



Master regulator genes and their impact on major diseases

Wanwan Cai^{1,*}, Wanbang Zhou^{2,*}, Zhe Han³, Junrong Lei², Jian Zhuang⁴, Ping Zhu⁴, Xiushan Wu¹ and Wuzhou Yuan¹

¹The Center for Heart Development, State Key Laboratory of Development Biology of Freshwater Fish, Key Laboratory of MOE for Development Biology and Protein Chemistry, College of Life Sciences, Hunan Normal University, Changsha, Hunan, China

²College of Physical Education, Hunan Normal University, Changsha, Hunan, China

³University of Maryland School of Medicine, Center for Precision Disease Modeling, Baltimore, MD, USA

⁴Guangdong Cardiovascular Institute, Guangdong Provincial People's Hospital, Guangdong Academy of Medical Sciences, Department of Cardiac Surgery, Guangzhou, Guangdong, China

*These authors contributed equally to this work.

ABSTRACT

Master regulator genes (MRGs) have become a hot topic in recent decades. They not only affect the development of tissue and organ systems but also play a role in other signal pathways by regulating additional MRGs. Because a MRG can regulate the concurrent expression of several genes, its mutation often leads to major diseases. Moreover, the occurrence of many tumors and cardiovascular and nervous system diseases are closely related to MRG changes. With the development in omics technology, an increasing amount of investigations will be directed toward MRGs because their regulation involves all aspects of an organism's development. This review focuses on the definition and classification of MRGs as well as their influence on disease regulation.

Subjects Molecular Biology, Clinical Trials, Oncology, Medical Genetics

Keywords Master regulator genes, Signal pathway, Tumor diseases, Cardiovascular disease, Nervous system disease

INTRODUCTION

Since the discovery of the master regulator genes (MRGs) and the powerful functions of these genes involved in all aspects of tissue and organ development, the study of MRGs have been more and more extensive, and an increasing number of new MRGs have been reported to play key roles in major clinical diseases. In the field of biomedicine, potential MRGs are generally analyzed based on the method of omic technologies, for instance, whole genome transcriptomics ChIPSeq and ATAC-Seq and well established bioinformatic analysis such as GSEA and its variants (*Alvarez et al., 2016; Boboila et al., 2018; Lefebvre et al., 2010; Tomljanovic et al., 2018*). Recent studies have pointed that the protein called myocyte enhancing factor 2C (MEF2C) is one of such master regulators involved in the pathogenesis of primary breast cancer. A systematic biological analysis of the transcriptional regulation activity of MEF2C and its target genes has revealed that this molecule induces collective responses leading to system-level gene expression deregulation and carcinogenesis (*Hernández-Lemus, Baca-López & Tovar, 2015*). A large number of clinical data from disease samples have been collected to calculate the potential MRGs

Submitted 16 June 2020
Accepted 25 August 2020
Published 6 October 2020

Corresponding author
Wuzhou Yuan, ywz@hunnu.edu.cn,
yuanwuzhou@aliyun.com

Academic editor
Amit Singh

Additional Information and
Declarations can be found on
page 13

DOI 10.7717/peerj.9952

© Copyright
2020 Cai et al.

Distributed under
Creative Commons CC-BY 4.0

OPEN ACCESS

in their pathological mechanisms. For example, in two breast cancer sample data sets, a systematic implementation of a series of algorithms is used to analyze the MRGs in potential primary breast cancer cells (Baca-López et al., 2012; Lim, Lyashenko & Califano, 2009; Tapia-Carrillo et al., 2019; Tovar et al., 2015). However, the definition of the MRG is still indistinct and imperfect, and a systematic and comprehensive review about MRGs is lacking. In this review, we proposed an updated definition and systematic classification of MRGs, and summarized the role of MRGs in major clinical diseases. The subject presented in this article is written in a descriptive manner instead of a systematic review so that clinicians outside our professional field can understand the basic characteristics of MRGs and their significant effects on clinical diseases.

WHAT IS THE MASTER REGULATOR GENE?

The term “master regulator gene” introduced by Susumu Ohno in 1978, refers to “the gene at the top of the regulatory hierarchy, which should not be affected by the regulation of any other genes” (Ohno, 1978). However, with the increasingly extensive and in-depth study of master regulator genes (MRGs) in recent decades, this definition is no longer an absolute. Many studies have shown that some MRGs can be regulated by others. For example, *mdm2* is the master regulator of tumor suppressor protein p53 (Momand, Wu & Dasgupta, 2000), while the *p53* gene is a master regulator of diverse cellular processes and a potential therapeutic target for cancer (Farnebo, Bykov & Wiman, 2010); and *snai1* is the master regulator of epithelial-mesenchymal transition, but it is regulated by *Pak1* through phosphorylation (Takahashi et al., 2013), which implicates *Pak1* as a master regulator of epithelial-mesenchymal transition (Yang et al., 2005).

It has been reported that MRGs play a key role via multiple signal pathways. For example, adenosine monophosphate-activated protein kinase (AMPK) regulates the energy balance inside cells by inhibiting adenosine triphosphate (ATP) consumption in the anabolic pathway and enhancing ATP synthesis in the catabolic pathway. When activated by external metabolic pressure, AMPK regulates a complex downstream signal cascade, promoting efficient energy production within the cells (Witczak, Sharoff & Goodyear, 2008). Another example is the phosphoinositide 3-kinase (PI3K)/protein kinase B (AKT)/mammalian target of rapamycin (mTOR) signaling pathway. Although this pathway is considered as a master regulator for cancer (Schaefer, Steiner & Lengerke, 2020; Xia & Xu, 2015), mTOR is also considered as a MRG of metabolism (Kim & Guan, 2015; Zeng, 2017). Furthermore, it has been reported that the genes for the three transcription factors Sox2, Oct3/4, and Nanog have been identified as the MRGs that regulate mammalian embryogenesis, embryonic stem cell self-renewal, and pluripotency. These MRGs can bind to enhancer elements in pluripotent embryonic stem cells (ESCs) and recruit mediators to form unusual enhancer domains, which are called super-enhancers. When the MRGs and mediators are simultaneously occupied, the expression programs for most genes in ESCs become co-activated (Rizzino, 2008; Whyte et al., 2013). Phenotypic conditions in living cells are largely determined by the interplay of a multitude of genes and their protein products, which form a gene regulatory network (GRN), and MRGs are the key players in GRNs. Gene regulatory

network analysis have shown that different levels of gene regulation are not only related but strongly coupled (*Hernández-Lemus, Baca-López & Tovar, 2015*). To summarize, MRGs can be updated as genes or signaling pathways that are expressed at the inception of a developmental lineage or a specific cell type, participate in the specification of that lineage by regulating multiple downstream genes' expression either directly or via interacting with other master regulator genes or signaling pathways to form super-enhancers, and critically, when misexpressed, will lead to uncontrolled expression of downstream target genes and MRGs, and have the ability to respecify the fate of cells destined to form other lineages, causing more abnormal development of tissues and organs.

SURVEY METHODOLOGY

A survey of >2,000 articles was carried out using the National Center for Biotechnology Information PubMed database (<https://www.ncbi.nlm.nih.gov/pubmed/>) by searching the keyword “master regulator gene”. After screening the contents of the abstracts of these literatures, we found that more than 900 articles quoting MRGs covered most species. Key words were extracted and recorded during the abstract reading, including the properties of the MRGs, the signaling pathways involved, the tissues or organs involved, and the diseases caused, etc. All the data was collated and considered effective. If multiple references mentioned a same MRG, we selected recently published papers or well-known journals for reference. These MRGs were systematically classified as either (1) whole-family MRGs, (2) signal pathway MRGs, or (3) tissue- or organ-specific MRGs.

OVERVIEW OF MRGS

Family MRGs refer to a gene family where all members are MRGs. There are two types: either all members have the same function, such as the HOX, MTA, and SREBP families; or different members in the same family may possess different functions, such as the GATA gene family. The HOX family MRGs are all involved in developmental processes, such as embryogenesis and hematopoiesis (*Candini et al., 2015; Grier et al., 2005; Magnusson et al., 2007; McGonigle, Lappin & Thompson, 2008; Rice & Licht, 2007; Vogel et al., 2016; Zhang et al., 2015*). In mammals, the HOX network consists of 39 genes that exhibit a high degree of sequence similarity, particularly in the homeobox domain. Homeobox genes function as master regulatory transcription factors during development, and their expression is often altered in cancer (*Brotto et al., 2020; Li et al., 2020; Qu et al., 2019*). Many of the chromosomal translocations associated with acute leukemias involve HOX genes, such as mixed lineage leukemia, which leads to the inappropriate expression of specific HOX gene subsets (*Collins & Thompson, 2018; Dickson, Lappin & Thompson, 2009*). In the GATA family, where each member has a different function, GATA1 and GATA2 regulate erythropoiesis and hematopoiesis as MRGs (*Bresnick & Johnson, 2019; Castaño et al., 2019; Gutiérrez et al., 2020; Kang et al., 2012; Katsumura et al., 2018; Katsumura et al., 2014; Leonards et al., 2020; Philipsen, 2013; Siegwart et al., 2020*), GATA3 is an immune response MRG (*El-Arabey et al., 2020; Li, Campos & Iida, 2015; Mirlekar, 2020; Nicol et al., 2016; Nomura et al., 2019*), and GATA4 regulates embryonic pancreas development

(Kondratyeva et al., 2017). Table 1 lists 18 major family MRGs. Among them, the CDX, CDK, HSF, MTA, SREBP, Rho, HNF, IL families and the Rab GTPase superfamily contain genes with the same functions. In the PLK, PAX, TBX, SOX, RUNX, IRF, BCL, and C/EBP families, each family member shares similar functions but also performs their own distinct role. In Fig. 1, we have summarized typical family MRGs involved in regulation at the cellular level, including CDK Family, Rho Family and PLK Family involved in cell cycle regulation, and BCL Family involved in cell apoptosis, etc. Figure 2 summarizes the Family MRGs involved in tissue and organ development, including PAX Family involved in eye development, TBX Family involved in heart development, etc.

The second type of MRGs is signaling pathways MRGs. In this type, either one of the members in the signal pathway is the MRG, such as AMPK from the AMPK signal pathway, which is known as a master regulator of cellular energy metabolism due to its role in regulating glucose, lipid, and protein metabolism. AMPK is an evolutionarily conserved master regulator of metabolism and a therapeutic target in type 2 diabetes. As an energy sensor, AMPK activity is responsive to both metabolic inputs, i.e., the ratio of AMP to ATP and numerous hormonal cues (Cunningham et al., 2014; Witczak, Sharoff & Goodyear, 2008). Or more commonly, members of the whole signaling pathway cooperate with each other as MRGs to regulate the development of a series of tissues and organs. For example, the mTOR signaling pathway is a master regulator of cell growth, proliferation and survival, metabolism, and skeletal muscle production in eukaryotes (Donnelly et al., 2017; Zeng, 2017). mTOR belongs to the PI3K-related protein kinase family. The mTOR signaling pathway plays a crucial role in the functional recovery of central nervous system trauma, especially for axon regeneration and autophagy, which has an extensive association with apoptosis. Significantly, this pathway is receiving novel concern for its role in the repair and regeneration of traumatic central nervous system injuries, such as traumatic brain injury and spinal cord injury (Lin, Huo & Liu, 2017a). The novel concern for mTOR is also because it is a master regulator of the inflammatory response in immune and non-immune cells and implicated in a number of chronic inflammatory diseases, especially rheumatic diseases, such as systemic lupus erythematosus, rheumatoid arthritis, systemic sclerosis, sjogren syndrome and seronegative spondyloarthropathy (Suto & Karonitsch, 2020). mTOR signaling pathway acts as a master regulator in memory CD8⁺ T⁻ cells, Th17, and NK cells development and their functional properties (Rostamzadeh et al., 2019). Researchers used RNAi system to specifically knockdown mTOR, raptor, S6K1, eIF4E, and FKBP12 expressions in antigenic CD8⁺ T⁻ cells and the results have demonstrated that mTOR acts as the key regulator of memory CD8⁺ T⁻ cells differentiation. When mTOR or raptor is knocked down, the expression levels of memory T⁻ cell markers CD127, CD62L, Bcl-2, and CD27 are remarkably elevated. Significant increases in memory CD8⁺ T⁻ cells differentiation after knockdown of S6K1 and eIF4E showed that mTOR exerted its effect through these two downstream molecules (Araki et al., 2009).

The major signaling pathways MRGs are presented in Table 2. For example, the transforming growth factor (TGF) β signaling pathway is the master regulator of the respiratory system, epithelial-mesenchymal transition and metastasis, and cancer development; Hedgehog signaling is the master regulator of cell differentiation; and the

Table 1 Summary of family MRGs and their related functions.

Family MRGs	Members	Functions
CDX Family	all	master regulator of HOX family gene (<i>Frohling et al., 2007; Rawat, Humphries & Buske, 2012; Shiotani et al., 2008</i>)
CDK Family	CDK1, CDK2, Cdc5	master regulator of cell cycle regulation (<i>Botchkarev & Haber, 2018; Hinds, 2003; Satyanarayana & Kaldis, 2009</i>)
HSFs Family	HSF1, HSF2, HSFA1	master regulator of heat shock reaction (<i>Filone et al., 2014; Liu & Charng, 2012; Qiao et al., 2017; Shinkawa et al., 2011</i>)
MTA Family	all	master regulator of the occurrence and metastasis of cancer (<i>Du et al., 2017; PA, 2014; Zhu et al., 2009</i>)
SREBP Family	all	master regulator of lipid homeostasis (<i>Gong et al., 2016; Krycer et al., 2010; Madison, 2016</i>)
Rho Family	Including RhoA, Rac1 and Cdc42 proteins, and so on	master regulator for a large number of cell functions, including control of cell morphology, cell migration and polarity, transcriptional activation and cell cycle progression (<i>Bai et al., 2015; Colomba & Ridley, 2014; Costa et al., 2011; PA, 2014; Singh et al., 2019; Watanabe, Takano & Endo, 2006; Zago et al., 2019</i>)
HNF (Hepatocyte nuclear factor) Family	Including HNF1A/B, HNF4alpha, HNF6	master regulator of pancreas and liver differentiation (<i>Alder et al., 2014; Janky et al., 2016; Kondratyeva et al., 2017; Odom et al., 2004; Sandovici et al., 2013</i>)
IL(interleukin) Family	Including IL-1, IL-2, IL-6, IL-7, IL-10, IL-12, IL-21, IL-23, IL-27, ILC3, and so on	master regulator of inflammation or immunity (<i>Fry & Mackall, 2001; Langrish et al., 2004; Neurath, 2007; Qin et al., 2017; Rojas et al., 2017; Sharma, Fu & Ju, 2011; Waldner & Neurath, 2014; Wilson & Esposito, 2009; Zhou & Sonnenberg, 2020</i>)
Rab GTPases Superfamily	Including Rab5, Rab7b, Rab11 GTPase, and so on	master regulator of cell membrane transport (<i>Distefano et al., 2015; Ishida, E. Oguchi & Fukuda, 2016; Pfeffer, 2017; Qi et al., 2015; Wu et al., 2015</i>)
The MiTF/TFE Family of Transcription Factors	MITF, TFEB, TFE3, TFEC	Master Regulators of Organelle Signaling, Metabolism, and Stress Adaptation (<i>Slade & Pulimilkunnil, 2017</i>)
	all	master regulator of cell division
PLK Family	PLK1	master regulator of mitotic related kinases (<i>Combes et al., 2017</i>)
	PLK4	master regulator of the formation of centrioles (<i>Levine & Holland Andrew, 2014; Shaheen et al., 2014</i>)
	all	master regulator of development and tissue homeostasis (<i>Relaix, 2015</i>)
PAX Family	Pax5	master regulator of b-cell development and leukemia (<i>Medvedovic et al., 2011; Nebral et al., 2009</i>)
	Pax6	master regulator of ganglion cells of the retina and eye development (<i>Albert et al., 2013; Shubham & Mishra, 2012</i>)
	TBX1	master regulator of muscle differentiation (<i>Chen et al., 2009</i>)
TBX Family	TBX5	master regulator of heart development (<i>Boogerd & Evans, 2016</i>)
	TBX21	master regulator of Th1 cell development (<i>Nicol et al., 2016; Stolarczyk, Lord & Howard, 2014; Wilkie et al., 2016</i>)

