

Genotypic characterization and genome comparison reveals insights into potential vaccine coverage and genealogy of *Neisseria meningitidis* in military camps in Vietnam

~~Genetic diversity and features of *Neisseria meningitidis* in military camps in Vietnam revealed by multi-locus sequence typing and genome comparison~~

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## 24 ABSTRACT

25 **Background.** *Neisseria meningitidis* remains the main cause of sporadic meningitis and sepsis in  
26 military camps in Vietnam. Yet, very limited molecular data of their genotypic and  
27 epidemiological characteristics are available from Vietnam, and particularly the military  
28 environment. Whole genome sequencing (WGS) has proven useful for meningococcal disease  
29 surveillance and guiding preventative vaccination programs. Previously, we characterized key  
30 genetic and epidemiological features of an invasive *N. meningitidis* B isolate from a military unit  
31 in Vietnam. Here, we extend these findings by sequencing two [additional invasive \*N.\*](#)  
32 [meningitidis B isolates isolated from cerebrospinal fluid \(CSF\) of two meningitis cases at](#) another  
33 military unit and compared their genomic sequences and features. We also report the sequence  
34 types and antigenic profiles of 25 historical and more recently emerged *N. meningitidis* isolates  
35 from these units and other units in proximity.

36 **Methods.** Strains were sequenced using the Illumina HiSeq platform, *de novo* assembled and  
37 annotated. Genomes were compared within and between military units, as well as against the  
38 global *N. meningitidis* collection and other isolates from the Southeast Asia region [using](#)  
39 [PubMLST](#). Variations at the nucleotide level were determined, and phylogenetic relationships  
40 [were](#) estimated. [Antigenic genotypes and vaccine coverage were analyzed using gMATS and](#)  
41 [PubMLST](#). Susceptibility of isolates against commonly used antibiotic agents was examined  
42 [using E-test](#).

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**Results.** Genome comparison revealed a high level of similarity among isolates both within and between units. All isolates showed resistance to chloramphenicol and carried identical *catP* gene with other Southeast Asian isolates, suggesting a common lineage. Their antigenic [profiles](#) [genotypes](#) predicted no coverage by either Bexsero® or Trumenba®, and nucleotide variation analysis revealed diverse new, unassigned alleles at multiple virulence loci of all strains. Groups of singleton and unique novel sequence types extending beyond individual camps were found from epidemiological data of 25 other isolates. Our results add to the sparse published molecular data of *N. meningitidis* in the military units in Vietnam, highlight their diversity, distinct genetic features and antibiotic resistance pattern, and emphasize the need for further studies on the molecular characteristics of *N. meningitidis* in Vietnam.

## INTRODUCTION

*Neisseria meningitidis* is an encapsulated Gram-negative bacterium that asymptotically colonizes the human nasopharynx but can cause serious septicemia and meningitis upon entering the blood stream and passing through the blood-meningeal barrier (Rosenstein et al. 2001). Carriage rate is age and setting dependent, among other factors, with high prevalence found in the age groups of infants (4.5%) and young adults (23.7%) (Christensen et al. 2010). Congregated living environment is another risk factor, as shown in increased carriage rate among university students living in dormitories (Peterson et al. 2018; Breakwell et al. 2018) or military recruits in camp sites (Sim et al. 2013; Keiser, Hamilton, and Broderick 2011). In these environments, meningococcal meningitis can sometimes become an outbreak, and sporadic cases often show recurring and cluster characteristics (Peterson et al. 2018). Hence, in many countries, preventative vaccination program is recommended for these high-risk groups (Yezli, Wilder-

66 Smith, and Bin Saeed 2016) and implementations have shown significant impact (Broderick,  
67 Phillips, and Faix 2015). However, to enable successful preventative implementation and reduce  
68 the risk related to these environments, as well as to aid future cases' diagnosis and treatment, it is  
69 crucial to have reliable and accurate monitoring data of sporadic cases and carriages in these  
70 groups.

71         Recent advances in genome sequencing technology allows for a significant volume of  
72 genomic data to be generated and made public. Such data also provide an unprecedented power  
73 of discrimination that is invaluable for studies of the relationship of closely related strains. The  
74 widely utilized *Neisseria* database PubMLST (<https://pubmlst.org/neisseria/>), at the time of  
75 preparation of this manuscript, hosts more than 22,000 genomes and 60,000 isolate records for  
76 *Neisseria*. Employing sequence data deposited to PubMLST, a core genome of 1605 loci was  
77 determined for *N. meningitidis* (Bratcher et al. 2014). Analysis of sequence variations at these  
78 loci has furthered our understanding of genomic variation within *N. meningitidis* population  
79 (Harrison et al. 2017) and enabled the study of closely related but distinct strains present in  
80 outbreaks (Jolley et al. 2012). Despite the usefulness of [whole genome sequencing \(WGS\)](#)  
81 analysis for epidemiological surveillance, data from Vietnam, and Southeast Asian region in  
82 general, are extremely limited. Previously, we have described for the first time the genome of a  
83 chloramphenicol-resistant invasive *N. meningitidis* B isolated from a military unit in Vietnam  
84 (Tran et al. 2019). Later, a study conducted by the Mahidol-Oxford Tropical Health Network  
85 (MORU) in Thailand, Laos and Cambodia identified eight additional *N. meningitidis* isolates  
86 carrying the identical chloramphenicol-resistant gene along other acquired resistance to multiple  
87 antibiotics (Batty et al. 2019), suggesting the existence and expansion of a lineage in this region.  
88 Here we extend the previous studies by describing the genomic characteristics of two additional

invasive *N. meningitidis* B isolates isolated from CSF of two meningitis cases at another Vietnamese military unit in proximity with the previously reported, and conduct a global analysis utilizing the PubMLST database and associated analytical tools. We show that besides the chloramphenicol non-susceptible lineage, reservoirs of known sequence types that cannot be assigned to lineages, novel emerging sequence types make up a significant part of both invasive and carriage strains found in the military camp environments in Vietnam.

