

Genome-wide identification of human genes involved in the evolution of human intelligence evolution through comparative genomics analysis combination of inter-species and intra-species genetic variations

Mengjie Li, Wenting Zhang, Xiaoyi Zhou*

College of Life Sciences, Shanghai Normal University, Shanghai, China

Corresponding Author:

Xiaoyi Zhou

No.100 Guilin Road, Shanghai, 200234, China

Email address: zhouxy@shnu.edu.cn

Abstract

Understanding the evolution of human intelligence ~~from great apes to humans~~ is an important undertaking in the science of human genetics. ~~Recently, a~~ great deal of biological research has been conducted to search for ~~the human-specific genes and variations that have resulted in which are related to~~ the significant increase in human ~~brain volume and cerebral cortex complexity intelligence over that of apes during the~~ hominid evolution. However, genetic changes affecting intelligence in hominid evolution have remained elusive. We supposed that a subset of intelligence-related genes, which harbored intra-species variations in human populations, may also be evolution-related genes which harbored inter-species variations between humans and the great apes. It is very important, yet extremely difficult, to discover additional genes ~~involved in the evolution of human intelligence and various approaches need to be taken to further explore the issue~~. We ~~designed a new strategy combined~~ inter-species and intra-species genetic variations to discover genes involved in the evolution of human intelligence. Information was collected from published GWAS works on intelligence and ~~from these~~ a total of 549 genes located within the intelligence-associated loci were identified. The intelligence-related genes containing ~~a human-specific variations~~ were detected based on the latest high-quality genome assemblies of three great apes. ~~Finally, we identified including~~ 40 strong candidates involved in human intelligence evolution. Expression analysis using RNA-Seq data revealed that most of the genes displayed a relatively high expression in the cerebral

cortex. ~~However, there~~ There is a distinct expression pattern between humans and other species, especially in ~~the tissues of the~~ neocortex tissues. Our work ~~may provided~~ a list of strong candidates for the evolution of human intelligence, which may also imply that some intelligence-related genes may undergo inter-species evolution and contain intra-species variation. ~~More importantly, the work provides a new method in searching for the key genes in human evolutionary genetics.~~

Introduction

~~The rapid change in intelligence from great apes to humans is one of the greatest mysteries in evolutionary genetics (Varki et al., 2008). Physically, h~~Humans have brains with significantly increased size and complexity ~~with a large expansion of the neocortex versus~~compared to their ape counterparts (Rakic., 2009; Chenn et al., 2002; Lui et al., 2011). Corresponding ~~to the increased size of the neocortex,~~ humans are ~~much more intelligent than chimpanzees, although chimpanzees are able to learn to use some specific tools and undertake certain tasks. The human advantage alterations~~ in intelligence has ~~helped this species~~humans compete with nature over the past few million years and survive, create tools, civilizations, and sciences (Deary et al., 2012). Inspection on the human genomic differences from our closest evolutionary relatives could help us to understand the intelligence-related genetic events during the hominid evolution. The ~~superior~~-intelligence difference is thought to be derived from changes in genetics, owing to a small fraction of the 1% of sequence differences between the human genome and the chimpanzee genome, in which the hominid-specific gene insertions, deletions, and duplications played a critical role (Cheng et al., 2015).

Various approaches in molecular biology have been used to search for the human-specific genes and mutations therein ~~that have led to the remarkable leap in~~ were related to the human intelligence. Several candidate genes involved in human intelligence evolution were identified based on hominid-human comparative genomic analysis, gene expression profiling and other functional evidences. For example, in a very recent study, the information from gene expression profiling was integrated with the information from gene duplications in the hominid and human lineages, which was then used to search for the human-specific genes that were highly expressed during human corticogenesis. In >35 candidates obtained through bioinformatics analysis, *NOTCH2NL* was functionally investigated and found to be able to promote the expansion of cortical progenitors, serving as an important gene contributing to the evolution of the human brain (Fiddes, et al., 2018; Suzuki et al., 2018). More recently, several human-specific genes ~~and variations~~ have been identified in which the genetic changes often occurred in gene regulation regions or resulted from the hominid-specific gene duplications; these include the *NOTCH2NL* gene, as well as *FZD8*, *SRGAP2*, *ARHGAP11B*, and *TBC1D3*, (Boyd et al., 2015; Dennis et al., 2012; Charrier et al., 2012; Florio et al., 2015; Ju et al., 2016).

As a highly heritable trait, intelligence has been intensively investigated using

Commented [FB1]: We don't really know this

forward genetic approaches (Davies et al., 2015; Davies et al., 2016; Sniekers et al., 2017; Trampush et al., 2017; Zabaneh et al., 2018; Savage et al., 2018; Davies et al., 2018; Hill et al., 2016; Hill et al., 2019). Several genome-wide association studies (GWAS) and meta-analyses using very large human populations have been performed to identify the genomic loci and related genes underlying intelligence. Despite a significant enrichment in the nervous system, the functional links of the identified genes are diverse and a wide variety of genes are involved (Davies et al., 2015; Davies et al., 2016; Sniekers et al., 2017; Trampush et al., 2017; Zabaneh et al., 2018; Savage et al., 2018). This suggests that the evolution of human intelligence ~~from great apes to humans~~ should also be a complicated process ~~and that~~. While ~~the~~ causative genes may be derived mainly from those in the central nervous system, ~~however~~, genes from many related biological processes may be also involved.

Investigating the human-specific variations (genetic differences between humans and the great apes) ~~would could~~ provide key clues for understanding the process of the evolution of human intelligence. As the assembling gaps and errors in the previous reference genomes of the great apes (e.g., the human sequence guided assembling from short reads). ~~However, the previous reference genomes of great apes (e.g., the human sequence guided assembling from short reads) the previous genomes were~~ are not qualified enough for the detection of complex structural variations (Prüfer et al., 2012; Scally et al., 2012; Prado-Martinez et al., 2013) such as tandem repeats, large-scale inversions, and duplications. ~~However, t~~These structural variations usually play important roles in human evolution (McLean et al., 2011). Hence, comparative genomic analysis from the complete genome sequences of both humans and great apes is needed to comprehensively mine the genetic variation. Recently, the high-quality genome sequences of three of human's closest relatives, chimpanzee, orangutan, and gorilla, were generated from long-read sequencing (PacBio technology) and *de novo* assembly (Kronenberg et al., 2018; Gordon et al., 2016). The chromosome-level contiguous genome assemblies facilitate a deeper understanding of the genomic differences between these species. These differences are responsible for all phenotypic differences between humans and apes but it is difficult to know which variants are specific to intelligence.

