

A taxonomic review of the genus *Astrocladus* (Echinodermata, Ophiuroidea, Euryalida, Gorgonocephalidae) from Japanese coastal waters

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Abstract

Japanese species of the genus *Astrocladus* (Echinodermata, Ophiuroidea, Euryalida, Gorgonocephalidae) are reviewed. Three species, *A. annulatus* Matsumoto, *A. coniferus* Döderlein, and *A. exiguus* Lamarck, are currently known from Japanese waters. *Astrocladus coniferus* has two junior synonyms, *A. dofleini* Döderlein and *A. pardalis* Döderlein, however, status of these species has long been questioned. Their type specimens had not been examined in recent years and no molecular phylogenetic analyses have been performed.

Observations of the lectotype of *A. coniferus*, the lectotype and four paralectotypes of *A. dofleini* and the holotype of *A. pardalis* revealed that *A. coniferus* and *A. pardalis* were conspecific and morphologically distinguishable from *A. dofleini*. The interspecific difference between *A. coniferus* and *A. dofleini* was also supported by molecular phylogenetic analysis. Additionally, we re-describe *A. exiguus* and *A. annulatus*, based on recently collected specimens and the holotype.

Running title: A taxonomic review of Japanese *Astrocladus*

Key words: Taxonomy, Basket stars, *Astrocladus coniferus*, *Astrocladus dofleini*, *Astrocladus pardalis*, Japan

Introduction

The gorgonocephalid brittle stars of the genus *Astrocladus* Verrill, 1899 (Ophiuroidea: Euryalida)

are characterized by having cone-shaped external ossicles on the disc; a madreporite placed on inner edge of interradial lateral disc; no calcareous plates on the lateral disc margin; and no arm spines before first branch in adults (Fell, 1960; Baker, 1980). The genus is distributed from the Indo-Pacific to southern Africa (e.g. A. M. Clark & Courtman-Stock, 1976; Baker, 1980; McKnight, 2000). It was established by Verrill (1899) for *Euryale verrucosum* Lamarck, 1816 (= *Astrocladus exiguus* (Lamarck, 1816)) and it currently comprises 10 species: *Astrocladus africanus* Mortensen, 1933 from southern Africa; *A. annulatus* Matsumoto, 1915 from Japan; *A. coniferus* (Döderlein, 1902) from Japan; *A. euryale* (Retzius, 1783) from southern Africa; *A. exiguus* (Lamarck, 1816) from the Indo-West Pacific; *A. goodingi* Baker, Okanishi & Pawson, 2018 from the western Indian Ocean; *A. hirtus* Mortensen, 1933 from South Africa; *A. ludwigi* (Döderlein, 1896) from Indo-Western Pacific; *A. socotrana* Baker, Okanishi & Pawson, 2018 from northern Indian Ocean, and *A. tonganus* Döderlein, 1911 from Tonga and New Zealand (Retzius, 1783; Lamarck, 1816; Lyman 1875, 1882; Koehler, 1897, 1905, 1907, 1930; H. L. Clark, 1911, 1915, 1923; Döderlein, 1896, 1911, 1927; Matsumoto, 1915; Mortensen, 1933; Murakami, 1944a, 1944b; Djakonov, 1954; A. M. Clark, 1951, 1974; A. M. Clark & Rowe, 1971; A. M. Clark & Courtman-Stock, 1976; Irimura, 1969, 1981, 1982; Cherbonnier & Guille, 1978; Baker, 1980; McKnight, 1989; 2000; Liao & A. M. Clark, 1995; Rowe & Gates, 1995; Irimura & Tachikawa, 2002; Baker, Okanishi & Pawson, 2018). Of these, three species are currently known from Japanese waters: *A. annulatus*, *A. coniferus*, and *A. exiguus* (e.g. Döderlein, 1911; Matsumoto, 1915; Irimura, 1981).

Taxonomic works on *Astrocladus* are scarce in Japan: *A. annulatus* was described, based on the holotype, from central Japan (Matsumoto, 1915, 1917) and subsequently recorded based on a specimen from western Japan (Irimura, 1981); *A. exiguus* was recorded from central and western Japan, respectively (H. L. Clark, 1915; Irimura, 1981); and *Astrocladus coniferus* was recorded from Japanese waters except Hokkaido Island (Döderlein, 1902, 1910, 1911; H. L. Clark, 1911; Matsumoto, 1917; Murakami, 1944b; Irimura, 1979, 1981, 1982, 1990; Yi & Irimura, 1987; Fujita & Kohtsuka, 2003; Okanishi, Yamaguchi, Horii & Fujita, 2011; Kohtsuka, Sekifuji, Omori & Okanishi, 2017).

In these three species, the taxonomic status of *A. coniferus*, which is currently synonymized with *A. dofleini* and *A. pardalis*, has been controversial. *Astrocladus coniferus* and *A. pardalis* were both originally described by Döderlein (1902) in the genus *Astrophyton* and they were mainly distinguished by presence/absence of large conical tubercles on radial shield and dorsal proximal portion of arms (Döderlein, 1902). Subsequently, Döderlein (1910) moved the two species to *Astrocladus* Verrill, 1899 and synonymized *A. pardalis* with *A. coniferus*. In the same paper, Verrill described *Astrocladus dofleini*, which is distinguished from *A. coniferus* in having numerous granular large tubercles on the dorsal disc. Since then, although some taxonomists considered *A. coniferus* (with synonymy of *A. pardalis*) and *A. dofleini* to be separate taxa (Döderlein, 1910; Saba, Tomida & Kimoto, 1982; Irimura, 1990), others regarded the three taxa as distinct (sub)species (Matsumoto, 1917; Fedotov, 1926; Irimura, 1982; Shin & Rho, 1996) or united them as *A. coniferus* without any (sub)specific division (Murakami, 1944a, b; Fujita & Kohtsuka, 2003).

Since the series of descriptions by Döderlein (1902, 1910, 1911), the type specimens of *A. coniferus*, *A. dofleini* and *A. pardalis* had not been formally re-examined.

The other two Japanese species, *A. annulatus* and *A. exiguus*, has been distinguished from *A. coniferus* (and also from *A. dofleini* and *A. pardalis*) for they possess a covering of external ossicles on the dorsal body. However, no detailed description, including photographs of the body and/or SEM images of ossicles has been provided for *A. annulatus* and *A. exiguus*.

All of the previous studies of *Astrocladus* were based on morphological characters alone. Molecular data for *Astrocladus* has been published in only three papers for *Astrocladus coniferus*, *A. exiguus* and *A. hirtus* Mortensen, 1933 (Okanishi, O'Hata & Fujita, 2011; Okanishi & Fujita, 2013; O'Hara et al., 2017; Christodoulou, O'Hara, Hugall & Arbizu, 2019).

In this study, to address the taxonomic status of *Astrocladus coniferus*, we compared the morphology of the specimens including type specimens, of *A. coniferus*, *A. pardalis* and *A. dofleini*, and we obtained partial mitochondrial COI genes for 7 specimens which are morphologically consistent with type specimens of the three species. Partial mitochondrial COI of the two specimens of *A. exiguus* are also included in the phylogenetic analysis to estimate species delimitation by measuring genetic distance within *Astrocladus*. Additionally, we re-describe *A. annulatus* based on specimens, including type specimens, and we re-describe *A. exiguus* based on newly collected specimens.

Materials & Methods

Specimens examined

The 7 type specimens examined in this study are deposited in The University Museum, The University of Tokyo, Japan (UMUT), the Zoologische Staatssammlung München, Germany (ZSM), and Museum für Naturkunde der Humboldt-Universität zu Berlin, Germany (ZMB). Other newly collected specimens for molecular analysis are deposited in the National Museum of Nature and Science, Tsukuba, Japan (NSMT) (Table 1).

One lectotype of *Astrophyton coniferum* (ZSM 453/1), one lectotype (one of four specimens of ZSM 440/1), four paralectotypes (three specimens of ZSM 440/1 and one specimens of ZMB 5923) of *Astrocladus dofleini* (ZSM 440/1 and), the holotype of *Astrocladus pardalis* (ZSM 453/2) and the holotype of *Astrocladus annulatus* (UMUTZ-Echi-Oph-26) are preserved in ethanol, but their fixation methods are unknown.

The specimens for molecular analysis were collected by commercial fishing lines of at Minabe Port, Wakayama Prefecture (2 specimens), by commercial dredge at Mogi, Nagasaki Prefecture (4 specimens), and by scuba diving near Misaki Marine Biological Station, Kanagawa Prefecture (3 specimens). The specimens were fixed and preserved in 70–99% ethanol (Fig. 1).

Morphological observation and terminology

Ossicles from *Astrocladus coniferus* (MO 2018-118A), *A. dofleini* (MO 2018-118B) and *A. exiguus* (MO-2019-19) were isolated by immersion in domestic bleach (approximately 5% sodium hypochlorite solution), washed in deionized water, air-dried, and mounted on SEM stubs using double-sided conductive tape. The ossicles were observed and photographed with a Jeol JSM 5200LV SEM at Misaki Marine Biological Station, The University of Tokyo. Photographs (Fig. 13) were focus-stacked using the software CombineZM v. 1.0.0 (<https://www.softpedia.com/get/Multimedia/Graphic/Graphic-Editors/CombineZM.shtml>). The size of external ossicles, represented by the length of the longest axis, it is referred to as “length” in this study.

The morphological terms used to describe euryalid brittle stars follow Thuy & Stöhr, (2011), Stöhr, O'Hara & Thuy, (2012), Okanishi (2016) and Hendler (2018). The granular external ossicle is referred to as a “granule” in this paper. Taxonomic arrangement follows Christodoulou, O'Hara, Hugall & Arbizu (2019).

DNA extraction, amplification and sequencing

Genomic DNA was extracted using DNeasy Blood and Tissue extraction kit (Qiagen) according to the manufacturer's protocol. We sequenced mitochondrial COI gene for phylogenetic analysis. The method of DNA extraction and PCR parameters followed Okanishi & Fujita (2013). A primer set of COI005 (5'-TTAGGTAAHWAAACCAVYTKCCTTCAAAG-3') and COI008 (5'-CCDTANGMDATCATDGCRTACATCATTC-3') (Okanishi & Fujita,2013) was used for PCR of COI. The optimum cycling parameters for those COI primers consisted of an initial denaturation step of 94°C/2 min followed by 41 cycles of 94°C/30 s,48°C/90 s and 72°C/60 s with final extension step at 72°C/10 min was followed by storage at 4°C. The PCR products were separated from excess primers and oligonucleotides using Exo-SAP-IT (GEHealthcare) by following manufacturer's protocol. All samples were sequenced bidirectionally and sequence products were run on the 3730xI DNA Analyzer of Misaki Marine Biological Station. The accession numbers of the DNA Data Bank of Japan (DDBJ) are shown in Table 1.

Phylogenetic analysis

We newly sequenced 2 specimens of *Astrocladus coniferus*, 5 specimens of *Astrocladus dofleini* and 2 specimens of *A. exiguus*, and the sequence data of single specimens of *A. dofleini* (referred as *A. coniferus* in Okanishi & Fujita, [2013, 2018]) and *A. exiguus* obtained by Okanishi & Fujita (2013; 2018) were also used. *A. exiguus* was added to the analysis to compare genetic distances and determine the taxonomic rank of each phylogenetic group within *Astrocladus*. For outgroups, we selected 6 species of the subfamily Gorgonocephalinae with the shortest genetic distance from *Astrocladus* to avoid long branch attraction (Bergsten, 2005; Okanishi & Fujita, 2018). These species were also used in previous molecular

phylogeny (Okanishi & Fujita, 2013, 2018).

All sequences were aligned using the Clustal W algorithm in MEGA X (Thompson, Higgins & Gibson, 1994; Kumar et al. 2018). All missing sequences were scored as gaps. Overall average of substitution rate was computed using MEGA X according to the Kimura 2-parameter model (Kimura, 1980) to compare the evolutionary rate of each gene. The K2P, HKY and TN93 with gamma distributions were selected as best-fit models of the first, second and third codons, respectively (Kimura, 1980; Hasegawa M, Kishino H, Yano T. 1985; Tamura & Nei, 1993), by using the “Find best fit models” option of MEGA X. Seaview ver. 4.3.0 (Gouy, Guindon & Gascuel, 2010) were used in preparing the data matrices in PHYLIP format.

Figtree v1.4.3 (<http://tree.bio.ed.ac.uk/software/figtree/>) was used in exploring tree files, in preparing NEWICK format and exploring alternative tree topologies. The phylogenetic tree was constructed with RAxML ver. 8.1.20 (Stamatakis, 2014) for maximum likelihood analysis (ML) to obtain bootstrap support values and with MrBayes v. 3.1.2 (Ronquist & Huelsenbeck, 2003) to obtain Bayesian posterior probabilities (BPP). The Markov-Chain Monte-Carlo (MCMC) process was run with four chains for 3,000,000 generations, with trees being sampled every 100 generations. The first 7,500 trees were discarded as burn-in. Data sets were partitioned by each codon for the maximum likelihood analysis to allow for separate optimization per-site substitution rates. The best-supported likelihood tree was found by performing 1000 replications.

Genetic distance

K2P genetic distances were computed within each clade and between clades using MEGA X according to the K2P model to compare the evolutionary rate of COI gene. Standard error of each clade was found by performing 100 bootstrap replications.

Results & Discussion

Taxonomy

Order Euryalida Gray, 1840

Family Gorgonocephalidae Ljungman, 1867

Genus *Astrocladus* Verrill, 1899

Type species: *Euryale verrucosum* Lamarck, 1816 (= *Astrocladus exiguus* (Lamarck, 1816))

Diagnosis

Arms five, branching. Number of arm segments less than six before first branch. No rows of calcareous plates on edge of disc margin. One madreporite situated on innermost portion of interrational

lateral disc. Arm spines present before the first branch. Hooklets on dorsal arms with one secondary tooth. Disc covered by variously shaped external ossicles and/or large tubercles (Döderlein, 1927; Baker, 1980; McKnight, 2000).

Remarks

In this study, our molecular and morphological examination confirmed that *Astrocladus pardalis* (Döderlein, 1902) is a junior synonym of *A. coniferus* (Döderlein, 1902) which can be separated from *A. dofleini* (Döderlein, 1910). Therefore, 11 species are now known in this genus (see list of “Included species” below); *A. annulatus*, *A. coniferus*, *A. dofleini* and *A. exiguus* are distributed in Japan.

Included species

A. africanus Mortensen, 1933; *A. annulatus* (Matsumoto, 1912); *A. coniferus* (Döderlein, 1902); *A. dofleini* Döderlein, 1910; *A. euryale* (Retzius, 1783); *A. exiguus* (Lamarck, 1816); *A. goodingi* Baker, Okanishi & Pawson, 2018; *A. hirtus*, Mortensen, 1933; *A. ludwigi* (Döderlein, 1896); *A. socotrana* Baker, Okanishi & Pawson, 2018; *A. tonganus* Döderlein, 1911.

Astrocladus coniferus (Döderlein, 1902) (Figs. 2–7)

Astrophyton coniferum Döderlein, 1902, 325, 326; Jangoux, de Ridder, Fechter, 1987, 306.

Astrocladus coniferus.—Döderlein, 1911, 46–49, Taf. 2, fig. 7, 7a; Taf. 4, Figs. 1–3a; Taf. 7, 5, 6a, 16; 1912; Clark, H.L., 1915, 186; Fedotov, 1926, 473–477; Murakami, 1944a, 247–248; 1944b, 262; Djakonov, 1949, 50; 1954, 20; Irimura, 1968, 32; 1969, 39; 1981, 18–19; Liao & Clark A.M., 1995, 170; Ishida et al., 2001, 8; Fujita, Ishida, Kato & Irimura, 2004, 192, 193; Okanishi, Yamaguchi, Horii & Fujita, 2011, 378, 379, Fig. 6G–J.

Astrocladus coniferus coniferus.—Irimura, 1982, 9–11, Fig. 5, Pl. 1(3); Rho & Shin, 1987, 209, 211; Shin, 1992, 118, 121; Shin & Rho, 1996, 389; Ishida et al., 2001, 8.

Astrocladus coniferus pardalis.—Saba, Tomida & Kimoto., 1982, 27. Pl. 14 (2, 3); Shin & Rho, 1996, 390; Ishida et al., 2001, 8.

Astrophyton cornutum.—Clark, H.L., 1911: 293.

Astrophyton pardalis Döderlein, 1902, 323; Clark, H.L., 1911, 293–294; Jangoux, de Ridder, Fechter, 1987, 308.

Astrocladus coniferus var. *pardalis*.—Matsumoto, 1917, 77; Fedotov, 1926, 473–477; Djakonov, 1954, 20. (Non) *Astrocladus coniferus coniferus*.—Saba, Tomida & Kimoto., 1982, 26–27, Pl. 14(1) (= *Astrocladus dofleini*)

(Non) *Astrocladus coniferus*.—Matsumoto, 1917, 77–79, Fig. 23c; Irimura, 1990, 75, an unnamed Pl.; Fujita & Kohestuka, 2003, 27, 28, Pl. 1B (= *Astrocladus dofleini*)

(Non) *Astrocladus coniferus* var. *pardalis*.—Irimura, 1982, 12–13, Fig. 7, Pl. 4(4) (= *Astrocladus dofleini*)

Notes on lectotype

In the original description (Döderlein, 1902), *Astrophyton coniferum* (= *Astrocladus coniferus*) was implied to be described based on two specimens which are listed in a table (Döderlein, 1902, P326). They were collected in Kagoshima Bay at, ca. 30 m depth and subsequently, one of them was figured by the same author in 1911 as “Typus” of *Astrocladus coniferus* (Döderlein, 1911, Taf 4, 2-2a). This is in accordance with §75.4 of the International Code of Zoological Nomenclature, and it can be regarded as the lectotype. The morphological traits of ZSM 453/1 concur with this figured specimen. Therefore, ZSM 453/1 is the lectotype.

Type material examined

ZSM 453/1, the lectotype of *Astrophyton coniferum* (Döderlein, 1902) (*Astrocladus coniferus*), Kagoshima Bay, ca 30 m, Japan, 1880 August (Fig. 2A, B). ZSM 453/2, the holotype of *Astrophyton pardalis* (Döderlein, 1902), collected by Karl Haberer, Sagami Bay, Japan, data unknown (Fig. 4B). In the original description (Döderlein, 1902), *Astrophyton pardalis* was implied to be described based on the single specimen, listed in a table (Döderlein, 1902, P326). Therefore, this specimen (ZSM 453/2) is the holotype by monotypy (ICZN Article 73.1.2.; see also Jangoux, de Ridder & Fechter, 1987)

Other material examined

MO 2018-118A, Moroiso, Kanagawa, Japan, 1.5 m, 26 April 2018, collected by Hisanori Kohtsuka, scuba. MO-2019-9, Hashiraguri, Oki Island, Shimane, Japan, 20 m, 15 July 2010, collected by Hisanori Kohtsuka, scuba.

External morphology of the lectotype (ZSM 453/1)

Disc. Disc five-lobed with notched interradiar edges, 60 mm in diameter (Fig. 2C). Dorsal disc wholly covered by external ossicles in contact with each other. Radial shields completely covered by granules and conical external ossicles, approximately 110–450 µm in length; other areas covered by smaller granules, approximately 100 µm in length (Fig. 2F). Radial shields tumid, bar-like, approximately 50 mm in length, width gradually decreasing from 10 mm at disc periphery to 2.5 mm almost at disc center (Fig. 2E). One large conical tubercle on peripheral edge of each radial shield, 2.5–3.4 mm in length (Fig. 2E).

Ventral surface of disc completely covered by skin and polygonal plate-like external ossicles, fully in contact, approximately 600 µm in length (Fig. 2G). Oral shields, adoral shields, oral plates and ventral arm plates completely concealed by ossicles (Fig. 2G). Teeth uniformly spiniform, situated on top

of dental plates (Fig. 2G). Teeth arranged in a cluster covering ventral-most part of dental plate approximately 10 in number (Fig. 2G), and in a vertical line, on other part of dental plates, 3 or 4 in number. Spiniform oral papillae situated in 1 or 2 transverse rows on ventral edge of each oral plate, 4 to 5 in number (Fig. 2G). Size of teeth varying in position on jaw, approximately 400–1000 μm in length and oral papillae approximately 400 μm in length (Fig. 2G).

