

Role of methyltransferase-like enzyme 3 and methyltransferase-like enzyme 14 of N6-methyladenosine in the urological cancers

Zijia Tao¹, Yiqiao Zhao¹, Xiaonan Chen^{Corresp. 1}

¹ Department of Urology, Shengjing Hospital of China Medical University, Shenyang, Liaoning, China

Corresponding Author: Xiaonan Chen
Email address: chenxn@cmu.edu.cn

N6-methyladenosine (m6A) modification can modify eukaryotic messenger RNA (mRNA), long non-coding RNAs (lncRNAs), microRNA (miRNA). A number of studies find that m6A show close relationship with cancer cells. Methyltransferase-like enzyme 3 (METTL3) and methyltransferase-like enzyme (METTL14), as two major enzymes in m6A modification, play a vital role in various cancers. However, the role and regulation mechanism of METTL3 and METTL14 in urological cancers are largely unknown. In this review, we summarize the present research results of METTL3/METTL14 and find potential pathways in kidney cancer, bladder cancer and prostate cancer. We found that METTL3 and METTL14 show different expression in three types of urological cancers. METTL3 is high expression in bladder/prostate cancer and has a positive regulatory effect on cancer cells. But its expression and role are opposite in kidney cancer. METTL14 is always low-expressed in kidney and bladder cancer, playing a negative regulatory role. When METTL3 or METTL14 is low expression in cancer cells, they always negatively regulate cell growth-pathways, such as mTOR, EMT, P2XR6, but positively regulate cell death-related pathways, such as P53, PTEN and Notch1. When METTL3 is high expression, it positively regulates NF- κ B/SHH-GL1 pathways and negatively regulates PTEN. These results suggest that although METTL3 and METTL14 show different expression and regulation mechanism in urological cancers. But they control cancer cell fate by cell growth- and cell death-related pathways. It provides a potential possibility for using m6A as a new therapeutic target in urological cancer.

1 **Role of methyltransferase-like enzyme 3 and**
2 **methyltransferase-like enzyme 14 of N6-**
3 **methyladenosine in the urological cancers**

4

5

6 Zijia Tao¹, Yiqiao Zhao¹, Xiaonan Chen^{1,*}

7

8 ¹Department of Urology, Shengjing Hospital of China Medical University, Shenyang, Liaoning
9 110004, People's Republic of China

10

11

12

13

14 * Corresponding Author:

15 Xiaonan Chen

16 Department of Urology, Shengjing Hospital of China Medical University, No. 36 Sanhao Street,
17 Heping District, Shenyang 110004, Liaoning, People's Republic of China

18 Email: chenxn@cmu.edu.cn

19

20

21

22

23

24

25

26

27

28 **Abstract**

29 N6-methyladenosine (m6A) modification can modify eukaryotic messenger RNA (mRNA), long
30 non-coding RNAs (lncRNAs), microRNA (miRNA). A number of studies find that m6A show
31 close relationship with cancer cells. Methyltransferase-like enzyme 3 (METTL3) and
32 methyltransferase-like enzyme (METTL14), as two major enzymes in m6A modification, play a
33 vital role in various cancers. However, the role and regulation mechanism of METTL3 and
34 METTL14 in urological cancers are largely unknown. In this review, we summarize the present
35 research results of METTL3/METTL14 and find potential pathways in kidney cancer, bladder
36 cancer and prostate cancer. We found that METTL3 and METTL14 show different expression in
37 three types of urological cancers. METTL3 is high expression in bladder/prostate cancer and has
38 a positive regulatory effect on cancer cells. But its expression and role are opposite in kidney
39 cancer. METTL14 is always low-expressed in kidney and bladder cancer, playing a negative
40 regulatory role. When METTL3 or METTL14 is low expression in cancer cells, they always
41 negatively regulate cell growth-pathways, such as mTOR, EMT, P2XR6, but positively regulate
42 cell death-related pathways, such as P53, PTEN and Notch1. When METTL3 is high expression,
43 it positively regulates NF-kB/SHH-GL1 pathways and negatively regulates PTEN. These results
44 suggest that although METTL3 and METTL14 show different expression and regulation
45 mechanism in urological cancers. But they control cancer cell fate by cell growth- and cell death-
46 related pathways. It provides a potential possibility for using m6A as a new therapeutic target in
47 urological cancer.

48

49 **Introduction**

50 Chemical modifications of eukaryotic RNAs are known from decades. However, their role in
51 tumor development was largely unknown until recent years. According to MODOMICS data,
52 163 different RNA chemical modifications have been identified in all living entities (Boccaletto
53 et al. 2018). N6-methyladenosine (m6A) modification is considered to be one of the most
54 common, invertible and ample modifications inside the RNA molecules, which can modify
55 eukaryotic messenger RNA (mRNA), microRNA (miRNA), long non-coding RNAs (lncRNAs)
56 and other RNA molecules, and affects the transcription, processing, translation and metabolism
57 of these molecules (Zheng et al. 2019). m6A modification is a dynamic process involving three
58 major classes of enzymes: 'Writers', 'Erasers' and 'Readers' (Vu et al. 2019). Writers include
59 methyltransferase-like enzyme 3 (METTL3), methyltransferase-like enzyme (METTL14),
60 Wilms tumor 1 associated protein (WTAP), RNA Binding Motif Protein 15/15B (RBM15/15B)

61 , and Vir Like M6A Methyltransferase Associated (VIRMA) , which are responsible for
62 catalyzing the process of m6A generation; Erasers include fat and obesity-related protein (FTO)
63 and alkB homolog 5 (ALKBH5) , which are responsible for catalyzing the demethylation
64 process; Readers include proteins containing the YT521-B homology (YTH) domain ,
65 eukaryotic initiation factor (eIF3) , the IGF2 mRNA binding protein (IGF2BP) family , and
66 heterogeneous nuclear ribonucleoproteins (HNRNP) protein family , they recognize m6A
67 methylation and generate functional signals(Chen et al. 2019b).

68 METTL3 was first identified as a 70 kDa protein from Hela cell lysate (Bokar et al. 1997),
69 subsequent research found that it contains two domains that bind S-adenosylmethionine (SAM)
70 and catalyzes the formation of m6A, and has the activity of independently catalyzing the
71 formation of m6A modification of RNA(Leach & Tuck 2001). WTAP promotes METTL3
72 localization in nuclear spots and greatly improves its catalytic activity (Ping et al. 2014). Studies
73 have also shown that METTL3 acts as a positive regulator of mRNA translation independent of
74 methyltransferase activity: promoting translation by interacting with translation initiation
75 mechanisms in the cytoplasm (Ke et al. 2017). It has been reported that METTL3 can play the
76 role without METTL14 and can promote translation of specific mRNAs independently of its
77 catalytic activity in vitro (Ke et al. 2017). METTL3 is the core of catalytic activity in the N6-
78 methyltransferase complex formed by the METTL3-METTL14 heterodimer. Adenosine residues
79 at the N (6) position of some RNAs are methylated this complex (5, 9-21). METTL14 is a
80 scaffold of binding RNA that identifies the substrate of N6-methyltransferase complex formed
81 by METTL3-METTL14 heterodimer (Liu et al. 2014; Liu et al. 2015; Ping et al. 2014; Scholler
82 et al. 2018; Sledz & Jinek 2016; Wang et al. 2016a; Wang et al. 2016b). METTL14 shares about
83 22% sequence identity and a nearly identical topology with the domains of METTL3. In the
84 METTL3-METTL14 heterodimer, METTL14 is thought to assume a pseudomethyltransferase
85 function that helps to bind with RNA and stabilize METTL3. However, it is possible that
86 methyltransferase activity of METTL14 manifests after binding additional factors (Wang et al.
87 2017). In mRNA, the methylation site is located in the 5 '-[AG] GAC-3' consensus site of some
88 mRNAs, and plays an important role in mRNA stability, processing, translation efficiency and
89 editing (Alarcon et al. 2015a; Alarcon et al. 2015b; Bokar et al. 1997; Dominissini et al. 2012;
90 Liu et al. 2015; Meyer et al. 2015; Wang et al. 2014; Xiang et al. 2017). It has been shown that
91 methylation is completed after the mRNA is released into the nucleoplasm and promotes mRNA
92 instability and degradation (Ke et al. 2017).

