols. When capturing live turtles, the following measures were taken to alleviate stress: 1) turtles were kept out of the water for a maximum of ten minutes; 2) work was performed in a shaded area; and 3) animals were released at the same location of capture.

We analyzed the mtDNA control region of 157 immature turtles morphologically identified as hawksbills from three important Brazilian hawksbill feeding grounds: (1) São Pedro and São Paulo Archipelago (SPSP; $n = 12$, Curved Carapace Length – CCL = 30 – 75 cm, mean 53.7 cm); (2) Bahia coast ($n = 32$, CCL = 21 – 72 cm, mean 39.7 cm), (3) Abrolhos National Marine Park ($n = 65$, CCL = 24.5 – 63.0 cm, mean 37.9 cm); as well as from three areas with sporadic occurrence of this species: (1) Arvoredo Biological Marine Biological Reserve ($n = 6$, CCL = 30 – 59.5 cm, mean 41.3 cm); (2) Ceará coast ($n = 23$, CCL = 22.4 – 57.5 cm, mean 37.8 cm); and (3) Cassino Beach ($n = 25$, CCL = 30 – 60 cm, mean 41 cm; Fig. 1). Loggerheads are not commonly observed at most of these areas (Reisser et al. 2008, Proietti, Reisser, & Secchi 2012), but occur at Ceará (Marcovaldi et al., 2012) and are frequently found at Cassino Beach (Bugoni et al., 2001; Monteiro et al., 2006). Samples were collected from turtles hand-captured in dives at SPSP, Abrolhos, and Arvoredo, and from

individuals incidentally caught in fishing nets or stranded on beaches (alive or dead) at Cear , Bahia, and Cassino.

Tissue samples were macerated and kept at 37 C in a lysis buffer containing Proteinase K until complete digestion (from 8 to 24 hours). DNA was extracted using Genomic DNA Extraction Kits (Norgen Biotek) or the phenol:chloroform method adapted from Hillis et al. (1996). mtDNA control region fragments of approximately 850 bp were amplified via Polymerase Chain Reaction (PCR) using primers LCM15382/H950 (Abreu-Grobois et al., 2006), under the following conditions: denaturation of 5' at 94 C; 36 cycles of 30'' at 94 C, 30'' at 50 C, 1' at 72 C; final extension of 10' at 72 C. Illustra GFX purification kits (GE Healthcare) were used for purification, and samples were sequenced in both directions through capillary electrophoresis using an Applied Biosystems  3130 Genetic Analyzer. Sequences were aligned and cropped to 740 bp using Clustal X 2.0 (Larkin et al., 2007), and classified according to GenBank  and the Atlantic Ocean hawksbill haplotype database (A. Abreu-Gobrois, pers. comm., 2013).

Biophysical modeling was performed using the particle-tracking tool ICHTHYOP-3.2 (<http://www.previmer.org/en/ichthyop>), see model description in Lett et al. (2008) for details. Surface velocity fields were extracted from the global HYbrid Coordinate Ocean Model (HyCOM) with 1/12  reanalysis outputs at daily intervals (<http://hycom.org>). We chose the fourth-order Runge-Kutta numerical scheme in ICHTHYOP-3.2 to simulate Lagrangian advection of individual particles. The numerical time step was set to 180 seconds and particle trajectory position outputs were set to daily intervals. Particles were released every 5 days from the Bahia rookery (12-13  S, 37-38  W) proportionally to the monthly amount of hatched loggerheads, hawksbills, and hybrids. Particles were tracked for three years (between May 2009 to June 2013) to encompass the oceanic phase of these sea turtles, following Putman and He (2013).

The monthly proportion of nesting loggerheads and "hawksbills" (including pure and hybrids) were obtained from Marcovaldi and Chaloupka (2007) and Marcovaldi et al. (1999). We then multiplied the monthly number of nesting animals identified as hawksbills (Marcovaldi et al., 1999) by the monthly percentage of genetically-confirmed hybrid and pure hawksbills (Lara-Ruiz et al., 2006; L. Soares, unpublished data). The hatching periods of loggerheads, hawksbills, and hybrids were calculated by adding 60 days (approximate incubation period; Godfrey et al., 1999, Marcovaldi et al., 1997) to their estimated nesting

164 periods. Finally, the proportion of particles dispersing southwards and northwards was
165 analyzed.

