

# KEYLINK: towards a more integrative soil representation for inclusion in ecosystem scale models. I. review and model concept

Gaby Deckmyn<sup>1</sup>, Omar Flores<sup>1,2</sup>, Mathias Mayer<sup>3,4</sup>, Xavier Domene<sup>5,6</sup>, Andrea Schnepf<sup>7</sup>, Katrin Kuka<sup>8</sup>, Kris Van Looy<sup>9</sup>, Daniel P. Rasse<sup>10</sup>, Maria J.I. Briones<sup>11</sup>, Sébastien Barot<sup>12</sup>, Matty Berg<sup>13,14</sup>, Elena Vanguelova<sup>15</sup>, Ivika Ostonen<sup>16</sup>, Harry Vereecken<sup>7</sup>, Laura M. Suz<sup>17</sup>, Beat Frey<sup>18</sup>, Aline Frossard<sup>18</sup>, Alexei Tiunov<sup>19</sup>, Jan Frouz<sup>20</sup>, Tine Grebenc<sup>21</sup>, Maarja Õpik<sup>16</sup>, Mathieu Javaux<sup>7,22</sup>, Alexei Uvarov<sup>19</sup>, Olga Vinduškova<sup>1</sup>, Paul Henning Krogh<sup>23</sup>, Oskar Franklin<sup>24,25</sup>, Juan Jiménez<sup>26</sup> and Jorge Curiel Yuste<sup>27,28</sup>

- <sup>1</sup> Department of Biology, Plants and Ecosystems (PLECO), Universiteit Antwerpen, Antwerpen, Belgium
- <sup>2</sup> Biogeography and Global Change, National Museum of Natural Sciences-Spanish National Research Council (MNCN-CSIC), Madrid, Spain
- <sup>3</sup> Institute of Forest Ecology, Department of Forest and Soil Sciences, University of Natural Resources and Life Sciences (BOKU), Vienna, Austria
- <sup>4</sup> Biogeochemistry Group, Forest Soils and Biogeochemistry, Swiss Federal Research Institute WSL, Birmensdorf, Switzerland
- <sup>5</sup> CREAM, Cerdanyola del Vallès, Spain
- <sup>6</sup> Universitat Autònoma de Barcelona, Cerdanyola del Vallès, Spain
- <sup>7</sup> Agrosphere Institute, IBG, Forschungszentrum Jülich GmbH, Jülich, Germany
- <sup>8</sup> Institute for Crop and Soil Science, Julius Kühn-Institut (JKI), Braunschweig, Germany
- <sup>9</sup> OVAM, Flemish Institute for Materials and Soils, Mechelen, Belgium
- <sup>10</sup> Department of Biogeochemistry and Soil Quality, Norwegian Institute of Bioeconomy Research (NIBIO), Aas, Norway
- <sup>11</sup> Departamento de Ecología y Biología Animal, Universidad de Vigo, Vigo, Spain
- <sup>12</sup> Institute of Ecology and Environmental Sciences, IRD, UPEC, CNRS, INRA, Sorbonne Université, Paris, France
- <sup>13</sup> Department of Ecological Science, Vrije Universiteit Amsterdam, Amsterdam, Netherlands
- <sup>14</sup> Groningen Institute for Evolutionary Life Sciences, University of Groningen, Groningen, Netherlands
- <sup>15</sup> Forest Research, Alice Holt Lodge, Farnham, UK
- <sup>16</sup> Institute of Ecology and Earth Sciences, University of Tartu, Tartu, Estonia
- <sup>17</sup> Comparative Plant and Fungal Biology, Royal Botanic Gardens, Kew, London, UK
- <sup>18</sup> Forest Soils and Biogeochemistry, Swiss Federal Research Institute WSL, Birmensdorf, Switzerland
- <sup>19</sup> A.N. Severtsov Institute of Ecology and Evolution RAS, Moscow, Russia
- <sup>20</sup> Institute for Environmental Studies, Charles University, Prague, Czech Republic
- <sup>21</sup> Slovenian Forestry Institute, Ljubljana, Slovenia
- <sup>22</sup> Earth and Life Institute, UCLouvain, Louvain-la-Neuve, Belgium
- <sup>23</sup> Department of Bioscience, Aarhus University, Silkeborg, Denmark
- <sup>24</sup> Department of Forest Ecology and Management, Swedish University of Agricultural Sciences, Umeå, Sweden
- <sup>25</sup> International Institute for Applied Systems Analysis IIASA, Laxenburg, Austria
- <sup>26</sup> Department of Biodiversity Conservation and Ecosystem Restoration, ARAID/IPE-CSIC, Jaca, Spain
- <sup>27</sup> BC3-Basque Centre for Climate Change, Scientific Campus of the University of the Basque Country, Bilbao, Bizkaia, Spain
- <sup>28</sup> IKERBASQUE, Basque Foundation for Science, Bilbao, Spain

Submitted 11 February 2020

Accepted 27 July 2020

Published 9 September 2020

Corresponding author

Gaby Deckmyn, gaby.

deckmyn@uantwerpen.be

Academic editor

Robert Björk

Additional Information and  
Declarations can be found on  
page 41

DOI 10.7717/peerj.9750

© Copyright

2020 Deckmyn et al.

Distributed under

Creative Commons CC-BY 4.0

**OPEN ACCESS**

## ABSTRACT

The relatively poor simulation of the below-ground processes is a severe drawback for many ecosystem models, especially when predicting responses to climate change and management. For a meaningful estimation of ecosystem production and the cycling of water, energy, nutrients and carbon, the integration of soil processes and the exchanges at the surface is crucial. It is increasingly recognized that soil biota play an important role in soil organic carbon and nutrient cycling, shaping soil structure and hydrological properties through their activity, and in water and nutrient uptake by plants through mycorrhizal processes. In this article, we review the main soil biological actors (microbiota, fauna and roots) and their effects on soil functioning. We review to what extent they have been included in soil models and propose which of them could be included in ecosystem models. We show that the model representation of the soil food web, the impact of soil ecosystem engineers on soil structure and the related effects on hydrology and soil organic matter (SOM) stabilization are key issues in improving ecosystem-scale soil representation in models. Finally, we describe a new core model concept (KEYLINK) that integrates insights from SOM models, structural models and food web models to simulate the living soil at an ecosystem scale.

**Subjects** Ecology, Ecosystem Science, Soil Science

**Keywords** Soil fauna, Model, Soil organic matter (SOM), Hydrology, Pore size distribution (PSD), Soil biota, Ecosystem

## INTRODUCTION

Soils are multi-scale complex systems that contribute to several ecosystem services such as food, fibre and fuel production, carbon sequestration or water regulation (*Adhikari & Hartemink, 2016*). Soil quality has many definitions (see review by *Bünemann et al. (2018)*) which however all agree on the interconnected importance of soil organic matter (SOM) and soil structure for soil functioning. Soil quality can decline rapidly in response to disturbance and management changes such as industrial and agricultural activities, deforestation, overgrazing, pollution, and overexploitation for fuelwood (*Oldeman, Hakkeling & Sombroek, 1991*, see also review by *Gregory et al. (2015)*). Soil can sometimes recover fast (*Hirsch et al., 2017*), but mismanagement can limit its regeneration and buffering capacity resulting in limited recovery after soil degradation (see review by *Gomiero (2016)*). Decline of SOM threatens soil functioning in many ways affecting soil fertility, productivity, and food security, as well as the stabilization or reduction of atmospheric CO<sub>2</sub> levels (*Gobin et al., 2011*).

The full impact of a range of management or environmental changes on soil and ecosystem functioning can only be predicted with mechanistic models in which key mechanisms are sufficiently represented (*Vereecken et al., 2016*). Mechanistic models allow us to integrate our knowledge of the soil system gained from numerous experiments and also to test its current level as reflected by the models' predictive capability.

For stand/ecosystem predictions, a very limited number of soil models are usually used, mainly based on CENTURY (*Parton et al., 1987; Paustian, Parton & Persson, 1992*), RothC (*Jenkinson & Rayner, 1977*), and few others (*Liski et al., 2005*; review: *Campbell & Paustian, 2015*) although a wide range of other models have been developed (*Manzoni & Porporato, 2009; Reichstein et al., 2003; Sándor et al., 2017*). Essentially, most models describe the soil as consisting of homogeneous horizons, where SOM transformation occurs in a cascade from easily degradable to passive or stable SOM based on its chemical complexity/degradability. Equations are based on first-order kinetics (depending on pool size) where decay-rate constants are controlled by the initial litter quality (mostly represented as CN ratio or recalcitrance) and modified by temperature (t) and humidity (h) (*Liski et al., 2005; Parton et al., 1987; Tupek et al., 2019*). This representation can adequately be parameterised to simulate a stable soil under unchanging conditions, but cannot explain differences in functioning between soils concerning C and nutrient cycling, plant nutrition and hydrological processes, nor represent changes due to climate, management or pollution (*Vereecken et al., 2016*). It is also more representative of well-mixed arable lands than of natural soils that have developed horizons, though models have been developed that simulate soil horizons (*Aitkenhead et al., 2011*).

Recently, research on SOM dynamics has made substantial progress by new conceptual approaches and methodological developments, for example, biogeochemical and physical analyses, molecular and microbial ecology, and novel visualization tools. *Vereecken et al. (2016)* reviewed key soil processes and existing models, covering different scales and the perspective of a wide range of soil science disciplines. They clearly demonstrate the need to include the contributions of the different ecological compartments involved in SOM dynamics, for example, microbes and fauna, and a revised and more realistic representation of SOM degradability and pools in order to obtain a wider understanding of the soil but they do not include a review of how the soil biota influence the soil.

The role of different functional traits and functional groups of soil biota has been described in the literature. For instance, *Schmidt et al. (2011)* highlighted the importance of the microbial biomass as key factor in SOM turnover and stabilization and *Deckmyn et al. (2014)* review the role of mycorrhizal fungi and how to model them. In addition, *Filser et al. (2016)* argued for the importance of including some representation of soil fauna in soil carbon models. The most important aspect appears to be the activity of ecosystem engineers such as earthworms, ants and termites (see also review by *Lavelle et al. (2016)*). Soil engineers not only incorporate plant residues into the soil and mix up soil layers (bioturbation) but also change the soil structure by creating biopores and biostructures (e.g., casts, aggregates) that greatly affect soil hydrology and/or the activity of other soil organisms (*Lavelle, 1997*). Furthermore, it is also increasingly evident that understanding the complexity of soil food-webs is key to determining the functioning of soil biota and their influence on SOM dynamics (*De Vries et al., 2013*).

The importance of soil structural modifications on SOM stabilization mediated by soil biota has stimulated the development of models including the explicit representation of structural effects on SOM, which improve predictive capacity without explicit representation of soil fauna (*Kuka, Franko & Rühlmann, 2007*). *Komarov et al. (2017)* and

*Chertov et al. (2017a, 2017b)* recently proposed a new mechanistic soil model which incorporates many of these ideas (ROMUL), which however, is quite complex and requires very detailed parameters and measurements.

In this manuscript, we aimed to review the main insights in soil science from different disciplines, with special emphasis on the role of soil biota as a major factor influencing soil C and N dynamics, as well as soil structure and hydrology. We discuss those key processes that can be included in ecosystem models in a mechanistic way. To that end, we review the latest knowledge of key soil processes in terms of chemical SOM concepts, more structurally based concepts, insights into the fine root and mycorrhizal fungal interactions, as well as the key soil faunal actors and how they interact in the soil food web, at a stand-scale. We assess existing models for nutrient (mainly nitrogen, N) and water availability to plants, as well as soil C sequestration and leaching. Finally, we propose a new model concept by extracting the most relevant processes and the minimal community complexity required to understand and predict the overall functioning of the soil concerning C and nutrient cycling and hydrological functioning. Prediction of the faunal food web or microbial biomass is not the goal of this model concept, but a means to improve predictions of soil C and nutrient cycling and hydrology, as well as our understanding of soil functioning in relation to climate change and management.

#### **Recommended literature:**

Soil health and degradation: *Gomiero (2016); Gregory et al. (2015)*

Importance of soil fauna: *Lavelle et al. (2016); Filser et al. (2016)*

Importance of Mycorrhizal fungi: *Deckmyn et al. (2014)*

Soil modelling: *Vereecken et al. (2016)*

#### **Reviewing key pools, processes and existing models**

##### **Survey methodology**

At present, there are different “schools” for representing SOM turnover and stabilization, with many overlapping views. We review the main concepts from all three main—chemical, physical and biological—aspects to ensure a comprehensive and unbiased approach: (1) The SOM pools-view, depicting SOM pools and their chemical characteristics as the central part of the soil (with structural and microbial effects as secondary determinants), (2) The soil structure view, emphasizing the soil structure and the role of the soil engineers thereon as the main determinant, and (3) The soil food web view, representing soil microbial and faunal food webs and their role in the flow of C and N. Finally, we will discuss the main interactions between SOM, soil structure and soil biota concerning soil aggregation, fate of earthworm casts, structural effects of soil engineers and the important interactions between fine roots, mycorrhizal fungi and SOM. This review was conducted by including leading authors from the different disciplines. Web of Science Core Collection ([www.webofknowledge.com](http://www.webofknowledge.com)), Google Scholar ([scholar.google.com](http://scholar.google.com)), ScienceDirect ([www.sciencedirect.com](http://www.sciencedirect.com)) and ResearchGate ([www.researchgate.net](http://www.researchgate.net)) were used to search for manuscripts covering soil models,

soil functioning and SOM stabilization/formation but also each of the different soil biota as well as reviews on soil hydrology and soil aggregation.

### **Review goal and limitations**

The aim of this extensive review is to identify the key processes and pools involved in soil C, N and water dynamics to form a basis for a new, integrative concept to represent soil in ecosystem models. We foresee a representation with stronger emphasis on mechanistic understanding of soil functioning (in contrast to a more empirical view that describes only the outcome of processes), which can be included in existing models to improve them. Because of the very strict relation between accessibility of SOM, structure and soil water we will also include a review on the soil water modelling.

In terms of nutrient cycling, we focus on nitrogen, because of its obvious link with SOM turnover and because nitrogen data are generally available at ecosystem scale (CN ratios of main pools such as SOM, microbial biomass or plant litter) (Cools *et al.*, 2014; Zhou *et al.*, 2019). Although many of the principles described below for N are relevant for any other element, other mechanisms such as weathering and adsorption/desorption become increasingly important for less mobile elements. Detailed models have been developed to describe these phenomena (Goddéris, Schott & Brantley, 2019; Steefel *et al.*, 2015) but are, at this moment, too complex and require too much detailed input (such as mineral composition, element composition of all soil C-pools), to be applicable at the scale we envisage for our model concept.

### **The chemical aspect: SOM pools and turnover**

Soil organic matter is derived from decomposition and transformation of plant (above- and below ground litter) and animal remains (detritus) and organic products (e.g., root exudates). The fate of SOM is primarily determined by a complex interplay of its chemical properties, the composition and activities of soil organisms, abiotic conditions, and different stabilization mechanisms in soil (see review by Stockmann *et al.* (2013) and Paul (2016)). Due to its mobility, particularly the dissolved form of soil organic matter (DOM) is important for the C and nutrient transport in and between ecosystems and for the contribution to soil forming processes (review: Kalbitz *et al.*, 2000, Kaiser & Kalbitz, 2012). Traditional and more recent perspectives on SOM turnover and its incorporation in SOM models is presented in this and the following section.

### **Classic chemical perspective of SOM**

Traditional soil biogeochemical models used in ecosystem models such as RothC (Jenkinson & Rayner, 1977), CENTURY (Parton *et al.*, 1987; Paustian, Parton & Persson, 1992) or Yasso (Liski *et al.*, 2005; Tuomi *et al.*, 2011) define SOM as a number of cascading pools with different intrinsic decomposition rates. Intrinsic decomposition rates can usually be associated with pools having specific chemical and physical properties, and are modified by abiotic parameters such as temperature and moisture (Liski *et al.*, 2005; Dungait *et al.*, 2012). Such models are good at describing the decay of litter and have been well validated with data derived from litter-bag studies (Liski *et al.*, 2005). While pools associated with labile, easy degradable compounds (e.g., sugars) have a fast decay, pools

associated with lignified compounds have a slow decay (Taylor, Parkinson & Parsons, 1989; see review by Krishna & Mohan, 2017). Several models assume SOM pools associated with the most recalcitrant compound groups (e.g., humic substances and lignin) and chemically protected (e.g., SOM-clay complexes) account for the long-term stabilization of organic matter in soil (Segoli et al., 2011; Tuomi et al., 2011; Smith et al., 1997).

However, the concept of long-term SOM stabilization due to chemical recalcitrance has increasingly been questioned (Schmidt et al., 2011; Dungait et al., 2012; Cotrufo et al., 2013; Lehmann & Kleber, 2015). There is a growing evidence showing that the formation of stable SOM is largely independent from molecular properties (Kleber et al., 2011; Schmidt et al., 2011; Lehmann & Kleber, 2015). Modern analytical methods could not prove humic substances to be persistent in soil (Schmidt et al., 2011; Lehmann & Kleber, 2015). It rather seems that SOM is a continuum of decomposing substances and even recalcitrant humic compounds can decay rather quickly (Lehmann & Kleber, 2015). In fact, it is increasingly accepted that chemical recalcitrance is primarily important in early stages of litter decomposition (review: Von Lützow et al., 2006, Marschner et al., 2008). Decay rates of plant litter for example, are usually inversely related to their lignin to N ratios, suggesting slow decomposition at high lignin contents (Melillo, Aber & Muratore, 1982; Zhang et al., 2008; Prescott, 2010).

Recent studies have furthermore highlighted that microbial products from the transformation of plant litter rather than plant litter per se are the largest contributors to stable SOM (Mambelli et al., 2011; Cotrufo et al., 2013; Gleixner, 2013).

#### **Recommended literature:**

Litter decomposition: Krishna & Mohan (2017)

Stability of SOM in relation to ecosystem: Schmidt et al. (2011); Von Lützow et al. (2006)

#### **The importance of dissolved organic matter (DOM) and nutrients**

Another key element of the SOM dynamics and the soil C and N cycle is the dissolved organic matter (DOM). The incorporation of DOM into microbial biomass is now considered an important pathway of stable SOM formation (Sokol & Bradford, 2019). Most DOM is derived from litter and humus degradation (see review by Kalbitz et al., 2000; Guggenberger & Kaiser, 2003). Recent studies show that while subsurface DOM is linked to recent plant material, DOM in deeper layers consists of older, more processed substrates, mainly derived from microbial turnover (Kaiser & Kalbitz, 2012).

Besides decaying litter and microbial turnover, direct exudation from plant roots (rhizodeposition) can be an important source of DOC in the soil (on average 5% but up to 20% of photosynthates in grassland species) (Pausch & Kuzyakov, 2018)), with very important effects on the surrounding zone. Recent evidence suggests that these belowground DOC inputs may be even more likely to contribute to the formation of stable SOM than litter derived DOM (Sokol & Bradford, 2019).

Because DOM can leach from soils and can move between soil layers, it is important to model DOM separately. A number of models such as LIDEL (Campbell et al., 2016) include the explicit simulation of DOM. A detailed dynamic model (DyDOC) for

predicting metabolic transformations of SOM components and the transport and sorption of DOM in different soil horizons with different soil properties was developed and tested by *Tipping et al. (2001, 2012)*, though it does not include soil biology. DOC can be controlled by sorption to minerals and co-precipitation with Fe, Al or Ca, all governed by the soil acidity (*Guggenberger & Kaiser, 2003*). For this reason, mineral weathering rate should be considered in the models predicting DOC solubility.

In general, the pathways, sorption and desorption processes of the different compounds of DOM and nutrients like phosphorus are extremely complex, and as such hard to include in a simple soil model. There are detailed surface complexation and ion-exchange models which deal with these processes (*Weng, Van Riemsdijk & Hiemstra, 2008; Duputel et al., 2013*). Models for soil weathering and for adsorption processes that ultimately explain the soluble nutrients available to plants exist, but are complex and require many parameters (for example, PhreeqC, *Parkhurst & Appelo, 2013*). In *Bortier et al. (2018)* a relatively simple empirical model within the soil model ANAFORE is used to distinguish adsorbed and soluble P based on pH, without concretely simulating different base cations. *Dzotsi et al. (2010)* developed a more complex model for P availability that goes beyond the scope of this paper as it requires extensive parameterization. The approaches to phosphorus modelling in ecosystem models have been comprehensive reviewed by *Pferdmenges et al. (2020)*.

Nitrogen is considered to be the most limiting nutrient for soil organisms and besides being part of SOM (and DOM), it is present in soil also in dissolved mineral forms ( $\text{NH}_4^+$ ,  $\text{NO}_3^-$ ,  $\text{NO}_2^-$ ) which are the primary source of nitrogen for plants, but also for the potential N losses via leaching or denitrification. The different approaches to modelling soil N dynamics have been reviewed by *Manzoni & Porporato (2009)*.

Each soil type has an associated a distinct physicochemical environment and development pathway of the soil profile, which affects the chemical composition and stability of SOC in mineral horizons (*Rumpel, Eusterhuesa & Kögel-Knabner, 2004; Rumpel & Kögel-Knabner, 2011*), by affecting both the living conditions and activity of soil decomposers but also through a distinctive physical and chemical protection. One of the main soil forming processes involved in chemical SOM stabilization especially in deep mineral soils is the “podzolization” which involves a transport of DOM, Al and Fe in solution from the surface to deeper horizons. The process consists of mobilization and immobilization of these compounds (*Lundström et al., 2000*). General conditions that favor podzolization are the absence of sufficient neutralizing divalent cations due to the presence of parent materials with low amounts of weatherable minerals ( $\text{Ca}^{+2}$ ,  $\text{Mg}^{+2}$ ), an impeded decomposition of plant litter due to low temperatures and high rainfall conditions that favor the transport of DOC (along with Al/Fe) down the profile (*Van Breemen & Buurman, 2002*). Moreover, the nutrient-poor status and high acidity typical of this soil type tends to decrease faunal activity which subsequently impedes vertical mixing of the soil and favors vertical differentiation and accumulation of partially decomposed plant residues in organic horizons (*Van Breemen et al., 2002; Rumpel, Kögel-Knabner & Bruhn, 2002*).

Although few studies have compared C stability among different soil types, some of them suggest that stabilization processes may be soil-type specific and therefore depend on pedogenic processes (*Rumpel, Eusterhuesa & Kögel-Knabner, 2004; Rumpel & Kögel-Knabner, 2011*). However, representation of pedogenic processes such as weathering or podzolization in mechanistic models is rare (*Minasny, Sulaeman & McBratney, 2011*) and may be relevant only for longer time scales.

**Recommended literature:**

DOC: *Kalbitz et al. (2000)*

N mineralization models: *Manzoni & Porporato (2009)*

**The physical aspect: new perspectives in SOM and water dynamics**

**Structural perspective on accessibility of SOM and diffusivity of water through pore space**

There is a close interaction between soil structure, SOM, water/gas balance, and the size and connectivity of pores as ecological habitats in soil. Recent advances in our understanding of SOM stabilization show that patterns of spatial inaccessibility against decaying soil organisms, or stabilization by interaction with mineral surfaces and metal ions (review: *Von Lützow et al., 2006*) seem to play a more important role in long term stabilization of SOM than chemical recalcitrance (*Dignac et al., 2017; Dungait et al., 2012*). These studies show that the main stabilization mechanisms that protect SOM from decomposition are physical protection by soil macro-(250–2,000  $\mu\text{m}$ ) and micro-(53–250  $\mu\text{m}$ ) aggregate formation and chemical protection associated with silt and clay particles and Fe- and Al- oxides (*Dwivedi et al., 2017*). The accessibility of SOM to microbes due to pore size and the capacity of microbes to oxidize SOM based on the strength of the organo-mineral associations are two different mechanisms involved in SOM stabilization and SOM dynamics. However, the separation of OM occluded in clay microstructures from “true” organo-mineral associations remains a methodological challenge (*Chenu & Plante, 2006; Von Lützow et al., 2008; Yudina & Kuzyakov, 2019*). Until this is possible, it might be possible to view organic matter stabilized in organo-mineral associations as in such close contact to the mineral that there is no space for microbes and microbial exoenzymes to physically reach the OM.

It can therefore be argued that the most important mechanism for SOM stabilization over longer time scales is the physical separation of organic compounds from the organisms able to degrade or transform them, for example, in anoxic or dry pore space areas or within aggregates and that this applies also to organo-mineral associations. (see review by *Von Lützow et al., 2008*). Soil structure and its dynamics are thus the most important factors controlling SOM turnover and sequestration, whereas chemical recalcitrance is only a secondary determinant (*Dungait et al., 2012*).

Soil structure also determines the soil water dynamics. Water is essential for all soil processes (chemistry, biology, physical transport of DOM and nutrients).

Water availability or water activity in soil is limited by water potential, which in soil is mainly controlled by the adhesion forces to solid particles (matric potential), which,



together with the cohesion forces between water molecules, drives capillarity. Water matrix potential is considered to be a major controlling factor of SOM turnover and microbial activity (Thomsen *et al.*, 1999). It affects the physiology of microorganisms and many critical mass transfer processes in the pore space: diffusion of soluble organic matter, exoenzymes and gasses, and motility of microbes (see review by Or *et al.* (2007)). These mass transfer processes can limit microbial access to organic matter at low water contents and, as a consequence, affect its turnover rate (Allison, 2005; Stark & Firestone, 1995). The physical separation of habitats at low water contents is likely what supports the vast diversity of soil microorganisms (Tecon & Or, 2017) as organisms have developed different strategies to mitigate the effect of these barriers (Allison, 2005; Mills, 2003; Torsvik & Ovreas, 2002). In turn, microorganism activities may stabilize (Six *et al.*, 2004) or destabilize aggregates and affect soil porosity (Crawford *et al.*, 2012; Helliwell *et al.*, 2014) or, under extensive microbial growth, may even result in pore clogging (Seki, Miyazaki & Nakano, 1998); providing a feedback to soil structural properties and consequently to SOM turnover. Many soil processes are thus closely interlinked (Six *et al.*, 2004).

#### **Recommended literature:**

Bacterial activity in soil pores: Or *et al.* (2007); Tecon & Or (2017)

Stabilisation of SOM through inaccessibility: Von Lützow *et al.* (2006, 2008)

#### **Modelling soil structural effects on SOM turnover**

Traditional ecosystem models represent physical and chemical stabilization of C in the soil as an implicit property of the most passive (inert) SOM pool and often relate it to clay content. Although clay content can be seen as a simplified proxy for both SOM stabilization mechanisms (i.e., adsorption and aggregate inclusion) (Rühlmann, 1999, Sulman *et al.*, 2014), it is clearly not their only driver. This has motivated the development of several new models that explicitly account for stabilization mechanisms for effects of either or both mechanisms on SOM turnover (Abramoff *et al.*, 2018; see review by Stockmann *et al.*, 2013; Wieder *et al.*, 2014, 2015; Sulman *et al.*, 2018). For example chemical protection by adsorption onto mineral surfaces is dynamically represented in the COMMISSION model (Ahrens *et al.*, 2015). The Struc-C model (inspired by Roth-C), on the other hand, is more aggregate-centric but incorporates both mechanisms by assuming organo-mineral associations are the smallest aggregates and describes the interaction among organic matter and soil structure through the incorporation of aggregation and porosity submodules (Malamoud *et al.*, 2009). Stamati *et al.* (2013) built on this effort and introduced another Roth-C based model CAST that simulates macro- and micro-aggregate formation and the stabilization of particulate organic matter. Abramoff *et al.* (2018) proposed to model mineral-associated organic matter (MAOM) and aggregate C as two separate measurable pools but did not actually propose how would they be analytically distinguished. Despite these advances, aggregate formation modelling remains a difficult issue at the stand scale because many of the processes occur at a much smaller scale (Yudina & Kuzyakov, 2019).

A different, more pore-based approach was introduced in the CIPS model (*Kuka, Franko & Rühlmann, 2007*) which modified the classic empirical SOM pools taking into account soil structure effects. It is based on a quality-driven primary stabilization mechanism (recalcitrance of SOM) and a process-driven secondary stabilization mechanism (site of turnover) of SOM in soil. In addition to the division of SOM into the qualitative pools on the basis of chemical measurability, it takes into account different turnover conditions depending on pore space and accessibility for microbial biomass. The main assumption of the CIPS model is that the biological activity is not evenly distributed through the whole pore space. The pore space classes (i.e., micro-, meso- and macropores) used in the model are marked by wilting point, field capacity and pore volume. Because of the poor aeration in the micropores they show very low biological activity, leading to a strong protection of the C localized in this pore space. This results in the reduction of the turnover activity, related to soil t, h, soil texture, relative air volume and distance to the soil surface. Simulation results show that the bulk density variations have a severe impact on C storage (*Kuka, Franko & Rühlmann, 2007*). Besides a validation of the CIPS model for longterm experiments representing a wide range of soils and site conditions, (*Kuka, Franko & Rühlmann, 2007*) show that the conceptual pool of inert SOM (used in many models) can also be described as the amount of C situated in micropores. Consequently this new approach seems more generally applicable than the soil texture based approaches applied so far.

**Recommended literature:**

Modelling soil structural effects on SOM: *Kuka, Franko & Rühlmann (2007)*;  
Modelling SOM effects on soil structure: *Malamoud et al. (2009)*;  
C sequestration modelling: *Stockmann et al. (2013)*

**Modelling soil hydrology and structure**

A large number of soil models of varying levels of complexity and dimensionality are now available to describe the basic physical and chemical processes affecting water flow and solute transport in the subsurface environment (*Vereecken et al., 2016*).

Many models that describe the soil-plant-atmosphere continuum still use simple capacity based soil water flow models to quantify the terms of the water balance (*Deckmyn et al., 2008*; *Farmer, Sivapalan & Jothityangkoon, 2003*; *Teshima et al., 2006*). The main motivation for using these capacity based models is their simple parameterization (*Romano, Palladino & Chirico, 2011*). They describe water flow in soils as mainly driven by gravitational forces where each soil layer spills over to the lower soil compartment once a critical soil moisture content has been reached (spilling bucket models). This critical soil moisture content is often defined as field capacity and is routinely measured in soil surveys. Soil water storage capacity of a specific compartment can be thus emptied by downward flow, surface runoff, deep drainage, and evapotranspiration processes. Since gravitation is the dominant potential controlling water flow, specific parameterization needs to be included in order to account for capillary rise from a groundwater table into the root zone and lateral flow processes (*Guswa, Celia & Rodriguez-Iturbe, 2002*).

However, this method tends to overestimate soil water in the top layer and underestimate drainage.

