

Trophic niches of a seabird assemblage in Bass Strait, south-eastern Australia

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The foraging niches of seabirds are driven by a variety of factors, including competition for prey that promotes divergence in foraging niches. Bass Strait, south-eastern Australia, is a key region for seabirds, with little penguins *Eudyptula minor*, short-tailed shearwaters *Ardenna tenuirostris*, fairy prions *Pachyptila turtur* and common diving-petrels *Pelecanoides urinatrix* being particularly abundant in the region. The trophic niches of these species were investigated using isotopic values in whole blood and by identifying prey remains in stomach contents. The four species occupied different isotopic niches that varied between 3 years and 3 regions. Little penguins consumed mainly fish whereas the three procellariiforms primarily consumed coastal krill *Nyctiphanes australis*. The dietary similarities between the procellariiforms suggest that food resources are segregated in other ways, with interspecific differences in isotope niches possibly reflecting differential consumption of key prey, divergent foraging locations and depth, and differences in breeding phenology. Because oceanographic changes predicted to occur due to climate change may result in reduced coastal krill availability, adversely affecting these seabird predators, further information on foraging zones and feeding behaviour of small procellariiform species is needed to elucidate more fully the segregation of foraging niches, the capacity of seabirds to adapt to climate change and the potential for interspecific competition in the region.

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15 ABSTRACT

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INTRODUCTION

Seabirds are major consumers of marine biomass, feeding on a variety of fish, cephalopods and crustaceans (Ridoux 1994, Brooke 2004). The foraging niche of seabirds is influenced by a range of factors, including environmental conditions (Waugh & Weimerskirch 2003, Amélineau et al. 2016, Jakubas et al. 2017), prey availability (Baird 1991, Camprasse et al. 2017), morphological characteristics and their influence on flight performance (Phillips et al. 2004, 2005, Navarro et al. 2013), and interspecific competition (González-Solís et al. 2008, Phillips et al. 2008, Kokubun et al. 2016). Competition is thought to promote foraging niche divergence since species occupying the same ecological niche cannot theoretically coexist through time (Schoener 1974). Seabirds may separate their resources on several dimensions, with studies showing divergence in foraging zone (González-Solís et al. 2008, Barger et al. 2016), diving depth (Mori & Boyd 2004), the timing of breeding (Granroth-Wilding & Phillips 2019), and seasonal patterns of activity (Phillips et al. 2008). In particular, divergence in diet has been proposed as an important mechanism in reducing niche overlap (Ridoux 1994, Surman & Wooller 2003, Pratte et al. 2017).

Bass Strait, the shallow continental shelf area between mainland Australia and Tasmania (Fig. 1), is a key region for Australian seabirds, supporting a large proportion of breeding populations of at least 11 major species (Table 1; Ross et al. 2001). This area is considered a region of low primary productivity (Gibbs et al. 1986, Gibbs et al. 1991) that occurs at the confluence of three main ocean currents. The warm, oligotrophic waters of the East Australian Current (EAC) flow southward along the eastern edge of Bass Strait (Ridgeway & Godfrey 1997, Sandery & Kämpf 2007) while the South Australian Current (SAC) advects warm water from the west onto the shelf which then flows eastward through Bass Strait (Sandery & Kämpf 2007). The latter is the

major source of Bass Strait water and is strongest in winter (Ridgeway & Condie 2004, Sandery & Kämpf 2007). In summer, a weakening or reversal of this eastward-flowing trend occurs (Gibbs et al. 1986, Sandery & Kämpf 2007). Finally, in winter, cold, nutrient-rich sub-Antarctic surface water (SASW) enters Bass Strait from the south (Gibbs 1992) where it mixes with the EAC and SAC along the sub-Tropical Convergence (STC, Prince 2001).

The relative influence of the currents and upwelling systems affecting Bass Strait varies spatially, seasonally and inter-annually (Prince 2001, Sandery & Kämpf 2005). This affects the reproductive success of seabirds in Bass Strait, presumably due to shifts in prey availability (Mickelson et al. 1992). Additionally, climate change is predicted to weaken the SAC (Feng et al. 2012) and increase the strength of the EAC, resulting in warming along the path of its strengthening (Cai et al. 2005). This is likely to have a considerable impact on the marine ecosystem of Bass Strait. Seabird assemblages in other parts of the world have shown differential responses to shifts in ocean regimes in parameters such as breeding success, population size and survivorship due, at least in part, to changes in prey availability (Croxall et al. 2002). Knowledge of the trophic relationships and diets of Bass Strait seabirds is crucial for predicting their population responses to environmental change. This information is important for the conservation of these marine predators and for the refinement of sustainable fisheries management practices. At present, the trophic structure of Bass Strait seabird community is poorly understood, with trophic niches of most pelagic species not yet described or based on a few localised studies that did not address spatial or temporal variation. Accordingly, it is not known whether, or how, they diverge in foraging niche.

The little penguin *Eudyptula minor*, short-tailed shearwater *Ardenna tenuirostris*, fairy prion *Pachyptila turtur* and common diving petrel *Pelecanoides urinatrix* are the most abundant and

ubiquitous seabirds in Bass Strait, breeding sympatrically on numerous offshore islands (Schumann et al. 2014). They are known to feed on a variety of fish, cephalopod and/or crustacean prey (Harper 1976, Montague et al. 1986, Schumann et al. 2008, Chiaradia et al. 2010). Previous foraging ecology studies indicate that the little penguin is an inshore forager relying mainly on small pelagic schooling fish (Cullen et al. 1992, Chiaradia et al. 2010), while the pelagic short-tailed shearwater, with a foraging range extending to the Antarctic waters (Woehler et al. 2006, Cleeland et al. 2014), and the smaller and more neritic fairy prion and common diving petrel, feed on a wide range of small prey, concentrating predominantly on coastal krill (*Nyctiphanes australis*) and myctophid fish (Harper 1976, Weimerskirch & Chérel 1998, Schumann et al. 2008). However, in Bass Strait, information on spatial and temporal variation in the ecology of these species is limited to the at-sea foraging behaviour of little penguins and short-tailed shearwaters (e.g. Collins et al. 1999, Chiaradia et al. 2010, Berlincourt & Arnould 2015a, b) and there is almost no information on the small procellariiforms (Underwood 2012).

The aims of the present study, therefore, were to: 1) determine the trophic niche of the study seabirds using both stomach contents and stable isotope analysis; 2) investigate temporal (inter-annual and seasonal) and geographic variation in their isotopic niche, and 3) assess the degree of niche segregation between these four abundant species within Bass Strait.

MATERIAL & METHODS

The study was conducted during the winters (July – August) of 2008 – 2010 and summers (January – February) of 2009 – 2011 in three regions of Bass Strait, south-eastern Australia (Fig.

1). In summer, the study species were sampled in mid, early and late chick-rearing, respectively for the little penguin (Reilly & Cullen 1981), the short-tailed shearwater (Vertigan 2010) and the fairy prion (Harper 1976) (Fig. 2). In winter, sampling occurred during the inter-breeding period for the little penguin (Reilly & Cullen 1981) and fairy prion (Harper 1976) and incubation for the common diving petrel (Schumann et al. 2008) (Fig. 2). The trophic niches of little penguins, short-tailed shearwaters, fairy prions and common diving petrels were determined using two complementary techniques. Trophic information was derived from stable isotope values in whole blood of each species in western, central and eastern Bass Strait, and stomach samples were collected from the seabirds in central Bass Strait to assess the relative importance of prey and inform interpretation of the stable isotope results. Procellariiform study species were banded and little penguins were micro-chipped to avoid sampling the same individual more than once. All research was conducted under permit from Deakin University (animal ethic permit: AWC A9-2008) and the Department of Sustainability and Environment (Permit No. 10004530), and access to the islands was provided by Parks Victoria.

Dietary analysis

Stomach contents analysis provides information on the composition and abundance of prey consumed (Duffy & Jackson 1986). Stomach samples were collected from the four seabird species on Notch (38°56'S, 146°37'E) and Kanowna (39°10'S, 148°16'E) Islands in central Bass Strait (Fig. 1). Adult little penguins were sampled in both winter and summer (n = 20 and 22, respectively), short-tailed shearwater and fairy prion diet samples were collected in summer only (n = 51 and 20, respectively) and common diving petrels were sampled in winter only (n = 6)

(Fig. 2). Due to logistical constraints and few individuals onshore in some years, it was not possible to sample all species in all years.

Adult birds were captured as they came ashore at night after foraging at sea. Diet samples were collected using the water-offloading technique. While it is possible that not all stomach contents were retrieved, stomach flushing is an effective technique for diet estimation in seabirds (Gales 1987) and, in most cases, the majority of prey remains were ejected on the second (final) flush, as evidenced by clear water being ejected. After flushing, birds were given an electrolyte solution of Vytrate or Lectade (Jurox Pty Ltd, NSW), and placed into boxes for recovery (Chiaradia et al. 2003). Little penguins and short-tailed shearwaters were also provided with a meal, delivered via a stomach tube, of homogenised pilchard (purchased snap-frozen and unsalted) immediately before release.

Diet samples were frozen (-20°C) or stored in 70% ethanol after collection. Crustacean remains were initially identified to lower taxonomic levels with reference to Ritz et al. (2003). Sagittal otoliths, scales, fish mouth parts and cephalopod beaks were identified by comparison to reference atlases (Neira et al. 1998, Lu & Ickeringill 2002, Furlani et al. 2007) and collections (held by Phillip Island Nature Parks and Deakin University).

Fresh prey items were washed with water and separated from accumulated ones. To estimate the numerical abundance of crustacean prey, the heads of amphipods and whole bodies of copepods, isopods and crab megalopa were counted directly while individual eyes of krill and stomatopods were counted and divided by two. Left and right otoliths were counted and the side comprising the greater number considered representative of the minimum number of each fish taxon per sample. Where otoliths were unidentifiable, their abundance was halved and rounded to the nearest number. Similarly, the highest number of upper or lower squid beaks in a sample was

used to estimate the abundance of cephalopods and only unbroken beaks were measured to estimate size (Tollit et al. 1997). Hard prey remains that could not be quantified, such as fish scales, were assigned a numerical abundance of one. The frequency of occurrence of prey remains was calculated as the proportion of samples containing identifiable remains in which a particular prey type occurred while the numerical abundance was expressed as the mean number of each prey taxon encountered in samples.