166 **Results**

167 Of the 157 individuals sampled along the coast, four were hawksbill x loggerhead
168 hybrids. Most of these hybrids presented the morphology of pure hawksbill turtles (Fig. 2)
169 and were identified as such, but their mtDNA haplotype was characteristic of nesting
170 loggerheads of the Bahia rookery (BR3). This haplotype was present in one of 23 samples
171 from Ceará (northeast Brazil), and in three of 19 samples from Cassino in the far South (Fig.
172 1). At Ceará, the hybrid was sampled after being incidentally caught in fisheries, and at
173 Cassino all three hybrids were found dead on the beach. At Cassino one hybrid displayed
174 carapace with overlapping scutes and serrated edges like hawksbills, but a short and thick
175 neck typical of loggerheads (Fig. 2a). This mixed morphology brings additional evidence of
176 this crossbreeding.

177 Trajectories of simulated virtual particles are shown in Fig. 3. A large proportion of
178 particles moved to the South when released during loggerhead hatching peak (72%;
179 December – March), reaching temperate waters of the western South Atlantic via the Brazil
180 current. Particles released during hybrid hatching peak (January – April) showed a higher
181 southwards displacement (44%) when compared to the hawksbill peak (37%; February –
182 May). Northwards dispersal was higher for particles released during hawksbill (63%),
183 followed by hybrid (56%) and loggerhead (26%) peak hatching seasons.

184 **Discussion**

185 In this work we begin to answer a fundamental question that arises when facing the
186 considerable portion of hybrids that nest in Brazil: where do their hatchlings go? Although
187 immature hybrids from the Bahia rookery remain highly undetected relative to the
188 considerable number that is generated, reporting their occurrence at loggerhead feeding
189 grounds (Cassino Beach and Ceará) and their absence at important hawksbill feeding grounds
190 (e.g. Abrolhos, SPSP) is an important step towards better understanding this phenomenon (see
191 Fig. 1). Our modeling approach also highlights the importance of sea turtle nesting season on
192 shaping the spatial distribution of post-hatchlings, with differences observed between
193 hawksbill, loggerhead and hybrid dispersal (see Fig. 3).

194 While immature hybrids were observed at areas uncommon for hawksbills, they were
195 absent at recognized high-occurrence feeding grounds such as Fernando de Noronha and
196 Abrolhos (this study; Vilaça et al., 2013). Despite the relatively large sample ($n = 65$) from

the tropical reefs of Abrolhos, located very close to the Bahia rookery (ca. 80 km), no hybrids were detected. This could indicate that while these hybrids are morphologically similar to hawksbills, they are not recruiting to the same feeding grounds of pure hawksbills. Three hybrids were found at Cassino Beach, a temperate sandy coast that lacks the optimal characteristics for hawksbill survival (e.g. abundance of preferred food items, relatively high temperatures; Davenport, 1997) and possess few records of this species (Monteiro et al., 2006). Loggerheads on the other hand are commonly found foraging at this region, suggesting that immature hybrids could be adopting the feeding and migration ecology of loggerheads. Similarly, Witzell and Schmid (2003) reported the occurrence of an immature hawksbill x loggerhead hybrid that established its home range in a loggerhead feeding ground.

Adult hawksbill x loggerhead hybrids from Bahia have also been shown to present a distinct ecology when compared to their pure hawksbill counterparts. Marcovaldi et al. (2012) tracked pure hawksbills and hawksbill x loggerhead hybrids after nesting in Bahia and showed different post-nesting migration patterns. Most tracked animals moved along the continental shelf, with all pure hawksbills occupying feeding areas along the eastern coast (Bahia and Alagoas states) while most hybrid females travelled to the northern coast, including Ceará where we detected an immature hybrid. Ceará is an important feeding ground for loggerheads that nest along the coast of Bahia as demonstrated by satellite tracking (Marcovaldi et al., 2010), indicating that the mature female hybrids adopt the behavior of loggerheads. This could also be a possibility for the immature hybrid we detected at the area.

