

Fauna associated with shallow-water methane seeps in the Laptev Sea

Andrey A Vedenin^{Corresp., 1}, Sergey V Galkin², Valentin N Kokarev^{3, 4}, Margarita V Chikina⁴, Alexander B Basin⁴, Andrey V Gebruk²

¹ Laboratory of plankton communities structure and dynamics, P.P. Shirshov Institute of Oceanology, Moscow, Russia

² Laboratory of Ocean Bottom Fauna, P.P. Shirshov Institute of Oceanology, Moscow, Russia

³ Faculty of Biosciences and Aquaculture, Nord University, Bodø, Norway

⁴ Laboratory of Ecology of Coastal Bottom Communities, P.P. Shirshov Institute of Oceanology, Moscow, Russia

Corresponding Author: Andrey A Vedenin

Email address: urasterias@gmail.com

Background. Methane seeps provide environment for unique benthic ecosystems in the deep-sea dependent on chemosynthetic organic matter. In contrast, in shallow waters there is little or no effect of methane seeps on macrofauna. In the present study we focused on the recently described methane discharge area at the northern Laptev Sea shelf. The aim of this work was to describe the shallow-water methane seep fauna and to understand whether the differences in community structure between the methane seep and background areas exist.

Methods. Samples of macrofauna were taken during three expeditions of RV *Akademik Mstislav Keldysh* in 2015, 2017 and 2018 using 0.1 m² grabs and the Sigsbee trawl. 21 grabs and two trawls in total were taken at two methane seep sites named *Oden* and *C15*, located at depths of 60-70 m. For control, three 0.1 m² grabs were taken in area without methane seepage.

Results. The abundance of macrofauna was higher at methane seep stations, compared to non-seep. Cluster analysis revealed five station groups corresponding to control area, *Oden* site and *C15* site (the latter represented by three groups). Taxa responsible for differences between the station groups were mostly widespread Arctic species, that were more abundant in samples from methane seep sites. However, large densities of symbiotrophic siboglinids *Oligobrachia* sp. were found exclusively at methane seep stations. In addition, several species presumably new to science were found at several methane seep stations, including the gastropod *Frigidalvania* sp. and the polychaete *Ophryotrocha* sp. The fauna at control stations was represented only by well-known and widespread Arctic taxa. Number of station groups revealed from *C15* stations and high species richness in *C15* trawl sample comparing to *Oden* indicated higher diversity of microniches within the *C15* site. The development of specific methane seep communities at such shallow depths can be related to pronounced oligotrophic environment on the northern Siberian shelf.

Fauna associated with shallow-water methane seeps in the Laptev Sea

Andrey A. Vedenin¹, Sergey V. Galkin¹, Valentin N. Kokarev^{1,2}, Margarita V. Chikina¹,
Alexander B. Basin¹, Andrey V. Gebruk¹

¹ – P.P. Shirshov Institute of Oceanology, Moscow, Russia

² – Nord University, Faculty of Biosciences and Aquaculture, Bodø, Norway

Corresponding Author:

Andrey A. Vedenin¹

Nahimovskiy Prospekt 36, Moscow, 117997, Moscow, Russia

Email address: urasterias@gmail.com

Abstract

Background. Methane seeps provide environment for unique benthic ecosystems in the deep-sea dependent on chemosynthetic organic matter. In contrast, in shallow waters there is little or no effect of methane seeps on macrofauna. In the present study we focused on the recently described methane discharge area at the northern Laptev Sea shelf. The aim of this work was to describe the shallow-water methane seep fauna and to understand whether the differences in community structure between the methane seep and background areas exist.

Methods. Samples of macrofauna were taken during three expeditions of RV *Akademik Mstislav Keldysh* in 2015, 2017 and 2018 using 0.1 m² grabs and the Sigsbee trawl. 21 grabs and two trawls in total were taken at two methane seep sites named *Oden* and *C15*, located at depths of 60-70 m. For control, three 0.1 m² grabs were taken in area without methane seepage.

Results. The abundance of macrofauna was higher at methane seep stations, compared to non-seep. Cluster analysis revealed five station groups corresponding to control area, *Oden* site and *C15* site (the latter represented by three groups). Taxa responsible for differences between the station groups were mostly widespread Arctic species, that were more abundant in samples from methane seep sites. However, large densities of symbiotrophic siboglinids *Oligobrachia* sp. were found exclusively at methane seep stations. In addition, several species presumably new to science were found at several methane seep stations, including the gastropod *Frigidalvania* sp. and the polychaete *Ophryotrocha* sp. The fauna at control stations was represented only by well-known and widespread Arctic taxa. Number of station groups revealed from *C15* stations and high species richness in *C15* trawl sample comparing to *Oden* indicated higher diversity of microniches within the *C15* site. The development of specific methane seep communities at such shallow depths can be related to pronounced oligotrophic environment on the northern Siberian shelf.

38

39 **Introduction**

40 Methane gas seeping from the seafloor, similar to hydrothermal vents, can provide
 41 environmental conditions for unique fauna largely independent of photosynthetic primary
 42 production (Van Dover, 2000). Distinct faunal response at methane seeps (also known as “cold
 43 seeps”) is associated with certain increase of total abundance/biomass and presence of unique
 44 taxa absent in background areas. This pattern has been described from many areas of the Ocean
 45 (Baker & German, 2004; Levin, 2005). These taxa either develop symbiotic relationships with
 46 methanotrophic or sulphide-oxidizing bacteria or feed directly on benthic or suspended bacterial
 47 matter. As a next trophic level, predators feeding exclusively on such taxa may be present
 48 (Gebruk, 2002; Dando, 2010).

49 In the Arctic Ocean, several methane seep ecosystems have been discovered and
 50 investigated. The most studied include the Håkon Mosby mud volcano in the Norwegian Sea
 51 (Gebruk et al., 2003) and several sites around Svalbard and at Vestnesa Ridge (Åström et al.,
 52 2016; Åström et al., 2018). Other described cold seeps include the Lofoten-Vesterålen
 53 continental margin area (Sen et al., 2019a) and mud volcanoes in the Beaufort Sea (Paull et al.,
 54 2015). The cold seeps inhabited by specific benthic macrofauna are mostly located below the
 55 photic zone (depth >200 m both around Svalbard and at Håkon Mosby) (Gebruk et al., 2003;
 56 Åström et al., 2016). At the same time, areas with extensive methane discharge located at
 57 shallow depths (e.g. in the Norwegian and White Seas at depths <100 m) have no or minor
 58 response of macrofauna (Savvichev et al., 2004; Levin, 2005). In general, a depth boundary is
 59 observed between shallow-water cold seeps and their “deep-sea” counterparts at approximately
 60 200 m (Tarasov et al., 2005; Dando, 2010). One of possible reasons for this boundary is the
 61 origin of organic matter: at depths <200 m photosynthetic organic matter is more available for
 62 benthic consumers due to stronger benthic-pelagic coupling. At greater depth, however, the
 63 amount of photosynthetic organic matter decreases and chemosynthesis starts to play a
 64 significant role for local organic matter production. Therefore, despite the presence of methane
 65 and sulfides (unfavorable for most organisms), unique and diverse ecosystems can develop at
 66 deep-sea cold seeps (summarized by Dando, 2010).

Fauna associated with cold seeps in the Arctic includes symbiotrophic siboglinid polychaetes and thyasirid bivalves, but mainly consists of species not unique for methane seeps. Widespread Arctic species tend to aggregate in habitats around methane seepage sites (Gebruk et al., 2003; Åström et al., 2016; Åström, Oliver & Carroll, 2017). Arctic cold seep assemblages are characterized by the dominance of frenulate siboglinid worms, while large chemosymbiotrophic methane seep taxa (vestimentiferan worms, bathymodioline and vesicomyid bivalves) are absent (Sen et al. 2018). Common effect of methane seeps on benthic communities around the Ocean is an increased abundance and biomass of regular allochthonous taxa compared to the background (Gebruk, 2002; Levin, 2005). The diversity values at cold seeps are not higher than in the background, though recent results obtained from the southwestern Barents Sea showed increased taxonomic richness within the seepage sites (Sen et al., 2019b).

In the Siberian Arctic, areas of intense methane discharge (methane seeps) were discovered on the outer shelf of the Laptev Sea in 2008 (Yusupov et al. 2010). Further research revealed numerous gas flares in the northern Laptev Sea shelf (Lobkovsky et al., 2015; Shakhova 2015). Within this area specific microbial communities based on methane oxidation were discovered (Savvichev et al. 2018). Characteristic features and structural control of methane seep fields in this area were given in Baranov et al. (2019). It is suggested that the methane seeping occurs through the fault system belonging to Laptev Sea Rift system and Khatanga-Lomonosov Fracture Zone located between the Eurasian and North American Tectonic Plates. The faults may conduit the gas from reservoirs deep in the sediment below the caprock formed by permafrost and gas hydrates (Baranov et al., 2019). Within the seeping area, multiple bacterial mats and occasional methane bubbles and carbonate crusts were observed (Baranov et al., 2019).

Notably, the methane associated fauna was registered on the Laptev Sea shelf and slope much earlier: during expeditions of RV *Polarstern* in 1993 and 1995 five species of siboglinids were found in this area in the depth range 50-2000 m (Sirenko et al., 2004), which is more than anywhere else in the high Arctic.

We examined benthic communities associated with methane seeps in the Laptev Sea at two sites: *C15*, centred around 76°47.4'N and 125°49.5'E with depths 70-73 m and *Oden*, centred around 76.894°N and 127.798°E, with depths 63-67 m. Preliminary description of benthic fauna observed on video was published by Baranov et al. (2019). The aim of this study is to describe further the biological peculiarities of the methane seep fauna and to reveal the differences in

either integral community characteristics or distribution of certain species between the methane seep and background areas. We hypothesized that the seep sites are different from the non-seep in terms of general community characteristics and certain species distribution.

Materials & Methods

Samples of macrofauna were taken during three expeditions of RV *Akademik Mstislav Keldysh* in 2015 (AMK-63), 2017 (AMK-69) and 2018 (AMK-72) on the northern Laptev Sea shelf, in the area of active methane discharge. The gears used for sampling were the *Okean* (in 2015) and *Van Veen* (in 2017-2018) grabs (0.1 m² sampling area) and the *Sigsbee* trawl (2 m frame width) (Eleftheriou & McIntyre, 2005). 21 grab and two trawl stations were performed in total at three sites: on two methane seep fields (12 grabs and one trawl at *C15* and six grabs and one trawl at *Oden*) and at the control site with no methane seeping (three grabs) (Fig. 1). A single trawl was taken at each seep site to minimize the possible ecosystem damage from this gear. In 2015 three seep stations were selected above the present gas flares visible on echo-sounder. Three more grabs were taken ~200 m away from the nearest gas flare to catch background community. In 2017 and 2018 station selection was based largely on the previously mapped methane flares. All the 2017 and 2018 grabs were taken above the gas flares (Fig. 1). Station data with coordinates and depths are shown in Table 1. For additional information on methane seep fields see Flint et al. (2018) and Baranov et al. (2019).

Table 1. Data on stations used in the present study. For trawl stations coordinates and depth of start and end are given.

Fig. 1. Study area.

Enlarged maps show sampling sites and corresponding stations. Detailed bathymetry is only available for *C15* and *Oden* sites; white circles indicate previously recorded gas flares (Baranov et al., 2019). Dotted line at *Oden* site enclosed map shows the approximate perimeter of seeping area.

Sediment from grab samples was washed by hand through the 0.5 mm mesh size sieve, and fixed with buffered 4% formalin solution afterwards. Two grab samples from the expedition in 2018 (Sts. 5947-3 at *C15* and 5953-2 at *Oden* site) were fixed with 96% ethanol. A 10-litre subsample of sediment taken from each trawl catch was washed through the 1 mm mesh size sieve and then fixed with neutralized 4% formalin. The material obtained was analyzed in the

laboratory; all macrofaunal organisms were identified to the lowest possible taxonomical level with the help of taxonomical experts (see Acknowledgements) and counted. Species from grab samples were weighed (wet weight, all specimens of each species at a time). Mollusks were weighed with shells, polychaetes with calcareous (spirorbids) or mucous tubes (*Spiochaetopterus typicus* and siboglinids) were weighed with tubes. Density and biomass were calculated per square meter in case of grabs. Dominant species were distinguished by biomass. For trawl samples we calculated the contribution (in %) of each species to abundance. Biomass was not measured for trawl samples due to poor state of preservation. For ethanol fixed samples from Sts. 5947-3 and 5053-2, the biomass loss was corrected using taxa-specific coefficients after Brotskaya & Zenkevich (1939).