Stable isotope analyses

Stable carbon ($\delta^{13}\text{C}$) and nitrogen ($\delta^{15}\text{N}$) values in tissues have been used to infer the diet of a range of marine species (Hobson & Welch 1992, Hobson 1993, Cherel et al. 2007). Stable isotope values of $\delta^{13}\text{C}$ values allow discrimination between benthic and pelagic prey (e.g., Cherel et al. 2007) and inshore and offshore feeding (Hobson et al. 1994b), while $\delta^{15}\text{N}$ in tissues show enrichment with increasing trophic levels (Hobson et al. 1994a). Information derived from whole blood, as used in the present study, reflects dietary integration of approximately four weeks (Bearhop et al. 2002).

Blood samples (< 0.2 ml) were collected from seabirds resident in western (WBS - Lady Julia Percy Island: 38°25'S, 142°00'E), central (CBS - Notch, Kanowna, Norman: 39°02'S, 146°12'E) and Anser Islands: 39°09'S, 146°18'E) and eastern Bass Strait (EBS - Gabo Island: 37°34'S, 149°55'E). A total of 278 (167 in summer, 111 in winter), 177 (summer only), 88 (66 in summer, 22 in winter) and 38 (winter only) stable isotope profiles were obtained from little penguins, short-tailed shearwaters, fairy prions and common diving petrels, respectively. Adult individuals (only birds that were not sampled for diet determination) were captured as they

returned to their nesting burrows at night or taken from their burrows during the day. Little penguins and short-tailed shearwaters were captured by hand, common diving-petrels were captured in mist nets, and fairy prions were captured by hand or using hand nets as they approached their burrows. Upon capture, blood was collected into a heparinised syringe via venipuncture of the tarsal vein or an inter-digital vein in the foot.

Blood samples were stored frozen (-20°C) and, prior to analysis, oven dried (60°C) and homogenised using a mortar and pestle. The low lipid content of whole blood does not typically necessitate lipid extraction (Cherel et al. 2005a). Indeed, all mean values of C:N mass ratio encompassed a narrow range (3.1-3.7) indicating low lipid content and, thus, allowing accurate comparisons of $\delta^{13}\text{C}$ values among groups (Bond & Jones 2009). Isotope ratios in whole blood were measured using a continuous-flow isotope ratio mass spectrometer, with analyses conducted by the Isotope Ratio Mass Spectrometry service in the Research School of Biology, Australian National University (Canberra, Australia). The values of stable isotope abundances were expressed in δ -notation as the deviation from standards in parts per thousand according to the equation:

$$\delta X = [(R_{\text{sample}}/R_{\text{standard}}) - 1]$$

where X is ^{15}N or ^{13}C and R represents the corresponding $^{15}\text{N}/^{14}\text{N}$ or $^{13}\text{C}/^{12}\text{C}$ ratios (Hobson et al. 1994a). R_{standard} values were based on Vienna Pee Dee Belemnite for ^{13}C , and atmospheric nitrogen (N_2) for ^{15}N . Based on variation between repeats of a standard material, measurement error was estimated to be ± 0.20 and ± 0.15 ‰ for $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$, respectively.

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190 *Statistical analyses*

191 All statistical analyses were conducted in the R statistical environment 3.5.1 (R Core Team
 192 2018). To investigate the effect of geographic, inter-annual and seasonal variations in stable
 193 isotope values, generalised linear models (GLM) were fitted using the *lme4* package (Bates et al.
 194 2014). Terms were added sequentially, model selection was based on Akaike's information
 195 criterion (AIC), and the global models were checked to ensure normality and homoscedasticity
 196 of the residuals. Post-hoc tests were conducted using analyses of variance (ANOVA) and *t*-tests,
 197 or Kruskal-Wallis and Wilcoxon rank sum tests where transformations did not improve data
 198 distributions. The stable isotope Bayesian ellipses in R (*SIBER* package; Jackson et al. 2011)
 199 were used to determine the isotopic niche width of each species in each region, for each year and
 200 season. The Standard Ellipse Area corrected (SEA_C; 40% probability of containing a
 201 subsequently sampled datum regardless of sample size) was used to quantify niche width. The
 202 Bayesian estimate of the standard ellipse and its area (SEA_B) were used to measure the overlap
 203 of the isotopic niches between groups (Jackson et al. 2011). The niche overlap was estimated as
 204 the isotopic area of overlap from the maximum likelihood fitted ellipses of two given groups.

205

206 **RESULTS**

207 *Diet*

208 Stomach content samples were obtained from individuals between August 2008 and January
 209 2011 in order to inform interpretation of the stable isotope results. Samples were collected from
 210 little penguins in both winter and summer (2008-2009), from short-tailed shearwaters in summer

2009 and 2010, from fairy prions in summer 2011 and from common diving petrels in winter 2008 and 2009. Overall, 79, 84, 95 and 40 % of little penguin, short-tailed shearwater, fairy prion and common diving-petrel samples, respectively, contained identifiable fresh prey remains. Stomach samples of studied seabird species contained remains of fish, cephalopods and crustaceans (Table 2). Not all taxa could be identified to species level. Little penguins ingested crustaceans, comprising isopods, amphipods and/or copepods, but consumed mainly jack mackerel in winter and Australian anchovy *Engraulis australis* in summer, though high numbers of post-larval fish were also ingested in summer (Table S1 in the Supplement). The diets of all three procellariiform species were dominated by euphausiids, particularly coastal krill (*Nyctiphanes australis*), representing 78 – 96 % of the mean number of prey items consumed by these species (Table S2 and Table S3 in the Supplement). Other important prey taxa included *Euphausia* sp. and the hyperiid amphipod *Themisto australis* for short-tailed shearwaters, the megalopa stage of a swimming crab species *Ovalipes* sp. for fairy prions and hyperiid amphipods for common diving-petrels. For short-tailed shearwaters, the abundance of crustaceans was higher in 2010 than in 2009 (Wilcoxon-test, $w = 126.5$, $p\text{-value} < 0.01$), mainly driven by the variation in number per samples of coastal krill ($n = 118.9 \pm 52.6$ and $n = 969.6 \pm 194.2$ in 2009 and 2010, respectively).

Stable isotopes analysis

Blood samples were collected from all four species in WBS and CBS, and from little penguins and short-tailed shearwaters in EBS (Table 3 and 4). Values of $\delta^{13}\text{C}$ ranged between -20.6 and -18.1 ‰ for little penguins, between -23.7 and -20.4 ‰ for short-tailed shearwaters, between -

233 21.0 and -18.3 ‰ for fairy prions and between -21.4 and -19.4 ‰ for common diving petrels.

234 Whole blood $\delta^{15}\text{N}$ values ranged between 10.8 and 16.0 ‰ for little penguins, between 7.8 and

235 11.4 ‰ for short-tailed shearwaters, between 8.8 and 14.8 ‰ for fairy prions and between 10.9

236 and 14.5 ‰ for common diving petrels (Fig. S1 in the Supplement).

237 For all four species, stable isotope values in whole blood showed intraspecific variation between

238 regions and years (Fig. 3 and 4). Inter-annual variations of $\delta^{13}\text{C}$ values were significant in all

239 species in most regions (Paired *t*-test or Wilcoxon-test: $P < 0.01$) except for short-tailed

240 shearwater and common diving petrel in CBS (*t*-test: $P > 0.07$). While there was no pattern in

241 $\delta^{13}\text{C}$ differences between regions for the short-tailed shearwater, for the three other species

242 values in CBS were generally lower than those from WBS (0.56 ‰ to 1.40 ‰ lower) (Tables 3

243 and 4). Indeed, for the little penguin, fairy prion and common diving petrel, the variable “region”

244 explained, respectively, 43, 37 and 58 % of the variance for the $\delta^{13}\text{C}$ model, but only 2 % for the

245 short-tailed shearwater. For $\delta^{15}\text{N}$, the best models retained, with interactions, the $\delta^{13}\text{C}$, the region

246 and the year (and season for the little penguin and fairy prion) as significant variables explaining

247 60 % to 93 % of the deviance (Table S4 in the Supplement). While “season” explained 27.1% of

248 the model for the fairy prion (winter data available only for WBS in 2009), this variable was not

249 significant for the little penguin (accounting for only 0.2% of the variation). No inter-seasonal

250 variations of $\delta^{13}\text{C}$ in blood of little penguin and fairy prion were found (except in CBS, paired *t*-

251 test or Wilcoxon-test: $P < 0.01$). Significant inter-annual differences were recorded in all regions

252 (Paired *t*-test or Wilcoxon-test: $P < 0.01$), but no clear pattern was detected in the values or the

253 isotopic niche metrics (Tables 3, 4 and Table S5 in the Supplement). Similarly, for each year, the

254 $\delta^{15}\text{N}$ values varied between region for the little penguin, fairy prion and common diving petrel

255 (Paired *t*-test or Wilcoxon-test: $P < 0.01$). For the short-tailed shearwater, a spatial difference in

$\delta^{15}\text{N}$ values was detected between WBS and EBS in 2010 (Paired t -test $t_{37} = -2.19$, $P < 0.05$), but no other differences were found. The models for $\delta^{13}\text{C}$ retained the region and year as main variables for all the study species, explaining 55 % to 70 % of the variance (Table S4 in the Supplement).