93 With the continuous research in recent years, the role of m6A in various tumors has been
94 revealed, such as leukemia, brain tumor, cervical tumor, endometrial tumor, breast tumor, liver
95 tumor and lung tumor etc.(Chen et al. 2018; Choe et al. 2018; Liu et al. 2018; Vu et al. 2017;

96 Weng et al. 2018; Zhang et al. 2016; Zhang et al. 2017). m6A shows a significant role of
97 regulatory function in tumorigenesis and development by modifying many target genes (Deng et
98 al. 2018; Liu et al. 2018). Interestingly, m6A may have carcinogenic or suppressive functions of
99 tumor in different environments in cells (Cui et al. 2017; Li et al. 2017b; Lin et al. 2016; Ma et
100 al. 2017; Visvanathan et al. 2018; Vu et al. 2017; Zhang et al. 2016; Zhang et al. 2017). A study
101 proved that METTL3 mediated post-transcriptional silencing of SOCS2 through YTHDF2
102 dependent way and promoted liver cancer progression (Chen et al. 2018). METTL14 was
103 confirmed that it lowly expresses in liver cancer cells and hematopoietic stem cells, and it
104 impairs the tumorigenesis of Acute Myelocytic Leukemia (AML) (Weng et al. 2018). It was also
105 revealed that METTL14 exhibits an inhibitory role in liver tumorigenesis and metastasis (Li et
106 al. 2017b). Some studies have shown that abnormal m6A modification is necessary for the
107 growth and progression in cancer (Cui et al. 2017; Li et al. 2017b; Lin et al. 2016; Ma et al.
108 2017; Visvanathan et al. 2018; Vu et al. 2017; Zhang et al. 2016; Zhang et al. 2017), suggesting
109 that the pathway of m6A modification may be a promising therapeutic target for malignant
110 tumors. In this review, we summarized the present research progress about METTL3 and
111 METTL14 in urological tumors.

112

113 **Survey methodology**

114 In order to search literatures exhaustively, we used keywords ‘METTL3’, ‘Methyltransferase-
115 like enzyme 3’, ‘METTL14’, ‘Methyltransferase-like enzyme 14’, ‘kidney cancer’, ‘renal cell
116 carcinoma’, ‘bladder cancer’, ‘prostate cancer’ to search articles in the PubMed, Web of Science
117 and CNKI. We excluded the articles which were not associated with METTL3, METTL14 and
118 urological cancers.

119

120 **Methyltransferase in urological cancer**

121

122 **1.Kidney cancer**

123 According to the data of GLOBOCAN statistics, 403,262 people were diagnosed with renal
124 cancer throughout the world and 175,098 people died, which made renal cancer become the 14th
125 most common cancer in the world. Besides, males have a higher incidence than females in renal
126 cancer (Bray et al. 2018). Many patients remain asymptomatic until the renal masses (RMs)
127 progress to advanced stages because of the special position of kidney in the body. In the World
128 Health Organization (WHO) 2016 classification, renal cancers are divided into three main sub

129 categories: clear cell renal cell carcinoma (ccRCC), which is the most common and aggressive
130 type; chromophobe renal cell carcinoma (chRCC); and papillary renal cell carcinoma (pRCC)
131 which has two styles, types 1 and 2(Hao et al. 2019; Moch et al. 2016). So far, there are four
132 articles studying methyltransferase in kidney cancer, one on METTL3, two on METTL14 and
133 one for both. All of them suggested that METTL3 and METTL14 are tumor suppressor in renal
134 cancer. In addition, two articles studied about ccRCC subtypes, others were unknown (Gong et
135 al. 2019; Li et al. 2017a; Wang et al. 2019; Zhou et al. 2019).

136

137 **1.1 METTL3 in kidney cancer**

138 METTL3 is more prone to copy number variations (CNV) or mutation than other genes in
139 ccRCC, and patients affected by METTL3 shallow deletions (a form of CNV) has poorer
140 disease-free survival (DFS) and overall survival (OS)(Zhou et al. 2019). METTL3 mRNA and
141 protein expression in RCC are lower. Its expression level is negatively related with higher
142 histological grade, larger tumor size, shorter OS and shorter DFS (Li et al. 2017a; Zhou et al.
143 2019). Study from cell line of RCC (CAKI-1/2 and ACHN) examined knocking down METTL3
144 can significantly promote cancer cell proliferation, migration and invasion (Li et al. 2017a). The
145 VHL-HIF-ZNF217-METTL3 pathway may be involved in m6A regulation in ccRCC cells by
146 mediating two downstream targets of m6A: the PI3K/AKT/mTOR and p53 signaling pathways
147 (Li et al. 2017a; Zhou et al. 2019). In various cellular processes, the PI3K/AKT/mTOR pathway
148 shows a significant role on cell proliferation, growth and survival (O'Reilly et al. 2006; Shaw &
149 Cantley 2006). In addition, METTL3 might be involved in down-regulated invasion and
150 migration of RCC by epithelial-mesenchymal transition (EMT) pathway. In the procedure of
151 EMT, epithelial cells gain mesenchymal fibroblast-like properties (Lim et al. 2014), which may
152 provide motility, migration and invasion functions for tumor cells (Fuchs et al. 2008; McConkey
153 et al. 2009) and contribute to the potential of tumor metastasis. Besides, it was demonstrated that
154 EMT is related to the prognosis of RCC patients (Chen et al. 2014). When METTL3 is down-
155 regulated, the expression of vimentin, β -catenin and N-cadherin are significantly at a higher
156 level, while E-cadherin is at a lower level. Based on the above results, pathway of EMT may be
157 involved in potential mechanisms. (Li et al. 2017a).

158 The GSEA analysis with ccRCC patients suggested that low expression level of METTL3 is also
159 related to some critical biological processes, such as mTOR pathway, adipogenesis and reactive
160 oxygen species (ROS), which partially validate the RCC cell line results. Therefore, evidences
161 from human tissues and cell line implied that the mTOR pathway, compared to other pathway,

162 may be the key target of m6A modification in kidney cancer. On the other hand, METTL3 can
163 also regulate cell cycle. It was indicated that down-regulation of METTL3 significantly decrease
164 arrest in G1 phase of cell cycle, whereas up-regulation of METTL3 increase arrest in G1 phase
165 (Zhou et al. 2019).

166

167 **1.2. METTL14 in kidney cancer**

168 Similar to METTL3, METTL14 are more predisposed to mutation or CNV , and patients
169 affected by shallow deletions of METTL14 have poorer OS and DFS(Zhou et al. 2019).
170 METTL14 is mainly located in the nucleus of ccRCC cells. Compared with normal kidney
171 tissues, the expression of METTL14 mRNA is significantly lower in ccRCC tissues. It was
172 shown that the expression level of METTL14 has a significantly negative correlation with RCC
173 pathological stages and clinical stages, while OS is opposite (Wang et al. 2019). For METTL14,
174 VHL-HIF-ZNF217-METTL14 pathway regulates m6A in ccRCC cells by two pathways: the
175 PI3K/AKT/mTOR and p53 signaling, which is same with METTL3(Li et al. 2017a; Zhou et al.
176 2019). Other two regulating pathways in METTL14 also have been found. The first one is
177 P2RX6, a non-selective cation channel protein which is a preferred receptor for ATP (Chadet et
178 al. 2014; North 2002). METTL14 expression is negatively correlated with P2RX6, and low
179 METTL14 expression shows shorter OS, while P2RX6 is opposite. METTL14 may increase the
180 pre-mRNA splicing of P2RX6 by increasing the methylation of P2RX6 mRNA and achieving
181 inhibition effect of P2RX6. Low METTL14 expression in cancer cells leads to high P2RX6
182 expression by acting on the ATP-P2RX6-Ca²⁺-p-ERK1/2-MMP9 signaling pathway and
183 promotes renal tumor cell metastasis and invasion (Gong et al. 2019). The second one is PTEN, a
184 tumor suppressor, whose duty is encoding phosphatidylinositol-3,4,5-trisphosphate 3-
185 phosphatase and preferentially dephosphorylating phosphoinositide substrates. The METTL14
186 mRNA expression level is positively related to PTEN. Low METTL14 and PTEN expression
187 show shorter OS. METTL14 stabilizes PTEN mRNA by regulating the m6A level of PTEN
188 mRNA. PTEN acts as a tumor suppressor through regulating the AKT/PKB signaling pathway
189 negatively. Besides, synergistic effects may be generated by the interaction between EIF3A and
190 METTL14, which regulate the progress of kidney cancer. In addition, further study showed that
191 24 circRNAs (such as circ-0023414, circ-0031772 etc.) interact with 4 miRNAs(miR-130a-3p,
192 miR-106b-5p , miR-130b-3p, and miR-301a-3p), which have a significantly negative relation
193 with METTL14 mRNA, and these circRNAs may act as 4 miRNAs sponge and regulate
194 METTL14 mRNA (Wang et al. 2019).