For grab samples total abundance, biomass, species richness (species number), Pielou evenness, Hurlbert rarefaction index and Shannon-Wiener diversity index ($H' \ln$) were calculated to get integral community characteristics. Abundance and biomass data from grab samples were square root transformed to increase the role of rare taxa. The similarity between grab samples and species was estimated using the quantitative index of Bray-Curtis. Clusters were built based on similarity matrices using the unconstrained tree routine (UNCTREE); results were verified by SIMPROF to distinguish different station groups with significant differences in species composition. The results from cluster analysis were verified by non-metric multidimensional scaling (n-MDS). Clusters revealed by these methods were defined as separate station groups in terms of quantitative taxonomical similarity. Shade plots were built to visualize the species abundance and biomass differences between the stations and species in clusters. The Kruskal-Wallis test was used to verify differences in certain taxa distribution between station groups. Results were corrected using the Tukey's pairwise post-hoc test. Species-individuals accumulation curves were plotted for each station group (McCune, Grace & Urban, 2002; Clarke & Gorley, 2015).

For all species present in any station group, an algorithm estimating the likelihood of accidental catch was applied. If a uniform distribution of species between two sampling efforts A and B is assumed, the probability of species absence at each station of B-sampling would be $(1 - P_A)^{N(B)}$, where $N(B)$ is the number of stations in B-sampling and P_A is the species occurrence (the proportion of stations where the species was present) in A-sampling. Using this equation, the likelihood of accidental absence of any species in either station group can be estimated. The

number of grabs required for species catch in B-sampling can be calculated by the equation: $n = \lg(\alpha) \lg(1-P_A)^{-1}$, where α is the likelihood of species finding in B-sampling taken as 0.99 (Azovsky, 2018; Vedenin et al., 2019).

For trawl samples, the species rank distributions were plotted. Species richness, Pielou evenness, Hurlbert rarefaction index and Shannon-Wiener diversity index were calculated using the taxa-percentage values. Differences between trawl catches were estimated by similarity percentage routine (SIMPER).

Statistical analyses were performed in Primer V6, V7 and Past 3.0 software (Clarke & Warwick, 2001; Hammer, 2013; Clarke & Gorley, 2015).

Results

A total of 289 taxa of benthic macrofauna were identified in grab and trawl samples. In grab samples, density varied in the wide range from 580 ind. m⁻² (St. 5624-3, Control site) to 9880 ind. m⁻² (St. seep-3, C15 site). Biomass ranged from 16.28 g ww m⁻² (St. seep-1, C15 site) to 405.79 g ww m⁻² (St. 5623-3, Oden site). The list of all identified taxa from trawl and grab samples, with values of abundance and biomass is given in the Supplementary 1.

Fig. 2. UNCTREE analysis with SIMPROF results (A) and non-metric multidimensional scaling plot (B) of grab stations using the Bray-Curtis similarity index. Square root transformed biomass data are used. Dashed lines connect statistically unreliable groupings ($p > 0.05$). Green lines indicate SIMPROF groups.

Grab samples

Unconstrained tree with SIMPROF analysis revealed five significantly distinct groups of samples at the similarity level of 50 (Fig. 2). The UNCTREE parameters are shown in Supplementary 2. The groups partly corresponded with the station locations and presence/absence of methane seeps (*Control*, *C15* and *Oden* sites). To avoid a mix-up between the station groups and seeping sites hereinafter the corresponding names are used with either – station group or –site ending.

Fig. 3. The species-individuals accumulation curves for the station groups. Colors are the same as in Figure 2.

194

195 *Characteristics of station groups*

196 The *Control* station group included three stations located between methane seep sites (Fig. 1).
 197 The bivalve *Portlandia arctica* was dominant by biomass at all three stations; the starfish
 198 *Ctenodiscus crispatus* and the bivalve *Macoma calcareo* played a secondary role. Due to the low
 199 number of samples, the species-individuals accumulation curve did not reach the saturation point
 200 (Fig. 3). Compared to other groups, the values of density and species richness were the lowest at
 201 *Control*, whereas the evenness was the highest (Fig. 4).

202 The *C15-seep a* station group included five stations, all within the *C15* seep site. In this
 203 group, biomass and diversity values were relatively low. Dominant species in this group were the
 204 bivalve *Nuculana pernula*, the siboglinid *Oligobrachia* sp. and the polychaete *Cistenides*
 205 *hyperborea*. Species-individuals accumulation curve in this group reached saturation due to the
 206 largest number of samples (Fig. 3).

207 The *C15-seep b* station group consisted of only two stations from *C15* site. This group
 208 demonstrated the highest abundance values and the lowest biomass values owing to high
 209 abundance of small polychaetes *Cossura longocirrata*, *Micronephthys minuta* and *Ophryotrocha*
 210 sp. at some stations (Fig. 4, Supplementary 1).

211 The *Oden* station group included six stations, all located within the *Oden* seep site. Values
 212 of biomass, species richness and diversity indices in this group were the highest among all
 213 station groups (Fig. 4). The main dominant species were the siboglinid *Oligobrachia* sp. and the
 214 polychaetes *Myriochele heeri* and *Nephtys ciliata*.

215
 216 Fig. 4. Univariate characteristics of identified clusters.

217 Mean values of total density, biomass, species richness, Pielou evenness, Hurlbert rarefaction
 218 index and Shannon-Wiener index with standard deviation are shown. Exact values of these
 219 characteristics are shown in Supplementary 3.

220

221 The last group *C15 background* contained five stations taken within the *C15* site. Three of
 222 these stations (Sts. background-1, 2 and 3) were taken several hundred meters away from active
 223 methane seeps. Two stations (5947-1 and 5947-3) were planned as active seep stations.
 224 Accidentally, they were taken in the seep background area according to taxa composition and
 225 following analysis. Taxonomical composition at these stations was similar to that in the *Control*
 226 group, with the bivalve *Portlandia arctica* being the dominant species. Bivalves *Yoldiella*

lenticula and *Y. solidula* were subdominant. In this station group, the biomass values were the lowest, other general community characteristics were intermediate (Fig. 4). As with the *Control* group, *C15 background* did not reach the saturation point at species-individuals accumulation plot (Fig. 3).

Comparison of seep and non-seep station groups

General community characteristics in the station groups appeared different in abundance, biomass and diversity (Fig. 4, Supplementary 3). The abundance of several taxa was significantly different in four station groups (Fig. 5). The Kruskal-Wallis test showed that differences in abundance of at least ten species are statistically reliable (Table 2). The seep sites were characterized by higher density of the polychaetes *Tharyx* sp. and *Cistenides hyperborea*, the bivalve *Macoma calcarea* and the ophiuroid *Ophiocten sericeum*. On the contrary, the bivalve *Portlandia arctica* was markedly more abundant in *Control* and, to a lesser extent, in *C15 background* station groups (Fig. 5). Notable were extreme densities of small polychaetes at some seep stations, including *Cossura longocirrata* and *Ophtyotrocha* sp. (Fig. 5A) at *C15 seep b*.

Fig. 5. Shade plot of species square root transformed abundance (A) and biomass (B) at stations arranged by clusters. The species list is reduced to 20 most important taxa. Order of stations and colors the same as in Figure 2. Taxa grouped in clusters using UPGMA algorithm based on index of association.

Certain species present at some methane seep sites were completely absent at the non-seep sites (Fig. 5). Among them, at least four species (the polychaete *Spiochaetopterus typicus*, the siboglinid *Oligobrachia* sp., the bivalve *Axinopsida orbiculata* and the amphipod *Pleusymtes pulchellus*) were present only at *C15* and *Oden* sites not randomly. At least one species, the undescribed gastropod *Frigidalvania* sp., was present only in *Oden* station group and absent elsewhere not randomly (Table 3). The estimated number of grabs required to catch the latter species was slightly lower than the number of grabs taken.

Table 2. Results of the Kruskal-Wallis and Tukey's post-hoc tests for taxa with different abundance values in five station groups. Mean abundance in each station group is shown. Taxa are arranged according to *p*-value. Pairs in post-hoc column indicate significant comparisons (Tukey's *p* < 0.05). 1 – *Control* group; 2 – *C15 background* group; 3 – *Oden* group; 4 – *C15-seep a* group; 5 – *C15-seep-b* group.

Table 3. Likelihood of not finding a species calculated for species present only at methane seep sites and only at the *Oden* site.

Trawl samples

The overall Bray-Curtis similarity between the two trawls was 65.6%. Species ranking graphs showed high level of dominance by abundance for both trawl stations (Fig. 6). The dominant species in both trawls was the ophiuroid *Ophiocten sericeum*: 37% of the total abundance at *C15* and 46% at *Oden*. The second most abundant species at *C15* was the gastropod *Frigidalvania* sp. (12%) and at *Oden* the bivalve *Yoldiella solidula* (11%). Ten most abundant species accounted for >70% of the total abundance in both trawls (Fig. 6).

Fig. 6. Species ranking for *C15* and *Oden* trawl samples.
The most numerous species are indicated. X-axis is logarithmic.

Species richness, Pielou evenness, Hurlbert rarefaction for 100 individuals and Shannon-Wiener index are shown in Table 4. Diversity was higher in the *Oden*-trawl than in the *C15*-trawl, similar to results based on grab samples. However, the species richness (as well as the total amount of individuals) in the *C15*-trawl was higher than in the *Oden*-trawl (Table 4, Supplementary 1).

Species responsible for taxonomical difference between the two trawl samples are shown in Table 5. Most notable is a high amount of the gastropod *Frigidalvania* sp. at *C15*. At *Oden* *Frigidalvania* sp. was also present, but in much smaller densities (only 2.3 % of the total abundance). In addition, *C15*-sample differs from *Oden* by high amount of various filter-feeders including 6 species of sponges (with *Craniella polyura* being most numerous), at least 6 species of cnidarians, 17 species of bryozoans and 3 species of tunicates (Supplementary 1).

At *C15* trawl sample, a large piece of carbonate crust was found. Cavities of its pores were inhabited by numerous polychaetes, typical also for the soft sediments around the seepage area (e.g. members of families Nephthyidae, Nereididae, Oweniidae and Terebellidae, see Supplementary 1), and by several filter-feeders (Hydrozoa).

Table 4. Species richness, Pielou evenness, Hurlbert rarefaction for 100 individuals and Shannon-Wiener index calculated for trawl samples.

Table 5. Similarity percentage routine for trawl samples.
Species with contribution >0.5% are shown. Species more abundant at *C15* are marked with bold.

Comparison of *C15* and *Oden* sites

All gears showed significant differences between the *C15* and *Oden* sites expressed in different taxonomical composition and quantitative characteristics. The Bray-Curtis similarity between the sites according to the grab samples and trawl samples was 26.2 and 65.6, respectively. The main differences in species composition included the high abundance of the sponge *Craniella polyura* and the gastropod *Frigidalvania* sp. at *C15* site and higher numbers of the ophiuroid *Ophiecten sericeum* at *Oden* site.

The grab data indicated a high level of heterogeneity of benthic fauna on the scale of several meters at both seep-sites. Some species formed patches, for example *Oligobrachia* sp., *Cossura longocirrata* and *Ophryotrocha* sp., being extremely numerous at several grab stations. There were also species with rather uniform distribution based on combined data, for example *Ophiecten sericeum*. According to the cluster analysis, the *C15* site is more heterogenic forming at least three different species complexes within its area (Fig. 2). Dissimilarity within the *C15* and *Oden* sites was 64.7 and 26, respectively (Supplementary 2).