The four study species occupied different isotopic niches in all years and each region (Fig. S1 in the Supplement). Values of $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ in whole blood of short-tailed shearwaters were lower (with no isotopic niche overlap) than those of the other species in each region in all three years (paired t -test or Wilcoxon-test: $p < 0.01$, Table 3 and Table S5 in the Supplement). In contrast, mean $\delta^{15}\text{N}$ values in blood of little penguins typically showed the greatest enrichment in both winter and summer, though this varied spatially and inter-annually (Table 3 and 4). Isotopic $\delta^{13}\text{C}$ values of little penguins and fairy prions were relatively similar, but tended to be higher than those of common diving-petrels in winter, particularly in CBS. While the isotopic niche of the fairy prion overlapped sparsely with the common diving petrel (SEA_B overlap $< 8.1\%$), and with little penguin in 2009 and 2011 (SEA_B overlap $< 4.9\%$), niche overlap with the little penguin was important in 2010 (maximum SEA_B overlap = 21.8% in winter 2010 in WBS, Table S5 in the Supplement).

DISCUSSION

Foraging niches of sympatric seabird species diverge in a variety of spatial and temporal ways (Waugh & Weimerskirch 2003, González-Solis et al. 2008, Davies et al. 2009). Bass Strait is occupied by an abundant marine avifauna (Ross et al. 2001), with little previously known of the trophic niches of most species. Combining stomach contents and stable isotope analyses, the

present study has shown that the four most abundant and ubiquitous species generally occupy different trophic niches that vary among regions, years and season.

Diet

The little penguin is considered an inshore generalist forager relying mostly on small pelagic prey such as Clupeiformes (Cullen et al. 1992, Chiaradia et al. 2010, Sutton et al. 2015). In the present study, stomach contents of little penguins in summer were similar to that previously reported, with Australian anchovy and post-larval fish contributing the majority of samples. In contrast, winter stomach contents were dominated by jack mackerel, highlighting a seasonal switch in the availability of the main prey of little penguins. Such differences have also been shown in little penguins from Albatross Island in southern Bass Strait (Gales & Pemberton 1990) and Phillip Island in northern Bass Strait (Cavallo et al. 2018). While recent studies have observed that jellyfish can contribute a substantial proportion of the little penguin diet (Sutton et al. 2015, Cavallo et al. 2018), no evidence of such prey were found in the present study. This could potentially be due to rapid digestion of gelatinous prey in comparison to fish or crustaceans (Cavallo et al. 2018), emphasising the limitation of traditional stomach content analyses, or reflect inter-annual differences in available prey types.

Stomach contents of short-tailed shearwaters in the present study were similar to those of individuals from Tasmania (Weimerskirch & Cherel 1998, Cherel et al. 2005b). The main identified prey was the coastal krill, indicating that sampled birds were mostly returning from short foraging trips over the continental shelf (Blackburn 1980, Weimerskirch & Cherel 1998). Indeed, during the breeding season, short-tailed shearwaters alternate between short (1-2 d) local trips within 35-70 km of the colony and long trips (10-20 d) to Antarctic waters (Weimerskirch

& Cherel 1998, Woehler et al. 2006, Raymond et al. 2010, Einoder et al. 2011, Berlincourt & Arnould 2015b) where they feed mainly on coastal krill, and myctophid fish and Antarctic krill, respectively (Montague 1986, Weimerskirch & Cherel 1998). In the 2010 samples, a limited number of birds (n = 4) had stomach contents dominated by stomach oil and digested *Euphausia* sp, suggesting they had returned from long trips to Antarctic waters (Weimerskirch & Cherel 1998).

Coastal krill occurs in neritic waters of eastern Australia, where other krill species are rare or absent (Blackburn 1980). Due to its abundance, it plays a key role in the coastal ecosystem, reflected by its dominance in the diets of various cetacean, seabirds and fish species (O'Brien 1988, Gill et al. 2011, Woehler et al. 2014). Despite limited data on the diet of fairy prions and common diving petrels in Bass Strait, their stomach contents confirmed the importance of coastal krill to these species in the Australasian region (Harper 1976, Schumann et al. 2008). These results, together with estimates of trip duration in previous studies (1-3 d trips at sea, Harper 1976, Bocher et al. 2000a, b, Navarro et al. 2013, Zhang et al. 2018), suggest that both breeding fairy prions and common diving petrels most likely forage within Bass Strait or in the vicinity of the continental shelf. This analysis emphasises the value of a multi-tools approach when considering niche segregation, as here, while stomach analysis can suggest substantial dietary overlap among the procellariiforms, isotopic and tracking analysis may be able to tease the species apart into separate foraging niches.

Spatial variability in isotopic niche

Since whole blood integrates dietary information over approximately four weeks (Bearhop et al. 2002), it might be expected that isotopic values for short-tailed shearwaters would reflect a

combination of both their local and Antarctic foraging areas (Berlincourt & Arnould 2015b), thereby masking any differences in blood isotope values between foraging zones. However, Cherel et al. (2005b) showed that while most of the food consumed by short-tailed shearwaters during short local trips is allocated to their chick, adults feed for themselves when foraging farther south and, therefore, have a truly Antarctic blood isotopic signature. In the present study, values of $\delta^{13}\text{C}$ in whole blood of short-tailed shearwaters were comparable to those of other procellariiforms foraging in subantarctic waters (Cherel et al. 2002a, Cherel et al. 2002b, Quillfeldt et al. 2005) but were slightly higher than those in plasma of short-tailed shearwaters from south Tasmania (Cherel et al. 2005b).

Interestingly, the $\delta^{13}\text{C}$ values of short-tailed shearwaters varied significantly between the three study regions in Bass Strait, indicating possible foraging spatial segregation within the Southern Ocean by individuals from the different regions. As $\delta^{13}\text{C}$ values are higher in subtropical than in Antarctic waters (Cherel & Hobson 2007, Cherel et al. 2007, 2010, Jaeger et al. 2010), this suggests a latitudinal segregation in the foraging areas between the three sampled populations. However, this variation was not consistent across years, with strong inter-annual variability in $\delta^{13}\text{C}$ values for birds from WBS and EBS compared to CBS. This could suggest not only colony-specific niche segregation but also density-dependent competition (Ainley et al. 2004, Wakefield et al. 2013), with the smallest colonies having a more flexible foraging area. Indeed, the CBS population is considerably bigger than the WBS and EBS populations, with 755,400, 30,000 and 6,000 breeding pairs, respectively (Bowker 1980, Pescott 1976, Fullagar & Heyligers 1996, Schumann et al. 2014). This is in accordance with previous tracking studies (Berlincourt & Arnould 2015b) that observed inter-annual longitudinal and latitudinal variation in the long trip foraging areas of short-tailed shearwater from the small populations in WBS and EBS. Despite

the geographic and temporal differences in $\delta^{13}\text{C}$ observed, there were no major differences in the $\delta^{15}\text{N}$ values between regions in the present study, highlighting the consistency of the diet of short-tailed shearwaters in the Southern Ocean.

For little penguins, fairy prions and common diving petrels, seasonal and geographic differences in isotopic signatures are likely to reflect differences in prey availability associated with the strength of the prevailing of ocean currents and upwelling systems in the different regions of Bass Strait. For example, the SAC may transport cold waters from the west into Bass Strait (Mickelson et al. 1992, Sandery & Kämpf 2007), weakening towards the east (Sandery & Kämpf 2007) where the EAC increases in prevalence, bringing warmer nutrient-poor water into north-eastern Bass Strait (Gibbs 1992). This was reflected in $\delta^{13}\text{C}$ values in the whole blood of little penguins and fairy prions, where values were systematically lower in CBS than in WBS and EBS. Similar observations have been reported for Australian fur seals where $\delta^{13}\text{C}$ values in blood plasma of individuals from CBS were consistently lower than those from EBS (Arnould et al. 2011). In winter, however, spatial differences in isotopic values declined. This may reflect the homogenization of Bass Strait waters in winter due to a greater influence of the SAC and SASW during this period (Prince 2001, Sandery & Kämpf 2007).

Inter-annual trophic variability

The isotope values in the whole blood of little penguins from WBS and CBS in summer are within the range previously reported from Phillip Island in CBS (Chiaradia et al. 2010, Chiaradia et al. 2012), with the exception of 2010 when $\delta^{15}\text{N}$ values were significantly lower in both regions. This could reflect fluctuations in isotopic baseline signatures due to different water masses and variable strength of the currents influencing the regions where individuals foraged.

Indeed, as reported by Polito et al. (2019), variations in oceanic factors such as chlorophyll-*a* concentration can substantially alter mean isotope values independently of any change in the diet of the species. However, inter-annual variation in $\delta^{15}\text{N}$ values could also reflect a variation in main prey species consumed, with little penguins known to have important inter-annual variability in their diet (Gales & Pemberton 1990, Cullen et al. 1992, Chiaradia et al. 2010). In the present study, individuals from CBS in summer 2009 consumed predominantly Australian anchovy, a species exploiting higher trophic levels than other prey targeted by little penguins (Espinoza et al. 2009, van der Lingen et al. 2009). As previously highlighted by Chiaradia et al. (2010), $\delta^{15}\text{N}$ values are higher during years with an important proportion of anchovy in little penguin diet. Therefore, it is likely that the low $\delta^{15}\text{N}$ values in summer 2010 in the present study was due to a depletion of Australian anchovy in the diet, potentially due to a reduced availability in the region. This is consistent with previous reports indicating inter-annual flexibility in little penguin at-sea foraging behaviour in relation to environmental conditions that directly influence prey abundance (Berlincourt & Arnould 2015a, Camprasse et al. 2017). Indeed, in WBS and CBS, the niche space occupied by little penguins in 2010 was much larger than in 2009 and 2011, indicating a larger trophic diversity (Layman et al. 2007), possibly due to the absence of the usual main prey. Similarly, for both fairy prions and common diving petrels, $\delta^{15}\text{N}$ values in the whole blood of both species in the region varied substantially between years, suggesting a potential variation in the importance of their main prey (coastal krill) in their respective diet. Significant inter-annual differences in the density and biomass of coastal krill in southern Bass Strait have previously been observed (Young et al. 1993).

