

A peer-reviewed version of this preprint was published in PeerJ on 11 August 2015.

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Göllner T, Fieder M. 2015. Selection in the dopamine receptor 2 gene: a candidate SNP study. PeerJ 3:e1149 <https://doi.org/10.7717/peerj.1149>

Selection in the dopamine receptor 2 gene: New candidate SNPs for disease-related studies

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Dopamine is a major neurotransmitter in the human brain and is associated with various diseases. Schizophrenia, for example, is treated by blocking the dopamine receptors type 2. In 2009, Shaner, Miller and Mintz stated that schizophrenia was the low fitness variant of a highly variable mental trait. We therefore explore whether the dopamine receptor 2 gene (*DRD2*) underwent any selection processes. We acquired genotype data of the 1000 Genomes project (phase I), which contains 1093 individuals from 14 populations. We included only single nucleotide polymorphisms (SNPs) with a minor allele frequency (MAF) of over 0.05 in the analysis. This is equivalent to 151 SNPs for *DRD2*. We used two different approaches (an outlier approach and a Bayesian approach) to detect loci under selection. The combined results of both approaches yielded nine candidate SNPs under balancing selection. While directional selection strongly favours one allele over all others, balancing selection favours more than one allele. All candidates are in the intronic region of the gene and only one (rs12574471) has been mentioned in the literature. Two of our candidate SNPs are located in specific regions of the gene: rs80215768 lies within a promoter flanking region and rs74751335 lies within a transcription factor binding site. We strongly encourage research on our candidate SNPs and their possible phenotypic effects.

2 Selection in the dopamine receptor 2 gene: New candidate SNPs for disease-related studies.

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6

7 ABSTRACT:

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9 Schizophrenia, for example, is treated by blocking the dopamine receptors type 2. In 2009,
10 Shaner, Miller and Mintz stated that schizophrenia was the low fitness variant of a highly
11 variable mental trait. We therefore explore whether the dopamine receptor 2 gene (*DRD2*)
12 underwent any selection processes. We acquired genotype data of the 1000 Genomes project
13 (phase I), which contains 1093 individuals from 14 populations. We included only single
14 nucleotide polymorphisms (SNPs) with a minor allele frequency (MAF) of over 0.05 in the
15 analysis. This is equivalent to 151 SNPs for *DRD2*. We used two different approaches (an outlier
16 approach and a Bayesian approach) to detect loci under selection. The combined results of both
17 approaches yielded nine candidate SNPs under balancing selection. While directional selection
18 strongly favours one allele over all others, balancing selection favours more than one allele. All
19 candidates are in the intronic region of the gene and only one (rs12574471) has been mentioned
20 in the literature. Two of our candidate SNPs are located in specific regions of the gene:
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22 factor binding site. We strongly encourage research on our candidate SNPs and their possible
23 phenotypic effects.

24 INTRODUCTION:

25 The catecholamine dopamine is a neurotransmitter in the human brain. Dopaminergic neurons
26 can be divided into four major pathways: nigrostriatal, mesolimbic, mesocortical and
27 tuberoinfundibular (Andén et al., 1964; Dahlstroem and Fuxe, 1964). These neurons play an
28 important role in voluntary movement, feeding, reward and learning, as well as certain other
29 functions. Outside of the brain, dopamine takes on a physiological role in cardiovascular
30 functions, hormonal regulation, renal functions and other (Snyder et al., 1970; Missale et al.,
31 1998; Sibley, 1999; Carlsson, 2001; Iversen and Iversen, 2007). Due to this involvement in many
32 different processes and systems, dopamine is also related to a variety of diseases. Parkinson's
33 disease, which is caused by a loss of dopaminergic innervations in the striatum, is a prominent
34 example (Ehringer and Hornykiewicz, 1960). Additionally, the expected associations between
35 the dopaminergic system and schizophrenia stem from the fact that various dopamine receptor 2
36 blockers are used as antipsychotics in treating that condition (Snyder et al., 1970; Creese et al.,
37 1976; Seeman et al., 1976; Carlsson et al., 2001). Further relationships with dopamine
38 dysregulation are expected in Tourette's syndrome and attention deficit hyperactivity disorder
39 (ADHD) (Mink, 2006; Swanson et al., 2007; Gizer et al., 2009). The strong involvement of
40 dopamine in the reward system suggests an association with drug abuse and addiction (Hyman et
41 al., 2006; Di Chiara and Bassareo, 2007; Koob and Volkow, 2010). Many more diseases and
42 conditions are expected to involve dopamine dysfunctions. (As reviewed by Beaulieu &
43 Gainetdinov, 2011)