More advanced soil models nowadays use Richards equation and the convection-dispersion equation (Jury & Horton, 2004) to describe water and solute movement through soil. Soil models describing water flow based on Richards equation provide more flexibility in incorporating the full complexity of water flow in the soil-plant-atmosphere continuum and its impact on spatially distributed abiotic and biotic processes, including capillary rise, though at a high computational cost (Kuraz, Mayer & Pech, 2014). Many of these processes are characterized by a large spatial and temporal variability with locally distributed hot spots and hot moments. However, these more advanced 3D features are harder to parameterize. To address parameterization difficulties, pedo transfer functions (PTFs) have been developed that allow predicting soil properties and soil parameters that control abiotic and biotic processes. Soil horizons, texture, qualitative structural and morphological information, organic matter content, pH, redox and mineral concentrations are soil properties that can be used in PTFs to quantify soil properties and gain information on functions (e.g., soil hydraulic functions, mineralization constants, sorption properties and ecosystem functions such as providing water and nutrients to plants and regulating biogeochemical cycles) (Bouma, 1989; McBratney et al., 2001; see review by Vereecken et al. (2016) and Van Looy et al. (2017)).

The presence of macropores and other structural heterogeneities can generate flow instabilities and cause preferential flow and transports (review: Jarvis, Koestel & Larsbo, 2016; Beven, 2018; Hendrickx & Flury, 2001). Due to preferential flow, water and solutes may move faster and deeper into the soil profile than what would be predicted by Richards equation, so models using this equation tend to underestimate leaching (Julich, Julich & Feger, 2017; Šimůnek et al., 2009). These macropores are in many cases the consequences of biotic processes, such as earthworms burrowing and root growth (Capowiez, Sammartino & Michel, 2014; Bastardie et al., 2002; review: Wilkinson, Richards & Humphreys, 2009). Modelling approaches for preferential and non-equilibrium flow and transport in the vadose zone were reviewed by Šimůnek et al. (2003). Extensions have been made to consider preferential flow and transport in models based on Richards equation (review: Köhne, Köhne & Šimůnek, 2009; Šimůnek et al., 2003). Yet, these models contain several uncertainties due to a lack of observational data at the pore scale and to the inherently dynamic macropore system in soils being subject to physical (swell/shrink, freeze/thaw), biological (variations in soil faunal and microbial activity, root growth, rhizosphere processes) and anthropogenic disturbances (e.g., tillage practices, Capowiez et al., 2009).

Continuous advances in both numerical techniques and computation power are now making it increasingly possible to perform comprehensive simulations of non-equilibrium flow processes in the vadose zone (review: Vereecken et al., 2019). Such simulations, especially if paired with exhaustive field data sets (e.g., by data assimilation), are vital for better understanding and quantifying the effects of heterogeneities, fractures and macropores on flow and transport at the field scale (Van Genuchten, Leij & Wu, 1999; review: Šimůnek et al., 2003).

Challenges in predicting soil water flow and solute transport beyond laboratory scale include: soil parameterization, handling structured soils including preferential flow, handling soil heterogeneity, temporally changing properties (e.g., soil bulk density, structural properties, etc.), and description of root water uptake (e.g., *Jury et al., 2011*; review: *Vereecken et al., 2019*). Thus, it is clear that although the importance of soil structure and water are proven, their inclusion in models is hampered because of the lack of data on soil structure and the difficulties in measuring and simulating soil water at the ecosystem scale.

#### **Recommended literature:**

Preferential flow: *Jarvis, Koestel & Larsbo (2016)*

Solute transport and preferential flow modeling: *Köhne, Köhne & Šimůnek (2009)*; *Šimůnek et al. (2003)*

Pedotransfer functions: *Van Looy et al. (2017)*

Infiltration: *Vereecken et al. (2019)*


Bioturbation: *Wilkinson, Richards & Humphreys (2009)*

#### **The biological aspect: the role of the soil food web**

The soil comprises a rich and very diverse community of organisms (*Bardgett, Usher & Hopkins, 2005*; *Bardgett & Van der Putten, 2014*; *Orgiazzi et al., 2016a*; *Ramirez et al., 2015*). To be able to cope with this high diversity, species can be grouped into functional groups, under the assumption that if species occur at the same location in the soil and share the same resources and predators they should perform the same function (review: *Briones, 2014*). Research has so far focused on the importance of each one of these functional groups to the ecosystem, but this highly specialised information is not integrated into the more plant-based ecosystem models (*Geisen et al., 2019*).

It has long been known that litter decay is faster in the presence of macro fauna (comparison between small and larger mesh size litterbags) (reviewed by *Frouz et al., 2015*). Also, the major roles of soil engineers for bioturbation are well described (see “Soil structural modifications by engineers”), which add to the effect of soil fauna to processes. Recent publications have shown the importance of the diversity of soil organisms in relation to soil functioning and stability, both in the laboratory and in the field (reviewed by *Deng (2012)*; *Wagg et al. (2013)*). Other studies have shown that an intact soil food web is important for ecosystem functioning because it influences key functions such as decomposition, nutrition retention and nutrient cycling (*Bengtsson, Setälä & Zheng, 1996*; *Philippot et al., 2013*). In addition, the soil food web is sensitive to management. Ploughing, soil compaction, litter removal and obviously the use of insecticides are practices that are deleterious to the soil faunal community (*Yeates et al., 1997*; *Wardle et al., 1995*), with repercussions for soil processes. Such major negative effects on soil organisms are ignored in the most widely used ecosystem models (ORCHIDEE—*Camino-Serrano et al. (2018)*, PaSim and Biome-BGC—*Sándor et al. (2016)*, ANAFORE—*Deckmyn et al. (2008)*) that thus cannot realistically simulate these management effects.

Biota	Main functions						
	micro- and mesoporosity	Macro porosity	SOM turnover	SOM input	fragmentation	Plant nutrient uptake	bioturbation
Bacteria	++		+++			***	
Fungi	++		+++		+	***	
Mycorrhizal fungi	++		++	++	+	+++	
Bacterivores	**		***			*	
Fungivores	**		***			*	
Predators	**	*	**	*	*	*	*
Engineers	++	+++	+		+++	*	+++
Detritivores			+		+++	*+	+
Fine roots	+	++		++		+++	
Herbivores		*		**	++		

**Figure 1** Importance of different functional groups of soil biota on key soil processes linked to ecosystem functioning. +, direct effects; \*, indirect effects. Full-size  DOI: 10.7717/peerj.9750/fig-1

### Recommended literature:

Role of macro-fauna: *Frouz et al. (2015)*

Role of soil fauna: *Briones (2014)*

Role of microbial community: *Deng (2012)*

### Defining and describing food web components

To develop a model that simplifies as much as possible the enormous complexity of the soil food web, it is important to review all soil biotic inhabitants and determine keystone species or functional groups within each trophic level. In the following sections, we therefore review the main soil organisms according to their size, trophic level and functional significance, that is, microorganisms (size 1–100  $\mu\text{m}$ ), microfauna (<0.1 mm), mesofauna (0.1–2 mm) and macrofauna (>2 mm), as well as fine roots (<2 mm) that are the main primary source of soil C. Simulating larger vertebrate fauna (mice, moles, rabbits, or birds) is beyond the scope of this paper. All size groups of soil fauna include organisms of different trophic level and functional significance. Nevertheless, microbivore soil fauna are usually small-sized members of micro- and mesofauna, whereas ecosystem engineers belong to the macrofauna. In this review we will classify the organisms mainly by function and food source, not by size but we describe for each functional group which organisms belong to it. All biotic effects on the main soil components necessary to simulate SOM as well as nutrient and water flows are described. [Figure 1](#) summarizes how the different functional groups impact on porosity as linked to aggregation (meso- and micropores), macroporosity, SOM turnover, nutrient availability, and C influx into the soil. Since the goal is to understand how to include these organisms in a model we also

review, where possible, data on the biomass of the group and of their contribution to the C cycle.

### Soil microorganisms

The soil microorganisms include bacteria, archaea and fungi which are the primary enzymatic degraders of organic matter ([Datta et al., 2017](#); [Johnston, Boddy & Weightman, 2016](#)) and protozoa which are microbivores strongly contributing to the control of the activities of other microorganisms (see review by [Gao et al. \(2019\)](#)). Microbial degradation activity ultimately determines both the rate at which nutrients become available to plants and the amount of C stored in soils ([Mambelli et al., 2011](#); [Cotrufo et al., 2013](#); [Gleixner, 2013](#)). Microbes are also known to induce weathering of minerals ([Uroz et al., 2009](#)). In this section, we take a functional approach and first discuss bacteria and non-mycorrhizal fungi as significant decomposers and deal with mycorrhizal fungi separately because of their other distinct functions.

### Bacteria and Archaea

Prokaryotic abundance can vary between 4 and  $20 \times 10^9$  cells  $\text{cm}^{-3}$  soil ([Bardgett & Van der Putten, 2014](#)). Several studies have shown that at least half of the soil microbial populations are respiratory active ([Lennon & Jones, 2011](#)). Bacteria were found to contribute about 35% of the total heterotrophic soil respiration ([Joergensen & Wichern, 2008](#)), and their contribution relative to fungi depends mainly on the chemical composition of the SOM.

The classic understanding about the distribution of the microorganisms (especially Bacteria and Archaea) has been that everything is everywhere ([Baas Becking, 1934](#)). However, recent studies showed that, in contrast with the classic understanding, bacterial species are biogeographically restricted due to variations in climatic, soil and plant conditions ([Bardgett & Van der Putten, 2014](#)). The common view is that there is a high functional redundancy within the soil communities for nutrient mineralization and that changes in community structure rather than changes in species richness play a role in soil and ecosystem functioning ([Bardgett & Van der Putten, 2014](#)). For most ecosystem scale purposes however, the classic understanding is adequate.

Bacteria also play a central role in the production and immobilization of inorganic and organic N. Moreover, microbial biomass contributes directly to the pool of soil organic N through its death and turnover ([Bottomley, Taylor & Myrold, 2012](#)). It is estimated that about 0.28–28  $\mu\text{g}$  N is assimilated into bacterial biomass (into protein) per g soil and per day ([Bottomley, Taylor & Myrold, 2012](#)).

Much of the organic material is degraded by microorganisms carrying out aerobic respiration. When organic matter is transported to zones in the soil where oxygen is low or inexistent, anaerobic respiration can be carried out by specialized bacteria that utilize terminal electron acceptors other than oxygen, such as nitrate, manganese, iron, sulphate or  $\text{CO}_2$  (resulting in methane emissions, the end product of  $\text{CO}_2$  reduction (review: [Dalal et al., 2008](#))). Global methane emissions reach 600 Tg  $\text{CH}_4$   $\text{year}^{-1}$  and it is estimated that water-saturated soils such as peat and rice soils contribute to about 55% of the total

methane emissions (see reviews by *Le Mer & Roger (2001)*; *Dalal et al. (2008)*). This process is however energetically less favorable and comparatively slower than aerobic respiration (*Keiluweit et al., 2017*). Recent evidence suggests that anaerobic microsites in which anaerobic respiration can take place such as insides of aggregates or peds comprise between 14% and 85% of the pore volume in well-drained soils at moderate moisture (*Keiluweit et al., 2018*). Mineralization rates in such microsites are reduced by ~90% compared to well-aerated soil compartments (*Keiluweit et al., 2017*).

Because of their size (0.3–5  $\mu\text{m}$ ), bacteria often reside in pores and inner surface of aggregates as micro-colonies of about 2–16 cells (*Gupta & Germida, 2015*). Higher colonization of bacterial cells is restricted to hot spots with higher available C, such as the rhizosphere or the outer surface of freshly formed aggregate (*Foster, 1988*). Several studies reported an influence of the physicochemical characteristics (water potential, nutrient and oxygen availability) on the ecology of the bacterial community (*Six et al., 2004*), which links well with the concepts of the structural availability of SOM as described in “Structural perspective on accessibility of SOM and diffusivity of water through pore space”.

### **Non-mycorrhizal fungi**

Fungi, are an important component of soil ecosystem functioning, especially regarding organic matter decomposition (*Van der Wal et al., 2013*). Their significance lies in their ability to degrade more recalcitrant SOM due to a combination of morphological (hyphal growth form) and physiological (extracellular enzymes) characteristics (*Van der Wal et al., 2013*).

Fungi can be of two distinct forms: spherical cells (yeasts) or long thread like structures called hyphae or mycelium (filamentous fungi). Filamentous fungi are of particular importance in terrestrial ecosystems as they allow an extended exploration of soil via their hyphal system, penetrating solid substrates (*Van der Wal et al., 2013*). Hyphae are also very efficient in the translocation of water by bridging air-filled pores (*Curiel Yuste et al., 2011*) and by bridging nutrient-poor patches by which they supply growth limiting elements to zones of metabolic activity (*Frey et al., 2000*; *Gupta & Germida, 2015*). Their average abundance has been estimated at 100 m of hyphal length per gram of soil (*Bardgett & Van der Putten, 2014*). It has also been estimated that about 1.3–10.9  $\mu\text{g}$  of fungal biomass is formed per g soil per day, corresponding to about 0.06–0.48  $\mu\text{g}$  N immobilized in to fungal biomass (*Bottomley, Taylor & Myrold, 2012*). Filamentous fungi are fundamental to C decomposition of terrestrial organic matter (review: *Geethanjali & Jayashankar, 2016*); it has been estimated that fungal respiration can account for up to 65% of the total microbial soil respiration (*Joergensen & Wichern, 2008*).

### **SOM mineralization: bacteria versus non-mycorrhizal fungi**

The ratio of fungal to bacterial biomass is highly variable (between 0.007 and 0.34) among different biomes (*Fierer et al., 2009*; *De Vries et al., 2006*). Generally, forest ecosystems have a higher fungal to bacterial (F:B) biomass ratio than grasslands. *Fierer et al. (2009)* found particularly high F:B in temperate coniferous forest soils, whereas deserts and grasslands had the lowest ratio.

Land-use changes, and agricultural intensification have been shown to shift a fungal-dominated to a bacterial-dominated food web (*De Vries et al., 2006*). For example, in a study comparing the resistance and resilience of the soil food web to drought, the fungal-based food-web of an extensively managed grassland and the processes of C and N it governs was more resistant to drought than the bacterial-based food web of an intensively managed wheat field (*De Vries et al., 2013*). Modelling of these two systems revealed that the fungal-based network had a greater evenness that mitigated C and N loss, which made the system more adaptable to drought than the bacterial-based food-web (*De Vries et al., 2013*).

Through evolution, bacteria and fungi have undergone niche differentiation in the decomposition of organic materials. Fungi have higher biomass C:N, broader enzymatic capabilities, and slower biomass turnover rates (*Waring, Averill & Hawkes, 2013*). Typically, fungal hyphae are better adapted to nutrient-poor niches in soil than bacteria because they can search for the heterogeneously distributed nutrient resources (review: *De Boer et al., 2005*). A classic view is that during evolution of terrestrial microbial life, fungi have become specialists in decomposing structurally complex organic matter, such as lignin (recalcitrant litter and SOM), while bacteria, on the other hand, have been able to maintain a significant role in the degradation of simple substrates (review: *De Boer et al., 2005*). However, fungi and bacteria compete for both complex and simple substrates (*Johnston, Boddy & Weightman, 2016*), especially for limiting nutrients such as N (*Bottomley, Taylor & Myrold, 2012*).

As mentioned in the DOM section, plant roots exude substantial amounts (up to 20–40% of their photosynthetically fixed C) of simple and easily degradable organic molecules (see reviews by *Badri & Vivanco (2009)* and *Canarini et al. (2019)*). Classically, due to the high abundance of bacteria in the rhizosphere, it was assumed that these easily degradable compounds were almost exclusively degraded by bacteria (*Jones, 1998*). However, stable isotope probing has revealed that a significant part is also degraded by fungi (*Treonis et al., 2004*). It has also been shown that fungi are the most active group in the degradation of easily degradable compounds in acid soils (*Rousk, Brookes & Bååth, 2009*) and at high substrate loading rates, probably due to their superior osmotic stress tolerance (*Griffiths et al., 1998*).

The degradation of cellulose, the most abundant organic compound on earth (30–50% of plant dry mass), can take place in both aerobic and anaerobic conditions. Aerobic cellulose degradation is widespread within the fungal and bacterial communities (review: *De Boer et al., 2005*, review: *Baldrian & Valášková, 2008*). Both aerobic bacteria and fungi produce hydrolytic enzymes, which convert cellulose into glucose (*Mansfield & Meder, 2003*). Competition for cellulose between fungi and bacteria is high (review: *De Boer et al., 2005*; *Johnston, Boddy & Weightman, 2016*). However, it is considered that most of the cellulose is degraded by fungi, because their hyphal growth strategy is better suited to access the cellulose fibres, which are often embedded in a matrix of other structural polymers, such as hemicellulose and lignin (*Van der Wal et al., 2013*). Contrastingly, in anoxic environments, some bacteria containing cellulosomes that allow enzyme activity to



take place directly in their cell are almost exclusively responsible for the cellulose degradation (Lynd *et al.*, 2002).

Lignin degradation is largely, but not exclusively, done by white-rot fungi (Leonowicz *et al.*, 1999) though ligninolytic capabilities have also been reported for Proteobacteria (Bandounas *et al.*, 2011; Tian *et al.*, 2014) and Actinobacteria (Abdel-Hamid, Solbiati & Cann, 2013). The decomposition of lignin needs specialized enzymes (Bödeker *et al.*, 2009; review: Datta *et al.*, 2017) and mostly occurs under aerobic conditions. However, most studies dealing with lignin degradation focus on single strains under laboratory conditions and therefore, a better understanding of lignin degradation and involved C-fluxes through the microbial food web in particular under field conditions is still needed (see review by Datta *et al.* (2017)).

### **Modelling perspectives for non-mycorrhizal fungi and bacteria**

Litter decay rates depend on litter chemistry (e.g., lignin content), but also on microbial activity and the amount of microbial biomass, it is therefore an active process which cannot be adequately represented as depending only on t and humidity of the litter (Deckmyn *et al.*, 2008). It has been long ago proposed to include microbial biomass and activity in soil carbon models (Hunt, 1977), but only the growing recognized importance of microbes in processes such as priming (Neill & Gignoux, 2006) and formation of stable SOM in recent decades has spiked the interest in this idea. This has given rise to a new generation of microbially-explicit biogeochemistry models (reviews Treseder *et al.*, 2012; Sulman *et al.*, 2018) such as MIMICS (Wieder *et al.*, 2014; Wieder *et al.*, 2015), CORPSE (Sulman *et al.*, 2014), LIDEL (Campbell *et al.*, 2016), MEMS (Robertson *et al.*, 2019) and others (Deckmyn *et al.*, 2011; Chertov *et al.*, 2017a). These models explicitly represent the soil microbial community and its role in SOM dynamics; dead microbial biomass is the main contributor to SOM and litter enters the SOM pool primarily via its transformation/incorporation by microbes (Wieder *et al.*, 2014; Wieder *et al.*, 2015; Campbell *et al.*, 2016; Grandy *et al.*, 2016). Microbial activity is modified by temperature and a variable growth efficiency parameter. Microbial biomass is assumed to be at any given time in balance with the available C-sources which is a reasonable assumption.

The modelled effects are usually limited to effects on OM decay/formation and N mineralization, but the important role of bacteria in the N-cycle as denitrifiers or N-fixing bacteria could also be modelled. This might be useful in ecosystems where these processes contribute significant fluxes of nitrogen and closing the N-budget is required (Treseder *et al.*, 2012; Levy-Booth, Prescott & Grayston, 2014). Besides denitrification, modelling other anaerobic processes carried out by bacteria (such as methanogenesis) might be necessary for some ecosystems. In terms of microbial diversity, models have so far mostly distinguished only between two functional groups, be it between fungi and bacteria (Riley *et al.*, 2014), copiotrophs and oligotrophs (Wieder *et al.*, 2014), SOM builders and decomposers (Perveen *et al.*, 2014) or litter-eating r-strategist and SOM-eating K-strategist (Fontaine & Barot, 2005). Bacteria and fungi are often modelled as separate pools because they differ in physiological traits relevant for C and N cycling and their relative abundance influences C and N dynamics at the ecosystem scale (Waring, Averill & Hawkes, 2013;

*Louis et al., 2016*). Even though within bacterial and fungal communities differences exist in terms of life strategies, when compared to each other, heterotrophic aerobic bacteria can be seen as copiotrophic—fast-growing in nutrient-rich environments—, while fungi as oligotrophs—slow-growing, better adapted to nutrient-poor environment (*Ho, Di Lonardo & Bodelier, 2017*).

Incorporating more detailed information about microbial diversity is controversially discussed (*Nannipieri et al., 2003; McGuire & Treseder, 2010*; review: *Nielsen et al., 2011; Graham et al., 2014*). The diversity of soil microorganisms (e.g. species richness and relative contribution of each species to the community composition), is vast, with a high level of functional redundancy in C and N transformations, which makes it difficult to explicitly integrate the microbial diversity in soil C and N models (*Louis et al., 2016*). Although including only two or three (if mycorrhizal and non-mycorrhizal fungi are distinguished) functional groups of microbes substantially underrepresents their observed functional diversity in soils, the use of multiple SOM decomposing microbial functional groups has not been explored to date (review: *De Graaff et al., 2015*).

In our view, in many cases it can be enough to distinguish between fungi and bacteria assuming the former are more oligotrophic and the latter copiotrophic. This approach is practical because of their differential contribution to SOM decay, and also because F:B ratio can be easily related to soil C:N ratio and/or to pH similar to the approach in Romul\_Hum (*Chertov et al., 2017a, 2017b; Fierer et al., 2009*).

Based on the very fast life cycle of bacteria, and the ‘everything is everywhere’ hypothesis that states that when conditions change the bacterial community will change as well, the bacterial community can for example switch quickly to an anaerobic life style. Building predictive models that link dynamically changing microbial communities to ecosystem function by explicitly calculating population dynamics is probably less necessary/relevant at the time scales interesting for ecosystem studies (*Widder et al., 2016*; review: *Succurro & Ebenhöh, 2018*).

### **Mycorrhizal fungi**

Mycorrhizal fungi are a group of soil dwelling fungi that form a symbiotic relationship with a vast majority of vascular plants (*Brundrett & Tedersoo, 2018*). Mycorrhizal fungi provide host plants with nutrients and improve biotic and abiotic stress tolerance (see review by *Smith, Anderson & Smith (2015)* and *Pozo et al. (2015)*), often leading to increased plant diversity and productivity of the host plants (*Van der Heijden, Bardgett & Van Straalen, 2008; Van der Heijden et al., 2015; Tedersoo, Bahram & Zobel, 2020*). Mycorrhizal fungi require C from their host plants to grow and form hyphae (mycelium) extending into the soil to take up water and nutrients (mainly N and P) that are subsequently transferred to their plant hosts (*Smith & Read, 2008*). While the nutrient to C exchange rates are highly variable, plants trade 15–30% of their C for gaining on average ca. 75% of their required N; for the fungi, this represents all of their required C at a cost of 40% of their N (*Hobbie & Hobbie, 2006; Smith & Read, 2008*). The C transfer from the plant to the mycorrhizal hyphae can occur quickly, contributing up to 30% of the total respiration in soil (*Söderström & Read, 1987*).

Structurally, there are several different types of mycorrhizal interactions (mycorrhizas). The most common types are the ectomycorrhizas (EcM fungi) with high number of taxa and a low number of plant partners but dominant in many ecosystems; arbuscular mycorrhizas (AM fungi) with a low number of taxa but a high number of plant partners and ericoid (ErM fungi) and orchid mycorrhizas (OrM fungi), which are restricted to plants in the Ericaceae and Orchidaceae families, respectively (review: [Frąc et al., 2018](#)). The C flux from plants to AM mycorrhiza has been estimated to 5 Pg per year ([Bago, Pfeffer & Shachar-Hill, 2000](#)) which represents about 10% of global NPP (50–60 Pg, [Nemani et al., 2003](#)). If we compare it with the 5% of NPP allocated to rhizodeposition ([Pausch & Kuzyakov, 2018](#)) this suggests that AM fungi receive a significant proportion of the belowground labile C inputs. In one gram of forest soil, tens to hundreds (50–800) of meters of EcM mycelia can be found, representing 20–30% of the total soil microbial biomass ([Söderström, 1979](#); [Leake et al., 2004](#), review: [Ekblad et al., 2013](#)). Mycelial biomass corresponding to EcM fungi can range from 100 to 600 kg ha<sup>-1</sup> ([Wallander, Goransson & Rosengren, 2004](#); [Cairney, 2012](#); [Hendricks et al., 2016](#)) or up to 1.5 Pg of AM fungal biomass globally ([Treseder & Cross, 2006](#)). Mycorrhizal fungi also contribute to soil structure and aggregation ([Lehmann & Rillig, 2015](#)) and their senescing hyphae provide C to the soil ([Wilson et al., 2009](#)). They also play a role in water absorption and transport ([Johnson et al., 2012](#)) even between multiple trees or seedlings ([Warren et al., 2008](#)).

For the plants, AM fungi are thought to be more important for uptake of P and mineral or other readily available N, whereas some EcM and ErM fungi are able to break down SOM to obtain nutrients, mainly N ([Moore et al., 2015](#); [De Vries & Caruso, 2016](#)) but also P (review: [Tedersoo & Bahram, 2019](#)). Thus, mycorrhizal fungi can play key roles in mobilizing organic N trapped in the SOM for plant primary production ([Rineau et al., 2013](#); [Shah et al., 2016](#)). The EcM fungal mycelium can retain in its biomass a high proportion of N ([Lindahl et al., 2007](#)) which can prevent up to 50% of nitrate leaching losses; reductions of organic N and P leaching have also been reported. The uptake and immobilization of N by EcM fungi may also aggravate and stabilize a state of strong N limitation in nutrient-poor forests ([Näsholm et al., 2013](#); [Franklin et al., 2014](#)). It has also been proposed that EcM fungi compete with the decomposer community for organic N and restrain activities of saprotrophs ([Bödeker et al., 2016](#)). This is known as the Gadgil effect ([Fernandez & Kennedy, 2016](#)) and results in a decrease of the nutrient content of SOM, reduced SOM decomposition and an increase in soil C ([Orwin et al., 2011](#); [Averill, Turner & Finzi, 2014](#); [Averill, 2016](#)).

### **Modelling perspectives for mycorrhizal fungi**

EcM and AM fungi form the most common types of mycorrhizas and it is therefore reasonable to include them in general soil/ecological models ([Treseder, 2016](#)). Several models have been developed to include the mycorrhizal symbiosis in individual plant models (reviewed by [Deckmyn et al. \(2014\)](#)), but they are rarely included in ecosystem models. Examples of such ecosystem models are the MoBiE and MYCOFON models ([Meyer et al., 2010](#); [Meyer, Grote & Butterbach-Bahl, 2012](#)), the C accumulation model MySCaN ([Orwin et al., 2011](#)), an AM fungal distribution model proposed by [Schnepp & Roose \(2006\)](#),

the mycorrhiza C partitioning model described by *Staddon (1998)*, and the EcM forest model by *Franklin et al. (2014)*. These models represent the symbiotic trade of C and mineral nutrients between plants and fungi, which is modelled in different ways. The most parsimonious approach is based on the assumption that fungi only transfer N that is taken up in excess of their own N demands to the plants (*Näsholm et al., 2013; Franklin et al., 2014*).

Recently *De Vries & Caruso (2016)* have developed a conceptual model for the soil food web considering the ability of EcM fungi to decompose SOM by extracellular enzymes (*Read & Perez-Moreno, 2003; Phillips, Ward & Jones, 2014*), previously only attributed to non-mycorrhizal fungi. Using a mechanistic model, *Baskaran et al. (2017)* showed that the capacity of EcM to decompose SOM leads to reduced soil C, increased tree growth and a shift in the balance between microbial groups.

In summary, while the key role of mycorrhizal fungi in providing nutrients to plants in exchange for C is relatively well understood, this is not true for the effects of mycorrhizal fungi on SOM decomposition. Because of the global importance of mycorrhizal symbiosis and the large C and nutrient fluxes involved, more research on these effects are urgently needed. As far as the uptake of nutrients is concerned, it is not unrealistic to simulate mycorrhizal fungi as ‘part’ of the plant fine roots. However, the main drawback is that only mineral N and P can be taken up by the plant, whereas in reality mycorrhizal fungi can also obtain nutrients from recalcitrant SOM and thus play a vital role in the SOM dynamics (*Deckmyn et al., 2014*).

#### **Recommended literature:**

Root exudates and microbes: *Badri & Vivanco (2009)*;

Enzymatic degradation of lignin: *Datta et al. (2017)*;

Fungi versus bacteria: *De Boer et al. (2005)*;

Importance of diversity of soil organisms: *De Graaff et al. (2015)*;

Protists: *Gao et al. (2019)*;

Fungal decomposition: *Geethanjali & Jayashankar (2016)*;

Mycorrhizal fungi: *Smith, Anderson & Smith (2015)*; *Deckmyn et al. (2014)*; *Tedersoo & Bahram (2019)*

Modelling of microbial systems: *Succurro & Ebenhöh (2018)*

#### **Microbivores**

Microbivores are animals that feed on the soil microflora (i.e., bacteria, Archea and fungi). In terms of size, they belong to both microfauna (protists, nematodes) and mesofauna (mites, collembolans, enchytraeids). Microbivores exert a primary control on bacterial and fungal biomass and activity, with cascading effects on soil carbon and nutrients (*Bardgett & Van der Putten, 2014; Blanc et al., 2006*; review: *Trap et al., 2016*; review: *Gao et al., 2019*). For example, a recent review revealed that, although on average, the presence of active bacterivores reduces soil microbial biomass by 16%, they increase soil respiration by 29%, plant biomass by 27%, and shoot N and P contents by 59% and 38%, respectively (*Trap et al., 2016*). In other words, the flow of C and N through soil,

and possibly other elements, from the bacterial and fungal pools to the SOM pool and to plants is controlled by the size, activity and efficiency of microbivores (Berg *et al.*, 2001; review: Frouz, 2018). Therefore, proper simulation of their effects in a food web SOM model is most likely crucial.

Microbivores are generally divided into bacterial and fungal feeders. Bacterial feeding organisms are generally small (mostly microfauna) and include notably nematodes such as Cephalobidae and free-living protozoans such as Amoebae and Flagellates (Blanc *et al.*, 2006). Fungal feeders include families of nematodes which use a stylet or spear to penetrate fungal hyphae of saprotrophic or mycorrhizal fungi (Yeates *et al.*, 1993). Mites and collembolans (mesofauna) are also important grazers of bacteria and fungi, but not exclusively so, as they also consume other food sources such as plant litter (Brussaard, 1997). In general, larger animals will tend to ingest plant litter and soil together with microbes. Not so many data are available concerning their abundance. Pausch *et al.* (2016), using  $^{13}\text{C}$  labelling, found  $51 \text{ mg C m}^{-2}$  in bacterial feeders and  $68 \text{ mg C m}^{-2}$  in fungal feeders in an arable maize field.