In order to further search out the candidate genes related to the evolution of human intelligence, we suppose that some intelligence genes may have both inter-species (between humans and the great apes) and intra-species (within humans)

variations. We collected genomic loci identified by several sets of GWAS on human intelligence. Genes in these loci (termed as intelligence-associated-genes) could be related to the intra-species intelligence differences. ~~DNA-sequence~~ Genomic variations differences between ~~the humans genome~~ and ~~that of~~ the great apes, revealed by recent high-quality sequencing (Kronenberg et al., 2018; Gordon et al., 2016), were considered to be the inter-species variations. Hence, intelligence evolution was integrated by the overlap of intelligence-associated genes and human-specific variations. We found that many of the intelligence-associated genes, including tens of strong candidate genes related to the evolution of human intelligence, contained human-specific structural variations, ~~including tens of strong candidate genes related to the evolution of human intelligence~~. Coupled with the expression profiling of the genes, this genome-wide analysis provided a useful resource for the evolutionary genetic studies on intelligence.

Materials & Methods

~~Identification of candidate genes from GWAS on human intelligence~~ Preparation of bait genes

To exploit the genes related to human intelligence, six major works were collected from GWAS or GWAS meta-analyses on human intelligence with a large sample size ~~from the last~~ in the recent five years (various intelligence related phenotypes including general cognitive, reaction time, verbal-numerical reasoning); ~~which~~ A total of ~~identified~~ 271 loci associated with intelligence in the human genome (Table S1) ~~were identified in these six studies (Table S1)~~.

The six GWAS studies include: (i) meta-analyses ~~for~~ of general cognitive function ($n=53,949$, Davies et al., 2015); (ii) GWAS of cognitive function and educational attainment ($n=112,151$, Davies et al., 2016); (iii) meta-analyses ~~for~~ of calculated Spearman's g or a primary measure of fluid intelligence ($n=78,308$, Sniekers et al., 2017); (iv) meta-analysis and gene-based analysis of human cognition using 24 cohorts ($n=35,298$, Zabaneh et al., 2018); GWAS using human populations with extremely high intelligence ($n=1,238$, Trampush et al., 2017); v) GWAS using human populations with extremely high intelligence ($n=1,238$, Trampush et al., 2017); meta-analysis and gene-based analysis for human cognition using 24 cohorts ($n=35,298$, Zabaneh et al., 2018); (vi) a recent meta-analysis of 14 independent epidemiological cohorts with intelligence assessed ($n=269,867$, Savage et al., 2018).

Commented [FB2]: The effort is appreciated but these two are still inverted. With "inverted" I mean the references are inverted. |

All the independent, significantly associated SNPs (IndSigSNPs) nearest genes (based on ANNOVAR annotations) were integrated with ~~the any~~ redundancies (the same gene identified in more than one study) removed. We take these 549 genes as “bait genes” (Table S2), which are candidates for human intelligence-related genes.

It should be noted that some GWAS works underlying human intelligence that have been published very recently may be not included in this study. This would not affect our analyses because this study aimed to provide some candidate genes for human intelligence evolution and cannot identify all related genes at one time. In addition,~~It should be noted here,~~ because genotype imputation was not performed for the X chromosome in some cohorts (e.g., the UKB cohort including 195,653 samples with the assessed phenotype verbal and mathematical reasoning)~~and GWAS have relatively low power in discovering the associations in the 23th chromosome,~~ the potential genes related to intelligence in the ~~sex X~~ chromosome~~s~~ were not included in our “bait genes”.

Preparation of variation pond

Comparative genomics analysis for the intelligence genes

The new genome assemblies of chimpanzees, gorillas, and orangutans have become fully available recently through single-molecule, real-time (SMRT) long-read sequencing, which improved the resolution of large and complex regions. We incorporated the human-specific structural variations (from intermediate size to large size) into a “variation pond”, including exon gain/loss, short tandem repeats (STRs), insertion/deletion (indels) of more than 50bp, and inversions. Moreover, considering the important role of the human-specific segmental duplications (HSDs, >1kb sequence with >90% similarity, indicating the large recent duplication events, Bailey et al., 2001) in new gene function and human evolution (Dennis et al., 2016), the HSDs identified recently from the comparative genomic information of both macaque and mouse (Dennis et al., 2017) were added into the “pond”. The “baits” and “ponds” were then integrated to detect whether there were human-specific variations hit by “bait genes”, thus generating the “prey genes” (Table S3-S5), which refer to the intelligence-related genes with human-specific variations. The human-specific variation that was located within each of the 549 genes was left for further analysis using a window of 1 Kb.

Previous studies (McLean et al., 2011; Kronenberg et al., 2018; Dennis et al., 2017)

Commented [FB3]: It is still unclear how this window is used. The authors should elaborate this further.

~~have identified a number of human-specific variants that were included in our work including exon gain and loss, STR, indels, hCONDEL, HSDs, and large structural variations. The human-specific variation that was located within each of the 549 genes was left for further analysis using a window of 1 Kb.~~ The genome sequences of three great apes were downloaded from the NCBI database. The reference genome sequences from Pan troglodytes (chimpanzee), Pongo abelii (Orangutan) and Gorilla gorilla (Western Lowland Gorilla) were downloaded from: ftp://ftp.ncbi.nlm.nih.gov/genomes/all/GCF/002/880/755/GCF_002880755.1_Clint_PTRv2, ftp://ftp.ncbi.nlm.nih.gov/genomes/all/GCF/002/880/775/GCF_002880775.1_Susie_PABv2, and ftp://ftp.ncbi.nlm.nih.gov/genomes/all/GCF/000/151/905/GCF_000151905.2_gorGor4, respectively.

Localized alignments of the target gene sequences were performed to filter out the false positives ~~of~~ human-specific variations from previous reports ([McLean et al., 2011](#); [Kronenberg et al., 2018](#); [Dennis et al., 2017](#)). The local sequences of the human genome were retrieved from chromosomes found in the GRCh38 version, and were then aligned with the great ape genomes using BLASTn (ncbi-blast-2.2.28+ version) with the parameters “-evalue 1E-50 -dust no”. The human-specific variation that was undetectable with a local BLAST was then removed for subsequent analyses, ~~thus generating the “prey genes” (Table 1 and Table S3–S5).~~