Dominant species were different in grab and trawl samples. The dominant species in trawls at both methane seep sites was the ophiuroid *Ophiecten sericeum*. Whereas based on grab data, the dominants at seep sites were the siboglinid *Oligobrachia* sp., the bivalve *Nuculana pernula* and the polychaete *Myriochele heeri*.

Discussion

Integral community parameters: methane seep vs. non-seep

The abundance of macrofauna was higher at the seep stations compared to the background. In addition, at *Oden* site the biomass was higher compared to non-seep sites. Increased values of

abundance and biomass have been reported from both hydrothermal vents and cold seeps all over the world. In the Arctic, a twofold increase of biomass compared to control sites was observed at cold seeps south off Svalbard (mean values of 20.7 vs. 9.8 g ww m⁻²), the abundance increase was less prominent (770 vs. 590 ind. m⁻²) (Åström, 2016). For the Håkon Mosby mud volcano, the comparison of abundance and biomass with the background is not available. In our study, the abundance at the methane seep sites *C15* and *Oden* was more than four times higher than at the control. However, differences in biomass although pronounced were not statistically reliable. Increased biomass in seep habitats is commonly explained by enhanced organic matter content and habitat heterogeneity (Gebruk et al. 2003; Sen et al. 2018).

Pielou's evenness was distinctly higher at the *Control* and *C15 background* station groups, which reflects the increased dominance of certain species at seep stations compared to non-seep. Many authors reported high abundance and biomass values of one to few dominant species at various cold seeps (Gebruk et al. 2003; Åström, 2016; Åström et al., 2018). This can be caused by conditions less favorable for some background species, but more favorable for symbiotrophs or grazers (summarized in Dando, 2010).

The cold seeps usually demonstrate lower diversity values compared to the background areas (Levin, 2005). However, combined species list from grab and trawl samples showed a big diversity taking into account that only two trawls were sampled. Our studies at Siberian shelf using the same gear under same conditions obtained less than 150 species per trawl (Galkin & Vedenin, 2015; Vedenin, Galkin & Kozlovsky, 2015), while a total of 203 species were found in a single *C15* sample. The unusually large diversity may reflect a higher amount of microniches within the *C15* site. This is indirectly confirmed by lower similarity values observed between all *C15* grab samples. Higher habitat heterogeneity at seep sites can increase the overall diversity of benthic fauna (Gebruk et al. 2003; Levin, 2005). The scale of heterogeneity is hard to assess, but based on stations coordinates and the fact that stations 5947-1 and 5947-2 from *C15*-site were grouped in *C15 background*, while station 5947-2 was grouped as *C15-seep* a we can assume that the scale is less than five meters (distance between these stations) (Fig. 1; Table 1).

In addition, the diversity values at *Oden* station group were significantly higher than at other sites. The reasons for this are unknown so far, since no environmental parameters measured directly at benthic stations are available. Interestingly, the peculiarly higher values of diversity

within the cold seeps are known only for the seep areas in the Arctic, e.g. for the Vestnesa Ridge (Åström et al., 2018) and for the South-Western Barents Sea (Sen et al., 2019b).

Common shelf taxa responsible for differences in station groups

The station groups revealed by UNKTREE and n-MDS analysis largely corresponded to the geographical position of the *C15*, *Oden* and control sites. A number of common species widely distributed across the Siberian shelf (see Supplementary 1, Sirenko, 2001) were largely responsible for increased integral community parameters in our study. Most of these taxa are listed in Table 2. Among such species (based on grab samples) were the polychaetes *Spiochaetopterus typicus*, *Cossura longocirrata* and *Tharyx* sp., the bivalve *Macoma calcarea*, the amphipod *Pleusymtes pulchella* and the ophiuroid *Ophiocten sericeum*. In addition, based on trawl data, the sponge *Craniella polyura* was present in high densities at the *C15* site, together with other filter-feeders including cnidarians and bryozoans. Apparently the same species aggregations were visible on the video reported by Baranov et al. (2019). All these species were previously reported from a wide range of areas of the Laptev Sea and adjacent regions (Sirenko et al., 2004).

The increased density of common taxa at deep-sea hydrothermal vents and cold seeps is a well-known phenomenon usually explained by increased availability of organic matter in these habitats (Hessler & Kaharl, 1995; Levin, 2005). In the Arctic, the increased biomass and abundance of common allochthonous species was reported for the Håkon Mosby mud volcano (Rybakova et al., 2013), Svalbard (Åström et al., 2016) and Vestnesa Ridge cold seeps (Åström et al., 2018). Also, a significant increase of abundance of filter feeders (especially sponges) was shown for the Aurora Seamount on the Gakkel Ridge, the only investigated hydrothermal vent in the Central Arctic Ocean (Boetius, 2015).

Fig. 7. Taxa found only at seep stations.

A – *Oligobrachia* sp. (left – tube with several fragments enlarged; center – complete specimen extracted from tube; right – anterior and posterior fragments of the specimen); B – *Frigidalvania* sp.; C – *Ophryotrocha* sp. (upper left – several specimens, total view; upper right – anterior fragment; lower – enlarged parapodia); D – *Axinopsida orbiculata*. Photos by A. Vedenin and V. Kokarev.

Taxa specific for methane seep sites

The main species marking the methane seeps in our study was the siboglinid *Oligobrachia* sp. (Fig. 7a). This species was present at every seep station and absent at every background/control station. This species is morphologically very close to *Oligobrachia haakonmosbiensis* originally described from the Håkon Mosby mud volcano from the depth of ~1200 m (Smirnov, 2000). Colonies of *O. haakonmosbiensis* with the biomass reaching 350 g ww m⁻² were reported from this area (Gebruk et al., 2003). Recent phylogenetical analyses showed that the species from the Laptev Sea belongs to a separate, undescribed species of *Oligobrachia* (Sen et al., 2018). In the Laptev Sea, *Oligobrachia* sp. is known from different localities, seep and noon-seep, occurring in a wide depth range 100-2166 m (Buzhinskaja, 2010). Our record at 63 m is the shallowest for this species, with high population density and biomass: >1000 ind. m⁻² and 45 g ww m⁻² at Sts. 2623-1 and 5953-2 (*Oden* site). Several specimens from 2015-samples (erroneously identified as *O. haakonmosbiensis*) were investigated using transmission electron microscopy (Savvichev et al., 2018). Interestingly, metanotrophic bacteria were found inside its trophosome. Usually the endosymbionts of Siboglinids are represented by sulphide-oxidizing bacteria (Rodrigues et al., 2011; Lee et al., 2019).

Some samples from the seep sites besides the siboglinids also were marked by several species of mollusks. The gastropod *Frigidalvania* sp. (Rissoidae) occurred in high density at *C15* site: up to 2340 ind. m⁻² and 25 g ww m⁻² at St. 5625-3 (Fig. 7b). According to trawl samples, this species occurs at *Oden* site, but in a few numbers. This species is new to science. Large numbers of unknown rissoid gastropods were previously reported from the Håkon Mosby, referred to as *Alvania* sp. in Gebruk et al. (2003). Later, the stable isotope analysis has shown that the rissoids at Håkon Mosby are grazing on bacterial mats (Decker & Olu, 2012). Another rissoid gastropod, *Pseudosetia griegi*, was observed grazing on bacterial mats at the hot vent Loki Castle on the Mohn's Ridge (Sweetman et al., 2013). At the recently investigated Lofoten canyon seep site dense aggregations of unidentified rissoids were observed from ROV (Sen et al., 2019a). Based on the details available from the published photo, we suggest that the gastropods are very likely to belong to genus *Frigidalvania*, based on the shell shape and rusty-brownish periostracum (Sen et al., 2019a, see Fig. 4b). Unfortunately, in our study we were not able to identify the behavior or lifestyle of *Frigidalvania* sp. This species remained unnoticed in the video data due to its small size (Baranov et al., 2019). However, multiple bacterial mats

observed from video-transects and caught by box corer provide an opportunity for such species to graze on them (Savvichev et al., 2018; Baranov et al., 2019). Other species common at seep sites and lacking in the background was the thyasirid bivalve *Axinopsida orbiculata* (Fig. 7d). Some species of thyasirids are known as symbiotrophic, however, the information on symbiotic bacteria in the gills of *A. orbiculata* is controversial: Zhukova, Kharlamenko & Gebruk (1991) have demonstrated the presence of bacteria in bivalve specimens from the Kraternaya Bight, the Kuril Islands, whereas according to Dufour (2005) this species lacks bacterial symbionts. It is possible that *A. orbiculata* is attracted by increased food availability at seep sites, same as another bivalve, *Macoma calcarea*, which is also common in seep background areas (Fig. 5). Overall, no bivalves restricted to cold seeps are known so far in the Arctic with the exception of two large thyasirids recently described by few empty shells (Åström, Oliver & Carroll, 2017) and Pleistocene subfossils (e.g. *Archivesica* spp., Sirenko et al., 2004; Hansen et al., 2017). The subfossils suggest that previously the Arctic cold seeps (and possibly hydrothermal vents) were inhabited by richer fauna that became extinct after Quaternary glaciation.

Notable is the high density (>3600 ind. m^{-2}) of the dorvilleid polychaete *Ophryotrocha* sp. in one grab sample at *C15-seep b* station group (Supplementary 1 (Fig. 7c). At least 15 species of *Ophryotrocha* have been described from reducing habitats (Taboada et al., 2013; Salvo et al., 2014; Ravara et al., 2015), including two species considered as obligate for cold seeps in the Kagoshima Bay, Japan (Miura, 1997). On the other hand, many species of this genus are common in regular marine ecosystems including Arctic seas (Sirenko, 2001).

Another taxon common in reducing habitats is tanaid crustaceans (Tanaidacea). In our material three species were present (Supplementary 1), all widely distributed in the Arctic (Sirenko, 2001). The density of tanaids in our samples was low, although this taxon was reported in high densities from the Håkon Mosby (Gebruk et al., 2003) and the Vestnesa Ridge (Åström et al., 2018) with several species (described as new) restricted to the methane seep habitats (Błazewicz-Paszkowycz and Bamber, 2011). It seems likely that many species of tanaids remain unidentified and diversity in this taxon remains underestimated owing to difficulties of identification of these small crustaceans (summarized by Błazewicz-Paszkowycz & Bamber, 2011). The low number of tanaids in our samples could be a result of a too large sieve mesh size used onboard (see Materials & Methods). Tanaids commonly are < 0.5 mm in size and require a corresponding mesh size to be found (Pavithran et al., 2009).

Overall, considering grab and trawl data combined, all the seep-specific taxa were the same at both seep sites. The only exception is the polychaete *Ophryotrocha* sp., which could be missed from the *Oden* trawl sample due to the large sieve mesh size (Supplementary 1).

Presence of specific benthic communities at C15 and Oden

Up to now no distinct macrofaunal changes in response to methane seeps were reported from the Arctic Ocean at depths < 80 m. In general, at depths <200 m both hydrothermal vents and cold-seeps are usually colonized by a subset of the local fauna (Tarasov et al., 2005; Dando, 2010). Some species notable at shallow-water methane seeps belong to opportunistic taxa common in various reducing habitats. These include siboglinid polychaetes and thyasirid bivalves reported from Skagerrak, Kattegat, coastal areas of Florida, Japan, New Zealand, New Guinea etc. (Southward & Culter, 1986; Schmaljohann & Flügel, 1987; Schmaljohann et al., 1990; Malakhov, Obzhairov & Tarasov, 1992; Gebruk, 2002). The isotope data suggest that food sources of macrofauna at shallow-water methane seeps are largely photosynthesis-based (Levin, 2005). It was suggested that the faunistic depth boundary between the deep-sea and shallow-water vents and seeps at approximately 200 m is controlled by the amount of organic matter input from the photosynthetic production (decreasing below the photic zone) and the greater number of predators at shallow depths. Definite seep-obligate species were not reported from depths <200 m (Tarasov et al., 2005; Dando, 2010).