195 In general, METTL3 and METTL4 show low expression in kidney cancer and play negative
196 effect on regulation of cancer cell. Low expression of METTL3 promotes cancer cell growth,
197 proliferation, motility, invasion and migration functions through negative regulation of mTOR
198 pathway, EMT pathway and positive regulation of P53. For the METTL4, in addition to
199 regulating mTOR pathway and P53 pathway as same as METTL3, METTL14 plays negative
200 role in kidney cancer also by negative regulation of P2RX6 pathway and positive regulation of
201 PTEN (Fig.1).

202

203 **2. Bladder Cancer**

204 Bladder cancer was the 12th most common cancer in the world, with 549,393 new diagnosed
205 cases and 199,922 deaths in 2018, according to the data of GLOBOCAN statistics. Besides,
206 incidence varies by gender, with males taking greater risks than females (Bray et al. 2018).
207 Urothelial carcinoma is the common histological style of bladder cancer. The non-papillary
208 muscle-invasive tumors and the papillary non-muscle-invasive ones are two main molecular,
209 clinical and pathological types of the disease (Sanli et al. 2017; Wu et al. 2019). So far, there are
210 seven articles studying methyltransferase in bladder cancer, six on METTL3 and one on
211 METTL14. Most of them suggested that METTL3 is tumor promoter in bladder cancer, and
212 METTL14 is tumor suppressor.

213

214 **2.1 METTL3 in bladder cancer**

215 METTL3 highly expresses in bladder cancer tissues (Chen et al. 2019a). METTL3 mRNA and
216 protein expression in bladder cancer significantly increase, Over-expressing METTL3
217 significantly promote the growth and invasion of bladder tumor cells. Knocking down of
218 METTL3 impairs bladder tumor cells' ability of proliferation, invasion and viability, reduces the
219 proportion of cell cycle S phase and raises the proportion of cell cycle G1 phase. Therefore,
220 METTL3 may maintain the characteristics of bladder cancer stem cells by inducing m6A
221 modification of SOX2, a marker of bladder cancer stem cells both in vivo and in vitro (Zhu et al.
222 2017). Patients with high METTL3 expression in bladder cancer have higher histological scores,
223 worse prognosis, and shorter survival time. METTL3 exhibits a carcinogenic role in bladder
224 cancer.

225 AFF4/NF- κ B/MYC signal network plays the important role in mediated up-regulation of
226 METTL3 on bladder cancer. This network contains three patterns: (1) METTL3 directly
227 increases the m6A site abundance of MYC mRNA to improve the stability of MYC transcripts,

228 and increase the expression of MYC protein; (2) METTL3 increases the m6A site abundance of
229 AFF4 mRNA to improve the stability of AFF4 transcripts and increase the expression of AFF4
230 protein. AFF4 protein directly combines with the promoter of MYC to promote the extension of
231 MYC transcription and up-regulate MYC expression; (3) METTL3 may promote the expression
232 of IKBKB and RELA (two key regulators of the NF- κ B pathway) by regulating translation
233 efficiency, and then induce MYC transcription. It is not difficult to find that METTL3-mediated
234 m6A modification of different signal pathways eventually converges to MYC expression.
235 Therefore, this m6A-regulated malignant regulatory network effectively increases the level of
236 MYC protein in bladder cancer and may lead to difficulties in reducing MYC results by blocking
237 single signaling (Cheng et al. 2019).

238 There is another pathway in mediating up-regulation of METTL3 in bladder cancer: METTL3-
239 DGCR8-pri-mi221/222-PTEN pathway. METTL3 can actively regulate pri-miR221/222 in a
240 m6A-dependent manner by interacting with the DGCR8, a micro-processor protein, and promote
241 pri-miR221/222 to become a mature miR221/222 in bladder cancer. Subsequently, miR221/222
242 binds to 3'-untranslated region (UTR) of PTEN mRNA, which leads to a decrease in mRNA
243 expression of PTEN and finally to a reduction in protein expression of PTEN (Han et al. 2019).

244 Two other pathways have also been identified for up-regulation of METTL3 on bladder cancer:
245 METTL3 -CDCP1 axis and METTL3-ITGA6 axis. METTL3 and CDCP1 are up-regulated in the
246 samples of bladder cancer patients. METTL3 and CDCP1 are related with the status of
247 progression in the bladder cancer. Inhibiting the METTL3-CDCP1 axis leads to reduced growth
248 and progression of bladder cancer cells and chemical-transformed cells. In vitro and vivo, it is
249 important that METTL3-CDCP1 axis and chemical carcinogens have synergistic effect in
250 promoting malignant transformation of uroepithelial cells and bladder cancer tumorigenesis
251 (Yang et al. 2019). For METTL3-ITGA6 axis, METTL3 highly enriches the m6A methylation
252 level of ITGA6 mRNA 3'-UTR region and promotes translation of ITGA6 mRNA, at the same
253 time, the binding of YTHDF1/ YTHDF3 to the m6A motif in ITGA6 3'UTR region also
254 promotes ITGA6 translation, resulting in increased expression of ITGA6 protein. The
255 overexpression of ITGA6 increases adhesion, proliferation, and migration of bladder tumor cells,
256 and enhances the growth and metastasis of bladder tumor cells. Therefore, ITGA6 as a
257 carcinogen is a key target of METTL3 function in bladder cancer (Jin et al. 2019).

258 However, there is a different viewpoint about the role of METTL3 in bladder cancer that
259 METTL3 is a bladder cancer suppressor gene. They worked out an integrated statistical model-
260 based method called driver MAPS that identifies METTL3 as driver gene in cohort bladder
261 cancer. Interestingly, in subsequent experimental verification, they found that knockdown of

262 METTL3 significantly increases cell proliferation, and METTL3 somatic mutations can promote
263 cancer cell growth by interrupting the methylation process of RNA. Therefore, they believed that
264 METTL3 acts as a gene of tumor suppressor for bladder cancer (Zhao et al. 2019).

265

266 **2.2 METTL14 in bladder cancer**

267 METTL14 has low expression level in bladder cancer and bladder tumor initiating cells (TICs).
268 TICs are a handful of cells in bladder cancer and have self-renewal, differentiation and tumor-
269 initiating abilities. METTL14 inhibits the proliferation, self-renewal, maintenance, metastasis
270 and bladder TICs tumor initiating ability. The expression of METTL14 is negatively associated
271 with the severity of bladder cancer and clinical outcome. METTL14 is significantly related to T
272 stage of TNM stage system (Chen et al. 2019a) and promotes the stability of some mRNA such
273 as MYC (Cheng et al. 2019).

274 Notch1 exhibits an important role in bladder tumorigenesis and TICs self-renewing. Notch1 is
275 downstream target of METTL14. m6A modification by METTL14 on Notch1 inhibits its RNA
276 stability, and then inhibits bladder cancer and bladder tumor initiating cells. Their work proved
277 that METTL14 is a gene of tumor suppressor in bladder cancer and plays a key role in the
278 occurrence of bladder tumors and bladder TICs through the METTL14-Notch1 pathway (Gu et
279 al. 2019).

280 In general, METTL3 shows high expression in bladder cancer and promotes cancer cell growth
281 through positive regulation of NF- κ B and negative regulation of PTEN. In contrast, low
282 expression level of METTL14 in bladder cancer plays negative effect in tumor cell by positive
283 regulation on Notch1, which also has a suppressive role in bladder TICs (Fig.2).

284

285 **3. Prostate Cancer**

286 Prostate cancer (PC) was the third most common cancer in the world, according to the data of
287 GLOBOCAN statistics, with 1,276,106 new diagnosed cases and 358,989 deaths in 2018 (Bray et
288 al. 2018). Because of the growth and aging of the population, prostate cancer has become a main
289 public health problem in men (Center et al. 2012). This tumor is often silent in clinical practice
290 and usually is found when it invades other tissue (Guo et al. 2019; Roobol & Carlsson 2013;
291 Shen & Abate-Shen 2010). Until now, there are only two articles studying methyltransferase in
292 prostate cancer, both are about METTL3. They suggested that METTL3 is tumor promotor in
293 prostate cancer.