44 In humans, five different dopamine receptors exist. They are classified into two categories based
45 on their structure and their pharmacological and biochemical properties. The D1-class includes
46 the dopamine receptors 1 and 5, while the D2-class consists of the dopamine receptors 2, 3 and 4
47 (Andersen et al., 1990; Niznik and Van Tol, 1992; Sibley and Monsma, 1992; Sokoloff et al.,
48 1992a; Civelli et al., 1993; Vallone et al., 2000). The focus of our study is on the dopamine
49 receptor 2 and its gene *DRD2*. The dopamine receptor 2 gene lies on the long arm of
50 chromosome 11 (11q23.1). It spans from 113,280,317 to 113,346,413 for a total of 66,096 base
51 pairs (bp) (information accessed on NCBI in the GnRH37 assembly). For the gene card, see
52 Figure 1 in Results. *DRD2* has six introns (Gingrich and Caron, 1993). Alternative splicing
53 between intron 4 and 5 of an 87 bp exon generates two variants of the dopamine receptor 2. The

54 difference between D2S (short) and D2L (long) is a 29-amino-acids-long chain in the third
55 intercellular loop of the protein (Giros et al., 1989; Monsma et al., 1989). While the short form
56 (D2S) is mainly expressed at the presynapse, the long form (D2L) is expressed postsynaptically
57 (Usiello et al., 2000; De Mei et al., 2009). The D2S are mainly autoreceptors, i.e. they reduce the
58 expression of dopamine when activated. This leads to an important negative feedback
59 mechanism (Wolf and Roth, 1990; Missale et al., 1998; Sibley, 1999). (Again, as reviewed by
60 Beaulieu & Gainetdinov, 2011)

61 Among the many single nucleotide polymorphisms (SNPs) of *DRD2*, one prominent example is
62 rs6277, also known as C957T. It has been associated with schizophrenia in Han Chinese in
63 Taiwan (Glatt et al., 2009), in Russians (Monakhov et al. 2008) and in Bulgarians (Betcheva et
64 al. 2009). Together with the -141C allele, the 957T allele is associated with the diagnosis of
65 anorexia nervosa (Bergen et al., 2005). A meta-analysis showed that the Ser311Cys
66 polymorphism (rs1801028) in *DRD2* is a risk factor for schizophrenia. The heterozygotes
67 (Ser/Cys) and the homozygotes for Cys were both at elevated risk for schizophrenia when
68 compared to the Ser/Ser genotypes (Glatt and Jönsson, 2006). In a study with alcoholic patients
69 and controls, the A allele of rs1076560 was more frequent in alcoholic patients (Sasabe et al.,
70 2007). In 2012, Mileva-Seitz et al. conducted a study with Caucasian mothers and their infants.
71 They taped mother-infant behaviour and genotyped various SNPs of *DRD2* and also *DRD1*.
72 Rs1799732 and the previously mentioned rs6277 were both associated with direct vocalization of
73 the mother towards the infant.

74 The body of literature on SNPs and their possible effects is growing rapidly. Considering the
75 influences those SNPs could have on human behaviour, and bearing in mind the different
76 ecological habitats of *Homo sapiens*, we aimed to explore if *DRD2* underwent any selection
77 processes. In 2009 an interesting proposal by Shaner, Miller and Mintz stated that schizophrenia
78 was the low fitness variant of a highly variable mental trait. Because of the connection between
79 dopamine receptor 2 and schizophrenia, as stated above, we focused our analysis on *DRD2*.

80 To reduce false-positives, we used two selection detection algorithms to explore *DRD2*. This is
81 an exploratory (“hypothesis-free”) approach in which we want to find candidate SNPs that were
82 under selection. The data basis of our analysis are the 1000 Genomes Project samples.