(continued on next page)

Table 1 (continued)

Family MRGs	Members	Functions
SOX Family	SOX2	master regulator of mammalian embryogenesis, embryonic stem cell self-renewal and pluripotency (<i>Rizzino, 2008; Whyte et al., 2013</i>)
	SOX3	master regulator of innate immunity (<i>Doostparast Torshizi & Wang, 2017</i>)
	SOX4	master regulator of EMT (epithelial-mesenchymal transition) (<i>Lourenço & Coffey, 2017; Tiwari et al., 2013</i>)
	SOX5, SOX6	the interaction with SOX9 is a master regulator of cartilage development (<i>Ma et al., 2016; Suzuki et al., 2012; Vivekanandan et al., 2015</i>)
	SOX9	master regulator of testis differentiation pathway (<i>Jakob & Lovell-Badge, 2011; Mork & Capel, 2010; Kozhukhar, 2012</i>) master regulator of fibroblast differentiation (<i>Noizet et al., 2016</i>) master regulator of pancreatic program (<i>Julian, McDonald & Stanford, 2017; Seymour, 2014</i>)
RUNX Family	SOXB1, SOXE, SOXF	master regulator of cell fate (<i>Julian, McDonald & Stanford, 2017</i>)
	RUNX1	master regulator of adult hematopoiesis (<i>Ichikawa et al., 2004; Wehrspaun, Haerty & Ponting, 2015; Wu et al., 2014</i>)
IRF Family	RUNX2	master regulator of osteoblast lineage (<i>Liu et al., 2017; Wysokinski, Pawlowska & Blasiak, 2015</i>)
	IRF-1	master regulator of cross talk between macrophage and L929 fibrosarcoma cells (<i>Nascimento et al., 2015</i>)
	IRF4	master regulator of human periodontitis (<i>Sawle et al., 2016</i>)
	IRF7	master regulator of IFN-I, virus-induced cytokine (<i>Hu et al., 2011; Lu et al., 2015; Wang et al., 2013</i>)
BCL Family	IRF8	master regulator of monocytes and dendritic cells development (<i>Tamura, 2017</i>)
	BCL-2	master regulator of apoptosis (<i>Chen et al., 2012; Häcker & Vaux, 1995</i>)
	BCL-6	master regulator of Tfh cell differentiation (<i>Matsumoto et al., 2017</i>)
C/EBP Family	BCL11B	master regulator of T cell (Th) differentiation (<i>Inoue et al., 2016</i>)
	BCL-2-like 10	master regulator of Aurora kinase mouse oocytes (<i>Lee et al., 2016</i>)
C/EBP Family	C/EBP α	master regulator of the bone marrow progenitor cells and fat formation (<i>Ding et al., 2011; Okuno, Inoue & Imai, 2013</i>)
	C/EBPbeta	master regulator of physiological cardiac hypertrophy (<i>Molkentin Jeffery, 2011</i>)

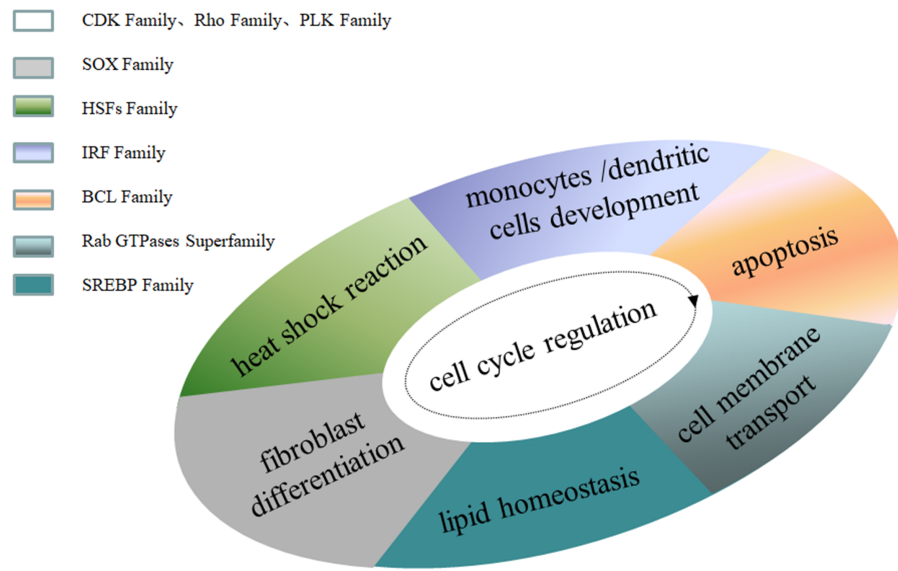


Figure 1 Family MRGs associated with cellular level regulation. .

Full-size DOI: 10.7717/peerj.9952/fig-1

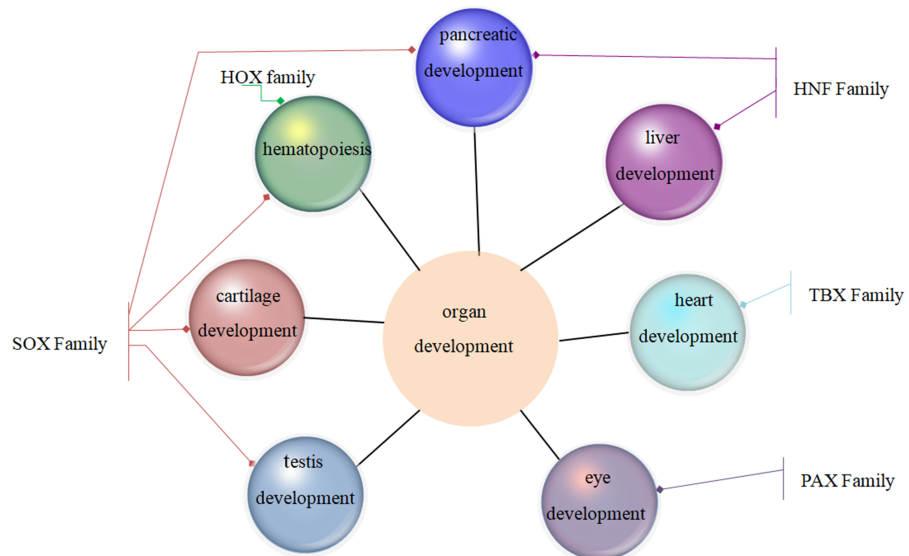


Figure 2 Family MRGs involved in tissue and organ development. .

Full-size DOI: 10.7717/peerj.9952/fig-2

NF- κ B signaling pathway is the master regulator of innate immune and inflammatory signals. It is noteworthy that the Wnt signaling pathway is not only the master regulator of cell development, cell polarization, and brain invasion but also the master regulator of liver-region and multiple renin-angiotensin system genes.

The third type of MRGs is tissue- or organ-specific MRGs that regulate the development of different tissue and organ systems. [Table 3](#) summarizes the MRGs associated with

Table 2 Summary of the important signaling pathways MRGs.

Signaling pathway	Master regulator gene	Functions
TGF- β signaling pathway	TGF- β signaling pathway	master regulator of the respiratory system, epithelial-mesenchymal transition and metastasis, and cancer development, etc (Fazilaty et al., 2013; Solomon et al., 2010; Zhou et al., 2014)
PI3K-AKT-mTOR signaling pathway	PI3K-AKT-mTOR signaling pathway	master regulator of cancer (Xia & Xu, 2015)
Hedgehog (Hh) signaling pathway	Hedgehog (Hh) signaling pathway	master regulator of cell differentiation (Peng & Joyner, 2015)
NF-kappaB signaling pathway	NF-kappaB signaling pathway	master regulator of innate immunity and inflammatory signaling (Krappmann et al., 2004; Matroule, Volanti & Piette, 2006; Schnappauf & Aksentijevich, 2020; Zeitz et al., 2017)
	Wnt signal pathway	master regulator of cell development and cell polarization (Gómez-Orte et al., 2013)
Wnt signaling pathway	Wnt5a	master regulator of brain invasion (Binda et al., 2017)
	Wnt/ β -catenin	master regulator of the liver region and multiple RAS (renin-angiotensin system) genes (Torre, Perret & Colnot, 2010)
	Notch	The fate of arteriovenous-lymphatic endothelial cells is regulated by the master regulator of Notch, COUP-TFII, and Prox1 (Kang et al., 2010)
Notch signaling pathway	Notch3	master regulator of neuroblastoma movement (Van Nes et al., 2013)
Yap signaling pathway	Yap1	master regulator of endometriosis (Lin et al., 2017b)
Hypoxia signaling pathway	HIF1, HIF-1 α	master regulators of the adaptive response to hypoxia (Lu & Kang, 2010; Schönenberger & Kovacs, 2015; Xiao, 2015; Zhao et al., 2020)
	HIF-2 α	

tissue/organ specificity, among which SCL/TAL1, VEGF, and PU.1 are the MRGs of hematopoiesis; Sim1 and Gcm are the MRGs of *Drosophila* neurodevelopment; FOXM1, Blimp1, Oct4, and Myc are the MRGs that regulate the cell cycle, B-cell differentiation to plasma cells, embryonic stem cells, and cell performance, respectively; CTCF is the MRG of human epigenetic and genomic spatial tissue; and FOXj1 is the MRG of the ciliary formation program. In bacteria, the MRGs include SinR, CtrA, FlhDC, Fur, CsgD, Spo0A, CcpA, LuxR, and WOR1. Details and other tissue- and organ-specific MRGs are listed in Table 3.

REGULATION OF MAJOR DISEASES BY THE MRGS

Since MRGs can concurrently regulate the expression of hundreds of genes, their expression levels must be tightly controlled, otherwise, misexpression or overexpression will exert a considerable impact on the development of affected organisms, resulting in runaway or uncontrolled metabolism and abnormal development in humans.

MRGs regulation of tumors

MRGs have been implicated in the occurrence of different tumors, including gum germ cell tumors, ovarian cancer, colon cancer, rectal cancer, and lung cancer. For example,

Table 3 Summary of reported MRGs and their related functions.

MRGs	Related functions
SCL/TAL1	master regulator of the adult hematopoietic (<i>Courtial et al., 2012; Wehrspaun, Haerty & Ponting, 2015</i>)
VEGF	master regulator of mucosal immunity driving angiogenesis (<i>Danese, 2008b</i>)
PU.1	master regulator of hematopoiesis and bone marrow (<i>Yang et al., 2012</i>)
Sim1	master regulator of <i>Drosophila</i> neurogenesis (<i>Eaton & Glasgow, 2006</i>)
Gcm	master regulator of nervous system development in <i>Drosophila</i> , parathyroid development, master regulator of expression and function regulation in mammals (<i>Cattenoz & Giangrande, 2016</i>)
FOXM1	master regulator of different stages of the cell cycle (<i>Jeffery et al., 2017; Zona et al., 2014</i>)
Blimp1	master regulator of B cell differentiation into plasma cells (<i>John & Garrett-Sinha, 2009; Vrzalikova, Woodman & Murray, 2012</i>)
Oct4	master regulator of embryonic stem cell self-renewal and pluripotency (<i>Samardzija et al., 2017a</i>)
Myc	master regulator of cell performance (growth, proliferation, stem cell pluripotency, ribosomal biogenesis, etc.) (<i>Grifoni & Bellosta, 2015; Holmberg Olausson, Nistér & Lindström, 2012; Kazan & Manners, 2013</i>)
HIF	master regulator of cellular responses to hypoxia (<i>Liu, Semenza & Zhang, 2015; Semenza, 2014; Semenza, 2017</i>)
CTCF	master regulator of human epigenetics and genomic spatial organization (<i>Golan-Mashiach et al., 2012</i>)
FOXj1	master regulator of cilia generation program (<i>Yu et al., 2008</i>)
SinR	master regulator of <i>Bacillus subtilis</i> biofilm formation (<i>Chu et al., 2006; Stowe et al., 2014</i>)
CtrA	master regulator of the cell cycle of the bacillus (<i>Gora et al., 2010; Laub et al., 2002; Pini et al., 2015</i>)
FlhDC	master regulator of flagellar genes (<i>Chatterjee, Cui & Chatterjee, 2015; Cui et al., 2008; Stafford, Ogi & Hughes, 2005</i>)
Fur	master regulator of iron metabolism in Gram-negative bacteria (<i>González et al., 2012; Huja et al., 2014</i>)
CsgD	master regulator of <i>E. coli</i> biofilm formation (<i>Ogasawara, Yamamoto & Ishihama, 2010; Wen et al., 2017</i>)
Spo0A	master regulator of the pathogenesis of <i>Bacillus subtilis</i> spore formation (<i>Fujita & Losick, 2005; Wolański & Jakimowicz, 2014</i>)

(continued on next page)

Table 3 (continued)

MRGs	Related functions
CcpA	master regulator of carbon catabolism regulation in <i>Bacillus</i> (Muscariello et al., 2013; Weeks et al., 2012a; Weeks et al., 2012b)
LuxR	master regulator of quorum sensing (Ball, Chaparian & van Kessel, 2017; Pompeani et al., 2008)
WOR1	master regulator of white and opaque phenotypes of <i>Candida albicans</i> (Zhang et al., 2014)
P53	master regulator of human malignant tumors (Farnebo, Bykov & Wiman, 2010; Resnick et al., 2005)
P63	master regulator of epidermal development and differentiation (Soares & Zhou, 2018)
Nrf2	master regulator of redox homeostasis (Basak et al., 2017; Cores et al., 2020; Hayes & Dinkova-Kostova, 2017)
MITF	master regulator of melanocyte development (Levy, Khaled & Fisher, 2006)
TFEB	master regulator of lysosomal biogenesis and autophagy (Medina et al., 2015; Settembre et al., 2011)
MyoD	master regulator of skeletal muscle gene expression programs (Aziz, Liu & Dilworth, 2010; Sunadome et al., 2014)
MicroRNAs (miR-10b*, miR21, miR-31, miR153, miR156, etc.)	master regulator of gene expression in many physiological and pathological processes (Biagioni et al., 2012; Datta & Paul, 2015; Kaul & Krams, 2015; Liang et al., 2020; Miranda et al., 2010; Schmittgen, 2010; Stief et al., 2014; Voorhoeve, 2010)
PGC-1 α	master regulator of mitochondrial gene expression (Fernandez-Marcos & Auwerx, 2011; Zhu et al., 2009)
Prox1	master regulator of lymphatic endothelial cell differentiation (Hong & Detmar, 2003; Kang et al., 2010; Ke & Yang, 2017)
AphA	master regulator of quorum sensing (Sun et al., 2012; Van Kessel et al., 2013)
PPARgamma	master regulator of fat formation (Lehrke & Lazar, 2005a; Sunadome et al., 2014)
foxp3	master regulator of regulatory T (Treg) cell development and function (Liston, 2010; Thornton & Shevach, 2019)
ComK	master regulator of late competence genes (Jaskólska & Gerdes, 2015; Ogura, Hashimoto & Tanaka, 2002)

SOX9, GATA4, PDX1, PTF1a, HNF1b, and GRP78 are master regulators of pancreatic cancer (*Kondratyeva et al., 2017*); while Srebp2 (*Krycer et al., 2010*) and E2F8 (*Rohde et al., 1996*) are MRGs of prostate cancer; and CDX2 is the master regulator of gastric cancer (*Shiotani et al., 2008*). Nuclear receptors are liver cancer-related (*Jakobsson et al., 2012*); PD-L1, TGF- β 1, and IL-10 are the master regulators of cervical cancer (*Qin et al., 2017*); and Oct4A is the master regulator of ovarian cancer (*Samardzija et al., 2017*). Analysis of master regulatory genes may help to understand the most upstream events in phenotypic development, particularly those related to cancer biology.