Although microbivores have probably little direct impact on soil structure (Lehmann, Zheng & Rilliga, 2017), the opposite is not true, as soil structure is thought to have a large influence on the predation potential of microbivores. For example, Cephalobidae nematodes have a much higher impact on bacterial community composition and biomass in large pores than in the bulk soil, presumably because they cannot access pores smaller than  $10 \mu\text{m}$  (Blanc *et al.*, 2006). Likewise, microbial biomass and diversity is highest in microaggregates while nematode abundance and diversity is highest in large macroaggregates (Zhang *et al.*, 2013). It is therefore likely that changes in soil structure with both SOM content and activities of soil fauna engineers induce a feedback mechanism on microbivores.

As far as DOM is concerned, there are several studies showing that microbivore soil fauna can increase the rate of N leaching (Setälä *et al.*, 1990; Toyota *et al.*, 2013; Williams & Griffiths, 1989). Similarly, Liao, Xu & Zhu (2015) compared litterbags accessible and not accessible to microbivores and found that microbivores decreased the CN ratio in DOM. One possible explanation is that faunal grazing can reduce microbial immobilization of N (Carrera *et al.*, 2011). This change in CN ratio of DOM can affect the rate of decomposition in the soil.

### **Modelling perspectives for microbivores**

Microbivore functions in soils should be taken into consideration in our efforts to improve SOM models for predicting soil fertility and C sequestration. Many of the needed parameters have been evaluated for some organisms, but the number of studies is still too limited to reliably quantify the overall effect of microbivores on ecosystem functioning (Trap *et al.*, 2016). Nonetheless, initial values from these studies might be enough to start exploring their effects on soil C, N and P dynamics. Predicting microbivore effects in specific environments remains difficult (Trap *et al.*, 2016) but a first effort targeting generic simulation of effects would be of great value.

The diversity of soil fauna feeding on the microorganisms and, at least for some of them, the non-specificity of their diet pose two challenges in terms of modelling. First, it is not clear if a common parameterization can be used for one generic pool of microbivores. For example, do fungal and bacterial feeders have a similar CN ratio, respiratory quotient, generation time and mortality rate? Although it is certainly not the case, standard parameters across a wide spectrum of organisms should be investigated. For example, microbivore composition has been reported to affect neither trophic-level biomass nor the response to increased resource availability (*Mikola, 1998*). The second challenge is that larger soil fauna, that is, mesofauna, do not feed exclusively on the soil microflora but might also digest litter, thereby creating an overlap between potential model pools of detritivores, on the one hand, and microbivores, on the other hand. The modelling concept based on nutrient stoichiometry (see Predators) developed by *Osler & Sommerkorn (2007)* is also relevant for microbivore microorganisms as well as for larger soil faunal predators.

It is clear that microbivores require more attention in our studies, so their role can be better understood and represented in more detail in SOM models if proven as beneficial for their predictive power. Given the current, limited, data, they can be simulated as a link between the microbial biomass and the larger predators and detritivores. Because these links and their importance in terms of SOM fluxes, are largely determined by pore size distribution, we would suggest to simulate only the micro faunal microbivores in simple models.

**Recommended literature:**

Protists: *Gao et al. (2019)*

Soil macro-and mesofauna effects on decomposition: *Frouz (2018)*

Bacterivores: *Trap et al. (2016)*

**Predators**

Soil predators are represented in each size class of soil fauna (micro-, meso- and macrofauna) and include predatory protists, nematodes, mites, centipedes, and others. The three size classes also form a hierarchy where larger animals prey on smaller animals as well as on prey of their own size. For instance, the main microfauna groups, nematodes and protists, have predators preying within and among them including Protozoa feeding on nematodes and vice-versa (*Geisen, 2016*). Isotopic studies have demonstrated that predators form a soil fauna group of their own, that is, an isotopic niche (*Korobushkin, Gongalsky & Tiunov, 2014*), including spiders, Gamasida and nematodes, preying on microbivores, detritivores and herbivores. Even the neanurid collembolans are classified as predators (*Hopkin, 1997; Potapov et al., 2016*), thus inhabiting the same isotopic niche as the afore mentioned predators.

Predation in soil challenges our conception of a boundary between aboveground and belowground biota. Aboveground predators, such as spiders, beetles and harvestmen in fact feed on prey traditionally considered to be soil organisms.

While predatory mites, spiders and beetles are ubiquitous, centipedes are rare in conventional agricultural systems, but enjoy the conditions offered in biological

agriculture. One of the consequences seems to be that under conventional agriculture there sometimes is a higher impact of pest species (herbivores) because of the lack of predators (*Kladivko, 2001*).

Soil predators can influence the entire food web by creating important secondary effects. For example, bacterivorous nematodes have been shown to increase plant P uptake by different mechanisms. Nematode predators can decrease bacterial grazing and thus increase mineralization by bacteria, because of the higher bacterial turnover (*Ferris et al., 1998*) They can also have a hormonal effect on plant roots increasing branching and therefore P-uptake capacity of the plants (*Ranoarisoa et al., 2018*).

### Modelling perspectives for predators

To our knowledge, there are no ecosystem models that include soil faunal predators, apart from the Romul-Hum extension to the ROMUL model (*Chertov et al., 2017a, 2017b*). In this model, predators are not a dynamic pool but a fixed part of the soil food web depending on soil characteristics. It is clear that more data are necessary to validate the population dynamics of predators and subsequently their effect on SOM dynamics. However, as suggested above, some important effects of differences in management cannot be simulated without including the predators.

The model framework described by *Osler & Sommerkorn (2007)* shows how using nutrient stoichiometry could be an effective and simple way to include the influence of predation on the C and N cycling. The main concept of their framework is that soil fauna with a high C-efficiency and prey with a similar CN ratio contribute to the mineral N, while inefficient assimilators that consume prey with a higher CN ratio would contribute more to the DOM pool.

Given the larger size and longer life-spans of many predators, simulating their effects as “in balance” with the environment seems unrealistic. To model the effects of land use changes (e.g., agricultural conversions, tillage, etc.) or drought periods/flooding in a more realistic fashion including a dynamic pool of predators seems a worthwhile extension to existing ecosystem models for many environments.

### Recommended literature:

ROMUL-Hum model: *Chertov et al. (2017a, 2017b)*

CN ratio's through soil faunal network: *Osler & Sommerkorn (2007)*

### Herbivores

Herbivores eat living plant material, such as leaves, flowers, stems and roots. Herbivores exert an influential role in plant community dynamics (*Bever, 2003*), which in turn determines the amount and quality of plant litter entering the soil and the density and tissue quality of roots. Herbivores have an effect on the amount of SOM via different actions. About 50% of net primary production occurs belowground, in the form of roots, while the largest part of aboveground primary production enters the soil in the form of litter (*Sinacore et al., 2017*). Root herbivory affects plants in all ecosystems (see meta-analysis by *Zvereva & Kozlov (2012)*). *Andersen (1987)* reported up to 30% of

root biomass is lost through herbivory. Accordingly, the invertebrates with a greater effect on carbon dynamics are root feeders (*Treonis et al., 2001*).

Root herbivores are a diverse soil fauna feeding group. Among microfauna they are represented by the plant-feeding and plant parasitic nematodes. These feed mainly on plant juices by tapping into the root. The density of plant-feeding nematodes varies greatly among ecosystems, but due to their short life cycle and fast reproduction they can significantly affect plant communities, including a severe reduction in the crop yields (*Yeates et al., 1993*). Symphyla and prostigmatid mites (partly) belong to the mesofauna and are also considered root feeders (*Striganova, 1980; Orgiazzi et al., 2016b*). However, the most influential root herbivores in terms of effects on the plants they feed on are found in the macrofauna, and include Diptera larvae (mainly midges), caterpillars and some major groups of beetles, such as click beetles and curculionids (mainly their larvae) (review: *Johnson & Rasmann, 2015*).

The highest recorded average density of Symphyla (plant-feeding Myriapoda) is around  $10.8 \times 10^3 \text{ m}^{-2}$  (*Belfield, 1956*). The few other sources generally report lower densities, around  $200 \text{ ind. m}^{-2}$ . With an average individual dry weight of  $81 \mu\text{g}$ , this translates in an annual mean biomass estimate of  $58 \text{ mg m}^{-2}$  (*Reichle, 1977*). Prostigmatid mites can be very abundant in temperate coniferous forest (up to  $2 \times 10^5 \text{ ind. m}^{-2}$ ;  $30 \text{ mg dry weight m}^{-2}$ ), and less abundant in tundra systems (around  $10^3\text{--}10^4 \text{ ind. m}^{-2}$ ;  $10 \text{ mg dry weight m}^{-2}$ , *Petersen, 1982*), with a mixed oak forest in between (*Lebrun, 1971*). An average dry weight of about  $0.5 \mu\text{g}$  (range  $0.2\text{--}4.0 \mu\text{g}$ ) is assumed in most data sets, resulting in an average biomass ranging between  $10 \text{ mg m}^{-2}$  (tundra and temperate deciduous forest) and  $50 \text{ mg m}^{-2}$  in tropical grasslands (*Petersen, 1982*).

Diptera larvae are probably the most important meso- and macrofauna root herbivores in terms of the overall effects on plant growth and physiology (but there are also caterpillars, wireworms, weevils and other insect herbivores), (*Stevens et al., 2000, Samson, 2001*). Their average biomass ranges between  $10 \text{ mg dry weight m}^{-2}$  in tropical grasslands to  $0.47 \text{ g m}^{-2}$  in tundra ecosystems (*Petersen, 1982*). Being of larger size, beetle densities will on average be much lower than Diptera densities. Based on average biomass estimations for predaceous beetles (Carabidae and Staphylinidae), that is, ranging between  $10 \text{ mg m}^{-2}$  to  $0.12 \text{ g m}^{-2}$  (*Petersen, 1982*), the biomass of root feeding beetles (Elateridae and Curculionidae) is probably in the same range (*Parker & Howard, 2001*).

### Modelling perspectives for root herbivores

The number of studies on consequences of root herbivore-plant interactions for the ecosystem is still quite limited (*Andersen, 1987; Eissenstat et al., 2000*, see also “Fine roots”). However, the available information from many studies on specific plant-root herbivore interactions (*Zvereva & Kozlov, 2012*) is enough to start exploring the effects of introducing root herbivores in SOM models on soil C and nutrient dynamics. Predicting root herbivore effects in a specific environment remains difficult, due to a number of often unknown factors, that is, species composition, actual density, ecological efficiencies (which can deviate considerably between modes of feeding), and population turn-over rates or generation times, but a first effort targeting generic simulation of effects would still



be of great value. At an ecosystem level fine root turnover is one of the most important C-sinks, and the fate of fine roots (whether they die or are eaten) could potentially have a major effect on the simulated C-balance ([Brunner et al., 2013](#)).

#### **Recommended literature:**

Root longevity: [Eissenstat et al. \(2000\)](#)

Root herbivory: [Zvereva & Kozlov \(2012\)](#)

Root-feeding insects: [Johnson & Rasmann \(2015\)](#)

#### **Detritivores**

##### **Mesofauna detritivores**

Mesofauna detritivores feeding on decomposing organic matter (plant and animal remains), also called saprophages, include enchytraeids, collembolans, large groups of mites, some small-sized Diptera larvae, Protura and Diplura. The first three groups have been recognized as having major ecological importance in terms of abundance and biomass whereas the rest have been subjected to very little specific research and will not be further included. As a whole, their primary role shifts between promoting physical (fragmentation) or chemical changes of the organic material ingested, depending on the group of species ([Wallwork, 1970](#), review [Dervash et al., 2018](#)). These transformations mainly occur at the top layers (organic soil horizons but also in the litter layer, under stones, etc.) due to their limited burrowing abilities. As explained earlier (see “Microbivores”) apart from ingesting litter, mesofaunal detritivores also graze on bacteria and fungi and thus belong also to microbivores.

##### **Enchytraeids**

General population density estimates range from 10,000 to 300,000 individuals  $m^{-2}$  ([O'Connor, 1967](#); [Briones, Ineson & Heinemeyer, 2007](#)), with the majority occupying the upper layers (the 0–4 cm can concentrate >70% of the total population; [Briones, Ineson & Pearce, 1997](#)). The main factors controlling their population sizes and vertical distribution are temperature and moisture.

There are no quantitative reliable estimates of enchytraeids' consumption and digestion rates or agreement on their preferred food sources. As a rule of thumb, it is believed that they feed on organic matter (20% of their diet), bacteria (40%) and fungi (40%) ([Didden, 1993](#)). Like earthworms they burrow through the soil and ingest the soil. More recently, C dating techniques performed on field populations have established that they feed on 5–10 year old organic matter ([Briones & Ineson, 2002](#)). Importantly, temperature-driven increases in their population size results in a greater competition and thus, when biomass reaches a value of  $2.1 \text{ gm}^{-2}$  ([Briones, Ineson & Heinemeyer, 2007](#)) consumption of older organic matter substrates increases and consequently, also a greater release of non-labile C occurs ([Briones, Ostle & Garnett, 2007](#)). Interestingly, in certain ecosystems, such as coniferous moder soils their metabolic contribution has been estimated to be 11% ([O'Connor, 1967](#)) and is comparable to that exhibited by woodland earthworm populations (8–10%; [Satchell, 1967](#)).

### Collembolans

Collembolans are important as epigeic decomposers (Ponge, 1991) and metabolic rates are comparable to those for enchytraeids and nematodes of similar weight (Mc Millan and Healey, 1971).

As many as 53,000 m<sup>-2</sup> (equivalent to 330 mg m<sup>-2</sup>) have been found in a limestone grassland (Hale, 1966). However, their numbers fluctuate seasonally and with food availability, and for example, 670,000 individuals m<sup>-2</sup> have been recorded in permanent moist soil in Antarctica covered by the alga *Prasiola crispa* (Collins, Baker & Tilbrook, 1975). Predation seems to be the primary regulatory factor of their population sizes (Wallwork, 1970).

As many hexapods, they accumulate a high proportion of fat in their bodies (54% of dry weight or 24% of live weight) which increases with age (Anderson & Healey, 1972). Importantly, they shed their exoskeleton several times as they grow (up to 60 times in their lives) and in exuvia representing 2–3% of body weight (Anderson & Healey, 1972) which could be an important source of nutrients for other soil organisms.

### Oribatid mites

Although the majority of oribatid mites are considered to be panphytophages (Luxton, 1972), more recent work (Schneider et al., 2004) indicated that besides fungal feeders and predators, there are larger groups that can be defined as primary and secondary decomposers and hence, having a preference for litter at different decomposition stages as well as being coprophagous (feeding on fecal material) (Petersen & Luxton, 1982). Their role in soil mixing is small compared to other invertebrates but they play an important role in humus formation and mineral turnover (Hoy et al., 2008). They produce fecal pellets, which help to distribute organic matter and are prone to microbial attack. Mites can colonise all soil horizons, including the mineral layers and can reach up to  $3 \times 10^5$  ind. m<sup>-2</sup> in temperate mixed forests (Petersen & Luxton, 1982). These high densities are the result of their fast life cycles, which in the case of small species could be several generations per year (Mitchell, 1977).

### Macrofauna detritivores

Macrofauna detritivores include soil organisms that are larger than 2 mm, such as isopods, millipedes, earthworms, ants, and termites. They either live in litter or excavate the soil in search for plant remains and SOM. The engineering capacities (burrowing and bioturbation) of some species in this group will be discussed further (see “Soil structural modifications by engineers”), but they also have an important role in the C-cycle.

Macrofauna detritivores can reach very high densities and biomasses. For example, earthworms are abundant as long as the climate is humid and warm enough, at least for a certain part of the year. When soils contain enough organic matter (for endogeic earthworms that ingest soil and digest SOM) and primary production is high enough (for epigeic and anecic earthworms that eat plant litter) earthworms can be very abundant (i.e., more than 10<sup>6</sup> individuals ha<sup>-1</sup>) and their biomass can be as high as 1,000 kg ha<sup>-1</sup>

(Lavelle & Spain, 2001). Endogeic earthworms may ingest more than their own weight of soil each day, so depending on their abundance and climate they may process all the soil in 5 years or less (review: Curry & Schmidt, 2006).

### Quantitative contribution of detritivores to SOM transformations

The bulk of plant-derived C enters the soil only when the vegetation dies. A fraction of it is transformed by the decomposers through breaking down the organic substrates and assimilated into their tissues, another fraction is released as fecal material and/or exuvia, respired as CO<sub>2</sub> and finally deposited as dead bodies (Petersen & Luxton, 1982).

There are very few estimates of how much organic material is ingested, digested, assimilated and respired by individual groups. In one year, detritivores (including earthworms) may consume 20–100% of the total annual input of litter (Frouz *et al.*, 2015). Certain species, such as blanket bog enchytraeids are responsible for processing 40% of the total litter input (Standen, 1973).

Even fewer attempts have been made to measure how much of the ingested organic matter has been assimilated. Overall, it has been suggested that the range of assimilation efficiencies is wide (1–65%), with oligochaetes being the least efficient (Petersen & Luxton, 1982). Under laboratory conditions, the measured metabolic activity of enchytraeids and collembolans per unit of dried weight seems to be twice that of oribatid mites (compiled by Wallwork, 1970). In certain ecosystems where these organisms are dominant, their contribution could have a great influence. For example, in moorland soils, 70–75% of the total energy is assimilated by the dominant enchytraeids (Heal, Jones & Whittaker, 1975).

A certain amount of energy ingested is metabolized and most of it is dissipated in respiration. Temperature has a strong influence on soil respiratory metabolism. For example, in a laboratory incubation of a grassland soil, Q<sub>10</sub> significantly increased and was 25% greater in the presence of enchytraeids (Q<sub>10</sub> = 3.4) than in their absence (Q<sub>10</sub> = 2.6) (Briones, Poskitt & Ostle, 2004) and even higher values were reported when enchytraeids were incubated in a peatland soil (Q<sub>10</sub> = 3.9; Carrera *et al.*, 2009).

In the field, the whole picture gets complicated because estimates change with population densities (and hence, with biomass and age structure) that are known to fluctuate with seasons (and thus, with variations in ambient temperature and moisture conditions).

A good quantitative assessment was provided by Petersen & Luxton (1982), who concluded that soil detritivores are reasonably efficient in assimilating organic matter (40–50%) and have a community growth efficiency of 10–20%; 4–85% of the assimilated energy is dissipated in respiration, with only 15–50% being allocated to growth and reproduction. In addition, coprophagy is important since it allows a better reutilization of organic substrates that were not fully digested on first consumption.

Furthermore, the role of soil animals on the retention of other nutrients can also be crucial: McBrayer (1977) estimated that 70% of the N released during litter decomposition is immobilized by soil invertebrates. Similarly, MacLean (1980) indicated that up to 1 mg P and 10 mg N m<sup>-2</sup> are found in dipteran adults emerging from tundra soils forming a major redistribution mechanism in these nutrient-poor soils.

On the other hand, detritivores can also increase the mobilization of C, N and P. For example, enchytraeids have been found to release significant amounts of CO<sub>2</sub> and dissolved organic C (DOC) (*Briones, Carreira & Ineson, 1998; Briones, Poskitt & Ostle, 2004; Carrera et al., 2009, 2011*). They also increase leaching of dissolved organic N (DON), ammonium and phosphorus (*Briones, Ineson & Poskitt, 1998*). Similarly, significant increases in the leaching of ammonium, nitrate and calcium occurred as a consequence of collembolan grazing (*Ineson, Leonard & Anderson, 1982*).

### **Modelling perspectives for detritivores**

Mesofauna detritivores have not been included into ecosystem scale models so far, and information at this scale is scarce. Nonetheless, their impact on the ecosystem has been shown to be significant (*Frouz et al., 2015; Filser et al., 2016*).

In our view, it is not possible (for lack of data) nor necessary for a simple SOM model (with the goal of modelling soil ecosystem functioning and C, not soil fauna) to distinguish the different mesofauna detritivores. However, parameterization of the saprotroph pool can mimic the differences between them. In the simplest case, this can be seen as a fixed relative abundance of the various species that determines the ‘average’ parameters. Besides maximal growth rate and respiration, CN ratio and response to temperature (Q<sub>10</sub>) are important to characterize this group, as is the production of excrements, exuvia and exoskeletons that need not be addressed separately but can be an important flux. From the review it seems clear that distinguishing only between C used for growth and C respired is not an adequate representation. Although the concept of recalcitrance has been questioned, it can still be used here to allow some chemical changes by detritivores that slow down decay and favor fungal decay above bacterial decay.

For macrofauna detritivores quite a number of models have been developed that often focus on their engineering capacity, these models are discussed in “Soil structural modifications by engineers”.

### **Recommended literature:**

Soil meso-fauna effects on SOM and litter: *Dervash et al. (2018)*;

Enchytraeids: *Briones & Ineson (2002)*

### **Fine roots**

The rhizosphere, the area of soils conformed by the fine roots and the microorganisms directly associated with them, has been shown to be of great importance to soil C and nutrient dynamics (*Adamczyk et al., 2019; Kriiska et al., 2019*). Fine root dynamics and activity includes the production of biomass and necromass as well as a continuous release of exudates from roots that is the base food for a large community of soil microorganisms and soil fauna (e.g., detritivores, herbivores) (*Juan-Ovejero et al., 2020*). Nowadays, the definition of ‘fine roots’ is under discussion, as the commonly used 2-mm threshold (*Finér et al., 2007*) is not a functional criterion and lumps together both thin and woody transport roots and absorptive roots with primary structure (*Ostonen et al., 2017*). Despite the fact that thin woody roots, absorptive roots and associated mycorrhizal mycelium cycle carbon at significantly different rates (*Godbold et al., 2006; Leppälammil-Kujansuu et al., 2014*),

fine root turnover is a significant and dynamic C sink. Furthermore, the direct input of DOC from fine roots is important for leaching and for all interactions with soil biota ([Juan-Ovejero et al., 2020](#)).

The root litter contribution to soil C is often underestimated ([Rasse, Rumpel & Dignac, 2005](#)). Live roots contain high concentrations of soluble and easily decomposable organic substrates (e.g., glucose, malate, cellulose, peptides such as glutamate), whereas root necromass is rich in organic constituents (lignin, suberin) characterized by lower decomposition rates (recalcitrant substances) ([Rasse, Rumpel & Dignac, 2005](#)). The composition of the roots is considered to be relatively similar to the above-ground parts, showing a similar pattern between deciduous (higher in nutrients and soluble compounds) and coniferous (higher in lignin and liposoluble compounds) species although this relationship has not been found within species ([Hobbie et al., 2010](#)). However, differences in fine root activity (production and mortality) and decomposition among ecosystem types are not well known ([Coleman & Hendrix, 2000](#)) and even less is known regarding the impact of species on the amount and composition of root exudates though it has been shown the impact on the ecosystem can be significant ([Yin, Wheeler & Phillips, 2014](#)). Root and hyphal exudates particularly rich in readily available constituents may induce a small but significant increase in litter decomposition indicating an active role of the rhizosphere in soil priming ([Grayston, Vaughan & Jones, 1997](#); [Kuzyakov, Friedel & Stahr, 2000](#); [Rasse, Rumpel & Dignac, 2005](#)). Moreover, root turnover can be increased by 50% by grazing ([Eissenstat et al., 2000](#)) as described in “Herbivores”.

### Modelling perspectives for fine roots

In many ecosystem models, fine roots are still simulated as a single pool with a single turnover rate. Though some data on fine root distribution are available ([Fan et al., 2016](#); [Finér et al., 2011](#)) this is not the case in the majority of ecosystem studies since fine root measurements are time-consuming. Furthermore, when root growth is not well defined over the soil layers, nutrient and water uptake is obviously not simulated realistically over the layers as well. The rate and description of fine root turnover in models (constant proportion of NPP, or following the seasonality of leaf area (constant leaf to fine root ratio—[Deckmyn et al. \(2008\)](#))) has a major effect on the model outcome in terms of total C budget ([McCormack et al., 2015](#)). Fine root production partially follows aboveground productivity ([Abramoff & Finzi, 2015](#)) but is not yet fully understood nor implemented in models.

To link fine root growth with soil hydrology (water and nutrient uptake), a more detailed approach is required. Novel root architecture models and tomography techniques have facilitated the development of three-dimensional functional-structural models as reviewed by [Dunbabin et al. \(2013\)](#). The description of root water uptake has been advanced through more complex approaches that explicitly describe water flow in both the soil and inside the root system ([Javaux et al., 2008](#); [Schröder et al., 2009](#); [Mai et al., 2019](#)). Yet the impact of specific rhizosphere hydraulic properties on the root water uptake at the plant scale is generally not considered, except for instance in [Schwartz, Carminati & Javaux \(2016\)](#). Models that simulate root growth and nutrient uptake processes, like

R-SWMS or OpenSimRoot, enable calculation of nutrient uptake as the roots grow and receive photosynthates from the shoot (*Postma et al., 2017*). Examples of coupling of the root growth model RootBox with soil models are presented for example, in *Schnepf, Leitner & Klepsch (2012)* who simulated root system phosphate uptake from a rhizotron as affected by root exudation. In most of those models, root architecture is used to compute volumetric sink terms for water or nutrient uptake. Few examples exist that explicitly simulate the roots as physical objects with uptake prescribed via the boundary conditions at the root surfaces (*Leitner et al., 2010; Daly et al., 2017; Schnepf et al., 2020*).

However, these improved descriptions are not yet sufficiently incorporated into larger scale models (*Hinsinger et al., 2011; Vereecken et al., 2016*). Recent initiatives in this way already include soil resistance, plant root distribution and climatic demand, to upscale to the macroscale (*Javaux et al., 2013*). There remains an overall lack of spatially explicit models that properly describe soil C and nutrient dynamics at different spatial scales (*Manzoni & Porporato, 2009*).

How macropores are used by roots and how roots create macropores or induce compaction are still challenging questions (*Lesturgez, Poss & Hartmann, 2004*) which only start to be included in models (*Landl et al., 2017*).

### **Modelling soil food webs**

Soil food web modelling has mainly been used to calculate the flow of C and nutrients through soil and to investigate the role of the various functional groups in these flows (*Scheunemann et al., 2016; Malard et al., 2020*). This kind of modelling requires knowledge about the architecture of the food web (“who eats who”), the biomass of the functional groups and physiological information, such as generation time, growth and death rates and metabolic efficiencies (see chapters above for details). The importance of these types of models in explaining N and C cycling was already shown in the late 80’s and 90’s (*Hunt et al., 1987; De Ruiter, Neutel & Moore, 1994; Berg et al., 2001*); however, this knowledge did not find its way into the basically plant-centred ecosystem models and relatively little advance in the domain has been made. Nonetheless, *Berg et al. (2001)* and *Schröter, Wolters & De Ruiter (2003)* used such food web models at a forest ecosystem scale to show the importance of functional groups for predicting C and N dynamics in the soil.

To model the C and nutrient fluxes, many food web models first calculate the feeding rates among the functional groups. Next, C and N mineralization are derived from the feeding rates of functional groups using metabolic efficiencies, that is, assimilation and production efficiencies, and CN ratios of consumer and resource. The equations used to calculate the feeding rates follow the approach of “inverse modelling”, which goes back to *O’Neill (1969)* based on the conservation of matter and energy and the assumption that the system is at steady-state. This approach has first been applied to soil food webs by *Hunt et al. (1987)* and later by *De Ruiter, Neutel & Moore (1994)*, *Berg et al. (2001)* and *Schröter, Wolters & De Ruiter (2003)*.

Alternatively to a steady-state description, different approaches exist for modelling the growth of a species population within a food web, more often applied to insect populations

(see review by [Ju & Shen \(2005\)](#)). The first approach is to simulate an increase in population towards the carrying capacity of the system. This yields stable and reliable results, but does not allow for a strong influence of management or climate on the carrying capacity, so it is not so different from assuming a steady state. Other models opt for a more Richards' shaped growth curve, where growth rate goes to a maximum, allowing a direct link between resource and species and a dynamic representation of climate and management effects. To be sensitive to climate change a daily time step is most appropriate at a stand scale. Daily faunal pool sizes can be calculated as a set of linear equations for each pool including growth, turnover and respiration. A dynamic representation of all populations is thus possible. We have found no models using such an approach at an ecosystem scale however, although current computational power should allow this. The new ROMUL model ([Chertov et al., 2017a, 2017b](#)) has a detailed representation of soil fauna in 15 groups. This is the first model (to our knowledge) simulating the effect of the faunal food web, including necromass and respiration, on the C and N cycle of a soil. The biota are assumed to be at steady state and climate and management affect them only empirically.

#### **Interactions between SOM, soil structure and soil biota**

The processes involved in SOM stabilization are strongly controlled by soil biota. Bacteria and fungi are considered to be the most important soil microorganisms involved in the formation and stabilization of aggregates, especially at the microscale ([Gupta & Germida, 2015](#), review: [Costa, Raaijmakers & Kuramae, 2018](#)), though there is still considerable debate ([Lehmann, Zheng & Rilliga, 2017](#); [Oades & Waters, 1991](#); [Six et al., 2004](#)). In fact, mycorrhizal fungi are known to influence the movement of SOM into mineral soil ([Frouz et al., 2001](#), [Ponge, 2003](#)) but also the formation and stabilization of aggregates. Ectomycorrhizal fungi affect soil aggregation (reviewed in [Rillig & Mummey, 2006](#)) through changes in the root architecture by (1) covering fine roots with fungal mantles ([Smith & Read, 2008](#)), (2) producing hydrophobins in the mycelium and rhizomorphs ([Tagu et al., 2001](#); [Mankel, Krause & Kothe, 2002](#)) that help adherence to different soil surfaces, (3) enmeshing and entangling soil primary particles, organic materials and small aggregates, and (4) oxidizing of biomolecules present in SOM that leads to the formation of aggregates of organic matter ([Kleber & Johnson, 2010](#); [Kleber et al., 2015](#)). In sandy soil, only hyphal networks are able to tie the abundant sand particles to form stable aggregates ([Six et al., 2004](#)).

Bacteria can also have a profound influence on soil aggregation ([Six et al., 2004](#)). Like fungi, bacteria produce exopolysaccharides, which act as glue and help organic residues to attach to clays, sands and other organic material, resulting in the formation of new microaggregates (review: [Degens, 1997](#)). In addition, other groups of soil biota, such as microarthropods, are assumed to affect SOM stabilization; most likely by influencing organo-mineral interactions (e.g., by effects on soil chemistry and leachate) and aggregate formation (e.g., by necromass, eggs as aggregate starting point) ([Maaß, Caruso & Rillig, 2015](#); [Soong & Nielsen, 2016](#)). Similarly, it has been shown that earthworms can play a central role in physical stabilization of newly generated organic matter through soil aggregate

formation (Pulleman *et al.*, 2005; Rillig & Mummey, 2006; Six & Paustian, 2014; Bottinelli *et al.*, 2015; Angst *et al.*, 2017, 2019) during cast formation (see below).

The link between aggregation and porosity is hard to quantify. Regelink *et al.* (2015) showed for different soils that overall soil porosity is the sum of the structural porosity (determined by clay, sand and silt fractions) and aggregation. They concluded that micropores (which they define <9 µm) are mainly situated within the aggregates, while mesopores are situated between dry-sieved aggregates. Total porosity increased with total aggregate content, and the fraction of micropores increased with increasing dry-sieved aggregate content. In this study, macropores were not studied but obviously, biopores are also part of the soil porosity.