Our biophysical simulations showed that post-hatchling dispersal from Bahia varied according to species: southwards dispersal was proportionally larger throughout loggerhead, followed by hybrid, and lowest during hawksbill peak hatching season. The factors influencing how hybrid sea turtles adopt different feeding and migration behaviors are unknown. Ocean currents influence the dispersal of sea turtle post-hatchlings and are believed to shape the posterior spatial distribution of juveniles and adults (Luschi et al., 2003; Amorocho et al., 2012; Proietti et al., 2012b; Putman et al., 2012, 2014; Putman and He, 2013). The model presented here shows that hybrids could have a higher chance of reaching the temperate waters of South Brazil when compared to pure hawksbills. This indicates that these hybrids could already be adopting loggerhead features once they reach the water after hatching. Although pure hawksbills also produce southwards-dispersing hatchlings, they could be limited to lower latitudes by food availability and water temperature, while hybrids

could present a behavioral pattern more similar to loggerheads and possibly occupy a wider niche.

The causes behind the extensive hybridization between hawksbills and loggerheads at the Bahia rookery are still unclear, but could be a result of anthropogenic population declines and uneven population sizes of different species (Lara-Ruiz et al., 2006; Vilaça et al., 2012). It is unknown if this hybridization is threatening the fitness and survival of animals, and the phenomenon should be further investigated for defining whether special measures should be taken when managing these populations. International collaboration might be necessary for determining such management approaches since our particle model shows that ocean currents could transport hybrid turtles from Bahia to distant areas such as Uruguay, Argentina, West African coast, and Western Indian region. Extensive genetic studies in areas of recognized and potential hybrid occurrence, such as loggerhead habitats, are of utmost importance. These studies should combine mtDNA with biparentally-inherited marker analyses for obtaining a better understanding of hawksbill x loggerhead hybrid distribution, parental species and generations. Studies on reproductive and survivorship parameters are also essential for verifying potential negative impacts of this process on long-term viability of local sea turtle populations. Satellite tracking, stable isotopes and diet analyses can also be used to confirm if their movements and feeding habits follow a distinctive pattern. Such studies would provide valuable insight on how the ecology and behavior of sea turtles are affected by hybridization, and consequently guide management practices and strategies to conserve their populations.

Acknowledgements

M.C.P. is a graduate student of the Programa de Pós-graduação em Oceanografia Biológica (FURG), and is sponsored by Coordenação de Aperfeiçoamento de Pessoal de Ensino Superior in Brazil (CAPES). J.R. is sponsored by the International Postgraduate Research Scholarship (IPRS) and CSIRO's Flagship Postgraduate Scholarship, and E.R.S. is sponsored by CNPq (307843/2011-4). This work was possible thanks to financial support from the Rufford Small Grants (RSG – UK, Grant #8110-2). We thank ICMBio, Pata da Cobra Diving, Brazilian Navy, CECIRM PRO-Arquipélago, Abrolhos Park coordination, and all field assistants (a special thanks to B. Barbosa) for logistic/field support. We acknowledge Núcleo de Educação e Monitoramento Ambiental (NEMA), Centro de Recuperação de Animais Marinhos (CRAM) and Projeto Tamar for providing samples. This is a contribution of the Research Group 'Ecologia e Conservação da Megafauna Marinha – EcoMega'.