At the same time, methane seep habitats even at shallow depths increase a number of microniches owing to increased organic matter availability, variety of substrates and repeated disturbance (Dando, 2010). Shallow cold-seeps may therefore support greater species diversity compared to the background. In our study at both methane seep sites, *C15* and *Oden*, integral community characteristics were significantly different from those in non-seep areas, among other things owing to presence of species obligate for reducing habitats. In addition, the communities found at *C15* site formed several station groups and were more scattered at the n-MDS plot (Fig. 2A, B) which could indicate a larger diversity of microniches within this site. Large amount of filter-feeders (Hydrozoa and Bryozoa) found in *C15*-trawl indicates the presence of hard substrata (including carbonate crusts). The larger amount of microniches is partly supported by

the video-data, where the landscape within the active seepages was more complex than in non-seep areas (Flint et al., 2018; Baranov et al., 2019).

Unfortunately, no environmental data except for the echo-sounding showing certain gas flares and CTD-measurements obtained from the area of the seeps from two points away from the benthic samples are available (Flint et al., 2018; Baranov et al., 2019). Nevertheless, we suggest that response of macrofauna to methane seeps at shallow depths 60-70 m can be related to very low primary productivity on the outer shelf of the Laptev Sea, dropping from ~720 mg C m⁻² per day at Lena river delta to <100 mg C m⁻² per day at 600 km (Sorokin & Sorokin, 1996) during September. Outside short Arctic summer months, these values tend to zero. In these extremely oligotrophic conditions, methane as a source of energy for the methane-oxidizing bacteria stimulates the development of local patchy benthic communities even at a shallow depth. As a comparison, the specific communities with siboglinids around Svalbard located at similar latitude are developed only at depths >200 m (Åström et al., 2016). Unlike the Laptev Sea shelf, the primary production south and west off Svalbard reaches much higher values up to 1800 mg C m⁻² per day during May blooms (Wassman et al., 2006). Furthermore, the Barents Sea remains uncovered with ice during most of the year, while the Laptev Sea shelf is ice-free during one to two months annually.

Conclusions

Our study is the first description of shallow-water methane seep communities in the Siberian Arctic. On the northern Laptev Sea shelf, significant differences were found between two methane seep sites (*C15* and *Oden*, located at depths of 63-73 m) and the background areas. The differences included integral community parameters and presence at seep sites of species typical for reducing habitats, such as siboglinids *Oligobrachia* sp. and thyasirid bivalves. Several species at methane seeps are presumably new to science, including the gastropod *Frigidalvania* sp. and the polychaete *Ophryotrocha* sp., found in large quantities at *C15* site. We suggest that response of macrofauna to methane seeps at shallow depths is related to very low primary productivity on the outer shelf of the Laptev Sea.

Acknowledgements

The authors would like to thank the Captain, crew members and shipboard parties of the RV *Akademik Mstislav Keldysh* for multiple help with the work onboard during the 63, 69 and 72 expeditions. Our special thanks to Dr. Michael Flint for organizing the expeditions and for informative discussions. We also thank Dr. Alexey Udalov and Dr. Elena Krylova for help in identifying mollusks and crustaceans. This work was partly funded by RFBR Grants 18-04-00206, 18-05-60053, 18-05-60228 and the State assignment of IORAS (theme No 0149-2019-0009).

References

1. Åström, E. K., Carroll, M. L., Ambrose Jr, W. G., & Carroll, J. (2016). Arctic cold seeps in marine methane hydrate environments: impacts on shelf macrobenthic community structure offshore Svalbard. *Marine Ecology Progress Series*, 552, 1-18.
2. Åström, E. K., Oliver, P. G., & Carroll, M. L. (2017). A new genus and two new species of Thyasiridae associated with methane seeps off Svalbard, Arctic Ocean. *Marine Biology Research*, 13(4), 402-416.
3. Åström, E. K., Carroll, M. L., Ambrose Jr, W. G., Sen, A., Silyakova, A., & Carroll, J. (2018). Methane cold seeps as biological oases in the high-Arctic deep sea. *Limnology and Oceanography*, 63(S1), S209-S231.
4. Azovsky, A.I. (2018) Analysis of long-term biological data series: methodological problems and possible solutions. *J Gene Biol (Moscow)* 79:329–341 [in Russian]
5. Baker, E. T., & German, C. R. (2004). On the global distribution of hydrothermal vent fields. *Mid-Ocean Ridges: Hydrothermal Interactions Between the Lithosphere and Oceans*, Geophys. Monogr. Ser, 148, 245-266.
6. Baranov B., Galkin S., Vedenin A., Dozorova K., Gebruk A., Flint M. 2019. Methane seeps on the outer shelf of the Laptev Sea: characteristic features, benthic fauna and structural control. *Geophysical Research Letters*. [in press]
7. Błażewicz-Paszkowycz, M., & Bamber, R. N. (2011). Tanaidomorph Tanaidacea (Crustacea: Peracarida) from mud-volcano and seep sites on the Norwegian Margin. *Zootaxa*, 3061, 1-35.
8. Boetius, A. (2015). The expedition PS86 of the Research Vessel POLARSTERN to the Arctic Ocean in 2014. *Berichte zur Polar-und Meeresforschung = Reports on polar and marine research*, 685, 1-132.
9. Brotskaya, V., Zenkevich, L., 1939. Quantitative account of the Barents Sea benthic fauna. *Trudy VNIRO*, 4, 55-120. [in Russian]
10. Buzhinskaja, G. N. (2010). Illustrated keys to free-living invertebrates of Eurasian Arctic seas and adjacent deep waters. Nemertea, Cephalorincha, Oligochaeta, Hirudinea, Pogonophora, Echiura, Sipuncula, Phoronida and Brachiopoda, 2. 186 p.
11. Clarke, K.R., Warwick, R.M., 2001. *Changes in Marine Communities: An Approach to Statistical Analysis and Interpretation* (2nd ed.). PRIMER-E, Plymouth
12. Clarke, K. R., Gorley, R. N. (2015). *Getting started with PRIMER v7. PRIMER-E: Plymouth, Plymouth Marine Laboratory.*
13. Dando, P.R., 2010. Biological communities at marine shallow-water vent and seep sites. In: Kiel S. (Ed.) *The Vent and Seep Biota*, Springer, Dordrecht, pp. 333-378.
14. Van Dover, C. (2000). *The ecology of deep-sea hydrothermal vents*. Princeton University Press.
15. Decker C., & Olu K. (2012). Habitat heterogeneity influences cold-seep macrofaunal communities within and among seeps along the Norwegian margin – Part 2: contribution of chemosynthesis and nutritional patterns. *Marine Ecology*, 33(2), 231-245.
16. Dufour, S. C. (2005). Gill anatomy and the evolution of symbiosis in the bivalve family Thyasiridae. *The Biological Bulletin*, 208(3), 200-212.
17. Eleftheriou A., McIntyre A. (2005). *Methods for the study of Marine benthos*. 3rd edn. Blackwell Science, Oxford, UK. 418 p.

18. Flint M., Arashkevich E., Artemyev V., Baranov B., Bezzubova E., Belevich T., ... & Shchuka S.A. Ecosystems of seas of Siberian Arctic. Materials of expeditionary research of 2015 and 2017. P.P. Shirshov Institute of oceanology, RAS. Moscow: 2018. 232 p. [in Russian]
19. Galkin, S. V., & Vedenin, A. A. (2015). Macrobenthos of Yenisei Bay and the adjacent Kara Sea shelf. *Oceanology*, 55(4), 606-613.
20. Gebruk A.V. Biology of hydrothermal vents. Moscow, KMK-Press, 2002. 543 p. [in Russian]
21. Gebruk, A. V., Krylova, E. M., Lein, A. Y., Vinogradov, G. M., Anderson, E., Pimenov, N. V., ... & Crane, K. (2003). Methane seep community of the Håkon Mosby mud volcano (the Norwegian Sea): composition and trophic aspects. *Sarsia*, 88(6), 394-403.
22. Hammer, Ø. 2013. PAST. Paleontological Statistics. Version 3.10. Reference manual. Natural History Museum. University of Oslo.
23. Hansen, J., Hoff, U., Szttybor, K., & Rasmussen, T. L. (2017). Taxonomy and palaeoecology of two Late Pleistocene species of vesicomyid bivalves from cold methane seeps at Svalbard (79° N). *Journal of Molluscan Studies*, 83(3), 270-279.
24. Hessler, R.R., Kaharl, V.A., 1995. The deep-sea hydrothermal vent community: an overview. Seafloor hydrothermal systems. In: Humphris, S.E., Zierenberg, R.A., Mullineaux, L.S., Thomson, R.E. (Eds.), *Geophysical Monograph*, vol. 91. American Geo-physical Union, pp. 72-84.
25. Lee, D. H., Kim, J. H., Lee, Y. M., Jin, Y. K., Paull, C., Kim, D., & Shin, K. H. (2019). Chemosynthetic bacterial signatures in Frenulata tubeworm *Oligobrachia* sp. in an active mud volcano of the Canadian Beaufort Sea. *Marine Ecology Progress Series*, 628, 95-104.
26. Levin, L. A. (2005). Ecology of cold seep sediments: interactions of fauna with flow, chemistry and microbes. In *Oceanography and Marine Biology* (pp. 11-56). CRC Press.
27. Lobkovsky, L. I., Nikiforov, S. L., Ananiev, R. A., Khortov, A. V., Semiletov, I. P., Jakobsson, M., & Dmitrievskiy, N. N. (2015). Recent geological-geomorphological processes on the east Arctic shelf: Results of the expedition of the icebreaker Oden in 2014. *Oceanology*, 55(6), 926-929.
28. Malakhov, V. V., Obzhairov, A. I., & Tarasov, V. G. (1992). Pogonophoran genus *Siboglinum* in relation to zones of high concentrations of methane. *Doklady Akademii Nauk*, 135, 195-197. [in Russian]
29. McCune, B., Grace, J.B., Urban, D.L., 2002. Analysis of Ecological Communities. Vol. Mjmm Software Design., Gleneden Beach, Oregon, p. 28.
30. Miura, T. (1997). Two new species of the genus *Ophryotrocha* (Polychaeta, Iphitimiidae) from Kagoshima Bay. *Bulletin of Marine Science*, 60(2), 300-305.
31. Paull, C. K., Dallimore, S. R., Caress, D. W., Gwiazda, R., Melling, H., Riedel, M., ... & Sherman, A. (2015). Active mud volcanoes on the continental slope of the Canadian Beaufort Sea. *Geochemistry, Geophysics, Geosystems*, 16(9), 3160-3181.
32. Pavithran, S., Ingole, B., Nanajkar, M., & Goltekar, R. (2009). Importance of sieve size in deep-sea macrobenthic studies. *Marine Biology Research*, 5(4), 391-398.
33. Ravara, A., Marçal, A. R., Wiklund, H., & Hilário, A. (2015). First account on the diversity of *Ophryotrocha* (Annelida, Dorvilleidae) from a mammal-fall in the deep-Atlantic Ocean with the description of three new species. *Systematics and biodiversity*, 13(6), 555-570.
34. Rodrigues, C. F., Hilário, A., Cunha, M. R., Weightman, A. J., & Webster, G. (2011). Microbial diversity in Frenulata (Siboglinidae, Polychaeta) species from mud volcanoes in the Gulf of Cadiz (NE Atlantic). *Antonie Van Leeuwenhoek*, 100(1), 83-98.