Investigation of HIEGs

Most “prey” genes contained the human-specific variation in introns, ~~with the variation~~ far away from the exon-intron junction site. This ~~are less likely to~~ may not affected the gene functions, so only ~~the~~ genes that contained variations in the coding regions were considered, named ~~as H~~ human intelligence evolution genes (HIEGs). These HIEGs included all the genes containing exon-gain/loss (2 genes), hCONDEL (28 genes), or HSD (1 gene), and genes with exon-located indels (8 genes) or STRs (4 genes). 40 non-redundant genes were finally identified, which were associated with human intelligence and which carried significant human-specific variations. The gene transcript information was obtained from Ensembl Release 95 ([Http://asia.ensembl.org/index.html](http://asia.ensembl.org/index.html)). For exon alignment, all transcript isoforms of one gene were compared both intra-species (within human) and inter-species (between human and the great apes) in order to confirm the specificity of the new

transcript in humans. ~~The human-specific transcripts and their most similar principle transcripts in humans and great apes are shown in Figure 3.~~ The human ~~principle~~principal isoform, the human variant isoform, the chimpanzee isoform, the gorilla isoform, and the orangutan isoform of the *PCCB* gene are ENST00000469217 (NM_001178014), ENST00000466072, ENSPTRT00000028811, ENSGGOT00000005484 and ENSPPYT00000016431, respectively. The human ~~principle~~principal isoform, the human variant isoform, the chimpanzee isoform, the gorilla isoform, and the orangutan isoform of the *STAU1* gene are ENST00000371856, ENST00000340954, ENSPTRT00000050938, ENSGGOT00000048652 and ENSPPYT00000012903, respectively.

The analysis for protein domain was performed using the Simple Modular Architecture Research Tool (SMART) in the normal mode ([Http://smart.embl-heidelberg.de](http://smart.embl-heidelberg.de)). The protein accession numbers in humans, chimpanzees, gorillas, and orangutans for KMT2D are NP_003473, XP_016778992, XP_018894141, XP_024112209, respectively. The protein accession numbers in human, chimpanzee, gorilla, and orangutan for TRIOBP are NP_001034230, XP_016794633, XP_004063488, and H2P4B5 (UniProt), respectively. We performed multiple alignments using the software Constraint-based Multiple Alignment Tool (COBALT, <ftp.ncbi.nlm.nih.gov/pub/agarwala/cobalt>).

Expression analysis for the candidates for intelligence evolution

We assessed the expression patterns of the HIEGs using transcriptome data for humans and their closest relatives. The HPA RNA-seq data ~~was-were~~ downloaded from the Human Protein Atlas ([Http://www.proteinatlas.org](http://www.proteinatlas.org)), including 102 samples of 37 tissues, in which TPM (transcripts per million) ~~was-were~~ used for the evaluation of expression levels. In order to compare the expression levels between humans and great apes, the RNA-Seq data (NCBI ID: GSE100796) from 107 samples ~~of-in~~ 8 brain regions ~~from-of~~ humans, chimpanzees, gorillas, and gibbons ~~was-were~~ used (Xu et al., 2018). The brain tissues included five neocortical areas ~~and three other brain tissues.~~ The five neocortical areas arenamely the anterior cingulate cortex (ACC), the dorsolateral prefrontal cortex (DPFC), the ventrolateral prefrontal cortex (VPFC), the premotor cortex (PMC), the primary visual cortex (V1C), ~~and. These t~~three other brain tissues including-are the hippocampus (HIP), the striatum, and the cerebellum (CB). Hierarchical cluster analysis was applied using the RPKM (reads per kilobase

per million) of 39 genes in 8 brain tissues of humans and the other three primates, and was displayed using the software MeV4.2 (<http://www.tm4.org/mev.html>).

Results

Identification of candidate A strategy for searching for genes in for human intelligence evolution

The ~~study~~ strategy that was used for detecting the candidate genes in the evolution of human intelligence is briefly described ~~in Figure 1~~ (Fig. 1). Hundreds of genetic loci for human intelligence and its related traits have been identified by GWAS ~~in the last ten years and studies have been conducted~~ using large population data ~~for human intelligence and its related traits (e.g., general cognitive ability, reaction time, and verbal-numerical reasoning). We integrated six high quality GWAS works and the large-scale GWAS meta-analyses from the last five years. This enabled the identification of a total of 271 loci in the human genome underlying intelligence related phenotypes (Table S1). A total of 549 human genes, which distributed across the whole human genome, were found to be located within the 271 intelligence-associated loci (Fig. 2, Table S2). We take the 549 genes implicated by these genetic loci as “bait genes”, which are related to human intelligence, constituted intelligence-related genes from GWAS, underlying the intra-species intelligence-related variations related to intelligence in human populations.~~

In the meantime, the recent long-read sequence assembly of chimpanzee, orangutan, and gorilla provided a large number of high-quality sequence differences between the human genome and the great ape genomes. We incorporated the human-specific structural variations, including exon gain and loss, STR, indels, hCONDEL, HSDs, and large structural variations, into a “variation pond” which~~In the meantime, comparative genomics analyses between the human genome and the great ape genomes (including those of the chimpanzee, gorilla, and orangutan) were used to identify kinds of specific human variations. The human-specific variations are taken as “variation ponds” that are related to human evolution.~~

The “bait genes” and “variation pond ponds” were then integrated to detect whether there were “bait genes” hit by the human-specific variation. After putting the “bait genes” into the “variation pond”, we found 406 sequence variations physically

located within 213 genes related to intelligence, which is considered to be “prey genes” (Fig. 2). The 213 “prey genes” linked the inter-species and intra-species variations and may be related to the evolution of human intelligence. Additionally, the potential effects of the human-specific variation through local sequence comparisons and gene structure analyses were carefully checked. Following in-depth analyses, there were 40 strong candidate genes that were identified as containing human-specific variations, probably changing either the coding or the expression of intelligence related genes, which were named human intelligence evolution genes (HIEGs, Table 1). intelligence-related genes with human specific variations as “prey genes” which are related to the evolution of human intelligence. Furthermore, based on the potential influence of the sequence variations and other gene function studies, the strong candidates in the “prey genes” was highlighted as potential human intelligence evolution related genes (HIEGs). The expression profiles of HIEGs were also investigated.

Preparation of bait genes

We integrated six high quality GWAS works and the large scale meta-analyses from the last five years that were found through publication searches. This enabled the identification of a total of 271 associated loci in the human genome underlying intelligence-related phenotypes (Table S1). It should be noted that some GWAS works underlying human intelligence that have been published very recently may be not included in this study. This would not affect our analyses because this study aimed to provide some candidate genes for human intelligence evolution and cannot identify all related genes at one time. According to the human gene annotation information, a total of 549 human genes were found to be located within the 271 associated loci. The 549 human genes, used as “bait genes” (Table S2), were distributed across the whole human genome as indicated by red dots in Fig. 2.