#### **Recommended literature:**

Microbial effects on aggregation: Costa, Raaijmakers & Kuramae (2018);

Mycorrhizal effects on soil structure: Rillig & Mummey (2006)

#### **Faeces**

When macrofauna is present, a substantial part of litter is turned into macrofauna excrements that are either holo-organic (such as millipede fecal pellets) or in the form of organo-mineral aggregates (such as earthworm casts) (Frouz & Kuraz, 2013). They can be deposited in the soil or at the surface in large quantities and in the case of some species of earthworms the surface aggregations of intact and fragmented litter together with defecated soil around the openings of the earthworm burrows are called “middens” and represent important microhabitats for microbial activities (Brown, 1995; Orazova, Semenova & Tiunov, 2003).

Several authors have shown that microbial activity increases during and shortly after faunal feeding but then decreases and may be lower in faunal faeces than in the non-ingested litter (Frouz & Šimek, 2009; Frouz, Santruckova & Elhottova, 1999; Lavelle & Martin, 1992; Tiunov & Scheu, 2000). The increase in microbial activity in fresh faeces is often attributed to litter fragmentation (Gunnarsson, Sundin & Tunlid, 1988; Kaneda *et al.*, 2013) which increases surface area and may thereby increase microbial access to the litter. Artificial litter fragmentation experiments have shown, however, that litter fragmentation alone may both enhance or suppress microbial activity (Gunnarsson, Sundin & Tunlid, 1988; Kaneda *et al.*, 2013).

The reasons for the decrease in decomposition rate and hence in the stabilization of SOM in the older faeces of soil fauna are also variable. Some macrofauna species, such as earthworms, consume organic matter together with soil mineral particles (Schulmann & Tiunov, 1999; review: Curry & Schmidt, 2006). This results in the binding of SOM in aggregates, which may slow decomposition and help stabilize SOM (Gunina & Kuzyakov, 2014; Lavelle, 1988; Six *et al.*, 2004). In the case of macrofauna that mainly consumes litter without soil, the reduced decomposability of their faeces is associated with changes in their chemistry compared to that of the original litter. The faeces are usually depleted in easily available polysaccharides, degraded by invertebrate enzymes (Frouz, Novakova & Jones, 2002), and are enriched in lignin (Frouz *et al.*, 2015; Hopkins, Wheatley & Robinson, 1998).



Because the easily available substances are not present in faeces, the decomposition rate is reduced (McInerney, Little & Bolger, 2001; Bossuyt, Six & Hendrix, 2005). The content of soluble phenols decreases after passage through the gut of litter-feeding fauna (Coulis *et al.*, 2009; Frouz *et al.*, 2015; Špaldoňová & Frouz, 2014), which may be caused by precipitation with proteins, making phenols insoluble (Frouz *et al.*, 2015) but at the same time also reduce N availability. Although earthworms are typically the main group contributing to faunal-mediated aggregation (Marashi & Scullion, 2003), faecal pellets produced by micro-arthropods have also been recognized as important contributors to aggregate formation (Maaß, Caruso & Rillig, 2015), either by promoting porosity or by filling the pore space between particles and hence, impairing fungal growth and decomposition.

For earthworm casts at the surface, aggregate degradation by rain can have a significant impact on their stability and the subsequent leaching of nutrients (Decaëns *et al.*, 1999) and similar effects have been found for termite mounds (review: Jouquet *et al.*, 2011).

#### **Recommended literature:**

Earthworms: Curry & Schmidt (2006);

Macro-aggregation: Degens (1997)

Termites: Jouquet *et al.* (2011)

#### **Soil structural modifications by engineers**

By definition, ecosystem engineers are organisms that have a measurable impact on the physical properties of their environment either through their activities or their mere presence (Jones, Lawton & Shachack, 1994; review: Jiménez & Lal, 2006).

Such organisms are thus often very influential for the functioning of ecosystems and tend to affect all organisms and their activities with which they share a common environment (Lavelle *et al.*, 2016). Note that engineers are also important because they can create heterogeneity in physical, chemical and biological features at various spatial scales (Barot *et al.*, 2007; Jiménez, Decaëns & Rossi, 2012; Jouquet *et al.*, 2007; Raynaud, Jones & Barot, 2013) and likely strongly influence the functioning of food webs (Sanders *et al.*, 2014). Three concurrent and interrelated processes are behind the engineering capacity of soil organisms but are generally considered separately for practical reasons: (i) biopore formation, (ii) bioturbation (soil mixing) and, (iii) fauna-mediated aggregation (see above “Faeces”).

#### **Biopore formation**

Many soil organisms can be considered as ecosystem engineers and are very influential for soil processes (Lavelle, Bignell & Lepage, 1997; Lavelle *et al.*, 2007). Indeed, soil biota require space and connectivity between pores to move through the soil, to forage for nutrients and/or carbon-based energy sources, water and living space (e.g., plant large roots and macrofauna such as earthworms, ants or termites). This can be achieved either by pushing aside soil aggregates or by ingesting soil (e.g., in earthworms), creating the so-called biopores that remain after roots death or the passage of fauna (Schneider *et al.*, 2018).

Some soil macrofauna is particularly influential for soil structure through their engineering activities, such as ants (review: [Folgarait, 1998](#)), termites ([Dangerfield, McCarthy & Ellery, 1998](#)) and earthworms ([Lavelle, 1988](#), [Lavelle et al., 2007](#)). As an example, values between 0.013 and 0.024 m<sup>3</sup> earthworm burrows m<sup>-3</sup> of soil have been reported ([Bastardie, Capowiez & Cluzeau, 2005](#)), that can persist for very long periods in the soil.

### **Bioturbation**

By burrowing through the soil and dragging litter, soil engineers mix mineral and organic materials from the different horizons in a process known as bioturbation. The extent and type of bioturbation largely depend on the ecological behaviour, body size and population density of the different species, and earthworms are a good example to illustrate this. Earthworms are traditionally classified into three main ecological groupings ([Bouché, 1977](#); [Brown, 1995](#)): epigeic, endogeic and anecic species. Epigeic and anecic earthworms consume fresh litter at the soil surface, whereas endogeic earthworms ingest more mineral soil creating a network of galleries and soil aggregates of various sizes (earthworm casts). While epigeics and endogeics mostly move horizontally in their respective layers, anecic earthworms create permanent or semi-permanent vertical galleries. Therefore, the latter group plays a more important role in mixing the soil and incorporating litter into the soil profile. Taken together, earthworms are thus very influential for soil structure ([Blanchart et al., 1999](#)) and subsequently for water drainage, aggregate stability, mineralization and leaching of mineral nutrients ([Edwards et al., 1989](#); [Jouquet et al., 2008](#); [Lavelle et al., 2020](#)).

It is generally considered that bioturbation tends to stabilize SOM by promoting physical protection (see [Filser et al., 2016](#)), although the deep burial of litter or casts is an often overlooked mechanism that could significantly contribute to carbon persistence in soils, also favoured by the more stable conditions ([Špaldoňová & Frouz, 2014](#)). However, some authors have highlighted that in some systems, wetter conditions in the deeper layers might accelerate SOM turnover ([Rasse et al., 2006](#)). To elucidate this, more information is needed regarding the decomposition rates of buried casts and C sequestration processes in earthworm burrow walls ([Zhang et al., 2013](#)).

Similarly, ants and termites build nests by gathering different organic and mineral materials, creating SOM hotspots. This creates soil physical and chemical heterogeneity ([Lovegrove, 1989](#); [Jouquet et al., 2002](#); [Dean, Milton & Klotz, 1999](#)). Little is known of the horizontal transportation carried out by termites during the construction of their fungus-growing chambers or those by ants with their anthills. Both ants and termites bring food to their nests (which are locally partially returned to the soil as faeces) and create fungal gardens in some chambers so that these nests often constitute patches enriched in organic matter and mineral nutrients ([Folgarait, 1998](#); [Dangerfield, McCarthy & Ellery, 1998](#)).

In agroecosystems, plant residues are artificially incorporated in soil by tillage but in natural ecosystems, besides bioturbation by fauna, the processes incorporating those materials into the soil are rather limited (i.e., soil flooding and consequent burial by mud,

burial by mineral particles brought by wind or water erosion, or cryoturbation). This is why, when macrofauna is absent, litter mostly accumulates at the soil surface, and can only reach deep soil after its physical fragmentation into small pieces and washing down by percolating water (*Bohlen et al., 2004; Hale et al., 2005*). Hence, faunal activity determines to a large extent if organic matter and processes such as decomposition mostly happens on the soil surface or in deeper soil horizons, and thus affects the amount and quality of organic matter incorporated into the soil.

### Soil engineer models

Most models on soil engineers focus on the effect of earthworms on mineral soils. Some models only tackle the demography of earthworms or their movements (*Martin & Lavelle, 1992; Klok, Van der Holt & Bodt, 2006; Pelosi et al., 2008; Vorpahl, Moenickes & Richter, 2009*), to predict their impact on soil functioning. Other models such as the Multi Agent System model, SWORM, simulate the movements of individual earthworms within a soil profile and the consequences for soil structure (*Blanchart et al., 2009*). *Barot et al. (2007)* modeled at a larger scale (about 100 m<sup>2</sup>) the feedbacks between earthworm demography and soil aggregates. Another analytical model (*Barot, Rossi & Lavelle, 2007*) allows predicting the impact of earthworm on mineral nutrient stocks and primary production from the impact of earthworms on fluxes of mineral nutrients within the ecosystems and losses of nutrients from the ecosystem (e.g., through leaching). More recently, a simulation model was developed to predict the impact of an invasive earthworm on the dynamics of soil C taking into account earthworm effects on microorganisms (*Huang et al., 2010*). In the future, this model may help predicting the speed of earthworm invasion. The activities of anecic earthworms are incorporated in the ROMUL\_Hum model (*Komarov et al., 2017; Chertov et al. (2017a, 2017b)*). There are few models tackling the impact of other soil engineers such as ants or termites on soils, except for the work by *Dangerfield, McCarthy & Ellery (1998)* on termites.

### Recommended literature:

Ants: *Folgarait (1998)*;

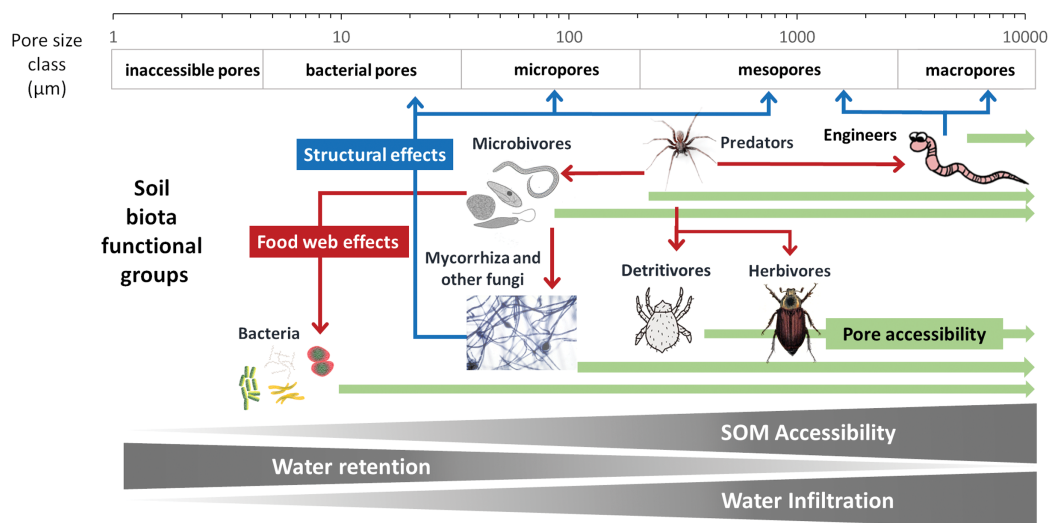
Soil fauna effects on bioturbation and aggregation: *Jiménez & Lal (2006)*;

### Model concept

#### KEYLINK general concepts

The goal of this effort is to integrate the current views on the central role of soil biota in SOM and soil water dynamics into a mechanistic model. The challenge faced was to minimize model complexity while retaining enough detail to predict and analyse effects of changes in climate and management of a very wide range of soils (grasslands, forest, agricultural soil, organic and more mineral soils) including the key processes and the key species according to the most recent insights.

From our extensive review our main conclusion is that placing chemical recalcitrance at the center of a soil model is not the best representation of soil functioning. Instead we propose soil structure as the central part of our soil model, since structure determines 'accessibility' for the dynamic soil faunal pools in terms of pore sizes and body sizes of soil



**Figure 2** The KEYLINK model representation of the soilbiota functional groups and their relationship with soil structure. Pore size distribution is represented by five classes. Blue arrows indicate effects on soil structure; green arrows indicate the range of pore classes accessible to each group; red arrows indicate trophic effects by grazing and predation. Grey triangles show the expected responses of water retention, infiltration and SOM stabilization. Image credits: spider by [Kratochvil \(2019\)](#), CC0; bacteria by [Fondren \(https://pixy.org/4793065/\)](https://pixy.org/4793065/), CC0; mite by unknown (<https://pixy.org/2176637/>), CC0; protists by [Klapper \(2012\)](#), CC0; herbivores by [Hochdanz \(1876\)](#), CC0; fungi by [CDC & Georg \(2012\)](#), CC0. Earthworm drawn by co-author (OV). Full-size DOI: 10.7717/peerj.9750/fig-2

fauna (Fig 2), but also the hydrological properties (soil water flow) and the associated temperature flow of a soil. Our key assumptions are:

- Litter and SOM decomposition are active processes, conducted by microbes and soil fauna and thus dependent on the consumer pool size.
- Decomposition depends on accessibility (function of pore size distribution and the related local soil water content and aeration) and secondly on the quality of the decomposing material.
- Pore size distribution determines the accessibility to all soil biota, but also the hydrology and the availability of  $O_2$ .
- Soil water flow depends on soil pore distribution which is also a function of the activity of soil engineers and aggregation by soil biota.
- In soils where soil engineers are important (most mineral soils), it is essential to simulate their effect on bio pore formation and bioturbation, for some organic soils their effect is less important.
- Mycorrhizal fungi need to be represented in the model regarding their interaction with the plant (important input of C to the soil), decay of SOM and effect on soil aggregation.
- In many cases a real food web, with dynamic faunal and microbial pools is necessary for example, to simulate management or climate change effects. The diversity and number of trophic levels changes with soil types/ecosystems. When there are not enough

data however, and when changes are slow (stable situation) a representation with constant pools of soil fauna can be considered.

- Special attention needs to be paid to the simulation of fine root turnover which should either include herbivory or herbivory should be simulated.
- Modelling aggregation in detail is beyond the scope of an ecosystem model, the most important effects of aggregation can be included through the concept of the pores (aggregation increasing micropore fraction and reducing mesopore fraction) as influenced by engineers (casts), bacteria and fungi.

To use the KEYLINK concept, a good hydrology model with multiple soil layers is necessary. For soils where, besides the water availability, distinct horizons are present with very different characteristics, each horizon should be simulated separately, but in other cases it can be adequate to use layers only for the hydrological calculations.

### **KEYLINK soil structure representation**

We define different pore sizes, based on measurability and accessibility by soil fauna as well as hydrological concepts. The initial pore size distribution can be calculated from water retention measurements.

Soil structure is dynamic: it can be modified by engineers, by aggregation (by bacteria and fungi which glue soil particles together), by organo-mineral interactions (function of clay content and SOM), but also by precipitation (destroying macropores and aggregates) and management (increasing bulk density). In a multi-layer soil system, bioturbation by soil engineers can be a major factor.

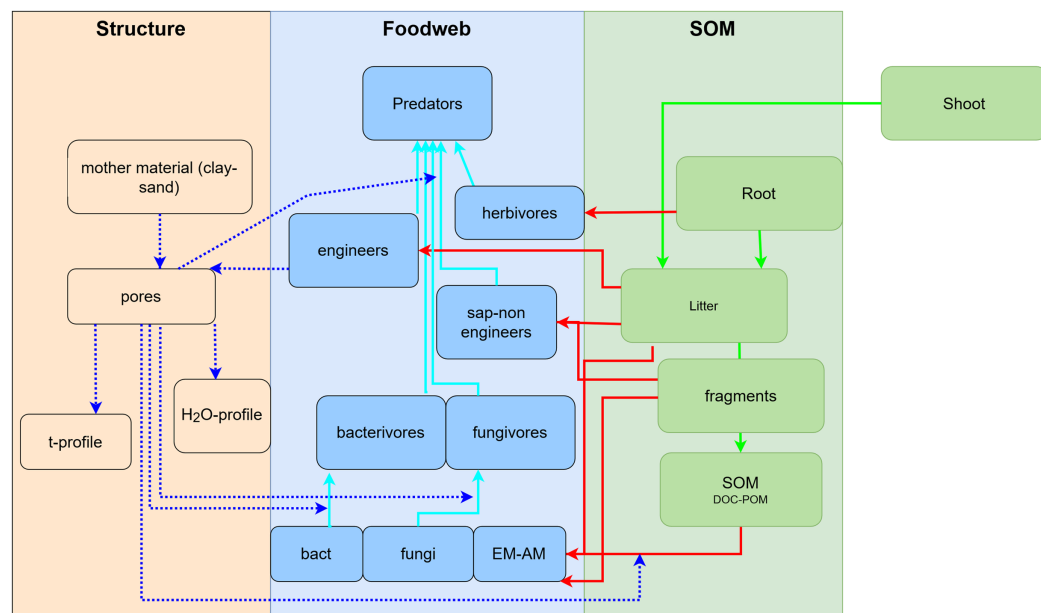
### **Description of SOM pools**

Concerning size and the main decomposing biota, a distinction between larger particulate material (fresh litter, fragments, and necromass) and SOM is required. Within SOM dissolved DOM and particulate POM need to be simulated separately to allow leaching, but can be simulated as in balance with each other. Mineral associated C is not a separate pool but a fraction of SOM depending on porosity/clay content.

Fungi and bacteria have different capabilities to decay litter. Therefore, we need to add enough description of the initial litter quality. The average recalcitrance (defined here as % non-hydrolysable compounds) and CN ratio are enough for a main division between these three pathways. SOM need not be further divided into pools. However, SOM is distributed across the pore space and depending on the pore size distribution, it is more or less accessible to decomposers. Accessibility is defined by pore size distribution by calculating the surface area of each pore fraction at each time step, and distributing the soil SOM across this area.

### **Soil biota**

We opted for a minimal complexity with which we can still include the best understood faunal and food web effects, the important distinction between the bacterial and fungal



**Figure 3** KEYLINK model concept. Full arrows, fluxes; dotted arrows, influences.

Full-size  DOI: 10.7717/peerj.9750/fig-3

pathway as well as the potential feedback effects of management in reducing food web complexity. The main division is based on function, not family or size.

- Non-mycorrhizal fungi
- Bacteria
- Mycorrhizal fungi
- Fungivores and bacterivores (or total microbivores)
- Predators
- Root herbivores
- Detritivores (non-engineers)
- Engineer detritivores

The different roles of all biota are summarized in Fig. 1. Engineers are part of the food web, and in addition create biopores and casts (changing accessibility by reducing pore size within the cast), and bioturbate the soil.

In our view, the most simple soil model can ignore all changes in chemistry apart from the initial litter quality, and decay is calculated from pore size distribution and environmental parameters (in combination with consumer pool size) (Negassa *et al.*, 2015; Strong *et al.*, 2004; Toosi *et al.*, 2017). However, for a more complete model, all biota can change “recalcitrance” and CN ratio of the material they consume by producing faeces that are more stable. All biota respire and become necromass that enters the SOM. The interaction between the biota is shown in Fig. 3. Since the goal is to simulate the response of the soil functioning to climate and management, the soil fauna need to be responsive to both. We suggest calculating the faunal pools as a set of linear equations with

the change in the pool size dependent on growth, respiration (depending on  $t$ ), faeces (including exoskeletons), and turnover (natural death and predation). Growth can be calculated as a function of maximal growth rate, resource availability (as a function of pore sizes) and quality, and environmental parameters ( $t$  and pH). The CN ratio and sensitivity to pH and  $t$ , as well as respiration rates and fecal production need to be included for each biota.

### **Model application and parameterisation**

The KEYLINK model has been implemented (Flores et al., 2020, unpublished data) and data are available to allow its development. However, full validation of the concept requires some crucial data which are missing in many experiments. For example, many ecosystem studies do not include soil fauna data at all, or only the diversity but not the abundance or biomass. While earthworms have been quite intensively studied the effect of termites and ants on soil C dynamics is less known. On the other hand, experiments focusing on soil fauna often do not include crucial data concerning the ecosystem such as litter quantity and quality, and fine root biomass and turnover. Soil structure and hydrology are very seldom described in detail, in many cases limited to sand and clay content and bulk density. Concerning hydrology, preferential flow through biopores is seldom taken into account. For a better representation of N availability, models on nitrifying/nitrogen-fixing bacteria would be necessary. For many other nutrients (including P), representation of the mineral weathering and the adsorption/desorption including a dynamic pH model would be required but in many cases data are lacking to parameterise such models. To evaluate our concept, data from isotope studies could be of great value, especially if they include the faunal food web as well as the microbial composition, the fine roots and the mycorrhizal fungi.

This very general model concept should be parameterized and implemented differently according to the specific ecosystem but will allow comparison across these different systems (which is not possible using most current models that focus on specific ecosystems (ANAFOR—[Deckmyn et al. \(2008\)](#), [Franklin et al. \(2014\)](#)) or grasslands (PaSim—[Sándor et al. \(2016\)](#))).

In organic soils, a focus on chemical decomposition can yield adequate results if the different pathways are included in an active way (microbes divided between bacteria, fungi and mycorrhizal fungi with different characteristics and efficiencies for transforming different food sources). For such soils, it is important to know at least the CN ratio and the ‘recalcitrance’ and to include the interaction between mycorrhizal fungi and plants. Inclusion of faunal effects (the composition will depend on C content and hence pH) and improved hydrological description (requiring structural description of the soil) should be able to improve the modelling results. For very wet soils (e.g., peatlands), it is clear that a correct distinction between anaerobic and aerobic processes should be included.

In the case of mineral or organo-mineral soils, the incorporation of pore-size distribution in the mineral layers will better describe the (in)accessibility of SOM due to physical inaccessibility (only bacteria can access the smallest pores, and they cannot be consumed by bacterivores in these pores) or due to water or oxygen availability. Here, the

role of soil ecosystem engineers would be crucial. In reality, the structural diversity of a soil is extremely important. A precise model would need to include a full 3D description of the rhizosphere which is beyond the scope of an ecosystem model at the scale we envisage. However, some aspects can be included by simulating root exudates as 100% accessible.

Concerning nutrients, the described model concept is limited to the nutrients available from SOM decay and ignores mineral weathering. Improved understanding of the interactions between the different soil biota and the soil geochemistry could enhance this concept, for example including the weathering effect of mycorrhizal fungi ([Andrews et al., 2011](#)), but available studies are as yet limited. For less soluble nutrients such as P depending on the parent material, pH and concentration of base cations, a more chemical approach (including the simulation of pH depending on parent material) might be necessary but hard to parameterize at an ecosystem scale, although [Yang et al. \(2019\)](#) showed this can give good results at a regional scale. In many cases an empirical approach as used in [Bortier et al. \(2018\)](#) could be added, for example for podzol soils where nutrient availability is low.

For the faunal food web, we have chosen to represent functional groups, instead of species. For the parameterization of these groups, average values of the main species can be used, as described in the sections above.

We describe a single layer here, but it is the goal to simulate the distinct horizons of a soil, since using average values when the soil horizons are strongly differentiated induces large errors. For hydrological simulations distinct soil layers need to be distinguished even if their composition is similar.

## CONCLUSIONS

Recent technological advances such as high-throughput DNA sequencing and stable isotopes analyses have greatly increased our knowledge and understanding of the key soil processes and how they interlink. Yet, the key interactions between major actors in the soil are often ignored in widely used soil models, and are only represented in complex models, focusing only on specific processes but not on ecosystem functioning.

Our model concept KEYLINK is a novel and simple yet integrative representation of the latest insights from different ‘schools’ of soil description and analyses. By including and linking the major faunal groups, the description of the soil pore space and the active decomposition of SOM, a dynamic link between management, climate and soil functioning is attainable. More insight into the interaction between the different soil biota, soil chemistry and soil structure is required to improve and validate this concept.

The strength of our concept goes beyond getting a more reliable prediction of soil processes. It is clear that, due to the limited available data for many sites, in many cases a very simplistic representation of the soil can, with site-specific parameterization, yield a reasonable fit to measured data. Indeed, given enough parameters and pools, and limited validation data, almost any model can “fit”. However, existing models, in which the growth of plants is limited by soil nutrient and water content only, create the false impression that adding nutrients and water is enough to have a well-functioning ecosystem. This is in contrast to all recent findings concerning the importance of a



well-functioning soil ecosystem including a diverse soil fauna that efficiently buffers the nutrient and water availability. Therefore, we believe that our model concept stimulates viewing the soil as a complex living system that needs to be protected in its diversity so it can fulfil all ecosystem functions.

## ADDITIONAL INFORMATION AND DECLARATIONS

### Funding

This work was conducted within COST Actions BIOLINK (FP1305) and KEYSOM (ES 1406). Gaby Deckmyn received funding from the Belgian Science Policy MASC (BR/121/A2/MASC ) and ECORISK (SD/RI/06A). This study was also co-financed by the research project J4-1766 “Methodology approaches in genome-based diversity and ecological plasticity study of truffles from their natural distribution areas” and the Research Program in Forest Biology, Ecology and Technology (P4-0107) of the Slovenian Research Agency. Jorge Curiel Yuste received funding by the Basque Government through the BERC 2018-2021 program, and by the Spanish Ministry of Science, Innovation and Universities through the BC3 María de Maeztu excellence accreditation (MDM-2017-0714). Oskar Franklin was funded by Knut and Alice Wallenberg Foundation. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

### Grant Disclosures

The following grant information was disclosed by the authors:

COST Actions BIOLINK (FP1305).

KEYSOM (ES 1406).

Belgian Science Policy MASC: BR/121/A2/MASC.

ECORISK: SD/RI/06A.

Slovenian Research Agency: J4-1766 and P4-0107.

Basque Government: BERC 2018-2021.

Spanish Ministry of Science, Innovation and Universities: MDM-2017-0714.

Knut and Alice Wallenberg Foundation.

### Competing Interests

Jorge Curiel Yuste is an Academic Editor for PeerJ. Andrea Schnepf, Harry Vereecken, and Mathieu Javaux are employed by Forschungszentrum Jülich GmbH.

### Author Contributions

- Gaby Deckmyn conceived and designed the experiments, performed the experiments, prepared figures and/or tables, and approved the final draft.
- Omar Flores conceived and designed the experiments, prepared figures and/or tables, and approved the final draft.
- Mathias Mayer performed the experiments, analyzed the data, prepared figures and/or tables, and approved the final draft.

- Xavier Domene performed the experiments, analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Andrea Schnepf analyzed the data, prepared figures and/or tables, and approved the final draft.
- Katrin Kuka analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Kris Van Looy analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Daniel P. Rasse analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Maria J.I. Briones analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Sébastien Barot analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Matty Berg analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Elena Vanguelova performed the experiments, analyzed the data, prepared figures and/or tables, and approved the final draft.
- Ivika Ostonen analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Harry Vereecken analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Laura M. Suz performed the experiments, analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Beat Frey analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Aline Frossard analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Alexei Tiunov analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Jan Frouz analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Tine Grebenc analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Maarja Õpik analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Mathieu Javaux analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Alexei Uvarov analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Olga Vindušková conceived and designed the experiments, analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.

- Paul Henning Krogh analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Oskar Franklin analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Juan Jiménez analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.
- Jorge Curiel Yuste conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, and approved the final draft.

### Data Availability

The following information was supplied regarding data availability:

This is a model concept presented in a literature review, there is no code and no data.

### REFERENCES

- Abdel-Hamid AM, Solbiati JO, Cann IKO. 2013.** Insights into lignin degradation and its potential industrial applications. In: Sariaslani S, Geoffrey MG, eds. *Advances in Applied Microbiology*. Waltham: Academic Press, 1–28.
- Abramoff RZ, Finzi AC. 2015.** Are above- and below-ground phenology in sync? *New Phytologist* **205**(3):1054–1061.
- Abramoff R, Xu X, Hartman M, O'Brien S, Feng W, Davidson E, Finzi A. 2018.** The Millennial model: in search of measurable pools and transformations for modeling soil carbon in the new century. *Biogeochemistry* **137**(1–2):51–71 DOI [10.1007/s10533-017-0409-7](https://doi.org/10.1007/s10533-017-0409-7).
- Adamczyk B, Sietio OM, Strakova P, Prommer J, Wild B, Hagner M, Pihlatie M, Fritze H, Richter A, Heinonsalo J. 2019.** Plant roots increase both decomposition and stable organic matter formation in boreal forest soil. *Nature Communications* **10**(1):3982 DOI [10.1038/s41467-019-11993-1](https://doi.org/10.1038/s41467-019-11993-1).
- Adhikari K, Hartemink AE. 2016.** Linking soils to ecosystem services—A global review. *Geoderma* **262**:101–111 DOI [10.1016/j.geoderma.2015.08.009](https://doi.org/10.1016/j.geoderma.2015.08.009).
- Ahrens B, Braakhekke MC, Guggenberger G, Schrumpf M, Reichstein M. 2015.** Contribution of sorption, DOC transport and microbial interactions to the 14C age of a soil organic carbon profile: Insights from a calibrated process model. *Soil Biology and Biochemistry* **88**:390–402 DOI [10.1016/j.soilbio.2015.06.008](https://doi.org/10.1016/j.soilbio.2015.06.008).
- Aitkenhead MJ, Albanito F, Jones MB, Black HIJ. 2011.** Development and testing of a process-based model (MOSES) for simulating soil processes, functions and ecosystem services. *Ecological Modelling* **222**(20–22):3795–3810 DOI [10.1016/j.ecolmodel.2011.09.014](https://doi.org/10.1016/j.ecolmodel.2011.09.014).
- Allison SD. 2005.** Cheaters, diffusion and nutrients constrain decomposition by microbial enzymes in spatially structured environments. *Ecology Letters* **8**(6):626–635 DOI [10.1111/j.1461-0248.2005.00756.x](https://doi.org/10.1111/j.1461-0248.2005.00756.x).
- Andersen DC. 1987.** Below-ground herbivory in natural communities: a review emphasizing fossorial animals. *Quarterly Review of Biology* **62**(3):261–286 DOI [10.1086/415512](https://doi.org/10.1086/415512).
- Anderson JM, Healey IN. 1972.** Seasonal and inter-specific variation in major components of the gut contents of some woodland Collembola. *Journal of Animal Ecology* **41**(2):359–368 DOI [10.2307/3473](https://doi.org/10.2307/3473).
- Andrews MY, Leake JR, Palmer BG, Banwart SA, Beerling DJ. 2011.** Plant and mycorrhizal driven silicate weathering: quantifying carbon flux and mineral weathering processes at the laboratory mesocosm scale. *Applied Geochemistry* **26**:314–316.