263 References

- 264 Abreu-Grobois F, Horrocks J, Formia A, Dutton P, LeRoux R, Vélez-Zuazo X, Soares L,
265 Meylan P (2006) New mtDNA Dloop primers which work for a variety of marine turtle
266 species may increase the resolution of mixed stock analyses. In: Frick M, Panagopoulou
267 A, Rees A, Williams K (eds) Book of Abstracts, Twenty-sixth Annual Symposium on
268 Sea Turtle Biology and Conservation. International Sea Turtle Society, Athens, Greece,
269 p 179
- 270 Allendorf FW, Leary RF, Spruell P, Wenburg JK (2001) The problems with hybrids: setting
271 conservation guidelines. *Trends Ecol Evol* 16:613–622
- 272 Amorocho DF, Abreu-Grobois FA, Dutton PH, Reina RD (2012) Multiple distant origins for
273 green sea turtles aggregating off Gorgona Island in the Colombian eastern Pacific. *PLoS*
274 *One* 7:e31486
- 275 Barton NH (2001) The role of hybridization in evolution. *Mol Ecol* 10:551–68
- 276 Bérubé M, Aguilar A (1998) A new hybrid between a blue whale, *Balaenoptera musculus*,
277 and a fin whale, *B. physalus*: frequency and implications of hybridization. *Mar Mammal*
278 *Sci* 14:82–98
- 279 Bolten A (2003) Variation in sea turtle life history patterns: neritic vs. oceanic developmental
280 stages. In: Lutz P, Musick J, Wyneken J (eds) *The Biology of Sea Turtles*, Vol 2. CRC
281 Press, Boca Raton, FL, p 243–257
- 282 Bugoni L, Krause L, Petry M V (2001) Marine debris and human impacts on sea turtles in
283 southern Brazil. *Mar Pollut Bull* 42:1330–4
- 284 Bugoni L, Krause L, Petry MV (2003) Diet of sea turtles in southern Brazil. *Chelonian*
285 *Conserv Biol* 4
- 286 Conceição M, Levy J, Marins L, Marcovaldi M (1990) Electrophoretic characterization of a
287 hybrid between *Eretmochelys imbricata* and *Caretta caretta* (Cheloniidae). *Comp*
288 *Biochem Physiol B Biochem Mol Biol* 97B:275–278
- 289 Davenport J (1997) Temperature and the life-history strategies of sea turtles. *J Therm Biol*
290 22:479–488
- 291 Glover KA, Kanda N, Haug T, Pastene LA, Øien N, Seliussen BB, Sørvik AGE, Skaug HJ
292 (2013) Hybrids between common and Antarctic minke whales are fertile and can back-
293 cross. *BMC Genet* 14:25
- 294 Godfrey MH, Amato AFD, Marcovaldi MÂ, Mrosovsky N (1999) Pivotal temperature and
295 predicted sex ratios for hatchling hawksbill turtles from Brazil. *Can J Zool* 77:1465–
296 1473

- 297 Hillis D, Mable B, Larson A, Davis S, Zimmer E (1996) Nucleic acids IV: sequencing and
298 cloning. In: Hillis D, Moritz C, Mable B (eds) Molecular systematics, 2nd edition.
299 Sinauer Associates, Sunderland, MA, p 321–381
- 300 Hubbs CL (2013) Hybridization between fish species in nature. *Syst Zool* 4:1–20
- 301 IUCN (2012) The IUCN Red List of Threatened Species, version 2012.2 <iucnredlist.org>
302 Accessed on July 21, 2013.
- 303 James M, Martin K, Dutton P (2004) Hybridization between a green turtle, *Chelonia mydas*,
304 and a loggerhead turtle, *Caretta caretta*, and the first record of a green turtle in Atlantic
305 Canada. *Can F Nat* 118:579–582
- 306 Karl S, Bowen B, Avise J (1995) Hybridization among the ancient mariners: characterization
307 of marine turtle hybrids with molecular genetic assays. *J Hered* 86:262–8
- 308 Kovacs K (1997) A harp seal x hooded seal hybrid. *Mar Mammal Sci* 13:460–468
- 309 Lara-Ruiz P, Lopez GG, Santos FR, Soares LS (2006) Extensive hybridization in hawksbill
310 turtles (*Eretmochelys imbricata*) nesting in Brazil revealed by mtDNA analyses. *Conserv*
311 *Genet* 7:773–781
- 312 Larkin MA, Blackshields G, Brown NP, Chenna R, McGettigan PA, McWilliam H, Valentin
313 F, Wallace IM, Wilm A, Lopez R, Thompson JD, Gibson TJ, Higgins DG (2007) Clustal
314 W and Clustal X version 2.0. *Bioinformatics* 23:2947–8
- 315 León YM, Bjørndal KA (2002) Selective feeding in the hawksbill turtle, an important
316 predator in coral reef ecosystems. *Mar Ecol Prog Ser* 245:249–258
- 317 Lett C, Verley P, Mullon C, Parada C, Brochier T, Penven P, Blanke B (2008) A Lagrangian
318 tool for modelling ichthyoplankton dynamics. *Environ Model Softw* 23:1210–1214
- 319 Luschi P, Hays GC, Papi F (2003) A review of long-distance movements by marine turtles,
320 and the possible role of ocean currents. *Oikos* 103:293–302
- 321 Mallet J (2005) Hybridization as an invasion of the genome. *Trends Ecol Evol* 20:229–237
- 322 Marcovaldi MÂ, Baptistotte C, Castilhos JC, Gallo BMG, Lima EHSM, Sanches TM, Vieitas
323 CF (1998) Activities by Project TAMAR in Brazilian Sea Turtle Feeding Grounds. *Mar*
324 *Turt Newsl* 80:5–7
- 325 Marcovaldi M, Chaloupka M (2007) Conservation status of the loggerhead sea turtle in
326 Brazil: an encouraging outlook. *Endanger Species Res* 3:133–143
- 327 Marcovaldi MA, Godfrey MH, Mrosovsky N (1997) Estimating sex ratios of loggerhead
328 turtles in Brazil from pivotal incubation durations. *Can J Zool* 75:755–770