The “baits” constituted intelligence-related genes from GWAS underlying intra-species intelligence variations in human populations. A subset of these may be involved in the evolution of intelligence from great apes to humans. Hence, the comparative genomics analyses between great apes and humans were further added to examine whether there was a human-specific variation hit by our “bait genes”.

Human-specific variation on the intelligence related genes

The nearly complete genome sequences from chimpanzees, gorillas, and orangutans

have become fully available recently through single-molecule, real-time (SMRT) long-read sequencing, providing a large number of high-quality sequence differences between the human genome and great ape genomes. The new genome assemblies had improved the resolution of large and complex regions. We incorporated the human-specific structural variations (from intermediate size to large size) into a “variation pond”, including exon gain/loss, short tandem repeats (STRs), insertion/deletion (indels) of more than 50bp, and inversions. Moreover, considering the important role of the human-specific segmental duplications (HSDs, >1kb sequence with >90% similarity, indicating the large recent duplication events, Bailey et al., 2001) in new gene function and human evolution (Dennis et al., 2016), the HSDs identified recently from the genomic information of both the macaque and mouse (Dennis et al., 2017) were added into the “pond”. After putting the “bait genes” into the “variation pond”, we found 406 sequence variations physically located within 213 genes related to intelligence, which is considered to be “prey genes” (Fig. 2). The 213 “prey genes” linked the inter-species and intra-species variations and may be related to the evolution of human intelligence. Additionally, the potential effects of the human-specific variation through local sequence comparisons and gene structure analyses were carefully checked. Following in-depth analyses, there were 40 strong candidate genes that were identified as containing human-specific variations, probably changing either the coding or the expression of intelligence-related genes, which were named Human intelligence evolution genes (HIEGs, Table 1).

Exon gain and loss on the intelligence related genes

There were only two genes classified as the “prey genes” with exon gain or loss events during the evolution of the human lineage, *PCCB* and *STAU1* (Table 1).

The *PCCB* gene encoding the propionyl-CoA carboxylase subunit beta, was located in the locus on chromosome 3 identified by GWAS for intelligence ($P = 1.956 \times 10^{-9}$, Savage et al., 2018). Human *PCCB* contained 15 protein coding isoforms according to the Ensembl release 95. The principal isoforms generated proteins of 539 aa, 559 aa and 570 aa, respectively, all of which have identical isoforms in the chimpanzee. However, a new transcript variant was comparative genomics analysis showed human-specific variation led to the generation of a new transcript variant. Compared with the principal isoform, the variant lost a 60-bp exon (exon 4 of principal isoform) but gained another new 60-bp exon (exon 11 of the variational isoform). The gained

60-bp exon did not appear in any transcripts of the *PCCB* of the three great apes (Fig. 3A). This human-specific transcript variant can be detected in the human cerebral cortex at a lower level than that of the principal isoform (~~note: transcriptome data was~~ were obtained from the Human Protein Atlas, HPA). However, the relative cerebral cortex expression level of the *PCCB* variational transcript was higher than that of the principal transcript in the cerebral cortex, (relative to the average expression level of ~~this gene~~ the transcript in 37 tissues), ~~of the *PCCB* variant was higher than the principal transcript~~ (Fig. 3B). These result suggested the generally low expression level and the relative enrichment of the *PCCB* variant in the cerebral cortex, implying a potential role of the human-specific *PCCB* transcript variant in the cerebral cortex. Although there were ~~few~~ no reports about the mechanism of *PCCB* function in ~~on~~ neuron system development ~~for *PCCB*~~, mutations in *PCCB* are one of the major causes of the genetic disease propionic academia (PA). Neurological complications, such as intellectual disability, brain structural abnormalities, optic neuropathy, and cranial nerve abnormalities are significant symptoms of PA (Schreiber et al., 2012). Moreover, there were reports that patients carrying *PCCB* mutations exhibited intellectual disabilities (Witters et al., 2016).

STAUI was also located in a locus identified by GWAS for intelligence (Savage et al., 2018). *STAUI* encoding the double-stranded RNA-binding protein which regulates RNA metabolism. There ~~were~~ was a total of 10 protein-coding isoforms of human *STAUI*. Compared with the longest principal isoform, a human-specific insertion resulted in a gain of 123-bp exon (exon 2, located within the 5'UTR of the gene) in one transcript variant. The isoform, with the addition of a 123-bp exon, ~~was~~ not detected in any transcripts of the chimpanzee, gorilla, or orangutan. ~~The isoform, with the addition of a 123-bp exon, is a new isoform in the human transcriptome~~ (Fig. 3C). The human-specific isoform was expressed in many human tissues. In the human cerebral cortex, the expression of the new isoform was equivalent to ~21% of that of the principal isoform of the gene (Fig. 3D). Previous functional studies found that *STAUI* plays a role on mRNA transport in neuronal dendrites (Broadus et al., 1998).

STR variations on the intelligence related genes

In the variation pond, there were a total of 1,465 human-specific STR contractions and 4,921 human-specific STR expansions. These STRs hit 100 “~~prey~~ bait genes” (26 genes containing the STR contractions and 74 genes containing the STR expansions,

Table S3). Most of the human-specific STR variations ~~were~~ located within the intron regions or intergenic regions. Only four human-specific STRs were located within the exonic regions, highlighted as HIEGs (Table1). Among them, three STR expansions were located within the exon of non-~~protein~~ coding isoforms (~~processed transcripts~~) of three genes (*ARIH2*, *STAB1* and *TSNARE1*), and one STR contraction was located within the 39th exon of the *KMT2D* gene.

KMT2D, also known as *MLL2*, encodes an H3K4 histone methyltransferase ~~in humans~~ made up of 5537 amino acids. GWAS for intelligence detected one associated locus ($P = 2.518 \times 10^{-14}$, Savage et al., 2018) on chromosome 12 containing the candidate gene *KMT2D*. Compared with the *KMT2D* sequence in ~~the~~ great apes, the STR region contracted for 60-bp in the c.11838 ~~of human KMT2D, and~~. This region was also polymorphic among the human population. The contraction led to a 16-aa discontinuous deletion for the human *KMT2D* protein from the p.3958 position (Fig. 3E). The ~~peptide coding by this~~ STR region ~~is~~ in the coiled coil region for *KMT2D*, which could affect the protein structure through wrapping the hydrophobic residues and forming an amphipathic surface (Mason et al., 2004). Mutations in *KMT2D* are the main cause of the genetic disease Kabuki syndrome, and several mutations in the 39th exon have been found in Kabuki patients. Kabuki syndrome affects mental capabilities and most of these patients show various levels of intellectual disability (Lehman et al., 2017).