- Angst Š, Mueller CW, Cajthaml T, Angst G, Lhotáková Z, Bartuška M, Špaldonová A, Frouz J. 2017. Stabilization of soil organic matter by earthworms is connected with physical protection rather than with chemical changes of organic matter. *Geoderma* **289**:29–35 DOI [10.1016/j.geoderma.2016.11.017](https://doi.org/10.1016/j.geoderma.2016.11.017).
- Angst G, Mueller CW, Prater I, Angst Š, Frouz J, Jílková V, Peterse F, Nierop KGJ. 2019. Earthworms act as biochemical reactors to convert labile plant compounds into stabilized soil microbial necromass. *Communications Biology* **2**(1):441 DOI [10.1038/s42003-019-0684-z](https://doi.org/10.1038/s42003-019-0684-z).
- Averill C. 2016. Slowed decomposition in ectomycorrhizal ecosystems is independent of plant chemistry. *Soil Biology and Biochemistry* **102**:52–54 DOI [10.1016/j.soilbio.2016.08.003](https://doi.org/10.1016/j.soilbio.2016.08.003).
- Averill C, Turner BL, Finzi AC. 2014. Mycorrhiza-mediated competition between plants and decomposers drives soil carbon storage. *Nature* **505**(7484):543–545 DOI [10.1038/nature12901](https://doi.org/10.1038/nature12901).
- Baas Becking LGM. 1934. *Geobiologie of inleiding tot de milieukunde*. Hague: W.P. Van Stockum & Zoon.
- Badri DV, Vivanco JM. 2009. Regulation and function of root exudates. *Plant, Cell & Environment* **32**(6):666–681 DOI [10.1111/j.1365-3040.2009.01926.x](https://doi.org/10.1111/j.1365-3040.2009.01926.x).
- Bago B, Pfeffer PE, Shachar-Hill Y. 2000. Carbon metabolism and transport in arbuscular mycorrhizas. *Plant Physiology* **124**(3):949–957 DOI [10.1104/pp.124.3.949](https://doi.org/10.1104/pp.124.3.949).
- Baldrian P, Valášková V. 2008. Degradation of cellulose by basidiomycetous fungi. *FEMS Microbiology Reviews* **32**(3):501–521 DOI [10.1111/j.1574-6976.2008.00106.x](https://doi.org/10.1111/j.1574-6976.2008.00106.x).
- Bandounas L, Wierckx NJ, De Winde JH, Ruijsenaars HJ. 2011. Isolation and characterization of novel bacterial strains exhibiting ligninolytic potential. *BMC Biotechnology* **11**(1):94 DOI [10.1186/1472-6750-11-94](https://doi.org/10.1186/1472-6750-11-94).
- Bardgett RD, Usher MB, Hopkins DW. 2005. *Biological diversity and function in soils*. Cambridge: Cambridge University Press, 411.
- Bardgett RD, Van der Putten WH. 2014. Belowground biodiversity and ecosystem functioning. *Nature* **515**:505–511.
- Barot S, Blouin M, Fontaine S, Jouquet P, Lata J-C, Mathieu J. 2007. A tale of four stories: soil ecology, theory, evolution and the publication system. *PLOS ONE* **2**(11):1248 DOI [10.1371/journal.pone.0001248](https://doi.org/10.1371/journal.pone.0001248).
- Barot S, Rossi JP, Lavelle P. 2007. Self-organization in a simple consumer–resource system, the example of earthworms. *Soil Biology and Biochemistry* **39**(9):2230–2240 DOI [10.1016/j.soilbio.2007.03.021](https://doi.org/10.1016/j.soilbio.2007.03.021).
- Baskaran P, Hyvönen R, Berglund SL, Clemmensen KE, Ågren GI, Lindahl BD, Manzoni S. 2017. Modelling the influence of ectomycorrhizal decomposition on plant nutrition and soil carbon sequestration in boreal forest ecosystems. *New Phytologist* **213**(3):1452–1465 DOI [10.1111/nph.14213](https://doi.org/10.1111/nph.14213).
- Bastardie F, Cannavacciuolo M, Capowiez Y, Dreuzy J-R, Bellido A, Cluzeau D. 2002. A new simulation for modelling the topology of earthworm burrow systems and their effects on macropore flow in experimental soils. *Biology and Fertility of Soils* **36**(2):161–169 DOI [10.1007/s00374-002-0514-0](https://doi.org/10.1007/s00374-002-0514-0).
- Bastardie F, Capowiez Y, Cluzeau D. 2005. 3D characterisation of earthworm burrow systems in natural soil cores collected from a 12-year-old pasture. *Applied Soil Ecology* **30**(1):34–46 DOI [10.1016/j.apsoil.2005.01.001](https://doi.org/10.1016/j.apsoil.2005.01.001).
- Belfield W. 1956. The arthropoda of the soil in a West African pasture. *Journal of Animal Ecology* **25**(2):275–287 DOI [10.2307/1926](https://doi.org/10.2307/1926).

- Bengtsson J, Setälä H, Zheng DW. 1996.** Food webs and nutrient cycling in soils: interactions and positive feedbacks. In: Polis GA, Winemiller KO, eds. *Food Webs*. Boston: Springer.
- Berg M, De Ruiter PC, Didden WAM, Janssen MPM, Schouten AJ, Verhoef HA. 2001.** Community food web, decomposition and nitrogen mineralisation in a stratified Scots pine forest soil. *Oikos* **94**(1):130–142 DOI [10.1034/j.1600-0706.2001.09121.x](https://doi.org/10.1034/j.1600-0706.2001.09121.x).
- Beven K. 2018.** A century of Denial: preferential and nonequilibrium water flow in soils, 1864–1984. *Vadose Zone Journal* **17**(1):180153 DOI [10.2136/vzj2018.08.0153](https://doi.org/10.2136/vzj2018.08.0153).
- Bever JD. 2003.** Soil community feedback and the coexistence of competitors: conceptual frameworks and empirical tests. *New Phytologist* **157**(3):465–473 DOI [10.1046/j.1469-8137.2003.00714.x](https://doi.org/10.1046/j.1469-8137.2003.00714.x).
- Blanc C, Sy M, Djigal D, Brauman A, Normand P, Villenave C. 2006.** Nutrition on bacteria by bacterial-feeding nematodes and consequences on the structure of soil bacterial community. *European Journal of Soil Biology* **42**:70–78 DOI [10.1016/j.ejsobi.2006.06.003](https://doi.org/10.1016/j.ejsobi.2006.06.003).
- Blanchart E, Albrecht A, Alegre J, Dubois A, Giloe C, Pashanasf B, Lavelle P, Brussaard L. 1999.** Effects of earthworms on soil structural and physical properties. In: Lavelle P, Brussaard L, Hendrix P, eds. *Earthworm management in tropical agroecosystems*. New York: CABI Publishing, 149–172.
- Blanchart E, Marilleau N, Chotte JL, Drogoul A, Perrier E, Cambier C. 2009.** SWORM: an agent-based model to simulate the effect of earthworms on soil structure. *European Journal of Soil Science* **60**(1):13–21 DOI [10.1111/j.1365-2389.2008.01091.x](https://doi.org/10.1111/j.1365-2389.2008.01091.x).
- Bohlen PJ, Groffman PM, Fahey TJ, Fisk MC, Suárez E, Pelletier DM, Fahey RT. 2004.** Ecosystem consequences of exotic earthworm invasion of north temperate forests. *Ecosystems* **7**(1):1–12 DOI [10.1007/s10021-003-0126-z](https://doi.org/10.1007/s10021-003-0126-z).
- Bortier M, Andivia E, Genon JG, Grebenc T, Deckmyn G. 2018.** Towards understanding the role of ectomycorrhizal fungi in forest phosphorus cycling: a modelling approach. *Central European Forestry Journal* **64**(2):79–95 DOI [10.1515/forj-2017-0037](https://doi.org/10.1515/forj-2017-0037).
- Bossuyt H, Six J, Hendrix PF. 2005.** Protection of soil carbon by microaggregates within earthworm casts. *Soil Biology and Biochemistry* **37**(2):251–258 DOI [10.1016/j.soilbio.2004.07.035](https://doi.org/10.1016/j.soilbio.2004.07.035).
- Bottinelli N, Jouquet P, Capowicz Y, Podwojewski P, Grimaldi M, Peng X. 2015.** Why is the influence of soil macrofauna on soil structure only considered by soil ecologists? *Soil and Tillage Research* **146**:118–124 DOI [10.1016/j.still.2014.01.007](https://doi.org/10.1016/j.still.2014.01.007).
- Bottomley PJ, Taylor AE, Myrold DD. 2012.** A consideration of the relative contributions of different microbial subpopulations to the soil N cycle. *Frontiers in Microbiology* **3**:373 DOI [10.3389/fmicb.2012.00373](https://doi.org/10.3389/fmicb.2012.00373).
- Bouché M. 1977.** Strategies lombriciennes. *Ecological Bulletins* **25**:122–132.
- Bouma J. 1989.** Using soil survey data for quantitative land evaluation. *Advanced Plant & Soil Science* **9**:177–213.
- Briones MJI. 2014.** Soil fauna and soil functions: a jigsaw puzzle. *Frontiers in Environmental Science* **2**:125 DOI [10.3389/fenvs.2014.00007](https://doi.org/10.3389/fenvs.2014.00007).
- Briones MJI, Carreira J, Ineson P. 1998.** Cognettia sphagnetorum (Enchytraeidae) and nutrient cycling in organic soils: a microcosm experiment. *Applied Soil Ecology* **9**(1–3):289–294 DOI [10.1016/S0929-1393\(97\)00055-3](https://doi.org/10.1016/S0929-1393(97)00055-3).
- Briones MJI, Ineson P. 2002.** Use of <sup>14</sup>C carbon dating to determine feeding behaviour of enchytraeids. *Soil Biology & Biochemistry* **34**(6):881–884 DOI [10.1016/S0038-0717\(02\)00010-X](https://doi.org/10.1016/S0038-0717(02)00010-X).

- Briones MJI, Ineson P, Heinemeyer A. 2007.** Predicting potential impacts of climate change on the geographical distribution of enchytraeids: a meta-analysis approach. *Global Change Biology* 13(11):2252–2269 DOI 10.1111/j.1365-2486.2007.01434.x.
- Briones MJI, Ineson P, Pearce TG. 1997.** Effects of climate change on soil fauna; responses of enchytraeids, Diptera larvae and tardigrades in a transplant experiment. *Applied Soil Ecology* 6(2):117–134 DOI 10.1016/S0929-1393(97)00004-8.
- Briones MJI, Ineson P, Poskitt J. 1998.** Climate change and *Cognettia sphagnetorum*: effects on carbon dynamics in organic soils. *Functional Ecology* 12(4):528–535 DOI 10.1046/j.1365-2435.1998.00218.x.
- Briones MJI, Poskitt J, Ostle N. 2004.** Influence of warming and enchytraeid activities on soil CO<sub>2</sub> and CH<sub>4</sub> fluxes. *Soil Biology and Biochemistry* 36(11):1851–1859 DOI 10.1016/j.soilbio.2004.04.039.
- Briones M, Ostle N, Garnett M. 2007.** Invertebrates increase the sensitivity of non-labile soil carbon to climate change. *Soil Biology and Biochemistry* 39:816–818 DOI 10.1016/j.soilbio.2006.09.007.
- Brown GG. 1995.** How do earthworms affect microfloral and faunal community diversity? *Plant and Soil* 170(1):209–231 DOI 10.1007/BF02183068.
- Brundrett MC, Tedersoo L. 2018.** Evolutionary history of mycorrhizal symbioses and global host plant diversity. *New Phytologist* 220(4):1108–1115 DOI 10.1111/nph.14976.
- Brunner I, Bakker MR, Björk RG, Hirano Y, Lukac M, Aranda X, Børja I, Eldhuset TD, Helmisaari HS, Jourdan C, Konôpka B, López BC, Miguel Pérez C, Persson H, Ostonen I. 2013.** Fine-root turnover rates of European forests revisited: an analysis of data from sequential coring and ingrowth cores. *Plant and Soil* 362(1–2):357–372 DOI 10.1007/s11104-012-1313-5.
- Brussaard L. 1997.** Biodiversity and ecosystem functioning in soil. *Ambio* 26:563–570.
- Bödeker ITM, Lindahl BD, Olson A, Clemmensen KE. 2016.** Mycorrhizal and saprotrophic fungal guilds compete for the same organic substrates but affect decomposition differently. *Functional Ecology* 30(12):1967–1978 DOI 10.1111/1365-2435.12677.
- Bödeker ITM, Nygren CM, Taylor AF, Olson Å, Lindahl BD. 2009.** ClassII peroxidase-encoding genes are present in a phylogenetically wide range of ectomycorrhizal fungi. *ISME Journal* 3(12):1387–1395 DOI 10.1038/ismej.2009.77.
- Bünemann EK, Giulia Bongiorno G, Zhanguo Bai Z, Rachel E, Creamer RE, De Deyn G, De Goede R, Luuk Flesskens L, Geissen V, Kuyper TW, Mäder P, Pulleman M, Sukkel W, Van Groenigen JW, Lijbert Brussaard L. 2018.** Soil quality—a critical review. *Soil Biology and Biochemistry* 120:105–125 DOI 10.1016/j.soilbio.2018.01.030.
- Cairney JWG. 2012.** Extramatrical mycelia of ectomycorrhizal fungi as moderators of carbon dynamics in forest soil. *Soil Biology and Biochemistry* 47:198–208 DOI 10.1016/j.soilbio.2011.12.029.
- Camino-Serrano M, Guenet B, Luyssaert S, Ciais P, Bastrikov V, DeVos B, Gielen B, Gleixner G, Jorner-Puig A, Kaiser K, Kothawala D, Lauerwald R, Peñuelas J, Schrumppf M, Vicca S, Vuichar N, Walmsley D, Janssens IA. 2018.** ORCHIDEE-SOM: modeling soil organic carbon (SOC) and dissolved organic carbon (DOC) dynamics along vertical soil profiles in Europe. *Geoscientific Model Development* 11(3):937–957 DOI 10.5194/gmd-11-937-2018.
- Campbell EE, Parton WJ, Soong JL, Paustian K, Hobb NT, Cotrufo MF. 2016.** Using litter chemistry controls on microbial processes to partition litter carbon fluxes with the Litter Decomposition and Leaching (LIDEL) model. *Soil Biology and Biochemistry* 100:160–174 DOI 10.1016/j.soilbio.2016.06.007.

- Campbell EE, Paustian K. 2015.** Current developments in soil organic matter modeling and the expansion of model applications: a review. *Environmental Research Letters* **10(12)**:123004 DOI [10.1088/1748-9326/10/12/123004](https://doi.org/10.1088/1748-9326/10/12/123004).
- Canarini A, Kaiser C, Merchant A, Richter A, Wanek W. 2019.** Root exudation of primary metabolites: mechanisms and their roles in plant responses to environmental stimuli. *Frontiers in Plant Science* **10**:157 DOI [10.3389/fpls.2019.00157](https://doi.org/10.3389/fpls.2019.00157).
- Capowiez Y, Cadoux S, Bouchand P, Ruy S, Roger-Estrade J, Richard G, Boizard H. 2009.** The influence of tillage type and compaction on earthworm communities and the consequences for macroporosity and water infiltration in crop fields. *Soil and Tillage Research* **105(2)**:209–216 DOI [10.1016/j.still.2009.09.002](https://doi.org/10.1016/j.still.2009.09.002).
- Capowiez Y, Sammartino S, Michel E. 2014.** Burrow systems of endogeic earthworms: effects of earthworm abundance and consequences for soil water infiltration. *Pedobiologia* **57(4–6)**:303–309 DOI [10.1016/j.pedobi.2014.04.001](https://doi.org/10.1016/j.pedobi.2014.04.001).
- Carrera N, Barreal ME, Gallego PP, Briones MJI. 2009.** Soil invertebrates control peatland C fluxes in response to warming. *Functional Ecology* **23(3)**:637–648 DOI [10.1111/j.1365-2435.2009.01560.x](https://doi.org/10.1111/j.1365-2435.2009.01560.x).
- Carrera C, Martínez MJ, Dardanelli J, Balzarini M. 2011.** Environmental variation and correlation of seed components in nontransgenic soybeans: protein, oil, unsaturated fatty acids, tocopherols, and isoflavones. *Crop Science* **51(2)**:800–809 DOI [10.2135/cropsci2010.06.0314](https://doi.org/10.2135/cropsci2010.06.0314).
- CDC, Georg LK. 2012.** Public Domain Picture: This micrograph reveals the terminal chlamydospores of the fungus *Microsporium audouinii*. Available at [http://www.publicdomainfiles.com/show\\_file.php?id=13529099413553](http://www.publicdomainfiles.com/show_file.php?id=13529099413553).
- Chenu C, Plante AF. 2006.** Clay-sized organo-mineral complexes in a cultivation chronosequence: revisiting the concept of the ‘primary organo-mineral complex’. *European Journal of Soil Science* **57(4)**:596–607 DOI [10.1111/j.1365-2389.2006.00834.x](https://doi.org/10.1111/j.1365-2389.2006.00834.x).
- Chertov O, Komarov A, Shaw C, Bykhovets S, Frolov P, Shanin V, Grabarnik P, Priputina I, Zubkova E, Shashkov M. 2017a.** Romul\_Hum—a model of soil organic matter formation coupling with soil biota activity: II. Parameterisation of the soil food web biota activity activity. *Ecological Modelling* **345**:140–149.
- Chertov O, Shaw C, Shashkov M, Komarov A, Bykhovets S, Shanin V, Grabarnik P, Frolov P, Kalinina O, Priputina I, Zubkova E. 2017b.** Romul\_Hum model of soil organic matter formation coupled with soil biota activity. III. Parameterisation of earthworm. *Ecological Modelling* **345**:125–139 DOI [10.1016/j.ecolmodel.2016.10.024](https://doi.org/10.1016/j.ecolmodel.2016.10.024).
- Coleman DC, Hendrix PF. 2000.** *Invertebrates as webmasters in ecosystems*. Wallingford: CABI Publishing.
- Collins NJ, Baker JH, Tilbrook PJ. 1975.** Signy Island, maritime Antarctic. In: Rosswall T, Heal OW, eds. *Structure and Function of Tundra Ecosystems*. Stockholm: Swedish Natural Research Council, 345–374.
- Cools N, Vesterdal L, De Vos B, Vanguelova E, Hansen K. 2014.** Tree species is the major factor explaining C:N ratios in European forest soils. *Forest Ecology and Management* **311**:3–16 DOI [10.1016/j.foreco.2013.06.047](https://doi.org/10.1016/j.foreco.2013.06.047).
- Costa OYA, Raaijmakers JM, Kuramae EE. 2018.** Microbial extracellular polymeric substances: ecological function and impact on soil aggregation. *Frontiers in Microbiology* **9**:1636 DOI [10.3389/fmicb.2018.01636](https://doi.org/10.3389/fmicb.2018.01636).
- Cotrufo MF, Wallenstein MD, Boot CM, Deneff K, Paul E. 2013.** The microbial efficiency-matrix stabilization (MEMS) framework integrates plant litter decomposition with soil organic matter

- stabilization: do labile plant inputs form stable soil organic matter? *Global Change Biology* **19**(4):988–995 DOI [10.1111/gcb.12113](https://doi.org/10.1111/gcb.12113).
- Coulis M, Hättenschwiler S, Rapior S, Coq S. 2009.** The fate of condensed tannins during litter consumption by soil animals. *Soil Biology and Biochemistry* **41**(12):2573–2578 DOI [10.1016/j.soilbio.2009.09.022](https://doi.org/10.1016/j.soilbio.2009.09.022).
- Crawford JW, Deacon L, Grinev D, Harris JA, Ritz K, Brajesh K, Singh BK, Young I. 2012.** Microbial diversity affects self-organization of the soil–microbe system with consequences for function. *Journal of the Royal Society Interface* **71**:1302–1310 DOI [10.1098/rsif.2011.0679](https://doi.org/10.1098/rsif.2011.0679).
- Curiel Yuste J, Penuels J, Estiarte M, Garcia-mas J, Mattana S, Ogaya R, Pujol M, Sardans J. 2011.** Drought-resistant fungi control soil organic matter decomposition and its response to temperature. *Global Change Biology* **17**(3):1475–1486 DOI [10.1111/j.1365-2486.2010.02300.x](https://doi.org/10.1111/j.1365-2486.2010.02300.x).
- Curry JP, Schmidt O. 2006.** The feeding ecology of earthworms—a review. *Pedobiologia* **50**(6):463–477 DOI [10.1016/j.pedobi.2006.09.001](https://doi.org/10.1016/j.pedobi.2006.09.001).
- Dalal RC, Allen DE, Livesley SJ, Richards G. 2008.** Magnitude and biophysical regulators of methane emission and consumption in the Australian agricultural, forest, and submerged landscapes: a review. *Plant and Soil* **309**(1–2):43–76 DOI [10.1007/s11104-007-9446-7](https://doi.org/10.1007/s11104-007-9446-7).
- Daly KR, Tracy SR, Crout NMJ, Mairhofer S, Pridmore TP, Mooney SJ, Roose T. 2017.** Quantification of root water uptake in soil using X-ray computed tomography and image-based modelling. *Plant, Cell & Environment* **41**(1):121–133 DOI [10.1111/pce.12983](https://doi.org/10.1111/pce.12983).
- Dangerfield JM, McCarthy TS, Ellery WN. 1998.** The mound-building termite *Macrotermes michaelseni* as an ecosystem engineer. *Journal of Tropical Ecology* **14**(4):507–520 DOI [10.1017/S0266467498000364](https://doi.org/10.1017/S0266467498000364).
- Datta R, Kelkar A, Baraniya D, Molaei A, Moulick A, Meena RS, Formánek P. 2017.** Enzymatic degradation of lignin in soil: a review. *Sustainability* **9**(7):1163 DOI [10.3390/su9071163](https://doi.org/10.3390/su9071163).
- De Boer W, Folman LB, Summerbell RC, Boddy L. 2005.** Living in a fungal world: impact of fungi on soil bacterial niche development. *FEMS Microbiology Reviews* **29**(4):795–811 DOI [10.1016/j.femsre.2004.11.005](https://doi.org/10.1016/j.femsre.2004.11.005).
- De Graaff M-A, Adkins J, Kardol P, Throop HL. 2015.** A meta-analysis of soil biodiversity impacts on the carbon cycle. *Soil* **1**(1):257–271 DOI [10.5194/soil-1-257-2015](https://doi.org/10.5194/soil-1-257-2015).
- De Ruiter PC, Neutel AM, Moore JC. 1994.** Food webs and nutrient cycling in agro-ecosystems. *Trends in Ecology & Evolution* **9**(10):378–383 DOI [10.1016/0169-5347\(94\)90059-0](https://doi.org/10.1016/0169-5347(94)90059-0).
- De Vries FT, Caruso T. 2016.** Eating from the same plate? Revisiting the role of labile carbon inputs in the soil food web. *Soil Biology and Biochemistry* **102**:4–9 DOI [10.1016/j.soilbio.2016.06.023](https://doi.org/10.1016/j.soilbio.2016.06.023).
- De Vries FT, Hoffland E, Van Eekeren N, Brussaard L, Bloem J. 2006.** Fungal/bacterial ratios in grasslands with contrasting nitrogen management. *Soil Biology and Biochemistry* **38**(8):2092–2103 DOI [10.1016/j.soilbio.2006.01.008](https://doi.org/10.1016/j.soilbio.2006.01.008).
- De Vries FT, Thebault E, Liiri M, Birkhofer K, Tsiafouli MA, Bjornlund L, Jorgensen HB, Brady MV, Christensen S, De Ruiter PC, d’Hertefeldt T, Frouz J, Hedlund K, Hemerik L, Hol WHG, Hotes S, Mortimer SR, Setälä H, Sgardelis SP, Uteseny K, Van der Putten WH, Wolters V, Bardgett RD. 2013.** Soil food web properties explain ecosystem services across European land use systems. *Proceedings of the National Academy of Sciences of the United States of America* **110**(35):14296–14301 DOI [10.1073/pnas.1305198110](https://doi.org/10.1073/pnas.1305198110).
- Dean WRJ, Milton SJ, Klotz S. 1999.** The role of ant nest-mounds in maintaining small-scale patchiness in dry grasslands in Central Germany. *Biodiversity and Conservation* **6**(9):1293–1307 DOI [10.1023/A:1018313025896](https://doi.org/10.1023/A:1018313025896).



- Decaëns T, Rangel AF, Asakawa N, Thomas RJ. 1999. Carbon and nitrogen dynamics in ageing earthworm casts in grasslands of the eastern plains of Colombia. *Biology and Fertility of Soils* 30(1–2):20–28 DOI 10.1007/s003740050582.
- Deckmyn G, Campioli M, Muys B, Kraigher H. 2011. Simulating C cycles in forest soils: Including the active role of micro-organisms in the ANAFORE forest model. *Ecological Modelling* 222(12):1972–1985 DOI 10.1016/j.ecolmodel.2011.03.011.
- Deckmyn G, Meyer A, Smits MM, Ekblad A, Grebenc T, Komarov A, Kraigher H. 2014. Simulating ectomycorrhizal fungi and their role in carbon and nitrogen cycling in forest ecosystems. *Canadian Journal of Forest Research* 44(6):535–553.
- Deckmyn G, Verbeeck H, Op de Beeck M, Vansteenkiste D, Steppe K, Ceulemans R. 2008. ANAFORE: a stand-scale process-based forest model that includes wood tissue development and labile carbon storage in trees. *Ecological Modeling* 215:345–368.
- Degens BP. 1997. Macro-aggregation of soils by biological bonding and binding mechanisms and the factors affecting these: a review. *Soil Research* 35(3):431–460 DOI 10.1071/S96016.
- Deng H. 2012. A review of diversity-stability relationship of soil microbial community: what do we not know? *Journal of Environmental Sciences* 24(6):1027–1035 DOI 10.1016/S1001-0742(11)60846-2.
- Dervash M, Bhat R, Mushtaq N, Singh V. 2018. Dynamics and importance of soil mesofauna. *International Journal of Advance Research in Science and Engineering* 7(4). Available at [https://www.researchgate.net/publication/325119590\\_Dynamics\\_and\\_Importance\\_of\\_Soil\\_Mesofauna](https://www.researchgate.net/publication/325119590_Dynamics_and_Importance_of_Soil_Mesofauna).
- Didden W. 1993. Ecology of terrestrial Enchytraeidae. *Pedobiologia* 37:2–29.
- Dignac M, Derrien D, Barré P, Barot S, Cécillon L, Chenu C, Chevallier T, Freschet GT, Garnier P, Guenet B, Hedde M, Klumpp K, Lashermes G, Maron P-A, Nunan N, Roumet C, Basile-Doelsch I. 2017. Increasing soil carbon storage: mechanisms, effects of agricultural practices and proxies—a review. *Agronomy for Sustainable Development* 37(2):14 DOI 10.1007/s13593-017-0421-2.
- Dunbabin VM, Postma JA, Schnepf A, Pagès L, Javaux M, Wu L, Leitner D, Chen YL, Rengel Z, Diggle AJ. 2013. Modelling root–soil interactions using three-dimensional models of root growth, architecture and function. *Plant and Soil* 372(1–2):93–124 DOI 10.1007/s11104-013-1769-y.
- Dungait JAJ, Hopkins DW, Gregory AS, Whitmore AP. 2012. Soil organic matter turnover is governed by accessibility not recalcitrance. *Global Change Biology* 18(6):1781–1796 DOI 10.1111/j.1365-2486.2012.02665.x.
- Duputel M, Van Hoyer F, Toucet J, Gérard F. 2013. Citrate adsorption can decrease soluble phosphate concentration in soil: experimental and modeling evidence. *Applied Geochemistry* 39:85–92 DOI 10.1016/j.apgeochem.2013.09.017.
- Dwivedi D, Riley WJ, Torn MS, Spycher N, Maggi F, Tang JY. 2017. Mineral properties, microbes, transport, and plant-input profiles control vertical distribution and age of soil carbon stocks. *Soil Biology and Biochemistry* 107:244–259 DOI 10.1016/j.soilbio.2016.12.019.
- Dzotsi KA, Jones JW, Adiku SGK, Naab JB, Singh U, Porter CH, Gijsman AJ. 2010. Modeling soil and plant phosphorus within DSSAT. *Ecological Modelling* 221(23):2839–2849 DOI 10.1016/j.ecolmodel.2010.08.023.
- Edwards WM, Shipitalo MJ, Owens LB, Norton LD. 1989. Water and nitrate movement in earthworm burrows within long-term no-till cornfields. *Journal of Soil Water Conservation* 44:240–243.