35. Rybakova, E., Galkin, S., Bergmann, M., Soltwedel, T., & Gebruk, A. (2013). Density and distribution of megafauna at the Håkon Mosby mud volcano (the Barents Sea) based on image analysis. *Biogeosciences*, 10, 3359-3374.
36. Salvo, F., Wiklund, H., Dufour, S. C., Hamoutene, D., Pohle, G., & Worsaae, K. (2014). A new annelid species from whalebones in Greenland and aquaculture sites in Newfoundland: *Ophryotrocha cyclops*, sp. nov. (Eunicida: Dorvilleidae). *Zootaxa*, 3887(5), 555-568.
37. Savvichev A.S., Rusanov I.I., Yusupov S.K., Pimenov N.V., Lein A.Y., & Ivanov M.V. 2004. The biogeochemical cycle of methane in the coastal zone and littoral of the Kandalaksha Bay of the White Sea. *Microbiology*, 73(4), 457-468.
38. Savvichev, A. S., Kadnikov, V. V., Kravchishina, M. D., Galkin, S. V., Novigatskii, A. N., Sigalevich, P. A., ... & Flint, M. V. (2018). Methane as an organic matter source and the trophic basis of a Laptev Sea cold seep microbial community. *Geomicrobiology journal*, 35(5), 411-423.
39. Schmaljohann, R., & Flügel, H. J. (1987). Methane-oxidizing bacteria in Pogonophora. *Sarsia*, 72(1), 91-98.
40. Schmaljohann, R., Faber, E., Whitticar, M. J., & Dando, P. R. (1990). Co-existence of methane-and sulphur-based endosymbioses between bacteria and invertebrates at a site in the Skagerrak. *Marine Ecology Progress Series*, 61, 119-124.
41. Sen A., Duperron S., Hourdez S., Piquet B., Le'ger N., Gebruk A., Le Port A.S., Svenning M.M., & Andersen A.C. (2018) Cryptic frenulates are the dominant chemosymbiotrophic fauna at Arctic and high latitude Atlantic cold seeps. *PLoS ONE*, 13(12): e0209273.
42. Sen A., Himmler T., Hong W.L., Chitkara C., Lee R.W., Ferré B., ... & Knies J. (2019a). Atypical biological features of a new cold seep site on the Lofoten-Vesterålen continental margin (northern Norway). *Scientific reports*, 9(1), 1762.
43. Sen, A., Chitkara, C., Hong, W. L., Lepland, A., Cochrane, S., di Primio, R., & Brunstad, H. (2019b). Image based quantitative comparisons indicate heightened megabenthos diversity and abundance at a site of weak hydrocarbon seepage in the southwestern Barents Sea. *PeerJ*, 7, e7398.
44. Shakhova, N., Semiletov, I., Sergienko, V., Lobkovsky, L., Yusupov, V., Salyuk, A., ... & Nicolsky, D. (2015). The East Siberian Arctic Shelf: towards further assessment of permafrost-related methane fluxes and role of sea ice. *Philosophical Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences*, 373(2052), 20140451.
45. Sirenko, B. I. (2001). List of species of free-living invertebrates of Eurasian Arctic seas and adjacent deep waters. *Explorations of the fauna of the seas*, 51(59), 5-131.
46. Sirenko B., Denisenko S., Deubel H., & Rachor E. (2004). Deep water communities of the Laptev Sea and adjacent parts of the Arctic Ocean. *Fauna and the ecosystems of the Laptev Sea and adjacent deep waters of the Arctic Ocean. Explorations of the fauna of sea*, 54(62), 28-73.
47. Smirnov, R. V. (2000). Two new species of Pogonophora from the arctic mud volcano off northwestern Norway. *Sarsia*, 85(2), 141-150.
48. Sorokin, Y. I., & Sorokin, P. Y. (1996). Plankton and primary production in the Lena River estuary and in the south-eastern Laptev Sea. *Estuarine, Coastal and Shelf Science*, 43(4), 399-418.
49. Southward, E. C., & Culter, J. K. (1986). Discovery of Pogonophora in warm shallow waters of the Florida Shelf. *Marine Ecology Progress Series*, 28, 287-89.

50. Sweetman, A. K., Levin, L., Rapp, H. T., & Schander, C., 2013. Faunal trophic structure at hydrothermal vents on the southern Mohn's Ridge, Arctic Ocean. *Marine Ecology Progress Series*, 473, 115-131.
51. Taboada, S., Wiklund, H., Glover, A. G., Dahlgren, T. G., Cristobo, J., & Avila, C. (2013). Two new Antarctic *Ophryotrocha* (Annelida: Dorvilleidae) described from shallow-water whale bones. *Polar Biology*, 36(7), 1031-1045.
52. Tarasov, V. G., Gebruk, A. V., Mironov, A. N., & Moskalev, L. I. (2005). Deep-sea and shallow-water hydrothermal vent communities: two different phenomena? *Chemical Geology*, 224(1-3), 5-39.
53. Vedenin, A. A., Galkin, S. V., & Kozlovskiy, V. V. (2015). Macrobenthos of the Ob Bay and adjacent Kara Sea shelf. *Polar Biology*, 38(6), 829-844.
54. Vedenin, A., Mokievsky, V., Soltwedel, T., & Budaeva, N. (2019). The temporal variability of the macrofauna at the deep-sea observatory HAUSGARTEN (Fram Strait, Arctic Ocean). *Polar Biology*, p. 1-14. doi.org/10.1007/s00300-018-02442-8.
55. Wassmann, P., Reigstad, M., Haug, T., Rudels, B., Carroll, M. L., Hop, H., ... & Slagstad, D. (2006). Food webs and carbon flux in the Barents Sea. *Progress in Oceanography*, 71(2-4), 232-287.
56. Yusupov, V. I., Salyuk, A. N., Karnaukh, V. N., Semiletov, I. P., & Shakhova, N. E. (2010). Detection of methane ebullition in shelf waters of the Laptev Sea in the Eastern Arctic Region. In: *Doklady Earth Sciences* (Vol. 430, No. 2, pp. 261-264). MAIK Nauka/Interperiodica.
57. Zhukova, N.V., V.I. Kharlamenko & Gebruk A.V., 1991. Fatty acids of *Axinopsida orbiculata* - potential for detection of symbiosis with chemoautotrophic bacteria. In, *Shallow Water Vents and Ecosystem of Kraternaya Bight (Volcano Ushishir. Kurile Islands)*, Vol. 1, Functional Characteristics, Part 11, Vladivostok, in press. [In Russian].

Table 1 (on next page)

Data on stations used in the present study

For trawl stations coordinates and depth of start and end are given.

Table 1. Data on stations used in the present study. For trawl stations coordinates and depth of start and end are given.

Expedition, year	Station	Site	Gear	Latitude	Longitude	Depth (m)
AMK-63, 2015	seep-1	C15	Okean-0.1	76°46.376' N	125°49.641' E	72.0
	seep-2	C15	Okean-0.1	76°46.376' N	125°49.664' E	72.3
	seep-3	C15	Okean-0.1	76°46.379' N	125°49.618' E	72.4
	background-1	C15	Okean-0.1	76°46.375' N	125°50.346' E	73.0
	background-2	C15	Okean-0.1	76°46.366' N	125°50.366' E	73.0
	background-3	C15	Okean-0.1	76°46.365' N	125°50.339' E	73.0
	C15 trawl	C15	Sigsbee	76°46.483' N 76°46.447' N	125°50.843' E 125°48.231' E	71.5 72.0
AMK-69, 2017	5623-1	Oden	Van Veen-0.1	76°53.624' N	127°48.110' E	63.0
	5623-2	Oden	Van Veen-0.1	76°53.608' N	127°48.114' E	63.1
	5623-3	Oden	Van Veen-0.1	76°53.632' N	127°48.219' E	63.0
	Oden trawl	Oden	Sigsbee	76°53,667' N	127°48,157' E	63.0
				76°53,566' N	127°49,075' E	63.0
	5624-1	Control	Van Veen-0.1	76°49.998' N	126°39.936' E	69.6
	5624-2	Control	Van Veen-0.1	76°50.003' N	126°39.896' E	69.7
	5624-3	Control	Van Veen-0.1	76°49.883' N	126°40.000' E	69.6
	5625-1	C15	Van Veen-0.1	76°46.438' N	125°49.417' E	70.8
	5625-2	C15	Van Veen-0.1	76°46.435' N	125°49.442' E	70.7
	5625-3	C15	Van Veen-0.1	76°46.413' N	125°49.437' E	70.6
AMK-72, 2018	5947-1	C15	Van Veen-0.1	76°46.847' N	125°49.085' E	72.3
	5947-2	C15	Van Veen-0.1	76°46.847' N	125°49.085' E	72.0
	5947-3	C15	Van Veen-0.1	76°46.848' N	125°49.097' E	72.0
	5953-1	Oden	Van Veen-0.1	76°53.554' N	127°48.405' E	63.0
	5953-2	Oden	Van Veen-0.1	76°53.551' N	127°48.409' E	63.0
	5953-3	Oden	Van Veen-0.1	76°53.567' N	127°48.400' E	63.0

Table 2 (on next page)

Results of the Kruskal-Wallis and Tukey's post-hoc tests for taxa with different abundance values in five station groups.

Mean abundance in each station group is shown. Taxa are arranged according to p -value. Pairs in post-hoc column indicate significant comparisons (Tukey's $p < 0.05$).

1 - *Control* group; 2 - *C15 background* group; 3 - *Oden* group; 4 - *C15-seep a* group; 5 - *C15-seep-b* group.

Table 2. Results of the Kruskal-Wallis and Tukey's post-hoc tests for taxa with different abundance values in five station groups. Mean abundance in each station group is shown. Taxa are arranged according to *p*-value. Pairs in post-hoc column indicate significant comparisons (Tukey's *p* < 0.05).

Species	Mean abundance in station groups					Kruskal-Wallis		Post-hoc
	1	2	3	4	5	<i>H</i> (chi ²)	<i>p</i>	
<i>Oligobranchia</i> sp.	0	0	63.0	14.6	53.5	15.86	0.00211	no values
<i>Yoldiella lenticula</i>	0.7	12.6	1.0	8.8	2.0	15.32	0.00362	2-1; 2-3; 2-5
<i>Cistenides hyperborea</i>	0	0.4	0.2	4.4	5.5	13.14	0.00378	no values
<i>Tharyx</i> sp.	2.3	6.0	32.5	3.8	2.5	14.70	0.00511	3-1; 3-2; 3-4; 3-5
<i>Spiochaetopterus typicus</i>	0	0	3.0	0.6	0	9.85	0.01151	no values
<i>Yoldiella solidula</i>	9.3	44.0	23.5	19.6	5.0	12.51	0.01391	no values
<i>Cossura longocirrata</i>	0	0	0.2	0.6	199.0	6.40	0.02177	no values
<i>Ophiosten sericeum</i>	0.3	3.8	6.3	4.8	26.5	10.38	0.03252	4-1; 4-2; 4-3; 4-5
<i>Portlandia arctica</i>	0	14.6	0	0.8	0.5	7.40	0.03257	no values
<i>Pleusymtes pulchella</i>	0	0	0.5	1.2	3.5	7.273	0.03463	no values
<i>Frigidaltia</i> sp.	0	0	0	59.8	29.5	5.15	0.05565	no values
<i>Anobothrus gracilis</i>	0	0	1.2	0.2	0	5.06	0.05839	no values
<i>Axinopsida orbiculata</i>	0	1.6	9.2	2.8	14.5	6.36	0.12860	no values
<i>Paroediceros lynceus</i>	0	0	0	0.8	4.0	2.57	0.13980	no values
<i>Haploops tubicola</i>	0.3	1.4	0.2	0	0	3.83	0.14310	no values

1 – Control group; 2 – C15 background group; 3 – Oden group; 4 – C15-seep a group; 5 – C15-seep-b group.

Table 3(on next page)

Likelihood of not finding a species calculated for species present only at methane seep sites and only at the *Oden* site.

¹ - Species absent not accidentally.

Table 3. Likelihood of not finding a species calculated for species present only at methane seep sites and only at the *Oden* site.