Interradial surface of lateral disc covered by granules fully in contact, approximately 100 μm in length (Fig. 2G, H). Two genital slits (5 mm long and 1–3 mm wide) in each interradius (Fig. 2G). One large, elliptical madreporite situated on ventral interradius, approximately 5.5 mm in width and 3.75 mm in length (Fig. 2H).

Arms. Arms branching. On proximal portion before first branch, arm 8.5 mm wide with arched dorsal surface and flattened ventral surface (Fig. 3A). Between first branch and second branch, arm width abruptly decreases to 5.8 mm. Subsequently, arms tapering gradually toward arm tip (Fig. 3).

On dorsal and lateral surface, each arm segment covered by single annular row of large hooklet-bearing plates (Fig. 3A, B). Before second branch, each plate separated by granules. Plates fully in contact from third branch and subsequent distal segments (Fig. 3B). With exception of hooklet-bearing plates, dorsal and lateral surface of arm completely covered by granules, fully in contact, approximately 200–400 μm in length (Fig. 3A). Before first branch, ventral surface covered by polygonal plate-like external ossicles, similar to those on ventral disc, approximately 100 μm in length (Fig. 3C). After first branch, ossicles become into round granules, slightly in contact, and decreasing in size gradually toward arm tip (Fig. 3D). Lateral arm plates and ventral arm plates completely concealed by external ossicles on entire arm (Fig. 3C, D). Tentacle pore with single arm spine before first branch; 2 or 3 spines after first branch; up to 4 spines on subsequent pores (Fig. 3C, D). Number of arm spines decrease gradually to 2 towards arm tip. Arm spines approximately one-seventh to one-eighth (ca. 12–13%) of length of the corresponding arm segment on proximal portion of arm, and one-thirds to one-fourth length of the corresponding arm segment on middle to distal region of arm (Fig. 3C, D).

Color. Uniformly dull brown in ethanol preserved specimen (Fig. 2C, D).

Description of other materials

External morphology of ZSM 453/2, the holotype of Astrophyton pardalis: Disc approximately 30 mm in diameter (Fig. 4A, C). The external ossicles on radial shield conical with acute tip (Fig. 4D). No large tubercle on periphery of radial shields (Fig. 4C), tubercle present in lectotype of *Astrophyton coniferum* (ZSM 453/1). Teeth and oral papillae not spiniform, but granular (Fig. 4E).

Living color of MO-2018-118A, d.d. = 30 cm: Dorsal disc vivid orange with yellow patches, arms with yellow transverse bands on dorsal side (Fig. 4G). Ventral side of arms and disc uniformly creamy white, with orange arm tip (Fig. 4H).

Ossicle morphology of MO-2018-118A: All arm hooklets with single inner tooth and reticular

structures (Fig. 5A–C). Inner tooth on distal portion of arm smaller and more rudimentary than those from proximal to middle portion of arm. Hooklet-bearing plates with 9 or 10 tubercle-shaped articulations for hooklets in proximal portion of arm; articulations forming 2 parallel rows (Fig. 5D). On proximal to middle portion of arm, lateral arm plates longer than high, curved to distal side (Figs. 5E, F, I, J). On proximal portion of arm, simple muscle openings besides border structures on distal edge (Fig. 5F), and on middle portion of arm, nerve openings on internal side of the muscle openings (Fig. 5I). One perforation present on internal side (Fig. 5J). On distal portion of arms, plate square, at least one nerve opening beside border structure and one articulation for hooklet on distal side (Fig. 6A). No perforations recognizable on internal side (Fig. 6B). On proximal and middle portion of arm, arm spines ovoid, with three or four terminal projections, approximately one-third to one-fourth length of the height of arm spine (Fig. 5G, H, K, L). In distal portion, arm spines transformed into hooks with one inner secondary tooth (Fig. 6C). Hook-shaped arm spines distinguished from hooklets by lack of reticular structure (Figs. 5A–C; 6C).

All vertebrae with hourglass-shaped streptospondylous articulations (Figs. 6D, F, G; 7C, D). In middle portion of arm, surfaces of lateral furrows smooth, with tubercle shaped ornamentations on dorsal side (Fig. 7B). In distal portion of arm, depressions for tube feet openings in distal part of ventral-lateral side of vertebrae but these are unrecognizable on proximal to middle portion of arm (Figs. 7E). A pair of channels for passage of lateral nerves opening inside ventral furrows (Figs. 6E; 7A, E). In middle portion of arm channels for passage of lateral canals also opening on distal side of canals of lateral nerves (Fig. 7A). Channels for lateral nerve obscured in distal portion of arm (Fig. 7E).

Distribution

Many records of *Astrocladus coniferus* have been confused with those of *A. dofleini*. Therefore, only those that can be identified as *A. coniferus* by their figures and/or photographs are listed here. JAPAN: Kagoshima Bay, southwestern Japan, ca. 20 m (Döderlein, 1902, 1911; type locality, Fig. 1), Sagami Bay, Kanagawa, central-eastern Japan, 1.5–130m (Döderlein, 1902, 1911; Irimura, 1982; This study, Fig. 1), Off Hachijojima Island and Ogasawara Islands, south-eastern Japan, 120–500 m (Okanishi, Yamaguchi, Horii & Fujita, 2011). Off Kii Nagashima, Mie, central Japan, depth unknown (Saba, Tomida & Kimoto., 1982); Hashiraguri, Oki Island, Shimane, western Japan, ca. 20 m (This study).

Remarks

In this study, we propose that *A. pardalis* is a junior subjective synonym of *A. coniferus* (see Remarks of *A. dofleini* for the details). *A. coniferus* can be distinguished from other congeners in having; granules and conical external ossicles on dorsal disc, and 0 or 1 conical large tubercles on the peripheral edge of each radial shield.

Astrocladus dofleini Döderlein, 1910

(Figs. 8–12)

Astrocladus dofleini Döderlein, 1910, 256; 1911, 41–46, 106, Fig. 9, Taf. 2, Fig. 6, Taf. 3 Figs. 1–4, Taf. 4, Figs. 4, 5, Taf. 7, 15, 15b; 1927, 35, 94, Taf. 3, Fig. 2.; Guille, 1981, 416, 417; Jangoux, de Ridder, Fechter, 1987, 306.

Astrocladus coniferus dofleini.—Rho & Shin, 1987, 209, 211; Yi & Irimura, 1987, 122, 123, fig. 2; Shin, 1992, 253; 1995, 117, 121; Shin & Rho, 1996, 391; Ishida et al., 2001, 8.

Astrocladus coniferus var. *dofleini*.—Matsumoto, 1917, 77–79, Fig. 23; Fedotov, 1926, 473–477; Djakonov, 1949, 50; 1954, 20; Irimura, 1968, 32; 1979, 2; 1981, 19; 1982: 11, 12, Fig. 6, Pl. 4, Figs. 5, 6.

Astrocladus coniferus.—Matsumoto, 1917, 77–79, Fig. 23; Irimura, 1990, 75, Pl; Fujita & Kohtsuka, 2003, 27, Pl. 1B; Kohtsuka, Sekifuji, Omori & Okanishi, 2017., 2017, 229–233, Figs. 2–5 (Non *Astrocladus coniferus*).

Astrocladus coniferus coniferus.—Saba, Tomida & Kimoto., 1982, 26–27, Pl. 14(1) (Non *Astrocladus coniferus*)

Astrocladus coniferus var. *pardalis*.— Irimura, 1982: 12, 13, Text-fig. 7, Pl. 4, Fig. 4 (Non *Astrocladus pardalis*).

(Non) *Astrocladus dofleini*.—Bomford, 1913, 220, 221, Pl. 13, Fig. 1 (= *Astrocladus exiguus*).

Notes on lectotype

In the original description (Döderlein, 1910), this species was based on a specimen in the Peabody Museum of Natural History (Yale University) which was reported by Verrill (1899), plus several specimens collected from Japan. Subsequently, one of them was figured by Döderlein (1911), and names the “Typus” of *Astrocladus coniferus*. In accordance with §75.4 of the International Code of Zoological Nomenclature, the author “unambiguously selected a particular syntype to act as the unique name-bearing type of the taxon”, namely the lectotype. The morphological traits of one of the 4 specimens of ZSM 440/1 concur with the specimen figured by Döderlein (1911). Therefore, the specimen of ZSM 440/1 is designated as the lectotype, and the other three specimens as paralectotypes.

Type material

ZSM 440/1, the lectotype and three paralectotypes of *Astrocladus dofleini* Döderlein, 1902, Okinose, Sagami Bay, ca 600 m, Japan, 1904–1905, collected by Franz Doflein (Fig. 8B). ZMB 5923, a paralectotype of *Astrocladus coniferus* Döderlein, 1902, Sagami Bay, depth unknown, Japan, collected by Karl Haberer.

Other material examined

MO 2019-15 to 18, 4 specimens, Mogi, Nagasaki, Japan, depth unknown, 7 February 2018, collected by Hatsuyuki Takeshita. NSMT E-5480 off Minabe, 1 specimen, Wakayama, Japan, ca. 80 m, 10 March 2006, collected by Hajime Watabe, gill net. NSMT E-10749, 1 specimen, Kuji Port, Hitachi, Ibaraki, Japan, 36 30.50N, 140.38.40.E, depth unknown, 30 September, 2016, collected by fishery boat *Daisan-shouei-Maru*, fishing net.

Description of external morphology of the lectotype (ZSM 440/1)

Disc. Disc circular with slightly notched interrarial edges, 53 mm in diameter (Fig. 8C). Radial shields tumid (Fig. 8C). Dorsal disc wholly covered by granules in contact each other and domed large tubercles (Fig. 8C). Radial shields covered by granules, approximately 200–330 µm in length (Fig. 8C). and large domed tubercles, 20–25 on each radial shield in number, each approximately 2.5–3.4 mm in length (Fig. 8C). Radial shields bar-like, approximately 25 mm in length, and the width gradually decreasing from 4.6 mm at disc periphery to 1.5 mm almost reaching to disc center (Fig. 8C).

Ventral surface of disc covered by polygonal plate-shaped external ossicles, fully in contact. Oral shields, adoral shields, oral plates and ventral arm plates concealed by ossicles (Fig. 8E). Teeth uniformly small, granule-like, situated on the top of dental plates, forming a cluster, approximately 6 to 8 in number (Fig. 8E). Oral papillae on the ventral edges of oral plates, forming a transverse row on each plate, 1 or 2 in number (Fig. 8E). Interrarial surface of lateral disc covered by granules fully in contact, approximately 170 µm in length, and domed tubercles, approximately 200–500µm in length (Fig. 8E). Two pore-like genital slits (6.5 mm long, 3 mm wide) in each interradius (Fig. 8E).

Arms. Arms branching. On proximal portion before first branch, arm 16 mm wide with an arched dorsal surface and flattened ventral surface (Fig. 8D, E). Between first branch and second branch, arm width abruptly decreasing to 10 mm. Subsequently, arms tapering gradually toward arm tip (Fig. 8F, H).

On dorsal and lateral surface, each arm segment covered by single annular row of hooklet-bearing plates. Before third branch, each plate separated by granules. The plates fully in contact from fourth branch and subsequent distal segments. With exception of hooklet-bearing plates, dorsal and lateral surface of arm completely covered by granules, fully in contact, approximately 100 µm in length. Before the first branch, ventral surface covered by polygonal and plate-shaped external ossicles, similar to those on ventral disc. After the first branch, the ossicles transforming granules, slightly in contact, and decreasing in size gradually toward the distal arm tip. Lateral arm plates and ventral arm plates concealed by external ossicles on entire arm (Fig. 8E, G, H). Tentacle pore with single arm spine after the first branch; 1 or 2 spines after second branch; and up to 4 spines for the subsequent pores (Fig. 8G, H). Distally, the number of arm spines decreasing gradually to 2 toward arm tip. Arm spines approximately one-fourth to one-fifth of length (ca. 20–25%) of corresponding arm segment on proximal portion of arm, and one-thirds to one-fourth length of corresponding arm segment on middle to distal arm segment (Fig. 8G, H).

Color. Uniformly dull brown with whitish large tubercles in ethanol preserved specimen (Fig. 8).

Description of other materials

Ossicle morphology of MO-2018-118B: Each hooklet on proximal and middle portion of arm with single inner secondary tooth, distal portion of arm with two inner secondary teeth. All hooklets with reticular structures (Figs. 9A, B, D). Hooklet-bearing plates with 6, 8 and 6 tubercle-shaped articulations for hooklets in proximal, middle and distal portion of the arm, respectively; articulations forming 2 parallel rows (Figs. 9C, E; 10B).

On proximal portion of arm, lateral arm plates long, with straight both proximal and distal edges (Fig. 9G, H); edges ellipse-shape in middle (Fig. 9J, K) and distal (Fig. 10B, C) portion of arms. No perforation observed on ventral side (Figs. 9H, K; 10C). Pairs of simple nerve and muscle openings with border structures on external edge on proximal and middle portion of arm (Fig. 9G, J), but only muscle opening present on distal portion of the arm (Fig. 10B). On middle to distal portion of arms, lateral arm plates carrying 5 or 6 articulations for hooklets on external edge (Fig. 9J; 10B). On proximal to middle portion of arm, arm spines ovoid, with three projections, approximately one-thirds to one-fifth to one-sixth length of the height of spine (Fig. 9F). In distal portion, arm spines transformed into hooks with two inner secondary teeth (Fig. 10A). Hook-shaped arm spines distinguished from hooklets on dorsal and lateral surface of arm by lack of reticular structure (Figs. 9A, B, D; 10A).

All vertebrae with hourglass-shaped streptospondylous articulations (Figs. 10D, E; 11A, 12F, G), and distal side of branching vertebra slightly wider than non-branching vertebra due to their possession of 2 articulation surfaces (Figs. 10H; 11C; 12C). Lateral furrows of vertebrae ornamented by tubercles in proximal to middle portion of arm, but smooth in distal portion of arm (Fig. 10F; 11D; 12B). Depressions for tube feet openings in distal part of ventral-lateral side of vertebrae (Figs. 10G, H; 11C; 12A). In proximal to middle arms, a pair of the channels for passages of lateral canals opening inside of ventral furrow, near depression of tube feet entire arms, and distal side of the channels, a pair of the channels for passage of lateral nerves opening (Figs. 10G, H; 11C). They are unrecognizable at distal portion of the arm (Figs. 10G, H; 11C; 12A, C).

Distribution

Unquestionable records of *Astrocladus dofleini* are: PHILIPPINES: Cabugan Grande Island, central Philippine, 135 m (Döderlein, 1927). JAPAN: Sagami Bay and Tokyo Bay, central-eastern Japan, 2–600 m (Döderlein, 1911; Irimura, 1982; Kohtsuka, Sekifuji, Omori & Okanishi, 2017; this study, type locality, Fig. 1); Toyama Bay, central Japan, 40–80 m (Fujita and Kohtsuka, 2003); Tachibana Bay, Nagasaki, western Japan, ca. 40 m (This study, Fig. 1); off Minabe, Shirahama, Wakayama, central Japan, depth unknown (This study, Fig. 1). KOREA: Huksando, southwestern Korea, depth unknown (Yi & Irimura, 1987).

Remarks

Döderlein (1902) described *A. coniferus* and *A. pardalis* on the basis of presence/absence of a large conical tubercle on the distal end of each radial shield. Subsequently, he determined that the presence/absence was an intra-specific character and the synonymized *A. pardalis* with *A. coniferus* (Döderlein, 1911).

In our study, although examinations of the lectotype of *A. coniferus* (ZSM 453/1) and the holotype of *A. pardalis* (ZSM 453/2) confirmed these morphological differences between the two specimens (Figs. 2E and 4C), monophyly of two additional specimens which morphologically agree with the lectotype of *A. coniferus* (MO-2018-118A) and the holotype of *A. pardalis* (MO-2019-9) was confirmed by our molecular phylogeny (see also “Molecular phylogeny” below). Thus, we follow Döderlein’s decision to synonymize these two species.

Döderlein (1911) also distinguished *Astrocladus coniferus* and *A. dofleini* as follow: *A. coniferus* possesses conical external ossicles but lacks large tubercles on dorsal surface of proximal portion of arms, whereas *A. dofleini* possesses only granules and large tubercles on the same position of the arms.

Matsumoto (1917) made the two species conspecific based on the existence of specimens showing intermediate features (Matsumoto, 1917) and Fujita & Kohtsuka (2003) followed this classification. Irimura (1982) distinguished *A. coniferus* (as “*A. coniferus coniferus*”) and *A. dofleini* (as “*A. coniferus* var. *dofleini*”) based on presence/absence of large tubercles on dorsal surface of proximal arms. However, Irimura (1982) did not recognize any morphological features between the “*A. coniferus* var. *dofleini*” and *A. pardalis* (as “*A. coniferus* var. *pardalis*”) except color differences.

In addition to the types of *A. coniferus* and *A. pardalis*, we also studied four paralectotypes (three specimens of ZSM 440/1 and ZMB 5923) of *A. dofleini* and confirmed that the Döderlein’s diagnostic characters can not distinguish these species, because:

Ossicles on dorsal surface of proximal portion of arms were granular and conical in *A. coniferus* (Figs. 3A; 4F) and granular in *A. dofleini* (Fig. 8D). Both *A. coniferus* (Fig. 3A) and *A. dofleini* (Fig. 8D) possess large tubercles on dorsal surface of proximal arms.

Instead, *A. coniferus* can be distinguished from *A. dofleini* by the following three morphological characters:

1) *Shape of ossicles*: Ossicles on periphery of radial shields were conical in *A. coniferus* (Figs. 2F; 4D) but those of *A. dofleini* (Fig. 8C) were granules.

2) *Shapes of large tubercles*: Large tubercles on dorsal disc were conical in *A. coniferus* (Fig. 2F), whereas those of *A. dofleini* were all domed (Fig. 8C).

3) *Distribution of the large tubercles*: Although the large tubercles were only on the peripheral edges of radial shields (Fig. 2F) or absent (Fig. 4C) in *A. coniferus*, those of *A. dofleini* were scattered on the dorsal surface of the disc (Fig. 8C).

These differences were also recognized in other examined materials: 2 specimens of *A. coniferus* (MO-2018-118A and MO-2019-9); and 6 specimens of *A. dofleini*. Therefore, we conclude that *A. coniferus* and *A. pardalis* are conspecific and distinct from *A. dofleini*. Our molecular phylogenetic analysis also supports this conclusion (see “Molecular phylogeny” below).

Additionally, color may also differ in these specimens. On the dorsal side, the two examined specimens of *A. coniferus* are vivid orange with yellow patches and arm bands, and the seven NSMT specimens of *A. coniferus* are uniformly black with small black patches and arm bands, or yellow with light yellowish small patches and arm bands (Fig. 7). However, we refrain from employing these color variations as diagnostic characters because other color patterns for these species have been recorded (e.g. Irimura, 1982).

Astrocladus exiguus (Lamarck, 1816) (Figs. 13–16)

Euryale exiguum Lamarck, 1816, 539.

Astrophyton exiguum.—Müller & Troschel, 1842, 125; Lyman, 1875, Pl. 4, Fig. 48; 1882, 257, Pl. 47, Fig. 1.

Astrocladus exiguus.—Doderlein, 1911, 76, 77, 106, 107, Pl. 9, Fig. 6; 1927, 34, 93, Pl. 5 Fig. 9; Clark H.L., 1915, 187; Koehler, 1931, 34, 35, Pl. 4, Figs. 1, 2; Chang, Liao, We, 1962, 59, 60, Pl. 1, Figs. 1, 2; Clark, A.M. & Rowe, 1971, 78, 79, 92, Fig. 21; Cherbonnier & Guille, 1978, 11, 12, Pl. 2, Figs. 1, 2; Baker, 1980, 63, Figs. 28, 33; Irimura, 1981, 19; Liao & Clark, A.M., 1995, 169, 170, Fig. 73, Pl. 19, Fig. 1; Rowe & Gates, 1995, 365; Baker, Okanishi & Pawson, 2018, 9, 10.

Gorgonocephalus cornutus Koehler, 1897, 368, 369, Pl. 9, Figs. 80, 81; 1899, 73, 74, Pl. 12, Figs. 95, 96, Pl. 13, Fig. 98.

Astrophyton cornutum.—Koehler, 1905, 127–129, Pl. 13, Fig. 1, pl. 18, Fig. 2.