The most extensively studied MRGs are associated with breast cancer and leukemia. Breast cancer is the most common malignant tumor in women. It has been reported that RUNX1 encodes the transcription factor of the RUNX family, a new mutation in RUNX gene was discovered in human breast cancer. It was reported that RUNX1 was expressed in all subpopulations of mouse mammary epithelial cells (MECs) except for secretory alveolar cells. The conditional knockout of RUNX1 in the MECs resulted in the reduction of luminal MECs. Mainly due to a significant reduction in estrogen receptors (ERs), this phenotype could be rescued by the absence of Trp53 or Rb1. The underlying molecular mechanism was explained by RUNX1 inhibiting the expression of *Elf5* (the dominant gene in alveolar cells) and regulating the involvement of mature transcription factor or cofactor genes (such as *Foxa1* and *Cited1*) in the processes of ER synthesis (*Van Bragt et al., 2014*). Many other MRGs have been reported to be associated with the development of breast cancer, including the HOX gene family, SOX4, RUNX2, AMPK, p53, TGF- β , microRNA, KDM4B, p16INK4A, BACH1, Snai1, HMGA1, SATB1, HSP90, TRB3, Ddx5 and Ddx17, FGFR2, and AGTR2 (*Table S1*).

Another type of widely studied cancer is leukemia, a malignant clonal disease of hematopoietic stem cells. Due to uncontrolled proliferation, differentiation disorder, and blocked apoptosis, clonal leukemia cells proliferate and accumulate in the bone marrow and other hematopoietic tissues, infiltrate other non-hematopoietic tissues and organs, and inhibit normal hematopoietic function. Acute lymphoblastic leukemia (ALL) is the most common form of childhood cancer and is characterized by impaired lymphocyte differentiation, resulting in the accumulation of immature progenitor cells in the bone marrow, peripheral blood, and occasionally the central nervous system. Although ALL cure rates are close to 90%, it remains the leading cause of cancer-related mortality in children and young adults. Another extremely prevalent form of leukemia is B-cell precursor (BCP)-ALL, which represents 85% of cases, while the remaining 15% involve T-cell precursors. It was reported that BCP-ALL might be caused by the synergistic regulation of transcription factors, such as RUNX1, IKZF1, E2A, EBF1, and PAX5 (*Tijchon et al., 2012*). The other MRGs associated with leukemia include HOX, GATA, CDX, Pax, C/EBPistic genetic lesions, and key transcriptional targets and pathways (*Table S1*).

Influence of MRGs on cardiovascular diseases

Because cardiovascular disease is the leading cause of death in humans, elucidation of the associated role of MRGs is of immense clinical and social value for the effective prevention and treatment of cardiovascular diseases. The MRGs related to heart disease (*Table 4*)

Table 4 Summary of MRGs related to heart disease.

MRGs	Cardiovascular disease type
TBX5, NuRD	Congenital heart disease (<i>Boogerd & Evans, 2016</i>)
SREBP	Treatment of cardiac metabolic diseases (<i>Krycer et al., 2010</i>)
VEGF	Vascular disease (<i>Danese, 2008b; Gianni-Barrera et al., 2014</i>)
MyoD	Heart disease (<i>Kojima & Ieda, 2017</i>)
PPAR γ	Obesity, diabetes and cardiovascular disease (<i>Lee & Ge, 2014; Lehrke & Lazar, 2005</i>)
PKC δ	Thrombosis complications (<i>Fischer, 2009</i>)
SCL/TAL1	Anemia patient (<i>Fujiwara, 2017</i>)
Class IB phosphoinositide 3-kinase p110s	Heart disease (<i>Perino, Ghigo & Hirsch, 2010</i>)
PI3K	Heart failure (<i>Weeks et al., 2012a</i>)
SOX9 and myocardin	Atherosclerosis, vascular calcification (<i>Xu et al., 2012</i>)
Klotho	Cardiovascular diseases (<i>Moe Sharon, 2012</i>)
PITX2	Atrial fibrillation (AF) is the most common persistent Arrhythmia (<i>Li, Dobrev & Wehrens, 2016</i>)
FLYWCH1, PSORSIC3, G3BP1	Coronary artery disease (CAD) (<i>Foroughi Asl et al., 2015</i>)
Thyroid hormones (THs)	Cardiovascular diseases (<i>Rajagopalan & Gerdes, 2015</i>)
CST	Cardiovascular diseases (CVD) (<i>Sushil, Malapaka & Nitish, 2018</i>)
Etv2	Chronic vascular disease (<i>Garry, 2016</i>)

include TBX5, NuRD, SREBP, MyoD, Class IB PI3K p110 genetic lesions, PI3K, and PITX2, which mainly regulate congenital heart disease, metabolic heart disease, heart failure, arrhythmia, etc. Vascular-related MRGs, which include PKC δ , VEGF, SCL/TAL1, PPAR gamma, PGC-1alpha, SOX9, myocardin, FLYWCH1, PSORSIC3, G3BP1, and Etv2, mainly regulate thrombosis, anemia, atherosclerosis, vascular calcification, coronary artery disease, chronic vascular disease, etc. Others, like, Klotho, thyroid hormones and thyroid-stimulating hormone, and CST were also reported as master regulators of cardiovascular disease.

Influence of MRGs on Nervous system diseases

Nervous system diseases refer to the diseases that occur in the central nervous system, peripheral nervous system and vegetative nervous system, with sensory, motor, consciousness and vegetative nervous dysfunction as the main manifestations, among which the central nervous system diseases are the most widely studied. The central nervous system disease generally refers to the central nervous system degenerative disease, which refers to a group of diseases produced by the chronic progressive degeneration of the central nervous system. Pathologically, there are neuronal degeneration and neuron loss in the brain and/or spinal cord. Major diseases include Parkinson's disease, the overall ischemia, stroke, epilepsy, Alzheimer's disease and Huntington's disease, etc. At present, many articles have clarified the important role of master regulator genes in neurodegenerative diseases. For example, REST, a major transcriptional regulator of neurodegenerative diseases, is

a transcriptional suppressor that silences target genes through epigenetic remodeling. REST and REST-dependent epigenetic remodeling provide a central mechanism critical to the progressive neuronal degeneration associated with neurologic disorders and diseases including global ischemia, stroke, epilepsy, Alzheimer's and Huntington's disease (*Hwang & Zukin, 2018*). NRF2 regulation processes as a source of potential drug targets against neurodegenerative diseases (*Buendia et al., 2016; Cores et al., 2020*). ZCCHC17 is a master regulator of synaptic gene expression in Alzheimer's disease (*Tomljanovic et al., 2018*). ATF2 and PARK2 are transcription factors that act as MRGs in Alzheimer's disease (*Vargas et al., 2018*). The ubiquitin-proteasome system is a master regulator of neural development and the maintenance of brain structure and function (*Luza et al., 2020*), etc. At present, it has not been reported that there is a specific drug effective for various neurological diseases in the world. For many patients, relevant drugs just only relieve symptoms rather than cure diseases, causing indelible damage to patients' physical and mental health. Exploring novel MRGs working on the nervous system and disclosing the molecular mechanism of nervous system diseases, may become the exciting expect to develop target drugs and therapeutic schedule to achieve special purpose for the treatment of patients.

There are still many references on the research of master regulatory genes and other human various diseases. For example, there are some reports on the progress of investigating the influence of MRGs on diseases such as inflammatory bowel disease (*Danese, 2008a*), cartilage disease (*Ma et al., 2016*), and human diseases related to fibroblasts (*Shenoy et al., 2014*). Thus, the influence of MRGs on human diseases has permeated every aspect, and MRGs play a vital role in the clinical research and treatment of human diseases. However, how the MRGs can be used more comprehensively to solve the therapy problems in human diseases is an arduous task at present.

OUTLOOK

With the sustained development in omics technologies, research pertaining to MRGs will continue getting more concern and progress because the involvement of MRGs in all aspects of an organism's development is becoming apparent. Here we demonstrated that MRGs fell within three operating motifs: (1) whole-family MRGs, (2) signaling pathway MRGs, and (3) tissue- or organ-specific MRGs and updated the definition of MRGs as genes or signaling pathways that are expressed at the inception of a developmental lineage or a specific cell type, participates in the specification of that lineage by regulating multiple downstream genes' expression either directly or via interacting with other master regulator genes or signaling pathways to form super-enhancers, and critically, when misexpressed, will lead to uncontrolled expression of downstream target genes and MRGs, and have the ability to respecify the fate of cells destined to form other lineages, causing more abnormal development of tissues and organs. The formidable function of an MRG lies not only in its regulation of the concurrent expression of hundreds of genes but also the diversity of its functions on human diseases.

MRGs play important roles in the occurrence of various human diseases (such as cancer, cardiovascular diseases and neurological diseases) and exhibit a great potential to

be targets of gene therapies and drugs. Therefore, exploring the MRGs corresponding to the pathological mechanisms of different diseases is particularly critical. At present, there have been many reports on the analysis of potential MRGs through different calculation methods, and subsequent experimental verification, which greatly improves the process of discovering and determining MRGs in the pathogenesis. Of course, the use of MRGs for gene therapy or targeted drugs is still a huge challenge, and its clinical application is also a long process, which requires unremitting efforts of the medical research team. We believe that the day of technological breakthroughs of MRGs will definitely come.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

This study was supported in part by grants from the National Natural Science Foundation of China (Nos.: 81370451, 81470449, 81670290, 81570279), the Cooperative Innovation Center of Engineering and New Products for Developmental Biology of Hunan Province (No. 2013-448-6). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors:

National Natural Science Foundation of China: 81370451, 81470449, 81670290, 81570279.
Cooperative Innovation Center of Engineering.

New Products for Developmental Biology of Hunan Province: 2013-448-6.

Competing Interests

The authors declare there are no competing interests.

Author Contributions

- Wanwan Cai, Wanbang Zhou, Xiushan Wu and Wuzhou Yuan conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
- Zhe Han performed the experiments, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
- Junrong Lei, Jian Zhuang and Ping Zhu performed the experiments, prepared figures and/or tables, and approved the final draft.

Data Availability

The following information was supplied regarding data availability:

The raw data are available in the [Table S1](#).

Supplemental Information

Supplemental information for this article can be found online at <http://dx.doi.org/10.7717/peerj.9952#supplemental-information>.

REFERENCES

- Albert M, Schmitz SU, Kooistra SM, Malatesta M, Torres CMorales, Reikling JC, Johansen JV, Abarrategui I, Helin K. 2013. The histone demethylase Jarid1b ensures faithful mouse development by protecting developmental genes from aberrant H3K4me3. *PLOS Genetics* 9:e1003461-e1003461 DOI 10.1371/journal.pgen.1003461.
- Alder O, Cullum R, Lee S, Kan AC, Wei W, Yi Y, Garside VC, Bilenky M, Griffith M, Morrissy AS, Robertson GA, Thiessen N, Zhao Y, Chen Q, Pan D, Jones SJM, Marra MA, Hoodless PA. 2014. Hippo signaling influences HNF4A and FOXA2 enhancer switching during hepatocyte differentiation. *Cell Reports* 9:261–271 DOI 10.1016/j.celrep.2014.08.046.
- Alvarez MJ, Shen Y, Giorgi FM, Lachmann A, Ding BB, Ye BH, Califano A. 2016. Functional characterization of somatic mutations in cancer using network-based inference of protein activity. *Nature Genetics* 48:838–847 DOI 10.1038/ng.3593.
- Araki K, Turner AP, Shaffer VO, Gangappa S, Keller SA, Bachmann MF, Larsen CP, Ahmed R. 2009. mTOR regulates memory CD8 T-cell differentiation. *Nature* 460:108–112 DOI 10.1038/nature08155.
- Aziz A, Liu Q-C, Dilworth FJ. 2010. Regulating a master regulator: establishing tissue-specific gene expression in skeletal muscle. *Epigenetics* 5:691–695 DOI 10.4161/epi.5.8.13045.
- Baca-López K, Mayorga M, Hidalgo-Miranda A, Gutiérrez-Nájera N, Hernández-Lemus E. 2012. The role of master regulators in the metabolic/transcriptional coupling in breast carcinomas. *PLOS ONE* 7:e42678–e42694 DOI 10.1371/journal.pone.0042678.
- Bai Y, Xiang X, Liang C, Shi L. 2015. Regulating Rac in the nervous system: molecular function and disease implication of Rac GEFs and GAPs. *BioMed Research International* 2015:632450–632466 DOI 10.1155/2015/632450.
- Ball AS, Chaparian RR, van Kessel JC. 2017. Quorum sensing gene regulation by LuxR/HapR master regulators in vibrios. *Journal of Bacteriology* 199:e00105–e00117 DOI 10.1128/JB.00105-17.
- Basak P, Sadhukhan P, Sarkar P, Sil PC. 2017. Perspectives of the Nrf-2 signaling pathway in cancer progression and therapy. *Toxicology Reports* 4:306–318 DOI 10.1016/j.toxrep.2017.06.002.
- Biagioni F, Ben-Moshe NBossel, Fontemaggi G, Canu V, Mori F, Antoniani B, Di Benedetto A, Santoro R, Germoni S, De Angelis F, Cambria A, Avraham R, Grasso G, Strano S, Muti P, Mottolose M, Yarden Y, Domany E, Blandino G. 2012. miR-10b*, a master inhibitor of the cell cycle, is down-regulated in human breast tumours. *EMBO Molecular Medicine* 4:1214–1229 DOI 10.1002/emmm.201201483.
- Binda E, Visioli A, Giani F, Trivieri N, Palumbo O, Restelli S, Dezi F, Mazza T, Fusilli C, Legnani F, Carella M, Di Meco F, Duggal R, Vescovi AL. 2017. Wnt5a drives an invasive phenotype in human glioblastoma stem-like cells. *Cancer Research* 77:996–1007 DOI 10.1158/0008-5472.CAN-16-1693.
- Boboila S, Lopez G, Yu J, Banerjee D, Kadenhe-Chiweshe A, Connolly EP, Kandel JJ, Rajbhandari P, Silva JM, Califano A, Yamashiro DJ. 2018. Transcription factor