## MATERIALS AND METHODS

### Bacterial isolation and typing

All invasive *N. meningitidis* strains were isolated from cerebrospinal fluid (CSF) and all carrier strains were isolated from mouthwash samples at the Laboratory of Microbiology, Military Institute of Preventive Medicine, Hanoi as described previously (Tran et al. 2019). Two isolates, NMB\_VN2013 and NMB\_VN2015, were from the CSF of two confirmed meningitis cases discovered in 2013 and 2015 at Military Unit 2, a camp in the geographical closeness to Military Unit 1, the camp where the previously reported DuyDNT isolate were identified. Both cases were treated at the military hospital and subsequently recovered. Serogroup typing identification and multi-locus sequence typing (MLST) were either done according to previously described standard methods (Organization 2011) or by manually extracting the corresponding sequences from WGS data.

DuyDNT isolate was renamed NMB\_VN2014 here for convenience and consistency. Available laboratory records of the Laboratory of Microbiology, Military Institute of Preventive Medicine, Hanoi were reviewed and a suitable collection of 25 isolates were chosen based on reported year

111 (before and after 2013 – 2015), location (Military Unit 1, 2, and two nearby units here named  
112 Unit 3 and Unit 4) and availability of molecular characterization data (Table 41). Serogroup data,  
113 MLST data, *fHbp* and *porA* allelic variants were obtained from laboratory records. Case-related  
114 metadata, including ~~demographic data and year~~, clinical status, and location were obtained where  
115 available.

#### 116 **Antibiotic susceptibility testing**

117 Susceptibility of isolates to seven antibiotics, namely ampicillin, ciprofloxacin,  
118 cefotaxime, ceftriaxone, rifampicin, meropenem and chloramphenicol, was examined using E-test  
119 strip (bioMérieux, France) following manufacturer's guideline and MIC values were determined.  
120 Susceptibility was interpreted according to CLSI 2018 breakpoints (CLSI 2018).

#### 121 **Genome sequencing and analysis**

122 Genomic DNA was extracted using GeneJET Genomic DNA Purification Kit  
123 (ThermoFisher Scientific) in accordance with the manufacturer's instruction. Samples' quality  
124 was checked before sequenced using the Illumina HiSeq 4000 system (Macrogen). Genome  
125 assembly and annotation were performed as previously described (Tran et al. 2019). Annotated  
126 amino acid sequences were used to identify genes involved in antibiotic resistance (Tran et al.  
127 2019). Additionally, allelic profile of relevant antibiotic resistant genes from PubMLST were  
128 extracted from WGS data of each genome, and where applied, PSI-BLAST were used to find  
129 homologous sequence. Antigenic profile, antibody cross reactivity prediction and allelic variants  
130 of virulent factors including capsular genes, Maf-toxin island, and outer membrane vesicle  
131 (OMV) genes were analyzed with gMATS (Muzzi et al. 2019) and the ~~at~~ PubMLST server using  
132 default parameters.

### Genome comparison and phylogenetic analysis

Assembled genomes were submitted to [the](#) PubMLST website and allelic variants were automatically assigned for each locus. Genomes were compared at the seven loci of MLST scheme, the 53 loci of ribosomal MLST scheme, and the 1605 loci of the core genome cgMLST scheme for *N. meningitidis*. Allelic variants were further processed in Excel and where necessary, manually removed from comparisons. Genomes were aligned using the progressive Mauve software (Darling, Mau, and Perna 2010), and the phylogenetic distance between strains was determined at the whole genome level. Neighbor-Net networks were constructed from various comparisons implemented on the PubMLST and visualized by SplitsTree (Huson 1998).

Raw sequence data of 18 Southeast Asian *N. meningitidis* was obtained from the European Nucleotide Archive (project PRJEB30968) (Batty et al. 2019), assembled using Spades V3.11.1 (Bankevich et al. 2012), and assembled contigs were used for phylogenetic network analysis by SplitsTree and other sequence comparisons.

## RESULTS [AND DISCUSSION](#)

### Characterization of NMB\_VN2013 and NMB\_VN2015 isolates

#### *Genome*

Both NMB\_VN2013 and NMB\_Vn2015 had a genome size of ~ 2.1 Mb and ~ 51.2% GC content, and contained 2390 and 2409 CDS each, respectively. Each genome had 3 rRNA and 53 tRNA coding sequences and contained ~ 400 repetitive sequences. Overall, their genome size and content matched closely with the typical genome of [the previously reported](#) *Neisseria* [representatives](#) [genus](#) such as *N. meningitidis* MC58 (Tettelin et al. 2000), [Z2491](#) (Parkhill et al.

155 2000), [FAM18](#) (Bentley et al. 2007), and *N. gonorrhoeae* [NCCP11945](#) (Chung et al. 2008).  
156 Assembly data and genomic sequences of both genomes were deposited to NCBI Genomes  
157 database under BioProject ID PRJNA523495.

158 ***Serogroup and Sequence type***

159 Both NMB\_VN2013 and NMB\_VN2015 were serogroup B, as inferred by the presence  
160 of [the](#) *csb* gene from their genome sequence and confirmed by Vitek®. Multi-locus sequence  
161 typing (MLST) profiles extracted from WGS data grouped NMB\_VN2013 and NMB\_VN2015  
162 into the same ST-1576, a singleton ST that had no known clonal complex. ST-1576 is closely  
163 related to ST-13074, which was assigned to NMB\_VN2014 before, with two STs differed at a  
164 single locus (*aroE*). Polymorphic site analysis revealed 46 nucleotide changes and no  
165 deletion/insertion between the two alleles, [aroE](#) 9 of ST-1576 and [aroE](#) 4 of ST-13074.