Astrocladus dofleini.—Bomford, 1913, 220, 221, Pl. 13, Fig. 1 (Non *Astrocladus dofleini*).

Material examined

MO 2019-11, 1 specimen, off Minabe, Wakayama, Japan, depth unknown, 11 November 2012, gill net. MO 2019-19, 1 specimen, off Minabe, Wakayama, Japan, depth unknown, 4 April 2019, gill net, collected by Sadao Inoue. NSMT E-6265, off Yaku-shima Island, Kagoshima, Japan, 29°47.00'N., 130°22.06'E. 155–170 m, 2 August 2008, 1 m biological dredge, R/V *Soyo-Maru* of Japan Fisheries Research and Education Agency.

Description of external morphology (MO-2019-19)

Disc. Disc five-lobed with notched interrational edges, approximately 26 mm in disc diameter. Radial shields and surrounding plates slightly tumid (Fig. 13B). Dorsal disc covered by variously sized

conical ossicles, which bear spiny projections on their apices (Fig. 13A, B). The larger conical external ossicles separated and scattered, approximately 140–1150 μm in length, having several thorny apical projections (Fig. 13A). Radial shields concealed by ossicles (Fig. 13B, C). One large tubercle situated on distal edge of each radial shield, approximately 1.7 mm in length (Fig. 13A).

Ventral surface of disc covered by polygonal plate-like ossicles, fully in contact, approximately 170–500 μm in length. Ossicles on ventral plates granule-shaped, approximately 130 μm in length (Fig. 13C). Oral shields, adoral shields, oral plates and ventral arm plates concealed by ossicles (Fig. 13C–E). Teeth uniformly spiniform, situated on top of dental plates and edges of ventral plates (Fig. 13C). Teeth arranged in 1 or 2 transverse rows on ventral plates, approximately 10 in number (Fig. 13C), in a cluster covering ventral-most part of dental plate, approximately 15 in number (Fig. 13C), and in a vertical line, on other areas of dental plates, 2 in number. Size of teeth varying in position on oral and dental plate, approximately 0.3–1 mm in length, 0.3 mm in greatest width on dental plates, and 1 mm in length, approximately 0.2 mm in width on ventral plates (Fig. 13C).

Interradial surface of lateral disc covered by conical ossicles similar to those on dorsal disc (Fig. 13D). They are fully in contact, approximately 40–100 μm in length (Fig. 13D). Two genital slits (0.9 mm long and 0.2 mm wide) in each interradius (Fig. 13D). One large, elliptical madreporite situated on ventral interradius, approximately 0.65 mm in width and 0.35 mm in length (Fig. 13D).

Arms. Arms branching. On proximal portion of arm, before first branch, arm 12.0 mm wide and 5.5 mm high, with an arched dorsal surface and flattened ventral surface. Between first branch and second branch, arm width and height abruptly decreasing to 4.3 mm in width and 3.0 mm in height. Subsequently, arms tapering gradually toward arm tip (Fig. 13E–J).

On dorsal and lateral surface of middle to distal portion of arms, each arm segment covered by single annular, ring-like row of large oblong plates, approximately 700 μm in transverse length (Fig. 13I, J). Before third branch, each plate separated by granules. Plates fully in contact from fourth branch and on subsequent distal segments (Fig. 13I). Before third branch, no hooklets (Fig. 13H), after fifth branch, plates appearing and forming an annual band (Fig. 13I). With exception of hooklet-bearing plates, dorsal and lateral surface of arm completely covered by conical, plate-shaped and domed granule-shaped ossicles (Fig. 13H–I). Proximal portion of dorsal arm covered by conical ossicles similar to those on dorsal disc, approximately 0.3–1.5 mm in length slightly separated, and plate-shaped external ossicles, fully in contact, approximately 200 μm in length (Fig. 13H). Middle portion of dorsal arm covered by domed granules, approximately 170–220 μm in length, and plate-shaped ossicles, approximately 110 μm in length (Fig. 13I). The larger conical ossicles sometimes carry spiny projections. Distal portion of dorsal arm covered by granule-shaped external ossicles, approximately 50 μm in length (Fig. 13J). In proximal to middle portion of arms, ventral surface covered by polygonal and plate-shaped ossicles, similar to those on ventral disc, fully in contact, approximately 150–250 μm in length at proximal region, and 60–260 in length distally (Fig. 13H, I). Distal portion of ventral arm covered by granule-shaped external ossicles, slightly in contact,

approximately 40 μm in length (Fig. 13J). Lateral arm plates and ventral arm plates concealed by skin and ossicles (Fig. 13E–G). First to fifth tentacle pore without single spine; sixth pores with 1 spine, seventh and subsequent pore with 2 or 3 spines (Fig. 13E). Distally, the number of arm spines decrease gradually to 2 toward arm tip (Fig. 13G). Arm spines approximately one-third to one-fourth of length (ca. 25–35%) of corresponding arm segment, covered by thin integument (Fig. 13E–G).

Color. Dorsal surface dark brown with whitish patches on disc and bands on arms. Ventral surface whitish but slightly brown on disc.

Ossicle morphology (MO-2019-19): Each hooklet with single inner tooth and reticular structure (Fig. 14A, C). Hooklet-bearing plates with 4 tubercle-shaped articulations for hooklets in proximal portion of the arm (Fig. 14B), approximately 5 articulations in distal portion (Fig. 14D); articulations forming 2 parallel rows (Fig. 14B, D). Lateral arm plates long, both distal and proximal edges straight (Fig. 14E). On proximal portion of arm, lateral arm plates without perforation-like structures on dorsal side and pairs of simple nerve and muscle openings on ventral-external side (Fig. 14E, F) and on distal portion of arms, no perforation-like structure on dorsal side and a pair of nerve and muscle openings beside dorsal lobe and 4 articulations for hooklets on ventral surfaces (Fig. 14G, H). Arm spines in proximal portion of arm ovoid, with four small projections, approximately one-thirds height of the height of spine (Fig. 14I). In distal portion, arm spines transformed into hooks with 2 inner secondary teeth, respectively (Fig. 15A).

All vertebrae with hourglass-shaped streptospondylous articulations (Fig. 15B, C, G, H), and distal side of branching vertebra slightly wider than in non-branching vertebra and with 2 articulation surfaces (Figs. 15E; 16B). Surfaces of lateral furrows smooth, with no special ornamentation (Figs. 15F; 16A). Depressions for tube feet openings in distal part of ventral-lateral side of vertebrae (Figs. 15D, E; 16C). Two pairs of the channels for passages of lateral canals and lateral nerves opening on ventral groove of vertebrae in proximal portion of arm (Fig. 15D, E). In distal portion of arm, only radial water canal observed (Fig. 16B). External ossicles on dorsal periphery of radial shields conical, approximately 150 μm in length and 200 μm in height with a spiny apical projection, approximately 100 μm in length (Fig. 16D).

Distribution. Widely distributed in Indo-Western Pacific Ocean. Depth range 18–494 m.

Remarks

Astrocladus exiguus can be distinguished from other congeners by its covering of large tubercles and ossicles on dorsal surface of disc and proximal regions of arms: large tubercles are conical and scattered; ossicles conical with acute thorny tips. Our molecular phylogeny showed that the two examined specimens of *A. exiguus* were monophyletic and distinguished from *A. coniferus* and *A. dofleini* (See “Molecular phylogeny” below).

Astrocladus annulatus Matsumoto, 1912

(Fig. 17)

Astrophyton annulatum Matsumoto, 1912a: 206, figs. 17–18.

Astrocladus annulatus.—Matsumoto, 1912b: 389; 1915: 56–57; 1917: 75–77, fig. 22; Clark, H. L., 1915: 187; Irimura, 1981: 19; Irimura & Kubodera, 1998: 138.

Type material examined

The holotype (UMUTZ-Oph-26): Off Misaki, Miura, Sagami Bay, Kanagawa, Japan, depth and collected data unknown, disc cut into two halves, probably done by Hikoshichiro Matsumoto (Fujita, 2006).

Description of holotype (UMUTZ-Ophi-26)

Disc. Disc five-lobed with notched interradial edges, 22 mm in diameter. Dorsal disc covered by granules, approximately 140–280 µm in length (Fig. 17B) Radial shields and their surrounds tumid, concealed by ossicles (Fig. 17A), approximately 1.1 mm in length, almost reaching disc center (Fig. 17A). Large domed tubercles, approximately 450 µm in length scattered on radial shields (Fig. 17B).

Ventral surface of disc covered by polygonal plate-like ossicles, fully in contact, approximately 160–200 µm in length (Fig. 17D). Oral shields, adoral shields, oral plates and ventral arm plates concealed by ossicles (Fig. 17D). Teeth uniformly spiniform, on top of dental plates and edges of ventral plates (Fig. 17D). Teeth approximately 8, arranged in 1 or 2 transverse rows on ventral plates in a cluster covering ventral-most part of dental plate, approximately 10 in number (Fig. 17D). Size of teeth variable, approximately 1 mm in greatest length on dental plates, approximately 0.5 mm on oral plates (Fig. 17D). Interradial surface of lateral disc covered by thick skin (Fig. 17E). Two genital slits (4.5 mm long and 1 mm wide) in each interradius (Fig. 17E). One small, elliptical madreporite on ventral interradius.

Arms. Arms branching. On the proximal portion, before first branch, arm 4.3 mm wide and 3.5 mm high, with an arched dorsal surface and flattened ventral surface (Fig. 17A, C). Between first and second branch, arm width and height abruptly decreasing to 3 mm in width and 1.8 mm in height. Subsequently, arms tapering gradually toward arm tip (Fig. 17A, C).

On dorsal and lateral surface, each arm segment covered by single annular row of large oblong plates (Fig. 17G, H). With exception of hooklet-bearing plates, dorsal and lateral surface of arm completely covered by polygonal plate-like ossicles, approximately 170–290 in length at proximal portion of arms, and subsequently decreasing in size to arm tip (Fig. 17G, H). Ventral side of arms covered by skin which completely conceals the external ossicles, lateral arm plates and ventral arm plates (Fig. 17I, J). Tentacle pores without arm spines before first branch; 3 or 4 spines after second branch. Distally, number of arm spines decrease gradually to 1 towards arm tip (Fig. 17J). In proximal portion, arm spines approximately one-fourth to one-fifth of length of corresponding arm segment, and covered by thin integument;

subsequently relative length increase, exceeding half length of corresponding arm segment on distal portion of arm (Fig. 17J).

Color. Uniformly creamy white (Fig. 17).

Distribution. JAPAN: Sagami Sea, Off Misaki, Kanagawa, central-eastern Japan depth unknown (Matsumoto, 1917); Seto, Wakayama, central Japan, depth unknown (Irimura, 1981); East China Sea, western Japan, 200 m (Irimura & Kubodera, 1998).

Remarks

Astrocladus annulatus was originally described by Matsumoto (1912) based on the holotype collected from off Misaki, Sagami Bay. It has never been re-collected from the type locality and never re-described so far. In our examination of the holotype, we confirmed the diagnostic character of this species, namely granules on dorsal surface of body (Fig. 17B) and continuous hooklet-bearing plates on proximal portions of arms (Matsumoto, 1912).

Molecular phylogeny

Phylogenetic tree and assignation of species to each detected clade

After removal of ambiguous aligned sites, 699 bp of COI were obtained for 10 specimens. The ML tree of concatenated sequence is shown in Fig. 18. The Bayesian tree also showed the same topology. In the ML analyses, monophyly of the genus *Astrocladus* was weakly supported (Fig. 18, Clade 1, bootstrap 77%, BPP 0.97). Within this clade, two clades (Fig. 18, Clade 2, bootstrap 99%, BPP 1.00; Clade 3, bootstrap 99%, BPP 1.00) were detected. The clade 3 was subdivided into two clades (Fig. 18, Clade 4, bootstrap 100%, BPP 1.00; Clade 5, bootstrap 99%, 0.99).

The specimens used in each clade were found to be identified as *A. exiguus* (Clade 2), *A. dofleini* (Clade 4) and *A. coniferus* (Clade 5), respectively (See also remarks of *A. dofleini* and *A. exiguus* above).

Genetic distances

Mean genetic distances within each clade were 0.67% in *A. exiguus* (Clade 2, 3 specimens), 0.7% in *A. dofleini* (Clade 4, 3 specimens) and 1.3% in *A. coniferus* (Clade 5, 3 specimens). Genetic distances were 13% between *A. dofleini* and *A. coniferus*, 14.7% between *A. dofleini* and *A. exiguus*, and 14.6% between *A. coniferus* and *A. exiguus*, respectively. Intra-clade distance (0.67 to 1.3%) was about ten folds smaller than inter-clade distance (13 to 14.7%).

Systematics

Our molecular phylogenetic analyses suggest that *A. exiguus*, *A. coniferus* and *A. dofleini* should be

assigned to separate taxa. Genetic distance analysis showed that the inter-clade distances exceed intra-clade values. In previous studies of ophiuroids, genetic distance corresponding to species differences range from approximately 2.2 to 23% (e.g. Okanishi, Sentoku, Martynov & Fujita, 2018). Therefore, the distances between current clades (13 to 14.7%) are within this range.

In our analysis, we found that *A. coniferus* and *A. dofleini* form a clade (Clade 3). Therefore, a possible classification would be to unite *A. coniferus* and *A. dofleini* as the same species (*A. coniferus*) and subdivide *A. coniferus coniferus* and *A. coniferus dofleini* under *A. coniferus*, as has been done in the past (e.g. Fedotov, 1926; Irimura, 1982). However, since the genetic distance between *A. dofleini* and *A. coniferus* is comparable to the distance of the two species from *A. exiguus*, which is considered to be a separate species in terms of morphology, *A. dofleini* and *A. coniferus* are herein judged to be separate species.

Conclusions

In the present study, morphological observations of type and non-type specimens revealed that *Astrocladus pardalis* (Döderlein, 1902) is a junior synonym of *A. coniferus* (Döderlein, 1902). Morphological observations and molecular phylogenetic analysis revealed that *A. coniferus* and *A. dofleini* (Döderlein, 1910) are different species. Therefore, 4 species, *A. annulatus*, *A. coniferus*, *A. dofleini* and *A. exiguus* are distributed in Japan. Additional molecular analyses including *A. annulatus* and examination of type specimens of *A. exiguus* are required to finally clarify the taxonomy of Japanese basket stars of the genus *Astrocladus*.

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References

- 681 Baker AN. 1980. Euryalinid Ophiuroidea (Echinodermata) from Australia, New Zealand, and the south-
682 west Pacific Ocean. *New Zealand Journal of Zoology* 7:73.
- 683 Baker AN, Okanishi M, Pawson DL. 2018. Euryalid brittle stars from the International Indian Ocean
684 Expedition 1963–64 (Echinodermata: Ophiuroidea: Euryalida). *Zootaxa* 4392:1–27.
- 685 Bergsten J. 2005. A review of long-branch attraction. *Cladistics* 21:163–193. DOI: 10.1111/j.1096-
686 0031.2005.00059.x LB
- 687 Bomford TL. 1913. A note on certain ophiurans in the Indian Museum. *Recor of the Indian Museum*
688 9:215–225.
- 689 Chang FY, Liao YL, We BL. 1962. Euryalae of the China Sea. *Acta Zoologica Sinica* 14:53–68.
- 690 Cherbonnier G, Guille A. 1978. *Echinodermes: Ophiurides*. Paris: Editions du Centre National de la
691 Recherche Scientifique.
- 692 Christodoulou M, O’Hara TD, Hugall AF, Arbizu PM. 2019. Dark ophiuroid biodiversity in a prospective
693 abyssal mine field. *Current Biology* 29:3909–3912.e3. DOI: 10.1016/j.cub.2019.09.012.
- 694 Clark HL. 1911. North Pacific Ophiurans in the collection of the United States National Museum.
695 *Smithsonian Institution United States National Museum Bulletin* 75:1–302.
- 696 Clark HL. 1915. Catalogue Of recent Ophiurans: Based on the Colletion of the Museum of Comparative
697 Zoology. *Memorians of the Museum of Comparative Zoölogy at Havard College XXV*:165–376.
- 698 Clark HL. 1923. The echinoderm fauna of South Africa. *Ann. S. A. Mus.* 13:221–435.
- 699 Clark AM. 1951. Some echinoderms from South Africa. *Transactions of the Royal Society of South*
700 *Africa* 33:193–221. DOI: 10.1080/00359195109519884.
- 701 Clark AM. 1974. Notes on some echinoderms from southern Africa. *Bulletin of the British Museum*
702 *(Natural History), Zoology* 26:421–487.
- 703 Clark AM, Courtman-Stock J. 1976. *The echinoderms of southern Africa*. London: British Museum
704 (Naturak History).
- 705 Clark AM, Rowe FWE. 1971. *Monograph of shallow-water Indo-West Pacific Echinoderms*. London:
706 British Museum (Natural History).
- 707 Djakonov AM. 1954. Ophiuroids of the USSR Seas. In: Pavlovskii EN ed. *Keys to the fauna of the USSR*.
708 Jerusalem: Wiener Bindery Ltd., 1–123.
- 709 Döderlein L. 1896. Bericht über die von Herrn Professor Semon bei Amboina und Thursday Island
710 gesammelten Ophiuroidea. *Denkschriften der Medicinisch-Naturwissenschaftlichen Gesellschaft zu*
711 *Jena* 8:279–300.
- 712 Döderlein L. 1902. Japanische Euryaliden. *Zool. Anze.* 25.
- 713 Döderlein L. 1911. Über japanische und andere Euryalae. *Abhandlungen der math. phys. Klasse der K.*
714 *Bayer Akademie der Wissenschaften, Suppl.* 5:1–123.
- 715 Döderlein L. 1912. Die arme der Gorgonocephalinae. *Zoologische Jahrbücher Supplement* 15:257–274.
- 716 Döderlein L. 1927. Indopacifische Euryalae. *Abhandlungen der Bayerische Akademie der*

- 717 *Wissenschaften, Mathematisch - naturwissenschaftliche Abteilung* 31:1–105.
- 718 Fedotov DM. 1926. Die morphologie der Euryalae. *Zeitschrift für wissenschaftliche Zoologie* 127:403–
- 719 528.
- 720 Fell HB. 1960. Synoptic key to the genera of Ophiuroidea. *Zoology Publications from Victoria*
- 721 *University, Wellington* 26:1–44.
- 722 Fujita T, Ishida Y, Kato T, Irimura S. 2004. Ophiuroids (Echinodermata) collected from the Oki Islands
- 723 in the Sea of Japan. *Ophiuroids (Echinodermata) Collected from the Oki Islands in the Sea of Japan*
- 724 30:191–218.
- 725 Fujita T, Kohtsuka H. 2003. Ophiuroids (Echinodermata) from Notojima Island and its adjacent waters in
- 726 the Sea of Japan. *Report of the Noto Marine Center* 9:25–38.
- 727 Gouy M, Guindon S, Gascuel O. 2010. Sea View version 4 : a multiplatform graphical user interface for
- 728 sequence alignment and phylogenetic tree building. *Molecular Biology and Evolution* 27:4.
- 729 Gray JE. 1840. Room II. In: *Synopsis of the Contents of the British Museum*. London, 57–65.
- 730 Guille A. 1981. Résultats des campagnes MUSORSTOM: 1. Philippines (18-28 Mars 1976). *Mémoires*
- 731 *du Muséum national d'Histoire naturelle. Série A, Zoologie* 91:413–456.
- 732 Hasegawa M, Kishino H, Yano T. 1985. Dating of the human-ape splitting by a molecular clock of
- 733 mitochondrial DNA. *Journal of Molecular Evolution* 22:160–174. DOI: 10.1007/BF02101694.
- 734 Hendler G. 2018. Armed to the teeth: A new paradigm for the buccal skeleton of brittle stars
- 735 (Echinodermata: Ophiuroidea). *Contributions in Science* 526:189–311.
- 736 Irimura S. 1968. Preliminary report of ophiuroids on Tanabe Bay and adjacent waters. *Nankiseibutu: The*
- 737 *Nanki Biological Society* 10:30–38.
- 738 Irimura S. 1969. Supplementary report of Dr. Murakami's paper on the ophiurans of Amakusa, Kyushu.
- 739 *Publications from the Amakusa Marine Biological Laboratory, Kyushu University* 2:37–48.
- 740 Irimura S. 1979. Ophiuroidea of Sado Island, the Sea of Japan. *Annual Report of the Sado Marine*
- 741 *Biological Station, Niigata University* 9:1–6.
- 742 Irimura S. 1981. Ophiurans from Tanabe Bay and its vicinity, with the description of a new species of
- 743 *Ophiocentrus*. *Publications of the Seto Marine Biological Laboratory* 26:15–49.
- 744 Irimura S. 1982. *The brittle-stars of Sagami Bay, collected by His Majesty the Emperor of Japan*. Japan:
- 745 Biological Laboratory, Imperial Household.
- 746 Irimura S. 1990. Echinoderms from continental shelf and slope around Japan Vol.I. In: *The Intensive*
- 747 *Research of Unexploited Fishery Resources on Continental Slopes*. Tokyo: Japan Fisheries
- 748 Resource Conservation Association, 65–100.
- 749 Irimura S, Kubodera T. 1998. Ophiuroidea in the East China Sea. *Memoirs of the National Science*
- 750 *Museum* 30:135–143.
- 751 Irimura S, Tachikawa H. 2002. Ophiuroids from the Ogasawara Islands (Echinodermata: Ophiuroidea).
- 752 *Ogasawara Research* 28:1–27.