- activating protein 4 is synthetically lethal and a master regulator of MYCN-amplified neuroblastoma. *Oncogene* **37**:5451–5465 DOI [10.1038/s41388-018-0326-9](https://doi.org/10.1038/s41388-018-0326-9).
- Boogerd CJ, Evans SM. 2016.** TBX5 and NuRD divide the heart. *Developmental Cell* **36**:242–244 DOI [10.1016/j.devcel.2016.01.015](https://doi.org/10.1016/j.devcel.2016.01.015).
- Botchkarev VV, Haber JE. 2018.** Functions and regulation of the Polo-like kinase Cdc5 in the absence and presence of DNA damage. *Current Genetics* **64**:87–96 DOI [10.1007/s00294-017-0727-2](https://doi.org/10.1007/s00294-017-0727-2).
- Bresnick EH, Johnson KD. 2019.** Blood disease-causing and -suppressing transcriptional enhancers: general principles and mechanisms. *Blood Advances* **3**:2045–2056 DOI [10.1182/bloodadvances.2019000378](https://doi.org/10.1182/bloodadvances.2019000378).
- Brotto DB, Siena ÁDD, De Barros II, Carvalho SdCES, Muys BR, Goedert L, Cardoso C, Praça JR, Ramão A, Squire JA, Araujo LF, WAd Silva. 2020.** Contributions of HOX genes to cancer hallmarks: enrichment pathway analysis and review. *Tumour Biology* **42**:1–16 DOI [10.1177/1010428320918050](https://doi.org/10.1177/1010428320918050).
- Buendia I, Michalska P, Navarro E, Gameiro I, Egea J, León R. 2016.** Nrf2-ARE pathway: an emerging target against oxidative stress and neuroinflammation in neurodegenerative diseases. *Pharmacology & Therapeutics* **157**:84–104 DOI [10.1016/j.pharmthera.2015.11.003](https://doi.org/10.1016/j.pharmthera.2015.11.003).
- Candini O, Spano C, Murgia A, Grisendi G, Veronesi E, Piccinno MS, Ferracin M, Negrini M, Giacobbi F, Bambi F, Horwitz EM, Conte P, Paolucci P, Dominici M. 2015.** Mesenchymal progenitors aging highlights a miR-196 switch targeting HOXB7 as master regulator of proliferation and osteogenesis. *Stem Cells* **33**:939–950 DOI [10.1002/stem.1897](https://doi.org/10.1002/stem.1897).
- Castaño J, Aranda S, Bueno C, Calero-Nieto FJ, Mejia-Ramirez E, Mosquera JL, Blanco E, Wang X, Prieto C, Zabaleta L, Mereu E, Rovira M, Jiménez-Delgado S, Matson DR, Heyn H, Bresnick EH, Göttgens B, Croce LDi, Menendez P, Raya A, Giorgetti A. 2019.** GATA2 promotes hematopoietic development and represses cardiac differentiation of human mesoderm. *Stem Cell Reports* **13**:515–529 DOI [10.1016/j.stemcr.2019.07.009](https://doi.org/10.1016/j.stemcr.2019.07.009).
- Cattenoz PB, Giangrande A. 2016.** Revisiting the role of the Gcm transcription factor, from master regulator to Swiss army knife. *Fly* **10**:210–218 DOI [10.1080/19336934.2016.1212793](https://doi.org/10.1080/19336934.2016.1212793).
- Chatterjee A, Cui Y, Chatterjee AK. 2009.** RsmC of *Erwinia carotovora* subsp. *carotovora* negatively controls motility, extracellular protein production, and virulence by binding FlhD and modulating transcriptional activity of the master regulator, FlhDC. *Journal of Bacteriology* **191**:4582–4593 DOI [10.1128/JB.00154-09](https://doi.org/10.1128/JB.00154-09).
- Chen L, Fulcoli FG, Tang S, Baldini A. 2009.** Tbx1 regulates proliferation and differentiation of multipotent heart progenitors. *Circulation Research* **105**:842–851 DOI [10.1161/CIRCRESAHA.109.200295](https://doi.org/10.1161/CIRCRESAHA.109.200295).
- Chen M, Guo Z, Ju W, Ryffel B, He X, Zheng SG. 2012.** The development and function of follicular helper T cells in immune responses. *Cellular & Molecular Immunology* **9**:375–379 DOI [10.1038/cmi.2012.18](https://doi.org/10.1038/cmi.2012.18).

- Chu F, Kearns DB, Branda SS, Kolter R, Losick R. 2006.** Targets of the master regulator of biofilm formation in *Bacillus subtilis*. *Molecular Microbiology* **59**:1216–1228 DOI [10.1111/j.1365-2958.2005.05019.x](https://doi.org/10.1111/j.1365-2958.2005.05019.x).
- Collins EM, Thompson A. 2018.** HOX genes in normal, engineered and malignant hematopoiesis. *The International Journal of Developmental Biology* **62**:847–856 DOI [10.1387/ijdb.180206at](https://doi.org/10.1387/ijdb.180206at).
- Colomba A, Ridley AJ. 2014.** Analyzing the roles of rho gtpases in cancer cell migration with a live cell imaging 3d-morphology-based assay. In: Trabalzini L, Retta S, eds. *Ras signaling. Methods in Molecular Biology (Methods and Protocols)*. Vol 1120. Totowa: Humana Press, 327–337 DOI [10.1007/9781627037914_21](https://doi.org/10.1007/9781627037914_21).
- Combes G, Alharbi I, Braga LG, Elowe S. 2017.** Playing polo during mitosis: PLK1 takes the lead. *Oncogene* **36**:4819–4827 DOI [10.1038/onc.2017.113](https://doi.org/10.1038/onc.2017.113).
- Cores Á, Piquero M, Villacampa M, León R, Menéndez JC. 2020.** NRF2 regulation processes as a source of potential drug targets against neurodegenerative diseases. *Biomolecules* **10**:904–941 DOI [10.3390/biom10060904](https://doi.org/10.3390/biom10060904).
- Costa C, Germena G, Martin-Conte EL, Molineris I, Bosco E, Marengo S, Az-zolino O, Altruda F, Ranieri VM, Hirsch E. 2011.** The RacGAP ArhGAP15 is a master negative regulator of neutrophil functions. *Blood* **118**:1099–1108 DOI [10.1182/blood.201012324756](https://doi.org/10.1182/blood.201012324756).
- Courtial N, Smink JJ, Kuvardina ON, Leutz A, Göthert JR, Lausen J. 2012.** Tall regulates osteoclast differentiation through suppression of the master regulator of cell fusion DC-STAMP. *The FASEB Journal* **26**:523–532 DOI [10.1096/fj.11-190850](https://doi.org/10.1096/fj.11-190850).
- Cui Y, Chatterjee A, Yang H, Chatterjee AK. 2008.** Regulatory network controlling extracellular proteins in *Erwinia carotovora* subsp. *carotovora*: FlhDC, the master regulator of flagellar genes, activates rsmB regulatory RNA production by affecting *gacA* and *hexA* (*lrhA*) expression. *Journal of Bacteriology* **190**:4610–4623 DOI [10.1128/JB.01828-07](https://doi.org/10.1128/JB.01828-07).
- Cunningham KA, Bouagnon AD, Barros AG, Lin L, Malard L, Romano-Silva MA, Ashrafi K. 2014.** Loss of a neural AMP-activated kinase mimics the effects of elevated serotonin on fat, movement, and hormonal secretions. *PLOS Genetics* **10**:e1004394–e1004409 DOI [10.1371/journal.pgen.1004394](https://doi.org/10.1371/journal.pgen.1004394).
- Danese S. 2008a.** VEGF in inflammatory bowel disease: a master regulator of mucosal immune-driven angiogenesis. *Digestive and Liver Disease* **40**:680–683 DOI [10.1016/j.dld.2008.02.036](https://doi.org/10.1016/j.dld.2008.02.036).
- Danese S. 2008b.** VEGF in inflammatory bowel disease: a master regulator of mucosal immune-driven angiogenesis. *Digestive and Liver Disease* **40**:680–683 DOI [10.1016/j.dld.2008.02.036](https://doi.org/10.1016/j.dld.2008.02.036).
- Datta R, Paul S. 2015.** Plant microRNAs: master regulator of gene expression mechanism. *Cell Biology International* **39**:1185–1190 DOI [10.1002/cbin.10502](https://doi.org/10.1002/cbin.10502).
- Dickson GJ, Lappin TR, Thompson A. 2009.** Complete array of HOX gene expression by RQ-PCR. In: Eric So CW, ed. *Leukemia. Methods in Molecular Biology* (Methods and Protocols). 538. Totowa: Humana Press, 369–393.

- Ding N, Gao Y, Wang N, Li H. 2011.** Functional analysis of the chicken PPAR γ gene 5'-flanking region and C/EBP α -mediated gene regulation. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology* **158**:297–303 DOI [10.1016/j.cbpb.2011.01.001](https://doi.org/10.1016/j.cbpb.2011.01.001).
- Distefano MB, Kjos I, Bakke O, Progida C. 2015.** Rab7b at the intersection of intracellular trafficking and cell migration. *Communicative & Integrative Biology* **8**:e1023492–e1023495 DOI [10.1080/19420889.2015.1023492](https://doi.org/10.1080/19420889.2015.1023492).
- Donnelly S, Huston WM, Johnson M, Tiberti N, Saunders B, O'Brien B, Burke C, Labbate M, Combes V. 2017.** Targeting the master regulator mTOR: a new approach to prevent the neurological consequences of parasitic infections? *Parasites & Vectors* **10**:581–581 DOI [10.1186/s13071-017-2528-3](https://doi.org/10.1186/s13071-017-2528-3).
- Doostparast Torshizi A, Wang K. 2017.** Deconvolution of transcriptional networks in post-traumatic stress disorder uncovers master regulators driving innate immune system function. *Scientific Reports* **7**:14486–14486 DOI [10.1038/s41598-017-15221-y](https://doi.org/10.1038/s41598-017-15221-y).
- Du L, Ning Z, Zhang H, Liu F. 2017.** Corepressor metastasis-associated protein 3 modulates epithelial-to-mesenchymal transition and metastasis. *Chinese Journal of Cancer* **36**:28–38 DOI [10.1186/s40880-017-0193-8](https://doi.org/10.1186/s40880-017-0193-8).
- Eaton JL, Glasgow E. 2006.** The zebrafish bHLH PAS transcriptional regulator, single-minded 1 (sim1), is required for isotocin cell development. *Developmental Dynamics* **235**:2071–2082 DOI [10.1002/dvdy.20848](https://doi.org/10.1002/dvdy.20848).
- El-Arabey AA, Denizli M, Kanlikilicer P, Bayraktar R, Ivan C, Rashed M, Kabil N, Ozpolat B, Calin GA, Salama SA, Abd-Allah AR, Sood AK, Lopez-Berestein G. 2020.** GATA3 as a master regulator for interactions of tumor-associated macrophages with high-grade serous ovarian carcinoma. *Cellular Signalling* **68**:109539–109563 DOI [10.1016/j.cellsig.2020.109539](https://doi.org/10.1016/j.cellsig.2020.109539).
- Farnebo M, Bykov VJN, Wiman KG. 2010.** The p53 tumor suppressor: A master regulator of diverse cellular processes and therapeutic target in cancer. *Biochemical and Biophysical Research Communications* **396**:85–89 DOI [10.1016/j.bbrc.2010.02.152](https://doi.org/10.1016/j.bbrc.2010.02.152).
- Fazilaty H, Gardaneh M, Bahrami T, Salmaninejad A, Behnam B. 2013.** Crosstalk between breast cancer stem cells and metastatic niche: emerging molecular metastasis pathway? *Tumor Biology* **34**:2019–2030 DOI [10.1007/s13277-013-0831-y](https://doi.org/10.1007/s13277-013-0831-y).
- Fernandez-Marcos PJ, Auwerx J. 2011.** Regulation of PGC-1 α , a nodal regulator of mitochondrial biogenesis. *The American Journal of Clinical Nutrition* **93**(884S-8890): DOI [10.3945/ajcn.110.001917](https://doi.org/10.3945/ajcn.110.001917).
- Filone CM, Caballero IS, Dower K, Mendillo ML, Cowley GS, Santagata S, Rozelle DK, Yen J, Rubins KH, Hacohen N, Root DE, Hensley LE, Connor J. 2014.** The master regulator of the cellular stress response (HSF1) is critical for orthopoxvirus infection. *PLOS Pathogens* **10**:e1003904–e1003917 DOI [10.1371/journal.ppat.1003904](https://doi.org/10.1371/journal.ppat.1003904).
- Fischer JW. 2010.** Protein kinase C delta: a master regulator of apoptosis in neointimal thickening? *Cardiovascular Research* **85**:407–408 DOI [10.1093/cvr/cvp390](https://doi.org/10.1093/cvr/cvp390).
- Foroughi Asl H, Talukdar HA, Kindt ASD, Jain Rajeev K, Ermel R, Ruusalepp A, Nguyen K-DH, Dobrin R, Reilly DF, Schunkert H, Samani NJ, Braenne I, Erdmann J, Melander O, Qi J, Ivert T, Skogsberg J, Schadt EE, Michoel T, Björkegren Johan**