Trophic and isotopic niche segregations

In the present study, interspecific comparisons of stomach contents and $\delta^{15}\text{N}$ values revealed that little penguins typically occupied the highest trophic positions of the four seabird species while short-tailed shearwaters always occupied the lowest. Little penguin $\delta^{15}\text{N}$ values were nevertheless lower than those of the top predators shy albatross and Australian fur seal (Arnould et al. 2011; Cherel et al. 2013), but were close to the values of the large Australasian gannet (Angel et al. 2016), that predominantly consumes pilchards and anchovy (Bunce 2001) (Fig. 5). In contrast, $\delta^{15}\text{N}$ values of short-tailed shearwaters, fairy prions and common diving petrels were remarkably lower than those of the much smaller white-faced storm petrels (Underwood 2012) (Fig. 5), which consume a significant proportion of fish in addition to coastal krill (Underwood 2012). These results, combined with the stomach content analysis, confirm that coastal krill was a key prey taxon in all three procellariiforms in central Bass Strait. During breeding, both fairy prions and common diving petrels return to the nest every night (Harper 1976, Payne & Prince 1979), suggesting that they forage mainly on the shelf near their colonies. Elsewhere, fairy prions take prey from the surface waters (Harper 1987, Prince & Morgan 1987) whereas common diving petrels exploit depths averaging 2-4m (Navarro et al. 2013, Navarro et al. 2014, Dunphy et al. 2015). Likewise, despite isotopic signatures showing self-maintenance feeding in the Southern Ocean, short-tailed shearwaters forage on coastal krill over the shelf near colonies during short trips (Einoder et al. 2011, Berlincourt & Arnould 2015b), resulting in the potential for interspecific overlap in the foraging zones of the three procellariiforms. However, short-tailed shearwaters forage at deeper depths (average 13 m) during local trips (Weimerskirch & Cherel 1998). While the foraging zones and dive depths of little penguins may overlap with those of short-tailed shearwaters (Berlincourt & Arnould 2015a, 2015b), the limited distance travelled per trip and fish-based diet of little penguins would reduce competition with procellariiforms.

In addition to segregation of diet and foraging behaviour, the four species differ in their breeding phenologies. Common diving petrels, fairy prions and short-tailed shearwaters lay their eggs in late July, late October and late November, respectively (Harris 1979, Marchant & Higgins 1990). Thus, there is limited overlap in the breeding periods of the three procellariiforms. However, the protracted and variable breeding season of little penguins (Reilly & Cullen 1981, Cullen et al. 1992) may overlap with the other species. Interspecific competition may, therefore, intensify in years of low fish prey availability. Indeed, previous studies have documented the presence of coastal krill in the diet of little penguin during years of low prey availability (Cullen et al. 1992). The dependence of these seabird species on relatively few prey types (such as coastal krill, pilchard or anchovy) may increase the impacts of reductions in prey abundance. Climate models have described an intensification of the EAC due to large-scale changes in ocean circulation in the Southern Hemisphere (Cai 2006), produced in association with an increasing trend in the Southern Annular Mode (Cai et al. 2005). During years of intensified EAC, Young et al. (1993) reported a dramatic drop in coastal krill biomass. This is likely to adversely affect seabirds in the region (Mills et al. 2008, Chambers et al. 2011). Similarly, significant mortality events, poor chick growth and population declines in short-tailed shearwaters in Tasmania have been previously attributed to local prey shortages (Vertigan 2010). Declines in coastal krill availability may also indirectly impact little penguins since this species is an important dietary component of several of their prey taxa (Harris et al. 1979, O'Brien 1988). The predicted increase in the strength of the EAC with climate change (Cai et al. 2005) could, therefore, have severe negative consequences for the Bass Strait seabird community (Chambers et al. 2011).

CONCLUSIONS

In summary, the present study has shown that the isotopic niches of seabirds in Bass Strait vary significantly between regions, years and seasons. These differences are likely to result from changes in prey availability driven by variations in ocean currents and local productivity. Despite interspecific similarities in diet, divergence in the relative foraging niche is likely to reduce interspecific competition for prey, though this may become more important in years of low prey availability. The low diversity of prey taxa ingested by these seabirds suggests that they are vulnerable to changes in the availability of key prey. In order to better understand the foraging niches of the Bass Strait community of seabirds, as well as their capacity to adapt to changing environmental conditions, more detailed information on their foraging zones and feeding behaviour is required. This is particularly important for the small procellariiforms in light of the paucity of information in south-eastern Australia, in contrast to the numerous studies that have been conducted on little penguins and short-tailed shearwaters in the region (*e.g.* Ropert-Coudert et al. 2004, 2009, Cleeland et al. 2014, Berlincourt & Arnould 2015a, b). Such information may help elucidate the likelihood of interspecific competition in this assemblage of seabirds.

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462 REFERENCES

463 Ainley DG, Ribic CA, Ballard G, Heath S, Gaffney I, Karl BJ, Barton KJ, Wilson PR, Webb S
464 (2004) Geographic structure of Adelie penguin populations: overlap in colony-specific
465 foraging areas. *Ecol Mono* 74(1):159-178

466 Alderman RL, Gales R, Tuck GN, Lebreton JD (2011) Global population status of shy albatross
467 and an assessment of colony-specific trends and drivers. *Wildl Res* 38: 672-686

468 Amélineau F, Grémillet D, Bonnet D Bot TL, Fort J (2016) Where to forage in absence of sea
469 ice? Bathymetry as a key factor for an Arctic seabird. *PLOSE ONE* 11:e0157764

470 Angel LP, Berlincourt M, Arnould JPY (2016) Pronouced inter-colony variation in the foraging
471 ecology of Australian gannets: influence of habitat differences. *Mar Ecol Prog Ser*
472 556:261-272

473 Arnould JPY, Cherel Y, Gibbens J, White JG, Littnan CL (2011) Stable isotopes reveal inter-
474 annual and inter-individual variation in the diet of female Australian fur seals. *Mar Ecol*
475 *Prog Ser* 422:291-302

476 Baird PH (1991) Optimal foraging and intraspecific competition in the tufted puffin. *Condor*
477 93:503-515

478 Barger CP, Young RC, Cable JM, Ito M, Kitaysky AS (2016) Resource partitioning between
479 sympatric seabird species increases during chick-rearing. *Ecosphere* 9:e01447

480 Bates D, Maechler M, Bolker B, Walker S (2014) lme4: Linear mixed-effects models using
481 Eigen and S4. R package version 1.1-6. <http://CRAN.R-project.org/package=lme4>

- 482 Bearhop S, Waldron S, Votier SC, Furness RW (2002) Factors that influence assimilation rates
- 483 and fractionation of nitrogen and carbon stable isotopes in avian blood and feathers.
- 484 *Physiol Biochem Zool* 75:451-458
- 485 Berlincourt M, Arnould JPY (2015a) Influence of environmental conditions on foraging
- 486 behaviour and its consequences on reproductive performance in little penguins. *Mar Biol*
- 487 162:1485-1501
- 488 Berlincourt M, Arnould JPY (2015b) Breeding short-tailed shearwaters buffer local
- 489 environmental variability in south-eastern Australia by foraging in Antarctic waters. *Mov*
- 490 *Ecol* 3:16
- 491 Blackburn M (1980) Observations on the distribution of *Nyctiphanes australis* Sars (Crustacea,
- 492 Euphausiidae) in Australian waters. *Rep Div Fish Oceanogr CSIRO* 119:1-10
- 493 Bocher P, Cherel Y, Hobson KA (2000a) Complete trophic segregation between south georgian
- 494 and common diving petrels during breeding at Iles Kerguelen. *Mar Ecol Prog Ser*
- 495 208:249-264
- 496 Bocher P, Labidoire B, Cherel Y (2000b) Maximum dive depths of common diving petrels
- 497 (*Pelecanoides urinatrix*) during the annual cycle at Mayes Island, Kerguelen. *J Zool*
- 498 *Lond* 251:517-524
- 499 Bond AL, Jones IL (2009) A practical introduction to stable-isotope analysis for seabird
- 500 biologists: approaches, cautions and caveats. *Mar Ornithol* 37:183-188
- 501 Bowker GM (1980) Griffiths Island. *Corella* 4:104-6

- Brooke MDL (2004) The food consumptions of the world's seabirds. PRSB Biol Sci 271:S246-S248
- Brothers N, Pemberton D, Pryor H, Halley V (2001) Tasmania's offshore islands: seabirds and other natural features. Tasmanian Museum and Art Gallery: Hobart, Tasmania
- Bunce A (2001) Prey consumption of Australasian gannets (*Morus serrator*) breeding in Port Phillip Bay, southeast Australia, and potential overlap with commercial fisheries. ICES J Mar Sci 58:904-915
- Bunce A, Norman FI, Brothers N, Gales R (2002) Long-term trends in the Australasian gannet (*Morus serrator*) population in Australia: the effect of climate change and commercial fisheries. Mar Biol 141: 263-269
- Cai W (2006) Antarctic ozone depletion causes an intensification of the Southern Ocean super-gyre circulation. Geophys Res Lett 33:L03712
- Cai W, Shi G, Cowan T, Bi D, Ribbe J (2005) The response of the Southern Annular Mode, the East Australian Current, and the southern mid-latitude ocean circulation to global warming. Geophys Res Lett 32:L23706
- Camprasse ECM, Sutton GJ, Berlincourt M, Arnould JPY (2017) Changing with the times: little penguins exhibit flexibility in foraging behaviour and low behavioural consistency. Mar Biol 164:169
- Cavallo C, Chiaradia A, Deagle BE, McInnes JC, Sánchez S, Hays GC, Reina RD (2018) Molecular analysis of predator scats reveals role of salps in temperate inshore food webs. Front Mar Sci 5:381