294 The expression level of METTL3 protein and mRNA in prostate cancer are significantly higher
295 than those in adjacent benign tissues. METTL3 is mainly localized in the nucleus of the prostate
296 cells, but distribute in a small amount in the cytoplasm. It has been reported that METTL3
297 mRNA and protein level are positively correlated with prostate specific antigen (PSA) value and
298 Gleason score. Therefore, METTL3 plays a carcinogenic role in prostate cancer and may be used
299 in combination with PSA as a diagnostic marker for prostate cancer (Xianyong et al. 2019).
300 Evidence from cell line (C4-2, C4-2B, LNCaP, PC3 and DU-145) revealed that knocking down
301 METTL3 reduces m6A content and inhibit survival, cell proliferation, colony formation and
302 invasion. Mechanistic analysis indicated that the decreased expression level of GLI1 after
303 METTL3 depletion. GLI1 is an important component of SHH-GLI signaling which is positively
304 correlated with the PC severity. GLI1 has been shown to act as a negative modulator for
305 androgen receptor and contribute to the androgen-independent growth of PC. The mRNA levels
306 of c-Myc and Cyclin D1, which are SHH signaling downstream targets, are also inhibited, and
307 finally result in cell apoptosis (Cai et al. 2019) (Fig.3).

308

309 **Conclusions**

310 In general, from the limited studies, we found that METTL3 and METTL14 show different
311 expression in three types of urological cancers. METTL3 is high expression in the bladder and
312 prostate cancers, playing the positive regulation for cancer cells. But for kidney cancer,
313 expression of METTL3 is low. METTL14 is always low expression in the kidney cancer and
314 bladder cancer, playing negative regulation in cancer cells. But high expression of METTL14 is
315 not found in the urological cancers. No matter which type of those three cancers, when METTL3
316 or METTL14 is low expression, they always negatively regulate these pathways that promote
317 cell growth, such as mTOR, EMT, P2XR6, but positively regulate cell death-related pathways or
318 tumor suppressors, such as P53, PTEN and Notch1. When METTL3 is high expression, it
319 positively regulates NF- κ B/SHH-GL1 pathways (two cell proliferation-related pathways) and
320 negatively regulates PTEN (see table). Compared to METTL14, METTL3 seemly shows
321 various expression pattern and different regulation pathways in urological cancers, suggesting
322 that METTL3 has organ-specific characteristics in expression and mechanism regulation. It
323 provides a research theoretical basis for using m6A regulator as a new therapeutic target.
324 However, because of limited number of available studies, we are not able to fully elucidate the
325 molecular mechanism of m6A in urological tumors. It calls for more studies to reveal the
326 mechanism and clinical application potential of m6A regulator in urological tumors. However,

327 according to existing research results, METTL3 and METTL14 control cancer cell fate by cell
328 growth- and cell death-related pathways.

329 In this review, the potential molecular pathway networks are described through organizing
330 the existing researches. Although m6A has just emerged in the field of urological tumors, it has
331 already shown researchers a promising way. As some researchers have reported, m6A regulator
332 may become new targets for treatment and act as a biomarker for diagnosis or prognosis of
333 urological cancers in the future.

334

335 **Acknowledgements**

336 Thanks to Professor Han Fang, who serves for PTSD Laboratory of Department of Histology and
337 Embryology in the Basic Medical College of China Medical University, for her strong guidance
338 and support for this review, she has put forward valuable suggestions for the improvement of this
339 paper.

340

341 **References**

- 342 Alarcon CR, Goodarzi H, Lee H, Liu X, Tavazoie S, and Tavazoie SF. 2015a. HNRNPA2B1 Is a
343 Mediator of m(6)A-Dependent Nuclear RNA Processing Events. *Cell* 162:1299-1308.
344 10.1016/j.cell.2015.08.011
- 345 Alarcon CR, Lee H, Goodarzi H, Halberg N, and Tavazoie SF. 2015b. N6-methyladenosine
346 marks primary microRNAs for processing. *Nature* 519:482-485. 10.1038/nature14281
- 347 Boccaletto P, Machnicka MA, Purta E, Piatkowski P, Baginski B, Wirecki TK, de Crecy-Lagard
348 V, Ross R, Limbach PA, Kotter A, Helm M, and Bujnicki JM. 2018. MODOMICS: a
349 database of RNA modification pathways. 2017 update. *Nucleic Acids Res* 46:D303-
350 d307. 10.1093/nar/gkx1030
- 351 Bokar JA, Shambaugh ME, Polayes D, Matera AG, and Rottman FM. 1997. Purification and
352 cDNA cloning of the AdoMet-binding subunit of the human mRNA (N6-adenosine)-
353 methyltransferase. *Rna* 3:1233-1247.
- 354 Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, and Jemal A. 2018. Global cancer
355 statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36
356 cancers in 185 countries. *CA Cancer J Clin* 68:394-424. 10.3322/caac.21492
- 357 Cai J, Yang F, Zhan H, Situ J, Li W, Mao Y, and Luo Y. 2019. RNA m(6)A Methyltransferase
358 METTL3 Promotes The Growth Of Prostate Cancer By Regulating Hedgehog Pathway.
359 *Onco Targets Ther* 12:9143-9152. 10.2147/ott.S226796

- 360 Center MM, Jemal A, Lortet-Tieulent J, Ward E, Ferlay J, Brawley O, and Bray F. 2012.
361 International variation in prostate cancer incidence and mortality rates. *Eur Urol* 61:1079-
362 1092. 10.1016/j.eururo.2012.02.054
- 363 Chadet S, Jelassi B, Wannous R, Angoulvant D, Chevalier S, Besson P, and Roger S. 2014.
364 The activation of P2Y2 receptors increases MCF-7 breast cancer cells migration through
365 the MEK-ERK1/2 signalling pathway. *Carcinogenesis* 35:1238-1247.
366 10.1093/carcin/bgt493
- 367 Chen D, Gassenmaier M, Maruschke M, Riesenberger R, Pohla H, Stief CG, Zimmermann W,
368 and Buchner A. 2014. Expression and prognostic significance of a comprehensive
369 epithelial-mesenchymal transition gene set in renal cell carcinoma. *J Urol* 191:479-486.
370 10.1016/j.juro.2013.08.052
- 371 Chen M, Nie ZY, Wen XH, Gao YH, Cao H, and Zhang SF. 2019a. m6A RNA methylation
372 regulators can contribute to malignant progression and impact the prognosis of bladder
373 cancer. *Biosci Rep* 39. 10.1042/bsr20192892
- 374 Chen M, Wei L, Law CT, Tsang FH, Shen J, Cheng CL, Tsang LH, Ho DW, Chiu DK, Lee JM,
375 Wong CC, Ng IO, and Wong CM. 2018. RNA N6-methyladenosine methyltransferase-
376 like 3 promotes liver cancer progression through YTHDF2-dependent posttranscriptional
377 silencing of SOCS2. *Hepatology* 67:2254-2270. 10.1002/hep.29683
- 378 Chen XY, Zhang J, and Zhu JS. 2019b. The role of m(6)A RNA methylation in human cancer.
379 *Mol Cancer* 18:103. 10.1186/s12943-019-1033-z
- 380 Cheng M, Sheng L, Gao Q, Xiong Q, Zhang H, Wu M, Liang Y, Zhu F, Zhang Y, Zhang X, Yuan
381 Q, and Li Y. 2019. The m(6)A methyltransferase METTL3 promotes bladder cancer
382 progression via AFF4/NF-kappaB/MYC signaling network. *Oncogene* 38:3667-3680.
383 10.1038/s41388-019-0683-z
- 384 Choe J, Lin S, Zhang W, Liu Q, Wang L, Ramirez-Moya J, Du P, Kim W, Tang S, Sliz P,
385 Santisteban P, George RE, Richards WG, Wong KK, Locker N, Slack FJ, and Gregory
386 RI. 2018. mRNA circularization by METTL3-eIF3h enhances translation and promotes
387 oncogenesis. *Nature* 561:556-560. 10.1038/s41586-018-0538-8
- 388 Cui Q, Shi H, Ye P, Li L, Qu Q, Sun G, Sun G, Lu Z, Huang Y, Yang CG, Riggs AD, He C, and
389 Shi Y. 2017. m(6)A RNA Methylation Regulates the Self-Renewal and Tumorigenesis of
390 Glioblastoma Stem Cells. *Cell Rep* 18:2622-2634. 10.1016/j.celrep.2017.02.059
- 391 Deng X, Su R, Weng H, Huang H, Li Z, and Chen J. 2018. RNA N(6)-methyladenosine
392 modification in cancers: current status and perspectives. *Cell Res* 28:507-517.
393 10.1038/s41422-018-0034-6
- 394 Dominissini D, Moshitch-Moshkovitz S, Schwartz S, Salmon-Divon M, Ungar L, Osenberg S,
395 Cesarkas K, Jacob-Hirsch J, Amariglio N, Kupiec M, Sorek R, and Rechavi G. 2012.
396 Topology of the human and mouse m6A RNA methylomes revealed by m6A-seq. *Nature*
397 485:201-206. 10.1038/nature11112
- 398 Fuchs BC, Fujii T, Dorfman JD, Goodwin JM, Zhu AX, Lanuti M, and Tanabe KK. 2008.
399 Epithelial-to-mesenchymal transition and integrin-linked kinase mediate sensitivity to