- LM. 2015.** Expression quantitative trait loci acting across multiple tissues are enriched in inherited risk for coronary artery disease. *Circulation: Cardiovascular Genetics* **8**:305–315 DOI [10.1161/CIRCGENETICS.114.000640](https://doi.org/10.1161/CIRCGENETICS.114.000640).
- Frohling S, Scholl C, Bansal D, Huntly BJP. 2007.** HOX gene regulation in acute myeloid leukemia: CDX marks the spot? *Cell Cycle* **6**:2241–2245 DOI [10.4161/cc.6.18.4656](https://doi.org/10.4161/cc.6.18.4656).
- Fry TJ, Mackall CL. 2001.** Interleukin-7: master regulator of peripheral T-cell homeostasis? *Trends in Immunology* **22**:564–571 DOI [10.1016/S1471-4906\(01\)02028-2](https://doi.org/10.1016/S1471-4906(01)02028-2).
- Fujita M, Losick R. 2005.** Evidence that entry into sporulation in *Bacillus subtilis* is governed by a gradual increase in the level and activity of the master regulator Spo0A. *Genes & Development* **19**:2236–2244 DOI [10.1101/gad.1335705](https://doi.org/10.1101/gad.1335705).
- Fujiwara T. 2017.** Transcriptional regulation of erythropoiesis. *Rinsho Ketsueki* **58**:643–648.
- Garry DJ. 2016.** Etv2 is a master regulator of hematoendothelial lineages. *Transactions of the American Clinical and Climatological Association* **127**:212–223.
- Gianni-Barrera R, Bartolomeo M, Vollmar B, Djonov V, Banfi A. 2014.** Split for the cure: VEGF, PDGF-BB and intussusception in therapeutic angiogenesis. *Biochemical Society Transactions* **42**:1637–1642 DOI [10.1042/BST20140234](https://doi.org/10.1042/BST20140234).
- Golan-Mashiach M, Grunspan M, Emmanuel R, Gibbs-Bar L, Dikstein R, Shapiro E. 2012.** Identification of CTCF as a master regulator of the clustered protocadherin genes. *Nucleic Acids Research* **40**:3378–3391 DOI [10.1093/nar/gkr1260](https://doi.org/10.1093/nar/gkr1260).
- Gómez-Orte E, Sáenz-Narciso B, Moreno S, Cabello J. 2013.** Multiple functions of the noncanonical Wnt pathway. *Trends in Genetics* **29**:545–553 DOI [10.1016/j.tig.2013.06.003](https://doi.org/10.1016/j.tig.2013.06.003).
- Gong X, Qian H, Shao W, Li J, Wu J, Liu J-J, Li W, Wang H-W, Espenshade P, Yan N. 2016.** Complex structure of the fission yeast SREBP-SCAP binding domains reveals an oligomeric organization. *Cell Research* **26**:1197–1211 DOI [10.1038/cr.2016.123](https://doi.org/10.1038/cr.2016.123).
- González A, Bes MT, Valladares A, Peleato ML, Fillat MF. 2012.** FurA is the master regulator of iron homeostasis and modulates the expression of tetrapyrrole biosynthesis genes in *Anabaena* sp, PCC 7120. *Environmental Microbiology* **14**:3175–3187 DOI [10.1111/j.1462-2920.2012.02897](https://doi.org/10.1111/j.1462-2920.2012.02897).
- Gora KG, Tsokos CG, Chen YE, Srinivasan BS, Perchuk BS, Laub MT. 2010.** A cell-type-specific protein-protein interaction modulates transcriptional activity of a master regulator in *Caulobacter crescentus*. *Molecular Cell* **39**:455–467 DOI [10.1016/j.molcel.2010.06.024](https://doi.org/10.1016/j.molcel.2010.06.024).
- Grier DG, Thompson A, Kwasniewska A, McGonigle GJ, Halliday HL, Lappin TR. 2005.** The pathophysiology of HOX genes and their role in cancer. *The Journal of Pathology* **205**:154–171 DOI [10.1002/path.1710](https://doi.org/10.1002/path.1710).
- Grifoni D, Bellosta P. 2015.** *Drosophila* Myc: a master regulator of cellular performance. *Biochimica et Biophysica Acta* **1849**:570–581 DOI [10.1016/j.bbagr.2014.06.021](https://doi.org/10.1016/j.bbagr.2014.06.021).
- Gutiérrez L, Caballero N, Fernández-Calleja L, Karkoulia E, Strouboulis J. 2020.** Regulation of GATA1 levels in erythropoiesis. *IUBMB Life* **72**(1):89–105 DOI [10.1002/iub.2192](https://doi.org/10.1002/iub.2192).

- Liu H-C, Charng Y-y. 2012.** Acquired thermotolerance independent of heat shock factor A1 (HsfA1), the master regulator of the heat stress response. *Plant Signaling & Behavior* 7:547–550 DOI [10.4161/psb.19803](https://doi.org/10.4161/psb.19803).
- Häcker G, Vaux DL. 1995.** Apoptosis: a sticky business. *Current Biology* 5:622–624.
- Hayes JD, Dinkova-Kostova AT. 2017.** Epigenetic control of NRF2-Directed cellular antioxidant status in dictating life-death decisions. *Molecular Cell* 68:5–7 DOI [10.1016/j.molcel.2017.09.023](https://doi.org/10.1016/j.molcel.2017.09.023).
- Hernández-Lemus E, Baca-López K, Tovar H. 2015.** What makes a transcriptional master regulator? *A Systems Biology Approach* 161–174 DOI [10.1007/978-3-319-21687-4_10](https://doi.org/10.1007/978-3-319-21687-4_10).
- Hinds PW. 2003.** Cdk2 dethroned as master of S phase entry. *Cancer Cell* 3:305–307 DOI [10.1016/S1535-6108\(03\)00084-9](https://doi.org/10.1016/S1535-6108(03)00084-9).
- Holmberg Olausson K, Nistér M, Lindström MS. 2012.** p53 -dependent and -independent nucleolar stress responses. *Cells* 1:774–798 DOI [10.3390/cells1040774](https://doi.org/10.3390/cells1040774).
- Hong Y-K, Detmar M. 2003.** Prox1, master regulator of the lymphatic vasculature phenotype. *Cell and Tissue Research* 314:85–92 DOI [10.1007/s00441-003-0747-8](https://doi.org/10.1007/s00441-003-0747-8).
- Hu Y, Wang J, Yang B, Zheng N, Qin M, Ji Y, Lin G, Tian L, Wu X, Wu L, Sun B. 2011.** Guanylate binding protein 4 negatively regulates virus-induced type I IFN and antiviral response by targeting IFN regulatory factor 7. *The Journal of Immunology* 187:6456–6462 DOI [10.4049/jimmunol.1003691](https://doi.org/10.4049/jimmunol.1003691).
- Huja S, Oren Y, Biran D, Meyer S, Dobrindt U, Bernhard J, Becher D, Hecker M, Sorek R, Ron EZ. 2014.** Fur is the master regulator of the extraintestinal pathogenic *Escherichia coli* response to serum. *mBio* 5:e01460–14 DOI [10.1128/mBio.01460-14](https://doi.org/10.1128/mBio.01460-14).
- Hwang J-Y, Zukin RS. 2018.** REST, a master transcriptional regulator in neurodegenerative disease. *Current Opinion in Neurobiology* 48:193–200 DOI [10.1016/j.conb.2017.12.008](https://doi.org/10.1016/j.conb.2017.12.008).
- Ichikawa M, Asai T, Chiba S, Kurokawa M, Ogawa S. 2004.** Runx1/AML-1 ranks as a master regulator of adult hematopoiesis. *Cell Cycle* 3:720–722 DOI [10.4161/cc.3.6.951](https://doi.org/10.4161/cc.3.6.951).
- Inoue J, Ihara Y, Tsukamoto D, Yasumoto K, Hashidume T, Kamimura K, Nakai Y, Hirano S, Shimizu M, Kominami R, Sato R. 2016.** Identification of BCL11B as a regulator of adipogenesis. *Scientific Reports* 6:32750–32750 DOI [10.1038/srep32750](https://doi.org/10.1038/srep32750).
- Ishida M, E. Oguchi M, Fukuda M. 2016.** Multiple types of guanine nucleotide exchange factors (GEFs) for Rab Small GTPases. *Cell Structure and Function* 41:61–79 DOI [10.1247/csf.16008](https://doi.org/10.1247/csf.16008).
- Jakob S, Lovell-Badge R. 2011.** Sex determination and the control of Sox9 expression in mammals. *The FEBS Journal* 278:1002–1009 DOI [10.1111/j.1742-4658.2011.08029.x](https://doi.org/10.1111/j.1742-4658.2011.08029.x).
- Jakobsson T, Treuter E, Gustafsson J-Å, Steffensen KR. 2012.** Liver X receptor biology and pharmacology: new pathways, challenges and opportunities. *Trends in Pharmacological Sciences* 33:394–404 DOI [10.1016/j.tips.2012.03.013](https://doi.org/10.1016/j.tips.2012.03.013).
- Janky Rs, Binda MM, Allemeersch J, Vanden Broeck A, Govaere O, Swinnen JV, Roskams T, Aerts S, Topal B. 2016.** Prognostic relevance of molecular subtypes and master regulators in pancreatic ductal adenocarcinoma. *BMC Cancer* 16:632 DOI [10.1186/s12885-016-2540-6](https://doi.org/10.1186/s12885-016-2540-6).

- Jaskólska M, Gerdes K. 2015. CRP-dependent positive autoregulation and proteolytic degradation regulate competence activator Sxy of *Escherichia coli*. *Molecular Microbiology* 95:833–845 DOI 10.1111/mmi.12901.
- Jeffery JM, Kalimutho M, Johansson P, Cardenas DG, Kumar R, Khanna KK. 2017. FBXO31 protects against genomic instability by capping FOXM1 levels at the G2/M transition. *Oncogene* 36:1012–1022 DOI 10.1038/onc.2016.268.
- John SA, Garrett-Sinha LA. 2009. Blimp1: a conserved transcriptional repressor critical for differentiation of many tissues. *Experimental Cell Research* 315:1077–1084 DOI 10.1016/j.yexcr.2008.11.015.
- Julian LM, McDonald ACH, Stanford WL. 2017. Direct reprogramming with SOX factors: masters of cell fate. *Current Opinion in Genetics & Development* 46:24–36 DOI 10.1016/j.gde.2017.06.005.
- Kang J, Yoo J, Lee S, Tang W, Aguilar B, Ramu S, Choi I, Otu HH, Shin JW, Dotto GP, Koh CJ, Detmar M, Hong Y-K. 2010. An exquisite cross-control mechanism among endothelial cell fate regulators directs the plasticity and heterogeneity of lymphatic endothelial cells. *Blood* 116:140–150 DOI 10.1182/blood-2009-11-252270.
- Kang Y-A, Sanalkumar R, O'Geen H, Linnemann AK, Chang C-J, Bouhassira EE, Farnham PJ, Keles S, Bresnick EH. 2012. Autophagy driven by a master regulator of hematopoiesis. *Molecular and Cellular Biology* 32:226–239 DOI 10.1128/MCB.06166-11.
- Katsumura KR, Mehta C, Hewitt KJ, Soukup AA, Fragade Andrade I, Ranheim EA, Johnson KD, Bresnick EH. 2018. Human leukemia mutations corrupt but do not abrogate GATA-2 function. *Proceedings of the National Academy of Sciences of the United States of America* 115:E10109–E10118 DOI 10.1073/pnas.1813015115.
- Katsumura KR, Yang C, Boyer ME, Li L, Bresnick EH. 2014. Molecular basis of crosstalk between oncogenic Ras and the master regulator of hematopoiesis GATA-2. *EMBO Reports* 15:938–947 DOI 10.15252/embr.201438808.
- Kaul V, Krams S. 2015. MicroRNAs as master regulators of immune responses in transplant recipients. *Current Opinion in Organ Transplantation* 20:29–36 DOI 10.1097/MOT.000000000000148.
- Kazan K, Manners JM. 2013. MYC2: the master in action. *Molecular Plant* 6:686–703 DOI 10.1093/mp/sss128.
- Ke ZY, Yang SJ. 2017. Role of master transcriptional factor Prox-1 in lymphatic endothelial differentiation of Kaposiform hemangioendothelioma. 46:176–181 DOI 10.3760/cma.j.issn.0529-5807.2017.03.007.
- Kim YC, Guan K-L. 2015. mTOR: a pharmacologic target for autophagy regulation. *The Journal of Clinical Investigation* 125:25–32 DOI 10.1172/JCI73939.
- Kojima H, Ieda M. 2017. Discovery and progress of direct cardiac reprogramming. *Cellular and Molecular Life Sciences* 74:2203–2215 DOI 10.1007/s00018-017-2466-4.
- Kondratyeva LG, Chernov IP, Zinovyeva MV, Kopantzev EP, Sverdlov ED. 2017. Expression of master regulatory genes of embryonic development in pancreatic tumors. *Doklady Biochemistry and Biophysics* 475:250–252 DOI 10.1134/S1607672917040020.

- Kozhukhar VG. 2012.** SRY and SOX9: the main genetic factors of mammalian sex determination. *Tsitologiya* **54**:390–404.
- Krappmann D, Wegener E, Sunami Y, Esen M, Thiel A, Mordmuller B, Scheidereit C. 2004.** The IkappaB kinase complex and NF-kappaB act as master regulators of lipopolysaccharide-induced gene expression and control subordinate activation of AP-1. *Molecular and Cellular Biology* **24**:6488–6500 DOI [10.1128/MCB.24.14.6488-6500.2004](https://doi.org/10.1128/MCB.24.14.6488-6500.2004).
- Krycer JR, Sharpe LJ, Luu W, Brown AJ. 2010.** The Akt–SREBP nexus: cell signaling meets lipid metabolism. *Trends in Endocrinology & Metabolism* **21**:268–276 DOI [10.1016/j.tem.2010.01.001](https://doi.org/10.1016/j.tem.2010.01.001).
- Langrish CL, McKenzie BS, Wilson NJ, DeWaal Malefyt R, Kastelein RA, Cua DJ. 2004.** IL-12 and IL-23: master regulators of innate and adaptive immunity. *Immunological Reviews* **202**:96–105 DOI [10.1111/j.0105-2896.2004.00214.x](https://doi.org/10.1111/j.0105-2896.2004.00214.x).
- Laub MT, Chen SL, Shapiro L, McAdams HH. 2002.** Genes directly controlled by CtrA, a master regulator of the Caulobacter cell cycle. *Proceedings of the National Academy of Sciences of the United States of America* **99**:4632–4637 DOI [10.1073/pnas.062065699](https://doi.org/10.1073/pnas.062065699).
- Lee J-E, Ge K. 2014.** Transcriptional and epigenetic regulation of PPAR γ expression during adipogenesis. *Cell & Bioscience* **4**:29–29 DOI [10.1186/2045-3701-4-29](https://doi.org/10.1186/2045-3701-4-29).
- Lee S-Y, Kim E-Y, Kim K-H, Lee K-A. 2016.** Bcl2l10, a new Tpx2 binding partner, is a master regulator of Aurora kinase A in mouse oocytes. *Cell Cycle* **15**:3296–3305 DOI [10.1080/15384101.2016.1243630](https://doi.org/10.1080/15384101.2016.1243630).
- Lefebvre C, Rajbhandari P, Alvarez MJ, Bandaru P, Lim WK, Sato M, Wang K, Sumazin P, Kustagi M, Bisikirska BC, Basso K, Beltrao P, Krogan N, Gautier J, Dalla-Favera R, Califano A. 2010.** A human B-cell interactome identifies MYB and FOXM1 as master regulators of proliferation in germinal centers. *Molecular Systems Biology* **6**:377 DOI [10.1038/msb.2010.31](https://doi.org/10.1038/msb.2010.31).
- Lehrke M, Lazar MA. 2005.** The many faces of PPARgamma. *Cell* **123**:993–999 DOI [10.1016/j.cell.2005.11.026](https://doi.org/10.1016/j.cell.2005.11.026).
- Leonards K, Almosailleakh M, Tauchmann S, Bagger FO, Thirant C, Juge S, Bock T, Méreau H, Bezerra MF, Tzankov A, Ivanek R, Losson R, Peters AHFM, Mercher T, Schwaller J. 2020.** Nuclear interacting SET domain protein 1 inactivation impairs GATA1-regulated erythroid differentiation and causes erythroleukemia. *Nature Communications* **11**:2807 DOI [10.1038/s41467-020-16179-8](https://doi.org/10.1038/s41467-020-16179-8).
- Levine MS, Holland Andrew J. 2014.** Polo-like Kinase 4 Shapes Up. *Structure* **22**:1071–1073 DOI [10.1016/j.str.2014.07.004](https://doi.org/10.1016/j.str.2014.07.004).
- Levy C, Khaled M, Fisher DE. 2006.** MITF: master regulator of melanocyte development and melanoma oncogene. *Trends in Molecular Medicine* **12**:406–414 DOI [10.1016/j.molmed.2006.07.008](https://doi.org/10.1016/j.molmed.2006.07.008).
- Li M, Alsager JS, Wang Z, Cheng L, Shan B. 2020.** Epigenetic upregulation of in non-small lung cancer cells. *Aging* **12**:16921–16935 DOI [10.18632/aging.103597](https://doi.org/10.18632/aging.103597).
- Li N, Dobrev D, Wehrens XHT. 2016.** PITX2: a master regulator of cardiac channelopathy in atrial fibrillation? *Cardiovascular Research* **109**:345–347 DOI [10.1093/cvr/cvw008](https://doi.org/10.1093/cvr/cvw008).