166 ***Antibiotic susceptibility***

167 Previously, NMB\_VN2014 was shown to carry a tetracycline (*rpsJ*) and chloramphenicol  
168 (*catP*) resistant genes. Identical *rpsJ* and *catP* genes were found in the genomes of  
169 NMB\_VN2013 and NMB\_VN2015. The 624 bp *catP* gene found in all three Vietnamese isolates  
170 was the same gene previously reported in France ([Galimand et al. 1998](#)), and Southeast Asia  
171 ([Batty et al. 2019](#)). Antibiotic susceptibility test confirmed NMB\_VN2013 and NMB\_VN2015's  
172 resistance to chloramphenicol, with the recorded MIC were 62 and 64 µg/ml, respectively (Table  
173 [42](#)).

174 From WGS data, of the 11 antibiotic susceptibility genes analyzed by PubMLST, eight  
175 were identical in all isolates, including *gyrA* (allele 2), *pen A* (allele 587) and *rpoB* (allele 42).  
176 Both *gyrA* 2 and *rpoB* 42 alleles were previously shown to confer no resistance to ciprofloxacin

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177 and rifampicin, respectively (Hong et al. 2013; Taha et al. 2007). Loci NEIS1609 (*folP*) from all  
178 isolates, NEIS1600 (*parE*) from NMB\_VN2014, and NEIS1753 (*penA*) from NMB\_VN2013 and  
179 NMB\_VN2014 had new allelic variants with no assigned numbers. NEIS1600 and NEIS1753,  
180 together with NEIS1635 (*mtrR*), represented three variable loci among the isolates of this study  
181 (Table 42). Notably, both allelic variants at *mtrR* locus (7 in NMB\_VN2013 and NMB\_VN2015,  
182 and 1086 in NMB\_VN2014) harbored the A39T mutation. This mutation was observed  
183 significantly more often in azithromycin-exposed *N. gonorrhoea* carriers (Wind et al. 2017), and  
184 may result in overexpression of the MtrCDE efflux pump and increased antibiotic resistance in *N.*  
185 *gonorrhoea* (Demczuk et al. 2017). Although azithromycin is not recommended by the Vietnam  
186 Ministry of Health for treatment of meningitis, it is recommended by the WHO for the dual  
187 therapy (along with ceftriaxone) to treat *N. gonorrhoea* infection. It is thus important to monitor  
188 the presence and spreading to potential azithromycin-resistant genetic features in *Neisseria*  
189 genus. Other well-known azithromycin-resistant mutations, 23S rRNA A2045G and C2597T,  
190 first identified in *N. gonorrhoea*, (Demczuk et al. 2017), were not found in any Vietnamese  
191 isolates in this study which all carried wild-type 23S rRNA.

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192 We confirmed antibiotic susceptibility of NMB\_VN2013 and NMB\_VN2015 by MIC  
193 test, and both strains showed sensitivity to ciprofloxacin, rifampicin, cefotaxime, and ceftriaxone;  
194 but diminished susceptibility toward ampicillin, and resistance toward chloramphenicol, though  
195 the recorded MICs were much lower than that of NMB\_VN2014. While NMB\_VN2014 showed  
196 reduced susceptibility to rifampicin (MIC = 1.5 µg/ml), both NMB\_VN2013 and NMB\_VN2015  
197 were still susceptible (MIC = 0.25 and 0.125 µg/ml, respectively) (Table 42).

198 ***Antigenic profiles***

Analysis of the deduced peptide sequence at antigenic loci using PubMLST showed all three Vietnamese isolate's genomes expressed-carried *fHbp* Peptide 31, *NhbA-nhbA* Peptide 16, and no *NadA-nadA* peptide, but carried different *PorA-porA* and *FetA-fetA* variants (Table 23). In detail, NMB\_VN2014 and NMB\_VN2015 both had *PorA-porA* VR1 22-25, but different *PorA-porA* VR2, 14-32 and 14, respectively. NMB\_VN2013 carried distinct *PorA-porA* variants (VR1 7-2, VR2 13) but shared the same *FetA-fetA* variant (F4-6) with NMB\_VN2014. NMB-VN2015 carried *FetA-fetA* variant 1-7. *fHbp* Peptide 31 belonged to subfamily A, 34 amino acid substituted from Peptide 19 (Trumenba® vaccine 2014) and 97 amino acid differed from Peptide 1 (BEXSERO® vaccine 2015). *NhbA-nhbA* Peptide 16 contained 79 amino acid substitutions from the Bexsero® component *NhbA-nhbA* (Peptide 2). Allele 22-25 of VR1 region of *PorA-porA* had eight amino acid substitutions and one deletion compared to allele 7-2, and both were not the 1.4 variant used in Bexsero®. ~~Antibody cross-reactivity prediction showed no protection of either Trumenba® or Bexsero® against the three isolates of this study.~~

When compared with clinical profiles of the Southeast Asian isolates [reported recently by Batty and colleagues](#) (Batty et al. 2019)([Table 2](#)), a chloramphenicol-resistant lineage specific features could be observed in sequence types, [fHbp](#) Peptide, and [NhbA-nhbA](#) Peptide variants of the Vietnamese and the chloramphenicol-resistant Southeast Asian isolates ([Table 3](#)). On the other hand, [FetA-fetA](#) variants showed more variables among groups, and the Vietnamese [PorA](#) [porA](#) loci shared variants with the chloramphenicol-susceptible groups instead of the resistant group.

According to gMATS, a recently developed genotyping tool that predicts strain coverage of 4CMenB (Bexsero®) based on *fHbp*, *nhbA*, and *porA* VR2 specific genotypes (Muzzi et al. 2019), all three Vietnamese isolates were *fHbp* and *porA* non-coverage and *nhbA* unpredictable.