- Chambers LE, Devney CA, Congdon BC, Dunlop N, Woehler E, Dann P (2011) Observed and predicted effects of climate on Australian seabirds. *Emu* 111:235-251
- Cherel Y, Bocher P, De Broyer C, Hobson KA (2002a) Food and feeding ecology of the sympatric thin-billed prion *Pachyptila belcheri* and Antarctic *P. Desolata* prions at Iles Kerguelen, Southern Indian Ocean. *Mar Ecol Prog Ser* 228:263-281
- Cherel Y, Bocher P, Trouvé C, Weimerskirch H (2002b) Diet and feeding ecology of blue petrels *Halobaena caerulea* at Iles Kerguelen, Southern Indian Ocean. *Mar Ecol Prog Ser* 228:283-299
- Cherel Y, Jaquemet S, Maglio A, Jaeger A (2014) Differences in $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values between feathers and blood of seabird chicks: implications for non-invasive isotopic investigations. *Mar Biol* 161:229-237
- Cherel Y, Hobson KA (2007) Geographical variation in carbon stable isotope signatures of marine predators: a tool to investigate their foraging areas in the Southern Ocean. *Mar Ecol Prog Ser* 329:281-287
- Cherel Y, Hobson KA, Guinet C, Vanpe C (2007) Stable isotopes document seasonal changes in trophic niches and winter foraging individual specialization in diving predators from the Southern Ocean. *J Anim Ecol* 76:826-836
- Cherel Y, Hobson KA, Hassani S (2005a) Isotopic discrimination between food and blood and feathers of captive penguins: implications for dietary studies in the wild. *Physiol Biochem Zool* 78:106-115
- Cherel Y, Hobson KA, Weimerskirch H (2005b) Using stable isotopes to study resource acquisition and allocation in procellariiform seabirds. *Oecologia* 145:533-540

- 544 Cherel Y, Jaeger A, Alderman R, Jaquemet S, Richard P, Wanless RM, Phillips RA, Thompson
545 DR (2013) A comprehensive isotopic investigation of habitat preferences in nonbreeding
546 albatrosses from the Southern Ocean. *Ecography* 36:277-286
- 547 Chiaradia A, Costalunga A, Kerry K (2003) The diet of little penguins (*Eudyptula minor*) at
548 Phillip Island, Victoria, in the absence of a major prey - pilchard (*Sardinops sagax*). *Emu*
549 103:43-48
- 550 Chiaradia A, Forero MG, Hobson KA, Cullen JM (2010) Changes in diet and trophic position of
551 a top predator ten years after a mass mortality of a key prey. *ICES J Mar Sci* 67:1710-
552 1720
- 553 Chiaradia A, Forero MG, Hobson KA, Swearer SE, Hume F, Renwick L, Dann P (2012) Diet
554 segregation between two colonies of little penguins *Eudyptula minor* in southeast
555 Australia. *Austral Ecol* 37:610-619
- 556 Clelland JB, Lea MA, Hindell MA (2014) Use of the Southern Ocean by breeding short-tailed
557 shearwaters (*Puffinus tenuirostris*). *J Exp Mar Biol Ecol* 450:109-117
- 558 Collins M, Cullen JM, Dann P (1999) Seasonal and annual foraging movements of little
559 penguins from Phillip Island, Victoria. *Wildl Res* 26:705-721
- 560 Croxall JP, Trathan PN, Murphy EJ (2002) Environmental change and Antarctic seabird
561 populations. *Science* 297:1510-1514
- 562 Cullen JM, Montague TL, Hull C (1992) Food of little penguins *Eudyptula minor* in Victoria:
563 Comparison of three localities between 1985 and 1988. *Emu* 91:318-341

- Dann P, Norman FI (2006) Population regulation in little penguins (*Eudyptula minor*): the role of intraspecific competition for nesting sites and food during breeding. *Emu* 106(4):269-289
- Davies WE, Hipfner JM, Hobson KA, Ydenberg RC (2009) Seabird seasonal trophodynamics: isotopic patterns in a community of pacific alcids. *Mar Ecol Prog Ser* 382:211-219
- Duffy DC, Jackson S (1986) Diet studies of seabirds: A review of methods. *Colonial Waterbirds* 9:1-17
- Dunphy BJ, Taylor GA, Landers TJ, Sagar RL, Chilvers BL, Ranjard L, Rayner MJ (2015) Comparative seabird diving physiology: first measures of haematological parameters and oxygen stores in three New Zealand Procellariiformes. *Mar Ecol Prog Ser* 523:187-198
- Einoder LD, Page B, Goldsworthy SD, De Little SC, Bradshaw CJA (2011) Exploitation of distant antarctic waters and close neritic waters by short-tailed shearwaters breeding in south australia. *Austral Ecol* 36:461-475
- Espinoza P, Bertrand A, van der Lingen CD, Garrido S, Mendiola BR (2009) Diet of sardine (*Sardinops sagax*) in the northern Humboldt Current system and comparison with the diets of clupeoids in this and other eastern boundary upwelling systems. *Prog in Oceano* 83:242-250
- Feng M, Caputi N, Pearce A (2012) The Leeuwin Current. In Poloczanska, ES, Hobday AJ, Richardson AJ (eds) A marine climate change impacts and adaptations report card for Australia 2012. Accessed 25 September 2012. www.oceanclimatechange.org.au
- Fullagar PJ, Heyligers PC (1996) Gabo Island Shearwater Surveys, 1995 and 1996.

- 584 Furlani D, Gales R, Pemberton D (2007) Otoliths of common australian temperate fish, Vol.
585 CSIRO Publishing, Collingwood
- 586 Gales R (1987) Validation of the stomach flushing technique for obtaining stomach contents of
587 penguins. Ibis 129:335-43
- 588 Gales R, Pemberton D (1990) Seasonal and local variation in the diet of the little penguin,
589 *Eudyptula minor*, in Tasmania. Aust Wildlife Res 17:231-259
- 590 Gibbs CF (1992) Oceanography of Bass Strait: Implications for the food supply of little penguins
591 *Eudyptula minor*. Emu 91:395-401
- 592 Gibbs CF, Arnott GH, Longmore AR, Marchant JW (1991) Nutrient and plankton distribution
593 near a shelf break front in the region of the Bass Strait cascade. Aust J Mar Fresh Res
594 42:201-217
- 595 Gibbs CF, Tomczak Jr M, Longmore AR (1986) The nutrient regime of Bass Strait. Aust J Mar
596 Fresh Res 37:451-466
- 597 Gill PC, Morrice MG, Page B, Pirzl R, Levings AH, Coyne M (2011) Blue whale habitat
598 selection and within-season distribution in a regional upwelling system off southern
599 Australia. Mar Ecol Prog Ser 421:243-263
- 600 González-Solís J, Croxall JP, Afanasyev V (2008) Offshore spatial segregation in giant petrels
601 *Macronectes* spp.: Differences between species, sexes and seasons. Aquat Conserv: Mar
602 Freshwat Ecosyst 17:S22-S36
- 603 Granroth-Wilding H MV, Phillips RA (2019) Segregation in space and time explains the
604 coexistence of two sympatric sub-Antarctic petrels. Ibis 161:101-116

- 605 Harper PC (1976) Breeding biology of the fairy prion (*Pachyptila turtur*) at the Poor Knights
606 Islands, New Zealand. N Z J Zool 3:351-371
- 607 Harper PC (1987) Feeding behaviour and other notes on 20 species of procellariiformes at sea.
608 Notornis 34:169-192
- 609 Harris GP, Griffiths FB, Clementson LA, Lyne V, Van der Doe H (1991) Seasonal and
610 interannual variability in physical processes, nutrient cycling and the structure of the food
611 chain in Tasmanian shelf waters. J Plankton Res 13:109-131
- 612 Harris MP (1979) The seabirds of the Victorian islands. A report to the Ministry for
613 Conservation, Victorian State Government. Institute for Terrestrial Ecology Project Report
614 No. 588. Natural Environment Research Council
- 615 Hedd A, Gales R (2001) The diet of shy albatrosses (*Thalassarche cauta*) at Albatross Island,
616 Tasmania. J. Zool. 253:69-90
- 617 Hobson KA (1993) Trophic relationships among high arctic seabirds: Insights from tissue-
618 dependent stable-isotope models. Mar Ecol Prog Ser 95:7-18
- 619 Hobson KA, Piatt JF, Pitocchelli J (1994a) Using stable isotopes to determine seabird trophic
620 relationships. J Anim Ecol 63:786-798
- 621 Hobson KA, Piatt JF, Pitocchelli J (1994b) Using stable isotopes to determine seabird trophic
622 relationships. J Anim Ecol 63:786-798
- 623 Hobson KA, Welch HE (1992) Determination of trophic relationships within a high Arctic
624 marine food web using $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ analysis. Mar Ecol Prog Ser 84:9-18

- 625 Jackson AL, Inger R, Parnell AC, Bearhop S (2011) Comparing isotopic niche widths among
626 and within communities: SIBER – Stable Isotope Bayesian Ellipses in R. *J Anim Ecol*
627 80:595-602
- 628 Jaeger A, Connan M, Richard P, Cherel Y (2010) Use of stable isotopes to quantify seasonal
629 changes of trophic niche and levels of population and individual specialisation in
630 seabirds. *Mar Ecol Prog Ser* 401:269-277
- 631 Jakubas D, Wojczulanis-Jakubas K, Iliszko LM, Strøm H, Stempniewicz L (2017) Habitat
632 foraging niche of a High Arctic zooplanktivorous seabird in a changing environment. *Sci*
633 *Rep* 7:16203
- 634 Kokubun N, Yamamoto T, Sato N, Watanuki Y, Will A, Kitaysky AS, Takahashi A (2016)
635 Foraging segregation of two congeneric diving seabird species breeding on St. George
636 Island, Bering Sea. *Biogeosciences* 13:2579-2591
- 637 Layman CA, Arrington DA, Montaña CG, Post DM (2007) Can stable isotope ratios provide for
638 community-wide measures of trophic structure? *Ecology* 88:42-48
- 639 Leitch TM, Dann P, Arnould JPY (2014) The diet of Pacific gulls (*Larus pacificus*) breeding at
640 Seal Island in northern Bass Strait. *Aus J Zool* 62:216-222
- 641 Lu CC, Ickeringill R (2002) Cephalopod beak identification and biomass estimation techniques:
642 Tools for dietary studies of southern Australian finfishes. Museum Victoria Science
643 Reports No. 6. Museum Victoria, Fisheries Research and Development Corporation,
644 Melbourne