- Li R, Campos J, Iida J. 2015.** A gene regulatory program in human breast cancer. *Genetics* **201**:1341–1348 DOI [10.1534/genetics.115.180125](https://doi.org/10.1534/genetics.115.180125).
- Liang Y, Liang Q, Qiao L, Xiao F. 2020.** MicroRNAs modulate drug resistance-related mechanisms in hepatocellular carcinoma. *Frontiers in Oncology* **10**:920 DOI [10.3389/fonc.2020.00920](https://doi.org/10.3389/fonc.2020.00920).
- Lim WK, Lyashenko E, Califano A. 2009.** Master regulators used as breast cancer metastasis classifier. *Pacific Symposium on Biocomputing Pacific Symposium on Biocomputing* **2009**:504–515.
- Lin J, Huo X, Liu X. 2017a.** mTOR signaling pathway: a potential target of curcumin in the treatment of spinal cord injury. *BioMed Research International* **2017**:1634801.
- Lin S-C, Lee H-C, Hou P-C, Fu J-L, Wu M-H, Tsai S-J. 2017b.** Targeting hypoxia-mediated YAP1 nuclear translocation ameliorates pathogenesis of endometriosis without compromising maternal fertility. *The Journal of Pathology* **242**:476–487 DOI [10.1002/path.4922](https://doi.org/10.1002/path.4922).
- Liston A. 2010.** Is foxp3 the master regulator of regulatory T cells? *Progress in Molecular Biology and Translational Science* **92**(10):315–317 DOI [10.1016/S1877-1173\(10\)92017-6](https://doi.org/10.1016/S1877-1173(10)92017-6).
- Liu C-F, Samsa WE, Zhou G, Lefebvre V. 2017.** Transcriptional control of chondrocyte specification and differentiation. *Seminars in Cell & Developmental Biology* **62**:34–49 DOI [10.1016/j.semcd.2016.10.004](https://doi.org/10.1016/j.semcd.2016.10.004).
- Liu Z-J, Semenza GL, Zhang H-F. 2015.** Hypoxia-inducible factor 1 and breast cancer metastasis. *Journal of Zhejiang University Science B* **16**:32–43 DOI [10.1631/jzus.B1400221](https://doi.org/10.1631/jzus.B1400221).
- Lourenço AR, Coffey PJ. 2017.** SOX4: joining the master regulators of epithelial-to-mesenchymal transition? *Trends in Cancer* **3**:571–582 DOI [10.1016/j.trecan.2017.06.002](https://doi.org/10.1016/j.trecan.2017.06.002).
- Lu L-F, Li S, Lu X-B, Zhang Y-A. 2015.** Functions of the two zebrafish MAVS variants are opposite in the induction of IFN1 by targeting IRF7. *Fish & Shellfish Immunology* **45**:574–582 DOI [10.1016/j.fsi.2015.05.019](https://doi.org/10.1016/j.fsi.2015.05.019).
- Lu X, Kang Y. 2010.** Hypoxia and hypoxia-inducible factors: master regulators of metastasis. *Clinical Cancer Research* **16**:5928–5935 DOI [10.1158/1078-0432.CCR-10-1360](https://doi.org/10.1158/1078-0432.CCR-10-1360).
- Luza S, Opazo CM, Bousman CA, Pantelis C, Bush AI, Everall IP. 2020.** The ubiquitin proteasome system and schizophrenia. *The Lancet Psychiatry* **7**:528–537 DOI [10.1016/S2215-0366\(19\)30520-6](https://doi.org/10.1016/S2215-0366(19)30520-6).
- Ma Y, Li J, Yao Y, Wei D, Wang R, Wu Q. 2016.** A controlled double-duration inducible gene expression system for cartilage tissue engineering. *Scientific Reports* **6**:26617 DOI [10.1038/srep26617](https://doi.org/10.1038/srep26617).
- Madison BB. 2016.** Srebp2: a master regulator of sterol and fatty acid synthesis. *Journal of Lipid Research* **57**:333–335 DOI [10.1194/jlr.C066712](https://doi.org/10.1194/jlr.C066712).
- Magnusson M, Brun ACM, Miyake N, Larsson J, Ehinger M, Bjornsson JM, Wutz A, Sigvardsson M, Karlsson S. 2007.** HOXA10 is a critical regulator for hematopoietic stem cells and erythroid/megakaryocyte development. *Blood* **109**:3687–3696 DOI [10.1182/blood-2006-10-054676](https://doi.org/10.1182/blood-2006-10-054676).

- Matroule J-Y, Volanti C, Piette J. 2006.** NF- κ B in photodynamic therapy: discrepancies of a master regulator. *Photochemistry and Photobiology* **82**:1241–1246 DOI [10.1562/2006-03-30-IR-862](https://doi.org/10.1562/2006-03-30-IR-862).
- Matsumoto Y, Nagoshi H, Yoshida M, Kato S, Kuroda J, Shimura K, Kaneko H, Horiike S, Nakamura S, Taniwaki M. 2017.** Expression of master regulators of T-cell, helper T-cell and follicular helper T-cell differentiation in angioimmunoblastic T-cell lymphoma. *Internal Medicine* **56**:2851–2856 DOI [10.2169/internalmedicine.8570-16](https://doi.org/10.2169/internalmedicine.8570-16).
- McGonigle GJ, Lappin TRJ, Thompson A. 2008.** Grappling with the HOX network in hematopoiesis and leukemia. *Frontiers in Bioscience* **4297–4308** DOI [10.2741/3006](https://doi.org/10.2741/3006).
- Medina DL, Di Paola S, Peluso I, Armani A, De Stefani D, Venditti R, Montefusco S, Scotto-Rosato A, Prezioso C, Forrester A, Settembre C, Wang W, Gao Q, Xu H, Sandri M, Rizzuto R, De Matteis MA, Ballabio A. 2015.** Lysosomal calcium signalling regulates autophagy through calcineurin and TFEB. *Nature Cell Biology* **17**:288–299 DOI [10.1111/j.1530-0277.2009.01126.x](https://doi.org/10.1111/j.1530-0277.2009.01126.x).
- Medvedovic J, Ebert A, Tagoh H, Busslinger M. 2011.** Pax5: a master regulator of B cell development and leukemogenesis. *Advances in immunology* **111**:179–206 DOI [10.1016/B9780123859914.000052](https://doi.org/10.1016/B9780123859914.000052).
- Miranda RC, Pietrzykowski AZ, Tang Y, Sathyan P, Mayfield D, Keshavarzian A, Sampson W, Hereld D. 2010.** MicroRNAs: master regulators of ethanol abuse and toxicity?. *Alcoholism, Clinical and Experimental Research* **34**:575–587 DOI [10.1111/j.1530-0277.2009.01126.x](https://doi.org/10.1111/j.1530-0277.2009.01126.x).
- Mirlekar B. 2020.** Co-expression of master transcription factors determines CD4 T cell plasticity and functions in auto-inflammatory diseases. *Immunology Letters* **222**:58–66 DOI [10.1016/j.imlet.2020.03.007](https://doi.org/10.1016/j.imlet.2020.03.007).
- Moe Sharon M. 2012.** Klotho. *Circulation* **125**:2181–2183 DOI [10.1161/CIRCULATIONAHA.112.104828](https://doi.org/10.1161/CIRCULATIONAHA.112.104828).
- Molkentin Jeffery D. 2011.** The transcription factor C/EBP β serves as a master regulator of physiologic cardiac hypertrophy. *Circulation Research* **108**:277–278 DOI [10.1161/RES.0b013e31820ff484](https://doi.org/10.1161/RES.0b013e31820ff484).
- Momand J, Wu H-H, Dasgupta G. 2000.** MDM2—master regulator of the p53 tumor suppressor protein. *Gene* **242**:15–29 DOI [10.1016/S0378-1119\(99\)00487-4](https://doi.org/10.1016/S0378-1119(99)00487-4).
- Mork L, Capel B. 2010.** Oestrogen shuts the door on SOX9. *BMC Biology* **8**:110–112 DOI [10.1186/1741-7007-8-110](https://doi.org/10.1186/1741-7007-8-110).
- Muscariello L, Marino C, Capri U, Vastano V, Marasco R, Sacco M. 2013.** CcpA and three newly identified proteins are involved in biofilm development in *Lactobacillus plantarum*. *Journal of Basic Microbiology* **53**:62–71 DOI [10.1002/jobm.201100456](https://doi.org/10.1002/jobm.201100456).
- Nascimento FRE, Gomes EA, Russo M, Lepique AP. 2015.** Interferon regulatory factor (IRF)-1 is a master regulator of the cross talk between macrophages and L929 fibrosarcoma cells for nitric oxide dependent tumoricidal activity. *PLOS ONE* **10**:e0117782 DOI [10.1371/journal.pone.0117782](https://doi.org/10.1371/journal.pone.0117782).
- Nebral K, Denk D, Attarbaschi A, König M, Mann G, Haas OA, Strehl S. 2009.** Incidence and diversity of PAX5 fusion genes in childhood acute lymphoblastic leukemia. *Leukemia* **23**:134–143 DOI [10.1038/leu.2008.306](https://doi.org/10.1038/leu.2008.306).

- Neurath MF. 2007.** IL-23: a master regulator in Crohn disease. *Nature Medicine* **13**:26–27 DOI [10.1038/nm0107-26](https://doi.org/10.1038/nm0107-26).
- Nicol L, Wilkie H, Gossner A, Watkins C, Dalziel R, Hopkins J. 2016.** Variations in T cell transcription factor gene structure and expression associated with the two disease forms of sheep paratuberculosis. *Veterinary Research* **47**:83–90 DOI [10.1186/s13567-016-0368-3](https://doi.org/10.1186/s13567-016-0368-3).
- Noizet M, Lagoutte E, Gratigny M, Bouschbacher M, Lazareth I, Roest Crollius H, Darzacq X, Dugast-Darzacq C. 2016.** Master regulators in primary skin fibroblast fate reprogramming in a human ex vivo model of chronic wounds. *Wound Repair and Regeneration* **24**:247–262 DOI [10.1111/wrr.12392](https://doi.org/10.1111/wrr.12392).
- Nomura S, Takahashi H, Suzuki J, Kuwahara M, Yamashita M, Sawasaki T. 2019.** Pyrrothiogatain acts as an inhibitor of GATA family proteins and inhibits Th2 cell differentiation in vitro. *Scientific Reports* **9**:17335 DOI [10.1038/s41598-019-53856-1](https://doi.org/10.1038/s41598-019-53856-1).
- Odom DT, Zizlsperger N, Gordon DB, Bell GW, Rinaldi NJ, Murray HL, Volkert TL, Schreiber J, Rolfe PA, Gifford DK, Fraenkel E, Bell GI, Young RA. 2004.** Control of pancreas and liver gene expression by HNF transcription factors. *Science* **303**:1378–1381 DOI [10.1126/science.1089769](https://doi.org/10.1126/science.1089769).
- Ogasawara H, Yamamoto K, Ishihama A. 2010.** Regulatory role of MlrA in transcription activation of csgD, the master regulator of biofilm formation in Escherichia coli. *FEMS Microbiology Letters* **312**:160–168 DOI [10.1111/j.1574-6968.2010.02112.x](https://doi.org/10.1111/j.1574-6968.2010.02112.x).
- Ogura M, Hashimoto H, Tanaka T. 2002.** Med, a cell–surface localized protein regulating a competence transcription factor gene, comK, in Bacillus subtilis. *Bioscience, Biotechnology, and Biochemistry* **66**:892–896 DOI [10.1271/bbb.66.892](https://doi.org/10.1271/bbb.66.892).
- Ohno S. 1978.** Major sex-determining genes. *Monographs on Endocrinology* **11**:1–140.
- Okuno Y, Inoue K, Imai Y. 2013.** Novel insights into histone modifiers in adipogenesis. *Adipocyte* **2**:285–288 DOI [10.4161/adip.25731](https://doi.org/10.4161/adip.25731).
- PA . 2014.** Atypical Rho GTPases RhoD and Rif integrate cytoskeletal dynamics and membrane trafficking. *Biological Chemistry* **395**:477–484 DOI [10.1515/hsz-2013-0296](https://doi.org/10.1515/hsz-2013-0296).
- Peng Y-C, Joyner AL. 2015.** Hedgehog signaling in prostate epithelial-mesenchymal growth regulation. *Developmental Biology* **400**:94–104 DOI [10.1016/j.ydbio.2015.01.019](https://doi.org/10.1016/j.ydbio.2015.01.019).
- Perino A, Ghigo A, Hirsch E. 2010.** Leukocyte and cardiac phosphoinositide 3-kinase γ activity in pressure overload–induced cardiac failure. *Trends in Cardiovascular Medicine* **20**:273–276 DOI [10.1016/j.tcm.2011.12.007](https://doi.org/10.1016/j.tcm.2011.12.007).
- Pfeffer SR. 2017.** Rab GTPases: master regulators that establish the secretory and endocytic pathways. *Molecular Biology of the Cell* **28**:712–715 DOI [10.1091/mbc.e16-10-0737](https://doi.org/10.1091/mbc.e16-10-0737).
- Philipsen S. 2013.** A new twist to the GATA switch. *Blood* **122**:3391–3392.
- Pini F, De Nisco NJ, Ferri L, Penterman J, Fioravanti A, Brillì M, Mengoni A, Bazzicalupo M, Viollier PH, Walker GC, Biondi EG. 2015.** Cell cycle control by the master regulator CtrA in sinorhizobium meliloti. *PLOS Genetics* **11**:e1005232–e1005255 DOI [10.1371/journal.pgen.1005232](https://doi.org/10.1371/journal.pgen.1005232).

