

1 **Mitochondrial phylogenomics and genetic population structure of anchovies**  
2 **(*Engraulis encrasicolus*) along the Moroccan coast using sequence analysis of**  
3 **the mitochondrial DNA cytochrome b**

4 Khalil Chahdi Ouazzani<sup>1</sup>, Touria Benazzou<sup>1</sup>, Naoki Tojo<sup>2</sup>, Malika Chlaida<sup>3</sup>

5 <sup>1</sup>Département de Biologie, Faculté des Sciences Rabat, Université Mohammed V, Rabat/  
6 Morocco.

7 <sup>2</sup>Japanese International Cooperation Agency/Japan.

8 <sup>3</sup>Institut National de Recherche Halieutique (INRH), Casablanca/ Morocco.

9 Corresponding author:

10 Malika Chlaida

11 Route Sidi Abderrahmane Club équestre Ould Jmel, Casablanca/Morocco

12 Email address: [ma\\_chlaida@hotmail.com](mailto:ma_chlaida@hotmail.com)

13 **Abstract** A fragment of 680 bp of mitochondrial cytochrome b locus of European anchovies,  
14 *Engraulis encrasicolus*, was sequenced for 138 individuals collected from three Moroccan  
15 Atlantic areas and from Moroccan Alboran Sea. These samples were surveyed for diversity and  
16 differentiation with a range of summary statistics. The results showed that the most dominant  
17 clade in Moroccan anchovy is Clade A with a percentage ranging from 89% in Alboran Sea to 91%  
18 - 95 % in the Moroccan Atlantic coast. Overall, there was a significant genetic differentiation  
19 among the 4 Moroccan anchovy zones ( $\Phi_{st} = 0.01283$ ;  $p = 0.03910$ ). Pairwise  $\Phi_{st}$  among  
20 populations and multidimensional scaling revealed a high homogeneity among Atlantic  
21 populations but some heterogeneity between Alboran population and Atlantic populations,  
22 mainly between the populations from Central Atlantic of Morocco with a significant difference.

23 **Keywords:** European anchovy (*Engraulis encrasicolus*), Morocco, mitochondrial cytochrome b,  
24 phylogeography, population structure

## 25 INTRODUCTION

26 The European anchovy, *Engraulis encrasicolus* (Linnaeus, 1758) is a small pelagic fish  
27 found in a wide range of temperatures (2–30 °C) and salinities (5–41 psu) in the eastern Atlantic,  
28 the Mediterranean Sea and the Black Sea (Whitehead, Nelson & Wongratana, 1988). This species

29 plays a major socio-economic role in all regions; it is one of the principal target species for  
30 commercial fisheries. Indeed, European anchovies with other Engraulidae fisheries representing  
31 14 percent of the world catch of fish (Ababouch & El Marrakchi, 2009). In regard to their  
32 economic importance, anchovies like other pelagic fish occupy crucial positions in the oceans  
33 ecosystems (Ganias, 2014) particularly in upwelling regions where it occupies fundamental  
34 intermediate trophic level (Bakun, 2006). As is known, small pelagic are characterized as ‘wasp-  
35 waist’, being considered as crucial components of pelagic ecosystems (Cury et al., 2000). In  
36 addition, anchovy as a pelagic species, it can achieve considerable biomass and undergoes crash-  
37 flush cycles (Uriarte, Prouzet & Villamor, 1996; Lavoué et al., 2007). For these reasons, the  
38 management of this precious resource is important. In fact, anchovies have been the subject of  
39 many genetic studies aimed a better understanding of the stock genetic structure as identifying  
40 stocks, discriminating among them, and determining the stock composition of mixed stocks are  
41 integral elements of fishery management (Waldman, 1999). Likewise, genetics and fishery  
42 management can interact in several ways. When the genetic population structure of a species is  
43 known, the distribution of subpopulations in mixed fisheries can be estimated (Utter, 1991).