Species	Species occurrence	Likelihood of not finding	Number of grabs	
			Required for finding ($\alpha = 0.99$)	Taken
Species present at C15-seep a, C15-seep b and Oden and absent at C15 background and Control sites				
<i>Spiochaetopterus typicus</i> ¹	0.62	0.000	4.8	8
<i>Cossura longocirrata</i>	0.38	0.021	9.5	8
<i>Anobothrus gracilis</i>	0.38	0.021	9.5	8
<i>Oligobranchia</i> sp. ¹	1.00	0.000	1.0	8
<i>Axinopsida orbiculata</i> ¹	0.77	0.000	3.1	8
<i>Paroedicerus lynceus</i>	0.23	0.123	17.6	8
<i>Pleusymtes pulchella</i> ¹	0.53	0.002	6.0	8
Species present at Oden and absent at C15-seep a and C15-seep b				
<i>Frigidaltvania</i> sp. ¹	0.57	0.006	5.4	6
<i>Portlandia arctica</i>	0.43	0.035	8.2	6
<i>Paroedicerus lynceus</i>	0.43	0.035	8.2	6

¹ - Species absent not accidentally

Table 4(on next page)

Species richness, Pielou evenness, Hurlbert rarefaction for 100 individuals and Shannon-Wiener index calculated for trawl samples.

1 Table 4. Species richness, Pielou evenness, Hurlbert rarefaction for 100 individuals and Shannon-Wiener
2 index calculated for trawl samples.

Trawl	Species richness	Pielou evenness	ES (100)	Shannon-Wiener index
<i>C15</i>	203	0.55	29.97	2.92
<i>Oden</i>	167	0.56	33.02	2.86

3

Table 5(on next page)

Similarity percentage routine for trawl samples. Species with contribution >0.5% are shown.

Species more abundant at *C15* are marked with bold.

1 Table 5. Similarity percentage routine for trawl samples. Species with contribution >0.5% are shown.

Species	Abundance (%)		Average dissimilarity	Contribution, %	Cumulative, %
	<i>C15</i>	<i>Oden</i>			
<i>Frigidavalvania</i> sp.	12.05	2.37	4.84	14.06	14.06
<i>Ophiocten sericeum</i>	37.32	45.55	4.11	11.94	26.00
<i>Yoldiella lenticula</i>	7.37	1.11	3.13	9.08	35.08
<i>Yoldiella solidula</i>	7.82	10.89	1.54	4.46	39.55
<i>Portlandia arctica</i>	2.56	0.15	1.21	3.51	43.05
<i>Laona finmarchica</i>	1.60	0.00	0.80	2.32	45.38
<i>Phascolion strombus</i>	1.95	0.36	0.80	2.31	47.69
<i>Myriochele heeri</i>	0.42	1.82	0.70	2.03	49.72
<i>Micronephthys minuta</i>	0.16	1.47	0.65	1.90	51.62
<i>Craniella polyura</i>	1.30	0.00	0.65	1.88	53.50
<i>Pholoe longa</i>	1.38	2.62	0.62	1.79	55.29
<i>Munnopsis typica</i>	0.67	1.84	0.59	1.70	56.99
<i>Scoletoma fragilis</i>	0.28	1.17	0.45	1.30	58.30
<i>Paraedicerus lynceus</i>	1.80	0.92	0.44	1.28	59.58
<i>Rostroculodes hansenii</i>	0.00	0.88	0.44	1.28	60.85
<i>Nothria hyperborea</i>	0.10	0.90	0.40	1.16	62.01
<i>Solariella obscura</i>	0.90	0.13	0.39	1.13	63.14
<i>Tharyx</i> sp.	0.04	0.67	0.31	0.91	64.05
<i>Axinopsida orbiculata</i>	0.00	0.61	0.30	0.88	64.93
<i>Brada villosa</i>	0.66	0.06	0.30	0.86	65.79
<i>Arrhis phyllonyx</i>	0.25	0.82	0.28	0.83	66.62
<i>Terebellides</i> aff. <i>stroemii</i>	0.86	1.42	0.28	0.82	67.44
<i>Similipecten</i>	0.68	0.15	0.27	0.78	68.22
<i>Cylichna occulta</i>	0.74	0.21	0.27	0.77	68.99
<i>Yoldiella frigida</i>	0.15	0.63	0.24	0.70	69.69
<i>Cossura longocirrata</i>	0.04	0.52	0.24	0.70	70.39
<i>Sabinea septemcarinata</i>	0.09	0.57	0.24	0.69	71.08
<i>Nymphon hirtipes</i>	0.10	0.57	0.23	0.67	71.76
<i>Cuspidaria glacialis</i>	0.80	0.38	0.21	0.61	72.37
<i>Brada incrustata</i>	0.04	0.46	0.21	0.61	72.98
<i>Lepidepecreum umbo</i>	0.42	0.02	0.20	0.58	73.56
<i>Ephesiella abyssorum</i>	0.01	0.40	0.19	0.56	74.12
<i>Nuculana pernula</i>	0.54	0.92	0.19	0.56	74.67
<i>Rozinante fragilis</i>	0.51	0.15	0.18	0.53	75.20
<i>Philine lima</i>	0.00	0.36	0.18	0.52	75.72
<i>Pleusymtes pulchellus</i>	0.36	0.71	0.17	0.51	76.22
<i>Owenia polaris</i>	0.07	0.42	0.17	0.50	76.72

2 Species more abundant at *C15* are marked with bold

Figure 1

Study area.

Enlarged maps show sampling sites and corresponding stations. Detailed bathymetry is only available for *C15* and *Oden* sites; white circles indicate previously recorded gas flares (Baranov et al., 2019). Dotted line at *Oden* site enclosed map shows the approximate perimeter of seeping area.

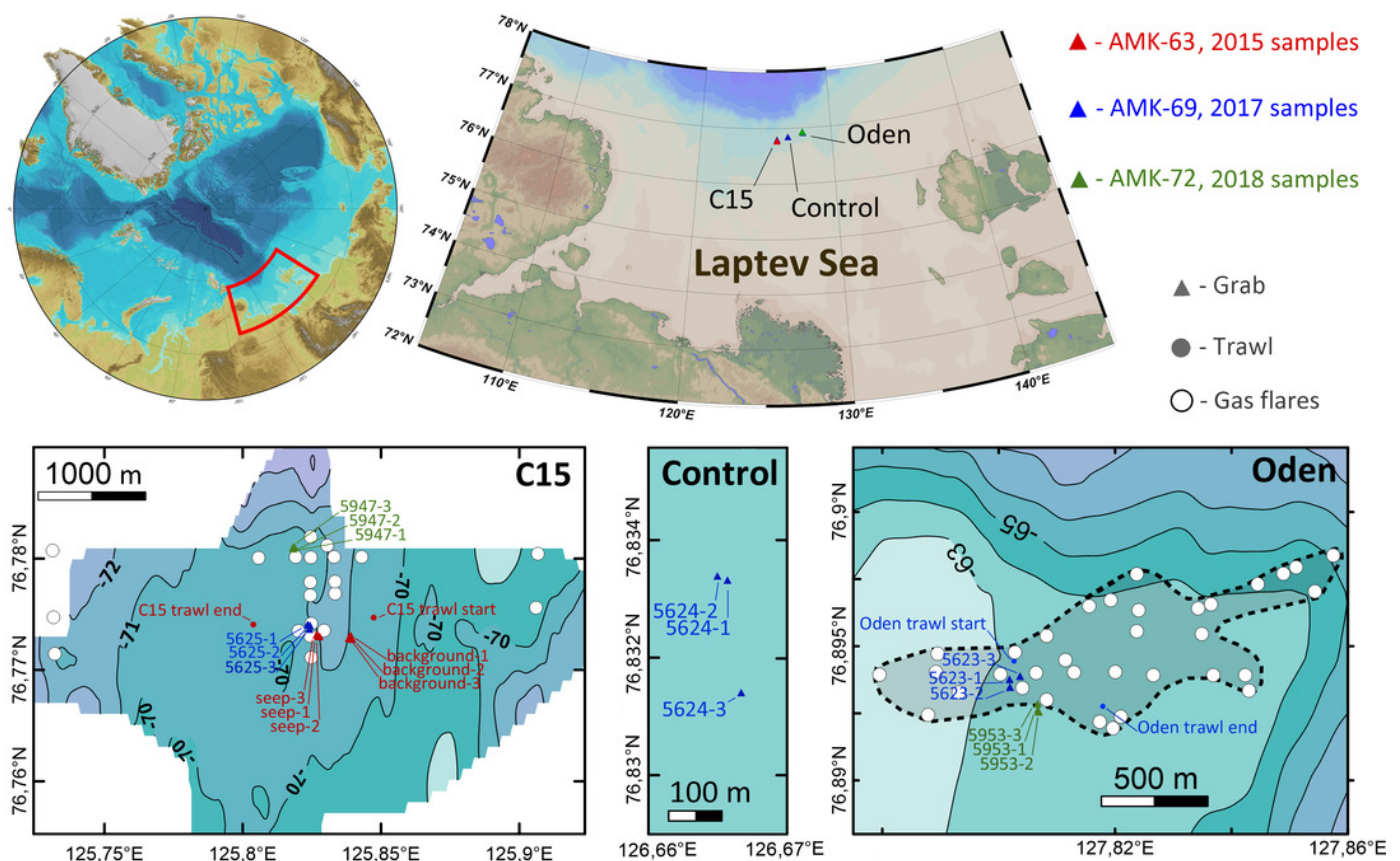


Figure 2

UNCTREE analysis with SIMPROF results (A) and non-metric multidimensional scaling plot (B) of grab stations using the Bray-Curtis similarity index.

Square root transformed biomass data are used. Dashed lines connect statistically unreliable groupings ($p > 0.05$). Green lines indicate SIMPROF groups.