Indels on the intelligence related genes

Among 5,894 ~~human-human~~-specific deletions and 11,899 ~~human-human~~-specific insertions in the “variation pond”, we found 94 “prey genes” with 148 deletions and 144 genes with 298 insertions (Table S4). Furthermore, it was found that there were ~~7~~ 8 insertions and 1 deletion for the exonic regions, as highlighted by the HIEGs (Table 1). The insertion in *ARIH2* was also been identified as STR expansion which has been described above. Insertions in *PDE4D*, *NRXN1*, *EXOC4*, *FUT8*, and *ZNF584*, and the deletion in *SLC27A5* affect the lengths of the non-~~protein~~ coding isoforms (~~processed transcripts~~) of the six genes. One insertion Two genes, *GNB5* and *TRIOBP*, carried one insertion in the 3’UTR region and the exon region respectively.

The ~~while one~~ insertion in *TRIOBP* resulted in a gain of 675-bp coding regions (c.887-1560) in the 5th exon of the longest isoform when compared with that in the chimpanzee. This variation resulted in a 238-aa discontinuous insertion in the region

p.296-811 of the human TRIOBP protein (Fig. 3F). The GWAS for intelligence ($P=3.582 \times 10^{-8}$, Savage et al., 2018) and the GWAS for underlying brain ventricular volume also identified the gene *TRIOBP* as a strong candidate (Vojinovic et al., 2018). Biochemistry experiments have shown that TRIOBP could physically interact with TRIO, which is an important ~~gene-protein~~ involved ~~with-in~~ neural tissue development (Seipel et al., 2001). Mutations in *TRIOBP* ~~could~~ ~~can~~ cause autosomal recessive deafness-28 (DFNB28), ~~which is a genetic disease~~, and surprisingly, several causal mutations ~~have been~~ ~~was~~ located within the human-specific insertion regions (e.g., R347X and Q297X) (Shahin et al., 2006). ~~Hence, the human-specific variation in TRIOBP is probably also involved in the evolution of human intelligence.~~

We also searched for human conserved deletions (hCONDELs) near the “bait genes” that had been previously identified (McLean et al., 2011; Kronenberg et al., 2018). These sequences are lost in the human genome but are highly conserved among other species (including ~~the~~ great apes, the macaque, and the mouse). In total, 28 ~~“prey genes” genes~~ were identified as containing the hCONDELs. ~~Although and all none of the 28 hCONDELs located in the coding regions, they genes~~ were ~~all~~ taken into HIEGs (Table 1), ~~with-as~~ the high lineage specificity of hCONDELs (Table 1) serving as important signs of intelligence evolution. The list included *NRXN1*, *GRID2* and *GRIA4*, which were all involved in neurotransmission and the formation of synaptic contacts.

The *GNB5* gene in the ~~intelligence~~-associated locus on chromosome 15 ($P=2.47 \times 10^{-11}$, Savage et al., 2018), which ~~is responsible for~~ ~~encoding~~ the beta subunit of ~~the~~ heterotrimeric GTP-binding proteins (G proteins), ~~was found to be one of the HIEGs. Aligned with the GNB5 sequences of the humans and the great apes, we found that there was a 292-bp human-specific insertion in the 3'UTR of the gene and a 1,472-bp hCONDEL in the intron (2.7 kb distance to the third exon) of the gene (Fig. 2).~~ *GNB5* was expressed in the brain tissues and participated in neurotransmitter signaling (e.g., through the dopamine D2 receptor) (Xie et al., 2012). In human populations, mutations in *GNB5* have been reported to cause several diseases affecting intelligence, including language delay, ADHD/cognitive impairment with or without cardiac arrhythmia, and intellectual developmental disorder with cardiac arrhythmia (Lodder et al., 2016). ~~Aligned with the genome sequences of great apes, we found that there was a 292-bp human-specific insertion in the 3'UTR of the gene and a 1,472-bp hCONDEL in the intron (2.7 kb distance to the third exon) of the gene~~

(Fig. 2). The human-specific variation in GNB5 might participate in the evolution of human intelligence as well.

Large structural variations on the intelligence related genes

The inversion variation is ~~a-the~~ rearrangement in which ~~a-the~~ genomic segment is reversed. Based on the previous report, there were a total of 625 inversions in our “variation pond” ranging from 9 kb to 8.4 Mb in size. Among these, 31 of them hit the “bait genes” (Table S5). However, none of these genes were located in the breakpoint of the human-specific inversions.

There were 218 human-specific duplications (HSD) ~~the length of which were more than of >5-kb that were also reported~~ (Prado-Martinez et al., 2013). Among them, one 24.6-kb HSD was detected to be overlapped with the *AFF3* gene. The *AFF3* ~~also~~ contained an hCONDEL around the intron-exon junction regions (31-bp distance to the exon). The region around *AFF3* has been identified to be an ~~intelligence~~-associated locus in found in two distinct meta-analyses conducted on intelligence ($P= 1.56\times 10^{-8}$ [16] or $P= 3.41\times 10^{-10}$, Savage et al., 2018), but no functional reports on neuron development are available.

Expression profiling analysis of the HIEGs

The transcriptome data ~~from of~~ 37 human tissues ~~in from~~ the Human Protein Atlas database (Uhlén et al, 2015) was used to investigate the tissue expression patterns of the 40 highlighted HIEGs (Fig. 4A). Of these there were 23 genes with higher expression ~~levels~~ in the cerebral cortex than ~~its their~~ average expression ~~levels~~ in all 37 tissues. Furthermore, there were 12 genes with more than twofold expression ~~levels~~ in the cerebral cortex versus ~~its their~~ average expression levels (Fig. 4A). These were regarded as genes that were potentially involved in the development of the cerebral cortex. Except ~~the case for~~ *FUT8*, the remaining 11 genes are the hCONDEL-containing genes. ~~The expression patterns of the 11 genes exhibited three types.~~ Four genes, including *GRIA4*, *NRXN1*, *CADM2*, and *CALN*, ~~showed the were~~ ~~most significant~~ highly ~~expressed-expressions~~ in ~~the cerebral cortex brain tissues but~~ ~~had and~~ low ~~expressions-expressions~~ in the other tissues; ~~SGCZ~~, *DCC*, and *GRID2* showed ~~only~~ relatively high expressions in ~~the cerebral cortex brain tissues compared~~ with ~~the~~ low expressions in all other tissues; ~~and~~ *NCAM1*, *FBXL17*, *FUT8* and *GNB5* were ~~generally~~ expressed ~~well~~ in all tissues (Fig. 4A).