- Eissenstat DM, Wells CE, Yanai RD, Whitback JL. 2000. Building roots in a changing environment: implications for root longevity. *New Phytologist* **147**(1):33–42 DOI [10.1046/j.1469-8137.2000.00686.x](https://doi.org/10.1046/j.1469-8137.2000.00686.x).
- Ekblad A, Wallander H, Godbold DL, Cruz C, Johnson D, Baldrian P, Björk G, Epron D, Kieliszewska-Rokicka B, Kjoller R, Kraigher H, Matzner E, Neumann J, Plassard C. 2013. The production and turnover of extramatrical mycelium of ectomycorrhizal fungi in forest soils: role in carbon cycling. *Plant and Soil* **366**(1–2):1–27 DOI [10.1007/s11104-013-1630-3](https://doi.org/10.1007/s11104-013-1630-3).
- Fan J, McConkey B, Wang H, Janzen H. 2016. Root distribution by depth for temperate agricultural crops. *Field Crops Research* **189**:68–74 DOI [10.1016/j.fcr.2016.02.013](https://doi.org/10.1016/j.fcr.2016.02.013).
- Farmer D, Sivapalan M, Jothityangkoon C. 2003. Climate, soil, and vegetation controls upon the variability of water balance in temperate and semiarid landscapes: downward approach to water balance analysis. *Water Resources Research* **39**(2):1035 DOI [10.1029/2001WR000328](https://doi.org/10.1029/2001WR000328).
- Fernandez CW, Kennedy PG. 2016. Revisiting the ‘Gadgil effect’: do interguild fungal interactions control carbon cycling in forest soils? *New Phytologist* **209**(4):1382–1394 DOI [10.1111/nph.13648](https://doi.org/10.1111/nph.13648).
- Ferris H, Venette R, Van der Meulen H, Lau SS. 1998. Nitrogen mineralization by bacterial-feeding nematodes: verification and measurement. *Plant and Soil* **203**(2):159–171 DOI [10.1023/A:1004318318307](https://doi.org/10.1023/A:1004318318307).
- Fierer N, Strickland MS, Liptzin D, Bradford MA, Cleveland CC. 2009. Global patterns in belowground communities. *Ecology Letters* **12**(11):1238–1249 DOI [10.1111/j.1461-0248.2009.01360.x](https://doi.org/10.1111/j.1461-0248.2009.01360.x).
- Filser J, Faber JH, Tiunov AV, Brussaard L, Frouz J, De Deyn G, Uvarov AV, Berg MP, Lavelle P, Loreau M, Wall DH, Querner P, Eijsackers H, Jiménez JJ. 2016. Soil fauna: key to new carbon models. *Soil* **2**(4):565–582 DOI [10.5194/soil-2-565-2016](https://doi.org/10.5194/soil-2-565-2016).
- Finér L, Helmisaari H-S, Löhmus K, Majdi H, Brunner I, Børja I, Eldhuset T, Godbold D, Grebenc T, Konôpka B, Kraigher H, Möttönen M-R, Ohashi M, Oleksyn J, Ostonen I, Uri V, Vanguelova E. 2007. Variation in fine root biomass of three European tree species: beech (*Fagus sylvatica* L.), Norway spruce (*Picea abies* L. Karst.), and Scots pine (*Pinus sylvestris* L.). *Plant Biosystems—An International Journal Dealing with all Aspects of Plant Biology* **141**(3):394–405 DOI [10.1080/11263500701625897](https://doi.org/10.1080/11263500701625897).
- Finér L, Ohashi M, Noguchi K, Hirano Y. 2011. Factors causing variation in fine root biomass in forest ecosystems. *Forest Ecology and Management* **261**(2):265–277 DOI [10.1016/j.foreco.2010.10.016](https://doi.org/10.1016/j.foreco.2010.10.016).
- Folgarait PJ. 1998. Ant biodiversity and its relationship to ecosystem functioning: a review. *Biodiversity and Conservation* **7**(9):1221–1244 DOI [10.1023/A:1008891901953](https://doi.org/10.1023/A:1008891901953).
- Fontaine S, Barot S. 2005. Size and functional diversity of microbe populations control plant persistence and long-term soil carbon accumulation. *Ecology Letters* **8**(10):1075–1087 DOI [10.1111/j.1461-0248.2005.00813.x](https://doi.org/10.1111/j.1461-0248.2005.00813.x).
- Foster R. 1988. Microenvironments of soil microorganisms, Fungal biodiversity and their role in soil health. *Biology and Fertility of Soils* **6**(3):189–203 DOI [10.1007/BF00260816](https://doi.org/10.1007/BF00260816).
- Fraç M, Hannula SE, Bełka M, Jędryczka M. 2018. Fungal biodiversity and their role in soil health. *Frontiers in Microbiology* **9**:707.
- Franklin O, Näsholm T, Högborg P, Högborg MN. 2014. Forests trapped in nitrogen limitation—an ecological market perspective on ectomycorrhizal symbiosis. *New Phytologist* **203**:657–666.
- Frey S, Elliott E, Paustian K, Peterson G. 2000. Fungal translocation as a mechanism for soil nitrogen inputs to surface residue decomposition in a no-tillage agroecosystem. *Soil Biology and Biochemistry* **32**(5):689–698 DOI [10.1016/S0038-0717\(99\)00205-9](https://doi.org/10.1016/S0038-0717(99)00205-9).

- Frouz J. 2018.** Effects of soil macro-and mesofauna on litter decomposition and soil organic matter stabilization. *Geoderma* **332**:161–172 DOI [10.1016/j.geoderma.2017.08.039](https://doi.org/10.1016/j.geoderma.2017.08.039).
- Frouz J, Keplin B, Pižl V, Tajovský K, Starý J, Lukešová A, Nováková A, Balík V, Háněl L, Materna J, Düker Ch, Chalupský J, Rusek J, Heinkele T. 2001.** Soil biota and upper soil layer development in two contrasting postmining chronosequences. *Ecological Engineering* **17**(2–3):275–284 DOI [10.1016/S0925-8574\(00\)00144-0](https://doi.org/10.1016/S0925-8574(00)00144-0).
- Frouz J, Kuraz V. 2013.** Soil fauna and soil physical properties. In: Frouz J, ed. *Soil Biota and Ecosystem Development in Post Mining Sites*. Boca Raton: CRC Press, 265–278.
- Frouz J, Novakova A, Jones TH. 2002.** The potential effect of high atmospheric CO<sub>2</sub> on soil fungi-invertebrate interactions. *Global Change Biology* **8**(4):339–344 DOI [10.1046/j.1354-1013.2001.00474.x](https://doi.org/10.1046/j.1354-1013.2001.00474.x).
- Frouz J, Roubíčková A, Heděnc P, Tajovský K. 2015.** Do soil fauna really hasten litter decomposition? A meta-analysis of enclosure studies. *European Journal of Soil Biology* **68**:264 DOI [10.1016/j.ejsobi.2015.03.002](https://doi.org/10.1016/j.ejsobi.2015.03.002).
- Frouz J, Santruckova H, Elhottova D. 1999.** The effect of bionid larvae feeding on the microbial community of litter and on reconsumed excrements. *Pedobiologia* **43**(3):221–230.
- Frouz J, Šimek M. 2009.** Short term and long term effects of bionid (Diptera: Bibionidae) larvae feeding on microbial respiration and alder litter decomposition. *European Journal of Soil Biology* **45**(2):192–197 DOI [10.1016/j.ejsobi.2008.09.012](https://doi.org/10.1016/j.ejsobi.2008.09.012).
- Gao Z, Karlsson I, Geisen S, Kowalchuk G, Jousset A. 2019.** Protists: puppet masters of the rhizosphere microbiome. *Trends in Plant Science* **24**(2):165–176 DOI [10.1016/j.tplants.2018.10.011](https://doi.org/10.1016/j.tplants.2018.10.011).
- Geethanjali P, Jayashankar M. 2016.** A review on litter decomposition by soil fungal community. *IOSR Journal of Pharmacy and Biological Sciences* **11**(4):1–3 DOI [10.9790/3008-1104030103](https://doi.org/10.9790/3008-1104030103).
- Geisen S. 2016.** The bacterial-fungal energy channel concept challenged by enormous functional versatility of soil protists. *Soil Biology and Biochemistry* **102**:22–25 DOI [10.1016/j.soilbio.2016.06.013](https://doi.org/10.1016/j.soilbio.2016.06.013).
- Geisen S, Briones MJ, Gan H, Behan-Pelletier VM, Friman VP, De Groot GA, Hannula SE, Lindo Z, Philippot L, Tiunov AV, Wall DH. 2019.** A methodological framework to embrace soil biodiversity. *Soil Biology and Biochemistry* **136**:107536 DOI [10.1016/j.soilbio.2019.107536](https://doi.org/10.1016/j.soilbio.2019.107536).
- Gleixner G. 2013.** Soil organic matter dynamics: a biological perspective derived from the use of compound-specific isotopes studies. *Ecological Research* **28**(5):683–695 DOI [10.1007/s11284-012-1022-9](https://doi.org/10.1007/s11284-012-1022-9).
- Gobin A, Campling P, Janssen L, Desmet N, Van Delden H, Hurkens J, Lavelle P, Berman S. 2011.** Soil organic matter management across the EU—best practices, constraints and trade-offs, final report for the European Commission’s DG environment. Available at [https://ec.europa.eu/environment/soil/som\\_en.htm](https://ec.europa.eu/environment/soil/som_en.htm).
- Godbold DL, Hoosbeek MR, Lukac M, Cotrufo MF, Janssens IA, Ceulemans R, Polle A, Velthorst EJ, Scarascia-Mugnozza G, De Angelis P, Miglietta F, Peressotti A. 2006.** Mycorrhizal hyphal turnover as a dominant process for carbon input into soil organic matter. *Plant and Soil* **281**(1–2):15–24 DOI [10.1007/s11104-005-3701-6](https://doi.org/10.1007/s11104-005-3701-6).
- Goddéris Y, Schott J, Brantley SL. 2019.** Reactive transport models of weathering. *Elements* **15**(2):103–106 DOI [10.2138/gselements.15.2.103](https://doi.org/10.2138/gselements.15.2.103).
- Gomiero T. 2016.** Soil degradation, land scarcity and food security: reviewing a complex challenge. *Sustainability* **8**(3):281 DOI [10.3390/su8030281](https://doi.org/10.3390/su8030281).
- Graham EB, Wieder WR, Leff JW, Weintraub SR, Townsend AR, Cleveland CC, Philippot L, Nemergut DR. 2014.** Do we need to understand microbial communities to predict ecosystem

- function? A comparison of statistical models of nitrogen cycling processes. *Soil Biology and Biochemistry* **68**:279–282 DOI [10.1016/j.soilbio.2013.08.023](https://doi.org/10.1016/j.soilbio.2013.08.023).
- Grandy AS, Wieder WR, Wickings K, Kyker-Snowman E. 2016.** Beyond microbes: are fauna the next frontier in soil biogeochemical models? *Soil Biology and Biochemistry* **102**:40–44 DOI [10.1016/j.soilbio.2016.08.008](https://doi.org/10.1016/j.soilbio.2016.08.008).
- Grayston SJ, Vaughan D, Jones D. 1997.** Rhizosphere carbon flow in trees, in comparison with annual plants: the importance of root exudation and its impact on microbial activity and nutrient availability. *Applied Soil Ecology* **5**(1):29–56 DOI [10.1016/S0929-1393\(96\)00126-6](https://doi.org/10.1016/S0929-1393(96)00126-6).
- Gregory AS, Ritz K, McGrath SP, Quinton JN, Goulding KWT, Jones RJA, Harris JA, Bol R, Wallace P, Pilgrim ES, Whitmore AP. 2015.** A review of the impacts of degradation threats on soil properties in the UK. *Soil Use and Management* **31**(Suppl. 1):1–15 DOI [10.1111/sum.12212](https://doi.org/10.1111/sum.12212).
- Griffiths B, Ritz K, Ebbelwhite N, Dobson G. 1998.** Soil microbial community structure: effects of substrate loading rates. *Soil Biology and Biochemistry* **31**(1):145–153 DOI [10.1016/S0038-0717\(98\)00117-5](https://doi.org/10.1016/S0038-0717(98)00117-5).
- Guggenberger G, Kaiser K. 2003.** Dissolved organic matter in soil: challenging the paradigm of sorptive preservation. *Geoderma* **113**(3–4):293–310 DOI [10.1016/S0016-7061\(02\)00366-X](https://doi.org/10.1016/S0016-7061(02)00366-X).
- Gunina A, Kuzyakov Y. 2014.** Pathways of litter C by formation of aggregates and SOM density fractions: implications from <sup>13</sup>C natural abundance. *Soil Biology and Biochemistry* **71**:95–104 DOI [10.1016/j.soilbio.2014.01.011](https://doi.org/10.1016/j.soilbio.2014.01.011).
- Gunnarsson T, Sundin P, Tunlid A. 1988.** Importance of leaf litter fragmentation for bacterial growth. *Oikos* **52**(3):303–308 DOI [10.2307/3565203](https://doi.org/10.2307/3565203).
- Gupta VVSR, Germida JJ. 2015.** Soil aggregation: influence on microbial biomass and implications for biological processes. *Soil Biology and Biochemistry* **80**:A3–A9 DOI [10.1016/j.soilbio.2014.09.002](https://doi.org/10.1016/j.soilbio.2014.09.002).
- Guswa AJ, Celia MA, Rodriguez-Iturbe I. 2002.** Models of soil moisture dynamics in ecohydrology: a comparative study. *Water Resources Research* **38**(9):5-1–5-15 DOI [10.1029/2001WR000826](https://doi.org/10.1029/2001WR000826).
- Hale WG. 1966.** A population study of moorland Collembola. *Pedobiologia* **6**:65–99.
- Hale CM, Frelich LE, Reich PB, Pastor J. 2005.** Effects of European earthworm invasion on soil characteristics in northern hardwood forests of Minnesota, USA. *Ecosystems* **8**:911–927 DOI [10.1007/s10021-005-0066-x](https://doi.org/10.1007/s10021-005-0066-x).
- Heal OW, Jones HE, Whittaker JB. 1975.** Structure and Function of tundra ecosystems. In: Rosswall T, Heal OW, eds. *Ecological Bulletins* **20**. Rosswall, Stockholm: Liber Tryck, 295–320.
- Helliwell JR, Miller AJ, Whalley WR, Mooney SJ, Sturrock CJ. 2014.** Quantifying the impact of microbes on soil structural development and behaviour in wet soils. *Soil Biology and Biochemistry* **74**:138–147 DOI [10.1016/j.soilbio.2014.03.009](https://doi.org/10.1016/j.soilbio.2014.03.009).
- Hendricks JJ, Mitchell RJ, Kuehn KA, Pecot SD. 2016.** Ectomycorrhizal fungal mycelia turnover in a longleaf pine forest. *New Phytologist* **209**(4):1693–1704 DOI [10.1111/nph.13729](https://doi.org/10.1111/nph.13729).
- Hendrickx JMH, Flury M. 2001.** *Uniform and preferential flow, mechanisms in the vadose zone, conceptual models of flow and transport in the fractured vadose zone*. Washington, D.C.: National Research Council, National Academy Press, 149–187.
- Hinsinger P, Betencourt E, Bernard L, Brauman A, Plassard C, Shen J, Tang X, Zhang F. 2011.** P for two, sharing a scarce resource: soil phosphorus acquisition in the rhizosphere of intercropped species. *Plant Physiology* **156**(3):1078–1086 DOI [10.1104/pp.111.175331](https://doi.org/10.1104/pp.111.175331).

- Hirsch PR, Jhurrea D, Williams JK, Philip J, Murray PJ, Scott T, Misselbrook TH, Goulding KWT, Clark IM. 2017. Soil resilience and recovery: rapid community responses to management changes. *Plant and Soil* 412:283–297.
- Ho A, Di Lonardo P, Bodelier P. 2017. Revisiting life strategy concepts in environmental microbial ecology. *FEMS Microbiology Ecology* 93(3) DOI 10.1093/femsec/fix006.
- Hobbie JE, Hobbie EA. 2006. N-15 in symbiotic fungi and plants estimates nitrogen and carbon flux rates in Arctic tundra. *Ecology* 87(4):816–822 DOI 10.1890/0012-9658(2006)87[816:NISFAP]2.0.CO;2.
- Hobbie SE, Oleksyn J, Eissenstat DM, Reich PB. 2010. Fine root decomposition rates do not mirror those of leaf litter among temperate tree species. *Oecologia* 162(2):505–513 DOI 10.1007/s00442-009-1479-6.
- Hochdanz E. 1876. Available at <https://commons.wikimedia.org/wiki/File:Melolontha.melolontha.female.-.calwer.19.04.jpg>.
- Hopkin S. 1997. *Biology of the springtails*. Oxford: Oxford University Press, 330.
- Hopkins DW, Wheatley RE, Robinson D. 1998. Stable isotope studies of soil nitrogen. In: Griffiths H, ed. *Stable Isotopes: Integration of Biological, Ecological and Geochemical Processes*. Oxford: BIOS Scientific Publishers, 75–88.
- Hoy CW, Grewal PS, Lawrence JL, Jagdale G, Acosta N. 2008. Canonical correspondence analysis demonstrates unique soil conditions for entomopathogenic nematode species compared with other free-living nematode species. *Biological Control* 46(3):371–379 DOI 10.1016/j.biocontrol.2008.06.001.
- Huang C-Y, Hendrix PF, Fahey TJ, Bohlen PJ, Groffman PM. 2010. A simulation model to evaluate the impacts of invasive earthworms on soil carbon dynamics. *Ecological Modelling* 221(20):2447–2457 DOI 10.1016/j.ecolmodel.2010.06.023.
- Hunt HW. 1977. A simulation model for decomposition in grasslands. *Ecology* 58(3):469–484 DOI 10.2307/1938998.
- Hunt HW, Coleman DC, Ingham ER, Ingham RE, Elliott ET, Moore JC, Reid CPP, Rose SL, Morley CR. 1987. The detrital food-web in a shortgrass prairie. *Biology and Fertility of Soils* 3:57–68.
- Ineson P, Leonard MA, Anderson JM. 1982. Effect of collembolan grazing upon nitrogen and cation leaching from decomposing leaf litter. *Soil Biology and Biochemistry* 14(6):601–605 DOI 10.1016/0038-0717(82)90094-3.
- Jarvis N, Koestel J, Larsbo M. 2016. Understanding preferential flow in the vadose zone: recent advances and future prospects. *Vadose Zone Journal* 15(12):75 DOI 10.2136/vzj2016.09.0075.
- Javaux M, Couvreur V, Vanderborght J, Vereecken H. 2013. Root water uptake: from three-dimensional biophysical processes to macroscopic modeling approaches. *Vadose Zone Journal* 12(4):1–16 DOI 10.2136/vzj2013.02.0042.
- Javaux M, Schröder T, Vanderborght J, Vereecken H. 2008. Use of a three-dimensional detailed modeling approach for predicting root water uptake all rights reserved: no part of this periodical may be reproduced or transmitted in any form or by any means, electronic or mechanical, including photocopying, recording, or any information storage and retrieval system, without permission in writing from the publisher. *Vadose Zone Journal* 7(3):1079–1088 DOI 10.2136/vzj2007.0115.
- Jenkinson DS, Rayner JH. 1977. The turnover of soil organic matter in some of the Rothamsted classical experiments. *Soil Science* 123(5):298–305 DOI 10.1097/00010694-197705000-00005.

- Jiménez JJ, Decaëns T, Rossi J-P. 2012.** Soil environmental heterogeneity allows spatial co-occurrence of competitor earthworm species in a gallery forest of the Colombian Llanos. *Oikos* **121**(6):915–926 DOI [10.1111/j.1600-0706.2012.20428.x](https://doi.org/10.1111/j.1600-0706.2012.20428.x).
- Jiménez JJ, Lal R. 2006.** Mechanisms of C sequestration in soils of Latin America. *Critical Reviews in Plant Sciences* **25**(4):337–365 DOI [10.1080/0735268060094240](https://doi.org/10.1080/0735268060094240).
- Joergensen RG, Wichern F. 2008.** Quantitative assessment of the fungal contribution to microbial tissue in soil. *Soil Biology and Biochemistry* **40**(12):2977–2991 DOI [10.1016/j.soilbio.2008.08.017](https://doi.org/10.1016/j.soilbio.2008.08.017).
- Johnson D, Martin F, Cairney JWG, Anderson IC. 2012.** The importance of individuals: intraspecific diversity of mycorrhizal plants and fungi in ecosystems. *New Phytologist* **194**(3):614–628 DOI [10.1111/j.1469-8137.2012.04087.x](https://doi.org/10.1111/j.1469-8137.2012.04087.x).
- Johnson SN, Rasmann N. 2015.** Root-feeding insects and their interactions with organisms in the Rhizosphere. *Annual Review of Entomology* **60**(1):517–535 DOI [10.1146/annurev-ento-010814-020608](https://doi.org/10.1146/annurev-ento-010814-020608).
- Johnston SR, Boddy L, Weightman AJ. 2016.** Bacteria in decomposing wood and their interactions with wood-decay fungi. *FEMS Microbiology Ecology* **92**(11):fw179 DOI [10.1093/femsec/fw179](https://doi.org/10.1093/femsec/fw179).
- Jones DL. 1998.** Organic acids in the rhizosphere—a critical review. *Plant and Soil* **205**(1):25–44 DOI [10.1023/A:1004356007312](https://doi.org/10.1023/A:1004356007312).
- Jones CG, Lawton JH, Shachack M. 1994.** Organisms as ecosystem engineers. *Oikos* **69**(3):373–386 DOI [10.2307/3545850](https://doi.org/10.2307/3545850).
- Jouquet P, Mamou L, Lepage M, Velde B. 2002.** Effects of termites on clay mineral soils: fungus-growing termites as weathering agents. *European Journal of Soil Science* **53**(4):521–527 DOI [10.1046/j.1365-2389.2002.00492.x](https://doi.org/10.1046/j.1365-2389.2002.00492.x).
- Jouquet P, Mathieu J, Barot S, Chossai C. 2007.** Soil engineers as ecosystem heterogeneity drivers. In: Munoz I, ed. *Ecology Research Progress*. Hauppauge: Nova Science Publishers.
- Jouquet P, Podwojewski P, Bottinelli N, Mathieu J, Ricoy M, Orange D, Tran TD, Valentin C. 2008.** Above-ground earthworm casts affect water runoff and soil erosion in Northern Vietnam. *Catena* **74**(1):13–21 DOI [10.1016/j.catena.2007.12.006](https://doi.org/10.1016/j.catena.2007.12.006).
- Jouquet P, Traoré S, Choosai C, Hartmann C, Bignell D. 2011.** Influence of termites on ecosystem functioning: ecosystem services provided by termites. *European Journal of Soil Biology* **47**(4):215–222 DOI [10.1016/j.ejsobi.2011.05.005](https://doi.org/10.1016/j.ejsobi.2011.05.005).
- Ju RH, Shen ZR. 2005.** Review on insect population dynamics simulation models. *Acta Ecologica Sinica* **25**:2709–2716.
- Juan-Ovejero R, Granjel RR, Ramil-Regoc P, Briones MJI. 2020.** The interplay between abiotic factors and below-ground biological interactions regulates carbon exports from peatlands. *Geoderma* **368**:114313 DOI [10.1016/j.geoderma.2020.114313](https://doi.org/10.1016/j.geoderma.2020.114313).
- Julich D, Julich S, Feger KH. 2017.** Phosphorus fractions in preferential flow pathways and soil matrix in hillslope soils in the Thuringian forest (Central Germany). *Journal of Plant Nutrition and Soil Science* **180**(3):407–417 DOI [10.1002/jpln.201600305](https://doi.org/10.1002/jpln.201600305).
- Jury WA, Horton R. 2004.** *Soil physics*. New York: John Wiley & Sons, 370.
- Jury WA, Or D, Pachepsky Y, Vereecken H, Hopmans JW, Ahuja LR, Clothier BE, Bristow KL, Kluitenberg GJ, Moldrup P, Simunek J, Van Genuchten MT, Horton R, Kalbitz KK. 2011.** Kirkham’s legacy and contemporary challenges in soil physics research, cycling downwards—dissolved organic matter in soils. *Soil Science society of America Journal* **75**(5):1589–1601 DOI [10.2136/sssaj2011.0115](https://doi.org/10.2136/sssaj2011.0115).

- Kaiser K, Kalbitz K. 2012. Cycling downwards—dissolved organic matter in soils. *Soil Biology and Biochemistry* 52:29–32.
- Kalbitz K, Solinger S, Park J-H, Michalzik B, Matzner E. 2000. Controls on the dynamics of dissolved organic matter in soils: a review. *Soil Science* 165(4):277–304  
DOI 10.1097/00010694-200004000-00001.
- Kaneda S, Frouz J, Baldrian P, Cajthaml T, Křišťůfek V. 2013. Does the addition of leaf litter affect soil respiration in the same way as addition of macrofauna excrements (of *Bibio marci* Diptera larvae) produced from the same litter? *Applied Soil Ecology* 72:7–13  
DOI 10.1016/j.apsoil.2013.05.011.
- Keiluweit M, Gee K, Denney A, Fendorf S. 2018. Anoxic microsites in upland soils dominantly controlled by clay content. *Soil Biology and Biochemistry* 118:42–50  
DOI 10.1016/j.soilbio.2017.12.002.
- Keiluweit M, Wanzek T, Kleber M, Nico P, Fendorf S. 2017. Anaerobic microsites have an unaccounted role in soil carbon stabilization. *Nature Communications* 8(1):1771  
DOI 10.1038/s41467-017-01406-6.
- Kladivko JE. 2001. Tillage systems and soil ecology. *Soil and Tillage Research* 61(1–2):61–76  
DOI 10.1016/S0167-1987(01)00179-9.
- Klapper. 2012. Variety of microbes as a graphic illustration. Available at <https://pixy.org/4831074/>.
- Kleber M, Johnson MG. 2010. Advances in understanding the molecular structure of soil organic matter. *Advances in Agronomy* 106:77–142 DOI 10.1016/S0065-2113(10)06003-7.
- Kleber M, Nico PS, Plante A, Filley T, Kramer M, Swanston C, Sollins P. 2011. Old and stable soil organic matter is not necessarily chemically recalcitrant: implications for modeling concepts and temperature sensitivity. *Global Change Biology* 17(2):1097–1107  
DOI 10.1111/j.1365-2486.2010.02278.x.
- Kleber M, Eusterhues K, Keiluweit M, Mikutta C, Mikutta R, Nico P. 2015. Mineral—organic associations: formation, properties, and relevance in soil environments. *Advances in Agronomy* 130:1–140 DOI 10.1016/bs.agron.2014.10.005.
- Klok C, Van der Holt A, Bodt J. 2006. Population growth and development of the earthworm *Lumbricus rubellus* in a polluted field soil: possible consequences for the godwit (*Limosa limosa*). *Environmental Toxicology and Chemistry* 25(1):213–219 DOI 10.1897/05-286R.1.
- Komarov A, Chertov O, Bykhovets S, Shaw C, Nadporozhskaya M, Frolov P, Shashkov M, Shanin V, Grabarnik P, Pripulina I, Zubkova E. 2017. Romul\_Hum model of soil organic matter formation coupled with soil biota activity: I. Problem formulation, model description, and testing. *Ecological Modelling* 345:113–124.
- Korobushkin DI, Gongalsky KB, Tiunov AV. 2014. Isotopic niche ( $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  values) of soil macrofauna in temperate forests. *Rapid Communications in Mass Spectrometry* 28(11):1303–1311 DOI 10.1002/rcm.6903.
- Kratochvil P. 2019. Spider isolated. Available at <https://www.publicdomainpictures.net/en/view-image.php?image=20174&picture=spider-isolated>.
- Kriiska K, Frey J, Asi E, Kabral N, Uri V, Aosaar J, Varik M, Napa Ü, Apuhtin V, Timmusk T, Ostonen I. 2019. Variation in annual carbon fluxes affecting the SOC pool in hemiboreal coniferous forests in Estonia. *Forest Ecology and Management* 433:419–430  
DOI 10.1016/j.foreco.2018.11.026.
- Krishna M, Mohan M. 2017. Litter decomposition in forest ecosystems: a review. *Energy, Ecology and Environment* 2(4):236–249 DOI 10.1007/s40974-017-0064-9.
- Kuka K, Franko U, Rühlmann J. 2007. Modelling the impact of pore space distribution on carbon turnover. *Ecological Modelling* 208(2–4):295–306 DOI 10.1016/j.ecolmodel.2007.06.002.

- Kuraz M, Mayer P, Pech P. 2014.** Solving the nonlinear Richards equation model with adaptive domain decomposition. *Journal of Computational and Applied Mathematics* **270**:2–11  
DOI [10.1016/j.cam.2014.03.010](https://doi.org/10.1016/j.cam.2014.03.010).
- Kuzyakov Y, Friedel JK, Stahr K. 2000.** Review of mechanisms and quantification of priming effects. *Soil Biology and Biochemistry* **32(11–12)**:1485–1498  
DOI [10.1016/S0038-0717\(00\)00084-5](https://doi.org/10.1016/S0038-0717(00)00084-5).
- Köhne JM, Köhne S, Simůnek J. 2009.** A review of model applications for structured soils: (a) Water flow and tracer transport. *Journal of Contaminant Hydrology* **104(1–4)**:4–35  
DOI [10.1016/j.jconhyd.2008.10.002](https://doi.org/10.1016/j.jconhyd.2008.10.002).
- Landl M, Huber K, Schnepf A, Vanderborght J, Javaux M, Glyn Bengough A, Vereecken H. 2017.** A new model for root growth in soil with macropores. *Plant and Soil* **415(1–2)**:99–116  
DOI [10.1007/s11104-016-3144-2](https://doi.org/10.1007/s11104-016-3144-2).
- Lavelle P. 1988.** Earthworm activities and the soil system. *Biology and Fertility of Soils* **6(3)**:237–251  
DOI [10.1007/BF00260820](https://doi.org/10.1007/BF00260820).
- Lavelle P. 1997.** Faunal activities and soil processes: adaptive strategies that determine ecosystem function. *Advances in Ecological Research* **27**:93–132.
- Lavelle P, Barot S, Blouin M, Decaëns T, Jimenez JJ, Jouquet P. 2007.** Earthworms as key actors in self-organized soil systems. In: Cuddington K, Byers JE, Wilson WG, Hastings A, eds. *Ecosystem Engineers: Plants to Protists*. Waltham: Academic Press, 405.
- Lavelle P, Bignell D, Lepage M. 1997.** Soil function in a changing world: the role of invertebrate ecosystem engineers. *European Journal of Soil Biology* **33(4)**:159–193.
- Lavelle P, Martin A. 1992.** Small-scale and large-scale effects of endogeic earthworms on soil organic matter dynamics in soils of the humid tropics. *Soil Biology and Biochemistry* **24(12)**:1491–1498 DOI [10.1016/0038-0717\(92\)90138-N](https://doi.org/10.1016/0038-0717(92)90138-N).
- Lavelle P, Spain A. 2001.** *Soil ecology*. Dordrecht: Kluwer Academic Publishers.
- Lavelle P, Spain A, Blouin M, Brown G, Decaëns T, Grimaldi M, Jiménez JJ, McKey D, Mathieu J, Velasquez E, Zangerlé A. 2016.** Ecosystem engineers in a self-organized soil: a review of concepts and future research questions. *Soil Science* **181(3–4)**:91–109  
DOI [10.1097/SS.0000000000000155](https://doi.org/10.1097/SS.0000000000000155).
- Lavelle P, Spain A, Fonte S, Bedano JC, Blanchart E, Galindo V, Grimaldi M, Jiménez JJ, Velasquez E, Zangerlé A. 2020.** Soil aggregation, ecosystem engineers and the C cycle. *Acta Oecologica* **105**:103561.
- Le Mer J, Roger P. 2001.** Production, oxidation, emission and consumption of methane by soils: a review. *European Journal of Soil Biology* **37(1)**:25–50  
DOI [10.1016/S1164-5563\(01\)01067-6](https://doi.org/10.1016/S1164-5563(01)01067-6).
- Leake J, Johnson D, Donnelly D, Muckle G, Boddy L, Read D. 2004.** Networks of power and influence: the role of mycorrhizal mycelium in controlling plant communities and agroecosystem functioning. *Canadian Journal of Botany* **82(8)**:1016–1045  
DOI [10.1139/b04-060](https://doi.org/10.1139/b04-060).
- Lebrun P. 1971.** *Ecologie et biocénotique de quelques peuplements d'arthropodes édaphiques*. Vol. 165. Bruxelles: Institut royal des sciences naturelles de Belgique, 1–203.
- Lehmann J, Kleber M. 2015.** The contentious nature of soil organic matter. *Nature* **528(7580)**:60–68 DOI [10.1038/nature16069](https://doi.org/10.1038/nature16069).
- Lehmann A, Rillig MC. 2015.** Understanding mechanisms of soil biota involvement in soil aggregation: a way forward with saprobic fungi? *Soil Biology and Biochemistry* **88**:298–302  
DOI [10.1016/j.soilbio.2015.06.006](https://doi.org/10.1016/j.soilbio.2015.06.006).