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**Commented [AW1]:** These should be protein names, ie NhbA as you had originally. I release that these have been deduced from the genome sequence, but when you are referring to peptides, these are peptides derived from the deduced protein sequence

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222 [Among the other 18 Southeast Asian isolates, three were covered by Bexsero® by all antigenic](#)  
223 [components \(NM03, NM14, and NM15\) while one \(NM23\) was covered by just \*nhbA\*.](#)  
224 [Altogether, this resulted in a coverage of 14.3%, 19%, and 14.3% at \*fHbp\*, \*nhbA\*, and \*porA\*](#)  
225 [among 21 isolates, respectively. Non-coverage was predicted at 71.4%, 19%, and 85.7% at \*fHbp\*,](#)  
226 [the \*nhbA\*, and \*porA\*, respectively. The rest of variants, including 14.3% of \*fHbp\*, 61.9% of \*nhbA\*, and](#)  
227 [none of \*porA\* were unpredictable by gMATs. Considering both expression level and genotyping,](#)  
228 [and extending vaccine coverage prediction to both Trumenba® and Bexsero®, PubMLST](#)  
229 [Antibody cross reactivity also prediction predicted showed no protection of either Trumenba®](#)  
230 [or Bexsero® vaccine against all the three Vietnamese isolates of this study. \(Muzzi et al.](#)  
231 [2019\) Notably, only three isolates \(NM03, NM14, and NM15\) could be predicted to be covered](#)  
232 [by Bexsero®.](#)

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#### 234 Inference of the Vietnamese isolates' genealogy

235 Neighbor-Net analysis was used to construct the phylogenetic networks of three  
236 Vietnamese isolates and the well-studied 108-isolate global *N. meningitidis* collection (Bratcher  
237 et al. 2014) based on the seven-locus (MLST), 53-locus (rMLST) and 1605-locus (cgMLST)  
238 comparisons. [This collection represents the major hyper-invasive lineages/sub-lineages recorded](#)  
239 [worldwide in the last 70 years.](#) All three methods grouped the Vietnamese isolates into a well-  
240 defined, separate clade from the rest of the network. While similar relationships for the  
241 Vietnamese isolates were maintained in all analyses, the 1605-locus cgMLST analysis was able  
242 to place the Vietnamese branch at the stem of the previously defined Lineage 3 (ST-41/44 clonal  
243 complex) (Figure 1). Additionally, only cgMLST analysis could separate the Vietnamese isolates  
244 into three unique strains; while MLST comparison grouped NMB\_VN2013 and NMB\_VN2015

245 into one ST, and rMLST identified NMB\_VN2013 and NMB\_VN2014 as a single strain;  
246 showing the close relationship among these strains.

247       —A more refined cluster was observed when the 18 Southeast Asian invasive *N.*  
248 *meningitidis* were added to the genealogical analysis. Eleven chloramphenicol-resistant isolates,  
249 including three Vietnamese isolates, four Thai isolates, and two each from Laos and Cambodia,  
250 formed a distinct group, diverged from the rest of Southeast Asian isolates. This lineage seemed  
251 to have rapidly expanded clonally in recent years, though this could partially be due to better case  
252 report and laboratory detection, since data from this region was scarce up until recently although  
253 still remains limited. Although NeighborNet analysis placed this lineage as a divergent branch  
254 from other chloramphenicol susceptible isolates in the region, due to the limited number of  
255 samples, it remains a possibility for their origin.

256       The rest of isolates also clustered into two groups, one included NM03, NM14, and  
257 NM15 that clustered to the ST-41 sub-lineage of ST-41/44 clonal complex, while the remaining  
258 isolates clustered together, seemingly formed a group connecting the chloramphenicol-resistant  
259 lineage with ST-44 sub-lineage of ST-41/44 clonal complex (Figure 1).

260

261 **Relationships among Vietnamese isolates revealed by genome comparisons**

262       Of the 1605 loci compared, Genome Comparator identified 1245 identical loci and 355  
263 variable loci in at least one genome of the three isolates in this study. Five loci were missing in  
264 all three isolates, four of those were pseudo-genes and one encoded for a phage-related protein.  
265 Fifteen loci were paralogous loci presumably resulting from assembly of repetitive sequences and  
266 thus were excluded from the final analysis. Resultant variable loci were further examined to

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267 exclude all variations from genes/pseudo\_genes encoding for hypothetical proteins.  
268 Consequently, from the initially identified 355 variable loci, we confirmed 264 loci that  
269 contained either point mutations, insertions/deletions, or allelic replacements from the three  
270 genomes. Among these, a number of sequence variables were observed in adjacent genes,  
271 suggesting frequent recombination events between genomes. Pairwise, the genomes of  
272 NMB\_VN2013 and NMB\_VN2014 showed the highest similarity, and NMB\_VN2014 and  
273 NMB\_VN2015 pair showed the lowest. However, pairwise phylogenetic distance between strains  
274 calculated from the progressiveMauve alignment using whole genome content was in the range of  
275 0.0158 to 0.0112, indicating a close relationship among them.

276 Besides core genome comparison, sequences of the known genetic determinants for the  
277 virulence of invasive *N. meningitidis*, namely the capsular gene cluster, the Maf-toxin island, and  
278 genes encoding for the outer membrane vesicle (OMV) peptides and pilin, from the genomes of  
279 three isolates were also compared. The resulting differences, after excluding all variables due to  
280 assembly or in the coding sequence of hypothetical proteins, are listed in Table 34. Overall,  
281 higher sequence variability could be observed among the three genomes at these loci. From 19  
282 loci of the capsular gene cluster being present for comparison, four confirmed variable loci were  
283 *rfbA*, *cssA*, *ctrF*, and *cnl*. Except for two assigned alleles found in two loci (allele 113, *cssA* locus  
284 of NMB\_VN2015's genome and allele 4, *ctrF* locus of NMB\_VN2013's genome), the rest of the  
285 alleles were newly identified and not yet assigned an allelic number. Four out of 45 loci of the  
286 Maf-toxin genomic island included in this comparison, namely *anmK*, *mafI*o2MGI-2,  
287 *mafA*\_MGI-3, and *mafB*\_MGI-3, appeared to be different. Sequences of the *anmK* locus of  
288 NMB\_VN2015 and the *mafB*\_MGI-3 locus of NMB\_VN2014 genome appeared to be novel,  
289 unassigned alleles. Detailed nucleotide changes analysis revealed a higher percentage of changed

290 nucleotides compared to any other single locus in both *mafA\_MGI-3* and *mafB\_MGI-3*, which  
291 were adjacent to each other, suggesting a recombination event could have happened in this  
292 region. Compared to the capsular gene cluster and Maf-toxin island, the CDS of OMV and pilin  
293 genes showed higher sequence diversity with variables scattered among loci.