- Marchant S, Higgins PJ (1990) Handbook of Australian, New Zealand and Antarctic birds, Vol. 1. Oxford University Press, Melbourne
- Mickelson MJ, Dann P, Cullen JM (1992) Sea temperature in bass strait and breeding success of the little penguin *Eudyptula minor* at Phillip Island, south-eastern australia. Emu 91:355-368
- Mills JA, Yarrall JW, Bradford-Grieve JM, Uddstrom MJ, Renwick JA, Merilä J (2008) The impact of climate fluctuation on food availability and reproductive performance of the planktivorous red-billed gull *Larus novaehollandiae scopulinus*. J Anim Ecol 77:1129-1142
- Montague TL, Cullen JM, Fitzherbert K (1986) The diet of the short-tailed shearwater *Puffinus tenuirostris* during its breeding season. Emu 86:207-213
- Mori Y, Boyd I (2004) Segregation of foraging between two sympatric penguin species: does rate maximization make the difference? Mar Ecol Prog Ser 275:241-249
- Navarro J, Votier SC, Aguzzi J, Chiesa JJ, Forero MG, Phillips RA (2013) Ecological segregation in space, time and trophic niche of sympatric planktivorous petrels. PLoS ONE 8(4): e62897
- Navarro J, Votier SC, Phillips RA (2014) Diving capabilities of diving petrels. Polar Biol 37:897-901
- Neira FJ, Miskiewicz AG, Trnski T (1998) Larvae of temperate australian fishes, Vol. University of Western Australia Press, Nedlands, Western Australia

- 665 O'Brien DP (1988) Surface schooling behaviour of the coastal krill *Nyctiphanes australis*
666 (Crustacea: Euphausiacea) off Tasmania, Australia. Mar Ecol Prog Ser 42:219-233
- 667 Payne MR, Prince PA (1979) Identification and breeding biology of the diving petrels
668 *Pelecanoides georgicus* and *P. urinatrix exsul* at South Georgia. NZ J Zool 6:299-318
- 669 Pescott TW (1976) Seabird islands No 27, Lady Julia Percy Island, Victoria. The Australian Bird
670 bander 14(1):29-31
- 671 Phillips RA, Croxall JP, Silk JRD, Briggs DR (2008) Foraging ecology of albatrosses and petrels
672 from South Georgia: Two decades of insights from tracking technologies. Aquat
673 Conserv: Mar Freshwat Ecosyst 17:S6-S21
- 674 Phillips RA, Silk JRD, Croxall JP (2005) Foraging and provisioning strategies of the light-
675 mantled sooty albatross at South Georgia: Competition and co-existence with sympatric
676 pelagic predators. Mar Ecol Prog Ser 285:259-270
- 677 Phillips RA, Silk JRD, Phalan B, Catry P, Croxall JP (2004) Seasonal sexual segregation in two
678 *Thalassarche* albatross species: Competitive exclusion, reproductive role specialization
679 or foraging niche divergence? Proc R Soc Lond, Ser B: Biol Sci 271:1283-1291
- 680 Polito MJ, Trivelpiece WZ, Reiss CS, Trivelpiece SG, Hinke JT, Patterson WP, Emslie SD
681 (2019) Intraspecific variation in a dominant prey species can bias marine predator dietary
682 estimates derived from stable isotope analysis. Limnol Oceanogr Methods 17(4):292-303
- 683 Pratte I, Robertson GJ, Mallory ML (2017) Four sympatrically nesting auks show clear resource
684 segregation in their foraging environment. MEPS 572:243-254

685 Prince JD (2001) Ecosystem of the South East Fishery (Australia), and fisher lore. Mar Freshw
686 Res 52:431-439

687 Prince PA, Morgan RA (1987) Diet and feeding ecology of procellariiformes. In: Croxall JP (ed)
688 Seabirds: Feeding ecology and role in marine ecosystems. Cambridge University Press,
689 Cambridge, p 135-171

690 Quillfeldt P, McGill RAR, Furness RW (2005) Diet and foraging areas of Southern Ocean
691 seabirds and their prey inferred from stable isotopes: Review and case study of Wilson's
692 storm-petrel. Mar Ecol Prog Ser 295:295-304

693 R Development Core Team (2018) R: A language and environment for statistical computing,
694 version 3.5.1. (R Foundation for Statistical Computing: Vienna)

695 Raymond B, Shaffer SA, Sokolov S, Woehler EJ, Costa DP, Einoder L, Hindell M, Hosie G,
696 Pinkerton M, Sagar PM, Scott D, Smith A, Thompson DR, Vertigan C, Weimerskirch H
697 (2010) Shearwater foraging in the Southern Ocean: the roles of prey availability and winds.
698 PLoS ONE 5(6):e10960

699 Reilly PN, Cullen JM (1981) The little penguin *Eudyptula minor* in Victoria, II: Breeding. Emu
700 81:1-19

701 Ridgeway KR, Condie SA (2004) The 5500-km-long boundary flow off western and southern
702 Australia. J Geophys Res 109:C04017

703 Ridgeway KR, Godfrey JS (1997) Seasonal cycle of the East Australian Current. J Geophys Res
704 102:22921-22936

- 705 Ridoux V (1994) The diets and dietary segregation of seabirds at the subantarctic Crozet Island.
706 Mar Ornithol 22:1-192
- 707 Ritz D, Swadling K, Hosie G, Cazassus F (2003) Guide to the zooplankton of south eastern
708 Australia. Fauna of Tasmania Committee, University of Tasmania, Hobart
- 709 Ropert-Coudert Y, Kato A, Chiaradia A (2009) Impact of small-scale environmental
710 perturbations on local marine food resources: a case study of a predator, the little
711 penguin. Proc Biol Sci 276:4105-4109
- 712 Ropert-Coudert Y, Wilson RP, Daunt F, Kato A (2004) Patterns of energy acquisition by a
713 central place forager: benefits of alternating short and long foraging trips. Behav Ecol
714 15:824-830
- 715 Ross GJB, Burbidge AA, Brothers N, Canty P, Dann P, Fuller PJ, Kerry KR, Norman FI,
716 Menkhorst PW, Pemberton D, Shaughnessy G, Shaughnessy PD, Smith GC, Stokes T,
717 Tranter J (2001) The status of Australia's seabirds. In: Zann LP, Kailola P (eds) The state
718 of the marine environment report for australia technical annex: 1 the marine environment.
719 Ocean Rescue 2000, Department of the Environment, Sport and Territories, Canberra, p
720 167-182
- 721 Sandery PA, Kämpf J (2005) Winter-spring flushing of Bass Strait, south-eastern Australia: A
722 numerical modelling study. Estuar Coast Shelf Sci 63:23-31
- 723 Sandery PA, Kämpf J (2007) Transport timescales for identifying seasonal variation in Bass
724 Strait, south-eastern Australia. Estuar Coast Shelf Sci 74:684-696
- 725 Schoener TW (1974) Resource partitioning in ecological communities. Science 185:27-39

726 Schumann N, Arnould JPY, Dann P (2008) Diet of common diving-petrels (*Pelecanoides*
727 *urinatrix urinatrix*) in southeastern Australia during chick rearing. *Waterbirds* 31:620-
728 624

729 Schumann N, Dann P, Arnould JPY (2014) The significance of northern-central Bass Strait in
730 south-eastern Australia as habitat for burrowing seabirds. *Emu* 114:234-240

731 Surman CA, Wooller RD (2003) Comparative foraging ecology of five sympatric terns at a sub-
732 tropical island in the eastern indian ocean. *J Zool Lond* 259:219-230

733 Sutton GJ, Hoskins AJ, and Arnould JP (2015) Benefits of group foraging depend on prey type
734 in a small marine predator, the little penguin. *PLoS One* 10:e0144297

735 Taylor AR, Dann P, Arnould JPY (2013) Timing of breeding and diet of the black-faced
736 cormorant *Phalacrocorax fuscescens*. *Mar Ornithol* 41:23-27

737 Tollit DJ, Steward MJ, Thompson PM, Pierce GJ, Santos MB, Hughes S (1997) Species and size
738 differences in the digestion of otoliths and beaks: Implications for estimates of pinniped
739 diet composition. *Can J Fish Aquat Sci* 54:105-119

740 Underwood MP (2012) Does size matter? Sex differences in white-faced storm petrels' ecology.
741 Deakin University, Australia

742 van der Lingen CD, Bertrand A, Bode A, Brodeur R, Cubillos LA, Espinoza P, Friedland K,
743 Garrido S, Irigoien X, Miller T, Mollmann C, Rodriguez-Sanchez R, Tanaka H,
744 Temming A (2009) Trophic dynamics in climate change and small pelagic fish, pp. 112-
745 157. Cambridge University Press, Cambridge, UK

- 746 Vertigan CA (2010) The life-history of short-tailed shearwaters (*Puffinus tenuirostris*) in
747 response to spatio-temporal environmental variation. University of Tasmania, Tasmania
- 748 Wakefield ED, Bodey TW, Bearhop S, Blackburn J, Colhoun K, Davies R, Dwyer RG, Green
749 JA, Gremillet D, Jackson AL, Jessopp MJ, Kane A, Langston RHW, Lescroel A, Murray
750 S, Le Nuz M, Patrick SC, Peron C, Soanes LM, Wanless S, Votier SC, Hamer KC (2013)
751 Space partitioning without territoriality in gannets. *Science* 341(6141):68-70
- 752 Waugh SM, Weimerskirch H (2003) Environmental heterogeneity and the evolution of foraging
753 behaviour in long ranging greater albatrosses. *Oikos* 103:374-384
- 754 Weimerskirch H, Cherel Y (1998) Feeding ecology of short-tailed shearwaters: Breeding in
755 Tasmania and foraging in the Antarctic? *Mar Ecol Prog Ser* 167:261-274
- 756 Woehler EJ, Raymond B, Watts DJ (2006) Convergence or divergence: where do short-tailed
757 shearwaters forage in the Southern Ocean? *MEPS* 324:261-270
- 758 Woehler EJ, Patterson TA, Bravington MV, Hobday AJ, Chambers LE (2014) Climate and
759 competition in abundance trends in native and invasive Tasmanian gulls. *MEPS* 511:249-
760 263
- 761 Young JW, Jordan AR, Bobbi C, Johannes RE, Haskard K, Pullen G (1993) Seasonal and
762 interannual variability in krill (*Nyctiphanes australis*) stocks and their relationship to the
763 fishery for jack mackerel (*Trachurus declivis*) off eastern Tasmania, Australia. *Mar Biol*
764 166:9-18

765 Zhang J, Rayner M, Vickers S, Landers T, Sagar R, Stewart J, Dunphy B (2018) GPS telemetry
 766 for small seabirds: using hidden Markov models to infer foraging behaviour of common
 767 diving petrels (*Pelecanoides urinatrix urinatrix*). Emu 119(2):126-137

Table 1(on next page)

Main species of seabirds breeding in Bass Strait, indicating the estimated number of breeding pairs, their proportion of the total Australian populations (based on Ross et al. 2001), and the major groups of prey consumed.