- Lehmann A, Zheng W, Rilliga MC. 2017.** Soil biota contributions to soil aggregation. *Nature Ecology & Evolution* **1**(12):1828–1835 DOI [10.1038/s41559-017-0344-y](https://doi.org/10.1038/s41559-017-0344-y).
- Leitner D, Klepsch S, Knieß A, Schnepf A. 2010.** The algorithmic beauty of plant roots – an L-System model for dynamic root growth simulation. *Mathematical and Computer Modelling of Dynamical Systems* **16**:575–587 DOI [10.1080/13873954.2010.491360](https://doi.org/10.1080/13873954.2010.491360).
- Lennon JT, Jones SE. 2011.** Microbial seed banks: the ecological and evolutionary implications of dormancy. *Nature Reviews Microbiology* **9**(2):119–130 DOI [10.1038/nrmicro2504](https://doi.org/10.1038/nrmicro2504).
- Leonowicz A, Matuszewska A, Luterek J, Ziegenhagen D, Wojtaś-Wasilewska M, Cho N-S, Hofrichter M, Rogalski J. 1999.** Biodegradation of lignin by white rot fungi. *Fungal Genetics and Biology* **27**(2–3):175–185 DOI [10.1006/fgbi.1999.1150](https://doi.org/10.1006/fgbi.1999.1150).
- Leppälampi-Kujansuu J, Salemaa M, Kleja DB, Linder S, Helmisaari HS. 2014.** Fine root turnover and litter production of Norway spruce in a long-term temperature and nutrient manipulation experiment. *Plant and Soil* **374**(1–2):73–88 DOI [10.1007/s11104-013-1853-3](https://doi.org/10.1007/s11104-013-1853-3).
- Lesturgez G, Poss R, Hartmann C. 2004.** Roots of *Stylosanthes hamata* create macropores in the compact layer of a sandy soil. *Plant and Soil* **260**(1/2):101–109 DOI [10.1023/B:PLSO.0000030184.24866.aa](https://doi.org/10.1023/B:PLSO.0000030184.24866.aa).
- Levy-Booth DJ, Prescott CE, Grayston SJ. 2014.** Microbial functional genes involved in nitrogen fixation, nitrification and denitrification in forest ecosystems. *Soil Biology and Biochemistry* **75**:11–25 DOI [10.1016/j.soilbio.2014.03.021](https://doi.org/10.1016/j.soilbio.2014.03.021).
- Liao K, Xu S, Zhu Q. 2015.** Development of ensemble pedotransfer functions for cation exchange capacity of soils of Qingdao in China. *Soil Use and Management* **31**(4):483–490 DOI [10.1111/sum.12207](https://doi.org/10.1111/sum.12207).
- Lindahl BD, Ihrmark K, Boberg J, Trumbore SE, Höglberg P, Stenlid J, Finlay RD. 2007.** Spatial separation of litter decomposition and mycorrhizal nitrogen uptake in a boreal forest. *New Phytologist* **173**(3):611–620 DOI [10.1111/j.1469-8137.2006.01936.x](https://doi.org/10.1111/j.1469-8137.2006.01936.x).
- Liski J, Palosuo T, Peltoniemi M, Sievänen R. 2005.** Carbon and decomposition model Yasso for forest soils. *Ecological Modelling* **189**(1–2):168–182 DOI [10.1016/j.ecolmodel.2005.03.005](https://doi.org/10.1016/j.ecolmodel.2005.03.005).
- Louis BP, Maron P-A, Viaud V, Leterme P, Menasseri-Aubry S. 2016.** Soil C and N models that integrate microbial diversity. *Environmental Chemistry Letters* **14**(3):331–344 DOI [10.1007/s10311-016-0571-5](https://doi.org/10.1007/s10311-016-0571-5).
- Lovegrove BG. 1989.** Spacing and origin(s) of mima-like earth mounds in the Cape Province of South Africa. *South African Journal of Science* **85**:108–112.
- Lundström US, Van Breemen N, Bain DC, Van Hees PAW, Giesler R, Gustafsson JP, Ilvesniemi H, Karlton E, Melkerud P-A, Olsson M, Riise G, Wahlberg O, Bergelin A, Bishop K, Finlay R, Jongmans AG, Magnusson T, Mannerkoski H, Nordgren A, Nyberg L, Starr M, Tau Strand L. 2000.** Advances in understanding the podzolization process resulting from a multidisciplinary study of three coniferous forest soils in the Nordic Countries. *Geoderma* **94**(2–4):335–353 DOI [10.1016/S0016-7061\(99\)00077-4](https://doi.org/10.1016/S0016-7061(99)00077-4).
- Luxton M. 1972.** Studies on the oribatid mites of a Danish beech wood soil: I. Nutritional biology. *Pedobiologia* **12**:434–463.
- Lynd LR, Weimer PJ, Van Zyl WH, Pretorius IS. 2002.** Microbial cellulose utilization: fundamentals and biotechnology. *Microbiology and Molecular Biology Reviews* **66**(3):506–577 DOI [10.1128/MMBR.66.3.506-577.2002](https://doi.org/10.1128/MMBR.66.3.506-577.2002).
- Maaß S, Caruso T, Rillig MC. 2015.** Functional role of microarthropods in soil aggregation. *Pedobiologia* **58**(2–3):59–63 DOI [10.1016/j.pedobi.2015.03.001](https://doi.org/10.1016/j.pedobi.2015.03.001).

- MacLean SF Jr. 1980.** The detritus-based trophic system. In: Brown J, Miller PC, Tieszen LL, Bunnell FL, eds. *An Arctic Ecosystem: The Coastal Tundra at Barrow, Alaska*. Stroudsburg: Dowden, Hutchinson & Ross, Inc, 411–457.
- Mai TH, Schnepf A, Vereecken H, Vanderborght J. 2019.** Continuum multiscale model of root water and nutrient uptake from soil with explicit consideration of the 3D root architecture and the rhizosphere gradients. *Plant and Soil* **439**(1–2):273–292 DOI [10.1007/s11104-018-3890-4](https://doi.org/10.1007/s11104-018-3890-4).
- Malamoud K, McBratney AB, Minasny B, Field DJ. 2009.** Modelling how carbon affects soil structure. *Geoderma* **149**(1–2):19–26 DOI [10.1016/j.geoderma.2008.10.018](https://doi.org/10.1016/j.geoderma.2008.10.018).
- Malard JJ, Adamowski JF, Díaz MR, Nassar JB, Anandaraja N, Tuy H, Arévalo-Rodríguez LA, Melgar-Quiñonez HR. 2020.** Agroecological food web modelling to evaluate and design organic and conventional agricultural systems. *Ecological Modelling* **421**:108961 DOI [10.1016/j.ecolmodel.2020.108961](https://doi.org/10.1016/j.ecolmodel.2020.108961).
- Mambelli S, Bird JA, Gleixner G, Dawson TE, Torn MS. 2011.** Relative contribution of foliar and fine root pine litter to the molecular composition of soil organic matter after in situ degradation. *Organic Geochemistry* **42**:1099–1108.
- Mankel A, Krause K, Kothe E. 2002.** Identification of a hydrophobin gene that is developmentally regulated in the ectomycorrhizal fungus *Trichoderma terreum*. *Applied and Environmental Microbiology* **68**(3):1408–1413 DOI [10.1128/AEM.68.3.1408-1413.2002](https://doi.org/10.1128/AEM.68.3.1408-1413.2002).
- Mansfield SD, Meder R. 2003.** Cellulose hydrolysis—the role of monocomponent cellulases in crystalline cellulose degradation. *Cellulose* **10**(2):159–169 DOI [10.1023/A:1024022710366](https://doi.org/10.1023/A:1024022710366).
- Manzoni S, Porporato A. 2009.** Soil carbon and nitrogen mineralization: theory and models across scales. *Soil Biology and Biochemistry* **41**(7):1355–1379 DOI [10.1016/j.soilbio.2009.02.031](https://doi.org/10.1016/j.soilbio.2009.02.031).
- Marashi ARA, Scullion J. 2003.** Earthworm casts form stable aggregates in physically degraded soils. *Biology and Fertility of Soils* **37**(6):375–380 DOI [10.1007/s00374-003-0617-2](https://doi.org/10.1007/s00374-003-0617-2).
- Marschner B, Brodowski S, Dreves A, Gleixner G, Gude A, Grootes PM, Hamer U, Heim A, Jandl G, Ji R, Kaiser K, Kalbitz K, Kramer C, Leinweber P, Rethemeyer J, Schäffer A, Schmidt MWI, Schwark L, Wiesenberger GLB. 2008.** How relevant is recalcitrance for the stabilization of organic matter in soils? *Journal of Plant Nutrition and Soil Science* **171**(1):91–110 DOI [10.1002/jpln.200700049](https://doi.org/10.1002/jpln.200700049).
- Martin S, Lavelle P. 1992.** A simulation model of vertical movements of an earthworm population (*Millsonia anomala* Omodeo, Megascolecidae) in an African savanna (Lamto, Ivory Coast). *Soil Biology and Biochemistry* **24**(12):1419–1424 DOI [10.1016/0038-0717\(92\)90127-J](https://doi.org/10.1016/0038-0717(92)90127-J).
- McBratney AB, Minasny B, Cattle SR, Vervoort RW. 2001.** From pedotransfer functions to soil inference systems. *Geoderma* **109**(1–2):41–73 DOI [10.1016/S0016-7061\(02\)00139-8](https://doi.org/10.1016/S0016-7061(02)00139-8).
- McBrayer JF. 1977.** Contributions of cryptozoa to forest nutrient cycles. In: Mattson WJ, ed. *The Role of Arthropods in Forest Ecosystems: Proceedings in Life Sciences*. Berlin: Springer.
- McCormack ML, Crisfield E, Raczka B, Schnekenburger F, Eissenstat DM, Smithwick EAH. 2015.** Sensitivity of four ecological models to adjustments in fine root turnover rate. *Ecological Modelling* **297**:107–117 DOI [10.1016/j.ecolmodel.2014.11.013](https://doi.org/10.1016/j.ecolmodel.2014.11.013).
- McGuire KL, Treseder K. 2010.** Microbial communities and their relevance for ecosystem models: decomposition as a case study. *Soil Biology and Biochemistry* **42**(4):529–535 DOI [10.1016/j.soilbio.2009.11.016](https://doi.org/10.1016/j.soilbio.2009.11.016).
- McInerney M, Little DJ, Bolger T. 2001.** Effect of earthworm cast formation on the stabilization of organic matter in fine soil fractions. *European Journal of Soil Biology* **37**(4):251–254 DOI [10.1016/S1164-5563\(01\)01092-5](https://doi.org/10.1016/S1164-5563(01)01092-5).
- Mc Millan and Healey. 1971.** A quantitative technique for the analysis of the gut contents of Collembola. *Revue d'Ecologie et de Biologie du Sol* **8**:295–300.

- Melillo JM, Aber JD, Muratore JF. 1982. Nitrogen and lignin control of hardwood leaf litter decomposition dynamics. *Ecology* 63:621–626.
- Meyer A, Grote R, Butterbach-Bahl K. 2012. Integrating mycorrhiza in a complex model system: effects on ecosystem C and N fluxes. *European Journal of Forest Research* 131(6):1809–1831 DOI 10.1007/s10342-012-0634-5.
- Meyer A, Grote R, Polle A, Butterbach-Bahl K. 2010. Simulating mycorrhiza contribution to forest C- and N- cycling – the MYCOFON model. *Plant and Soil* 327(1–2):493–517 DOI 10.1007/s11104-009-0017-y.
- Mikola J. 1998. Effects of microbivore species composition and basal resource enrichment on trophic-level biomasses in an experimental microbial-based soil food web. *Oecologia* 117(3):396–403 DOI 10.1007/s004420050673.
- Mills AL. 2003. Keeping in touch: microbial life on soil particle surfaces. *Advances in Agronomy* 78:1–43.
- Minasny B, Sulaeman Y, McBratney AB. 2011. Is soil carbon disappearing? *The Dynamics of Soil Organic Carbon in Java Global Change Biology* 17:1917–1924 DOI 10.1111/j.1365-2486.2010.02324.x.
- Mitchell MJ. 1977. Population dynamics of oribatid mites (Acari, Cryptostigmata) in an aspen woodland soil. *Pedobiologia* 17:305–319.
- Moore JAM, Jiang J, Post WM, Classen AT. 2015. Decomposition by ectomycorrhizal fungi alters soil carbon storage in a simulation model. *Ecosphere* 6(3):29 DOI 10.1890/ES14-00301.1.
- Nannipieri P, Ascher J, Ceccherini MT, Landi L, Pietramellara G, Renella G. 2003. Microbial diversity and soil functions. *European Journal of Soil Science* 54(4):655–670 DOI 10.1046/j.1351-0754.2003.0556.x.
- Negassa WC, Guber AK, Kravchenko AN, Marsh TL, Hildebrandt B, Rivers ML. 2015. Properties of soil pore space regulate pathways of plant residue decomposition and community structure of associated bacteria. *PLOS ONE* 10(4):e0123999 DOI 10.1371/journal.pone.0123999.
- Neill C, Gignoux J. 2006. Soil organic matter decomposition driven by microbial growth: a simple model for a complex network of interactions. *Soil Biology and Biochemistry* 38(4):803–811 DOI 10.1016/j.soilbio.2005.07.007.
- Nemani R, Keeling C, Hashimoto H, Jolly W, Piper S, Tucker C, Myneni R, Running S. 2003. Climate-driven increases in global terrestrial net primary production from 1982 to 1999. *Science* 300(5625):1560–1563 DOI 10.1126/science.1082750.
- Nielsen U, Ayres E, Wall D, Bardgett R. 2011. Soil biodiversity and carbon cycling: a review and synthesis of studies examining diversity-function relationships. *European Journal of Soil Science* 62(1):105–116 DOI 10.1111/j.1365-2389.2010.01314.x.
- Näsholm T, Högborg P, Franklin O, Metcalfe D, Keel SG, Campbell C, Hurry V, Linder S, Högborg MN. 2013. Are ectomycorrhizal fungi alleviating or aggravating nitrogen limitation of tree growth in boreal forests? *New Phytologist* 198:214–221.
- O'Connor FB. 1967. The enchytraeidae. In: Burges A, Raw F, eds. *Soil Biology*. London: Academic Press, 213–257.
- Oades JM, Waters AG. 1991. Aggregate hierarchy in soils. *Soil Research* 29(6):815–828 DOI 10.1071/SR9910815.
- Oldeman LR, Hakkeling RTA, Sombroek WG. 1991. *World map of the status of human-induced soil degradation: an explanatory note*. Second Edition. Nairobi: International Soil Reference and Information Center, Wageningen and United Nations Environment Programme.

- Or D, Smets BF, Wraith JM, Dechesne A, Friedman SP. 2007. Physical constraints affecting bacterial habitats and activity in unsaturated porous media—a review. *Advances in Water Resources* 30(6–7):1505–1527 DOI 10.1016/j.advwatres.2006.05.025.
- Orazova MK, Semenova TA, Tiunov AV. 2003. The microfungus community of Lumbricus terrestris middens in a linden (Tilia cordata) forest. *Pedobiologia* 47(1):27–32 DOI 10.1078/0031-4056-00166.
- Orgiazzi A, Bardgett RD, Barrios E, Behan-Pelletier V, Briones MJI, Chotte JL, De Deyn GB, Eggleton P, Fierer N, Fraser T, Hedlund K, Jeffrey S, Johnson NC, Jones A, Kandeler E, Kaneko N, Lavelle P, Lemanceau P, Miko L, Montanarella L, De Souza Moreira FM, Ramirez KS, Scheu S, Singh BK, Six J, Van der Putten WH, Wall DH. 2016a. *Global Soil Biodiversity Atlas*. Luxembourg: European Commission.
- Orgiazzi A, Panagos P, Yigini Y, Dunbar MB, Gardi C, Montanarella L, Ballabio C. 2016b. A knowledge-based approach to estimating the magnitude and spatial patterns of potential threats to soil biodiversity. *Science of the Total Environment* 545–546:11–20 DOI 10.1016/j.scitotenv.2015.12.092.
- Orwin KH, Kirschbaum MUF, St John MG, Dickie IA. 2011. Organic nutrient uptake by mycorrhizal fungi enhances ecosystem carbon storage: a model-based assessment. *Ecology Letters* 14(5):493–502 DOI 10.1111/j.1461-0248.2011.01611.x.
- Osler GH, Sommerkorn M. 2007. Toward a complete soil C and N cycle: incorporating the soil fauna. *Ecology* 88(7):1611–1621 DOI 10.1890/06-1357.1.
- Ostonen I, Truu M, Helmisaari H-S, Lukac M, Borken W, Vanguelova E, Godbold D, Lõhmus K, Zang U, Tedersoo L, Preem J-K, Rosenvald K, Aosaar J, Armolaitis K, Frey J, Kabral N, Kukumägi M, Leppälammikujansuu J, Lindroos A-J, Merilä P, Napa Ü, Nöjd P, Parts K, Uri V, Varik M, Truu M. 2017. Adaptive root foraging strategies along a boreal-temperate forest gradient. *New Phytologist* 215(3):977–991 DOI 10.1111/nph.14643.
- O’Neill RV. 1969. Indirect estimation of energy fluxes in animal food webs. *Journal of Theoretical Biology* 22(2):284–290 DOI 10.1016/0022-5193(69)90006-X.
- Parker WE, Howard JJ. 2001. The biology and management of wireworms (*Agriotes* spp.) on potato with particular reference to the UK. *Agricultural and Forest Entomology* 3(2):85–98 DOI 10.1046/j.1461-9563.2001.00094.x.
- Parkhurst DL, Appelo CAJ. 2013. Description of input and examples for PHREEQC version 3. A computer program for speciation, batch-reaction, one-dimensional transport, and inverse geochemical calculations. U.S. Geological Survey Techniques and Methods, book 6, chap. A43. Available at <https://pubs.usgs.gov/tm/06/a43/>.
- Parton WJ, Schimel DS, Cole CV, Ojima DS. 1987. Analysis of factors controlling soil organic matter levels in great plains grasslands. *Soil Science Society of America Journal* 51(5):1173–1179 DOI 10.2136/sssaj1987.03615995005100050015x.
- Paul EA. 2016. The nature and dynamics of soil organic matter: plant inputs, microbial transformations, and organic matter stabilization. *Soil Biology and Biochemistry* 98:109–126 DOI 10.1016/j.soilbio.2016.04.001.
- Pausch J, Kramer S, Scharroba A, Scheunemann N, Butenschön O, Kandeler E, Ruess L. 2016. Small but active—pool size does not matter for carbon incorporation in below-ground food webs. *Functional Ecology* 30(3):479–789 DOI 10.1111/1365-2435.12512.
- Pausch J, Kuzyakov Y. 2018. Carbon input by roots into the soil: quantification of rhizodeposition from root to ecosystem scale. *Global Change Biology* 24(1):1–12 DOI 10.1111/gcb.13850.

- Paustian K, Parton WJ, Persson J. 1992.** Modeling Soil Organic Matter in Organic-Amended and Nitrogen-Fertilized Long-Term Plots. *Soil Science of America Journal* **56**:476–488 DOI [10.2136/sssaj1992.03615995005600020023x](https://doi.org/10.2136/sssaj1992.03615995005600020023x).
- Pelosi C, Bertrand M, Makowski D, Roger-Estrade J. 2008.** WORMDYN: a model of Lumbricus terrestris population dynamics in agricultural fields. *Ecological Modelling* **218**(3–4):219–234 DOI [10.1016/j.ecolmodel.2008.07.002](https://doi.org/10.1016/j.ecolmodel.2008.07.002).
- Perveen N, Barot S, Alvarez G, Klumpp K, Martin R, Rapaport A, Herfurth D, Louault F, Fontaine S. 2014.** Priming effect and microbial diversity in ecosystem functioning and response to global change: a modeling approach using the SYMPHONY model. *Global Change Biology* **20**(4):1174–1190 DOI [10.1111/gcb.12493](https://doi.org/10.1111/gcb.12493).
- Petersen H. 1982.** The total soil fauna biomass and its composition. *Oikos* **39**:330–339.
- Petersen H, Luxton M. 1982.** A comparative analysis of soil fauna populations and their role in decomposition processes. *Oikos* **39**(3):288–388 DOI [10.2307/3544689](https://doi.org/10.2307/3544689).
- Pferdmenges J, Breuer L, Julich S, Kraft P. 2020.** Review of soil phosphorus routines in ecosystem models. *Environmental Modelling and Software* **126**:104639 DOI [10.1016/j.envsoft.2020.104639](https://doi.org/10.1016/j.envsoft.2020.104639).
- Philippot L, Spor A, Hénault C, Bru D, Bizouard F, Jones CM, Sarr A, Maron P-A. 2013.** Loss in microbial diversity affects nitrogen cycling in soil. *ISME Journal* **7**(8):1609–1619 DOI [10.1038/ismej.2013.34](https://doi.org/10.1038/ismej.2013.34).
- Phillips LA, Ward V, Jones MD. 2014.** Ectomycorrhizal fungi contribute to soil organic matter cycling in sub-boreal forests. *ISME Journal* **8**(3):699–713 DOI [10.1038/ismej.2013.195](https://doi.org/10.1038/ismej.2013.195).
- Ponge JF. 1991.** Succession of fungi and fauna during decomposition of needles in a small area of Scots pine litter. *Plant and Soil* **138**(1):99–113 DOI [10.1007/BF00011812](https://doi.org/10.1007/BF00011812).
- Ponge JF. 2003.** Humus forms in terrestrial ecosystems: a framework to biodiversity. *Soil Biology and Biochemistry* **35**(7):935–945 DOI [10.1016/S0038-0717\(03\)00149-4](https://doi.org/10.1016/S0038-0717(03)00149-4).
- Postma J, Kuppe C, Owen M, Mellor N, Griffiths M, Bennett J, Lynch J, Watt M. 2017.** OpenSimRoot: widening the scope and application of root architectural models. *New Phytologist* **215**(3):1274–1286 DOI [10.1111/nph.14641](https://doi.org/10.1111/nph.14641).
- Potapov AA, Semenina EE, Korotkevich AY, Kuznetsova NA, Tiunov AV. 2016.** Connecting taxonomy and ecology: trophic niches of collembolans as related to taxonomic identity and life forms. *Soil Biology and Biochemistry* **101**:20–31 DOI [10.1016/j.soilbio.2016.07.002](https://doi.org/10.1016/j.soilbio.2016.07.002).
- Pozo MJ, Lopez-Raez JA, Azcon-Aguilar C, Garcia-Garrido JM. 2015.** Phytohormones as integrators of environmental signals in the regulation of mycorrhizal symbioses. *New Phytologist* **205**:1431–1436.
- Prescott CE. 2010.** Litter decomposition: what controls it and how can we alter it to sequester more carbon in forest soils? *Biogeochemistry* **101**(1–3):133–149 DOI [10.1007/s10533-010-9439-0](https://doi.org/10.1007/s10533-010-9439-0).
- Pulleman MM, Six J, Uyl A, Marinissen JCY, Jongmans AG. 2005.** Earthworms and management affect organic matter incorporation and microaggregate formation in agricultural soils. *Applied Soil Ecology* **29**(1):1–15 DOI [10.1016/j.apsoil.2004.10.003](https://doi.org/10.1016/j.apsoil.2004.10.003).
- Ramirez KS, Döring M, Eisenhauer N, Gardi C, Ladau J, Leff JW, Lentendu G, Lindo Z, Rillig MC, Russell D, Scheu S, St. John MG, De Vries FT, Wubet T, Van der Putten WH, Wall DH. 2015.** Toward a global platform for linking soil biodiversity data. *Frontiers in Ecology and Evolution* **3**:91.
- Ranoarisoa M, Morel P, Andriamananjara CA, Bernard L, Becquer T, Rabeharisoa L, Rahajaharilaza K, Plassard C, Blanchart E, Trap J. 2018.** Effects of a bacterivorous nematode on rice 32P uptake and root architecture in a high P sorbing ferrallitic soil. *Soil Biology and Biochemistry* **122**:39–49 DOI [10.1016/j.soilbio.2018.04.002](https://doi.org/10.1016/j.soilbio.2018.04.002).

- Rasse D, Mulder J, Moni C, Chenu C. 2006. Carbon turnover kinetics with depth in a French loamy soil. *Soil Science Society of America Journal* 70(6):2097–2105 DOI 10.2136/sssaj2006.0056.
- Rasse DP, Rumpel C, Dignac M-F. 2005. Is soil carbon mostly root carbon? Mechanisms for a specific stabilization. *Plant and Soil* 269(1–2):341–356 DOI 10.1007/s11104-004-0907-y.
- Raynaud X, Jones CG, Barot S. 2013. Ecosystem engineering, environmental decay and environmental states of landscapes. *Oikos* 122(4):591–600 DOI 10.1111/j.1600-0706.2012.20283.x.
- Read DJ, Perez-Moreno J. 2003. Mycorrhizas and nutrient cycling in ecosystems—a journey towards relevance? *New Phytologist* 157:475–492.
- Regelink IC, Weng L, Lair GJ, Comans RNJ. 2015. Adsorption of phosphate and organic matter on metal (hydr)oxides in arable and forest soil: a mechanistic modelling study. *European Journal of Soil Science* 66(5):867–875 DOI 10.1111/ejss.12285.
- Reichle DE. 1977. The role of the soil invertebrates in nutrient cycling. In: Lohm U, Persson T, eds. *Soil Organisms as Components of Ecosystems. Proceedings of the VI International Soil Zoology Colloquium of the International Society of Soil Science*. Uppsala: Swedish Natural Science Research Council, 145–156.
- Reichstein M, Rey A, Freibauer A, John Tenhunen J, Valentini R, Banza J, Casals P, Cheng Y, Grünzweig JM, Irvine J, Joffre R, Law BE, Loustau D, Miglietta F, Oechel W, Ourcival J-M, Pereira JS, Peressotti A, Ponti F, Qi Y, Rambal S, Rayment M, Romanya J, Rossi F, Tedeschi V, Tirone G, Xu M, Yakir D. 2003. Modeling temporal and large-scale spatial variability of soil respiration from soil water availability, temperature and vegetation productivity indices. *Global Biogeochemical Cycles* 1(4):35 DOI 10.1029/2003GB002035.
- Riley WJ, Maggi F, Kleber M, Torn MS, Tang JY, Dwivedi D, Guerry N. 2014. Long residence times of rapidly decomposable soil organic matter: application of multi-phase, multicomponent, and vertically resolved model (BAMS1) to soil carbon dynamics. *Geoscience Model Development* 7:1335–1355.
- Rillig MC, Mummey DL. 2006. Mycorrhizas and soil structure. *New Phytologist* 171(1):41–53 DOI 10.1111/j.1469-8137.2006.01750.x.
- Rineau F, Shah F, Smits MM, Persson P, Johansson T, Carleer R, Troein C, Tunlid A. 2013. Carbon availability triggers the decomposition of plant litter and assimilation of nitrogen by an ectomycorrhizal fungus. *ISME Journal* 7(10):2010–2022 DOI 10.1038/ismej.2013.91.
- Robertson AD, Paustian K, Ogle S, Wallenstein MD, Lugato E, Cotrufo MF. 2019. Unifying soil organic matter formation and persistence frameworks: the MEMS model. *Biogeosciences* 16(6):1225–1248 DOI 10.5194/bg-16-1225-2019.
- Romano N, Palladino M, Chirico GB. 2011. Parameterization of a bucket model for soil-vegetation-atmosphere modeling under seasonal climatic regimes. *Hydrology and Earth System Sciences* 15:3877–3893.
- Rousk J, Brookes PC, Bååth E. 2009. Contrasting soil pH effects on fungal and bacterial growth suggests functional redundancy in carbon mineralisation. *Applied and Environmental Microbiology* 75(6):1589–1596 DOI 10.1128/AEM.02775-08.
- Rumpel C, Eusterhuesa K, Kögel-Knabner I. 2004. Location and chemical composition of stabilized organic carbon in topsoil and subsoil horizons of two acid forest soils. *Soil Biology and Biochemistry* 36(1):177–190 DOI 10.1016/j.soilbio.2003.09.005.
- Rumpel C, Kögel-Knabner I. 2011. Deep soil organic matter—a key but poorly understood component of terrestrial C cycle. *Plant and Soil* 338(1–2):143–158 DOI 10.1007/s11104-010-0391-5.

- Rumpel C, Kögel-Knabner I, Bruhn F. 2002. Vertical distribution, age, and chemical composition of organic carbon in two forest soils of different pedogenesis. *Organic Geochemistry* 33(10):1131–1142 DOI 10.1016/S0146-6380(02)00088-8.
- Rühlmann J. 1999. A new approach to estimating the pool of stable organic matter in soil using data from long-term field experiments. *Plant and Soil* 213(1/2):149–160 DOI 10.1023/A:1004552016182.
- Samson RI. 2001. Effect of feeding by larvae of *Inopus rubriceps* (Diptera: Stratiomyidae) on development and growth of sugarcane. *Journal of Economic Entomology* 94(5):1097–1103 DOI 10.1603/0022-0493-94.5.1097.
- Sanders D, Jones CG, Thébault E, Bouma TJ, Van der Heide T, Van Belzen J, Barot S. 2014. Integrating ecosystem engineering and food webs. *Oikos* 123(5):513–524 DOI 10.1111/j.1600-0706.2013.01011.x.
- Satchell JE. 1967. Lumbricidae. In: Burges A, Raw F, eds. *Soil Biology*. London: Academic Press, 259–352.
- Scheunemann N, Pausch J, Digel C, Kramer S, Scharroba A, Kuzyakov Y, Kandeler E, Ruess L, Butenschoen O, Scheu S. 2016. Incorporation of root C and fertilizer N into the food web of an arable field: variations with functional group and energy channel. *Food Webs* 9:39–45 DOI 10.1016/j.fooweb.2016.02.006.
- Schmidt MWI, Torn MS, Abiven S, Dittmar T, Guggenberger G, Janssens IA, Kleber M, Kögel-Knabner I, Lehmann J, Manning DAC, Nannipieri P, Rasse DP, Weiner S, Trumbore SE. 2011. Persistence of soil organic matter as an ecosystem property. *Nature* 478(7367):49–56 DOI 10.1038/nature10386.
- Schneider AK, Hohenbrink TL, Reck A, Zangerlé A, Schröder B, Zehe E, Van Schaik L. 2018. Variability of earthworm-induced biopores and their hydrological effectiveness in space and time. *Pedobiologia* 71:8–19 DOI 10.1016/j.pedobi.2018.09.001.
- Schneider KS, Migge S, Norton RA, Scheu S, Langel R, Reinekingd A, Maraun M. 2004. Trophic niche differentiation in soil microarthropods (Oribatida, Acari): evidence from stable isotope ratios ( $^{15}\text{N}/^{14}\text{N}$ ). *Soil Biology and Biochemistry* 36(11):1769–1774 DOI 10.1016/j.soilbio.2004.04.033.
- Schnepf A, Black CK, Couvreur V, Delory BM, Doussan C, Koch A, Koch T, Javaux M, Landl M, Leitner D, Lobet G, Hieu MT, Meunier F, Petrich L, Postma J, Priesack E, Schmidt V, Vanderborght J, Vereecken H, Weber M. 2020. Call for participation: collaborative benchmarking of functional-structural root architecture models—the case of root water uptake. *Frontiers in Plant Science* 11:316 DOI 10.3389/fpls.2020.00316.
- Schnepf A, Leitner D, Klepsch S. 2012. Modeling phosphorus uptake by a growing and exuding root system A. *Vadose Zone Journal* 11(3):1 DOI 10.2136/vzj2012.0001.
- Schnepf A, Roose T. 2006. Modelling the contribution of arbuscular mycorrhizal fungi to plant phosphate uptake. *New Phytologist* 171(3):669–682.
- Schröder T, Tang L, Javaux M, Vanderborght J, Korfgén B, Vereecken H. 2009. A grid refinement approach for a 3D soil-root water transfer model. *Water Resources Research* 45(10):1597 DOI 10.1029/2009WR007873.
- Schröter D, Wolters V, De Ruiter PC. 2003. C and N mineralisation in the decomposer food webs of a European forest transect. *Oikos* 102(2):294–308 DOI 10.1034/j.1600-0579.2003.12064.x.
- Schulmann OP, Tiunov AV. 1999. Leaf litter fragmentation by the earthworm *Lumbricus terrestris* L. *Pedobiologia* 43:453–458.