- 400 epidermal growth factor receptor inhibition in human hepatoma cells. *Cancer Res*
401 68:2391-2399. 10.1158/0008-5472.Can-07-2460
- 402 Gong D, Zhang J, Chen Y, Xu Y, Ma J, Hu G, Huang Y, Zheng J, Zhai W, and Xue W. 2019.
403 The m(6)A-suppressed P2RX6 activation promotes renal cancer cells migration and
404 invasion through ATP-induced Ca(2+) influx modulating ERK1/2 phosphorylation and
405 MMP9 signaling pathway. *J Exp Clin Cancer Res* 38:233. 10.1186/s13046-019-1223-y
- 406 Gu C, Wang Z, Zhou N, Li G, Kou Y, Luo Y, Wang Y, Yang J, and Tian F. 2019. Mettl14 inhibits
407 bladder TIC self-renewal and bladder tumorigenesis through N(6)-methyladenosine of
408 Notch1. *Mol Cancer* 18:168. 10.1186/s12943-019-1084-1
- 409 Guo L, Lin M, Cheng Z, Chen Y, Huang Y, and Xu K. 2019. Identification of key genes and
410 multiple molecular pathways of metastatic process in prostate cancer. *PeerJ* 7:e7899.
411 10.7717/peerj.7899
- 412 Han J, Wang JZ, Yang X, Yu H, Zhou R, Lu HC, Yuan WB, Lu JC, Zhou ZJ, Lu Q, Wei JF, and
413 Yang H. 2019. METTL3 promote tumor proliferation of bladder cancer by accelerating
414 pri-miR221/222 maturation in m6A-dependent manner. *Mol Cancer* 18:110.
415 10.1186/s12943-019-1036-9
- 416 Hao H, Wang Z, Ren S, Shen H, Xian H, Ge W, and Wang W. 2019. Reduced GRAMD1C
417 expression correlates to poor prognosis and immune infiltrates in kidney renal clear cell
418 carcinoma. *PeerJ* 7:e8205. 10.7717/peerj.8205
- 419 Jin H, Ying X, Que B, Wang X, Chao Y, Zhang H, Yuan Z, Qi D, Lin S, Min W, Yang M, and Ji
420 W. 2019. N(6)-methyladenosine modification of ITGA6 mRNA promotes the
421 development and progression of bladder cancer. *EBioMedicine* 47:195-207.
422 10.1016/j.ebiom.2019.07.068
- 423 Ke S, Pandya-Jones A, Saito Y, Fak JJ, Vagbo CB, Geula S, Hanna JH, Black DL, Darnell JE,
424 Jr., and Darnell RB. 2017. m(6)A mRNA modifications are deposited in nascent pre-
425 mRNA and are not required for splicing but do specify cytoplasmic turnover. *Genes Dev*
426 31:990-1006. 10.1101/gad.301036.117
- 427 Leach RA, and Tuck MT. 2001. Expression of the mRNA (N6-adenosine)-methyltransferase S-
428 adenosyl-L-methionine binding subunit mRNA in cultured cells. *Int J Biochem Cell Biol*
429 33:984-999. 10.1016/s1357-2725(01)00071-1
- 430 Li X, Tang J, Huang W, Wang F, Li P, Qin C, Qin Z, Zou Q, Wei J, Hua L, Yang H, and Wang Z.
431 2017a. The M6A methyltransferase METTL3: acting as a tumor suppressor in renal cell
432 carcinoma. *Oncotarget* 8:96103-96116. 10.18632/oncotarget.21726
- 433 Li Z, Weng H, Su R, Weng X, Zuo Z, Li C, Huang H, Nachtergaele S, Dong L, Hu C, Qin X,
434 Tang L, Wang Y, Hong GM, Huang H, Wang X, Chen P, Gurbuxani S, Arnovitz S, Li Y,
435 Li S, Strong J, Neilly MB, Larson RA, Jiang X, Zhang P, Jin J, He C, and Chen J. 2017b.
436 FTO Plays an Oncogenic Role in Acute Myeloid Leukemia as a N(6)-Methyladenosine
437 RNA Demethylase. *Cancer Cell* 31:127-141. 10.1016/j.ccell.2016.11.017
- 438 Lim SH, Becker TM, Chua W, Ng WL, de Souza P, and Spring KJ. 2014. Circulating tumour
439 cells and the epithelial mesenchymal transition in colorectal cancer. *J Clin Pathol*
440 67:848-853. 10.1136/jclinpath-2014-202499

- 441 Lin S, Choe J, Du P, Triboulet R, and Gregory RI. 2016. The m(6)A Methyltransferase METTL3
442 Promotes Translation in Human Cancer Cells. *Mol Cell* 62:335-345.
443 10.1016/j.molcel.2016.03.021
- 444 Liu J, Eckert MA, Harada BT, Liu SM, Lu Z, Yu K, Tienda SM, Chryplewicz A, Zhu AC, Yang Y,
445 Huang JT, Chen SM, Xu ZG, Leng XH, Yu XC, Cao J, Zhang Z, Liu J, Lengyel E, and
446 He C. 2018. m(6)A mRNA methylation regulates AKT activity to promote the proliferation
447 and tumorigenicity of endometrial cancer. *Nat Cell Biol* 20:1074-1083. 10.1038/s41556-
448 018-0174-4
- 449 Liu J, Yue Y, Han D, Wang X, Fu Y, Zhang L, Jia G, Yu M, Lu Z, Deng X, Dai Q, Chen W, and
450 He C. 2014. A METTL3-METTL14 complex mediates mammalian nuclear RNA N6-
451 adenosine methylation. *Nat Chem Biol* 10:93-95. 10.1038/nchembio.1432
- 452 Liu N, Dai Q, Zheng G, He C, Parisien M, and Pan T. 2015. N(6)-methyladenosine-dependent
453 RNA structural switches regulate RNA-protein interactions. *Nature* 518:560-564.
454 10.1038/nature14234
- 455 Ma JZ, Yang F, Zhou CC, Liu F, Yuan JH, Wang F, Wang TT, Xu QG, Zhou WP, and Sun SH.
456 2017. METTL14 suppresses the metastatic potential of hepatocellular carcinoma by
457 modulating N(6) -methyladenosine-dependent primary MicroRNA processing.
458 *Hepatology* 65:529-543. 10.1002/hep.28885
- 459 McConkey DJ, Choi W, Marquis L, Martin F, Williams MB, Shah J, Svatek R, Das A, Adam L,
460 Kamat A, Siefker-Radtke A, and Dinney C. 2009. Role of epithelial-to-mesenchymal
461 transition (EMT) in drug sensitivity and metastasis in bladder cancer. *Cancer Metastasis*
462 *Rev* 28:335-344. 10.1007/s10555-009-9194-7
- 463 Meyer KD, Patil DP, Zhou J, Zinoviev A, Skabkin MA, Elemento O, Pestova TV, Qian SB, and
464 Jaffrey SR. 2015. 5' UTR m(6)A Promotes Cap-Independent Translation. *Cell* 163:999-
465 1010. 10.1016/j.cell.2015.10.012
- 466 Moch H, Cubilla AL, Humphrey PA, Reuter VE, and Ulbright TM. 2016. The 2016 WHO
467 Classification of Tumours of the Urinary System and Male Genital Organs-Part A: Renal,
468 Penile, and Testicular Tumours. *Eur Urol* 70:93-105. 10.1016/j.eururo.2016.02.029
- 469 North RA. 2002. Molecular physiology of P2X receptors. *Physiol Rev* 82:1013-1067.
470 10.1152/physrev.00015.2002
- 471 O'Reilly KE, Rojo F, She QB, Solit D, Mills GB, Smith D, Lane H, Hofmann F, Hicklin DJ, Ludwig
472 DL, Baselga J, and Rosen N. 2006. mTOR inhibition induces upstream receptor tyrosine
473 kinase signaling and activates Akt. *Cancer Res* 66:1500-1508. 10.1158/0008-5472.Can-
474 05-2925
- 475 Ping XL, Sun BF, Wang L, Xiao W, Yang X, Wang WJ, Adhikari S, Shi Y, Lv Y, Chen YS, Zhao
476 X, Li A, Yang Y, Dahal U, Lou XM, Liu X, Huang J, Yuan WP, Zhu XF, Cheng T, Zhao
477 YL, Wang X, Rendtlew Danielsen JM, Liu F, and Yang YG. 2014. Mammalian WTAP is a
478 regulatory subunit of the RNA N6-methyladenosine methyltransferase. *Cell Res* 24:177-
479 189. 10.1038/cr.2014.3
- 480 Roobol MJ, and Carlsson SV. 2013. Risk stratification in prostate cancer screening. *Nat Rev*
481 *Urol* 10:38-48. 10.1038/nrurol.2012.225