294 [To better understand the probable evolutionary context of these strains, we have further](#)  
295 [collected and analyzed 25 isolates representing a collection of both historical and later emerged](#)  
296 [isolates from the same or nearby military units with the three in this study \(Table 1\).](#)  
297 Epidemiological data of [these 25 historical and later emerged isolates from the same or nearby](#)  
298 [military units with the three in this study](#) revealed the predominance of serogroup B *N.*  
299 *meningitidis* (78.5%) and the recurrence of two major lineages centered around ST-1576 and ST-  
300 4821 in all camp sites (Table 41). [The predominance of serogroup B agrees with the previous](#)  
301 [report from sporadic cases in the region \(Pancharoen et al. 2000\). The other serogroup observed](#)  
302 [was serogroup C \(21.5%\). Both ST-1576 and ST-4821 were major hyper-virulent STs with a long](#)  
303 [history and worldwide distribution. Antigenic profiles associated with these major STs show high](#)  
304 [similarity to strains described before in China during 1978 – 2013 \(Zhu et al. 2015\). Besides](#)  
305 [these, strains representing singleton and novel STs were also frequently observed throughout the](#)  
306 [years and locations. Available antigenic profiles suggested frequent exchange of genetic material](#)  
307 [via recombination among strains and reservoirs, with several alleles \(P1.20, P1.7-2\) associated](#)  
308 [with the major lineages frequently recurring in different combinations. Clonal expansion was also](#)  
309 [observed, indicated by the emergence of novel sequence types, and showed no distinct cluster in](#)  
310 [regard of geological locations, reflecting the close and frequent contact nature of the training](#)  
311 [units of new military recruits, suggesting close transmission extending beyond individual camps](#)  
312 [seemed to be the main driving force for \*N. meningitidis\* prevalence and expansion within the](#)

313 military environment in Vietnam. From searching through the PubMLST database, many  
314 emergent STs identified were limited to these units only, highlighting the niche characteristic of  
315 *N. meningitidis* population of the military camps.

316 *N. meningitidis* remains the main cause of sporadic meningitis and sepsis in military  
317 recruit camps (Tran et al. 2019; Sim et al. 2013; Keiser, Hamilton, and Broderick 2011).  
318 Accurate identification and characterization of the causative strain is crucial for the success of  
319 treatment for patients and prophylaxis for contact persons, as well as prevention of outbreaks.  
320 Records of Vietnamese and Southeast Asian *N. meningitidis* isolates are still extremely limited,  
321 thus it is not possible to determine the origin of these strains, or how they have arisen. Study at  
322 the genomic level of additional historical invasive and carriage strains collected at these and  
323 nearby camps, or nearby regions can help identify the phylogenetic routes that led to their  
324 emergence. On the other hand, since the military setting is among the highest risk group for  
325 meningococcal disease in adults, continual effort is needed to provide the surveillance data  
326 essential for effective policy making and preparation for response in case of potential outbreaks  
327 in the future.

328  
329 ~~Besides these, isolates with novel sequence types or sequence types that could not be~~  
330 ~~assigned to lineages were also frequently observed throughout the years and locations. Available~~  
331 ~~antigenic profiles suggested frequent exchange of genetic material via recombination among~~  
332 ~~strains and reservoirs, with several alleles (P1.20, P1.7-2) associated with the major lineages~~  
333 ~~frequently recurring in different combinations. Capsule switch could also be suggested for ST-~~  
334 ~~4821 serogroup B and C isolates. Clonal expansion was also observed, indicated by the~~  
335 ~~emergence of novel sequence types, and showed no distinct cluster in regard of geographical~~

336 locations, reflecting the close and frequent contact nature of the training units of new military  
337 recruits. From a search through PubMLST database, many emergent STs identified were limited  
338 to those units only, highlighting the niche characteristic of *N. meningitidis* population of the  
339 military camps.

341 **DISCUSSION**

342 *Neisseria meningitidis* remains the main cause of sporadic meningitis and sepsis in  
343 military recruit camps (Tran et al. 2019; Sim et al. 2013; Keiser, Hamilton, and Broderick 2011).  
344 Accurate identification and characterization of the causative strain is crucial for the success of  
345 treatment for patients and prophylaxis for contact persons, as well as prevention of outbreaks.

346 Previously, we have characterized the genomic features of a *N. meningitidis* strain causing a  
347 severe case of meningitis at a military camp in Vietnam in 2014 (Tran et al. 2019). In this study,  
348 we obtained two more isolates causing two meningitis cases at the nearby military camp in 2013  
349 and 2015. The isolates shared sequence type and were closely related to the previously reported  
350 isolate. The two most prominent features of their genomic sequences are variations at antigenic  
351 loci that are predicted not to be covered by Trumenba® and Bexsero®, the two currently used  
352 vaccines against NmB; and their similar antibiotic resistant profile, with notably high resistance  
353 for chloramphenicol. Genome comparison revealed a close phylogenetic relationship among all  
354 three Vietnamese isolates and suggesting all three strains were likely to originate from a common  
355 lineage.