44 Previous published data using allozymes, mitochondrial DNA (mtDNA) RFLPs, control  
45 region sequence, microsatellites and SNP marker (Bembo et al., 1996a; Bembo et al., 1996b;  
46 Bembo et al., 1996c; Magoulas, Tsimenides & Zouros, 1996; Magoulas et al., 2006;  
47 Kristoffersen & Magoulas, 2008; Tudela, Garcia-Marin & Pla, 1999; Sanz et al., 2008;  
48 Zarraonaindia et al., 2009; Borrell et al., 2012; Vinas et al., 2013; Silva, Horne & Castilho, 2014)  
49 suggest the presence of various genetically differentiated groups with spatial and ecological  
50 components. In addition to this, Magoulas et al. (1996; 2006) by analyzing mitochondrial DNA  
51 restriction fragment length polymorphism, reported a significant phylogeographic structure in  
52 both the Atlantic and Mediterranean populations of anchovy with two haplotype clades (A and B)  
53 separated by 3.2% sequence divergence. Grant et al. (2005); Vinas et al. (2013); Oueslati et al.  
54 (2014) and Silva, Horne & Castilho (2014) confirmed the existence of these clades whose  
55 frequency varies within samples across the Mediterranean and Northeast Atlantic.

56 Along the Moroccan coasts, anchovies fisheries are very important with thousands tons  
57 annually cashed. Despite the importance of this species, no widespread genetic studies have  
58 focused on the European anchovy population from Moroccan coasts. The objective of this work  
59 is to deal with genetic structure of anchovy from this region by using Mitochondrial DNA and to

60 examine the hypotheses of existence of tow clades (A and B) in the Moroccan coasts as it was  
61 reported in other region.

## 62 MATERIALS AND METHODS

### 63 Fish Sampling

64 Fish was carried out; using INRH RV “Amir Moulay Abdallah” during species spawning  
65 period in 2012, from three Moroccan Atlantic locations, the fourth sample was collected from  
66 Moroccan Alboran Sea (Figure 1). (See Field Study Permissions in additional information and  
67 declarations)

### 68 DNA extraction, amplification and sequencing

69 DNA was extracted from 25 mg of fin using the “QIAGEN DNeasy Blood & Tissue kit”  
70 following the manufacturer’s recommendations. A total of 138 individuals were used for mtDNA  
71 analyses (Table 1 and Figure 1). For PCR, a fragment of 680 bp was amplified with an initial  
72 denaturation at 95 °C for 10 min, followed by 35 cycles (95 °C 60 s; 52 °C 1 min, 72 °C 1 min)  
73 with a final extension at 72 °C for 10 min using the primers 5'-AACGACGCAGTAGTAGACC'  
74 and 5'-GAGGAAGTATCACTCAGGC- defined in positions 42 and 825 of the cyt-b locus of  
75 *E. encrasicolus*, respectively. The amplification products were sequenced at the  
76 CNRST/Morocco Platform and 680 bp aligned for individuals between positions 108 and 787 of  
77 the gene in BIOEDIT (Hall, 1999) with ClustalW, using the mtDNA cytochrome b sequence of  
78 the European anchovy (*Engraulis encrasicolus*) (sequence accession numbers GENBANK  
79 KF873783 and KF874179) as a reference with manual adjustments (Oueslati et al., 2014).

### 80 Data treatment

81 DnaSP version 5.10.1 (Librado, & Rozas, 2009) was used to calculate nucleotide diversity  
82 per site ( $\pi$ ) (Nei, 1987) and haplotype diversity (h) (Nei & Tajima, 1981) from haplotype  
83 frequencies and haplotype divergence. The Tajima’s D (Tajima, 1989) was also estimated to  
84 assess the likelihood that the DNA sequences have evolved in a neutral manner with significance  
85 tests (1 000 simulations) (significant,  $P < 0.05$ ).

86 The phylogenetic relationships among mDNA cyto b haplotypes were evaluated by  
87 neighbour-joining (NJ) analysis using MEGA 6.06 (Tamura et al., 2013) (Figure 2). Also, the  
88 relationship between haplotypes can be understood by Median joining haplotype networks  
89 estimated for European anchovy for each clade from four sites in the Moroccan coast (Figure 3).