A transcriptome dataset sampling eight brain regions (five neocortical areas, hippocampus, striatum, and cerebellum) of both humans and four primate species (chimpanzees, gorillas, gibbon, and macaques) was ~~very recently~~ reported previously (Xu et al., 2018), which enabled a comprehensive interspecies comparison. This dataset was used to examine whether the genes containing human-specific variation ~~led to the~~ showed expression changes in the cortex ~~of HIEGs~~ between humans and the great apes. The expression data ~~of for the other 39 genes of the 40 HIEGs~~, with the exception of *STAU1*, can be found in the transcriptome dataset. Among them, the expression levels of the genes *AFF3*, *SKAP1*, *REEP3*, *DCC*, and *SGCZ* in the human neocortical areas were much lower than those in the neocortical areas of the great apes (fold change <0.5), while the expressions of *STAB1* in the human neocortex were much higher than those in the great apes (fold change= 4.8). Hierarchical clustering was also performed for the 39 gene expressions in all samples. We found that 5 neocortice ~~ex~~ tissues in the same species could be always clustered (that is, one clade for one species), while CB, STR, and HIP were generally clustered based on their tissue types (Fig4 B). This result suggested that the expression profiles of the HIEGs in the neocortex tissues displayed a strong species specificity, which was in contrast to the profiles in these non-neocortex tissues (e.g, CB). Taken together, ~~it was possible to determine that the human-specific variation in the intelligence-related genes, containing the human-specific variations, may have effects on the changes in expressioned differently in the neocortex tissues between the humans and the great apes. in the neocortex tissues.~~

Discussion

The understanding of human intelligence from the view of evolutionary genetics is an important scientific undertaking. Science magazine posted 125 scientific questions ~~to cover over the next quarter century which were driving basic scientific research were selected to cover in~~ various disciplines ~~as a way to celebrate its 125th anniversary~~ (Kennedy et al., 2005); among the top 25 questions, human evolution was addressed with the question: “What Genetic Changes Made Us Uniquely Human?” (Culotta., 2005). Obviously, ~~a genetic rise~~ changes related to intelligence over the last million years is ~~a one of the~~ key steps in human evolution, making us uniquely Homo sapiens. However, it is still very challenging to identify the causative genes that are responsible for the intelligence difference between humans and the other species.

~~The high quality genome assemblies of three great apes were completed recently and the improved sequence contiguity enabled more comprehensive and accurate discoveries with complex variations.~~ With both the human genome sequence and those of our closest relatives becoming available, it is possible to pinpoint the genetic changes underlying the phenotypic differences between humans and the great apes. However, it is still very challenging to study the causative genetic changes that are responsible for the rise in human intelligence because the molecular mechanisms controlling human intelligence are largely unknown. One possible solution is to utilize the genetic findings on intelligence from GWAS. A subset of genes ~~with that~~ were reveals ~~related to the ape-human intelligence differences in the evolution of intelligence in the human chimpanzee~~ may also contained intraspecific allelic variation underlying the variation of intelligence levels in human populations. ~~The high quality genome assemblies of three great apes were completed recently and the improved sequence contiguity enabled more comprehensive and accurate discoveries with complex variations.~~ Hence, in this work we integrated both the latest GWAS information on intelligence and the latest advances in the great ape genomics, aiming to mine the gene clues to understand the evolution of human intelligence. ~~We found several strong candidates, for example, the genes *TRIOBP* and *GNB5* contain human-specific variations and have the genetic evidence to be involved in the development of intelligence (Vojinovic et al. 2018; Xie et al., 2012; Lodder et al., 2016), although more in-depth molecular evidence and validations are needed in future experiments. Although our work did find some candidate genes involved in human intelligence evolution, this strategy has its shortcomings. Some of intelligence-related genes with inter-species (between human and the great apes) variation may contain no intra-specific variation in human populations and cannot be identified by GWAS. Hence, the human genes involved in intelligence evolution but without intra-species variations within human populations were missed using our strategy.~~

We identified a total of 40 HIEGs, based on the location of the human-specific variations. Except for the hCONDELs as their high lineage specificity and importance in evolution, all the other human-specific variations in the HIEGs located within the exons or putative regulatory regions. However, some of them located within the exon of non-protein coding isoforms. There were five genes carried human-specific variations in exons or UTRs of protein coding isoforms. We only discussed these five

HIEGs in this paper. It was notable that two of the 5 genes were related to mRNA decay. One candidate gene with an exon gain, *Staufen1 (STAU1)*, were reported to be involved in the transport, relocation, translation of mRNA and mRNA decay (Paul et al., 2018). Loss of *STAU1* function in mice resulted in impaired mRNA transport and reduced synapse formation (Vessey et al., 2008). Another candidate gene, *KMT2D* harbored a human-specific STR contraction. Several truncating mutations within *KMT2D* resulting in mRNA degradation through nonsense-mediated mRNA decay, contributing to protein haploinsufficiency (Micale et al., 2014). It is unclear whether there are any functional links between the two genes for mRNA processes in brain developments.

~~We found several strong candidates, for example, the genes *TRIOBP* and *GNB5* contain human specific variations and have the genetic evidence to be involved in the development of intelligence (Vojinovic et al. 2018; Xie et al., 2012; Lodder et al., 2016), although more in depth molecular evidence and validations are needed in future experiments.~~

The associated loci from GWAS in humans often contain several candidate genes, which is one of the difficulties in our bioinformatics analyses. To avoid artificial bias, all the candidate genes around the associated loci were included in the collection of “bait genes”, although usually for each associated locus only a single one is causative. Consequently, a total of 549 human genes were included for the 271 intelligence-associated loci and “bait genes” contain many false positives. However, ~~we~~We cannot clearly distinguish the true one with the highly linked one, because other information (e.g., based on expression profiles or the distance to lead SNPs) is often misleading. As a result, HIEGs must contain many unrelated genes, although the intelligence related genes involved in human evolution have been partly enriched. Further experiments and analyses may include the validation of gene functions (whether and how these genes could influence ~~intelligence~~ the development of ~~intelligence in~~ human brains) and the assessment of the effects of the human-specific variation (whether and how these sequence variations could influence the gene coding or the gene expression patterns).

There are already several findings of the human-chimpanzee differences altering the development of the neocortex to date. The knowledge from the works of gene functional studies and evolutionary genetics studies greatly enhanced our understanding of intelligence and the brain (Boyd et al., 2015; Dennis et al., 2012;

Charrier et al., 2012; Florio et al., 2015; Ju et al., 2016). Certainly, the known genes (e.g., *NOTCH2NL*, *FZD8*, *SRGAP2*, *ARHGAP11B*, and *TBC1D3*) are only a small proportion of the whole gene set that encapsulates the vast differences in brain size and intelligence ~~levels from~~between the great apes ~~to and the~~ humans, leaving many remaining gaps in our knowledge. More integrated approaches incorporating genetics, genomics, bioinformatics, and development biology will be needed in future works.