83 MATERIAL AND METHODS:

84 We acquired data from the 1000 Genomes Project (phase I) through SPSmart engine v5.1.1
 85 (<http://spsmart.cesga.es/engines.php>; Amigo et al., 2008), using the search term “DRD2”. We
 86 included all single nucleotide polymorphisms (SNPs) with a minor allele frequency (MAF)
 87 greater than 0.05. Over the span of the whole *DRD2* gene (113,280,317 – 113,346,413 in the
 88 GnRH37.p13 primary assembly; gene card shown in Results, Figure 1), this amounts to 151
 89 SNPs. In total we included the following populations in our analysis.

| Superpopulation (Code) | Population Code | Population | Number of Individuals |
|---------------------------|--------------------|---|--------------------------|
| Africa (AFR) | ASW | African ancestry in Southwest USA | 61 |
| | LWK | Luhya in Webuye, Kenya | 97 |
| | YRI | Yoruba in Ibadan, Nigeria | 88 |
| Europe (EUR) | CEU | Utah residents with Northern and Western European ancestry | 87 |
| | FIN | Finnish from Finland | 93 |
| | GBR | British from England and Scotland | 88 |
| | IBS | Iberian populations in Spain | 14 |
| | TSI | Toscans in Italy | 98 |
| East Asia (ASN) | CHB | Han Chinese in Beijing, China | 97 |
| | CHS | Han Chinese South | 100 |
| | JPT | Japanese in Tokyo, Japan | 89 |
| America (AMR) | CLM | Colombians from Medellin, Colombia | 66 |
| | MXL | Mexican ancestry from Los Angeles USA | 60 |
| | PUR | Puerto Ricans from Puerto Rico | 55 |
| | All populations = | | |

90 Table 1. Populations of the 1000 Genomes Project.

91 The data was converted by hand into the CONVERT format. All further format conversions were
92 performed by PGD Spider 2.0.5.2 (Lischer and Excoffier, 2012).

93 Two different programs were used to detect selection. Both use F_{ST} approaches to detect outliers.
94 The program LOSITAN calculates $FDIST$, which uses F_{ST} and the expected heterozygosity. It
95 assumes an island model of migration with neutral markers. An expected distribution of Wright's
96 inbreeding coefficient is calculated and then outliers are identified. A neutral mean F_{ST} was
97 computed by the program before the 50,000 simulations were performed. The infinite alleles
98 model was used. To avoid false positive detection we set the significance level to $p < 0.01$
99 ($P(\text{Simulation } F_{ST} < \text{sample } F_{ST})$). (Antao et al., 2008)

100 BayeScan is a Bayesian statistics program. Basically it calculates two simulations for every loci:
101 one in which it assumes the locus is under selection and the other one in which this assumption is
102 dropped. It splits the F_{ST} coefficient into two parts. The alpha value is a locus-specific
103 component shared by all populations. The beta value is a population-specific component shared
104 by all loci. This is achieved via logistic regression and provides insight into selection. The alpha
105 value serves as an indicator for selection. Significant positive values of alpha indicate directional
106 selection, whereas significant negative values indicate balancing selection. The posterior
107 probabilities are estimated using a reversible-jump Markov Chain Monte Carlo (MCMC)
108 approach. The posterior probabilities are gained by counting how many times alpha is included
109 in the model. Before the Markov chains are computed, we calculate 20 pilot runs with 5000
110 iterations each. The initial burn-in is set to 50,000 steps and the chains are run with 5000
111 iterations and a thinning interval of 10. The program output consists of a posterior probability,
112 the logarithm (base 10) of the posterior odds and a q value. These three values are all for the
113 model with selection. Furthermore, the alpha value is reported and an F_{ST} coefficient average of
114 all population per locus. In BayeScan the threshold of a posterior P of > 0.99 and a $\log_{10}(PO)$ of
115 2 or higher is used. This threshold is labelled as "Decisive" by BayeScan (see the program
116 manual at http://cmpg.unibe.ch/software/BayeScan/files/BayeScan2.1_manual.pdf). (Foll and
117 Gaggiotti, 2008)

118

119 To compute linkage disequilibrium (LD) of the SNPs, we used the R "genetics package"
120 (<http://cran.r-project.org/web/packages/genetics/genetics.pdf>; Warnes et al., 2013). We used D'

121 as a measurement for LD. In most populations one or more SNPs had to be excluded to
122 successfully run the computation. The population IBS was excluded entirely from this
123 computation. IBS is a very small population ($n = 14$), and 30 SNPs caused the computation to
124 fail. For a detailed view on all excluded SNPs see Table S1 in the supplementary material.

125 We accessed information on the gene via NCBI (<http://www.ncbi.nlm.nih.gov/>) and on the
126 specific SNPs via Ensembl (<http://www.ensembl.org/>).