- 329 Marcovaldi M, Lopez G, Soares L, Lima E, Thomé J, Almeida A (2010) Satellite-tracking of
330 female loggerhead turtles highlights fidelity behavior in northeastern Brazil. *Endanger*
331 *Species Res* 12:263–272
- 332 Marcovaldi M, Lopez G, Soares L, López-Mendilaharsu M (2012) Satellite tracking of
333 hawksbill turtles *Eretmochelys imbricata* nesting in northern Bahia, Brazil: turtle
334 movements and foraging destinations. *Endanger Species Res* 17:123–132
- 335 Marcovaldi MA, Lopez GG, Soares LS, Santos AJB, Bellini C, Barata PCR (2007) Fifteen
336 years of hawksbill sea turtle (*Eretmochelys imbricata*) nesting in Northern Brazil.
337 *Chelonian Conserv Biol* 6:223–228
- 338 Marcovaldi M, Vieitas CF, Godfrey MH (1999) Nesting and conservation management of
339 hawksbill turtles (*Eretmochelys imbricata*) in northern Bahia, Brazil. *Chelonian Conserv*
340 *Biol* 3:301–307
- 341 Monteiro D, Bugoni L, Estima S (2006) Strandings and sea turtle fisheries interactions along
342 the coast of Rio Grande do Sul state, Brazil. In: Frick M, Panagopoulou A, Rees A,
343 Williams K (eds) *Book of Abstracts, Twenty-sixth Annual Symposium on Sea Turtle*
344 *Biology and Conservation*. International Sea Turtle Society, Athens, Greece, p 257
- 345 Monzón-Argüello C, Loureiro NS, Delgado C, Marco A, Lopes JM, Gomes MG, Abreu-
346 Grobois FA (2011) Principe island hawksbills: genetic isolation of an eastern Atlantic
347 stock. *J Exp Mar Bio Ecol* 407:345–354
- 348 Proietti M, Reisser J, Kinas P, Kerr R, Monteiro D, Marins L, Secchi E (2012) Green turtle
349 *Chelonia mydas* mixed stocks in the western South Atlantic, as revealed by mtDNA
350 haplotypes and drifter trajectories. *Mar Ecol Prog Ser* 447:195–209
- 351 Proietti MC, Reisser J, Secchi ER (2012) Foraging by immature hawksbill sea turtles at
352 Brazilian islands. *Mar Turt Newsl* 135:4–6
- 353 Putman NF, Abreu-grobois FA, Broderick AC, Cio C, Formia A, Godley BJ, Stroud S,
354 Pelembe T, Verley P, Williams N (2014) Numerical dispersal simulations and genetics
355 help explain the origin of hawksbill sea turtles in Ascension Island. *J Exp Mar Bio Ecol*
- 356 Putman N, He R (2013) Tracking the long-distance dispersal of marine organisms: sensitivity
357 to ocean model resolution. *J R Soc Interface* 10
- 358 Putman NF, Verley P, Shay TJ, Lohmann KJ (2012) Simulating transoceanic migrations of
359 young loggerhead sea turtles: merging magnetic navigation behavior with an ocean
360 circulation model. *J Exp Biol* 215:1863–1870
- 361 Reis EC, Soares LS, Lôbo-Hajdu G (2010) Evidence of olive ridley mitochondrial genome
362 introgression into loggerhead turtle rookeries of Sergipe, Brazil. *Conserv Genet*
363 11:1587–1591
- 364 Reis EC, Soares LS, Vargas SM, Santos FR, Young RJ, Bjørndal K a., Bolten AB, Lôbo-
365 Hajdu G (2009) Genetic composition, population structure and phylogeography of the