90 These haplotypes networks are made using the Network 4.1 software (available at: [www.fluxus-](http://www.fluxus-technology.com)  
91 [technology.com](http://www.fluxus-technology.com)). The construction of these networks is based on the theory of coalescence  
92 (Kingman, 2000) and combines features of Kruskal's algorithm for finding minimum spanning  
93 trees by choosing short connections, and Farris's maximum-parsimony (MP) heuristic algorithm  
94 (Bandelt, Forster & Röhl, 1999).

95 Pairwise genetic differentiation between samples ( $\Phi_{st}$ ) was estimated in Arlequin 3.5.3.1  
96 (Excoffier & Lischer, 2010) on 10 000 permutations. In addition, the relationships between  
97 samples were estimated by multidimensional scaling (MDS) analysis of the pairwise  $\Phi_{st}$  values.  
98 To determine homogenous groups of populations, analysis of molecular variance "AMOVA" of  
99 haplotypes was carried out in Arlequin 3.5.3.1 with 1 000 permutations to establish significance.

## 100 RESULTS

101 Among all the samples, the Clade A was dominant. Thus, 92% had a haplotype from Clade  
102 A and 8% had a haplotype from Clade B (Figure 2). The frequency of Clade A was high (89%) in  
103 the Alboran Sea, but its prevalence was lower than in the Moroccan Atlantic coast with a  
104 percentage that ranged from 91% in Southern Atlantic to 98% in the Central Atlantic. The  
105 median-joining haplotype network revealed two major clades (A and B) separated by 14  
106 mutational steps and showed a star-like genealogy (Figure 3), which means that the European  
107 Anchovy population increases. This appears consistent with the values of Tajima's D, significant  
108 negative for each population independently, or for all individuals (Table 1).

109 The polymorphisms investigation defined 118 distinct European anchovy mtDNA  
110 haplotypes and 147 segregating sites (S), 59 of which were parsimoniously informative sites.  
111 Higher levels of genetic variation were found for the haplotype diversity  $h = 0.9868 \pm 0.0061$ ,  
112 but a concatenated mean nucleotide diversity for all samples was  $0.008946 \pm 0.004740$  (Table 1).  
113 All locations had a close haplotype diversity, whereas for the nucleotide diversity the Moroccan  
114 Atlantic samples were less variable than the Alboran samples, which had the higher nucleotide  
115 diversity ( $\pi = 0.01185 \pm 0.00166$ ). Under the infinite mutation model, significant negative values  
116 of Tajima's D indicate an excess of low-frequency haplotypes that can arise from selection or  
117 rapid population growth (Tajima, 1989).

118 Overall, there was significant genetic differentiation among the 4 populations  
119 ( $\Phi_{st} = 0.01283$ ;  $p = 0.03519$ ). Pairwise  $\Phi_{st}$  revealed a strong homogeneity within Atlantic  
120 populations, but less heterogeneity between Alboran population and Atlantic populations,

121 especially between Moroccan Central Atlantic which showed a significant difference (Table 2).  
122 When using only Clade A haplotypes, the genetic structure stills significant between the  
123 populations  $F_{st}$  ( $\Phi_{st}$  0.01619,  $p$  very highly significant). Also the pairwise  $\Phi_{st}$  were significantly  
124 different between Alboran population and Moroccan Atlantic populations (Table 2). A multi-  
125 dimensional scaling (MDS) ordination, with a stress coefficient 0.001, using all dataset or  
126 sequences of Clade A, showed that the Northern Atlantic, Central Atlantic and Southern Atlantic  
127 populations of Morocco were structured as a unique unit separated from Moroccan Alboran  
128 population (Figure 5).

129 Using only Clade A sequences, the AMOVA test explained 3.29% of total variation among  
130 groups (group 1: Alboran samples and group 2: Atlantic samples) and was significant  
131 ( $p=0.00587$ ). This percentage decreases when we include all dataset (2.53 %,  $p=0.00489$ )  
132 (Table 3).