127 RESULTS:

128 The combined results of LOSITAN and BayeScan yielded nine candidate SNPs under balancing
129 selection:

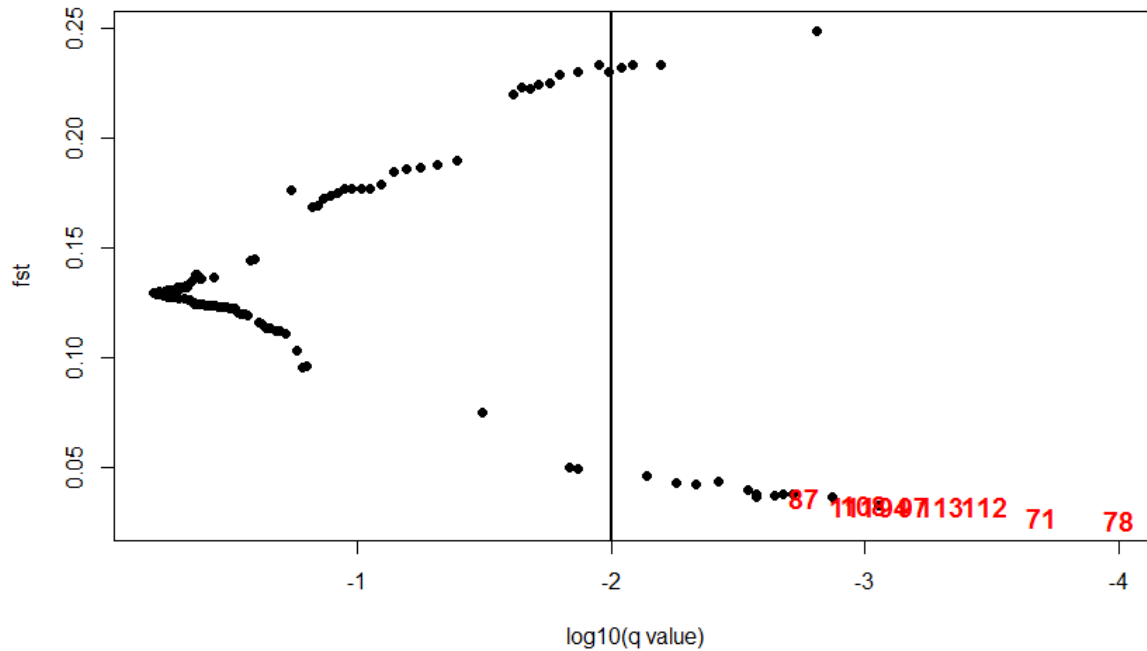
130

| Locus | Major Allele (Frequency) | Minor Allele (Frequency) | F_{ST} (Lositan) | F_{ST} (BayeScan) | Location |
|-------------------|-------------------------------------|-------------------------------------|-------------------------------------|--------------------------------------|----------------------------------|
| rs60599314 | C (0.871) | T (0.129) | 0.0110 | 0.0272 | 113,306,431 (Intronic region) |
| rs79549222 | T (0.87) | G (0.13) | 0.0106 | 0.0260 | 113,310,340 (Intronic region) |
| rs12574471 | C (0.891) | T (0.109) | 0.0172 | 0.0364 | 113,316,236 (Intronic region) |
| rs80215768 | G (0.925) | A (0.075) | 0.0304 | 0.0328 | 113,318,880 (Intronic region) |
| rs76581995 | C (0.925) | A (0.075) | 0.0304 | 0.0328 | 113,319,835 (Intronic region) |
| rs80014933 | T (0.923) | C (0.077) | 0.0304 | 0.0332 | 113,328,135 (Intronic region) |
| rs74751335 | G (0.915) | C (0.085) | 0.0266 | 0.0322 | 113,328,810 (Intronic region) |
| rs77264605 | A (0.915) | G (0.085) | 0.0266 | 0.0327 | 113,328,913 (Intronic region) |
| rs76499333 | G (0.925) | A (0.075) | 0.0299 | 0.0327 | 113,329,449 (Intronic region) |

131 Table 2. The dopamine receptor 2 gene's nine candidate SNPs for balancing selection.

132

150 Figure 2. LOSITAN graphical output. Yellow area = candidate loci for balancing selection. Grey
151 Area = neutral. Red area = candidate loci for positive selection.



152

153 Figure 3. Graphical output of BayeScan. The red numbers represent the overlapping SNPs with
154 LOSITAN. 71 = rs60599314, 78 = rs79549222, 87 = rs12574471, 94 = rs80215768, 97 =
155 rs76581995, 108 = rs80014933, 111 = rs74751335, 112 = rs77264605, 113 = rs76499333. For the
156 exact values see Table S2 in the supplementary material.