366 loggerhead sea turtle: colonization hypothesis for the Brazilian rookeries. *Conserv Genet*
367 11:1467–1477

368 Reisser J, Proietti M, Kinas P, Sazima I (2008) Photographic identification of sea turtles:
369 method description and validation, with an estimation of tag loss. *Endanger Species Res*
370 5:73–82

371 Rhymer M, Simberloff D (1996) Extinction by hybridization and introgression. *Annu Rev*
372 *Ecol Syst* 27:83–109

373 Sales G, Giffoni B, Barata P (2008) Incidental catch of sea turtles by the Brazilian pelagic
374 longline fishery. *J Mar Biol Assoc UK* 88:853–864

375 Seminoff JA, Karl SA, Schwartz T, Resendiz A (2003) Hybridization of the green turtle
376 (*Chelonia mydas*) and hawksbill turtle (*Eretmochelys imbricata*) in the Pacific Ocean:
377 indication of absence of gender bias in the directionality of crosses. *Bull Mar Sci*
378 73:643–652

379 Vilaça ST, Lara-Ruiz P, Marcovaldi MA, Soares LS, Santos FR (2013) Population origin and
380 historical demography in hawksbill (*Eretmochelys imbricata*) feeding and nesting
381 aggregates from Brazil. *J Exp Mar Bio Ecol* 446:334–344

382 Vilaça ST, Santos FR dos (2013) Molecular data for the sea turtle population in Brazil.
383 *Dataset Pap Sci* 2013:1–7

384 Vilaça ST, Vargas SM, Lara-Ruiz P, Molfetti É, Reis EC, Lôbo-Hajdu G, Soares LS, Santos
385 FR (2012) Nuclear markers reveal a complex introgression pattern among marine turtle
386 species on the Brazilian coast. *Mol Ecol* 21:4300–4312

387 Willis BL, Oppen MJH van, Miller DJ, Vollmer S V., Ayre DJ (2006) The role of
388 hybridization in the evolution of reef corals. *Annu Rev Ecol Evol Syst* 37:489–517

389 Witzell W (2002) Immature Atlantic loggerhead turtles (*Caretta caretta*): suggested changes
390 to the life history model. *Herpetol Rev* 33:266–269

391 Witzell WN, Schmid JR (2003) Multiple recaptures of a hybrid hawksbill-loggerhead turtle in
392 the Ten Thousand Islands, Southwest Florida. *Herpetol Rev* 34:323–325

393 Wood J, Wood F, Critchley K (1983) Hybridization of *Chelonia mydas* and *Eretmochelys*
394 *imbricata*. *Copeia* 1983:839–842

395 Yazdi P (2002) A possible hybrid between the dusky dolphin (*Lagenorhynchus obscurus*) and
396 the southern right whale dolphin (*Lissodelphis peronii*). *Aquat Mamm* 28:211–217