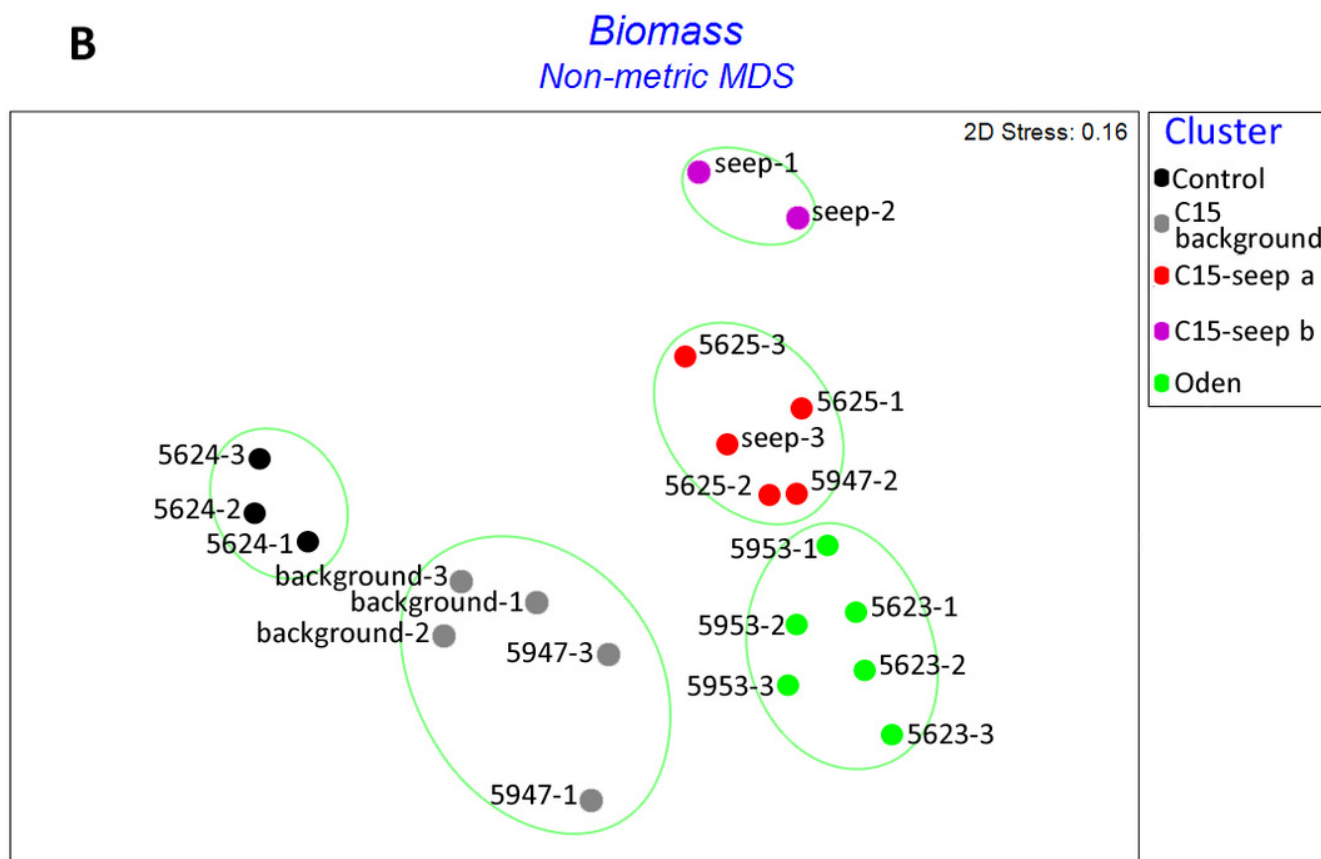
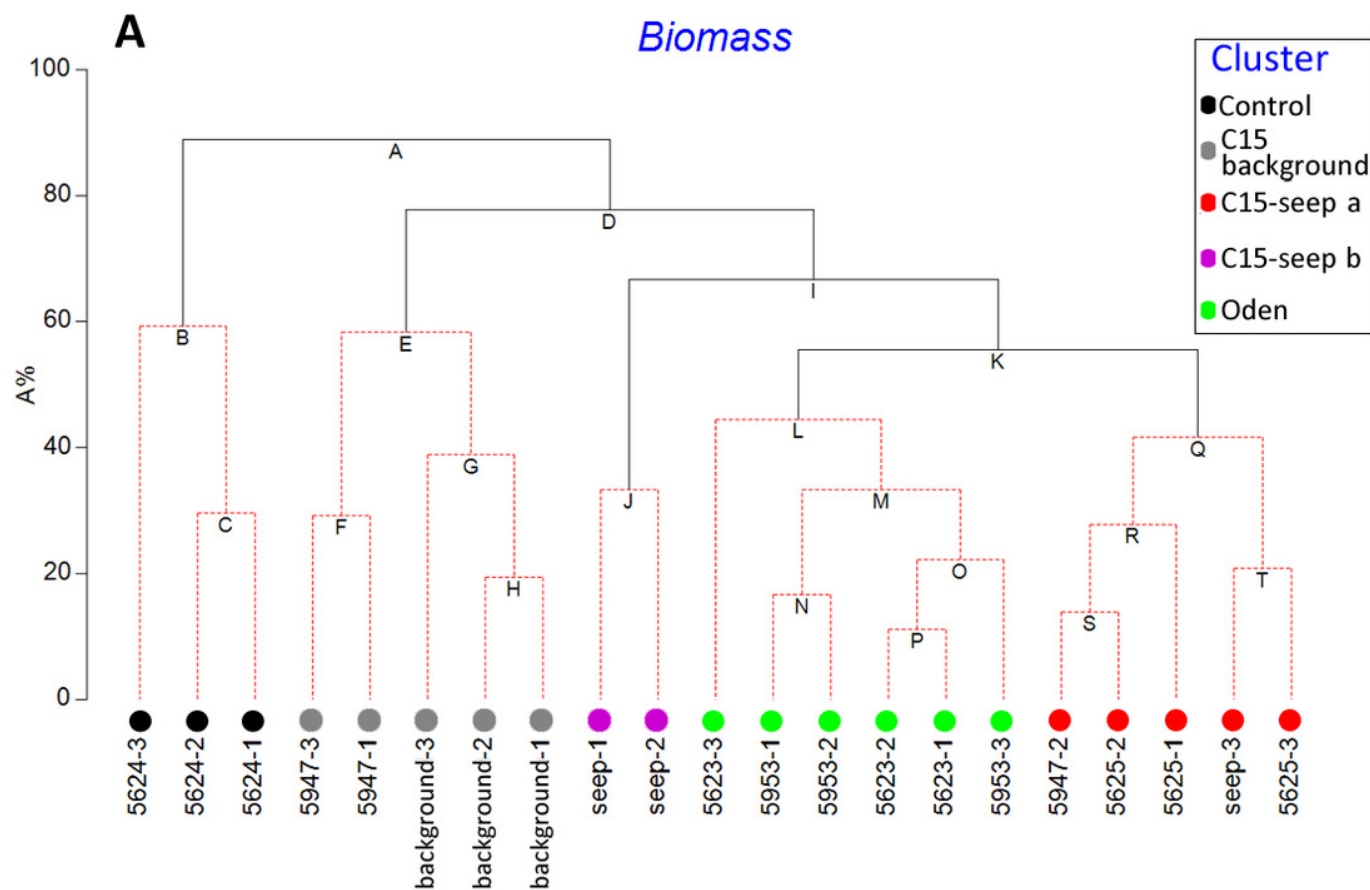


Figure 3

The species-individuals accumulation curves for the station groups.

Colors are the same as in Figure 2.

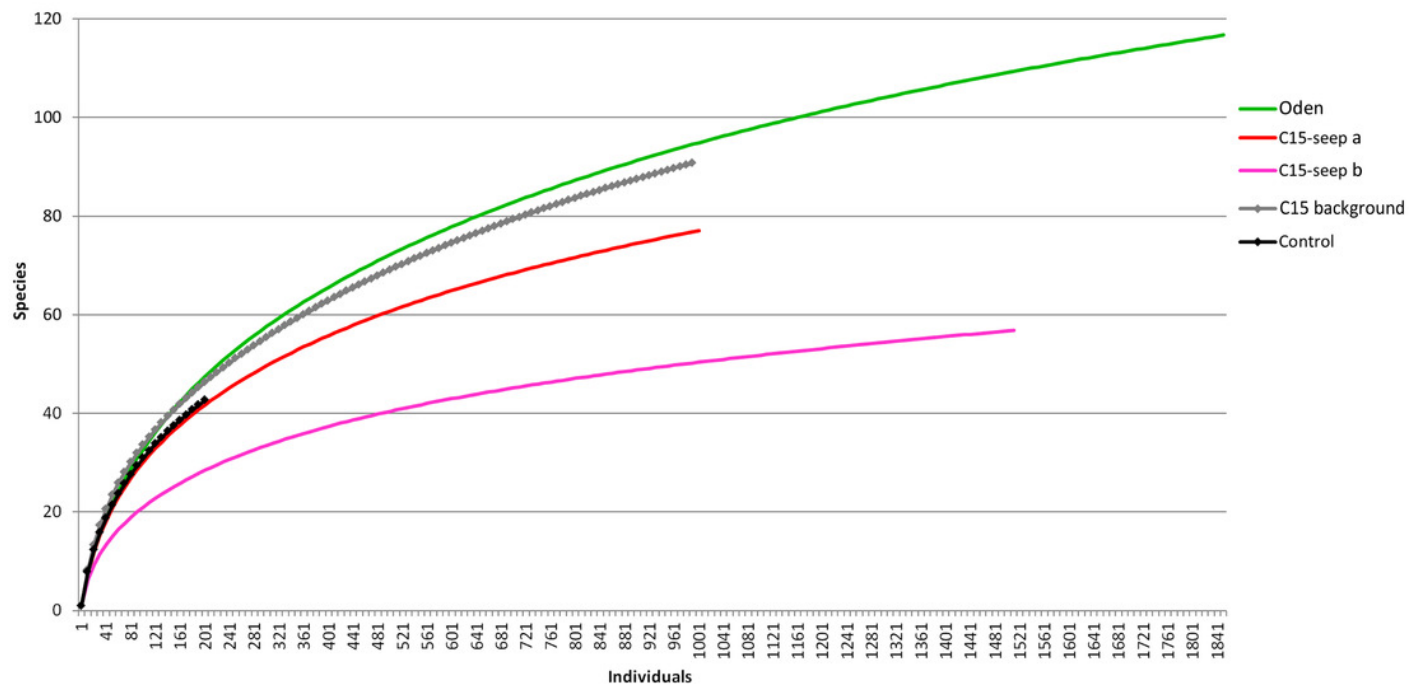


Figure 4

Univariate characteristics of identified clusters.

Mean values of total density, biomass, species richness, Pielou evenness, Hurlbert rarefaction index and Shannon-Wiener index with standard deviation are shown. Exact values of these characteristics are shown in Supplementary 3.

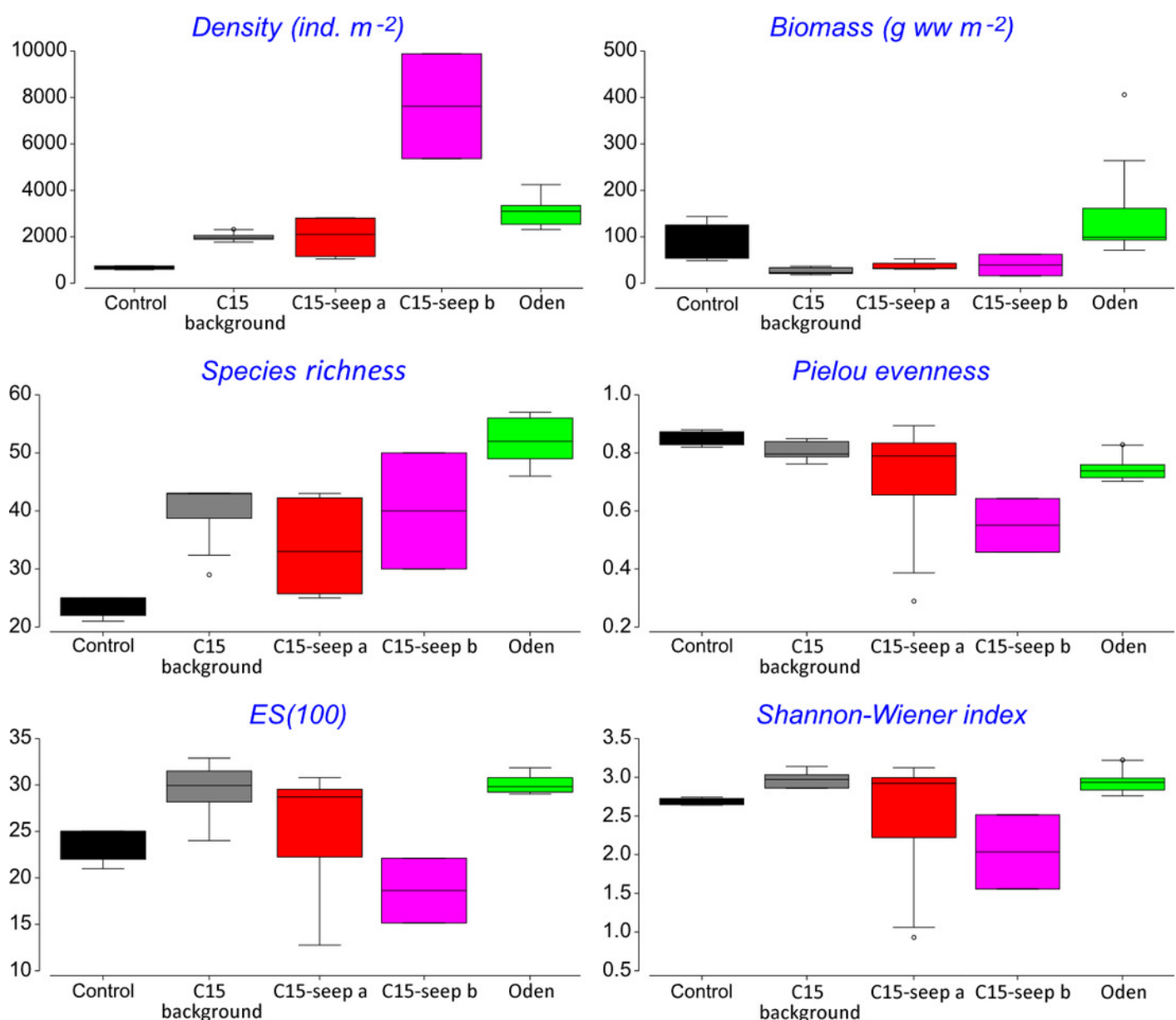


Figure 5

Shade plot of species square root transformed abundance (A) and biomass (B) at stations arranged by clusters.