753 Ishida Y, Ohtsuka S, Takayasu K, Kobayashi I, Lee Y-G, Tanaka H, Tamura K, Go A, Nakaguchi K.
754 2001. Preliminary Fauistic survey of Ophiuroids in the Westernmost Part of the Sea of Japan.
755 *Journal of the Faculty of Applied Biological Science, Hokkaido University* 40:14.

756 Jangoux M, de Ridder C, Fechter H. 1987. Annotated catalogue of recent echinoderm type specimens in
757 the collection of the Zoologische Staatssammlung München. *Spixiana* 10:295–311.

758 Kimura M. 1980. A simple method for estimating evolutionary rates of base substitutions through
759 comparative studies of nucleotide sequences. *Journal of Molecular Evolution* 16:111–120. DOI:
760 10.1007/BF01731581.

761 Koehler R. 1897. Échinoderms recueillis par “L’Investigator” dans l’Ocean Indian. *Annales des Scienses*
762 *Naturelles Zoologie et Paléontologie* 4:277–371.

763 Koehler R. 1905. Ophiures de l’Expédition du Siboga. Part II. *Ophiures de mer profonde. Siboga-*
764 *Expeditie* 45b:1–142.

765 Koehler R. 1907. Revision de la collection des Ophiures du Museum d’histoire Naturelle Paris. *Bulletin*
766 *Scientifique de la Franca et de la Belgique* 41:279–351.

767 Koehler R. 1930. Ophiures recueillies par le Docteur Th. Mortensen dans les Mers d’Australie et dans
768 l’Archipel Malais. Papers from Dr. Th. Mortensen’s Pacific Expedition 1914–16. LIV.
769 *Videnskabelige Meddelelser fra Dansk naturhistorisk Forening* 89:1–295.

770 Kohtsuka H, Sekifuji M, Omori A, Okanishi M. 2017. Records of young basket stars, *Astrocladus*
771 *coniferus* (Döderlein , 1902) from Misaki, Kanagawa Prefecture. *Bulletin of the Biogeographical*
772 *Society of Japan* 71:229–235.

773 Kumar S, Stecher G, Li M, Knyaz C, Tamura K. 2018. MEGA X: Molecular evolutionary genetics
774 analysis across computing platforms. *Molecular Biology and Evolution* 35:1547–1549. DOI:
775 10.1093/molbev/msy096.

776 Lamarck J-B. 1816. Ordre Second. Radiaires Échinodermes. *Histoire Naturelle des Animaux Sans*
777 *Vertèbres* 2:522–568.

778 Liao Y, Clark AM. 1995. *The echinoderms of southern China*. Beijing, New York: Science Press.

779 Ljungman A V. 1867. Ophiuroidea viventia huc usque cognita enumerat. In: *Öfversigt af Kgl.*
780 *Vetenskaps-Akademiens Förhandlingar*. 221–272.

781 Lyman T. 1875. Zoological Results of the Hassler Expedition. 2. Ophiuridae and Astrophytidae.
782 *Illustrated catalogue of the Museum of Comparative Zoology at Harvard College* 8:1–34.

783 Lyman T. 1882. Report on the Opiuroidea. *Report of the Scientific Results of the voyage of H.M.S.*
784 *Challenger 1873-76. Zoology* 5:1–386.

785 Matsumoto H. 1912a. Nihon san tedurumoduru ka ni tsuite [About Japanese Gorgonocephalidae].
786 *Dobutsu gaku zasshi [Zoological Magazine]* 282:198–206.

787 Matsumoto H. 1912b. Nihon san tedurumoduru rui no saisa [Revision of Japanese Euryalida]. *Dobutsu*
788 *gaku zasshi [Zoological Magazine]* 285:379–390.

- 789 Matsumoto H. 1915. A new classification of the Ophiuroidea: with descriptions of new genera and
790 species. *Proceedings of the Academy of Natural Sciences of Philadelphia* 67:43–92.
- 791 Matsumoto H. 1917. A monograph of Japanese Ophiuroidea, arranged according to a new classification.
792 *Journal of the College of Science Imperial University Tokyo* 38:1–408.
- 793 McKnight DG. 1989. Some echinoderm records from the tropical Southwest Pacific Ocean. *Department*
794 *of Scientific and Industrial Research, Division of Marine and Freshwater Science (Wellington)*
795 *Report* 3:19–30.
- 796 McKnight DG. 2000. The marine fauna of New Zealand: basket-stars and snake-stars (Echinodermata:
797 Ophiuroidea: Euryalinida). *NIWA Biodiversity Memoir* 115:79.
- 798 Mortensen T. 1933. Echinoderms of South Africa (Asteroidea: Ophiuroidea). *Veidenskabelige*
799 *Meddelelser fra Dansk Naturhistorisk Forening i Kjobenhavn* 93:215–400.
- 800 Müller J, Troschel FH. 1842. *System der Asteriden. 1. Asteriae. 2. Ophiuridae*. Braunschweig: Vieweg.
- 801 Murakami S. 1944a. Note on the ophiurans of Amakusa, Kyusyu. *Journal of the Department of*
802 *Agriculture, Kyushu Imperial University* 7:259–281.
- 803 Murakami S. 1944b. Report on the Ophiurans from off Ogasawara Islands and from off the Yaeyama
804 Group, Nippon. *Journal of the Department of Agriculture, Kyusyu Imperial University* 7:235–257.
- 805 O’Hara TD, Hugall AF, Thuy B, Stöhr S, Martynov A V. 2017. Restructuring higher taxonomy using
806 broad-scale phylogenomics: The living Ophiuroidea. *Molecular Phylogenetics and Evolution*
807 107:415–430. DOI: 10.1016/j.ympev.2016.12.006.
- 808 Okanishi M. 2016. Euryalida. *AccessScience*. DOI: <https://doi.org/10.1036/1097-8542.246500>.
- 809 Okanishi M, Fujita T. 2013. Molecular phylogeny based on increased number of species and genes
810 revealed more robust family-level systematics of the order Euryalida (Echinodermata: Ophiuroidea).
811 *Molecular Phylogenetics and Evolution* 69:566–580. DOI: 10.1016/j.ympev.2013.07.021.
- 812 Okanishi M, Fujita T. 2018. Description of a New Subfamily, Astrocloninae (Ophiuroidea: Euryalida:
813 Gorgonocephalidae), Based on Molecular Phylogeny and Morphological Observations. *Zoological*
814 *Science* 35. DOI: 10.2108/zs170090.
- 815 Okanishi M, O’Hara TD, Fujita T. 2011. Molecular phylogeny of the order Euryalida (Echinodermata:
816 Ophiuroidea), based on mitochondrial and nuclear ribosomal genes. *Molecular Phylogenetics and*
817 *Evolution* 61. DOI: 10.1016/j.ympev.2011.07.003.
- 818 Okanishi M, Yamaguchi K, Horii Y, Fujita T. 2011. Ophiuroids of the order Euryalida (Echinodermata)
819 from Hachijōjima Island and Ogasawara Islands, Japan. *Memoirs of the National Museum of Nature*
820 *and Science* 47:367–385.
- 821 Okanishi M, Sentoku A, Martynov A, Fujita T. 2018. A new cryptic species of *Asteronyx* Müller and
822 Troschel, 1842 (Echinodermata: Ophiuroidea), based on molecular phylogeny and morphology,
823 from off Pacific Coast of Japan. *Zoologischer Anzeiger* 274:14–33.
- 824 Retzius AJ. 1783. Anmärkningar vid. Asteriae genus. *Kungliga Svenska Vetenskaps Akademiens Nya*

- Handlingar* 4:230–248.
- Rho BJ, Shin S. 1987. Systematic study on the Ophiuroidea from Cheju Island, Korea. *The Korean Journal of Systematic Zoology* 3:208–224.
- Rowe FWE, Gates J. 1995. Echinodermata. In: Wellis A ed. *Zoological catalogue of Australia*. Australia, Melbourne: CSIRO, 1–510.
- Saba M, Tomida Y, Kimoto T. 1982. Echinoderms fauna of Ise Bay, and the northern and the middle parts of Kumano-nada. *Bulletin of the Mie Prefectural Museum, Natural Science* 4:1–82.
- Shin S. 1992. A Systematic Skdy on the Ophiuroidea in Korea II. Cheiu Island. *Korean Journal of Zoology* 35:350–361.
- Shin S, Rho BJ. 1996. *Echinodermata*. Seoul: Ministry of Education.
- Stamatakis A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* 30:1312–1313. DOI: 10.1093/bioinformatics/btu033 LB - Stamatakis2014.
- Stöhr S, O'Hara TD, Thuy B. 2012. Global diversity of brittle stars (Echinodermata: Ophiuroidea). *PLoS ONE* 7. DOI: 10.1371/journal.pone.0031940.
- Tamura K, Nei M. 1993. Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. *Molecular Biology and Evolution* 10. DOI: 10.1093/oxfordjournals.molbev.a040023.
- Thompson JD, Higgins DG, Gibson TJ. 1994. CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Research* 22:4673–4680.
- Thuy B, Stöhr S. 2011. Lateral arm plate morphology in brittle stars (Echinodermata: Ophiuroidea): new perspectives for ophiuroid micropalaeontology and classification. *Zootaxa* 3013:47.
- Verrill AE. 1899. North American Ophiuroidea. I. Revision of certain families and genera of West Indian ophiurans. *Transactions of the Connecticut Academy Arts and Sciences* 10:301–371.
- Yi SK, Irimura S. 1987. A taxonomic study on the Ophiuroidea from the Yellow Sea. *The Korean Journal of Systematic Zoology* 3:117–136.

Figure legends

Figure 1. Sampling sites of *Astrocladus annulatus*, *A. coniferus*, *A. dofleini* and *A. exiguus*. Numerals indicate serial specimen number in Table 1.

Figure 2. *Astrocladus coniferus*, lectotype (ZSM 453/1).

(A) External view of lectotype bottle. (B) Labels of the lectotype. (C) Dorsal view. (D) Ventral view. (E) Dorsal disc and proximal portion of arm. (F) Dorsal periphery of one radius of disc. (G) Ventral disc and

proximal to middle portion of arm. (H) Ventral interrarial disc. Abbreviations: G, genital slit; M, madreporite; T, large tubercle.

Figure 3. *Astrocladus coniferus*, lectotype (ZSM 453/1).

(A) Dorsal proximal portion of arm. (B) Dorsal middle portion of arm. (C) Ventral proximal portion of arm. (D) Ventral middle portion of arm. Arrowheads indicate rows of hooklets on dorsal and lateral side of the arms (A, B). Abbreviations: AS, arm spine; T, large tubercle.

Figure 4. *Astrocladus coniferus*, holotype of *Astrophyton pardalis* (ZSM 453/2) (A–F) and MO-2018-118A (G, H).

(A) Dorsal view. (B) Labels of the holotype. (C) Dorsal disc and proximal portion of arm. (D) Dorsal periphery of radial shield (E). Ventral disc. (F) Dorsal proximal portion of arm, partly enlarged. (G, H) live specimens, dorsal (G) and ventral (H) views.

Figure 5. *Astrocladus coniferus* (MO-2018-118A). SEM photographs of ossicles.

(A–C) Hooklets on proximal (A), middle (B) and distal (C) portion of arm, arcs indicate reticular structure. (D) Hooklet-bearing plate on proximal portion of arm. (E, F) Lateral arm plates on proximal portion of arm, internal (E) and external (F) views. (G, H) Arm spines from proximal portion of arm, inner most (G) and second inner most (H). (I, J) Lateral arm plates on middle portion of arm, distal (I) and internal (J) views. (K, L) Arm spines on middle portion of arms, inner most (K) and second inner most (L). Arrowheads indicate articulations for hooklets (D) and terminal projections (G, H, K, L). Orientations: dis, distal side; ex, external side; in, internal side; prox, proximal side. Abbreviations: MO, muscle opening; NO, nerve opening; P, perforation; ST, secondary tooth.

Figure 6. *Astrocladus coniferus* (MO-2018-118A). SEM photographs of ossicles.

(A, B) Lateral arm plates on distal portion of arm, external (A) and internal (B) views. (C) Hook-shaped arm spine on distal portion of arm. (D–G) Vertebrae from proximal (D, E) and middle (F, G) portion of arm, distal (D, G), ventral (E) and proximal (F) views. An arrowhead indicates articulation for hooklet (A). Abbreviations: B, border structure; LN, passage of lateral canal; NO, nerve opening; ST, secondary tooth.

Figure 7. *Astrocladus coniferus* (MO-2018-118A). SEM photographs of ossicles.

Vertebrae from middle (A, B) and distal (C–F) portion of arm, ventral (A, E), dorsal (B, F), proximal (C) and distal (D) views. Orientations: d, dorsal side; dis, distal side; prox, proximal side; v, ventral side. Abbreviations: DT, depression for tentacle; LC, passages of lateral canal; LN, passages of lateral nerve; T, tubercle.

Figure 8. *Astrocladus dofleini*, lectotype (440/1).

(A) External view of lectotype bottle. (B) Labels of the lectotype. (C) Dorsal disc and proximal portion of arm, periphery part of disc enlarged in upper-right. (D) Dorsal proximal portion of arm, partly enlarged in upper right. (E) Interradial ventral disc. (F) Dorsal middle to distal portion of arm. (G) Ventral disc and proximal portion of arm. (H) Ventral middle to distal tips of arm. Abbreviation: G, genital slit.

Figure 9. *Astrocladus dofleini* (MO-2018-118B). SEM photographs of ossicles.

(A, B, D) Hooklets on proximal (A), middle (B) and distal (D) portion of arms, arcs indicate reticular structure. (C, E) Hooket-bearing plate on proximal (C) and distal (E) portion of arm. (F, I) Arm spines on proximal (F) and middle (I) portion of arms. (G, H, J, K) Lateral arm plates on proximal (G, H) and middle (J, K) portion of the arms, distal (G, J) and internal (H, K) views. Arrowheads indicate articulations of hooklets (C, E, J) and terminal projections (F). Orientations: d, dorsal side; dis, distal side; ex, external side; in, internal side prox, proximal side; v, ventral side. Abbreviations: B, border structure; MO, muscle opening; NO, nerve opening; ST, secondary tooth.

Figure 10. *Astrocladus dofleini* (MO-2018-118B). SEM photographs of ossicles.

(A) An arm spine from distal portion of arm. (B, C) Lateral arm plates on distal portion of arm, distal (B) and internal (C) views. (D–H) Vertebrae from proximal portion of arm (H is branching vertebra), distal (D), proximal (E), dorsal (F) and ventral (G, H) views. Orientations: d, dorsal side; dis, distal side; prox, proximal side; v, ventral side. Arrowheads indicate articulations for hooklets. Abbreviations: B, boarder structure; DT, depression for tentacle; LC, passages of lateral canal; LN, passages of lateral nerve; MO, muscle opening; ST, secondary tooth; T, tubercle.

Figure 11. *Astrocladus dofleini* (MO-2018-118B). SEM photographs of ossicles.

(A–E) Vertebrae from middle portion of arm (C is branching vertebra), distal (A), ventral (B, C), dorsal (D) views, a part of (D) enlarged in (E). Vertebrae from distal portion of arm (F, G), distal (F) and proximal (G) views. Orientations: d, dorsal side; dis, distal side; prox, proximal side; v, ventral side. Arrowheads indicate tubercles on lateral furrow of vertebra. Abbreviations: DT, depression for tentacle; LC, passages of lateral canal; LN, passages of lateral nerve; T, tubercle.

Figure 12. *Astrocladus dofleini* (MO-2018-118B).

SEM photographs of vertebrae from distal portion of arm (C is branching vertebra), ventral (A, C) and dorsal (B) views. Orientations: dis, distal side; prox, proximal side. Abbreviation: DT, depression for tentacle.

Figure 13. *Astrocladus exiguus* (MO-2019-19).

(A) Periphery of dorsal disc. (B) Central view of dorsal disc. (C) Ventral disc. (D) Interradial ventral disc. (E–G) Ventral surfaces of arms, proximal (E), middle (F) and distal (G) portion of arm. Dorsal surfaces of arms, proximal (H), middle (I) and distal (J) portion of arm. Arrowheads indicate rows of hooklets on dorsal and lateral side of the arms (I, J). Abbreviations: AS, arm spine; G, genital slit; M, madreporite; T, large tubercle.

Figure 14. *Astrocladus exiguus* (MO-2019-19). SEM photographs of ossicles.

(A, C) Hooklets on proximal (A) and distal (C) portion of arms, arcs indicate reticular structure. (B, D) Hooket-bearing plate on proximal (B) and distal (D) portion of arm. (E–H) Lateral arm plates on proximal (E, F) and distal (G, H) portion of arms. (I) An arm spine on proximal portion of arm. Arrowheads indicate articulations for hooklets (B, D, H) and terminal projections (I). Orientations: d, dorsal side; dis, distal side; ex, internal side; in, internal side; prox, proximal side; v, ventral side. Abbreviations: B, border structure; MO, muscle opening; NO, nerve opening; ST, secondary tooth.

Figure 15. *Astrocladus exiguus* (MO-2019-19). SEM photographs of ossicles.

(A) An arm spine on distal portion of arm. (B–F) Vertebrae from proximal portion of arm (E is branching vertebra), proximal (B), distal (C), ventral (D, E) and dorsal (F) views. (G, H) Vertebrae from distal portion of arm, proximal (G) and distal (H) views. Orientations: d, dorsal side; dis, distal side; prox, proximal side; v, ventral side. Abbreviations: DT, depression for tentacle; LC, passages of lateral canal; LN, passages of lateral nerve; ST, secondary tooth.

Figure 16. *Astrocladus exiguus* (MO-2019-19). SEM photographs of ossicles.

(A–C) Vertebrae from distal portion of arm (B is branching vertebra), dorsal (A), ventral (B, C) views. (D) An conical external ossicle on proximal portion of arm, lateral view, an arc indicates a terminal projection. Orientations: ba, basal side; dis, distal side; ex, external side; prox, proximal side. Abbreviations: DT, depression for tentacle; LC, passages for lateral canals.

Figure 17. *Astrocladus annulatus*, holotype (UMUTZ-Ophi-26).

(A) Dorsal view. (B) Dorsal surface of periphery disc (C) Ventral view. (D) Jaws. (E) Interradial ventral disc. (F–H) Dorsal surface of arms, proximal (F), middle (G) and distal (H) portion of arm. (I, J) Ventral surface of arms, proximal (I) and distal (J) portion of arm. Arrowheads indicate rows of hooklets on dorsal and lateral side of the arms (B, G, H). Abbreviations: AS, arm spine; G, genital slit.

Figure 18. Maximum likelihood tree based on a partial sequence of mitochondrial COI gene (699 bp).

Support values for each node are shown by maximum likelihood bootstrap values (%) and Bayesian

posterior probabilities. Numerals (1–5) in circles at nodes refer to the clade number discussed in the text. Bootstrap value less than 74% and Bayesian posterior probability value less than 0.97 and for each node were shown by as “-”.

Table 1:

Examined specimens of *Astrocladus* species including outgroup.

COI accession numbers are lodged at the DNA Data Bank of Japan. See referees for the detailed information. Unknown data are shown by “-”. Abbreviations: NSMT, the National Museum of Nature and Science, Tsukuba, Japan; UMUT, The University Museum, The University of Tokyo, Japan; ZMB, Museum für Naturkunde der Humboldt-Universität zu Berlin, Germany; ZSM, the Zoologische Staatssammlung München, Germany.