397

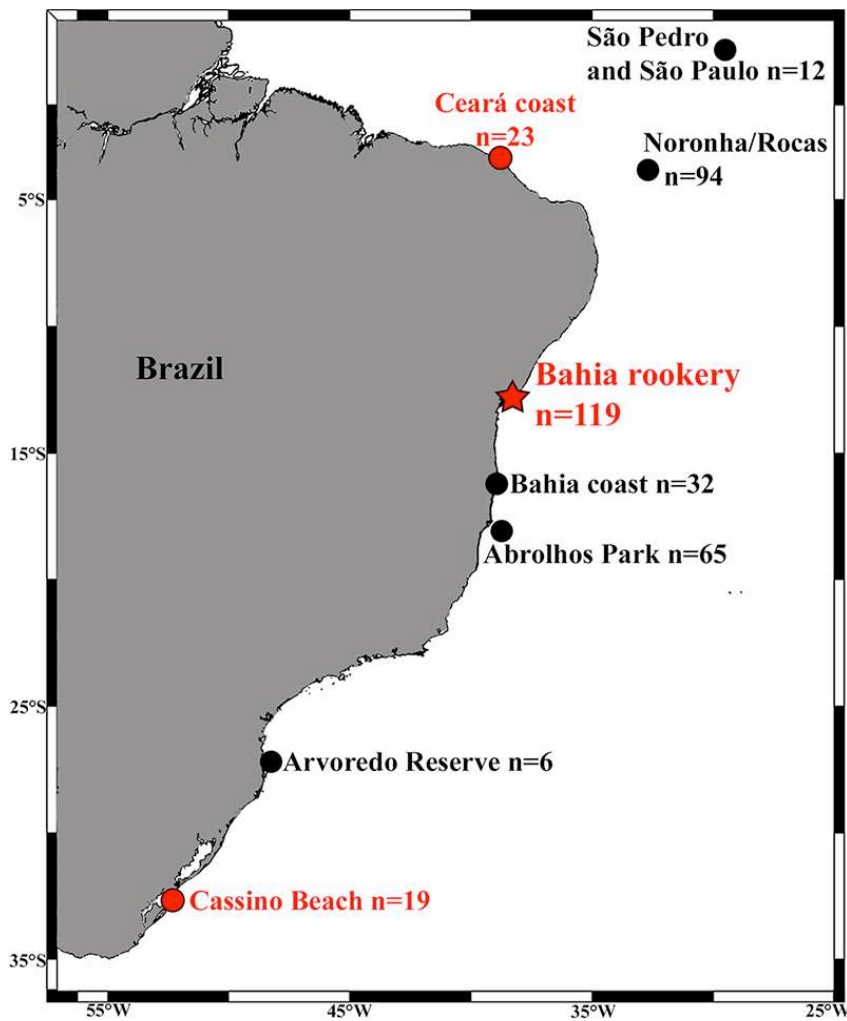


Figure 1. Locations and sample sizes of genetically-described immature hawksbill areas (dots) and the Bahia rookery (red star), in Brazil. Red dots indicate detection of hawksbill x loggerhead sea turtle hybrids from the Bahia rookery.

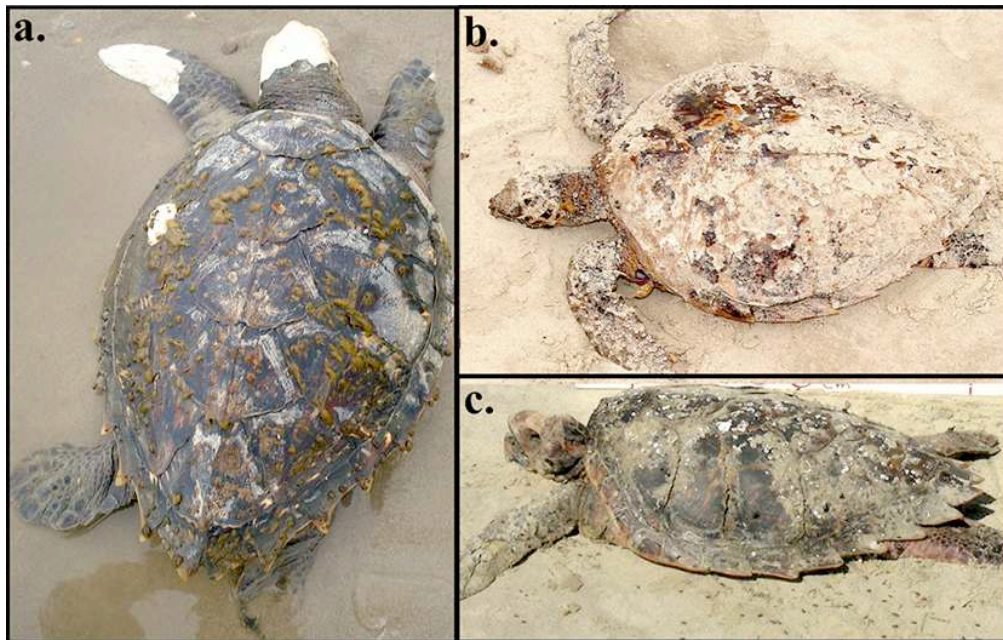


Figure 2. Sampled hawksbill x loggerhead sea turtles at Cassino Beach, South Brazil. Note the relatively large head and thick neck of the individual in a. Photo credits: Nema archive (a,b) and Jonatas H. Prado (c).