The species list is reduced to 20 most important taxa. Order of stations and colors the same as in Figure 2. Taxa grouped in clusters using UPGMA algorithm based on index of association.

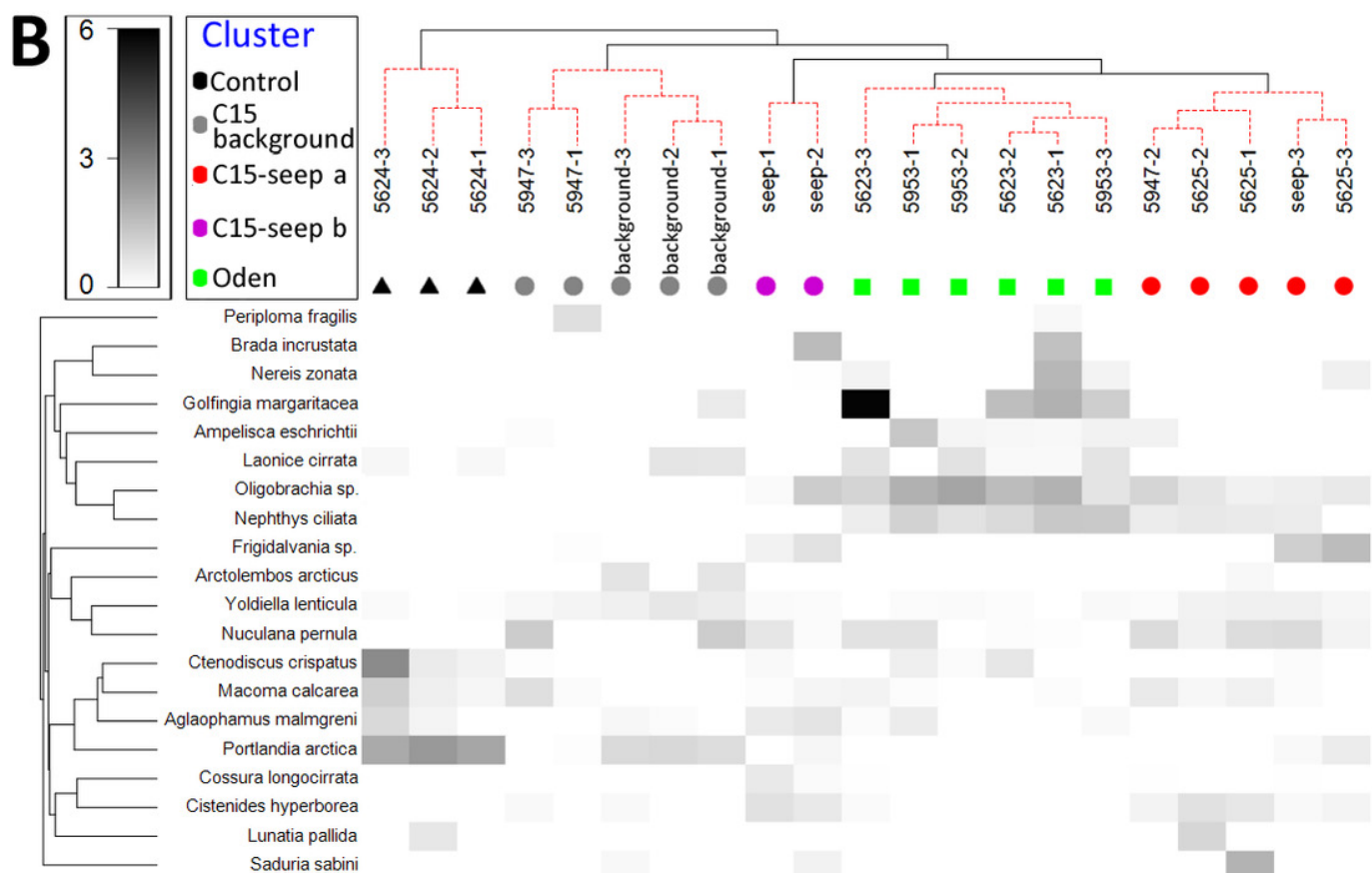
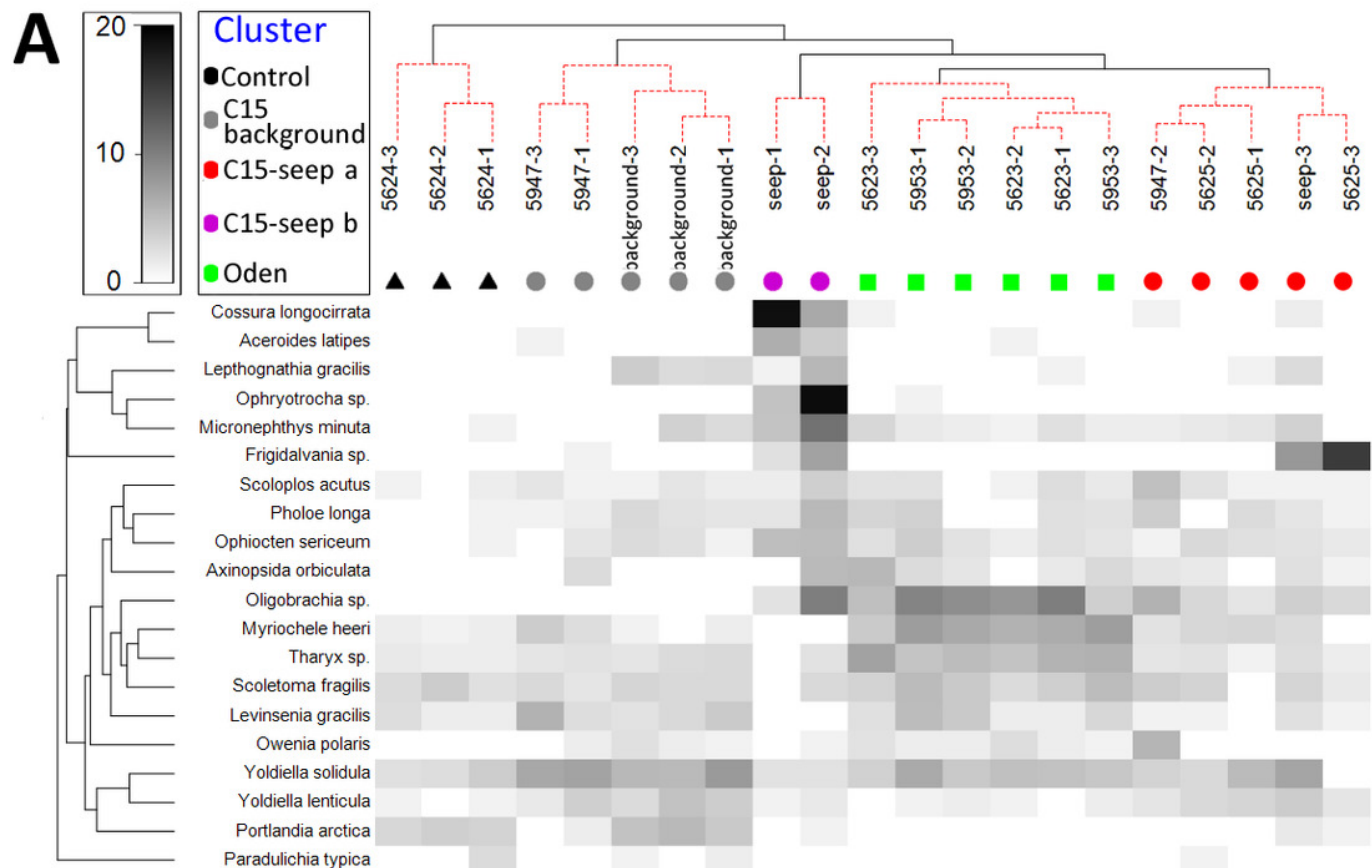


Figure 6

Species ranking for *C15* and *Oden* trawl samples.

The most numerous species are indicated. X-axis is logarithmic.

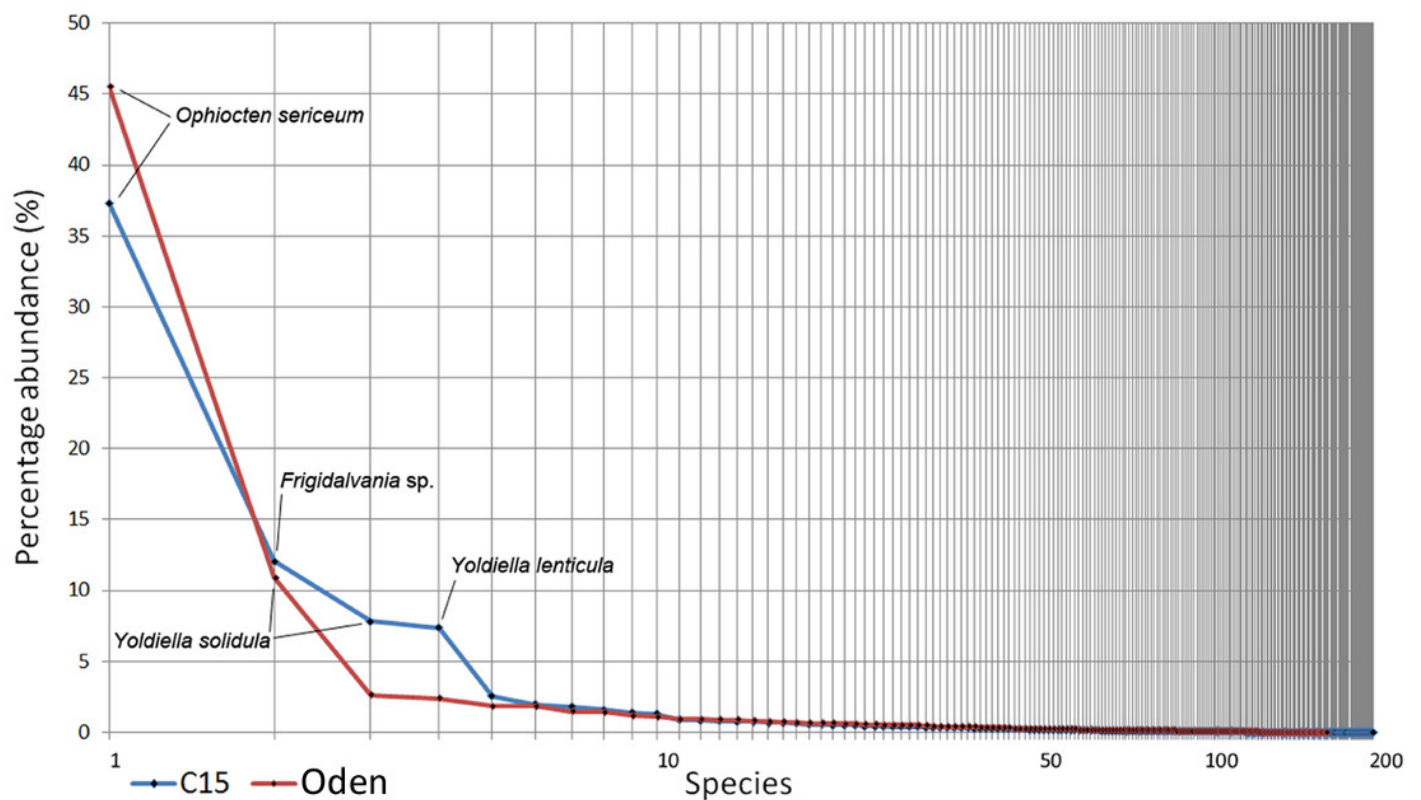


Figure 7

Taxa found only at seep stations.

A - *Oligobrachia* sp. (left - tube with several fragments enlarged; center - complete specimen extracted from tube; right - anterior and posterior fragments of the specimen); B - *Frigidalvania* sp.; C - *Ophryotrocha* sp. (upper left - several specimens, total view; upper right - anterior fragment; lower - enlarged parapodia); D - *Axinopsida orbiculata*. Photos by A. Vedenin and V. Kokarev.