Figure 1

Figure 1. Sampling sites of *Astrocladus annulatus*, *A. coniferus*, *A. dofleini* and *A. exiguus*.

Numerals indicate serial specimen number in Table 1.

FIGURE 1

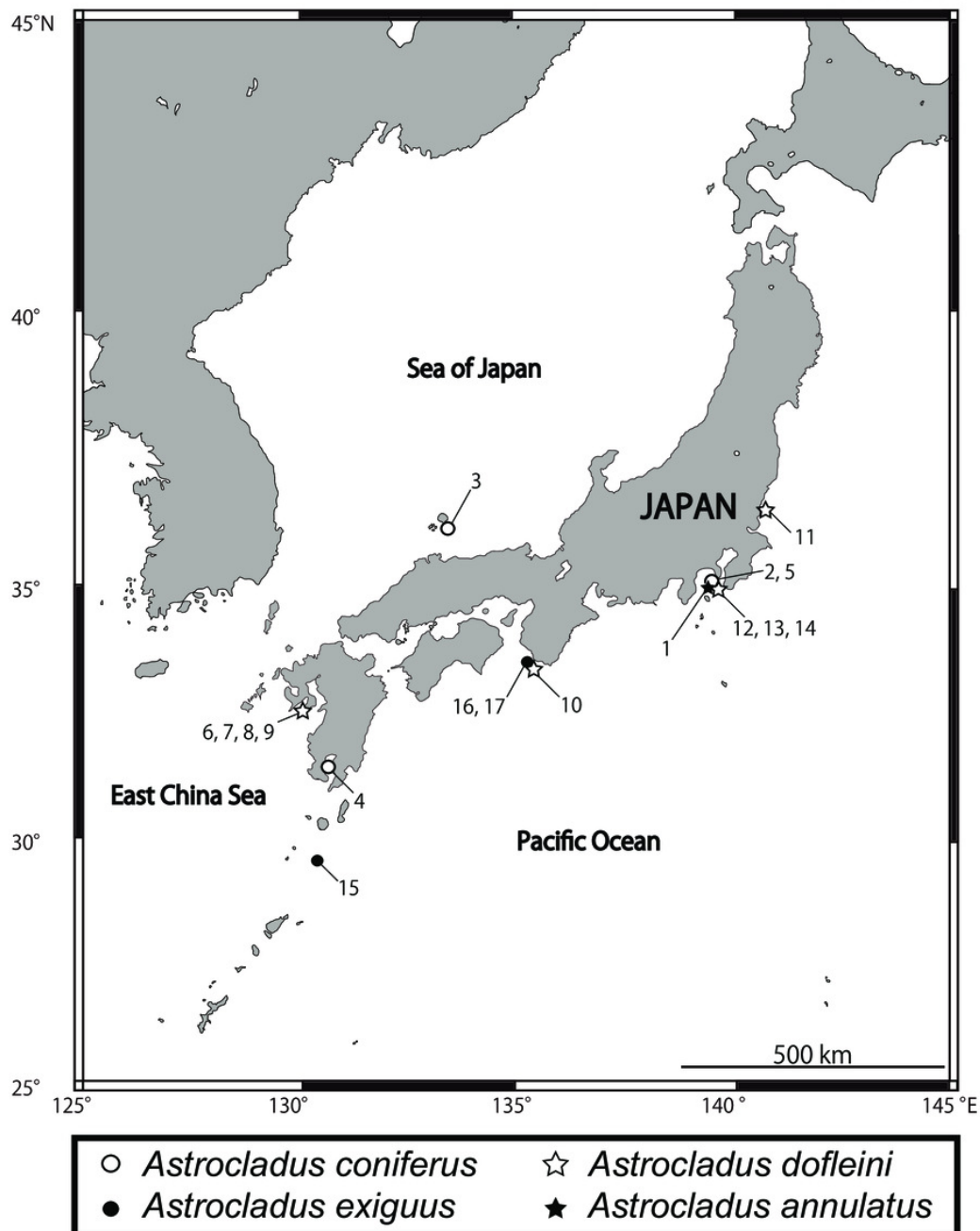


Figure 2

Figure 2. *Astrocladus coniferus*, lectotype (ZSM 453/1).

(A) External view of lectotype bottle. (B) Labels of the lectotype. (C) Dorsal view. (D) Ventral view. (E) Dorsal disc and proximal portion of arm. (F) Dorsal periphery of one radius of disc. (G) Ventral disc and proximal to middle portion of arm. (H) Ventral interrarial disc.

Abbreviations: G, genital slit; M, madreporite; T, large tubercle.

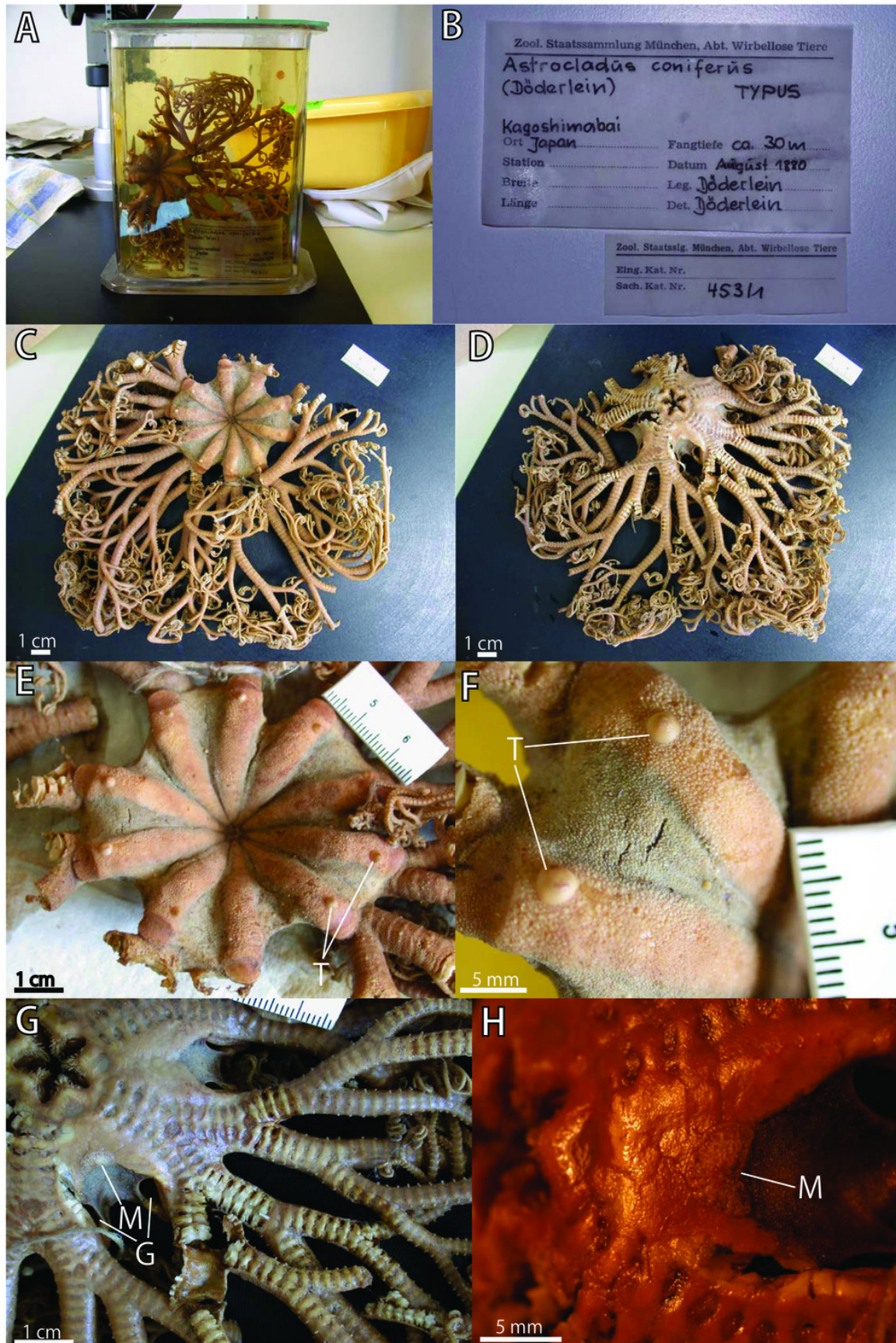


Figure 3

Figure 3. *Astrocladus coniferus*, lectotype (ZSM 453/1).

(A) Dorsal proximal portion of arm. (B) Dorsal middle portion of arm. (C) Ventral proximal portion of arm. (D) Ventral middle portion of arm. Arrowheads indicate rows of hooklets on dorsal and lateral side of the arms (A, B). Abbreviations: AS, arm spine; T, large tubercle.

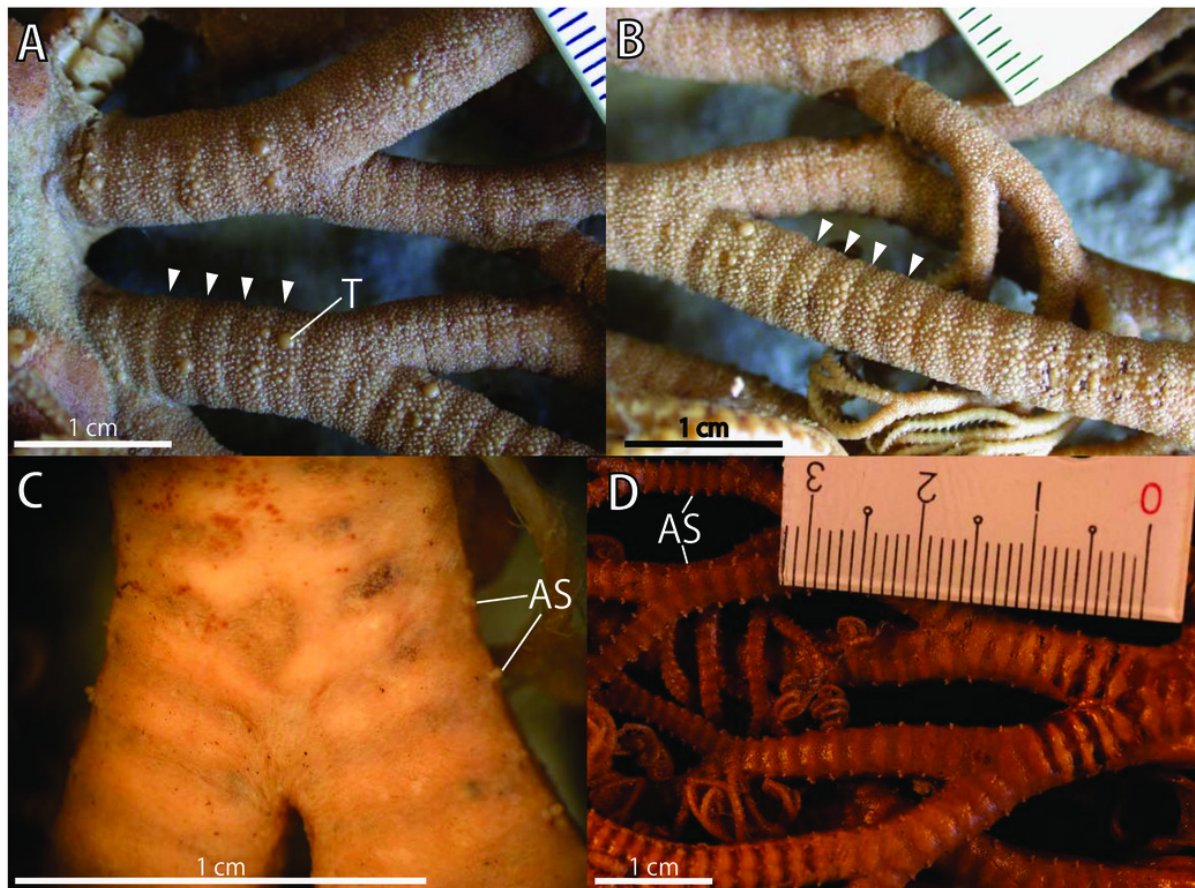


Figure 4

Figure 4. *Astrocladus coniferus*, holotype of *Astrophyton pardalis* (ZSM 453/2) (A-F) and MO-2018-118A (G, H).

(A) Dorsal view. (B) Labels of the holotype. (C) Dorsal disc and proximal portion of arm. (D) Dorsal periphery of radial shield (E). Ventral disc. (F) Dorsal proximal portion of arm, partly enlarged. (G, H) live specimens, dorsal (G) and ventral (H) views.

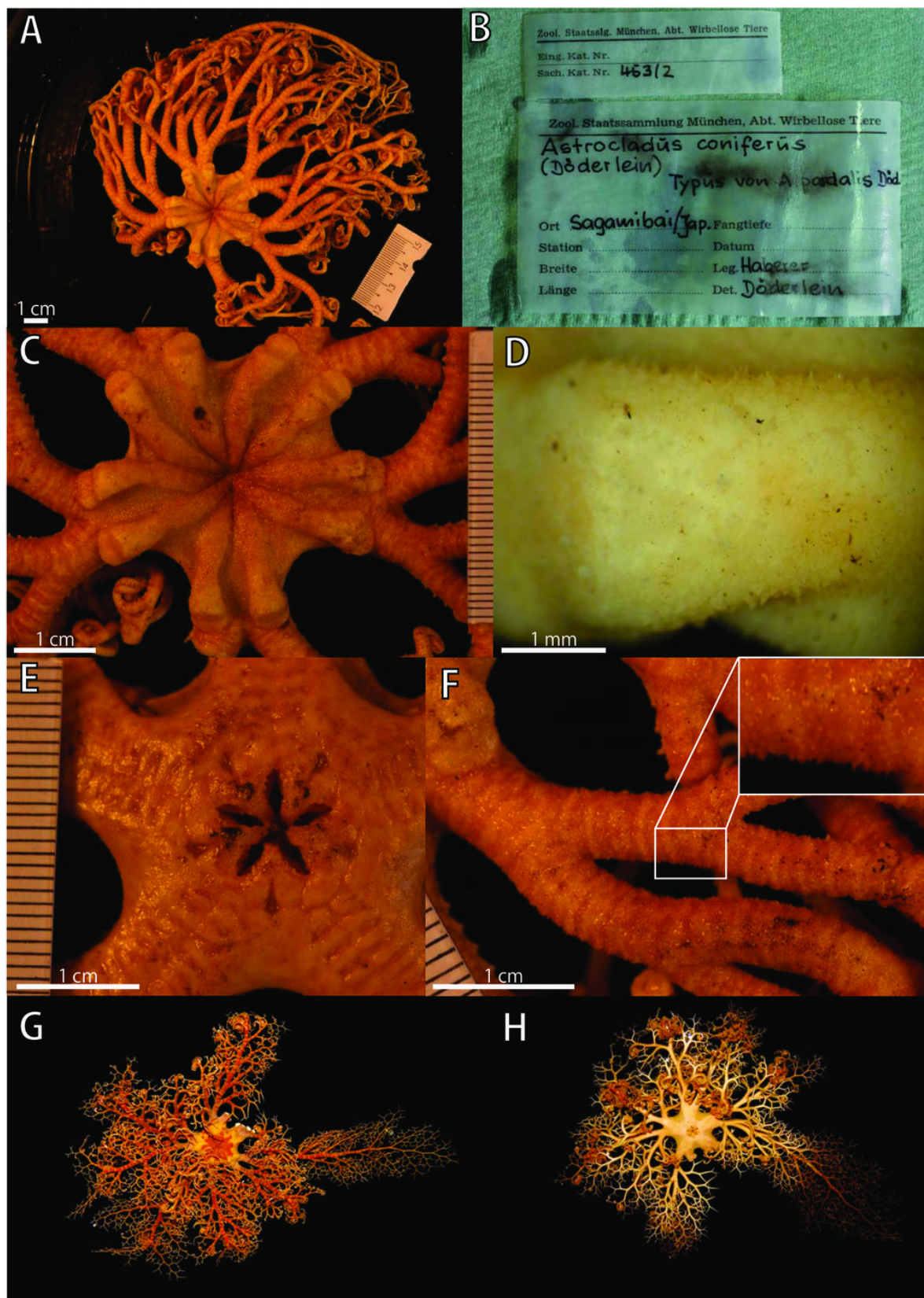


Figure 5

Figure 5. *Astrocladus coniferus* (MO-2018-118A). SEM photographs of ossicles.

(A–C) Hooklets on proximal (A), middle (B) and distal (C) portion of arm, arcs indicate reticular structure. (D) Hooket-bearing plate on proximal portion of arm. (E, F) Lateral arm plates on proximal portion of arm, internal (E) and external (F) views. (G, H) Arm spines from proximal portion of arm, inner most (G) and second inner most (H). (I, J) Lateral arm plates on middle portion of arm, distal (I) and internal (J) views. (K, L) Arm spines on middle portion of arms, inner most (K) and second inner most (L). Arrowheads indicate articulations for hooklets (D) and terminal projections (G, H, K, L). Orientations: dis, distal side; ex, external side; in, internal side; prox, proximal side. Abbreviations: MO, muscle opening; NO, nerve opening; P, perforation; ST, secondary tooth.

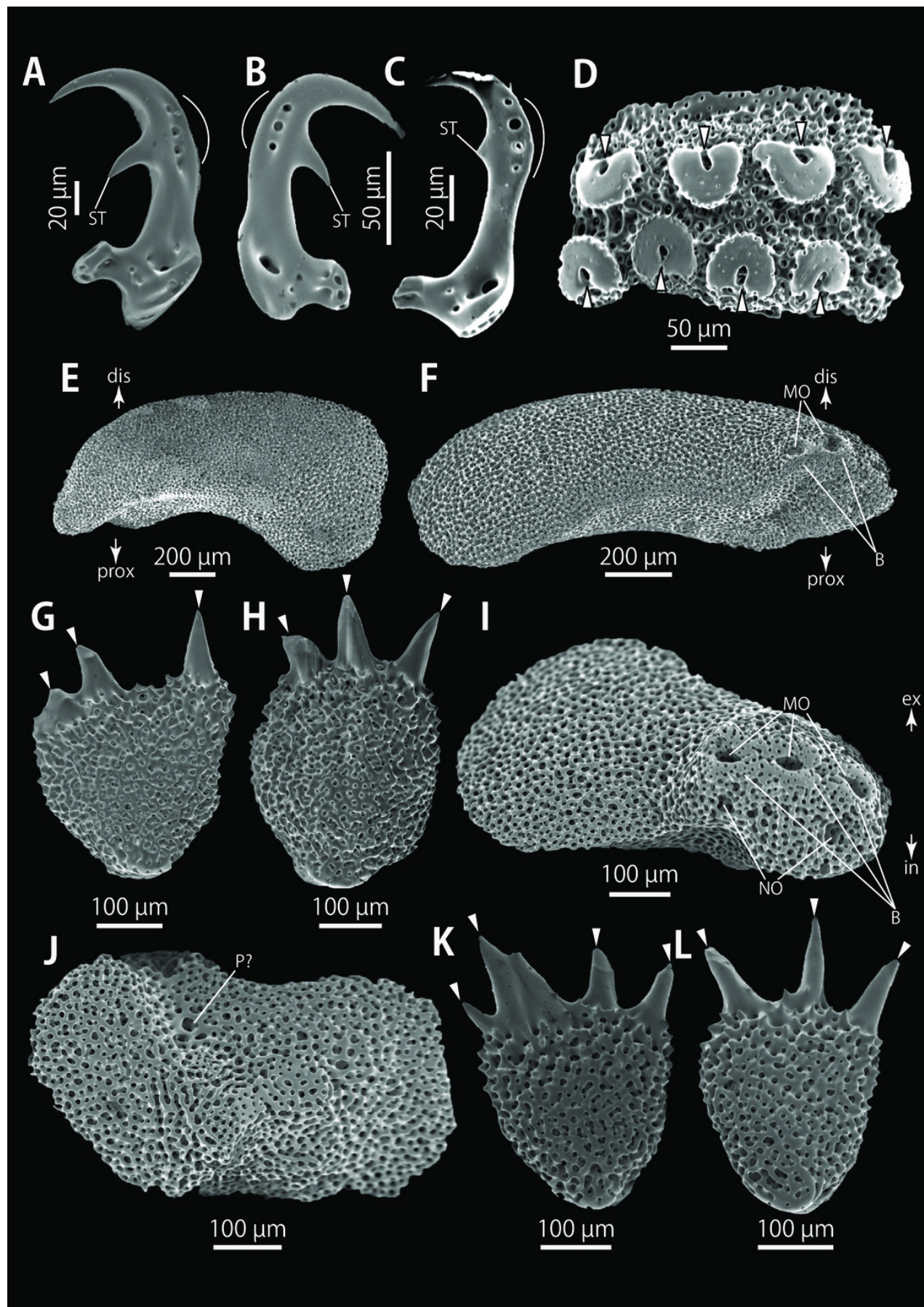


Figure 6

Figure 6. *Astrocladus coniferus* (MO-2018-118A). SEM photographs of ossicles.