356 Recently, Batty et al sequenced the genomes of 18 invasive *N. meningitidis* isolates from  
357 Thailand, Laos and Cambodia and revealed a group of eight chloramphenicol resistant isolates  
358 carrying identical *catP* gene, the same variant also found in NMB\_VN2014, suggesting a

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359 common origin (Batty et al. 2019). The two new isolates from Vietnam in this study also  
360 expressed resistance to chloramphenicol and carried the same *catP* variant. We reconstructed  
361 their phylogenetic relationship based on core genome comparison using NeighborNet analysis  
362 and revealed a chloramphenicol-resistant lineage clustered separately from the rest of isolates.  
363 This lineage seemed to have rapidly expanded clonally in recent years, though this could partially  
364 be due to better case report and laboratory detection, since data from this region was scarce up  
365 until recently although still remains limited. Although NeighborNet analysis placed this lineage  
366 as a divergent branch from other chloramphenicol-susceptible isolates in the region, due to the  
367 limited number of samples, it remains a possibility for their origin.

368 To better understand the probable evolutionary context of these strains, we have further  
369 collected and analyzed 25 isolates representing a collection of both historical and later-emerged  
370 isolates from the same or nearby military units with the three in this study (Table 4). Serogroup B  
371 was predominantly observed, besides serogroup C, which agrees with previous reports from  
372 sporadic cases in the region (Pancharoen et al. 2000). Though limited in number, this collection  
373 revealed the existence of two recurring lineages in all military units, centered around ST-1576  
374 and ST-4821, both were major hypervirulent STs with a long history and worldwide distribution.  
375 Antigenic profiles associated with these major STs show high similarity to strains described  
376 before in China during 1978–2013 (Zhu et al. 2015). Besides these, strains representing  
377 singleton and novel STs were also frequently observed, many limited only to these units,  
378 suggesting close transmission extending beyond individual camps seemed to be the main driving  
379 force for *N. meningitidis* prevalence and expansion within the military environment in Vietnam.

380 Records of Vietnamese and Southeast Asian *N. meningitidis* isolates are still extremely  
381 limited, thus it is not possible to determine the origin of these strains, or how they have arisen.

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Study at the genomic level of additional historical invasive and carriage strains collected at these and nearby camps, or nearby regions can help identify the phylogenetic routes that led to their emergence. On the other hand, since the military setting is among the highest risk group for meningococcal disease in adults, continual effort is needed to provide the surveillance data essential for effective policy making and preparation for response in case of potential outbreaks in the future.

## CONCLUSIONS

In a previous study, we described for the first time the genome of a chloramphenicol-resistant invasive *N. meningitidis* B isolate from a military unit in Vietnam. In this study, using WGS analysis, we characterized the genetic features of two additional *N. meningitidis* B isolates causing sporadic meningitis in another military camp in Vietnam. Core genome comparisons highlights the close phylogenetic relationship of isolates both within and between camps, with emphasis on their shared antibiotic resistant genes and antigenic profiles that are likely yet covered by current meningococcal B vaccines, Trumenba® and Bexsero®. Another notable shared feature of these isolates was their high resistance against chloramphenicol, likely attributed by but not limited to the 624 bp *catP* variant that were previously found in chloramphenicol-resistant isolates in France (Galimand et al. 1998) and Southeast Asia (Batty et al. 2019). A phylogenetic network reconstructed from core genome comparison suggests a common lineage of chloramphenicol resistant isolates in the military camps of Vietnam and other Southeast Asian countries that seemed to be expanding in this region.

Since molecular knowledge of the epidemiological characteristics of *N. meningitidis* in Vietnam remains limited, we also reported epidemiological analysis of 25 invasive and carriage

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405 strains from these Vietnamese military camps. Besides the major lineages, additional groups of  
406 singleton and unique novel sequence types extending beyond individual camps were observed,  
407 indicating close transmission is likely the main driving force for *N. meningitidis* prevalence and  
408 expansion within the military environment in Vietnam. Taken together, our results provide useful  
409 information for further understanding the molecular epidemiology of *N. meningitidis* in the  
410 military units in Vietnam, aiding future meningococcal meningitis monitoring and surveillance in  
411 the country.

412

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415 research utilized the PubMLST database (<https://pubmlst.org/>) developed by Keith Jolley (Jolley  
416 and Maiden 2010).

417 **Competing interests**

418 The authors declare that they have no competing interests.

419 **Availability of data and materials**

420 Assembly data and genomic sequences were deposited to NCBI Genomes database under  
421 BioProject ID PRJNA523495. Isolate records and novel sequence types were submitted to  
422 PubMLST.

423 **Authors' contributions**

424 TTL, TXT, and LPT performed experiments. LPT cultured isolates and provide laboratory  
425 records of additional isolates. CMA supported data analysis and contributed to revising and  
426 proofreading the manuscript. DVQ contributed to study design and manuscript drafting and

427 revision. HMN designed experiments, interpreted data, wrote and revised the manuscript. All  
428 authors read and approved the final manuscript.

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**Table 1.** Epidemiological characterization of historical and emerging *N. meningitidis* isolates from military camps in Vietnam