## 133 DISCUSSION AND CONCLUSION

### 134 Historical phylogeography

135 The result of this study showed that the most dominant clade in Moroccan anchovy was  
136 Clade A (about 90 %). The origin and expansion of this Clade was previously described by  
137 several authors. Magoulas et al. (1996; 2006), by using RFLPs in mtDNA reported two divergent  
138 clades (A and B), which differed by 3.2% sequence divergence. The authors postulated that  
139 Clade A originated in the Atlantic Sea in the West African coast, in which anchovy was possibly  
140 restricted during glacial periods. At a certain time later, approximately 100 000 years ago, under  
141 favorable climatic conditions, Clade A anchovy should have migrated to the north east Atlantic  
142 up to the west Portuguese coast, while it also entered the Mediterranean and colonized the Black  
143 Sea. On the other hand, the different composition of clade in the northwestern Mediterranean and  
144 Bay of Biscay populations dominated by Clade B haplotypes, suggest that this clade was  
145 restricted to a Mediterranean refuge during the last glacial maximum (LGM), likely the  
146 easternmost Mediterranean, from which it expanded to the rest of the Mediterranean during the  
147 deglaciation period (Magoulas et al., 2006). Grant (2005) has argued against this hypothesis and  
148 suggested that the genetic imprints in Mediterranean populations of European anchovy indicate  
149 colonization dynamics on a similar time scale. Extreme climates during glaciations 350 000 and  
150 450 000 years ago likely extirpated populations in the entire Mediterranean Basin, and this was  
151 followed by a subsequent colonization of phylogroup 'B' fish. A later invasion by phylogroup 'A'

152 fish occurred during one of the pleniglacial cycles (50–25 k years ago) of the Weichselian ice age.  
153 The common ancestor of both clades was unknown, but most certainly lay outside the  
154 Mediterranean Basin complex from the Atlantic. Because of the salinity and temperature during  
155 the Pleistocene, Grant (2005) thinks that the European anchovy was not continuously present in  
156 the Black Sea which connects several times with the basins of the Mediterranean during the mid  
157 to late Pleistocene (Grant, 2005).

158 In keeping with distribution of clades found by Magoulas et al. (1996; 2006). In the present  
159 study, the clade composition of the Alboran and Moroccan Atlantic coast is almost exclusively  
160 from Clade A. This clade composition is similar to the clade composition in the western  
161 Portuguese coast, Bay of Cadiz, Canary Islands, the central African coast and the Alboran  
162 Spanish Sea and Algerian coast (Magoulas, Tsimenides & Zouros, 1996; Magoulas et al., 2006;  
163 Borrell et al., 2012; Viñas et al., 2013; Oueslati et al., 2014; Silva, Horne & Castilho, 2014). Also,  
164 in our sequences, we found a very low frequency of mitochondrial Clade B with nucleotide  
165 divergence of Clade A around 2.2%.

#### 166 **Population structure**

167 This work showed a heterogeneity among the 4 populations analyzed ( $\Phi_{st} = 0.01283$ ;  $p =$   
168  $0.03519$ ). In fact, there is a small signal, which may assume the presence of a population in  
169 Alboran Sea genetically different from Atlantic populations, particularly when we use dominant  
170 clade (Clade A) data only. This observation was indicated in a recent study of Viñas et al. (2013)  
171 by surveying sequence variability in the mitochondrial (mtDNA) control region. Viñas et al.,  
172 (2013) found that the Alboran population was genetically distinct from other Mediterranean and  
173 Northeast Atlantic populations, including neighboring populations; Cadiz. Viñas et al., (2013)  
174 explained the genetic distinctness of this population might result from the endogenous recovery  
175 of the population after the fishery collapse in the 1990s (Irazola et al., 1996; Pertierra & Lleonart,  
176 1996 in Viñas et al., 2013) that led to the accumulation of rare alleles, such as those detected in  
177 allozymes (Sanz et al., 2008). Alternatively, Viñas et al. (2013) think that the genetic distinctness  
178 might reflect the influx of both Clade A haplotypes from the Atlantic and Clade B haplotypes  
179 from the northwestern Mediterranean populations, resulting in a mixed population that is distinct  
180 from both sources. However, when we use only the clade A sequences, genetic distinctness  
181 increases and becomes more significant, which allows us to reject this hypothesis.