(A, B) Lateral arm plates on distal portion of arm, external (A) and internal (B) views. (C) Hook-shaped arm spine on distal portion of arm. (D–G) Vertebrae from proximal (D, E) and middle (F, G) portion of arm, distal (D, G), ventral (E) and proximal (F) views. An arrowhead indicates articulation for hooklet (A). Abbreviations: B, border structure; LN, passage of lateral canal; NO, nerve opening; ST, secondary tooth.

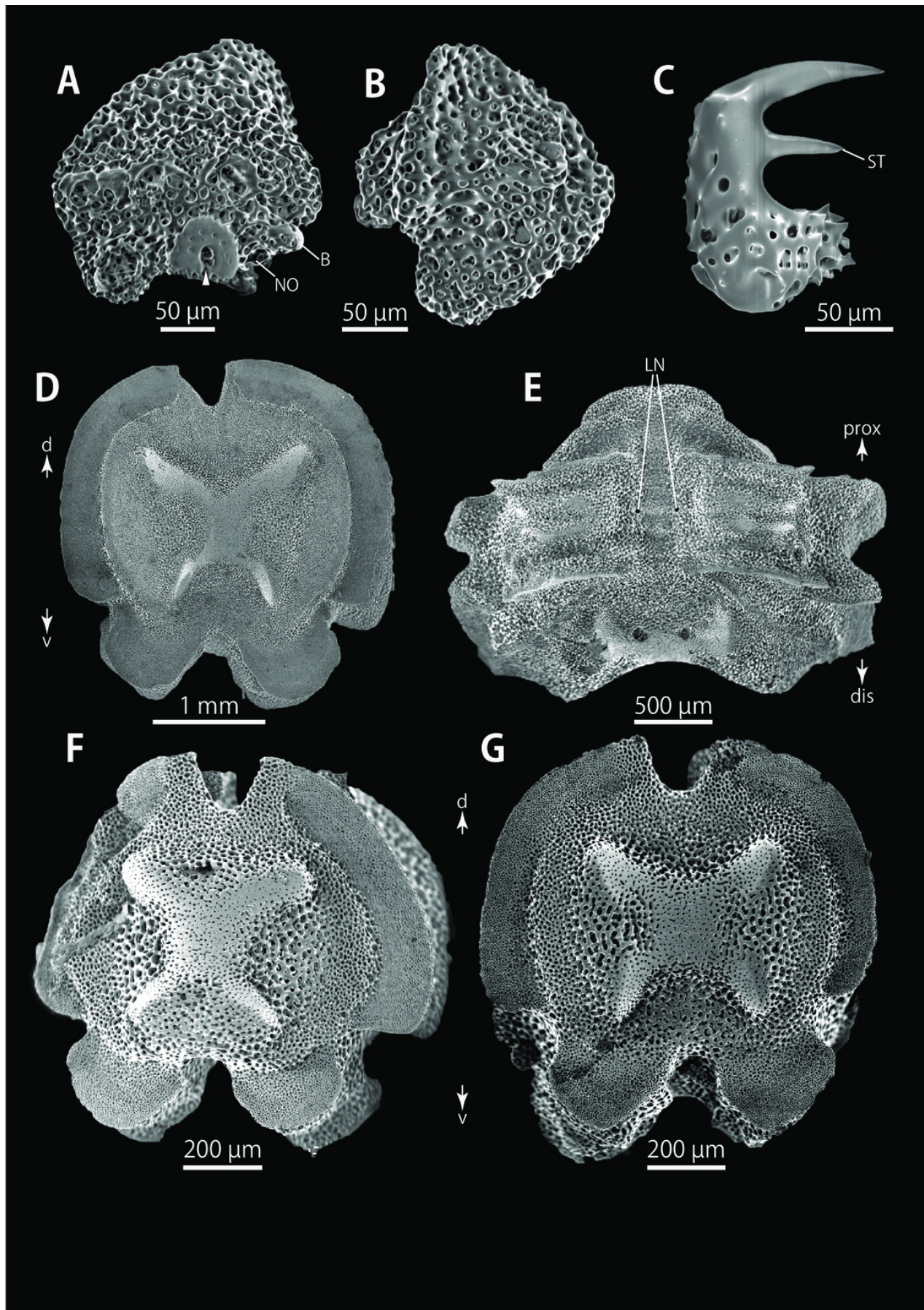


Figure 7

Figure 7. *Astrocladus coniferus* (MO-2018-118A). SEM photographs of ossicles.

Vertebrae from middle (A, B) and distal (C–F) portion of arm, ventral (A, E), dorsal (B, F), proximal (C) and distal (D) views. Orientations: d, dorsal side; dis, distal side; prox, proximal side; v, ventral side. Abbreviations: DT, depression for tentacle; LC, passages of lateral canal; LN, passages of lateral nerve; T, tubercle.

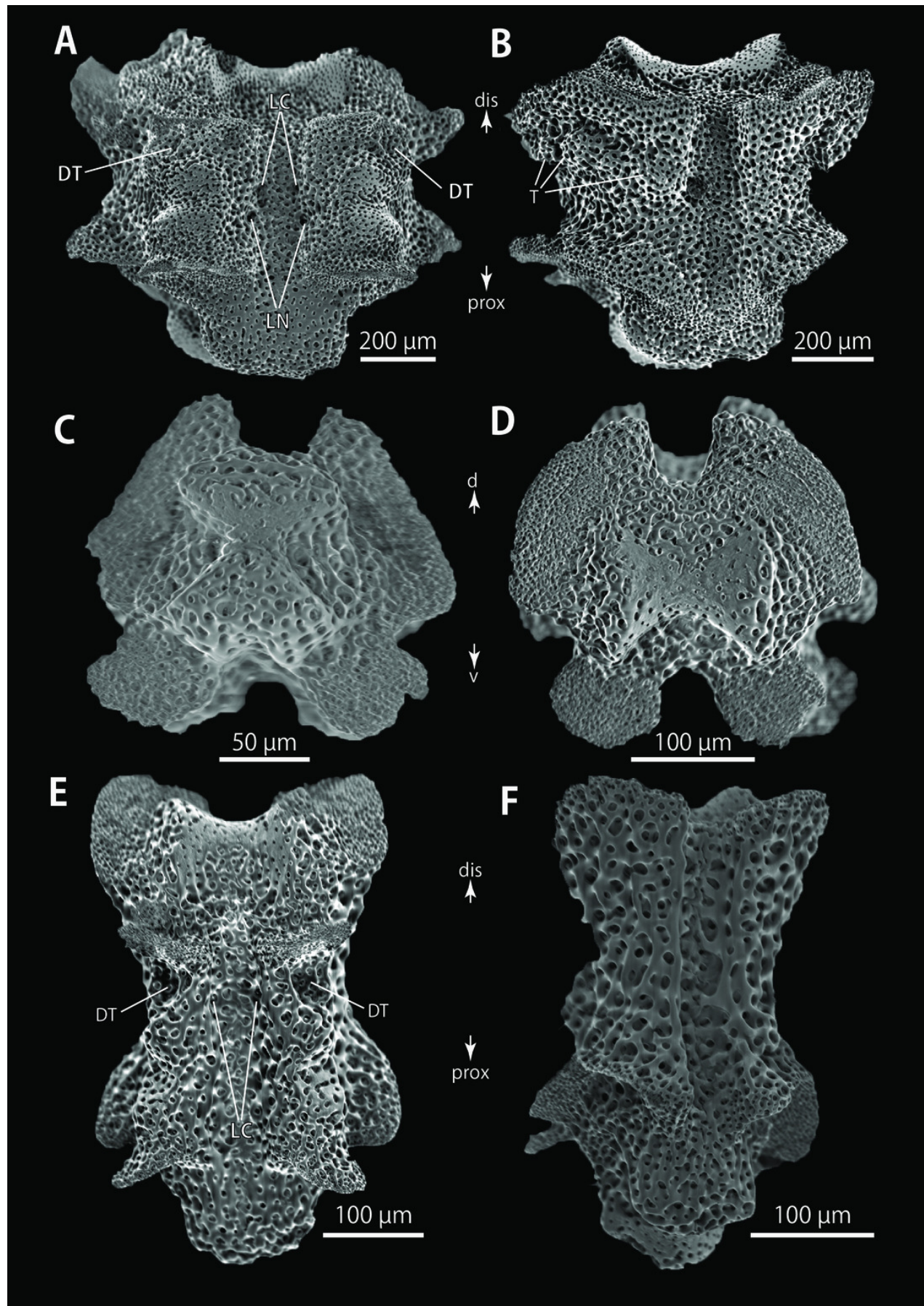


Figure 8

Figure 8. *Astrocladus dofleini*, lectotype (440/1).

(A) External view of lectotype bottle. (B) Labels of the lectotype. (C) Dorsal disc and proximal portion of arm, periphery part of disc enlarged in upper-right. (D) Dorsal proximal portion of arm, partly enlarged in upper right. (E) Interradial ventral disc. (F) Dorsal middle to distal portion of arm. (G) Ventral disc and proximal portion of arm. (H) Ventral middle to distal tips of arm. Abbreviation: G, genital slit.

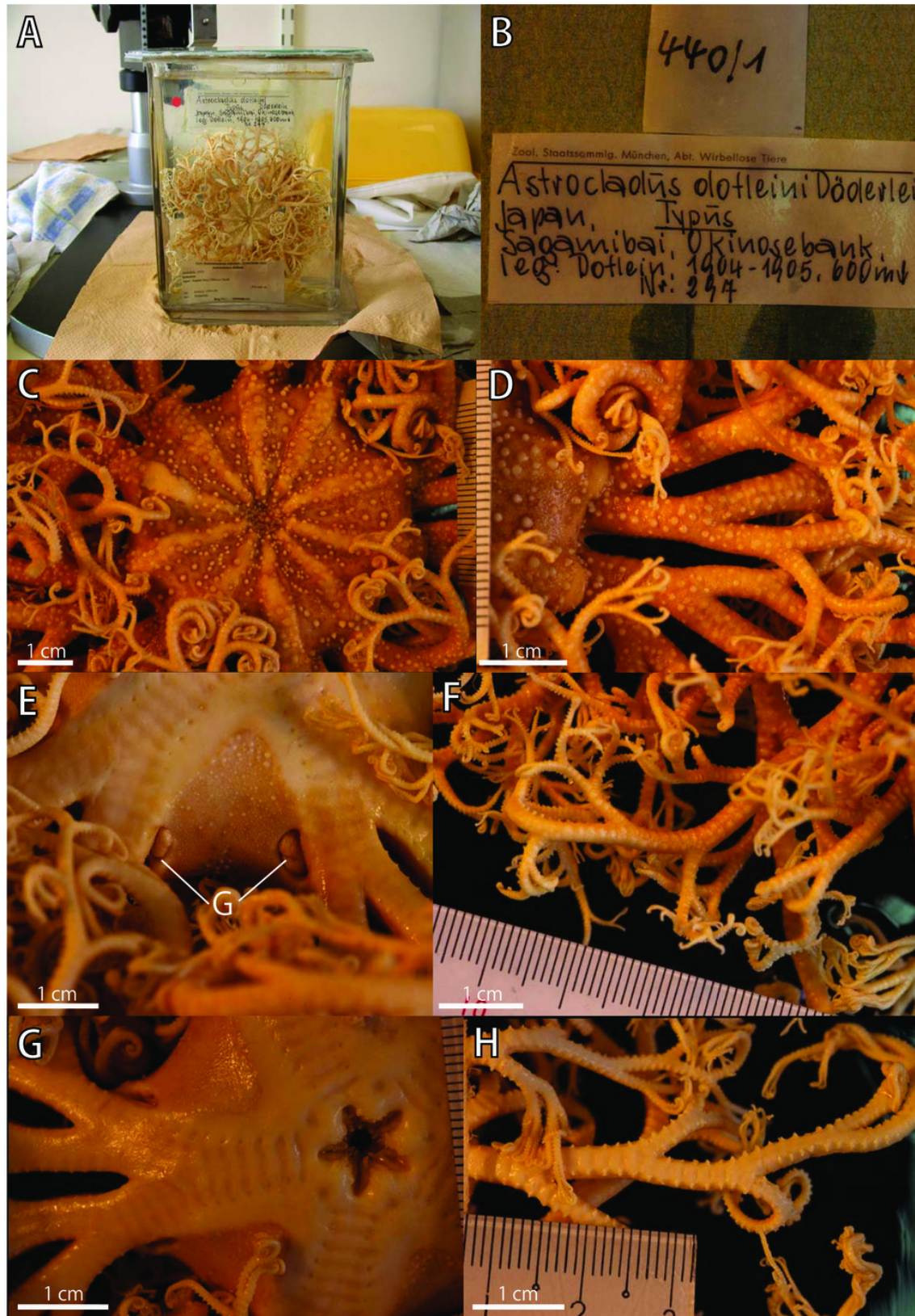


Figure 9

Figure 9. *Astrocladus dofleini* (MO-2018-118B). SEM photographs of ossicles.

(A, B, D) Hooklets on proximal (A), middle (B) and distal (D) portion of arms, arcs indicate reticular structure. (C, E) Hooket-bearing plate on proximal (C) and distal (E) portion of arm. (F, I) Arm spines on proximal (F) and middle (I) portion of arms. (G, H, J, K) Lateral arm plates on proximal (G, H) and middle (J, K) portion of the arms, distal (G, J) and internal (H, K) views. Arrowheads indicate articulations of hooklets (C, E, J) and terminal projections (F).

Orientations: d, dorsal side; dis, distal side; ex, external side; in, internal side prox, proximal side; v, ventral side. Abbreviations: B, border structure; MO, muscle opening; NO, nerve opening; ST, secondary tooth.

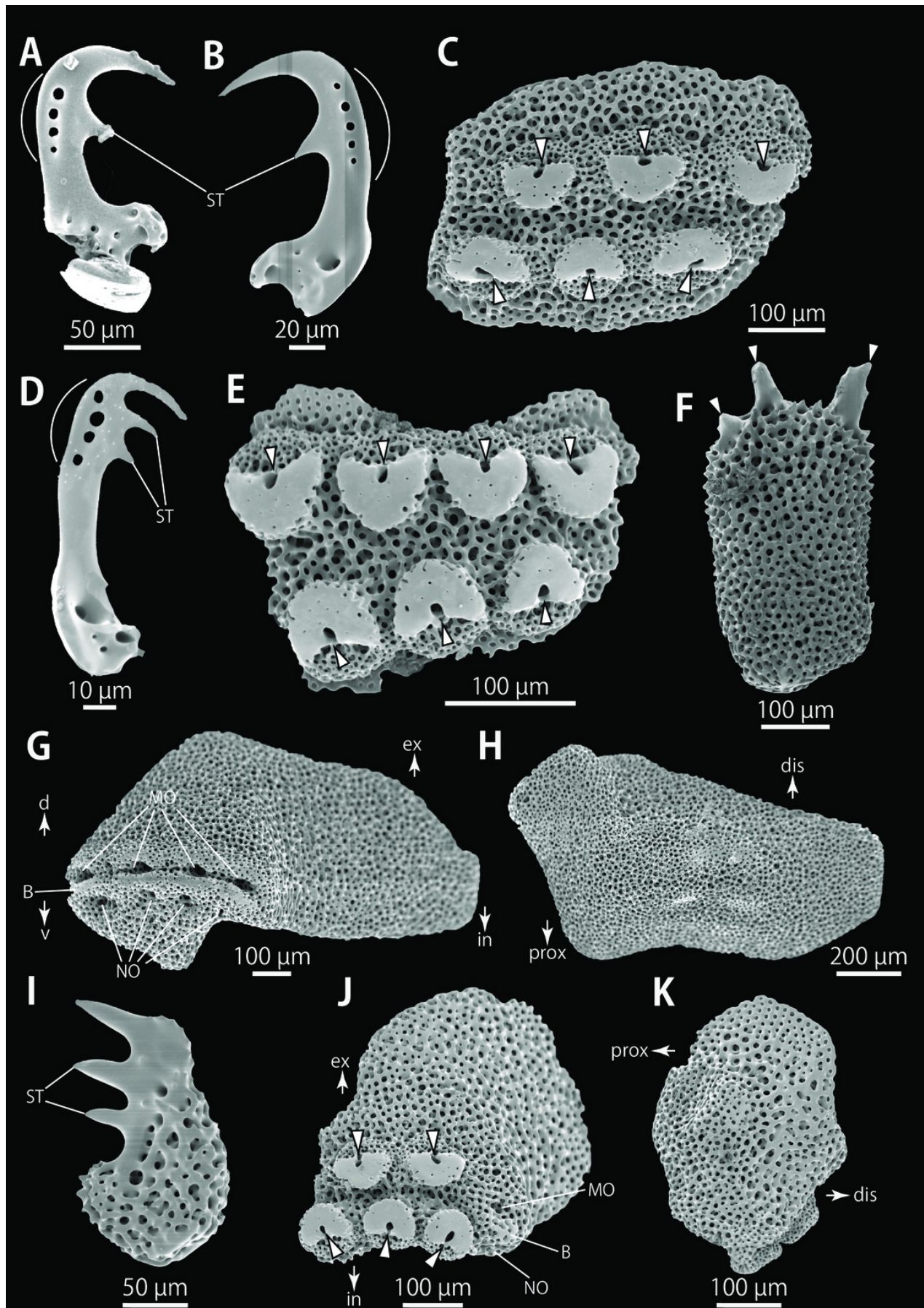


Figure 10

Figure 10. *Astrocladus dofleini* (MO-2018-118B). SEM photographs of ossicles.

(A) An arm spine from distal portion of arm. (B, C) Lateral arm plates on distal portion of arm, distal (B) and internal (C) views. (D-H) Vertebrae from proximal portion of arm (H is branching vertebra), distal (D), proximal (E), dorsal (F) and ventral (G, H) views. Orientations: d, dorsal side; dis, distal side; prox, proximal side; v, ventral side. Arrowheads indicate articulations for hooklets. Abbreviations: B, boarder structure; DT, depression for tentacle; LC, passages of lateral canal; LN, passages of lateral nerve; MO, muscle opening; ST, secondary tooth; T, tubercle.