- Schwartz N, Carminati A, Javaux M. 2016.** The impact of mucilage on root water uptake—a numerical study. *Water Resources Research* **52(1)**:264–277 DOI [10.1002/2015WR018150](https://doi.org/10.1002/2015WR018150).
- Segoli M, De Gryze S, Post W, Six J. 2011.** Soil aggregate dynamics: a mechanistic model. AGU Fall Meeting Abstracts. 03-. Available at [https://www.researchgate.net/publication/258459551\\_Soil\\_Aggregate\\_Dynamics\\_A\\_Mechanistic\\_Model](https://www.researchgate.net/publication/258459551_Soil_Aggregate_Dynamics_A_Mechanistic_Model).
- Seki K, Miyazaki T, Nakano M. 1998.** Effects of microorganisms on hydraulic conductivity decrease in infiltration. *European Journal of Soil Science* **49(2)**:231–236 DOI [10.1046/j.1365-2389.1998.00152.x](https://doi.org/10.1046/j.1365-2389.1998.00152.x).
- Setälä H, Martikainen E, Tynnismaa M, Huhta V. 1990.** Effects of soil fauna on leaching of N and P from experimental systems simulating coniferous forest floor. *Biology and Fertility of Soils* **10**:170–177.
- Shah F, Nicolás C, Bentzer J, Ellström M, Smits MM, Rineau F, Canbäck B, Floudas D, Carleer R, Lackner G, Braesel J, Hoffmeister DC, Henrissat B, Ahrén DG, Johansson T, Hibbett DS, Martin FM, Persson P, Tunlid A. 2016.** Ectomycorrhizal fungi decompose soil organic matter using oxidative mechanisms adapted from saprotrophic ancestors. *New Phytologist* **209(4)**:1705–1719 DOI [10.1111/nph.13722](https://doi.org/10.1111/nph.13722).
- Šimůnek J, Jarvis NJ, Van Genuchten MT, Gärdenäs A. 2003.** Review and comparison of models for describing non-equilibrium and preferential flow and transport in the vadose zone. *Journal of Hydrology* **272(1–4)**:14–35 DOI [10.1016/S0022-1694\(02\)00252-4](https://doi.org/10.1016/S0022-1694(02)00252-4).
- Šimůnek J, Jacques D, Twarakavi NKC, van Genuchten MTh. 2009.** Modeling subsurface flow and contaminant transport as influenced by biological processes at various scales using selected HYDRUS modules. *Biologia* **64(3)**:465–469.
- Sinacore K, Hall JS, Potvin C, Royo AA, Ducey MJ, Ashton MS. 2017.** Unearthing the hidden world of roots: root biomass and architecture differ among species within the same guild. *PLOS ONE* **12(10)**:e0185934 DOI [10.1371/journal.pone.0185934](https://doi.org/10.1371/journal.pone.0185934).
- Six J, Bossuyt H, Degryze S, Deneff K. 2004.** A history of research on the link between (micro) aggregates, soil biota, and soil organic matter dynamics. *Soil and Tillage Research* **79(1)**:7–31 DOI [10.1016/j.still.2004.03.008](https://doi.org/10.1016/j.still.2004.03.008).
- Six J, Paustian K. 2014.** Aggregate-associated soil organic matter as an ecosystem property and a measurement tool. *Soil Biology and Biochemistry* **68**:A4–A9 DOI [10.1016/j.soilbio.2013.06.014](https://doi.org/10.1016/j.soilbio.2013.06.014).
- Smith SE, Anderson IC, Smith FA. 2015.** Mycorrhizal associations and phosphorus acquisition: from cells to ecosystems. In: Plaxton WC, Lambers H, eds. *Annual Plant Reviews*. Vol. 48. John Wiley & Sons Ltd, 409–439.
- Smith P, Powlson DS, Glendining MJ, Smith JU. 1997.** Potential for carbon sequestration in European soils: preliminary estimates for five scenarios using results from long-term experiments. *Global Change Biology* **3(1)**:67–79 DOI [10.1046/j.1365-2486.1997.00055.x](https://doi.org/10.1046/j.1365-2486.1997.00055.x).
- Smith SE, Read DJ. 2008.** *Mycorrhizal Symbiosis*. Amsterdam: Academic Press.
- Sokol NW, Bradford MA. 2019.** Microbial formation of stable soil carbon is more efficient from belowground than aboveground input. *Nature Geoscience* **12(1)**:46–53 DOI [10.1038/s41561-018-0258-6](https://doi.org/10.1038/s41561-018-0258-6).
- Soong JL, Nielsen UN. 2016.** The role of microarthropods in emerging models of soil organic matter. *Soil Biology and Biochemistry* **102**:37–39 DOI [10.1016/j.soilbio.2016.06.020](https://doi.org/10.1016/j.soilbio.2016.06.020).
- Špaldoňová A, Frouz J. 2014.** The role of *Armadillidium vulgare* (Isopoda: Oniscidea) in litter decomposition and soil organic matter stabilization. *Applied Soil Ecology* **83**:186–192 DOI [10.1016/j.apsoil.2014.04.012](https://doi.org/10.1016/j.apsoil.2014.04.012).



- Staddon PL. 1998.** Insights into mycorrhizal colonisation at elevated CO<sub>2</sub>: a simple carbon partitioning model. *Plant and Soil* **205**(2):171–180 DOI [10.1023/A:1004388605110](https://doi.org/10.1023/A:1004388605110).
- Stamati FE, Nikolaidis NP, Banwart S, Blum WEH. 2013.** A coupled carbon, aggregation, and structure turnover (CAST) model for topsoils. *Geoderma* **211–212**:51–64 DOI [10.1016/j.geoderma.2013.06.014](https://doi.org/10.1016/j.geoderma.2013.06.014).
- Standen V. 1973.** The life cycle and annual production of *Trichoniscus pusillus pusillus* (Crustacea: Isopoda) in a Cheshire Wood. *Pedobiologia* **13**:273–291.
- Stark JM, Firestone MK. 1995.** Mechanisms for soil moisture effects on activity of nitrifying bacteria. *Applied and Environmental Microbiology* **61**(1):218–221 DOI [10.1128/AEM.61.1.218-221.1995](https://doi.org/10.1128/AEM.61.1.218-221.1995).
- Steeffel CI, Appelo CAJ, Arora B, Jacques D, Kalbacher T, Kolditz O, Lagneau V, Lichtner PC, Mayer KU, Meeussen JCL, Molins S, Moulton D, Shao H, Šimůnek J, Spycher N, Yabusaki SB, Yeh GT. 2015.** Reactive transport codes for subsurface environmental simulation. *Computational Geosciences* **19**(3):445–478 DOI [10.1007/s10596-014-9443-x](https://doi.org/10.1007/s10596-014-9443-x).
- Stevens MM, Fox KM, Warren GN, Cullis BR, Coombes NE, Lewina LG. 2000.** An image analysis technique for assessing resistance in rice cultivars to root-feeding chironomid midge larvae (Diptera: Chironomidae). *Field Crops Research* **66**(1):25–36 DOI [10.1016/S0378-4290\(99\)00074-X](https://doi.org/10.1016/S0378-4290(99)00074-X).
- Stockmann U, Adams MA, Crawford JW, Field DJ, Henakaarchchi N, Jenkins M, Minasny B, McBratney AB, De Remy deCourcellesa V, Singh K, Wheeler I, Abbott L, Angers DA, Baldock J, Bird M, Brookes PC, Chenu C, Jastrow JD, Lal R, Lehmann J, O'Donnell AG, Parton WJ, Whitehead D, Zimmermann M. 2013.** The knowns, known unknowns and unknowns of sequestration of soil organic carbon. *Agriculture, Ecosystems and Environment* **164**:80–99.
- Striganova BR. 1980.** *Nutrition of soil saprophages*. Moscow: Nauka Science Publications[in Russian], 243.
- Strong DT, De Wever H, Merckx R, Recous S. 2004.** Spatial location of carbon decomposition in the soil pore system. *Soil Science* **5**:739–750.
- Succurro A, Ebenhöf O. 2018.** Review and perspective on mathematical modeling of microbial ecosystems. *Biochemical Society Transactions* **46**(2):403–412 DOI [10.1042/BST20170265](https://doi.org/10.1042/BST20170265).
- Sulman B, Moore J, Abramoff R, Averill C, Kivlin S, Georgiou K, Sridhar B, Hartman MD, Wang G, Wieder WR, Bradford MA, Luo Y, Mayes MA, Morrison E, Riley WJ, Salazar A, Schimel JP, Tang J, Classen AT. 2018.** Multiple models and experiments underscore large uncertainty in soil carbon dynamics. *Biogeochemistry* **141**(2):1–15 DOI [10.1007/s10533-018-0509-z](https://doi.org/10.1007/s10533-018-0509-z).
- Sulman BN, Phillips RP, Oishi AC, Shevliakova E, Pacala SW. 2014.** Microbe-driven turnover offsets mineral-mediated storage of soil carbon under elevated CO<sub>2</sub>. *Nature Climate Change* **4**(12):1099–1102 DOI [10.1038/nclimate2436](https://doi.org/10.1038/nclimate2436).
- Sándor R, Barcza Z, Acutis M, Doro L, Hidy D, Köchy M, Minet J, Lellei-Kovács E, Ma S, Perego A, Rolinski S, Ruget F, Sanna M, Seddaiu G, Wu L, Bellocchi G. 2017.** Multi-model simulation of soil temperature, soil water content and biomass in Euro-Mediterranean grasslands: uncertainties and ensemble performance. *European Journal of Agronomy* **88**:22–40 DOI [10.1016/j.eja.2016.06.006](https://doi.org/10.1016/j.eja.2016.06.006).
- Sándor R, Barcza Z, Hidy D, Lellei-Kovács E, Ma S, Bellocchi G. 2016.** Modelling of grassland fluxes in Europe: evaluation of two biogeochemical models. *Agriculture, Ecosystems & Environment* **215**:1–19 DOI [10.1016/j.agee.2015.09.001](https://doi.org/10.1016/j.agee.2015.09.001).

- Söderström B. 1979.** Seasonal fluctuations of active fungal biomass in horizons of a podzolized pine-forest soil in central Sweden. *Soil Biology and Biochemistry* **11**(2):149–154  
DOI [10.1016/0038-0717\(79\)90093-2](https://doi.org/10.1016/0038-0717(79)90093-2).
- Söderström B, Read DJ. 1987.** Respiratory activity of intact and excised ectomycorrhizal mycelial systems growing in unsterilized soil. *Soil Biology and Biochemistry* **19**(3):231–236  
DOI [10.1016/0038-0717\(87\)90002-2](https://doi.org/10.1016/0038-0717(87)90002-2).
- Tagu D, De Bellis R, Balestrini R, De Vries OMH, Piccoli G, Stocchi V, Bonfante P, Martin F. 2001.** Immunolocalization of hydrophobin HYDpt-1 from the ectomycorrhizal basidiomycete *Pisolithus tinctorius* during colonization of *Eucalyptus globulus* roots. *New Phytologist* **149**(1):127–135 DOI [10.1046/j.1469-8137.2001.00009.x](https://doi.org/10.1046/j.1469-8137.2001.00009.x).
- Taylor BR, Parkinson D, Parsons WFJ. 1989.** Nitrogen and lignin content as predictors of litter decay rates: a microcosm test. *Ecology* **70**(1):97–104 DOI [10.2307/1938416](https://doi.org/10.2307/1938416).
- Tecon R, Or D. 2017.** Biophysical processes supporting the diversity of microbial life in soil. *FEMS microbiology reviews* **41**(5):599–623 DOI [10.1093/femsre/fux039](https://doi.org/10.1093/femsre/fux039).
- Tedersoo L, Bahram M. 2019.** Mycorrhizal types differ in ecophysiology and alter plant nutrition and soil processes. *Biological Reviews* **94**(5):1857–1880 DOI [10.1111/brv.12538](https://doi.org/10.1111/brv.12538).
- Tedersoo L, Bahram M, Zobel M. 2020.** How mycorrhizal associations drive plant population and community biology. *Science* **367**(6480):eaba1223 DOI [10.1126/science.aba1223](https://doi.org/10.1126/science.aba1223).
- Teshima J, Iwanaga R, Kobayashi T, Ikegami D, Yasutake D, Cho H. 2006.** An improvement on the BBH model of soil hydrology made to incorporate bioprocesses. *Journal of the Faculty of Agriculture, Kyushu University* **51**(2):399–402.
- Thomsen IK, Schjønning P, Jensen B, Kristensen K, Christensen BT. 1999.** Turnover of organic matter in differently textured soils. II. Microbial activity as influenced by soil water regimes. *Geoderma* **89**(3–4):199–218 DOI [10.1016/S0016-7061\(98\)00084-6](https://doi.org/10.1016/S0016-7061(98)00084-6).
- Tian J-H, Pourcher A-M, Bouchez T, Gelhaye E, Peu P. 2014.** Occurrence of lignin degradation genotypes and phenotypes among prokaryotes. *Applied Microbiology and Biotechnology* **98**(23):9527–9544 DOI [10.1007/s00253-014-6142-4](https://doi.org/10.1007/s00253-014-6142-4).
- Tippling E, Michalzik B, Mulder J, Gallardo LJF, Matzner E, Bryant C, Clarke N, Lofts SV, Esteban A. 2001.** DyDOC, a model to describe carbon dynamics in forest soils. In: *Borggaard, Ole K. (Hrsg.): the proceedings of 8th Nordic IHSS Symposium on humic substances - characterisation, dynamics, transport and effects*. 122–126.
- Tippling E, Rowe EC, Evans CD, Mills RTE, Emmett BA, Chaplow JS, Hall JR. 2012.** N14C: a plant–soil nitrogen and carbon cycling model to simulate terrestrial ecosystem responses to atmospheric nitrogen deposition. *Ecological Modelling* **247**:11–26  
DOI [10.1016/j.ecolmodel.2012.08.002](https://doi.org/10.1016/j.ecolmodel.2012.08.002).
- Tiunov AV, Scheu S. 2000.** Microfungal communities in soil, litter and casts of *Lumbricus terrestris* L. (Lumbricidae): a laboratory experiment. *Applied Soil Ecology* **14**(1):17–26  
DOI [10.1016/S0929-1393\(99\)00050-5](https://doi.org/10.1016/S0929-1393(99)00050-5).
- Toosi ER, Kravchenko AN, Guber AK, Rivers ML. 2017.** Pore characteristics regulate priming and fate of carbon from plant residue. *Soil Biology and Biochemistry* **113**:219–230  
DOI [10.1016/j.soilbio.2017.06.014](https://doi.org/10.1016/j.soilbio.2017.06.014).
- Torsvik V, Ovreas L. 2002.** Microbial diversity and function in soil: from genes to ecosystems. *Current Opinion in Microbiology* **5**(3):240–245 DOI [10.1016/S1369-5274\(02\)00324-7](https://doi.org/10.1016/S1369-5274(02)00324-7).
- Toyota A, Hynst J, Cajthaml T, Frouz J. 2013.** Soil fauna increase nitrogen loss in tilled soil with legume but reduce nitrogen loss in non-tilled soil without legume. *Soil Biology and Biochemistry* **60**:105–112 DOI [10.1016/j.soilbio.2013.01.017](https://doi.org/10.1016/j.soilbio.2013.01.017).

- Trap J, Bonkowski M, Plassard C, Villenave C, Blanchart E. 2016.** Ecological importance of soil bacterivores for ecosystem functions. *Plant and Soil* **398**(1–2):1–24 DOI [10.1007/s11104-015-2671-6](https://doi.org/10.1007/s11104-015-2671-6).
- Treonis AM, Grayston SJ, Murray PJ, Cook R, Currie AF, Dawson LA, Gange AC. 2001.** Impact of invertebrate root feeders on rhizosphere C flow and soil microbial communities. In: *Abstracts 9th International Symposium on Soil Microbial Ecology (ISME 9), 26 August–1 September 2001, Amsterdam*. 270.
- Treonis AM, Ostle NJ, Stott AW, Primrose R, Grayston SJ, Ineson P. 2004.** Identification of groups of metabolically-active rhizosphere microorganisms by stable isotope probing of PLFAs. *Soil Biology and Biochemistry* **36**(3):533–537 DOI [10.1016/j.soilbio.2003.10.015](https://doi.org/10.1016/j.soilbio.2003.10.015).
- Treseder KK. 2016.** Model behavior of arbuscular mycorrhizal fungi: predicting soil carbon dynamics under climate change. *Botany* **94**(6):417–423 DOI [10.1139/cjb-2015-0245](https://doi.org/10.1139/cjb-2015-0245).
- Treseder KK, Baiser TC, Bradford MA, Brodie EL, Dubinsky EA, Eviner VT, Hofmockel KS, Lennon JT, Levine UY, MacGregor BJ, Pett-Ridge J, Waldrop MP. 2012.** Integrating microbial ecology into ecosystem models: challenges and priorities. *Biogeochemistry* **109**(1–3):7–18 DOI [10.1007/s10533-011-9636-5](https://doi.org/10.1007/s10533-011-9636-5).
- Treseder KK, Cross A. 2006.** Global distributions of arbuscular mycorrhizal fungi. *Ecosystems* **9**(2):305–316 DOI [10.1007/s10021-005-0110-x](https://doi.org/10.1007/s10021-005-0110-x).
- Tuomi M, Rasinmäki J, Repo A, Vanhala P, Liski J. 2011.** Soil carbon model Yasso07 graphical user interface. *Environmental Modelling & Software* **26**(11):1358–1362 DOI [10.1016/j.envsoft.2011.05.009](https://doi.org/10.1016/j.envsoft.2011.05.009).
- Tupek B, Launiainen S, Peltoniemi M, Sievänen R, Perttunen J, Kulmala L, Penttilä T, Lindroos A-J, Hashimoto S, Lehtonen A. 2019.** Evaluating CENTURY and Yasso soil carbon models for CO<sub>2</sub> emissions and organic carbon stocks of boreal forest soil with Bayesian multi-model inference. *European Journal of Soil Science* **14**:2636 DOI [10.1111/ejss.12805](https://doi.org/10.1111/ejss.12805).
- Uroz S, Calvaruso C, Turpault MP, Frey-Klett P. 2009.** Mineral weathering by bacteria: ecology, actors and mechanisms. *Trends in Microbiology* **17**(8):378–387 DOI [10.1016/j.tim.2009.05.004](https://doi.org/10.1016/j.tim.2009.05.004).
- Van Breemen N, Boyer EW, Goodale CL, Jaworski NA, Paustian K, Seitzinger SP, Lajtha K, Mayer B, Van Dam D, Howarth RW, Nadelhoffer KJ, Eve M, Billen G. 2002.** Where did all the nitrogen go? Fate of nitrogen inputs to large watersheds in the northeastern U.S.A. *Biogeochemistry* **57**(1):267–293 DOI [10.1023/A:1015775225913](https://doi.org/10.1023/A:1015775225913).
- Van Breemen N, Burman P. 2002.** *Soil formation*. The Netherlands: Kluwer Academic Publishers.
- Van der Heijden MGA, Bardgett RD, Van Straalen NM. 2008.** The unseen majority: soil microbes as drivers of plant diversity and productivity in terrestrial ecosystems. *Ecology Letters* **11**(3):296–310 DOI [10.1111/j.1461-0248.2007.01139.x](https://doi.org/10.1111/j.1461-0248.2007.01139.x).
- Van der Heijden MGA, Martin FM, Selosse MA, Sanders IR. 2015.** Mycorrhizal ecology and evolution: the past, the present, and the future. *New Phytologist* **205**(4):1406–1423 DOI [10.1111/nph.13288](https://doi.org/10.1111/nph.13288).
- Van der Wal A, Geydan TD, Kuyper TW, De Boer W. 2013.** A thready affair: linking fungal diversity and community dynamics to terrestrial decomposition processes. *FEMS Microbiology Reviews* **37**(4):477–494 DOI [10.1111/1574-6976.12001](https://doi.org/10.1111/1574-6976.12001).
- Van Genuchten MT, Leij FJ, Wu L. 1999.** *Characterization and measurement of the hydraulic properties of unsaturated porous media*. Riverside: University of California.

- Van Looy K, Bouma J, Herbst M, Koestel J, Minasny B, Mishra U, Vereecken H. 2017. Pedotransfer functions in earth system science: challenges and perspectives. *Reviews of Geophysics* 55(4):1199–1256 DOI 10.1002/2017RG000581.
- Vereecken H, Schnepf A, Hopmans J, Javaux M, Or D, Roose T, Vanderborght J, Young M, Amelung W, Aitkenhead M, Allison S, Assouline S, Baveye P, Berli M, Brüggemann N, Finke P, Flury M, Gaiser T, Govers G, Young I. 2016. Modeling soil processes: review, key challenges and new perspectives. *Vadose Zone Journal* 15:1–57.
- Vereecken H, Weihermuller L, Assouline SI, Verhoef A, Herbst M, Archer N, Mohanty B, Montzka C, Vanderborght J. 2019. Infiltration from the Pedon to global grid scales: an overview and outlook for land surface modeling. *Vadose Zone Journal* 18(1):1–53 DOI 10.2136/vzj2018.10.0191.
- Von Lützw M, Kögel-Knabner I, Ekschmitt K, Matzner E, Guggenberger G, Marschner B, Flessa H. 2006. Stabilization of organic matter in temperate soils: mechanisms and their relevance under different soil conditions—a review. *European Journal of Soil Science* 57(4):426–445 DOI 10.1111/j.1365-2389.2006.00809.x.
- Von Lützw M, Kögel-Knabner I, Ludwig B, Matzner E, Flessa H, Ekschmitt K, Guggenberger G, Marschner B, Kalbitz K. 2008. Stabilization mechanisms of organic matter in four temperate soils: development and application of a conceptual model. *Journal of Plant Nutrition and Soil Science* 171(1):111–124 DOI 10.1002/jpln.200700047.
- Vorpahl P, Moenickes S, Richter O. 2009. Modelling of spatio-temporal population dynamics of earthworms under wetland conditions—an integrated approach. *Ecological Modelling* 220(24):3647–3657 DOI 10.1016/j.ecolmodel.2009.03.012.
- Wagg CS, Benders F, Widmer F, Van der Heijden MGA. 2013. Soil biodiversity and soil community composition determine ecosystem multifunctionality. *Proceedings of the National Academy of Sciences of the United States of America* 111(14):5266–5270 DOI 10.1073/pnas.1320054111.
- Wallander H, Goransson H, Rosengren U. 2004. Production, standing biomass and natural abundance of <sup>15</sup>N and <sup>13</sup>C in ectomycorrhizal mycelia collected at different soil depths in two forest types. *Oecologia* 139(1):89–97 DOI 10.1007/s00442-003-1477-z.
- Wallwork JA. 1970. *Ecology of soil animals*. London: McGraw-Hill.
- Wardle DA, Yeates GW, Watson RN, Nicholson KS. 1995. The detritus food-web and the diversity of soil fauna as indicators of disturbance regimes in agro-ecosystems. *Plant and Soil* 170(1):35–43 DOI 10.1007/BF02183053.
- Waring BG, Averill C, Hawkes CV. 2013. Differences in fungal and bacterial physiology alter soil carbon and nitrogen cycling: insights from meta-analysis and theoretical models. *Ecology Letters* 16(7):887–894 DOI 10.1111/ele.12125.
- Warren JM, Brooks JR, Meinzer FC, Eberhart JL. 2008. Hydraulic redistribution of water from *Pinus ponderosa* trees to seedlings: evidence for an ectomycorrhizal pathway. *New Phytologist* 178(2):382–394 DOI 10.1111/j.1469-8137.2008.02377.x.
- Weng L, Van Riemsdijk WH, Hiemstra T. 2008. Humic nanoparticles at the oxide-water interface: interactions with phosphate ion adsorption. *Environmental Science & Technology* 42(23):8747–8752 DOI 10.1021/es801631d.
- Widder S, Allen RJ, Pfeiffer T, Curtis TP, Wiuf C, Sloan WT, Cordero OX, Brown SP, Momeni B, Shou W, Kettle H, Flint HJ, Haas AF, Laroche B, Kreft J-U, Rainey PB, Freilich S, Schuster S, Milferstedt K, van der Meer JR, Großkopf T, Huisman J, Free A, Picioreanu C, Quince C, Klapper I, Labarthe S, Smets BF, Wang H, Soyer OS, Isaac Newton Institute. 2016.

- Challenges in microbial ecology: building predictive understanding of community function and dynamics. *ISME Journal* **10**(11):2557–2568 DOI [10.1038/ismej.2016.45](https://doi.org/10.1038/ismej.2016.45).
- Wieder WR, Grandy AS, Kallenbach CM, Bonan GB. 2014.** Integrating microbial physiology and physio-chemical principles in soils with the MICrobial-MINeral Carbon Stabilization (MIMICS) model. *Biogeosciences* **11**(14):3899–3917 DOI [10.5194/bg-11-3899-2014](https://doi.org/10.5194/bg-11-3899-2014).
- Wieder WR, Grandy AS, Kallenbach CM, Taylor PG, Bonan GB. 2015.** Representing life in the Earth system with soil microbial functional traits in the MIMICS model. *Geoscientific Model Development* **8**(6):1789–1808 DOI [10.5194/gmd-8-1789-2015](https://doi.org/10.5194/gmd-8-1789-2015).
- Wilkinson MT, Richards PJ, Humphreys GS. 2009.** Breaking ground: pedological, geological, and ecological implications of soil bioturbation. *Earth-Science Reviews* **97**(1–4):257–272 DOI [10.1016/j.earscirev.2009.09.005](https://doi.org/10.1016/j.earscirev.2009.09.005).
- Williams BL, Griffiths BS. 1989.** Enhanced nutrient mineralization and leaching from decomposing sitka spruce litter by enchytraeid worms. *Soil Biology and Biochemistry* **21**(2):183–188 DOI [10.1016/0038-0717\(89\)90093-X](https://doi.org/10.1016/0038-0717(89)90093-X).
- Wilson GWT, Rice CW, Rillig MC, Springer A, Hartnett DC. 2009.** Soil aggregation and carbon sequestration are tightly correlated with the abundance of arbuscular mycorrhizal fungi: results from long-term field experiments. *Ecology Letters* **12**(5):452–461 DOI [10.1111/j.1461-0248.2009.01303.x](https://doi.org/10.1111/j.1461-0248.2009.01303.x).
- Yang X, Ricciuto D, Thornton P, Shi X, Xu M, Hoffman F, Norby R. 2019.** The effects of phosphorus cycle dynamics on carbon sources and sinks in the amazon region: a modeling study using ELM v1. *Journal of Geophysical Research: Biogeosciences* **124**(12):3686–3698 DOI [10.1029/2019JG005082](https://doi.org/10.1029/2019JG005082).
- Yeates GW, Bardgett RD, Cook R, Hobbs PJ, Bowling PJ, Potter JF. 1997.** Faunal and microbial diversity in three welsh grassland soils under conventional and organic management regimes. *Journal of Applied Ecology* **34**(2):453–470 DOI [10.2307/2404890](https://doi.org/10.2307/2404890).
- Yeates GW, Bongers T, De Goede RGM, Freckman DW, Georgieva SS. 1993.** Feeding habits in soil nematode families and genera—an outline for soil ecologists. *Journal of Nematology* **25**(3):315.
- Yin H, Wheeler E, Phillips RP. 2014.** Root-induced changes in nutrient cycling in forests depend on exudation rates. *Soil Biology and Biochemistry* **78**:213–221 DOI [10.1016/j.soilbio.2014.07.022](https://doi.org/10.1016/j.soilbio.2014.07.022).
- Yudina A, Kuzyakov Y. 2019.** Saving the face of soil aggregates. *Global Change Biology* **25**(11):3574–3577 DOI [10.1111/gcb.14779](https://doi.org/10.1111/gcb.14779).
- Zhang D, Hui D, Luo Y, Zhou G. 2008.** Rates of litter decomposition in terrestrial ecosystems: global patterns and controlling factors. *Journal of Plant Ecology* **1**(2):85–93 DOI [10.1093/jpe/rtn002](https://doi.org/10.1093/jpe/rtn002).
- Zhang XK, Li Q, Liang WJ, Zhang M, Bao XL, Xie ZB. 2013.** Soil nematode response to biochar addition in a Chinese wheat field. *Pedosphere* **23**(1):98–103 DOI [10.1016/S1002-0160\(12\)60084-8](https://doi.org/10.1016/S1002-0160(12)60084-8).
- Zhou G, Xu S, Ciais P, Manzoni S, Fang J, Yu G, Tang X, Zhou P, Wang W, Yan J, Wang G, Ma K, Li S, Du S, Han S, Ma Y, Zhang D, Liu J, Liu S, Chu G, Zhang Q, Li Y, Huang W, Ren H, Lu X, Chen X. 2019.** Climate and litter C/N ratio constrain soil organic carbon accumulation. *National Science Review* **6**(4):746–757 DOI [10.1093/nsr/nwz045](https://doi.org/10.1093/nsr/nwz045).
- Zvereva EL, Kozlov MV. 2012.** Sources of variation in plant responses to belowground insect herbivory: a meta-analysis. *Oecologia* **169**(2):441–452 DOI [10.1007/s00442-011-2210-y](https://doi.org/10.1007/s00442-011-2210-y).