182 Finally, the observed genetic structure might be the reason of the presence of cryptic species.



183 Indeed, the geographical isolation, as shelters of the European anchovy during episodes of  
184 glaciation, followed by a secondary contact may be at the origin of the formation of cryptic  
185 species, that it is often difficult to distinguish from simple populations genetically structured.  
186 Bembo et al. (1996a) by analyzing allozymic markers and morphological differences, observed a  
187 genetic structuration between coastal anchovy (> 50 m) and open waters anchovy in the Adriatic  
188 Sea differences that Borsa, Collet & Durand (2004) confirmed by studying the polymorphism of  
189 the nuclear EPIC marker CK6-2. The existence of coastal forms of anchovies in the Moroccan  
190 Alboran Sea had already been reported in the literature on the basis of morpho-anatomical and  
191 behavioural differences. Thus, Kada et al. (2009) showed that an anchovy of Nador lagoon in  
192 Morocco displays some clear morphological and biological differences from marine *Engraulis*  
193 *encrasicolus*. Lastly, Oueslati et al. (2014) by analyzing mtDNA and six nuclear microsatellite  
194 loci, found two strongly differentiated loci and nucleo-cytoplasmic disequilibrium supported the  
195 existence of a coastal genetic entity in Nador lagoon in Morocco that is distinct from the more  
196 offshore marine populations.

197 The lack of genetic structure at the Atlantic populations, seems to be related to the fact that  
198 this pelagic fish presents relatively high migratory capacities. Indeed, this species is likely  
199 characterized by important gene flow, accelerated by a strong dispersion, a large effective  
200 population size countering genetic drift (Waples, 1989; Gonzalez & Zardoya, 2007), and the  
201 homogenizing forces related to the hydrodynamic environment, such as in the case of the north  
202 western African coasts with the Canaries current and the upwelling phenomenon (Le Floch, 1974;  
203 Belvèze & Erzini, 1983). However, more suitable sampling methods and more sensitive genetic  
204 markers is needed to confirm our results. We believe that additional genetic data can provide  
205 insights that will improve the sustainable management of European anchovy fisheries in the  
206 Moroccan fishery stocks.

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### 216 Field Study Permissions

217 Sampling and analysis techniques used in this work are consistent with the Moroccan  
218 legislation Article 14, Section 3, Bill 122.22 and the *Cartagena Protocol on Biosafety to the*  
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361 **Table 1** List of sample and genetic statistics within each of the sample sites for cytochrome b mitochondrial gene in *E.*  
 362 *encrasicolus*.

Region sampling locality	Sample Code	FAO fishery area	Date	N	$h \pm \text{s.d.}$	$\pi \pm \text{s.d.}$	Tajima's D	D p-value
Moroccan Alboran Sea	Albo	37.1.1	oct-12	37	0.998 $\pm 0.007$	0.01185 $\pm 0.00166$	-1.88700 $\pm 0.91417$	0.01000
Moroccan North Atlantic	N_ATL	34.1.11	nov-12	25	0.967 $\pm 0.029$	0.00704 $\pm 0.00135$	-1.97692 $\pm 0.89397$	0.01600
Moroccan Center Atlantic	C_ATL	34.1.12 & 34.1.13	Jan-13	44	0.993 $\pm 0.008$	0.00702 $\pm 0.00082$	-2.43168 $\pm 0.92737$	0.00000
Moroccan South Atlantic	S_ATL	34.1.3	mars-12	32	0.968 $\pm 0.024$	0.00934 $\pm 0.00183$	-2.20225 $\pm 0.91166$	0.00100
				138	0.9868 $\pm 0.0061$	0.008946 $\pm 0.004740$	-2.12446 $\pm 0.24399$	0.00675

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386 **Table 2** Pairwise  $\Phi_{st}$  among the 4 samples. Below the diagonal using the whole dataset samples and above using only Clade A  
 387 sequences.