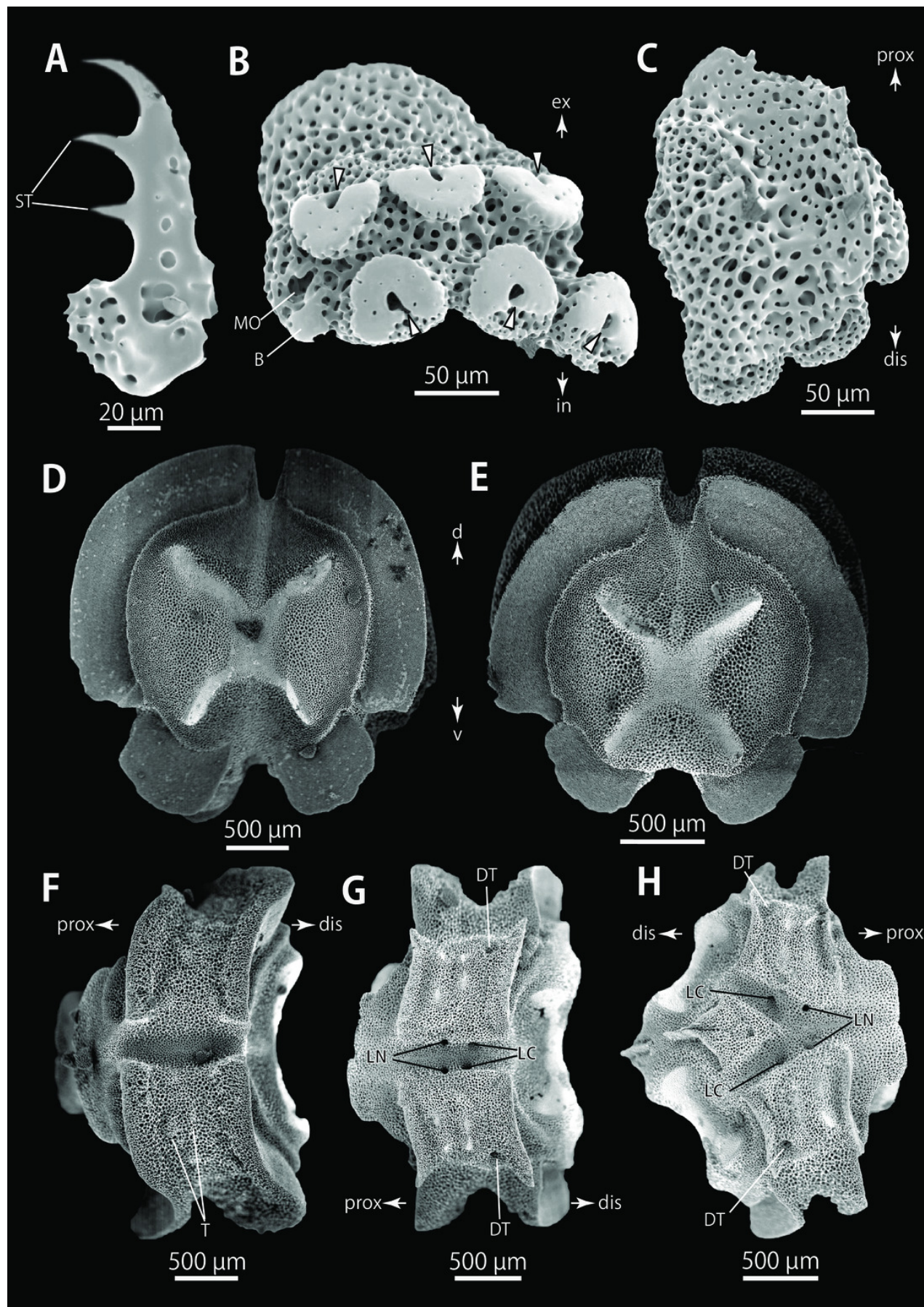


Figure 11

Figure 11. *Astrocladus dofleini* (MO-2018-118B). SEM photographs of ossicles.

(A-E) Vertebrae from middle portion of arm (C is branching vertebra), distal (A), ventral (B, C), dorsal (D) views, a part of (D) enlarged in (E). Vertebrae from distal portion of arm (F, G), distal (F) and proximal (G) views. Orientations: d, dorsal side; dis, distal side; prox, proximal side; v, ventral side. Arrowheads indicate tubercles on lateral furrow of vertebra.

Abbreviations: DT, depression for tentacle; LC, passages of lateral canal; LN, passages of lateral nerve; T, tubercle.

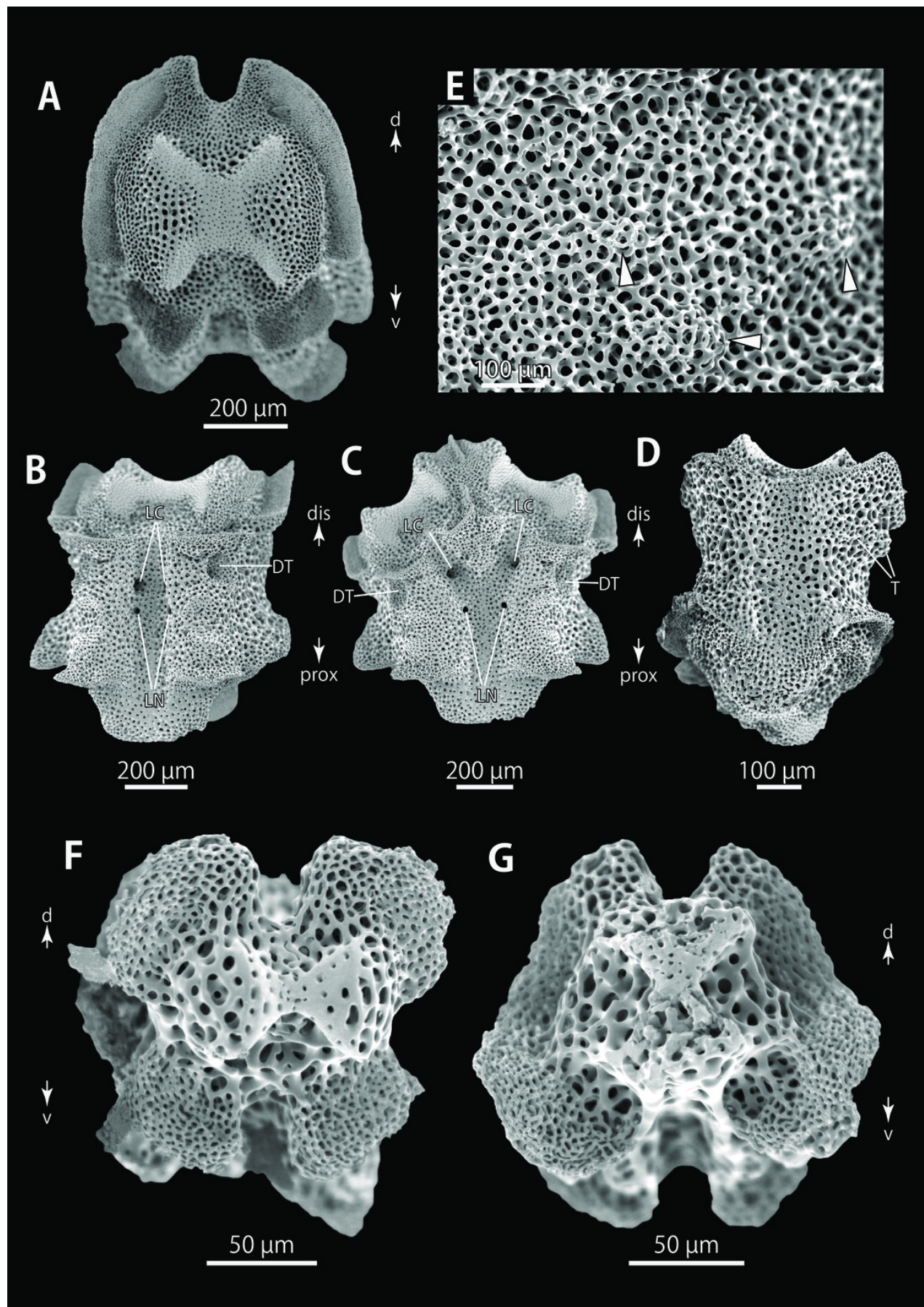


Figure 12

Figure 12. *Astrocladus dofleini* (MO-2018-118B).

SEM photographs of vertebrae from distal portion of arm (C is branching vertebra), ventral (A, C) and dorsal (B) views. Orientations: dis, distal side; prox, proximal side. Abbreviation: DT, depression for tentacle.

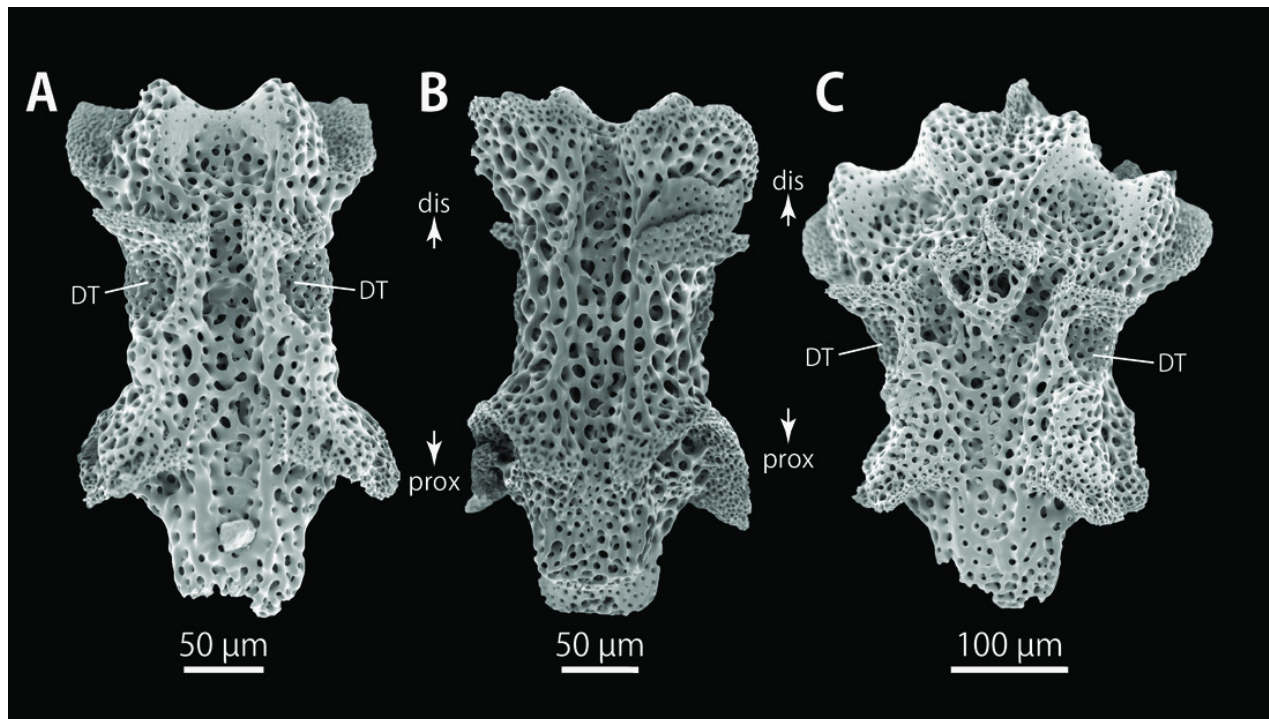


Figure 13

Figure 13. *Astrocladus exiguus* (MO-2019-19).

(A) Periphery of dorsal disc. (B) Central view of dorsal disc. (C) Ventral disc. (D) Interradial ventral disc. (E–G) Ventral surfaces of arms, proximal (E), middle (F) and distal (G) portion of arm. Dorsal surfaces of arms, proximal (H), middle (I) and distal (J) portion of arm. Arrowheads indicate rows of hooklets on dorsal and lateral side of the arms (I, J). Abbreviations: AS, arm spine; G, genital slit; M, madreporite; T, large tubercle.

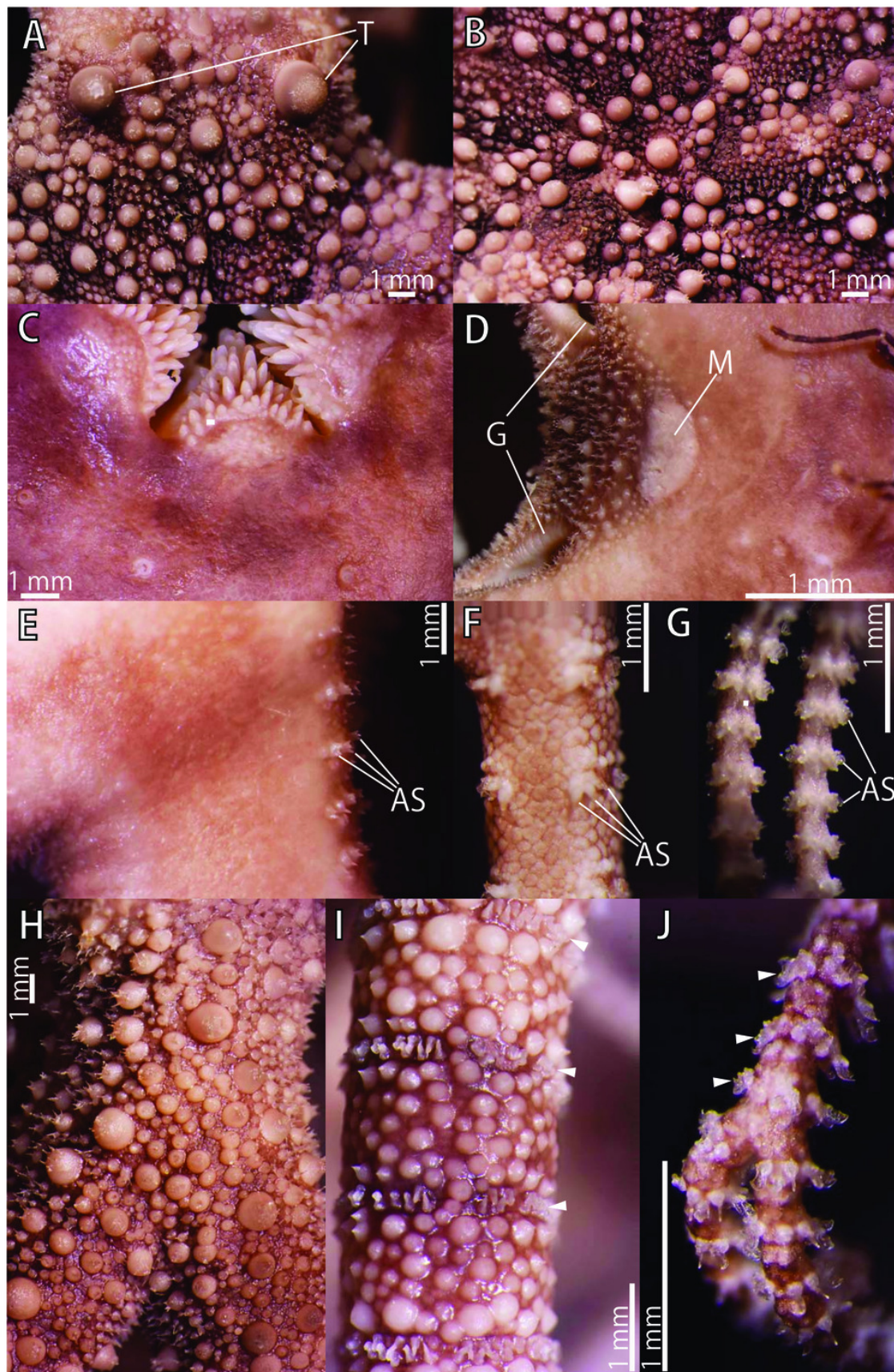


Figure 14

Figure 14. *Astrocladus exiguus* (MO-2019-19). SEM photographs of ossicles.

(A, C) Hooklets on proximal (A) and distal (C) portion of arms, arcs indicate reticular structure. (B, D) Hocket-bearing plate on proximal (B) and distal (D) portion of arm. (E-H) Lateral arm plates on proximal (E, F) and distal (G, H) portion of arms. (I) An arm spine on proximal portion of arm. Arrowheads indicate articulations for hooklets (B, D, H) and terminal projections (I). Orientations: d, dorsal side; dis, distal side; ex, internal side; in, internal side; prox, proximal side; v, ventral side. Abbreviations: B, border structure; MO, muscle opening; NO, nerve opening; ST, secondary tooth.

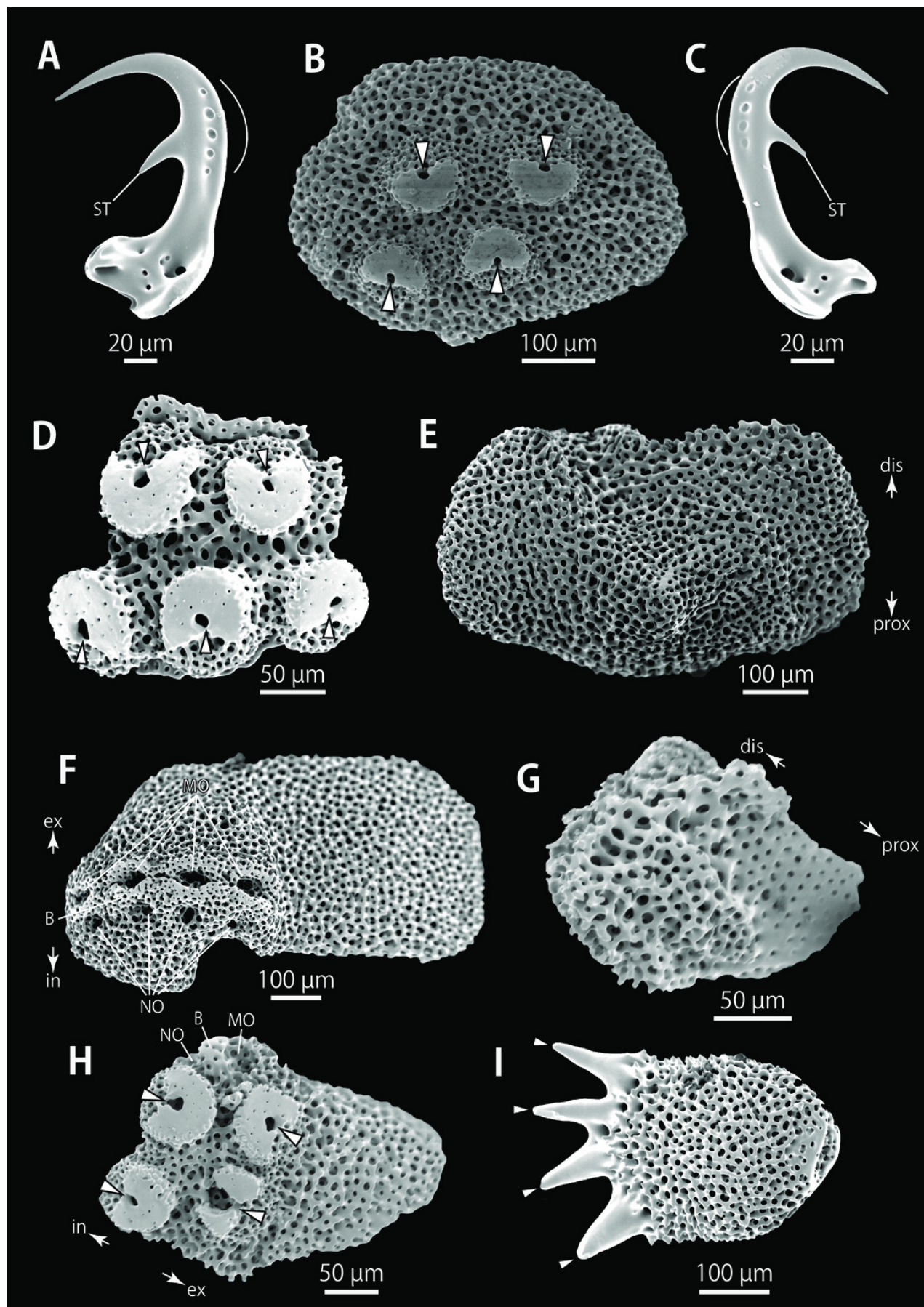


Figure 15

Figure 15. *Astrocladus exiguus* (MO-2019-19). SEM photographs of ossicles.

(A) An arm spine on distal portion of arm. (B–F) Vertebrae from proximal portion of arm (E is branching vertebra), proximal (B), distal (C), ventral (D, E) and dorsal (F) views. (G, H) Vertebrae from distal portion of arm, proximal (G) and distal (H) views. Orientations: d, dorsal side; dis, distal side; prox, proximal side; v, ventral side. Abbreviations: DT, depression for tentacle; LC, passages of lateral canal; LN, passages of lateral nerve; ST, secondary tooth.