- 482 Sanli O, Dobruch J, Knowles MA, Burger M, Alemozaffar M, Nielsen ME, and Lotan Y. 2017.
483 Bladder cancer. *Nat Rev Dis Primers* 3:17022. 10.1038/nrdp.2017.22
- 484 Scholler E, Weichmann F, Treiber T, Ringle S, Treiber N, Flatley A, Feederle R, Bruckmann A,
485 and Meister G. 2018. Interactions, localization, and phosphorylation of the m(6)A
486 generating METTL3-METTL14-WTAP complex. *Rna* 24:499-512.
487 10.1261/rna.064063.117
- 488 Shaw RJ, and Cantley LC. 2006. Ras, PI(3)K and mTOR signalling controls tumour cell growth.
489 *Nature* 441:424-430. 10.1038/nature04869
- 490 Shen MM, and Abate-Shen C. 2010. Molecular genetics of prostate cancer: new prospects for
491 old challenges. *Genes Dev* 24:1967-2000. 10.1101/gad.1965810
- 492 Sledz P, and Jinek M. 2016. Structural insights into the molecular mechanism of the m(6)A
493 writer complex. *Elife* 5. 10.7554/eLife.18434
- 494 Visvanathan A, Patil V, Arora A, Hegde AS, Arivazhagan A, Santosh V, and Somasundaram K.
495 2018. Essential role of METTL3-mediated m(6)A modification in glioma stem-like cells
496 maintenance and radioresistance. *Oncogene* 37:522-533. 10.1038/onc.2017.351
- 497 Vu LP, Cheng Y, and Kharas MG. 2019. The Biology of m(6)A RNA Methylation in Normal and
498 Malignant Hematopoiesis. *Cancer Discov* 9:25-33. 10.1158/2159-8290.Cd-18-0959
- 499 Vu LP, Pickering BF, Cheng Y, Zaccara S, Nguyen D, Minuesa G, Chou T, Chow A, Saletore Y,
500 MacKay M, Schulman J, Famulare C, Patel M, Klimek VM, Garrett-Bakelman FE,
501 Melnick A, Carroll M, Mason CE, Jaffrey SR, and Kharas MG. 2017. The N(6)-
502 methyladenosine (m(6)A)-forming enzyme METTL3 controls myeloid differentiation of
503 normal hematopoietic and leukemia cells. *Nat Med* 23:1369-1376. 10.1038/nm.4416
- 504 Wang P, Doxtader KA, and Nam Y. 2016a. Structural Basis for Cooperative Function of Mettl3
505 and Mettl14 Methyltransferases. *Mol Cell* 63:306-317. 10.1016/j.molcel.2016.05.041
- 506 Wang Q, Zhang H, Chen Q, Wan Z, Gao X, and Qian W. 2019. Identification of METTL14 in
507 Kidney Renal Clear Cell Carcinoma Using Bioinformatics Analysis. *Dis Markers*
508 2019:5648783. 10.1155/2019/5648783
- 509 Wang X, Feng J, Xue Y, Guan Z, Zhang D, Liu Z, Gong Z, Wang Q, Huang J, Tang C, Zou T,
510 and Yin P. 2016b. Structural basis of N(6)-adenosine methylation by the METTL3-
511 METTL14 complex. *Nature* 534:575-578. 10.1038/nature18298
- 512 Wang X, Huang J, Zou T, and Yin P. 2017. Human m(6)A writers: Two subunits, 2 roles. *RNA*
513 *Biol* 14:300-304. 10.1080/15476286.2017.1282025
- 514 Wang X, Lu Z, Gomez A, Hon GC, Yue Y, Han D, Fu Y, Parisien M, Dai Q, Jia G, Ren B, Pan T,
515 and He C. 2014. N6-methyladenosine-dependent regulation of messenger RNA stability.
516 *Nature* 505:117-120. 10.1038/nature12730
- 517 Weng H, Huang H, Wu H, Qin X, Zhao BS, Dong L, Shi H, Skibbe J, Shen C, Hu C, Sheng Y,
518 Wang Y, Wunderlich M, Zhang B, Dore LC, Su R, Deng X, Ferchen K, Li C, Sun M, Lu
519 Z, Jiang X, Marcucci G, Mulloy JC, Yang J, Qian Z, Wei M, He C, and Chen J. 2018.
520 METTL14 Inhibits Hematopoietic Stem/Progenitor Differentiation and Promotes
521 Leukemogenesis via mRNA m(6)A Modification. *Cell Stem Cell* 22:191-205.e199.
522 10.1016/j.stem.2017.11.016

- 523 Wu G, Wang F, Li K, Li S, Zhao C, Fan C, and Wang J. 2019. Significance of TP53 mutation in
524 bladder cancer disease progression and drug selection. *PeerJ* 7:e8261.
525 10.7717/peerj.8261
- 526 Xiang Y, Laurent B, Hsu CH, Nachtergaele S, Lu Z, Sheng W, Xu C, Chen H, Ouyang J, Wang
527 S, Ling D, Hsu PH, Zou L, Jambhekar A, He C, and Shi Y. 2017. RNA m(6)A methylation
528 regulates the ultraviolet-induced DNA damage response. *Nature* 543:573-576.
529 10.1038/nature21671
- 530 Xianyong L, Runyun G, Kun-bin K, Peng G, Tonghai L, Qingpeng C, Yin C, and Hui Z. 2019.
531 Expression and clinical significance of METTL3 in prostate cancer. *The Journal of*
532 *Practical Medicine* 35:75-79.
- 533 Yang F, Jin H, Que B, Chao Y, Zhang H, Ying X, Zhou Z, Yuan Z, Su J, Wu B, Zhang W, Qi D,
534 Chen D, Min W, Lin S, and Ji W. 2019. Dynamic m(6)A mRNA methylation reveals the
535 role of METTL3-m(6)A-CDCP1 signaling axis in chemical carcinogenesis. *Oncogene*
536 38:4755-4772. 10.1038/s41388-019-0755-0
- 537 Zhang C, Samanta D, Lu H, Bullen JW, Zhang H, Chen I, He X, and Semenza GL. 2016.
538 Hypoxia induces the breast cancer stem cell phenotype by HIF-dependent and ALKBH5-
539 mediated m(6)A-demethylation of NANOG mRNA. *Proc Natl Acad Sci U S A* 113:E2047-
540 2056. 10.1073/pnas.1602883113
- 541 Zhang S, Zhao BS, Zhou A, Lin K, Zheng S, Lu Z, Chen Y, Sulman EP, Xie K, Bogler O,
542 Majumder S, He C, and Huang S. 2017. m(6)A Demethylase ALKBH5 Maintains
543 Tumorigenicity of Glioblastoma Stem-like Cells by Sustaining FOXM1 Expression and
544 Cell Proliferation Program. *Cancer Cell* 31:591-606.e596. 10.1016/j.ccell.2017.02.013
- 545 Zhao S, Liu J, Nanga P, Liu Y, Cicek AE, Knoblauch N, He C, Stephens M, and He X. 2019.
546 Detailed modeling of positive selection improves detection of cancer driver genes. *Nat*
547 *Commun* 10:3399. 10.1038/s41467-019-11284-9
- 548 Zheng W, Dong X, Zhao Y, Wang S, Jiang H, Zhang M, Zheng X, and Gu M. 2019. Multiple
549 Functions and Mechanisms Underlying the Role of METTL3 in Human Cancers. *Front*
550 *Oncol* 9:1403. 10.3389/fonc.2019.01403
- 551 Zhou J, Wang J, Hong B, Ma K, Xie H, Li L, Zhang K, Zhou B, Cai L, and Gong K. 2019. Gene
552 signatures and prognostic values of m6A regulators in clear cell renal cell carcinoma - a
553 retrospective study using TCGA database. *Aging (Albany NY)* 11:1633-1647.
554 10.18632/aging.101856
- 555 Zhu F, Qian W, Zhang H, Liang Y, Wu M, Zhang Y, Zhang X, Gao Q, and Li Y. 2017. SOX2 Is a
556 Marker for Stem-like Tumor Cells in Bladder Cancer. *Stem Cell Reports* 9:429-437.
557 10.1016/j.stemcr.2017.07.004
558

Figure 1

METTL3 and METTL14 regulatory network in kidney cancer

The red arrow in the figure represents the promoting effect, and the yellow arrow represents the inhibiting effect. PI3K / AKT / mTOR, EMT and P2RX6 play a pro-cancer role in bladder cancer, while p53 and PTEN play a tumor-suppressive role. METTL3 and METTL14 play a role in suppressing kidney cancer by inhibiting or promoting some pathways, respectively. At the same time, they also accept regulation from upstream molecules or pathways.