157

158 The Linkage Disequilibrium measurement D' was used. The heat maps for all nine populations
159 are shown in the supplementary material (Figure S1 – S13). The relative position of the marked
160 SNPs change because different populations had different SNPs excluded (see Table S1 for the list).
161

162 DISCUSSION:

163 We found nine SNPs to be candidates for balancing selection and none for directional selection.
164 Checking those SNPs with Genome Browser reveals that they are all intronic region variants.
165 rs80215768 (4) lies within a promoter flanking region and rs74751335 (7) lies within a
166 transcription factor binding site (TFBS). There have been many studies on the possible effects of
167 mutations in such regions (Hayashi, Watanabe and Kawajiri, 1991; and In et al. 1997; or for a
168 more general review on the topic, Jaenish & Bird, 2003). We strongly encourage further analysis
169 of those two SNPs in order to analyse their possible effects. Nonetheless, the SNPs show low F_{ST}
170 values, which is congruent with the finding of balancing selection. Sewall Wright's guidelines
171 for interpreting F_{ST} values suggest little genetic differentiation in our populations (as cited by
172 Jobling, 2013; Chapter 5, Box 5.2). As silent mutations in *DRD2* are known to alter the mRNA
173 stability and even the synthesis of the receptor itself (Duan et al., 2003), we should be eager to
174 explore the possible effects of these SNPs.

175 Additionally, the levels of the linkage disequilibrium measurement D' are typical for the
176 respective populations: African populations show a dispersed pattern and no clear LD blocks
177 (Figure S1, S9, and S13). While the LD blocks are visible in American populations (Figure S5,
178 S10 and S11), they are not as clear as in Asian (Figure S3, S4 and S8) or European populations
179 (Figure S2, S6, S7 and S12). Looking up the SNPs that are in high D' (> 0.8) with our nine
180 candidate loci revealed no new information as to the nature of those loci.

181 All of our SNPs have a $MAF > 0.05$ and there are some individuals (0.8 – 2.3% per SNP, over
182 all populations) that are homozygote for the minor allele. The finding of balancing selection
183 suggests that in our sample the minor alleles bear some fitness disadvantage. Fitness is altered if
184 survival or reproduction of an organism is affected. This raises the possibility of a connection
185 between our candidate SNPs and diseases or malfunctions of dopamine receptor 2. In the list of
186 diseases associated with dopamine (see Introduction) the most striking example is schizophrenia
187 because dopamine receptor 2 blockers can successfully treat patients. Leaving the biochemical
188 field, we can also find connections with diseases on a molecular level. Rs6277 is associated with
189 schizophrenia in Han Chinese in Taiwan (Glatt et al., 2009), in Russians (Monakhov et al. 2008)
190 and in Bulgarians (Betcheva et al. 2009). Rs1801028 was also described as a risk factor for
191 schizophrenia (Glatt and Jönsson, 2006). Bergen et al. (2005) found an association between the

192 alleles -141C and 957T and the diagnosis of anorexia nervosa. The A allele of rs1076560 was
193 more frequent in alcoholic patients when compared with a control group (Sasabe et al., 2007). In
194 a meta-analysis, Le Foll et al. (2009) concluded that there are associations between variants of
195 *DRD2* and alcohol dependency.

196 The question is whether these conditions could potentially affect fitness. Bassett et al. (1996)
197 showed that reproductive fitness is reduced in groups of familial schizophrenia, which suggests a
198 selection process. Puzzlingly enough, they also found some evidence for an increased fitness of a
199 small subsample of sisters. Shaner, Miller and Mintz in 2009 proposed that schizophrenia is the
200 low-fitness trait of a highly variable mental trait. They argue that the persistence of the illness at
201 about 1% globally is too high for new mutations. Balancing selection would fit this hypothesis
202 very well and our candidate SNPs could be viable indicators for this. Interestingly, rs6277 and
203 rs1801028, which are both associated with schizophrenia (Glatt et al., 2009; Monakhov et al.
204 2008; Betcheva et al. 2009; Glatt and Jönsson, 2006), are candidates for neutral loci in our study.
205 Note that also the other SNPs that are associated with alcohol dependency, anorexia nervosa or
206 even the behaviour of mothers with their infants, are all candidates for neutral loci. This method
207 does not allow us to make direct inferences about a phenotype (e.g. schizophrenia). We can only
208 make assumptions about the phenotype based on our genotype data. This study is a potential
209 precursor to future studies on the subject.