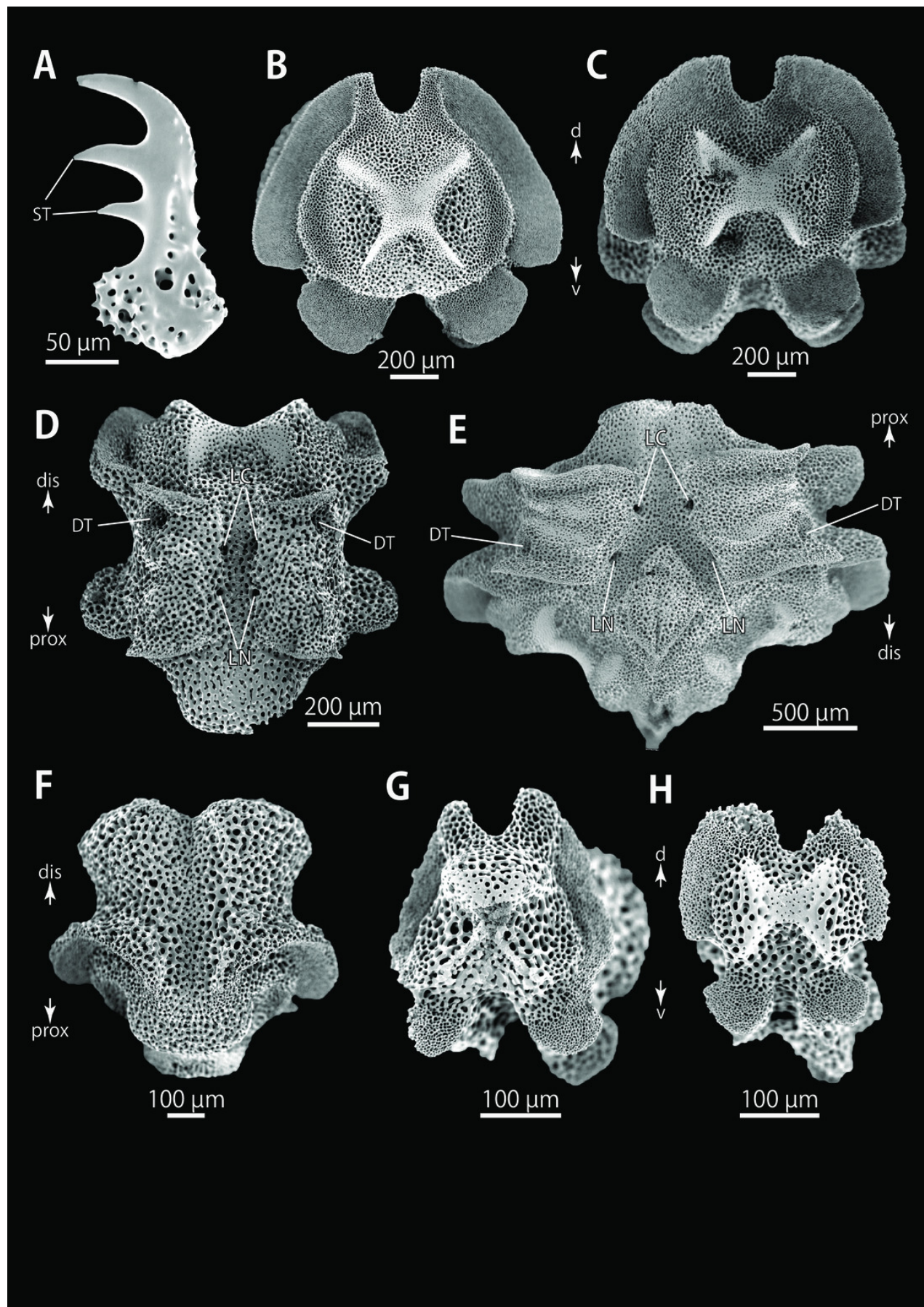


Figure 16

Figure 16. *Astrocladus exiguus* (MO-2019-19). SEM photographs of ossicles.

(A-C) Vertebrae from distal portion of arm (B is branching vertebra), dorsal (A), ventral (B, C) views. (D) A conical external ossicle on proximal portion of arm, lateral view, an arc indicates a terminal projection. Orientations: ba, basal side; dis, distal side; ex, external side; prox, proximal side. Abbreviations: DT, depression for tentacle; LC, passages for lateral canals.

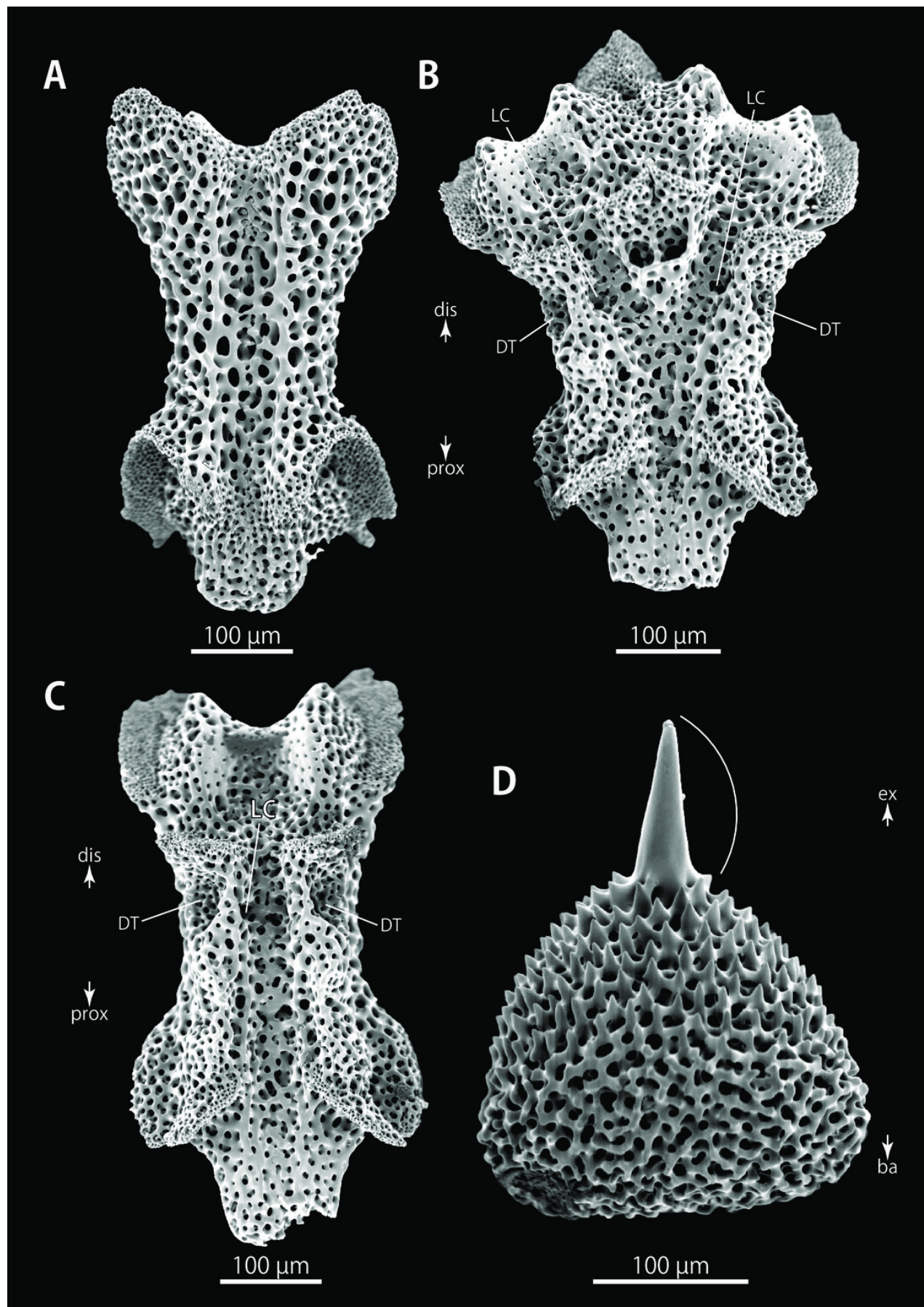


Figure 17

Figure 17. *Astrocladus annulatus*, holotype (UMUTZ-Ophi-26).

(A) Dorsal view. (B) Dorsal surface of periphery disc (C) Ventral view. (D) Jaws. (E) Interradial ventral disc. (F–H) Dorsal surface of arms, proximal (F), middle (G) and distal (H) portion of arm. (I, J) Ventral surface of arms, proximal (I) and distal (J) portion of arm. Arrowheads indicate rows of hooklets on dorsal and lateral side of the arms (B, G, H). Abbreviations: AS, arm spine; G, genital slit.

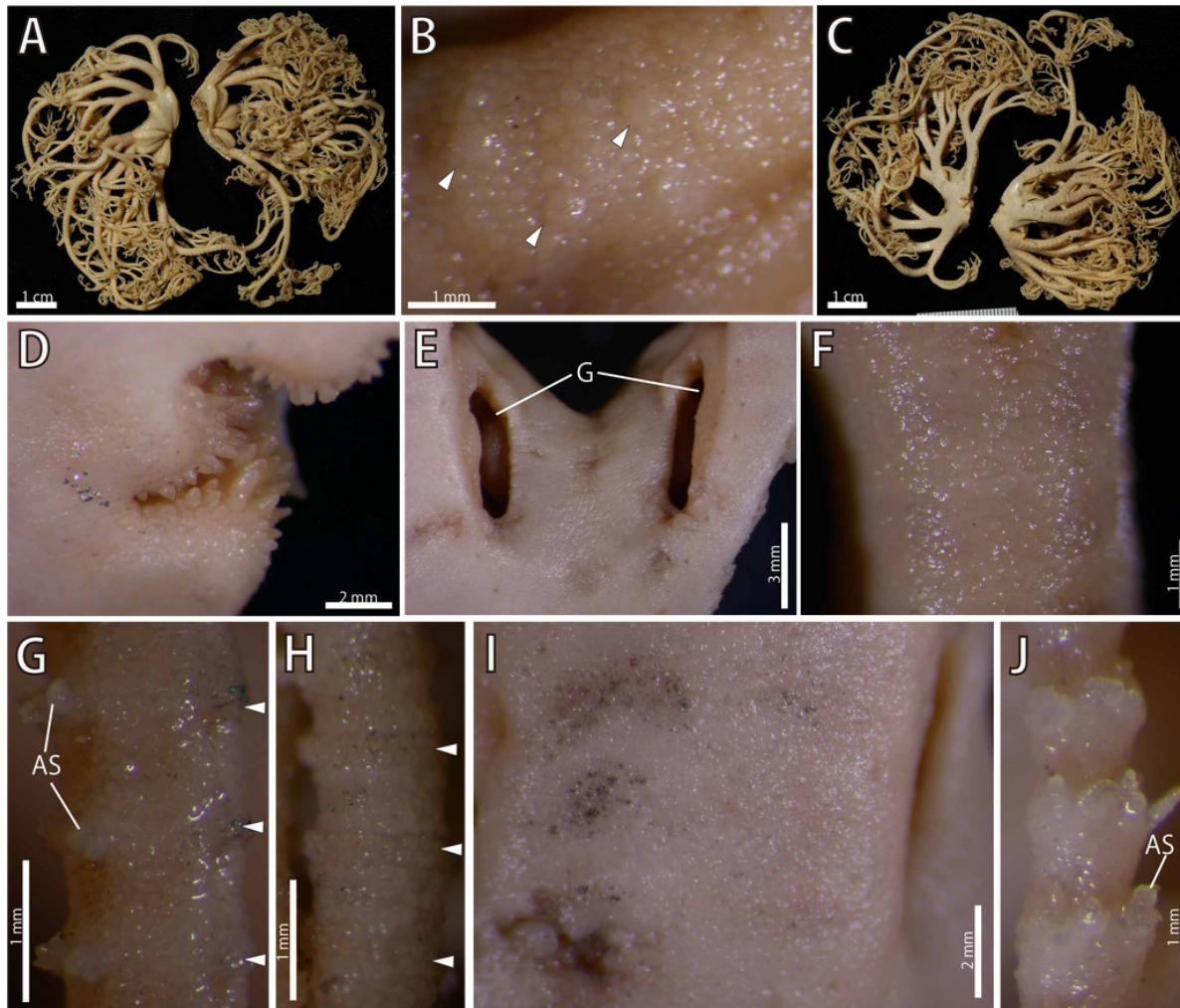


Figure 18

Figure 18. Maximum likelihood tree based on a partial sequence of mitochondrial COI gene (699 bp).

Support values for each node are shown by maximum likelihood bootstrap values (%) and Bayesian posterior probabilities. Numerals (1-5) in circles at nodes refer to the clade number discussed in the text. Bootstrap value less than 74% and Bayesian posterior probability value less than 0.97 and for each node were shown by as "-".

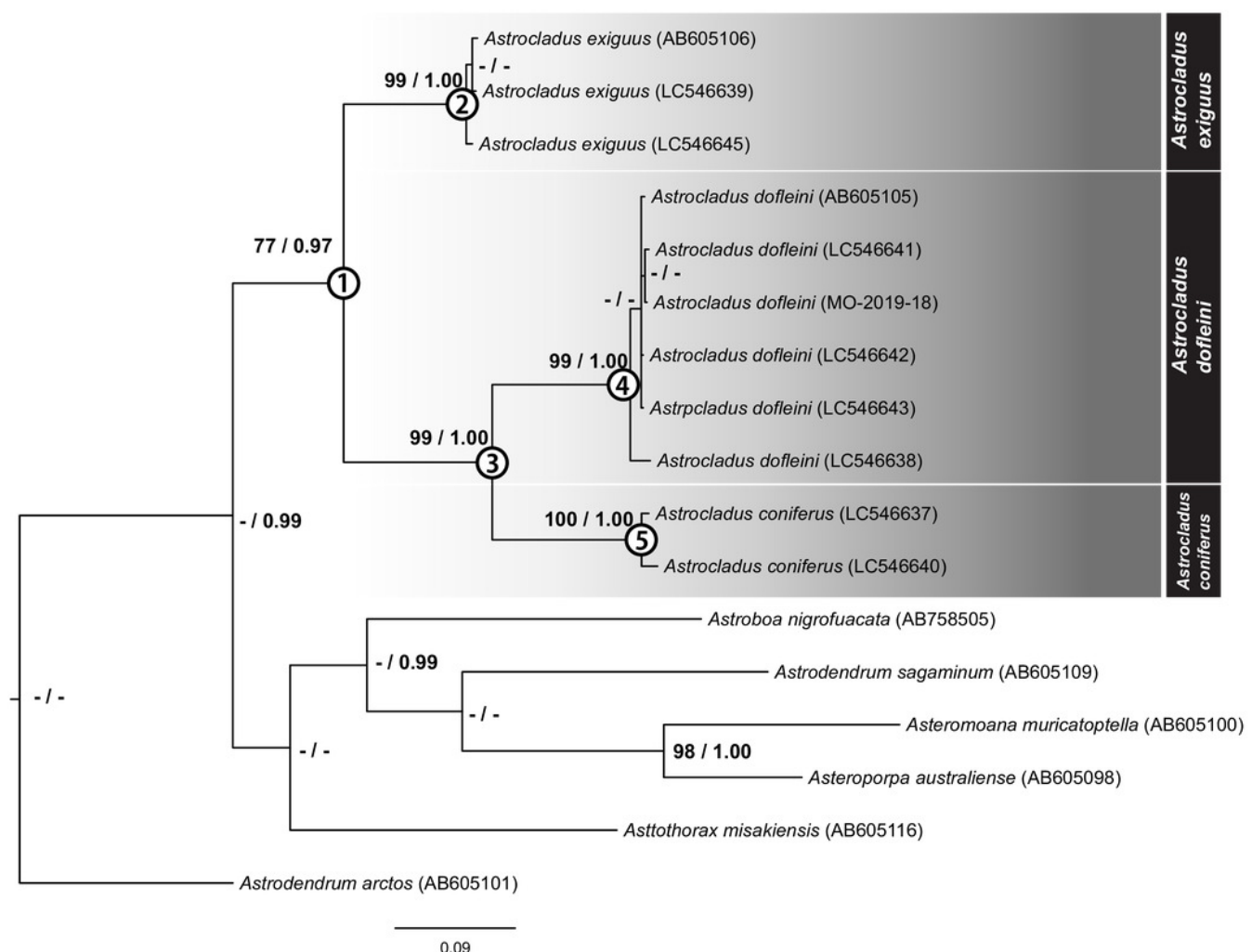


Table 1(on next page)

Table 1: Examined specimens of *Astrocladus* species including outgroup.

COI accession numbers are lodged at the DNA Data Bank of Japan. See referees for the detailed information. Unknown data are shown by “-”. Abbreviations: NSMT, the National Museum of Nature and Science, Tsukuba, Japan; UMUT, The University Museum, The University of Tokyo, Japan; ZMB, Museum für Naturkunde der Humboldt-Universität zu Berlin, Germany; ZSM, the Zoologische Staatssammlung München, Germany.

Specimen No.	Species	Catalogue. number	Locality	Depth (m)	Date of sampling	Type status	16S Access. no.	References
1	<i>Astrocladus annulatus</i>	UNUTZ-Ooh-26	Off Misaki, Sagami Bay, Kanagawa	-	-	Holotype	-	Matsumoto, 1912
2	<i>Astrocladus confierus</i>	MO-2018-118A	Koajiro Bay, Sagami Bay, Kanagawa	1.5	2018/4/26	Non type	LC546637	This study
3	<i>Astrocladus confierus</i>	MO-2019-9	Hashiraguri, Oki Island, Shimane	ca. 20	2010/7/15	Non type	LC546640	This study
4	<i>Astrocladus confierus</i>	ZSM 453/1	Kagoshima Bay, Kagoshima	ca. 30	1880/8	Lectotype	--	Döderlein, 1902
5	<i>Astrocladus pardaris</i>	ZSM 453/2	Sagami Bay, Kanagawa	-	-	Holotype	--	Döderlein, 1902
6	<i>Astrocladus dofleini</i>	MO-2019-15	Tachibana Bay, Mogi, Nagasaki	ca. 40	2019/2/7	Non type	LC546641	This study
7	<i>Astrocladus dofleini</i>	MO-2019-16	Tachibana Bay, Mogi, Nagasaki	ca. 40	2019/2/7	Non type	LC546642	This study
8	<i>Astrocladus dofleini</i>	MO-2019-17	Tachibana Bay, Mogi, Nagasaki	ca. 40	2019/2/7	Non type	LC546643	This study
9	<i>Astrocladus dofleini</i>	MO-2019-18	Tachibana Bay, Mogi, Nagasaki	ca. 40	2019/2/7	Non type	LC546644	This study
10	<i>Astrocladus dofleini</i>	NSMT E-5480	Off Minabe, Wakayama	ca. 80	2006/3/10	Non type	AB605105	Okanishi & Fujita, 2013
11	<i>Astrocladus dofleini</i>	NSMT E-10749	Off Kuji, Hitachi, Ibaraki	-	2016/9/30	Non type	LC546638	This study
12	<i>Astrocladus dofleini</i>	MO-2018-118B	Koajiro Bay, Sagami Bay, Kanagawa	15	2009/4/12	Non type	-	This study
13	<i>Astrocladus dofleini</i>	ZSM 440/1	Okinose, Sagami Bay, Kanagawa	600	1904-1905	Lectotype	-	Döderlein, 1910
14	<i>Astrocladus dofleini</i>	ZSM 440/1	Okinose, Sagami Bay, Kanagawa	600	1904-1905	Paralectotype	-	Döderlein, 1910
15	<i>Astrocladus exiguus</i>	NSMT E-6265	Off Yaku-shima Island, Kagoshima	155–170	2008/8/2	Non type	AB605106	Okanishi & Fujita, 2013
16	<i>Astrocladus exiguus</i>	MO-2019-11	Off Minabe, Wakayama	-	2012/11/22	Non type	LC546639	This study
17	<i>Astrocladus exiguus</i>	MO-2019-19	Off Minabe, Wakayama	-	2019/4/4	Non type	LC546645	This study
18	<i>Asteromoana muricatopatella</i>	NSMT E-5619B	Off Yaku-shima Isl., Kagoshima.	140	2007/9/26	Non type	AB605100	Okanishi & Fujita, 2013
19	<i>Asteropora australiense</i>	MV F99691	Wanganella Bank, New Zealand.	254–259	2003/5/28	Non type	AB605098	Okanishi & Fujita, 2013
20	<i>Astroboa arctos</i>	NSMT E-6718	Off Minabe, Wakayama	ca. 30	2009/2/26	Non type	AB605101	Okanishi & Fujita, 2013
21	<i>Astroboa nigrofurcata</i>	MNHN IE-2013-8003	Northern Pacific	143–173	1998/8/7	Non type	AB758505	Okanishi & Fujita, 2013
22	<i>Astrodendrum sagaminum</i>	NSMT E-5645	Sagami Sea, Kanagawa	681–716	2007/11/28	Non type	AB605109	Okanishi & Fujita, 2013

23	<i>Astrothorax misakiensis</i>	NSMT E-6266	Off Toshima Isl., Tokyo	266–312	2008/8/6	Non type	AB605116	Okanishi & Fujita, 2013
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