<u>Isolate</u>	<u>Year</u> <sup>a</sup>	<u>Description</u> <sup>a</sup>	<u>Military unit</u> <sup>a</sup>	<u>Sequence type (ST)</u> <sup>b</sup>	<u>Strain designation</u> <sup>b</sup>
<a href="#">Khoa</a>	2008	meningitis	Unit 2	1576	B: P1.19,15-39: ST-1576
<a href="#">37C</a>	2012	carrier	Unit 3 <sup>c</sup>	4821	C: P1.20,2: ST-4821 (cc4821)
<a href="#">40C</a>	2012	carrier	Unit 3 <sup>c</sup>	4821	C: P1.20,2: ST-4821 (cc4821)
<a href="#">Bach</a>	2013	meningitis	Unit 1	13455	B: P1.19,15: ST-13455
<a href="#">NMB_VN2013</a>	2013	meningitis	Unit 2	1576	B: P1.7-2,13: F4-6: ST-1576
<a href="#">14072</a>	2014	carrier	Unit 4 <sup>d</sup>	13065	B: P1.18,Δ: F18: ST-13065
<a href="#">14075</a>	2014	carrier	Unit 4 <sup>d</sup>	13065	B: P1.Δ,Δ: ST-13065
<a href="#">14089</a>	2014	carrier	Unit 4 <sup>d</sup>	13065	B: P1.Δ,Δ: F18: ST-13065
<a href="#">14155</a>	2014	carrier	Unit 1	4821	C: P1.20,2: ST-4821 (cc4821)
<a href="#">14156</a>	2014	carrier	Unit 1	4821	C: P1.20,2: ST-4821 (cc4821)
<a href="#">14157</a>	2014	carrier	Unit 1	4821	C: P1.20,2: F22: ST-4821 (cc4821)
<a href="#">14196</a>	2014	carrier	Unit 2	1576	B: P1.7-2,13-2: ST-1576
<a href="#">NMB_VN2014</a>	2014	meningitis	Unit 1	13074	B: P1.22-25,14-32: F4-6: ST-13074
<a href="#">15020</a>	2015	carrier	Unit 1	1576	B: P1.7-2,13-1: ST-1576
<a href="#">1513</a>	2015	carrier	Unit 1	13056	B: P1.Δ,Δ: F18: ST-13065
<a href="#">1523</a>	2015	carrier	Unit 1	1576	B: P1.7-2,13-2: ST-1576
<a href="#">1530</a>	2015	carrier	Unit 1	1576	B: P1.22-25,14: ST-1576
<a href="#">1533</a>	2015	carrier	Unit 1	1576	B: P1.7-2,13: ST-1576
<a href="#">1535</a>	2015	carrier	Unit 1	44	B: P1.7-2,13-1: ST-44 (cc41/44)
<a href="#">NMB_VN2015</a>	2015	meningitis	Unit 2	1576	B: P1.22-25,14: F1-7: ST-1576
<a href="#">1237C</a>	2016	carrier	Unit 3 <sup>c</sup>	4821	C: P1.20,2: ST-4821 (cc4821)
<a href="#">16005</a>	2016	carrier	Unit 2	1576	B: P1.22-25,14: ST-1576
<a href="#">16016</a>	2016	carrier	Unit 2	1576	B: P1.22-25,14: ST-1576
<a href="#">16406</a>	2016	carrier	Unit 1	4821	B: P1.7-2,14: ST-4821 (cc4821)
<a href="#">16408</a>	2016	carrier	Unit 1	4821	B: P1.7-2,14: ST-4821 (cc4821)
<a href="#">16416</a>	2016	carrier	Unit 1	4821	B: P1.7-2,14: F80: ST-4821 (cc4821)
<a href="#">17088</a>	2017	carrier	Unit 1	13074	B: P1.22-25,Δ: ST-13074
<a href="#">17090</a>	2017	carrier	Unit 1	13074	B: P1.Δ,Δ: ST-13074

<sup>a</sup> Isolates' metadata (year, clinical description, and location) were obtained from laboratory records of the Laboratory of Microbiology, Military Institute of Preventive Medicine, Hanoi.

<sup>b</sup> Molecular data was extracted from genomic sequence (NMB\_VN2013, NMB\_VN2014, NMB\_VN2015) or laboratory records (other isolates). Strain designation was based on the previously recommended nomenclature (Jolley, Brehony, and Maiden 2007), comprising of serogroup, *porA* type (Px), *fHbp* type (Fx), and sequence type (STx) (clonal complex (ccx))

<sup>c</sup> Unit 3 is geologically close to Unit 1, <sup>d</sup> Unit 4 is geologically close to Unit 2

**Table 2.** Allelic profiles of antibiotic resistant genes <sup>a</sup> and antibiotic susceptibility of the Vietnamese isolates <sup>b</sup>

	Locus											Antibiotic susceptibility						
	<i>gyrA</i>	<i>penA</i>	<i>tpoB</i>	NEIS0123	NEIS0414	NEIS1320	NEIS1525	NEIS1600	NEIS1609	NEIS1635	NEIS1753	AM	CIP	CTX	CRO	RI	MRP	CL
<b>NMB_VN2013</b>	2	587	42	1446	1	32	1338	1315	NA*	7	NA*	I	S	S	S	S	S	R
												0.42	0.004	0.016	0.002	0.25	0.064	62
<b>NMB_VN2014</b>	2	587	42	1446	1	32	1338	NA	NA*	1086	NA*	I	S	S	S	I	S	R
												0.5	0.008	0.016	0.004	1.5	0.094	256
<b>NMB_VN2015</b>	2	587	42	1446	1	32	1338	1315	NA*	7	2242	I	S	S	S	S	S	R
												0.62	0.008	0.023	0.002	0.125	0.064	64

<sup>a</sup> An allele number was assigned to each locus based on its DNA sequence using PubMLST database (<https://pubmlst.org/>) (Jolley and Maiden 2010)

<sup>b</sup> Antibiotic susceptibility of isolates was examined using E-test strip (bioMerieux, France) and interpreted according to CLSI 2018 breakpoints (CLSI 2018).

NA\*: new, unassigned alleles identical at said locus; I: Intermediate, S: Susceptible, R: Resistance. Numbers below each susceptibility interpretation indicate MIC (µg/ml) values.



**Table 3.** Antigenic profile of Vietnamese and Southeast Asian isolates, with middle line separate the chloramphenicol-resistant (above) and susceptible (below) isolates