Location	albo	N_ATL	C_ATL	S_ATL
Albo	0.00000	0.02004*	0.02764*	0.03181*
N_ATL	0.01015	0.00000	0.00087	0.00069
C_ATL	0.02990*	0.00409	0.00000	0.00277
S_ATL	0.01291	0.00000	0.00702	0.00000

388 Asterisks indicate significant values after Bonferroni correction (Sample codes as in Table 1).

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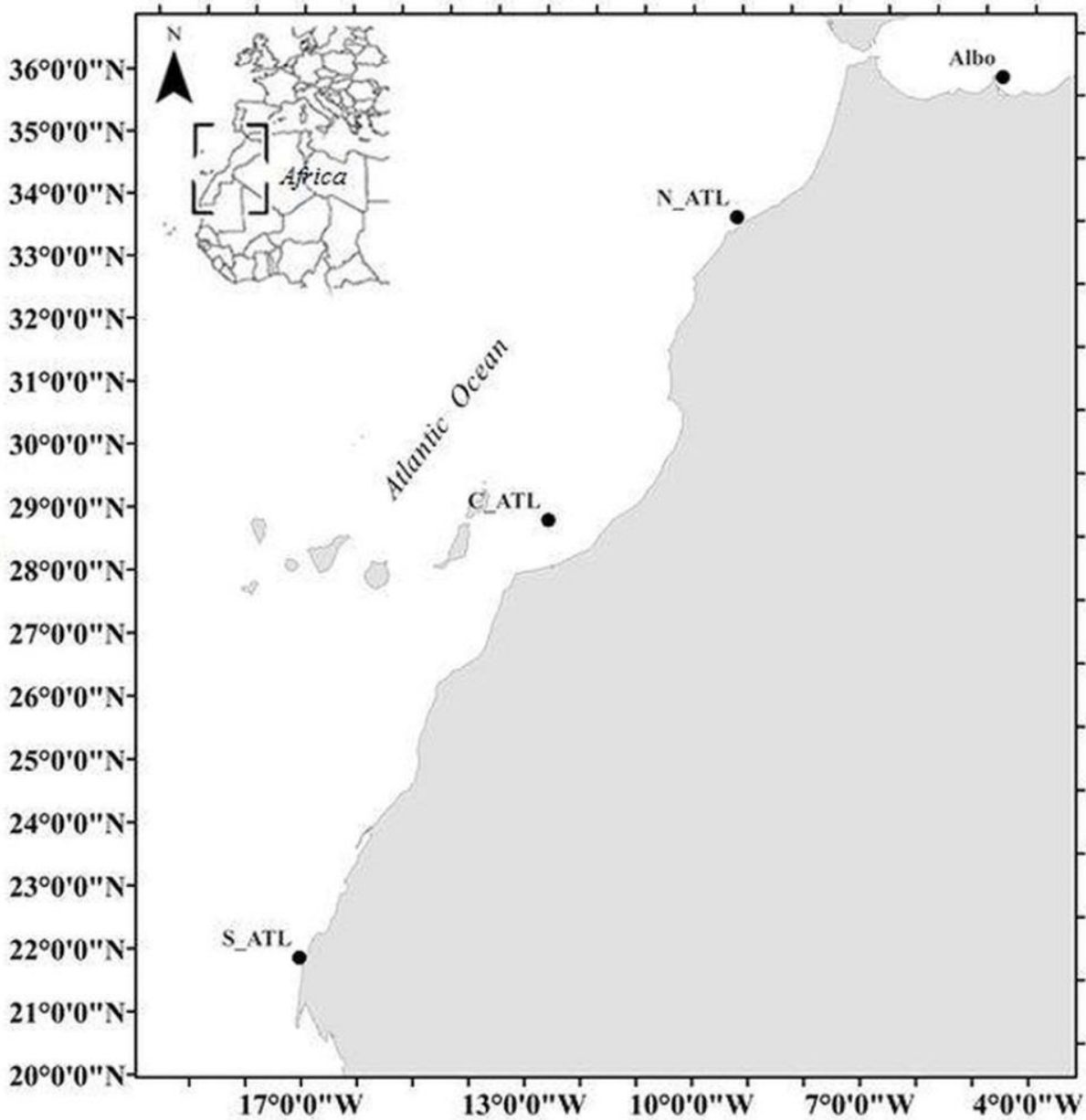


413 **Table 3** Analysis of the genetic variability among samples from *E. encrasicolus* using AMOVA .