210 Lastly, what also should pique our interest is the fact that we only found candidates for balancing
211 selection. We extended our analysis to the other 4 dopamine receptors and again only found
212 candidates for balancing selection and none for directional selection (data not shown). Note that
213 this applies only to SNPs. There has been evidence for directional selection in *DRD4*, by
214 analysing a tandem repeat in the coding region (Ding et al. 2001). Recently, *DRD1* has been
215 analysed as one of ten neurochemical genes that are responsible for the Social-Decision Making
216 System (O'Connell & Hofmann, 2012). The study finds that the system underwent very little
217 change during evolution. The authors wanted to include *DRD2* but too little data for different
218 species were available. The fact that we find only balancing selection and no directional
219 selection acting upon *DRD2* hints that it is a conserved region within the human genome.

220 To untangle the possible effects of our SNPs we propose a study in which our candidate SNPs
221 are investigated in schizophrenic and non-schizophrenic persons. A simple comparison of the

222 SNPs and the different haplotypes between the two groups should efficiently assess our findings.
223 If this proposed study finds differences in those two groups, the mechanisms of those SNPs and
224 their possible haplotypes must be investigated.

225 CONCLUSION:

226 We found nine candidates for balancing selection on *DRD2* but no evidence for directional
227 selection. Some of the nine candidate SNPs under selection are potentially associated with various
228 diseases. These SNPs could be important as biomarkers due to their very low F_{ST} values: the
229 genetic differentiation of one population compared with the whole sample is very small. While all
230 candidate SNPs may be worth exploring, we definitely recommend using rs80215768 and
231 rs74751335 in further studies on *DRD2* because rs80215768 is within a promoter flanking region
232 and rs74751335 lies in a transcription factor binding site. The dopaminergic system, and thus also
233 its genes, seem to play a very basal role in animals. Accordingly, our finding of balancing selection
234 is in line with our expectations, i.e. a finding of directional selection within a current sample of
235 humans would have indeed been very striking.

236

237 ACKNOWLEDGEMENTS:

238 The authors would like to thank the R Core Team for the statistical computing environment, the
239 1000 Genomes Project, as well as the creators of LOSITAN, BAYESCAN and PGD Spider. The
240 first author would like to thank Martin Fieder for a great cooperation and support during the
241 writing of this publication. Additional thanks go to Bernard Wallner, Philipp Gewessler and
242 Matthias Hirschmanner.

243 REFERENCES:

- 244 Amigo, J., Salas, A., Phillips, C. & Carracedo, Á. (2008). SPSmart: adapting population based
245 SNP genotype databases for fast and comprehensive web access. *BMC bioinformatics*, 9(1), 428.
- 246 Andén, N. E., Carlsson, A., Dahlström, A., Fuxe, K., Hillarp, N. Å., & Larsson, K. (1964).
247 Demonstration and mapping out of nigro-neostriatal dopamine neurons. *Life sciences*, 3(6), 523-
248 530.
- 249 Andersen, P. H., Gingrich, J. A., Bates, M. D., Dearry, A., Falardeau, P., Senogles, S. E., &
250 Caron, M. G. (1990). Dopamine receptor subtypes: beyond the D 1/D 2 classification. *Trends in*
251 *pharmacological sciences*, 11(6), 231-236.
- 252 Antao, T., Lopes, A., Lopes, R. J., Beja-Pereira, A., & Luikart, G. (2008). LOSITAN: a
253 workbench to detect molecular adaptation based on a Fst-outlier method. *BMC bioinformatics*,
254 9(1), 323.
- 255 Bassett, A. S., Bury, A., Hodgkinson, K. A., & Honer, W. G. (1996). Reproductive fitness in
256 familial schizophrenia. *Schizophrenia research*, 21(3), 151-160.
- 257 Bertram, L. (2008). Genetic research in schizophrenia: new tools and future perspectives.
258 *Schizophrenia bulletin*, 34(5), 806-812.
- 259 Carlsson, A. (2001). A paradigm shift in brain research. *Science*, 294(5544), 1021-1024.
- 260 Carlsson, A., Waters, N., Holm-Waters, S., Tedroff, J., Nilsson, M., & Carlsson, M. L. (2001).
261 Interactions between monoamines, glutamate, and GABA in schizophrenia: new evidence.
262 *Annual review of pharmacology and toxicology*, 41(1), 237-260.
- 263 Civelli, O., Bunzow, J. R., & Grandy, D. K. (1993). Molecular diversity of the dopamine
264 receptors. *Annual review of pharmacology and toxicology*, 33(1), 281-307.
- 265 Creese, I., Burt, D. R., & Snyder, S. H. (1976). Dopamine receptor binding predicts clinical and
266 pharmacological potencies of antischizophrenic drugs. *Science*, 192(4238), 481-483.
- 267 Dahlström, A., & Fuxe, K. (1964). Evidence for the existence of monoamine-containing neurons
268 in the central nervous system. I. Demonstration of monoamines in the cell bodies of brain stem
269 neurons. *Acta Physiologica Scandinavica. Supplementum*, SUPPL-232.