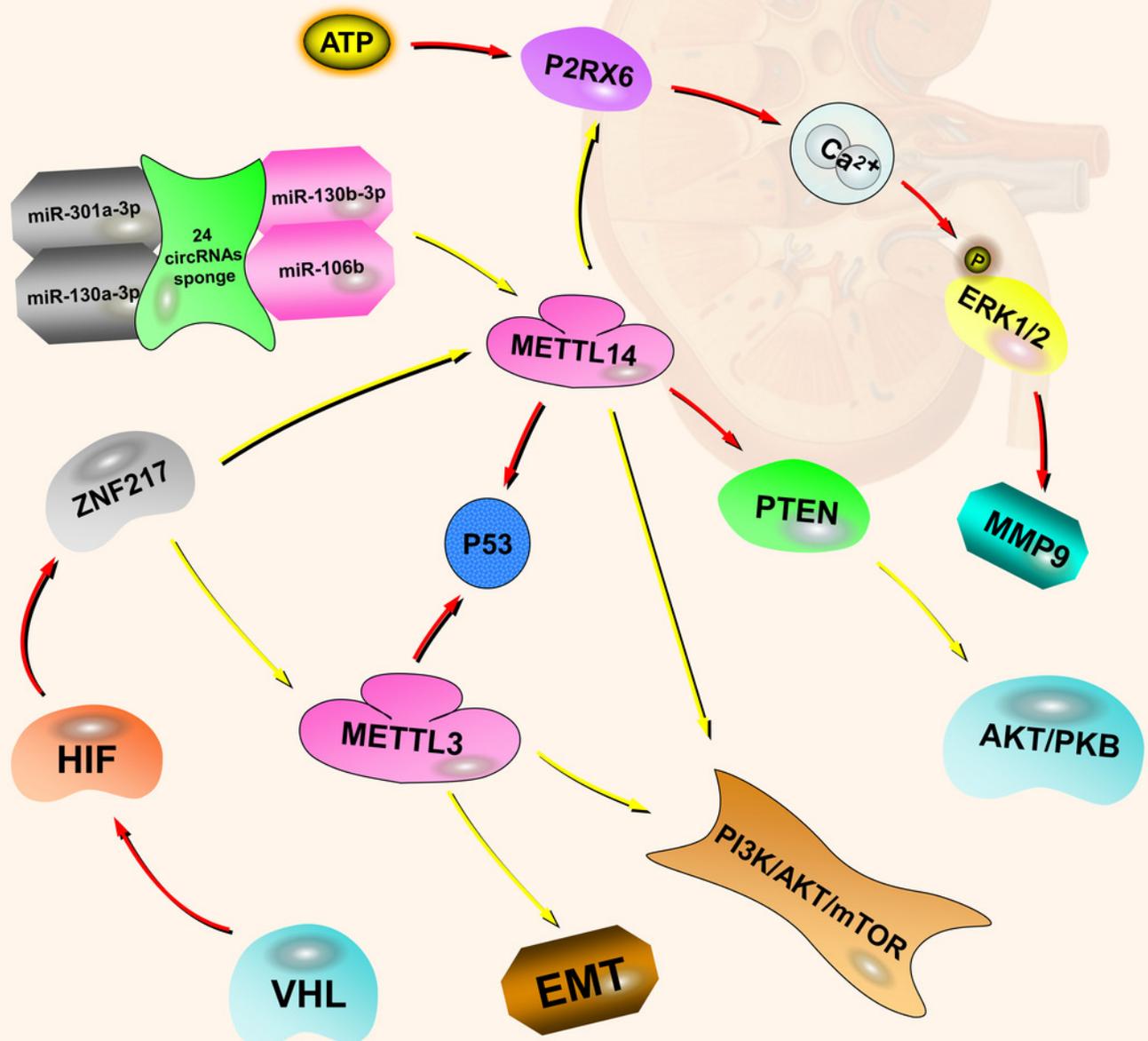
METTL3/14 regulatory network in kidney cancer.

Figure 2

METTL3 and METTL14 regulatory network in bladder cancer.

The red arrow in the figure represents the promoting effect, and the yellow arrow represents the inhibiting effect. AFF4 /NF- κ B/ MYC, DGCR8-pri-miR221/222-PTEN, CDCP1, ITGA6 and Notch1 play carcinogenic role in bladder cancer. METTL3 can play a carcinogenic role in bladder cancer through a variety of pathways. METTL14 is the opposite, although there are not many related studies, it is certain that it plays a tumor suppressive role in bladder cancer.

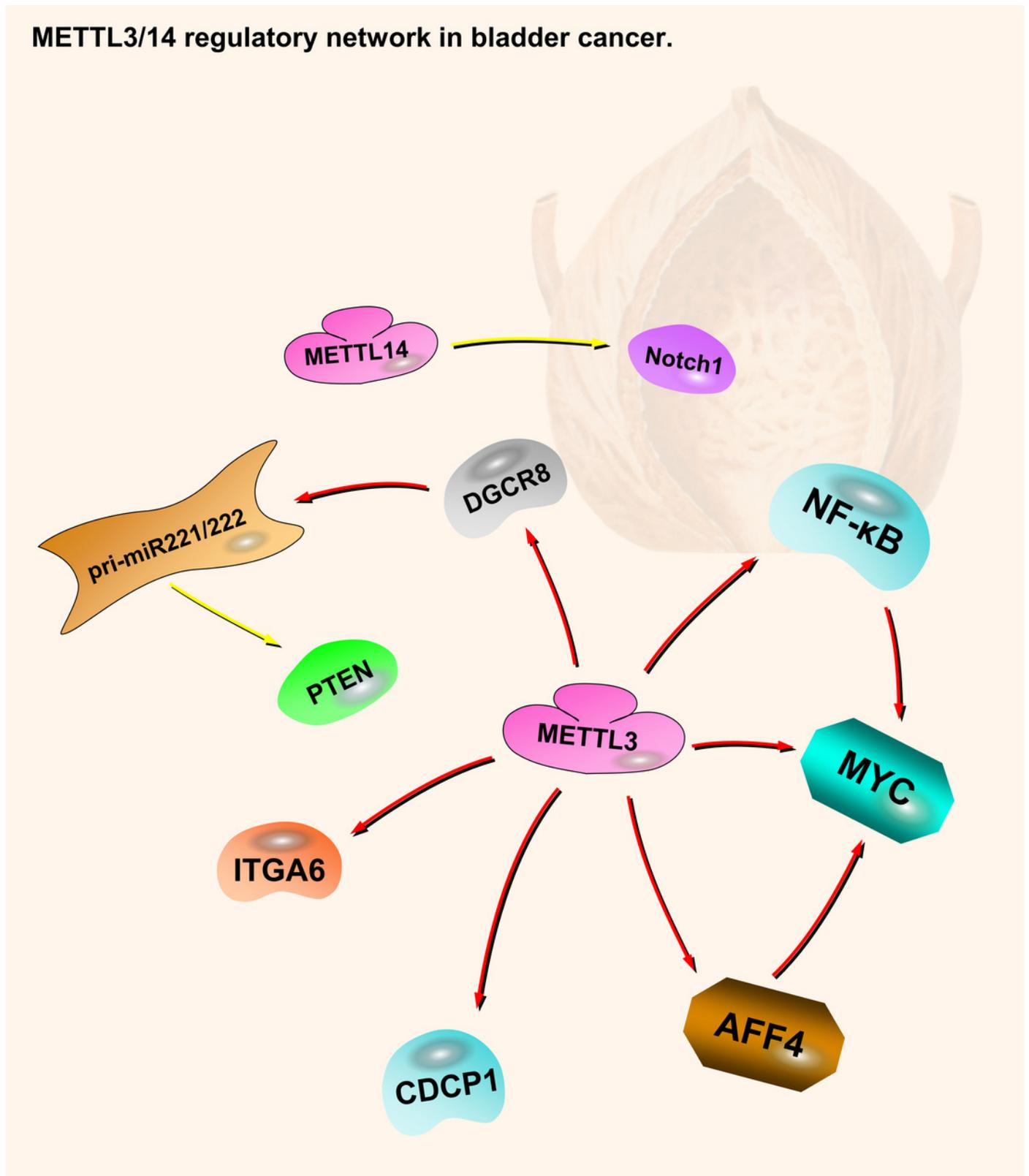
METTL3/14 regulatory network in bladder cancer.