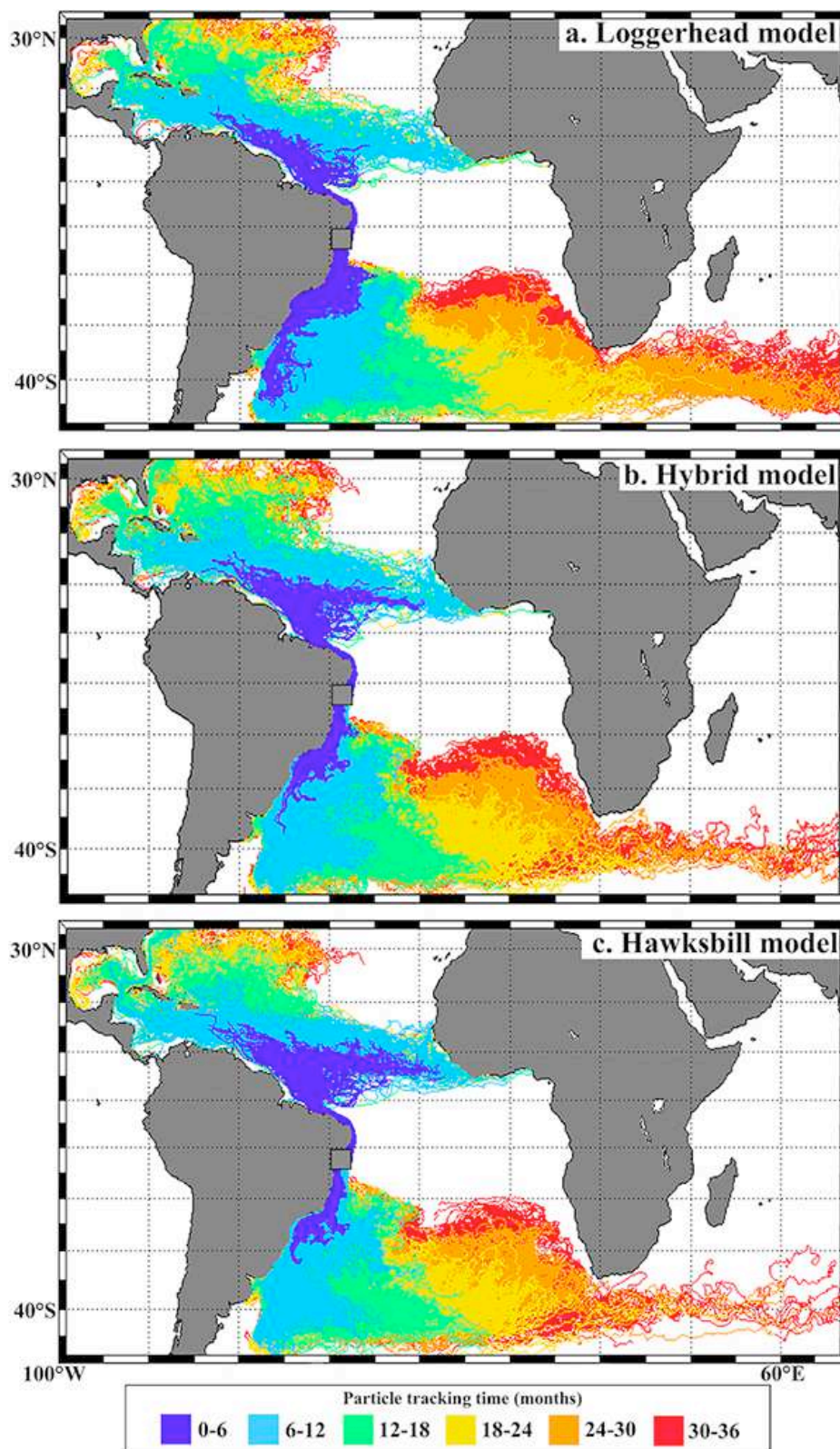


Figure 3. Virtual particles leaving the Bahia rookery during loggerhead (a), hybrid (b) and hawksbill (c) hatching seasons.