- 270 De Mei, C., Ramos, M., Iitaka, C., & Borrelli, E. (2009). Getting specialized: presynaptic and
271 postsynaptic dopamine D2 receptors. *Current opinion in pharmacology*, 9(1), 53-58.
- 272 Di Chiara, G., & Bassareo, V. (2007). Reward system and addiction: what dopamine does and
273 doesn't do. *Current opinion in pharmacology*, 7(1), 69-76.
- 274 Ding, Y. C., Chi, H. C., Grady, D. L., Morishima, A., Kidd, J. R., Kidd, K. K., ... & Moyzis, R.
275 K. (2002). Evidence of positive selection acting at the human dopamine receptor D4 gene locus.
276 *Proceedings of the National Academy of Sciences*, 99(1), 309-314.
- 277 Duan, J., Wainwright, M. S., Comeron, J. M., Saitou, N., Sanders, A. R., Gelernter, J., &
278 Gejman, P. V. (2003). Synonymous mutations in the human dopamine receptor D2 (DRD2)
279 affect mRNA stability and synthesis of the receptor. *Human molecular genetics*, 12(3), 205-216.
- 280 Ehringer, H., & Hornykiewicz, O. (1960). Distribution of noradrenaline and dopamine (3-
281 hydroxytyramine) in the human brain and their behavior in diseases of the extrapyramidal
282 system. *Klinische Wochenschrift*, 38, 1236.
- 283 Foll, M., & Gaggiotti, O. (2008). A genome-scan method to identify selected loci appropriate for
284 both dominant and codominant markers: a Bayesian perspective. *Genetics*, 180(2), 977-993.
- 285 Gingrich, J. A., & Caron, M. G. (1993). Recent advances in the molecular biology of dopamine
286 receptors. *Annual review of neuroscience*, 16(1), 299-321.
- 287 Giros, B., Sokoloff, P., Martres, M. P., Riou, J. F., Emorine, L. J., & Schwartz, J. C. (1989).
288 Alternative splicing directs the expression of two D2 dopamine receptor isoforms. *Nature*, 342,
289 923 – 926.
- 290 Gizer, I. R., Ficks, C., & Waldman, I. D. (2009). Candidate gene studies of ADHD: a meta-
291 analytic review. *Human genetics*, 126(1), 51-90.
- 292 Hayashi, S. I., Watanabe, J., & Kawajiri, K. (1991). Genetic polymorphisms in the 5'-flanking
293 region change transcriptional regulation of the human cytochrome P450IIE1 gene. *Journal of*
294 *biochemistry*, 110(4), 559-565.
- 295 Hyman, S. E., Malenka, R. C., & Nestler, E. J. (2006). Neural mechanisms of addiction: the role
296 of reward-related learning and memory. *Annual review of neuroscience*, 29, 565-598.

297 In, K. H., Asano, K., Beier, D., Grobholz, J., Finn, P. W., Silverman, E. K., ... & Drazen, J. M.
298 (1997). Naturally occurring mutations in the human 5-lipoxygenase gene promoter that modify
299 transcription factor binding and reporter gene transcription. *Journal of clinical investigation*,
300 99(5), 1130.

301 Iversen, S. D., & Iversen, L. L. (2007). Dopamine: 50 years in perspective. *Trends in*
302 *neurosciences*, 30(5), 188-193.

303 Jaenisch, R., & Bird, A. (2003). Epigenetic regulation of gene expression: how the genome
304 integrates intrinsic and environmental signals. *Nature genetics*, 33, 245-254.