- Pompeani AJ, Irgon JJ, Berger MF, Bulyk ML, Wingreen NS, Bassler BL. 2008.** The *Vibrio harveyi* master quorum-sensing regulator, LuxR, a TetR-type protein is both an activator and a repressor: DNA recognition and binding specificity at target promoters. *Molecular Microbiology* **70**:76–88 DOI [10.1111/j.1365-2958.2008.06389](https://doi.org/10.1111/j.1365-2958.2008.06389).
- Qi Y, Liang Z, Wang Z, Lu G, Li G. 2015.** Determination of Rab5 activity in the cell by effector pull-down assay. *Methods in Molecular Biology* **1298**:259–270 DOI [10.1007/978-1-4939-2569-8_22](https://doi.org/10.1007/978-1-4939-2569-8_22).
- Qiao A, Jin X, Pang J, Moskophidis D, Mivechi NF. 2017.** The transcriptional regulator of the chaperone response HSF1 controls hepatic bioenergetics and protein homeostasis. *The Journal of Cell Biology* **216**:723–741 DOI [10.1083/jcb.201607091](https://doi.org/10.1083/jcb.201607091).
- Qin Y, Ekmekcioglu S, Forget M-A, Szekvolgyi L, Hwu P, Grimm EA, Jazaeri AA, Roszik J. 2017.** Cervical cancer neoantigen landscape and immune activity is associated with human papillomavirus master regulators. *Frontiers in Immunology* **8**:689–689 DOI [10.3389/fimmu.2017.00689](https://doi.org/10.3389/fimmu.2017.00689).
- Qu X, Alsager S, Zhuo Y, Shan B. 2019.** HOX transcript antisense RNA (HOTAIR) in cancer. *Cancer Letters* **454**:90–97 DOI [10.1016/j.canlet.2019.04.016](https://doi.org/10.1016/j.canlet.2019.04.016).
- Rajagopalan V, Gerdes AM. 2015.** Role of thyroid hormones in ventricular remodeling. *Current Heart Failure Reports* **12**:141–149 DOI [10.1007/s11897-014-0246-0](https://doi.org/10.1007/s11897-014-0246-0).
- Rawat VPS, Humphries RK, Buske C. 2012.** Beyond Hox: the role of ParaHox genes in normal and malignant hematopoiesis. *Blood* **120**:519–527 DOI [10.1182/blood.201202385898](https://doi.org/10.1182/blood.201202385898).
- Relaix F. 2015.** Pax genes: master regulators of development and tissue homeostasis. *Seminars in Cell & Developmental Biology* **44**:62–63 DOI [10.1016/j.semcdb.2015.10.036](https://doi.org/10.1016/j.semcdb.2015.10.036).
- Resnick MA, Tomso D, Inga A, Menendez D, Bell D. 2005.** Functional diversity in the gene network controlled by the master regulator p53 in humans. *Cell Cycle* **4**:1026–1029 DOI [10.4161/cc.4.8.1904](https://doi.org/10.4161/cc.4.8.1904).
- Rice KL, Licht JD. 2007.** HOX deregulation in acute myeloid leukemia. *The Journal of Clinical Investigation* **117**:865–868 DOI [10.1172/JCI31861](https://doi.org/10.1172/JCI31861).
- Rizzino A. 2008.** Transcription factors that behave as master regulators during mammalian embryogenesis function as molecular rheostats. *Biochemical Journal* **411**:e5–e7 DOI [10.1042/BJ20080479](https://doi.org/10.1042/BJ20080479).
- Rohde M, Warthoe P, Gjetting T, Lukas J, Bartek J, Strauss M. 1996.** The retinoblastoma protein modulates expression of genes coding for diverse classes of proteins including components of the extracellular matrix. *Oncogene* **12**:2393–2401.
- Rojas JM, Avia M, Martín V, Sevilla N. 2017.** IL-10: a multifunctional cytokine in viral infections. *Journal of Immunology Research* **2017**:6104054–6104067 DOI [10.1155/2017/6104054](https://doi.org/10.1155/2017/6104054).
- Rostamzadeh D, Yousefi M, Haghshenas MR, Ahmadi M, Dolati S, Babaloo Z. 2019.** mTOR signaling pathway as a master regulator of memory CD8 T-cells, Th17, and NK cells development and their functional properties. *Journal of Cellular Physiology* **234**:12353–12368 DOI [10.1002/jcp.28042](https://doi.org/10.1002/jcp.28042).
- Samardzija C, Greening DW, Escalona R, Chen M, Bilandzic M, Luwor R, Kannourakis G, Findlay JK, Ahmed N. 2017.** Knockdown of stem cell regulator Oct4A in ovarian cancer reveals cellular reprogramming associated with key regulators of

- cytoskeleton-extracellular matrix remodelling. *Scientific Reports* 7:46312–46329 DOI 10.1038/srep46312.
- Sandovici I, Hammerle CM, Ozanne SE, Constância M. 2013.** Developmental and environmental epigenetic programming of the endocrine pancreas: consequences for type 2 diabetes. *Cellular and Molecular Life Sciences* 70:1575–1595 DOI 10.1007/s00018-013-1297-1.
- Satyanarayana A, Kaldis P. 2009.** A dual role of Cdk2 in DNA damage response. *Cell Division* 4:9–12 DOI 10.1186/1747-1028-4-9.
- Sawle AD, Kebschull M, Demmer RT, Papapanou PN. 2016.** Identification of master regulator genes in human periodontitis. *Journal of Dental Research* 95:1010–1017 DOI 10.1177/0022034516653588.
- Schaefer T, Steiner R, Lengerke C. 2020.** SOX2 and p53 expression control converges in PI3K/AKT signaling with versatile implications for stemness and cancer. *International Journal of Molecular Sciences* 21(14):4902–4922 DOI 10.3390/ijms21144902.
- Schmittgen TD. 2010.** miR-31: a master regulator of metastasis? *Future Oncology* 6:17–20 DOI 10.2217/fon.09.150.
- Schnappauf O, Aksentijevich I. 2020.** Mendelian diseases of dysregulated canonical NF- κ B signaling: from immunodeficiency to inflammation. *Journal of Leukocyte Biology* 108:573–589 DOI 10.1002/JLB.2MR0520-166R.
- Schönenberger MJ, Kovacs WJ. 2015.** Hypoxia signaling pathways: modulators of oxygen-related organelles. *Frontiers in Cell and Developmental Biology* 3:42–60 DOI 10.3389/fcell.2015.00042.
- Semenza GL. 2014.** Hypoxia-inducible factor 1 and cardiovascular disease. *Annual Review of Physiology* 76:39–56 DOI 10.1146/annurev-physiol-021113-170322.
- Semenza GL. 2017.** A compendium of proteins that interact with HIF-1 α . *Experimental Cell Research* 356:128–135 DOI 10.1016/j.yexcr.2017.03.041.
- Settembre C, Malta CDi, Polito VA, Arencibia MGarcia, Vetrini F, Erdin S, Erdin SU, Huynh T, Medina D, Colella P, Sardiello M, Rubinsztein DC, Ballabio A. 2011.** TFEB links autophagy to lysosomal biogenesis. *Science* 332:1429–1433 DOI 10.1126/science.1204592.
- Seymour PA. 2014.** Sox9: a master regulator of the pancreatic program. *The Review of Diabetic Studies: RDS* 11:51–83 DOI 10.1900/RDS.2014.11.51.
- Shaheen R, Tala SA, Almoisheer A, Alkuraya FS. 2014.** Mutation in encoding a master regulator of centriole formation, defines a novel locus for primordial dwarfism. *Journal of Medical Genetics* 51:814–816 DOI 10.1136/jmedgenet-2014-102790.
- Sharma R, Fu SM, Ju S-T. 2011.** IL-2: a two-faced master regulator of autoimmunity. *Journal of Autoimmunity* 36:91–97 DOI 10.1016/j.jaut.2011.01.001.
- Shenoy PS, Bose B, Sharma M, McFarlane C, Kambadur R. 2014.** Lack of myostatin reduces MyoD induced myogenic potential of primary muscle fibroblasts. *Journal of Cellular Biochemistry* 115:1908–1917 DOI 10.1002/jcb.24860.
- Shinkawa T, Tan K, Fujimoto M, Hayashida N, Yamamoto K, Takaki E, Takii R, Prakasam R, Inouye S, Mezger V, Nakai A. 2011.** Heat shock factor 2 is required for maintaining proteostasis against febrile-range thermal stress

- and polyglutamine aggregation. *Molecular Biology of the Cell* **22**:3571–3583 DOI [10.1091/mbc.e11-04-0330](https://doi.org/10.1091/mbc.e11-04-0330).
- Shiotani A, Kamada T, Yamanaka Y, Manabe N, Kusunoki H, Hata J, Haruma K. 2008.** Sonic hedgehog and CDX2 expression in the stomach. *Journal of Gastroenterology and Hepatology* **23**:S161–S166 DOI [10.1111/j.1440-1746.2008.05406.x](https://doi.org/10.1111/j.1440-1746.2008.05406.x).
- Shubham K, Mishra R. 2012.** Pax6 interacts with SPARC and TGF- β in murine eyes. *Molecular Vision* **18**:951–956.
- Siegwart LC, Schwemmers S, Wehrle J, Koellerer C, Seeger T, Gründer A, Pahl HL. 2020.** The transcription factor NFE2 enhances expression of the hematopoietic master regulators SCL/TAL1 and GATA2. *Experimental Hematology* **87**:1–48 DOI [10.1016/j.exphem.2020.06.004](https://doi.org/10.1016/j.exphem.2020.06.004).
- Singh V, Davidson AC, Hume PJ, Humphreys D, Koronakis V. 2019.** Arf GTPase interplay with Rho GTPases in regulation of the actin cytoskeleton. *Small GTPases* **10**:411–418 DOI [10.1080/21541248.2017.1329691](https://doi.org/10.1080/21541248.2017.1329691).
- Slade L, Pulinilkunnil T. 2017.** The MiTF/TFE family of transcription factors: master regulators of organelle signaling. *Metabolism, and Stress Adaptation. Molecular Cancer Research: MCR* **15**:1637–1643 DOI [10.1158/15417786.MCR170320](https://doi.org/10.1158/15417786.MCR170320).
- Soares E, Zhou H. 2018.** Master regulatory role of p63 in epidermal development and disease. *Cellular and Molecular Life Sciences* **75**:1179–1190 DOI [10.1007/s00018-017-2701-z](https://doi.org/10.1007/s00018-017-2701-z).
- Solomon E, Li H, Duhachek Muggy S, Syta E, Zolkiewska A. 2010.** The role of SnoN in transforming growth factor beta1-induced expression of metalloprotease-disintegrin ADAM12. *The Journal of Biological Chemistry* **285**:21969–21977 DOI [10.1074/jbc.M110.133314](https://doi.org/10.1074/jbc.M110.133314).
- Stafford GP, Ogi T, Hughes C. 2005.** Binding and transcriptional activation of non-flagellar genes by the Escherichia coli flagellar master regulator FlhD2C2. *Microbiology* **151**:1779–1788 DOI [10.1099/mic.0.27879-0](https://doi.org/10.1099/mic.0.27879-0).
- Stief A, Altmann S, Hoffmann K, Pant BD, Scheible W-R, Bäurle I. 2014.** Arabidopsis miR156 regulates tolerance to recurring environmental stress through SPL transcription factors. *The Plant Cell* **26**:1792–1807 DOI [10.1105/tpc.114.123851](https://doi.org/10.1105/tpc.114.123851).
- Stolarczyk E, Lord GM, Howard JK. 2014.** The immune cell transcription factor T-bet: a novel metabolic regulator. *Adipocyte* **3**:58–62 DOI [10.4161/adip.26220](https://doi.org/10.4161/adip.26220).
- Stowe SD, Olson AL, Losick R, Cavanagh J. 2014.** Chemical shift assignments and secondary structure prediction of the master biofilm regulator, SinR, from Bacillus subtilis. *Biomolecular NMR Assignments* **8**:155–158 DOI [10.1007/s12104-013-9473-7](https://doi.org/10.1007/s12104-013-9473-7).
- Sun F, Zhang Y, Wang L, Yan X, Tan Y, Guo Z, Qiu J, Yang R, Xia P, Zhou D. 2012.** Molecular characterization of direct target genes and cis-acting consensus recognized by quorum-sensing regulator AphA in Vibrio parahaemolyticus. *PLOS ONE* **7**:e44210–e44221 DOI [10.1371/journal.pone.0044210](https://doi.org/10.1371/journal.pone.0044210).
- Sunadome K, Suzuki T, Usui M, Ashida Y, Nishida E. 2014.** Antagonism between the master regulators of differentiation ensures the discreteness and robustness of cell fates. *Molecular Cell* **54**:526–535 DOI [10.1016/j.molcel.2014.03.005](https://doi.org/10.1016/j.molcel.2014.03.005).