	<b>source of variation</b>	<b>Percentage of Variation among groups</b>	<b>Variation Among populations within groups</b>
<b>dataset</b>	Group1 : Albo	2.53178	0.00%
	Group2: N_ATL , C_ATL and S_ATL	P-value = 0.00489	P-value = 0.17204
<b>Clade A</b>	Group1 : Albo	3.29	0.00%
	Group2: N_ATL , C_ATL and S_ATL	P-value = 0.00587	P-value = 0.24829

414 Sample codes as in Table 1.

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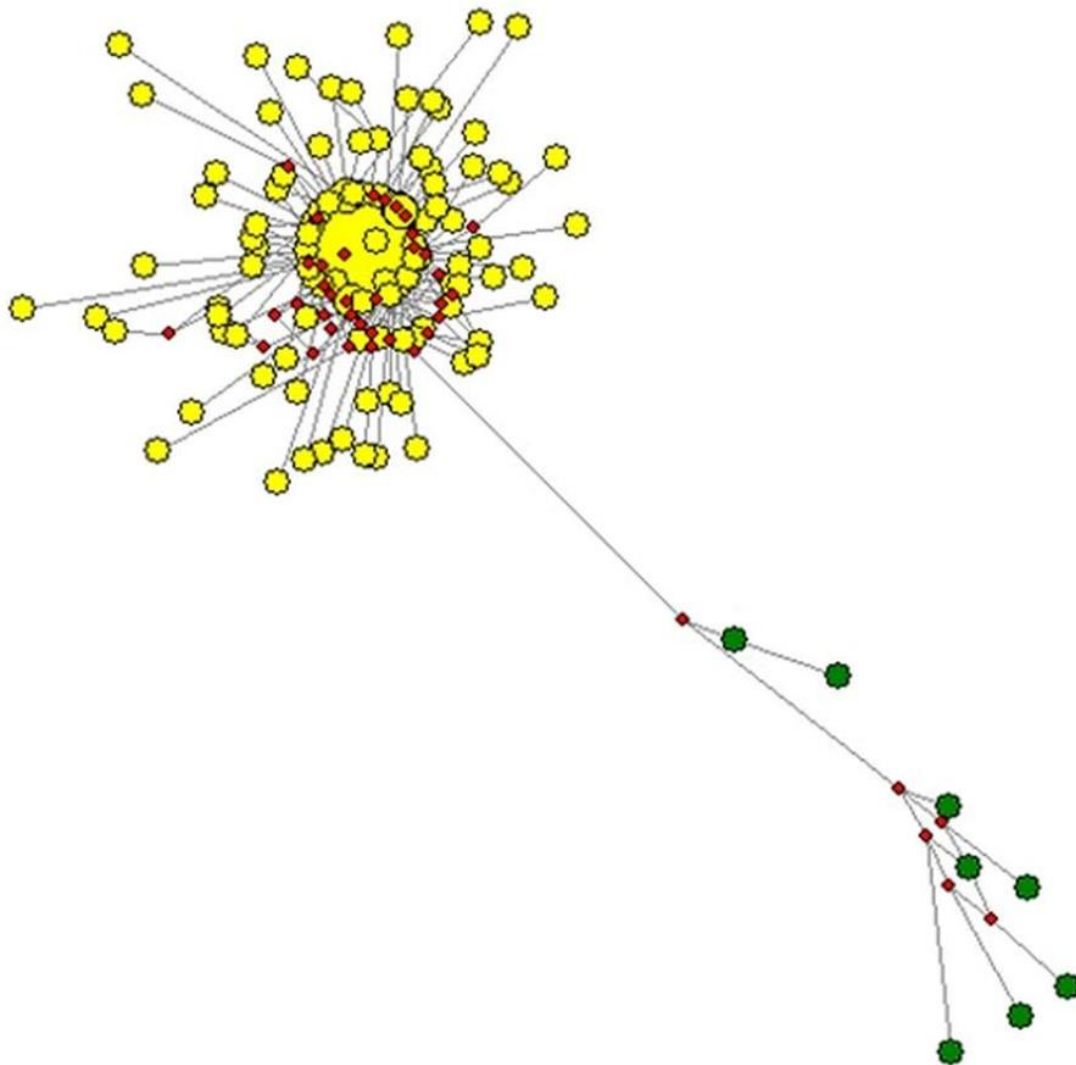