Figure 3

METTL3 regulatory network in prostate cancer.

The red arrow in the figure represents the promoting effect, and the yellow arrow represents the inhibiting effect. There are not many researches on METTL3 in prostate cancer. It is certain that it plays a role in promoting cancer by affecting downstream pathway SHH-GLI1.

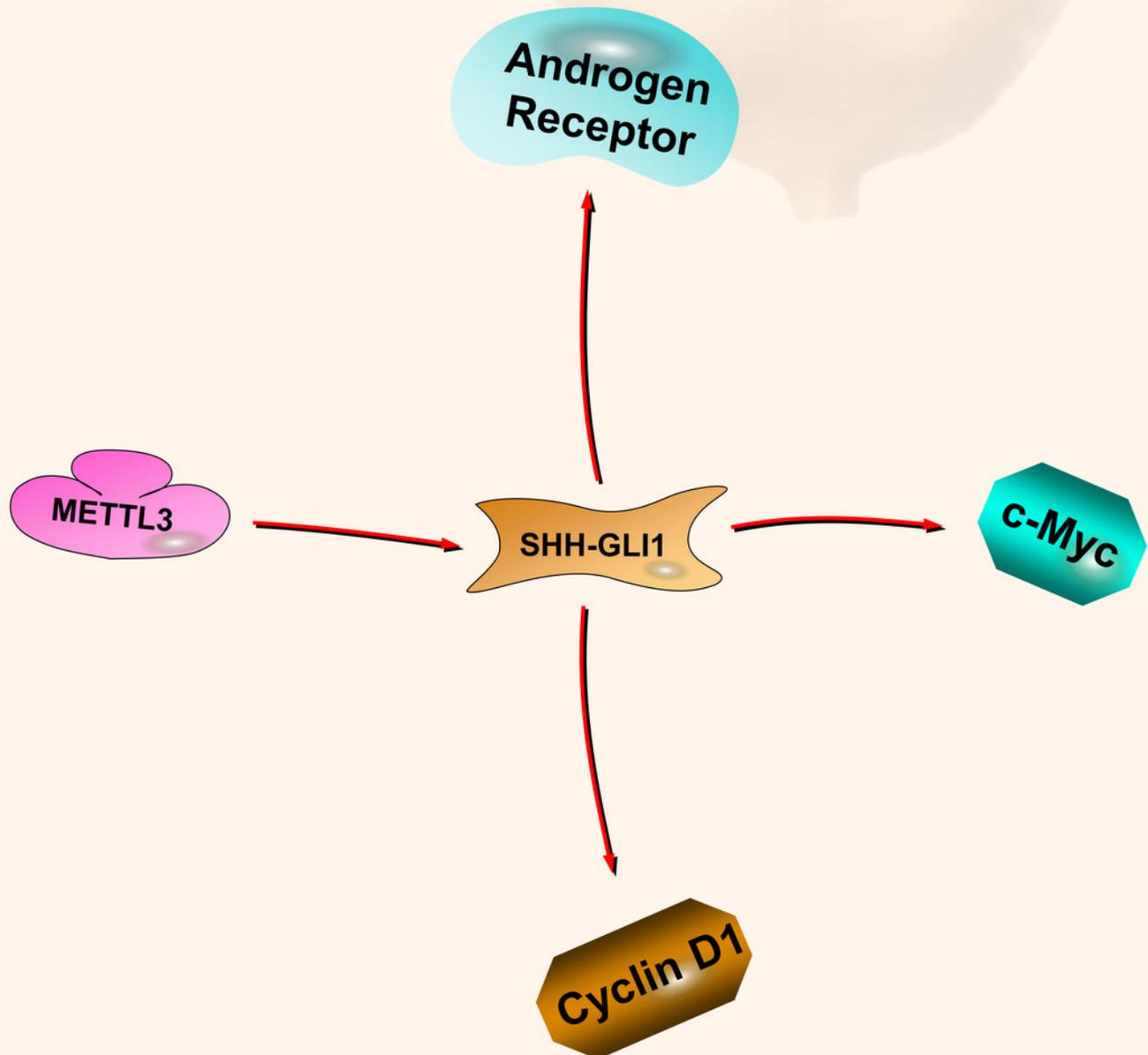
METTL3 regulatory network in prostate cancer.

Table 1 (on next page)

Multiple functions exerted by Methyltransferase in kidney, bladder and prostate cancers

1

Table 1: Multiple functions exerted by Methyltransferase in kidney, bladder and prostate cancers

Diseases	component	Role	Source of experimental evidence	Regulation	Potential Signal pathway	Author & Refs
	METTL3	Anti-oncogene	RCC and matched histologically-normal renal tissue are from 145 RCC patients; RCC cell lines (CAKI-1, CAKI-2 and ACHN); a normal renal tubular epithelial cell line (HK-2); BALB/c nude mice	Down-regulation	METTL3-PI3K/AKT/mTOR METTL3-EMT	Li et al. 2017
	METTL3	Anti-oncogene	528 ccRCC patients with CNV data and pathology reports from the TCGA database; GSEA database	Up-regulation	VHL-HIF-ZNF217-METTL3-PI3K/AKT/mTOR VHL-HIF-ZNF217-METTL3-p53	Zhou et al. 2019
Kidney cancer	METTL14	Anti-oncogene	528 ccRCC patients with CNV data and pathology reports from the TCGA database; GSEA database	Up-regulation	VHL-HIF-ZNF217-METTL14-PI3K/AKT/mTOR VHL-HIF-ZNF217-METTL14-p53	Zhou et al. 2019
	METTL14	Anti-oncogene	Online databases (TCGAportal,GTExPortal,UALCAN,GEPIA2,MEXPRESS,RMBasev2.0,OncoLnc,starBase,circBank,STRING...)	Down-regulation	circRNAs-miRNAs-METTL14-PTEN-AKT/PKB	Wang et al. 2019
	METTL14	Anti-oncogene	17 groups of renal cell carcinoma tissues and adjacent tissues received in patients with partial or complete kidney resection; Renal cancer cell line(OS-RC-2,786-O,HEK-293, SN12-PM6, SW839, A498); Human cortical proximal tubule epithelial cell line (HK-2); Nude mice; Online databases (TCGA, UALCAN...)	Down-regulation	METTL14-P2RX6-Ca ²⁺ -p-ERK1/2-MMP9	Gong et al. 2019
	METTL3	Oncogene	Human/mouse bladder cancer samples; bladder cancer cell lines (5637, UM-UC-3); Immortalized epithelial cells (SV-HUC-1); GSEA database	Up-regulation	METTL3-AFF4 /NF-κB/ MYC	Cheng et al. 2019
	METTL3	Oncogene	Human/mouse bladder cancer samples; bladder cancer cell lines (EJ, T24)	Up-regulation	METTL3-DGCR8-pri-miR221/222-PTEN	Han et al. 2019
Bladder cancer	METTL3	Oncogene	Formalin-fixed paraffin-embedded (FFPE) tissue from 114 cases of bladder cancer and 30 cases of cystitis with radical cystectomy and bladder biopsy; Human prostate	Up-regulation	METTL3 -CDCP1	Yang et al. 2019

epithelial cell line(RWPE-1); Human bladder cancer cell line(T24, UM-UC-3); urethral epithelial cells (SV-HUC-1); 3-methylcholesterol transformed urethral epithelial cells(MC-SV-HUC T2);NOD / SCID mice

	METTL3	Oncogene	TCGA GDAC Firehose; bladder cancer cell lines(T24)	Up-regulation	METTL3- ITGA6	Jin et al. 2019
	METTL14	Anti-oncogene	Primary bladder cancer specimens from 6 bladder cancer patients	Down-regulation	METTL14-Notch1	Gu et al. 2019
Prostate cancer	METTL3	Oncogene	Human prostate cancer cell lines(LNCaP, PC3, C4-2, C4-2B, DU-145); Human normal prostate epithelial cell line(RWPE-1); Six-week-old male NOD/SCID mice	Up-regulation	METTL3-SHH/GLI1-C-Myc/cyclin D1	Cai et al. 2019

2

3

4