- Sushil KM, Malapaka K, Nitish RM. 2018.** Catestatin: a master regulator of cardiovascular functions. *Current Medicinal Chemistry* **25**:1352–1374
DOI [10.2174/0929867324666170425100416](https://doi.org/10.2174/0929867324666170425100416).
- Suto T, Karonitsch T. 2020.** The immunobiology of mTOR in autoimmunity. *Journal of Autoimmunity* **110**:102373–102381 DOI [10.1016/j.jaut.2019.102373](https://doi.org/10.1016/j.jaut.2019.102373).
- Suzuki T, Kusakabe M, Nakayama K, Nishida E. 2012.** The protein kinase MLTK regulates chondrogenesis by inducing the transcription factor Sox6. *Development* **139**:2988–2998 DOI [10.1242/dev.078675](https://doi.org/10.1242/dev.078675).
- Takahashi R-U, Takeshita F, Honma K, Ono M, Kato K, Ochiya T. 2013.** Ribophorin II regulates breast tumor initiation and metastasis through the functional suppression of GSK3 β . *Scientific Reports* **3**:2474–2486 DOI [10.1038/srep02474](https://doi.org/10.1038/srep02474).
- Tapia-Carrillo D, Tovar H, Velazquez-Caldelas TE, Hernandez-Lemus E. 2019.** Master regulators of signaling pathways: an application to the analysis of gene regulation in breast cancer. *Frontiers in Genetics* **10**:1180–1190 DOI [10.3389/fgene.2019.01180](https://doi.org/10.3389/fgene.2019.01180).
- Thornton AM, Shevach EM. 2019.** Helios: still behind the clouds. *Immunology* **158**:161–170 DOI [10.1111/imm.13115](https://doi.org/10.1111/imm.13115).
- Tijchon E, Havinga J, Van Leeuwen FN, Scheijen B. 2012.** B-lineage transcription factors and cooperating gene lesions required for leukemia development. *Leukemia* **27**:541–552 DOI [10.1038/leu.2012.293](https://doi.org/10.1038/leu.2012.293).
- Tiwari N, Tiwari Vijay K, Waldmeier L, Piotr J Balwierz, Arnold P, Pachkov M, Meyer-Schaller N, Schübeler D, Van Nimwegen E, Christofori G. 2013.** Sox4 is a master regulator of epithelial-mesenchymal transition by controlling ezh2 expression and epigenetic reprogramming. *Cancer Cell* **23**:768–783 DOI [10.1016/j.ccr.2013.04.020](https://doi.org/10.1016/j.ccr.2013.04.020).
- Tomljanovic Z, Patel M, Shin W, Califano A, Teich AF. 2018.** ZCCHC17 is a master regulator of synaptic gene expression in Alzheimer's disease. *Bioinformatics* **34**:367–371 DOI [10.1093/bioinformatics/btx608](https://doi.org/10.1093/bioinformatics/btx608).
- Torre C, Perret C, Colnot S. 2010.** Molecular determinants of liver zonation. *Progress in molecular biology and translational science* **97**:127–150
DOI [10.1016/B9780123852335.000052](https://doi.org/10.1016/B9780123852335.000052).
- Tovar H, García-Herrera R, Espinal-Enríquez J, Hernández-Lemus E. 2015.** Transcriptional master regulator analysis in breast cancer genetic networks. *Computational Biology and Chemistry* **59**:67–77 DOI [10.1016/j.compbiolchem.2015.08.007](https://doi.org/10.1016/j.compbiolchem.2015.08.007).
- Tamura T. 2017.** Regulation of mononuclear phagocyte development by IRF8. [*Rinsho Ketsueki*] *The Japanese journal of clinical hematology* **58**:798–805
DOI [10.11406/rinketsu.58.798](https://doi.org/10.11406/rinketsu.58.798).
- Van Bragt MPA, Hu X, Xie Y, Li Z. 2014.** RUNX1, a transcription factor mutated in breast cancer, controls the fate of ER-positive mammary luminal cells. *eLife* **3**:e03881–e03903 DOI [10.7554/eLife.03881](https://doi.org/10.7554/eLife.03881).
- Van Kessel JC, Rutherford ST, Shao Y, Utria AF, Bassler BL. 2013.** Individual and combined roles of the master regulators AphA and LuxR in control of the *Vibrio harveyi* quorum-sensing regulon. *Journal of Bacteriology* **195**:436–443
DOI [10.1128/JB.01998-12](https://doi.org/10.1128/JB.01998-12).

- Van Nes J, Chan A, Van Groningen T, Va Sluis P, Koster J, Versteeg R. 2013. A NOTCH3 transcriptional module induces cell motility in neuroblastoma. *Clinical Cancer Research* 19:3485–3494 DOI 10.1158/1078-0432.CCR-12-3021.
- Vargas DM, De Bastiani MA, Zimmer ER, Klamt F. 2018. Alzheimer's disease master regulators analysis: search for potential molecular targets and drug repositioning candidates. *Alzheimer's Research & Therapy* 10:59–70 DOI 10.1186/s13195-018-0394-7.
- Vivekanandan S, Moovarkumudalvan B, Lescar J, Kolatkar PR. 2015. Crystallization and X-ray diffraction analysis of the HMG domain of the chondrogenesis master regulator Sox9 in complex with a ChIP-Seq-identified DNA element. *Acta Crystallographica Section F, Structural Biology Communications* 71:1437–1441 DOI 10.1107/S2053230X1501969X.
- Vogel M, Velleuer E, Schmidt-Jiménez LF, Mayatepek E, Borkhardt A, Alawi M, Kutsche K, Kortüm F. 2016. Homozygous HOXB1 loss-of-function mutation in a large family with hereditary congenital facial paresis. *American Journal of Medical Genetics Part A* 170:1813–1819 DOI 10.1002/ajmg.a.37682.
- Voorhoeve PM. 2010. MicroRNAs: Oncogenes, tumor suppressors or master regulators of cancer heterogeneity? *Biochimica et Biophysica Acta* 1805:72–86 DOI 10.1016/j.bbcan.2009.09.003.
- Vrzalikova K, Woodman CBJ, Murray PG. 2012. BLIMP1 α . *Biomedical Papers of the Medical Faculty of the University Palacky, Olomouc, Czechoslovakia* 156:1–6 DOI 10.5507/bp.2012.003.
- Waldner MJ, Neurath MF. 2014. Master regulator of intestinal disease: IL-6 in chronic inflammation and cancer development. *Seminars in Immunology* 26:75–79 DOI 10.1016/j.smim.2013.12.003.
- Wang J, Yang B, Hu Y, Zheng Y, Zhou H, Wang Y, Ma Y, Mao K, Yang L, Lin G, Ji Y, Wu X, Sun B. 2013. Negative regulation of Nmi on virus-triggered type I IFN production by targeting IRF7. *The Journal of Immunology* 191:3393–3399 DOI 10.4049/jimmunol.1300740.
- Watanabe H, Takano K, Endo T. 2006. Rho family: master regulators of cytoskeleton and cell migration. *Tanpakushitsu Kakusan Koso Protein, Nucleic Acid, Enzyme* 51:683–692.
- Weeks KL1 GX, Du XJ, Boey EJ, Matsumoto A, Bernardo BC, Kiriazis H, Cemerlang N, Tan JW, Tham YK, Franke TF, Qian H, Bogoyevitch MA, Woodcock EA, Febbraio MA, Gregorevic P, McMullen JR. 2012a. Phosphoinositide 3-kinase p110 α is a master regulator of exercise-induced cardioprotection and PI3K gene therapy rescues cardiac dysfunction. *Circ Heart Fail* 5:523–534 DOI 10.1161/CIRCHEARTFAILURE.112.966622.
- Weeks KL, Gao X, Du X-J, Boey EJH, Matsumoto A, Bernardo BC, Kiriazis H, Cemerlang N, Tan JW, Tham YK, Franke TF, Qian H, Bogoyevitch MA, Woodcock EA, Febbraio MA, Gregorevic P, McMullen JR. 2012b. Phosphoinositide 3-kinase p110 α is a master regulator of exercise-induced cardioprotection and PI3K gene therapy rescues cardiac dysfunction. *Circulation Heart Failure* 5:523–534 DOI 10.1161/CIRCHEARTFAILURE.112.966622.

- Wehrspaun CC, Haerty W, Ponting CP. 2015. Microglia recapitulate a hematopoietic master regulator network in the aging human frontal cortex. *Neurobiology of Aging* 36:2443.e9–2443.e20 DOI 10.1016/j.neurobiolaging.2015.04.008.
- Wen Y, Ouyang Z, Devreese B, He W, Shao Y, Lu W, Zheng F. 2017. Crystal structure of master biofilm regulator CsgD regulatory domain reveals an atypical receiver domain. *Protein Science: a Publication of the Protein Society* 26:2073–2082 DOI 10.1002/pro.3245.
- Whyte WA, Orlando DA, Hnisz D, Abraham BJ, Lin CY, Kagey MH, Rahl PB, Lee TI, Young RA. 2013. Master transcription factors and mediator establish super-enhancers at key cell identity genes. *Cell* 153:307–319 DOI 10.1016/j.cell.2013.03.035.
- Wilkie H, Gossner A, Bishop S, Hopkins J. 2016. Variations in T Cell transcription factor sequence and expression associated with resistance to the sheep nematode *Teladorsagia circumcincta*. *PLOS ONE* 11:e0149644–e0149657 DOI 10.1371/journal.pone.0149644.
- Wilson SE, Esposito A. 2009. Focus on molecules: interleukin-1: a master regulator of the corneal response to injury. *Experimental Eye Research* 89:124–125 DOI 10.1016/j.exer.2009.02.011.
- Witczak CA, Sharoff CG, Goodyear LJ. 2008. AMP-activated protein kinase in skeletal muscle: from structure and localization to its role as a master regulator of cellular metabolism. *Cellular and Molecular Life Sciences* 65:3737–3755 DOI 10.1007/s00018-008-8244-6.
- Wolański M, Jakimowicz D. 2014. Fifty years after the replicon hypothesis: cell-specific master regulators as new players in chromosome replication control. *Journal of Bacteriology* 196:2901–2911 DOI 10.1128/JB.01706-14.
- Wu B, Wang J, Zhao Y, Guo W. 2015. Chapter 5 - Biochemical analysis of Rabin8, the guanine nucleotide exchange factor for Rab8.. In: Guo W, ed. *Methods in Cell Biology*. New York: Academic Press, 59–68.
- Wu W, Morrissey CS, Keller CA, Mishra T, Pimkin M, Blobel GA, Weiss MJ, Hardison RC. 2014. Dynamic shifts in occupancy by TAL1 are guided by GATA factors and drive large-scale reprogramming of gene expression during hematopoiesis. *Genome Research* 24:1945–1962 DOI 10.1101/gr.164830.113.
- Wysokinski D, Pawlowska E, Blasiak J. 2015. RUNX2: a master bone growth regulator that may be involved in the DNA damage response. *DNA and Cell Biology* 34:305–315 DOI 10.1089/dna.2014.2688.
- Xia P, Xu X-Y. 2015. PI3K/Akt/mTOR signaling pathway in cancer stem cells: from basic research to clinical application. *American Journal of Cancer Research* 5:1602–1609.
- Xiao W. 2015. The hypoxia signaling pathway and hypoxic adaptation in fishes. *Science China Life Sciences* 58:148–155 DOI 10.1007/s11427-015-4801-z.
- Xu Z, Ji G, Shen J, Wang X, Zhou J, Li L. 2012. SOX9 and myocardin counteract each other in regulating vascular smooth muscle cell differentiation. *Biochemical and Biophysical Research Communications* 422:285–290 DOI 10.1016/j.bbrc.2012.04.149.

- Yang H, Liang H, Yan J-S, Tao R, Hao S-G, Ma L-Y. 2012.** Down-regulation of hematopoiesis master regulator PU.1 via aberrant methylation in chronic myeloid leukemia. *International Journal of Hematology* **96**:65–73 DOI [10.1007/s12185-012-1106](https://doi.org/10.1007/s12185-012-1106).
- Yang Z, Rayala S, Nguyen D, Vadlamudi RK, Chen S, Kumar R. 2005.** Pak1 phosphorylation of snail, a master regulator of epithelial-to-mesenchyme transition, modulates snail's subcellular localization and functions. *Cancer Research* **65**:3179–3184 DOI [10.1158/0008-5472.CAN-04-3480](https://doi.org/10.1158/0008-5472.CAN-04-3480).
- Yu X, Ng CP, Habacher H, Roy S. 2008.** Foxj1 transcription factors are master regulators of the motile ciliogenic program. *Nature Genetics* **40**:1445–1453 DOI [10.1038/ng.263](https://doi.org/10.1038/ng.263).
- Zago G, Biondini M, Camonis J, Parrini MC. 2019.** A family affair: a Ral-exocyst-centered network links Ras, Rac, Rho signaling to control cell migration. *Small GTPases* **10**:323–330 DOI [10.1080/21541248.2017.1310649](https://doi.org/10.1080/21541248.2017.1310649).
- Zeitz JO, Kaltenböck S, Most E, Eder K. 2017.** Antioxidant status and expression of inflammatory genes in gut and liver of piglets fed different dietary methionine concentrations. *Journal of Animal Physiology and Animal Nutrition* **101**:1166–1174 DOI [10.1111/jpn.12633](https://doi.org/10.1111/jpn.12633).
- Zeng H. 2017.** mTOR signaling in immune cells and its implications for cancer immunotherapy. *Cancer Letters* **408**:182–189 DOI [10.1016/j.canlet.2017.08.038](https://doi.org/10.1016/j.canlet.2017.08.038).
- Zhang M-L, Nie F-Q, Sun M, Xia R, Xie M, Lu K-H, Li W. 2015.** HOXA5 indicates poor prognosis and suppresses cell proliferation by regulating p21 expression in non small cell lung cancer. *Tumor Biology* **36**:3521–3531 DOI [10.1007/s13277-014-2988-4](https://doi.org/10.1007/s13277-014-2988-4).
- Zhang S, Zhang T, Yan M, Ding J, Chen J. 2014.** Crystal structure of the WOPR-DNA complex and implications for Wor1 function in white-opaque switching of *Candida albicans*. *Cell Research* **24**:1108–1120 DOI [10.1038/cr.2014.102](https://doi.org/10.1038/cr.2014.102).
- Zhao C, Zeng C, Ye S, Dai X, He Q, Yang B, Zhu H. 2020.** Yes-associated protein (YAP) and transcriptional coactivator with a PDZ-binding motif (TAZ): a nexus between hypoxia and cancer. *Acta Pharmaceutica Sinica B* **10**:947–960 DOI [10.1016/j.apsb.2019.12.010](https://doi.org/10.1016/j.apsb.2019.12.010).
- Zhou F, Li F, Xie F, Zhang Z, Huang H, Zhang L. 2014.** TRAF4 mediates activation of TGF- β signaling and is a biomarker for oncogenesis in breast cancer. *Science China Life Sciences* **57**:1172–1176 DOI [10.1007/s11427-014-4727-x](https://doi.org/10.1007/s11427-014-4727-x).
- Zhou W, Sonnenberg GF. 2020.** Activation and suppression of group 3 innate lymphoid cells in the gut. *Trends in Immunology* **41**:721–733 DOI [10.1016/j.it.2020.06.009](https://doi.org/10.1016/j.it.2020.06.009).
- Zhu L, Sun G, Zhang H, Zhang Y, Chen X, Jiang X, Jiang X, Krauss S, Zhang J, Xiang Y, Zhang C-Y. 2009.** PGC-1 α is a key regulator of glucose-induced proliferation and migration in vascular smooth muscle cells. *PLOS ONE* **4**:e4182 DOI [10.1371/journal.pone.0004182](https://doi.org/10.1371/journal.pone.0004182).
- Zona S, Bella L, Burton MJ, Nestalde Moraes G, Lam EWF. 2014.** FOXM1: an emerging master regulator of DNA damage response and genotoxic agent resistance. *Biochimica et Biophysica Acta* **1839**:1316–1322 DOI [10.1016/j.bbagr.2014.09.016](https://doi.org/10.1016/j.bbagr.2014.09.016).