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417 **Figure. 1** Map of Moroccan Coast showing sampling sites. (Albo: Moroccan Alboran Sea; N\_ATL: Moroccan North Atlantic;  
418 C\_ATL: Moroccan Center Atlantic; S\_ATL: Moroccan South Atlantic).

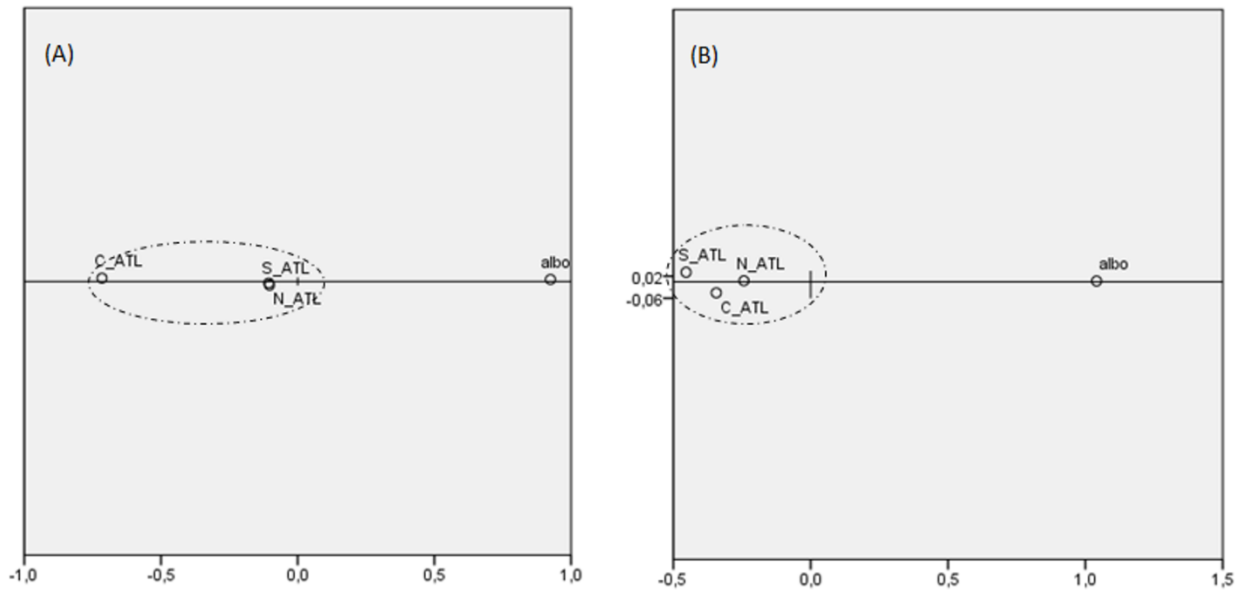


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**Figure 3** Haplotype analysis for Cytb mitochondrial genes in *E. encrasicolus* by Median-joining network showing the presence of two main clades (A and B). Yellow nodes represent Clade A, green nodes represent Clade B and red nodes represent undetected haplotypes in the study but necessary to most parsimonious network construction.



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433 **Figure 4** Multidimensional scaling of  $\Phi_{st}$  distance between 4 samples. (A) using the whole dataset samples and (B) using only  
434 Clade A sequences (Sample codes as in Table 1).

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