~~One candidate gene with an exon gain in human evolution, Staufeni1 (*STAU1*), which is involved in the transport, relocation, translation of mRNA and mRNA decay is known to regulate the post transcription phase (Paul et al., 2018). However, the loss of *STAU1* function in mice resulted in impaired mRNA transport and reduced synapse formation (Vessey et al., 2008). Another candidate gene, *KMT2D* with STR contractions, showed a number of truncating mutations within *KMT2D* resulting in mRNA degradation through nonsense mediated mRNA decay, contributing to protein haploinsufficiency (Micale et al., 2014). It is unclear whether there are any functional links between the two genes for mRNA processes in brain developments.~~

Conclusion

GWAS has identified hundreds of genes associated with intelligence variation in human populations. Through inter-species genome comparisons with the great apes, we found a small proportion of intelligence-related genes that also contained a human-specific variation which were detected in multiple high-quality genome assemblies of humans and its closest relatives. Through integrated analytical approaches, especially the careful checking of sequence alignments and gene annotations, we identified 40 ~~strong~~ candidates in which human-specific variation may have effects on gene coding or expressions. Transcriptome-wide comparison between humans and four primate species for the 40 candidate genes suggests that several of them displayed a different expression pattern among these species. The results implied that at least a few of the intelligence-related genes may contain both intra-species variations and inter-species variations. The intra-species variation underlies the small variation of intelligence levels for different human individuals while the inter-species variation controlled the large genetic differences of intelligence between the great apes and humans. This work may provide a list of candidate genes to be used in subsequent studies ~~as well as a new route for discovering genes that are important in the study of human intelligence evolution.~~

620

621 **Acknowledgments**

622 We thank Professor Xuehui Huang from Shanghai Normal University for providing
623 computer servers.

624

625

References

- Bailey JA, Yavor AM, Massa HF, Trask BJ, and Eichler EE. 2001.** Segmental duplications: organization and impact within the current human genome project assembly. *Genome Res* 11:1005-1017.
- Boyd JL, Skove SL, Rouanet JP, Pilaz LJ, Bepler T, Gordan R, et al. 2015.** Human-Chimpanzee Differences in a FZD8 Enhancer Alter Cell-Cycle Dynamics in the Developing Neocortex. *Current Biology* 25:772-779.
- Broadus J, Fuerstenberg S, and Doe CQ. 1998.** Stufen-dependent localization of prospero mRNA contributes to neuroblast daughter-cell fate. *Nature* 391:792-795.
- Charrier C, Joshi K, Coutinho-Budd J, Kim JE, Lambert N, de Marchena J, et al. 2012.** Inhibition of SRGAP2 Function by Its Human-Specific Paralogs Induces Neoteny during Spine Maturation. *Cell* 149:923-935.
- Cheng Z, Ventura M, She X, Khaitovich P, Graves T, Osoegawa K, et al. 2005.** A genome-wide comparison of recent chimpanzee and human segmental duplications. *Nature* 437:88-93.
- Chenn A, and Walsh CA. 2002.** Regulation of cerebral cortical size by control of cell cycle exit in neural precursors. *Science* 297:365-369.
- Culotta E. 2005.** What genetic changes made us uniquely human? *Science* 309: 91.
- Davies G, Armstrong N, Bis JC, Bressler J, Chouraki V, Giddaluru S, et al. 2015.** Genetic contributions to variation in general cognitive function: a meta-analysis of genome-wide association studies in the CHARGE consortium (N=53 949). *Molecular Psychiatry* 20:183-192.
- Davies G, Lam M, Harris SE, Trampush JW, Luciano M, Hill WD, et al. 2018.** Study of 300,486 individuals identifies 148 independent genetic loci influencing general cognitive function. *Nat Commun* 9:2098.
- Davies G, Marioni RE, Liewald DC, Hill WD, Hagenaars SP, Harris SE, et al. 2016.** Genome-wide association study of cognitive functions and educational attainment in UK Biobank (N=112151). *Mol Psychiatry* 21:758-767.
- Deary IJ. 2012.** Intelligence. *Annu Rev Psychol* 63:453-482.
- Dennis MY, and Eichler EE. 2016.** Human adaptation and evolution by segmental duplication. *Curr Opin in Genet & Dev* 41:44-52.
- Dennis MY, Harshman L, Nelson BJ, Penn O, Cantsilieris S, Huddleston J, et al. 2017.** The evolution and population diversity of human-specific segmental duplications. *Nat Ecol Evol* 1:69.

Formatted: Norwegian (Bokmål)

660 **Dennis MY, Nuttle X, Sudmant PH, Antonacci F, Graves TA, Nefedov M, et al.**
661 **2012.** Evolution of human-specific neural SRGAP2 genes by incomplete segmental
662 duplication. *Cell* 149:912-922.

663 **Fiddes IT, Lodewijk GA, Mooring M, Bosworth CM, Ewing AD, Mantalas GL,**
664 **et al. 2018.** Human-Specific NOTCH2NL Genes Affect Notch Signaling and Cortical
665 Neurogenesis. *Cell* 173:1356-1369.

666 **Florio M, Albert M, Taverna E, Namba T, Brandl H, Lewitus E, et al. 2015.**
667 Human-specific gene ARHGAP11B promotes basal progenitor amplification and
668 neocortex expansion. *Science* 347:1465-1470.

669 **Gordon D, Huddleston J, Chaisson MJ, Hill CM, Kronenberg ZN, Munson KM,**
670 **et al. 2016.** Long-read sequence assembly of the gorilla genome. *Science* 352:
671 aae0344.

672 **Hill WD, Davies G, Harris SE, Hagenaars SP, Liewald DC, Penke L, et al. 2016.**
673 Molecular genetic aetiology of general cognitive function is enriched in evolutionarily
674 conserved regions. *Transl Psychiatry* 6:e980.

675 **Hill WD, Marioni RE, Maghzian O, Ritchie SJ, Hagenaars SP, McIntosh AM, et**
676 **al. 2019.** A combined analysis of genetically correlated traits identifies 187 loci and a
677 role for neurogenesis and myelination in intelligence. *Mol Psychiatry* 24:169-181.

678 **Ju XC, Hou QQ, Sheng AL, Wu KY, Zhou Y, Jin Y, et al. 2016.** The
679 hominoid-specific gene TBC1D3 promotes generation of basal neural progenitors and
680 induces cortical folding in mice. *Elife* 5: e18197

681 **Kennedy D, and Norman C. 2005.** What don't we know? *Science* 309:75.