Some of the population estimates were not updated for at least three decades (e.g. Brothers et al. 2001) and may represent a source of error.

1

Species	Abundance (number of breeding pairs)	% of the Australian population	Groups of main prey	Reference
Shy albatross	5 200	35 %	Fish/cephalopods	Alderman et al. 2011, Hedd & Gales 2001
Short-tailed shearwater*	14 600 000	75 %	Crustaceans/Fish	Weimerskirch & Cherel 1998, Brothers et al. 2001, Schumann et al. 2014
Common diving petrel*	98 500	63 %	Crustaceans	Brothers et al. 2001, Schumann et al. 2008, Schumann et al. 2014
Fairy prion*	97 000	7 %	Crustaceans	Brothers et al. 2001, Schumann et al. 2014
White-faced storm petrel	94 500	25 %	Crustaceans	Brothers et al. 2001, Underwood 2012
Little penguin*	353 000	82 %	Fish	Cullen et al. 1992, Brothers et al. 2001, Dann & Norman 2006, Schumman et al. 2014
Australasian gannet	16 800	85 %	Fish	Bunce et al. 2002, Bunce 2001
Black faced cormorant	4 400	55 %	Fish	Brothers et al. 2001, Taylor et al. 2013
Pacific gull	1 500	82 %	Scavenge - polyvorous	Brothers et al. 2001, Leitch et al. 2014
Silver gull	50 000	35 %	Scavenge - polyvorous	Brothers et al. 2001, Leitch et al. 2014
Crested tern	10 400	13 %	Fish	Brothers et al. 2001, Chiaradia et al. 2012

*study species

2

Table 2(on next page)

Percentage of numerical abundance of the main groups of prey recovered from stomach contents of little penguins, short-tailed shearwaters, fairy prions and common diving petrels in Central Bass Strait.

1

	Little penguin (n = 42)	Short-tailed shearwater (n = 51)	Fairy prion (n = 20)	Common diving petrel (n = 6)
Fish (%)	74.1	1.8	>0.1	-
Cephalopods (%)	4.8	0.2	-	-
Crustaceans (%)	21.1	98.0	99.9	100

2

Table 3(on next page)

Summer mean (\pm SD) $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values (‰) in whole blood and isotopic niche metrics of little penguins, short-tailed shearwaters and fairy prions from western, central and eastern Bass Strait.

The samples were collected in summer (January-February) 2009, 2010 and 2011.

1

		Western Bass Strait			Central Bass Strait			Eastern Bass Strait		
		Little penguin	Short-tailed shearwater	Fairy prion	Little penguin	Short-tailed shearwater	Fairy prion	Little penguin	Short-tailed shearwater	Fairy prion
$\delta^{13}\text{C}$ (‰)	2009	-19.1 ± 0.4 (n = 10)	-21.9 ± 0.4 (n = 16)	-19.2 ± 0.2 (n = 8)	-19.8 ± 0.1 (n = 18)	-22.2 ± 0.3 (n = 20)	-	-19.0 ± 0.3 (n = 18)	-22.9 ± 0.5 (n = 20)	-
	2010	-19.5 ± 0.2 (n = 20)	-23.1 ± 0.3 (n = 20)	-19.5 ± 0.6 (n = 10)	-20.1 ± 0.4 (n = 19)	-22.2 ± 0.3 (n = 20)	-20.5 ± 0.7 (n = 6)	-19.0 ± 0.2 (n = 20)	-22.1 ± 0.4 (n = 20)	-
	2011	-18.9 ± 0.3 (n = 20)	-21.9 ± 0.4 (n = 20)	-18.8 ± 0.4 (n = 18)	-19.9 ± 0.2 (n = 10)	-22.0 ± 0.3 (n = 20)	-19.8 ± 0.3 (n = 17)	-18.5 ± 0.2 (n = 10)	-21.1 ± 0.5 (n = 13)	-
$\delta^{15}\text{N}$ (‰)	2009	15.5 ± 0.3 (n = 10)	9.5 ± 0.6 (n = 16)	13.7 ± 0.6 (n = 8)	14.4 ± 0.2 (n = 18)	9.3 ± 0.7 (n = 20)	-	12.9 ± 0.9 (n = 18)	9.4 ± 0.6 (n = 20)	-
	2010	13.2 ± 0.9 (n = 20)	8.5 ± 0.4 (n = 20)	13.6 ± 0.9 (n = 10)	13.4 ± 0.5 (n = 19)	8.6 ± 0.5 (n = 20)	11.7 ± 0.8 (n = 6)	13.6 ± 0.3 (n = 20)	8.8 ± 0.3 (n = 20)	-
	2011	15.0 ± 0.4 (n = 20)	9.7 ± 0.6 (n = 20)	12.5 ± 0.9 (n = 18)	14.9 ± 0.3 (n = 10)	9.9 ± 0.6 (n = 20)	13.2 ± 0.5 (n = 17)	13.6 ± 0.2 (n = 10)	9.9 ± 0.7 (n = 13)	-

2

Table 4(on next page)

Winter mean (\pm SD) $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values (‰) in whole blood and isotopic niche metrics of little penguins, fairy prions and common diving petrels from western, central and eastern Bass Strait.

The samples were collected in winter (July-August) 2008, 2009 and 2010.

1

		Western Bass Strait			Central Bass Strait			Eastern Bass Strait		
		Little penguin	Fairy prion	Common diving petrel	Little penguin	Fairy prion	Common diving petrel	Little penguin	Fairy prion	Common diving petrel
$\delta^{13}\text{C}$ (‰)	2008	-19.2 ± 0.4 (n = 3)	-	-	-19.0 ± 0.3 (n = 7)	-	-20.8 ± 0.8 (n = 10)	-	-	-
	2009	-20.0 ± 0.4 (n = 2)	-	-	-19.7 ± 0.1 (n = 20)	-	-20.8 ± 0.2 (n = 4)	-19.3 ± 0.5 (n = 20)	-	-
	2010	-19.3 ± 0.6 (n = 16)	-19.2 ± 0.4 (n = 18)	-19.6 ± 0.1 (n = 8)	-19.8 ± 0.1 (n = 20)	-19.5 ± 0.3 (n = 3)	-20.9 ± 0.3 (n = 15)	-19.2 ± 0.3 (n = 20)	-	-
$\delta^{15}\text{N}$ (‰)	2008	13.0 ± 0.1 (n = 3)	-	-	14.4 ± 0.4 (n = 7)	-	12.0 ± 1.1 (n = 10)	-	-	-
	2009	13.3 ± 0.4 (n = 2)	-	-	15.0 ± 0.4 (n = 20)	-	14.3 ± 0.2 (n = 4)	13.1 ± 1.2 (n = 20)	-	-
	2010	13.2 ± 0.6 (n = 16)	11.4 ± 1.4 (n = 18)	11.6 ± 0.3 (n = 8)	14.2 ± 0.3 (n = 20)	12.2 ± 0.3 (n = 3)	12.2 ± 0.2 (n = 15)	13.3 ± 0.6 (n = 20)	-	-

2

Figure 1

Simplified representation of the three study areas and the major water masses influencing the region.

Western Bass Strait (WBS); Central Bass Strait (CBS); Eastern Bass Strait (EBS); Lady Julia Percy Island (LJPI); Norman Island (NorI); Kanowna Island (KI); Answer Island (AI); Notch Island (NotI); Gabo Island (GI); South Australian Current (SAC); Sub-Antarctic Surface Water (SASW); East Australian Current (EAC) from Sandery & Kämpf 2005. The solid line indicates the location of the 300 m isobath.