Isolate	Sequence type and clonal complex (ST (cc)) <sup>a</sup>	Antigenic profile <sup>b</sup>				
		<i>porA</i>		<i>fHbp</i>	<i>nhbA</i>	<i>fetA</i>
		VR1	VR2	Peptide	Peptide	
NMB_VN2013	1576	7-2	13	31	16	4-6
NMB_VN2014	13074	22-25	14-32	31	16	4-6
NMB_VN2015	1576	22-25	14	31	16	1-7
NM01	14487	19	15	283	16	1-20
NM11	14496	19	15	31	-	3-7
NM12	1576	19-1	15-31	31	16	4-6
NM13	1576	19	15	-	16	5-88
NM16	1576	19	15	31	-	-
NM18	1576	19	15	1035	16	3-31
NM20	11005	19	15-39	31	16	5-135
NM25	1576	19	15	31	16	-
NM14	1145 (cc41/44)	7-2	4	14	2	1-20
NM15	41 (cc41/44)	7-2	4	14	2	1-19
NM19	14503 (cc4821)	20	23-7	141	669	-
NM21	12811	12-1	13-1	18	945	1-19
NM23	14507	22	23-1	-	21	4-21
NM03	14488 (cc41/44)	7-2	4	14	2	1-49
NM04	14489	22-15	-	5	-	-
NM06	32 (cc32)	18	-	101	-	1-21
NM07	3256	7-1	-	24	1086	3-1
NM09	5604	22-1	26	-	1068	3-2

<sup>a</sup> Sequence type (ST) and clonal complex (cc) determined by the sequence of seven house-keeping genes (*abcZ*, *adk*, *aroE*, *fumC*, *gdh*, *pdhC*, and *pgm*)

<sup>b</sup> Allele number assigned to each locus based on its DNA (*porA*) or protein (*fHbp*, *nhbA*, and *fetA*) sequences

<sup>a, b</sup> Analyses was performed using PubMLST database (<https://pubmlst.org/>) (Jolley and Maiden 2010)

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**Table 4.** Diversity of virulent determining factors from genome sequence of the Vietnamese isolates

<u>Locus</u>	<u>Gene</u>	<u>Product</u>	<u>NMB VN</u> <u>2013</u>	<u>NMB VN</u> <u>2014</u>	<u>NMB VN</u> <u>2015</u>
<b><u>Capsular gene cluster<sup>a, b</sup></u></b>					
<u>NEIS0046</u>	<u><i>rfbA</i></u>	glucose-1-phosphate thymidyltransferase	<u>NA</u>	<u>NA</u>	<u>NA</u>
<u>NEIS0054</u>	<u><i>cssA</i></u>	N-acetylglucosamine-6-P 2-epimerase	<u>NA*</u>	<u>NA*</u>	<u>113</u>
<u>NEIS0067</u>	<u><i>ctrF</i></u>	capsule translocation	<u>4</u>	<u>NA*</u>	<u>NA*</u>
<b><u>Maf-toxin genomic island<sup>a</sup></u></b>					
<u>NEIS1788</u>	<u><i>anmK</i></u>	anhydro-N-acetylmuramic acid kinase	<u>25</u>	<u>25</u>	<u>NA</u>
<u>NEIS1795</u>	<u><i>mafI<sub>Δ2MGL-2</sub></i></u>	MafI immunity protein	<u>4;186</u>	<u>4</u>	<u>4;186</u>
<u>NEIS2083</u>	<u><i>mafA<sub>MGL-3</sub></i></u>	MafA3 lipoprotein	<u>252</u>	<u>252</u>	<u>252;890</u>
<u>NEIS2084</u>	<u><i>mafB<sub>MGL-3</sub></i></u>	MafB3 toxin protein	<u>31</u>	<u>NA</u>	<u>31</u>
<b><u>Outer membrane vesicle (OMV)peptide<sup>c</sup></u></b>					
<u>NEISp0653</u>	<u>:</u>	Competence lipoprotein	<u>NA*</u>	<u>NA*</u>	<u>1</u>
<u>NEISp0275</u>	<u>:</u>	Organic solvent tolerance protein	<u>NA*</u>	<u>NA</u>	<u>NA*</u>
<u>NEISp0923</u>	<u>:</u>	Antioxidant AhpC TSA family glutaredoxin	<u>2</u>	<u>2</u>	<u>NA</u>
<u>NEISp1364</u>	<u>:</u>	Outer membrane protein PorA	<u>155</u>	<u>NA</u>	<u>NA</u>
<u>NEISp1687</u>	<u>:</u>	Phospholipase A1	<u>254</u>	<u>254</u>	<u>NA</u>
<u>NEISp1690</u>	<u>:</u>	Transferrin-binding protein 1	<u>NA*</u>	<u>NA*</u>	<u>NA</u>
<u>NEISp1963</u>	<u>:</u>	Iron-regulated outer membrane protein FrpB	<u>NA*</u>	<u>NA*</u>	<u>NA</u>
<b><u>Pilin genes<sup>a</sup></u></b>					
<u>NEIS0020</u>	<u><i>pilB/msrAB</i></u>	peptide methionine sulfoxide reductase MsrA/MsrB	<u>379</u>	<u>11</u>	<u>NA</u>
<u>NEIS0021</u>	<u><i>pilA/ftsY</i></u>	probable signal recognition particle protein	<u>NA</u>	<u>1778</u>	<u>1978</u>
<u>NEIS0036</u>	<u><i>pilTI</i></u>	type IV pilus retraction ATPase PilT	<u>200</u>	<u>11</u>	<u>11</u>
<u>NEIS0210</u>	<u><i>pilE</i></u>	PilE	<u>NA</u>	<u>NA</u>	<u>NA</u>

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<u>NEIS0213</u>	<u><i>pglA</i></u>	<u>pilin glycosyltransferase</u>	<u>NA</u>	<u>1053</u>	<u>1053</u>
<u>NEIS0830</u>	<u><i>pilK</i></u>	<u>type IV biogenesis protein</u>	<u>1905</u>	<u>1618</u>	<u>1618</u>
<u><i>pilS</i></u>	<u><i>pilS</i> cassette -</u>		<u>NA</u>	<u>NA</u>	<u>NA</u>

<sup>a</sup> Allele numbers assigned to each locus based on DNA sequence using PubMLST database  
(<https://pubmlst.org/>) (Jolley and Maiden 2010)

<sup>b</sup> Comparison result at *cnl* (capsule null locus) was omitted since all isolates were capsulated

<sup>c</sup> Allele numbers assigned to each locus based on protein sequence using PubMLST

NA: new, unassigned alleles; NA\*: new, unassigned alleles identical at said locus

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