682 **Kronenberg ZN, Fiddes IT, Gordon D, Murali S, Cantsilieris S, Meyerson OS, et**
683 **al. 2018.** High-resolution comparative analysis of great ape genomes. *Science*. 360

684 **Lehman N, Mazery AC, Visier A, Baumann C, Lachesnais D, Capri Y, et al. 2017.**
685 Molecular, clinical and neuropsychological study in 31 patients with Kabuki
686 syndrome and KMT2D mutations. *Clin Genet* 92:298-305.

687 **Lodder EM, De Nittis P, Koopman CD, Wiszniewski W, De Souza CFM,**
688 **Lahrouchi N, et al. 2016.** GNB5 Mutations Cause an Autosomal-Recessive
689 Multisystem Syndrome with Sinus Bradycardia and Cognitive Disability. *Am J Hum*
690 *Genet* 99: 786-786.

691 **Lui JH, Hansen DV, and Kriegstein AR. 2011.** Development and Evolution of the
692 Human Neocortex. *Cell* 146: 18-36.

693 **Mason JM, and Arndt KM. 2004.** Coiled coil domains: stability, specificity, and

Formatted: Norwegian (Bokmål)

biological implications. *Chembiochem* 5:170-176.

McLean CY, Reno PL, Pollen AA, Bassan AI, Capellini TD, Guenther C, et al. 2011. Human-specific loss of regulatory DNA and the evolution of human-specific traits. *Nature* 471:216-219.

Micale L, Augello B, Maffeo C, Selicorni A, Zucchetti F, Fusco C, et al. 2014. Molecular Analysis, Pathogenic Mechanisms, and Readthrough Therapy on a Large Cohort of Kabuki Syndrome Patients. *Human Mutat* 35:841-850.

Paul S, Dansithong W, Figueroa KP, Scoles DR, and Pulst SM. 2018. Staufen1 links RNA stress granules and autophagy in a model of neurodegeneration. *Nat Commun* 9:3648.

Prado-Martinez J, Sudmant PH, Kidd JM, Li H, Kelley JL, Lorente-Galdos B, et al. 2013. Great ape genetic diversity and population history. *Nature* 499:471-475.

Prufer K, Munch K, Hellmann I, Akagi K, Miller JR, Walenz B, et al. 2012. The bonobo genome compared with the chimpanzee and human genomes. *Nature* 486:527-531.

Rakic P. 2009. Evolution of the neocortex: a perspective from developmental biology. *Nat Rev Neurosci* 10:724-735.

Savage JE, Jansen PR, Stringer S, Watanabe K, Bryois J, De Leeuw CA, et al. 2018. Genome-wide association meta-analysis in 269,867 individuals identifies new genetic and functional links to intelligence. *Nat Genet* 50:912-919.

Scally A, Dutheil JY, Hillier LW, Jordan GE, Goodhead I, Herrero J, et al. 2012. Insights into hominid evolution from the gorilla genome sequence. *Nature* 483:169-175.

Schreiber J, Chapman KA, Summar ML, Ah Mew N, Sutton VR, MacLeod E, et al. 2012. Neurologic considerations in propionic acidemia. *Mol Genet Metab* 105:10-15.

Seipel K, O'Brien SP, Iannotti E, Medley QG, and Streuli M. 2001. Tara, a novel F-actin binding protein, associates with the Trio guanine nucleotide exchange factor and regulates actin cytoskeletal organization. *J Cell Sci* 114:389-399.

Shahin H, Walsh T, Sobe T, Abu Sa'ed J, Abu Rayan A, Lynch ED, et al. 2006. Mutations in a novel isoform of TRIOBP that encodes a filamentous-actin binding protein are responsible for DFNB28 recessive nonsyndromic hearing loss. *Am J Hum Genet* 78:144-152.

Sniekers S, Stringer S, Watanabe K, Jansen PR, Coleman JRI, Krapohl E, et al.

Formatted: Norwegian (Bokmål)

Formatted: Norwegian (Bokmål)

2017. Genome-wide association meta-analysis of 78,308 individuals identifies new loci and genes influencing human intelligence. *Nat Genet* 49:1107-1112.

Suzuki IK, Gacquer D, Van Heurck R, Kumar D, Wojno M, Bilheu A, et al. 2018. Human-Specific NOTCH2NL Genes Expand Cortical Neurogenesis through Delta/Notch Regulation. *Cell* 173:1370-1384e1316.

Trampush JW, Yang MLZ, Yu J, Knowles E, Davies G, Liewald DC, et al. 2017. GWAS meta-analysis reveals novel loci and genetic correlates for general cognitive function: a report from the COGENT consortium. *Mol Psychiatry* 22:1651-1652.

Uhlen M, Fagerberg L, Hallstrom BM, Lindskog C, Oksvold P, Mardinoglu A, et al. 2015. Proteomics. Tissue-based map of the human proteome. *Science* 347:1260419.

~~Varki A, Geschwind DH, and Eichler EE. 2008. Explaining human uniqueness: genome interactions with environment, behaviour and culture. *Nat Rev Genet* 9:749-763.~~

Vessey JP, Macchi P, Stein JM, Mikl M, Hawker KN, Vogelsang P, et al. 2008. A loss of function allele for murine *Staufen1* leads to impairment of dendritic *Staufen1*-RNP delivery and dendritic spine morphogenesis. *Proc Natl Acad Sci USA* 105:6374-16379.

Vojinovic D, Adams HH, Jian X, Yang Q, Smith AV, Bis JC, et al. 2018. Genome-wide association study of 23,500 individuals identifies 7 loci associated with brain ventricular volume. *Nat Commun* 9:3945.

Witters P, Debbold E, Crivelly K, Vande Kerckhove K, Corthouts K, Debbold B, et al. 2016. Autism in patients with propionic acidemia. *Mol Genet Metab* 119:317-321.

Xie K, Ge S, Collins VE, Haynes CL, Renner KJ, Meisel RL, et al. 2012. Gβ5-RGS complexes are gatekeepers of hyperactivity involved in control of multiple neurotransmitter systems. *Psychopharmacology (Berl)* 219:823-834.

Xu C, Li Q, Efimova O, He L, Tatsumoto S, Stepanova V, et al. 2018. Human-specific features of spatial gene expression and regulation in eight brain regions. *Genome Res* 28:1097-1110

Zabaneh D, Krapohl E, Gaspar HA, Curtis C, Lee SH, Patel H, et al. 2018. A genome-wide association study for extremely high intelligence. *Mol Psychiatry* 23:1226-1232.

Formatted: Norwegian (Bokmål)

Formatted: Norwegian (Bokmål)