305 Jobling, M., Hollox, E., Hurles, M., Kivisild, T. & Tyler-Smith, C. (2013). *Human evolutionary*
306 *genetics*. New York and London: Garland Science.

307 Koob, G. F., & Volkow, N. D. (2010). *Neurocircuitry of addiction*. *neuropsychopharmacology*,
308 35(1), 217-238.

309 Le Foll, B., Gallo, A., Le Strat, Y., Lu, L., & Gorwood, P. (2009). Genetics of dopamine
310 receptors and drug addiction: a comprehensive review. *Behavioural pharmacology*, 20(1), 1-17.

311 Lischer, H. E. L., & Excoffier, L. (2012). PGDSpider: an automated data conversion tool for
312 connecting population genetics and genomics programs. *Bioinformatics*, 28(2), 298-299.

313 Mink, J. W. (2006). Neurobiology of basal ganglia and Tourette syndrome: basal ganglia circuits
314 and thalamocortical outputs. *Advances in neurology*. New York: Raven Press, 99, 89.

315 Missale, C., Nash, S. R., Robinson, S. W., Jaber, M., & Caron, M. G. (1998). Dopamine
316 receptors: from structure to function. *Physiological reviews*, 78(1), 189-225.

317 Monsma, F. J., McVittie, L. D., Gerfen, C. R., Mahan, L. C., & Sibley, D. R. (1989). Multiple
318 D2 dopamine receptors produced by alternative RNA splicing. *Nature*, 342, 926 – 929.

319 Niznik, H. B., & Van Tol, H. H. (1992). Dopamine receptor genes: new tools for molecular
320 psychiatry. *Journal of psychiatry and neuroscience*, 17(4), 158.

321 O’Connell, L. A., & Hofmann, H. A. (2012). Evolution of a vertebrate social decision-making
322 network. *Science*, 336(6085), 1154-1157.

323 Seeman, P., Lee, T., Chau-Wong, M., & Wong, K. (1976). Antipsychotic drug doses and
324 neuroleptic/dopamine receptors. *Nature*, 261(5562), 717-719.

325 Shaner, A., Miller, G., & Mintz, J. (2004). Schizophrenia as one extreme of a sexually selected
326 fitness indicator. *Schizophrenia research*, 70(1), 101-109.

327 Sibley, D. R. (1999). New insights into dopaminergic receptor function using antisense and
328 genetically altered animals 1. *Annual review of pharmacology and toxicology*, 39(1), 313-341.

329 Sibley, D. R., & Monsma, F. J. (1992). Molecular biology of dopamine receptors. *Trends in*
330 *pharmacological sciences*, 13, 61-69.

331 Snyder, S. H., Taylor, K. M., Coyle, J. T., & Meyerhoff, J. L. (1970). The role of brain dopamine
332 in behavioral regulation and the actions of psychotropic drugs. *American journal of psychiatry*,
333 127(2), 199-207.

334 Sokoloff, P., Andrieux, M., Besançon, R., Pilon, C., Martres, M. P., Giros, B., & Schwartz, J. C.
335 (1992). Pharmacology of human dopamine D 3 receptor expressed in a mammalian cell line:
336 comparison with D 2 receptor. *European journal of pharmacology: molecular pharmacology*,
337 225(4), 331-337.

338 Swanson, J. M., Kinsbourne, M., Nigg, J., Lanphear, B., Stefanatos, G. A., Volkow, N., ... &
339 Wadhwa, P. D. (2007). Etiologic subtypes of attention-deficit/hyperactivity disorder: brain
340 imaging, molecular genetic and environmental factors and the dopamine hypothesis.
341 *Neuropsychology review*, 17(1), 39-59.

342 Usiello, A., Baik, J. H., Rougé-Pont, F., Picetti, R., Dierich, A., LeMeur, M., ... & Borrelli, E.
343 (2000). Distinct functions of the two isoforms of dopamine D2 receptors. *Nature*, 408(6809),
344 199-203.

345 Vallone, D., Picetti, R., & Borrelli, E. (2000). Structure and function of dopamine receptors.
346 *Neuroscience & biobehavioral reviews*, 24(1), 125-132.

347 Warnes, G., Leisch, F., Man, M., & Warnes, M. G. Package 'genetics' (2013).

348 Wolf, M. E., & Roth, R. H. (1990). Autoreceptor regulation of dopamine synthesis. *Annals of the*
349 *New York academy of sciences*, 604(1), 323-343.