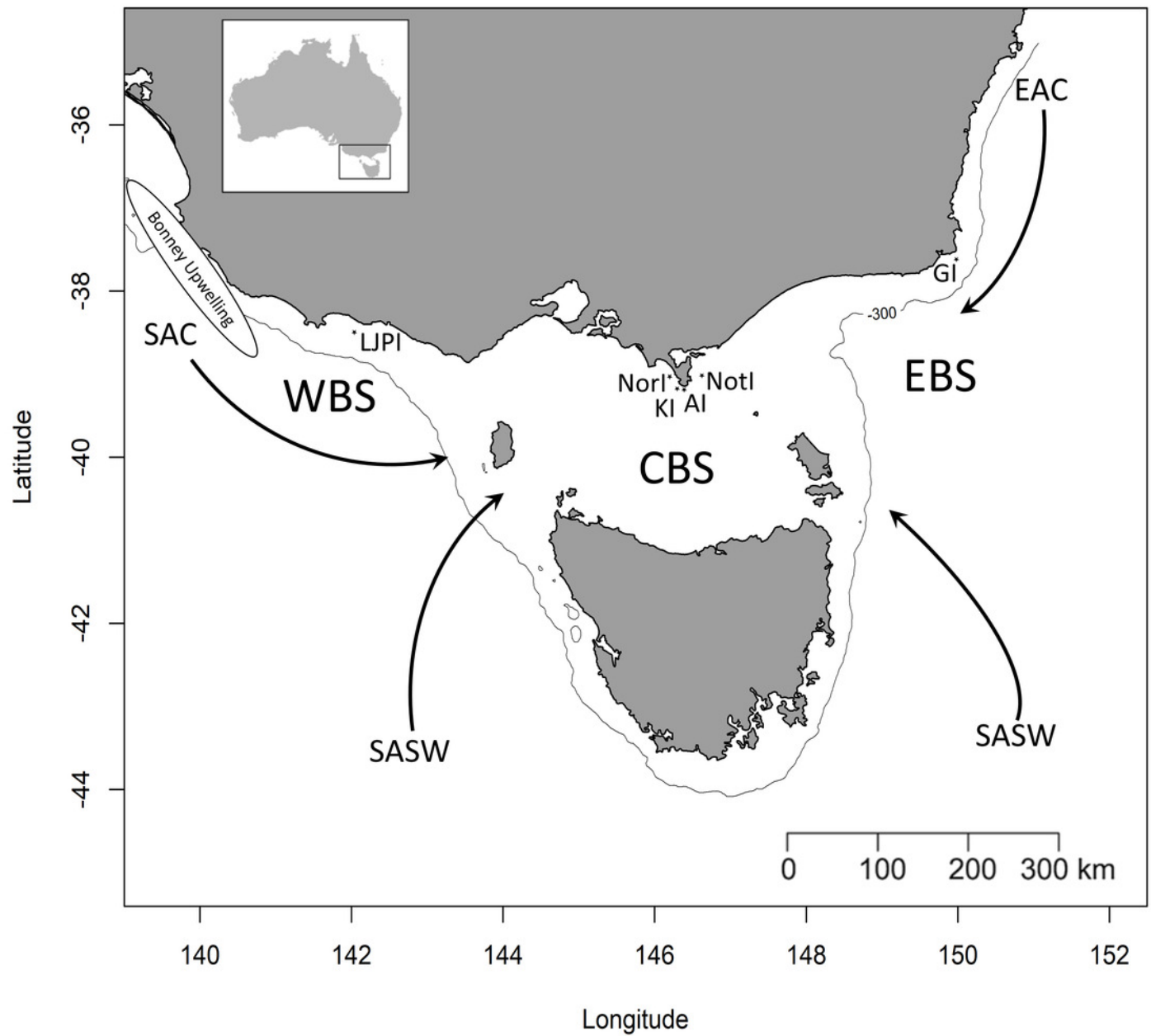


Figure 2

Phenology and sampling period of little penguin (LP), short-tailed shearwater (STSW), fairy prion (FP) and common diving petrel (CDP) in Bass Strait.

Blocks with vertical and horizontal lines correspond to incubation and chick-rearing period, respectively. Grey shaded blocks correspond to the winter and summer sampling periods.

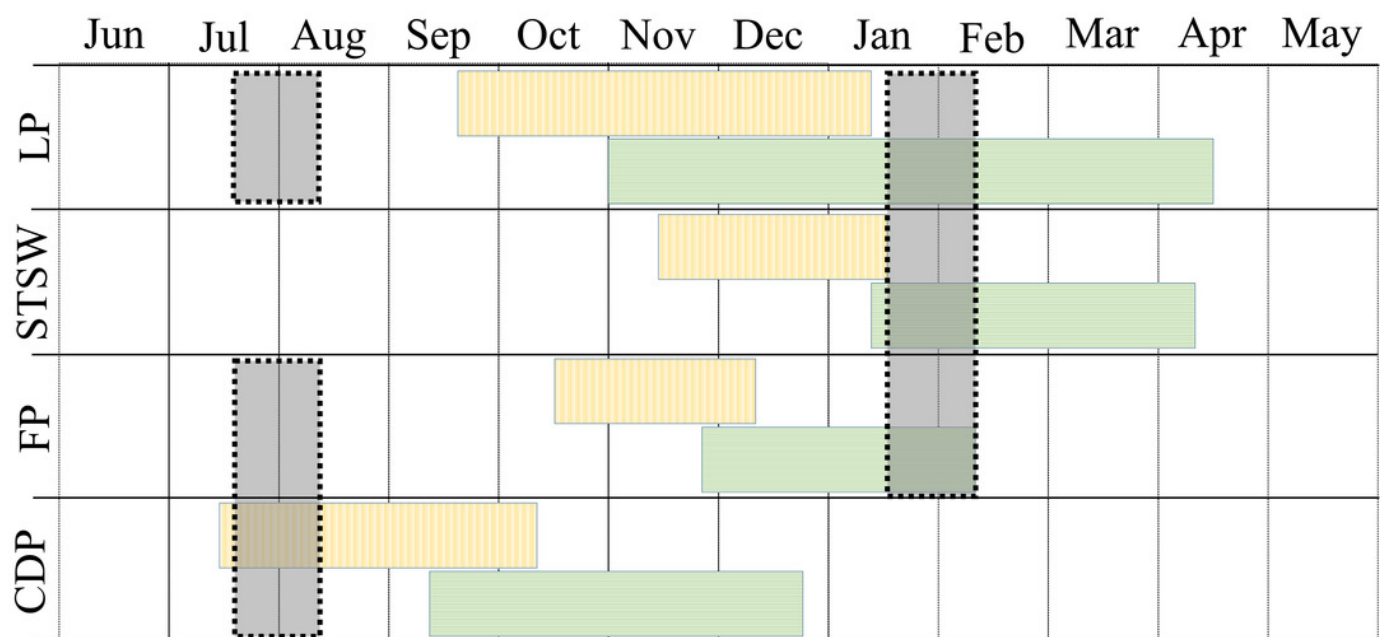


Figure 3

Summer inter-annual variation of $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values (‰) in whole blood of little penguins (A,B,C), short-tailed shearwaters (D,E,F) and fairy prions (G,H): western Bass Strait (WBS), central Bass Strait (CBS) and eastern Bass Strait

Solid lines represent the standard ellipses corrected for sample size (SEAc) based on $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values in summer 2009, 2010 and 2011. Note that the ranges for x and y axes are different for each species.

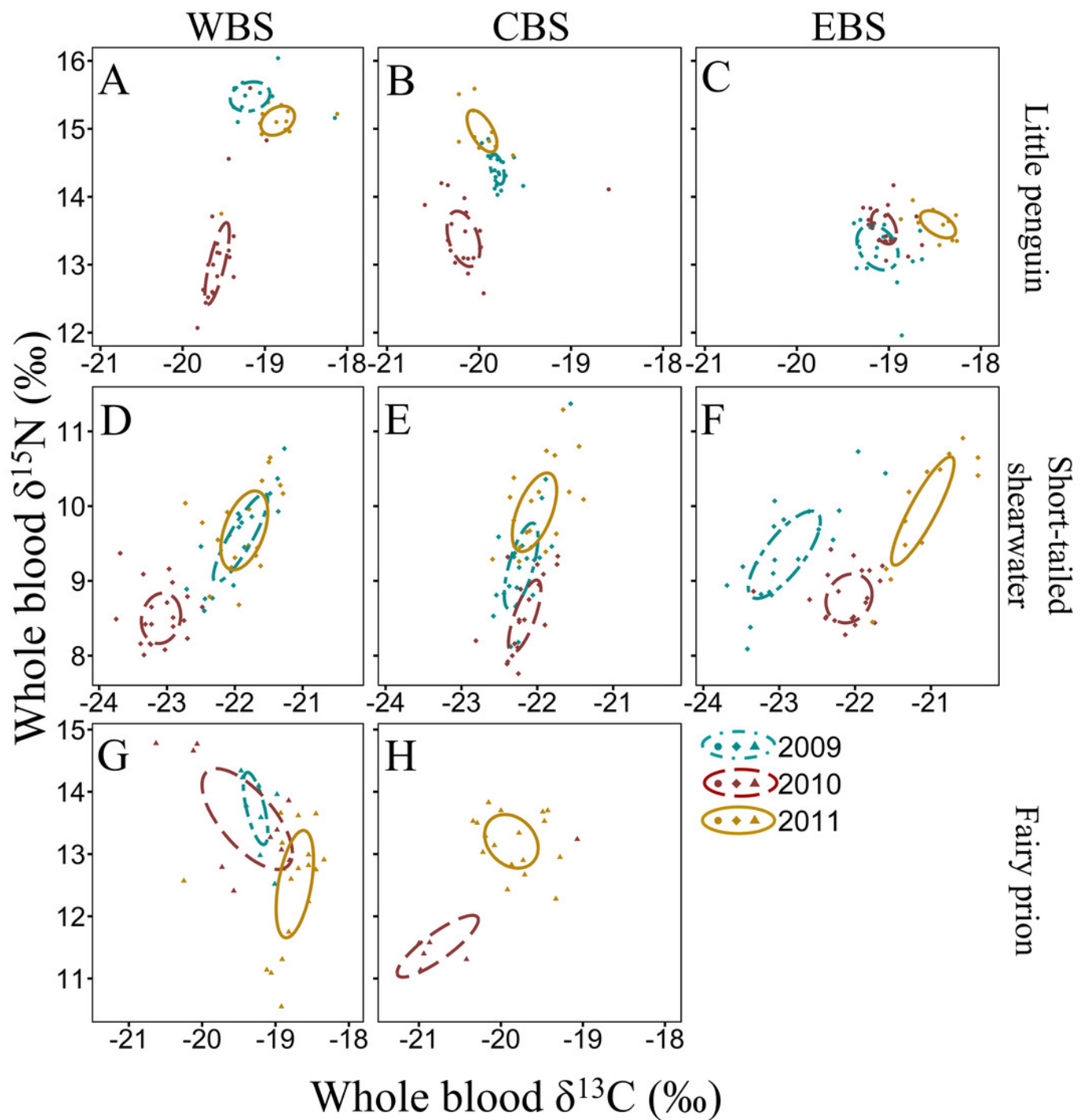


Figure 4

Winter inter-annual variation of $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values (‰) in whole blood of little penguins (A,B,C) and common diving petrels (D,E): western Bass Strait (WBS), central Bass Strait (CBS) and eastern Bass Strait (EBS).

Solid lines represent the standard ellipses corrected for sample size (SEAc) based on $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values in winter 2008, 2009 and 2010. Note that the range for x and y axes are different for each species.

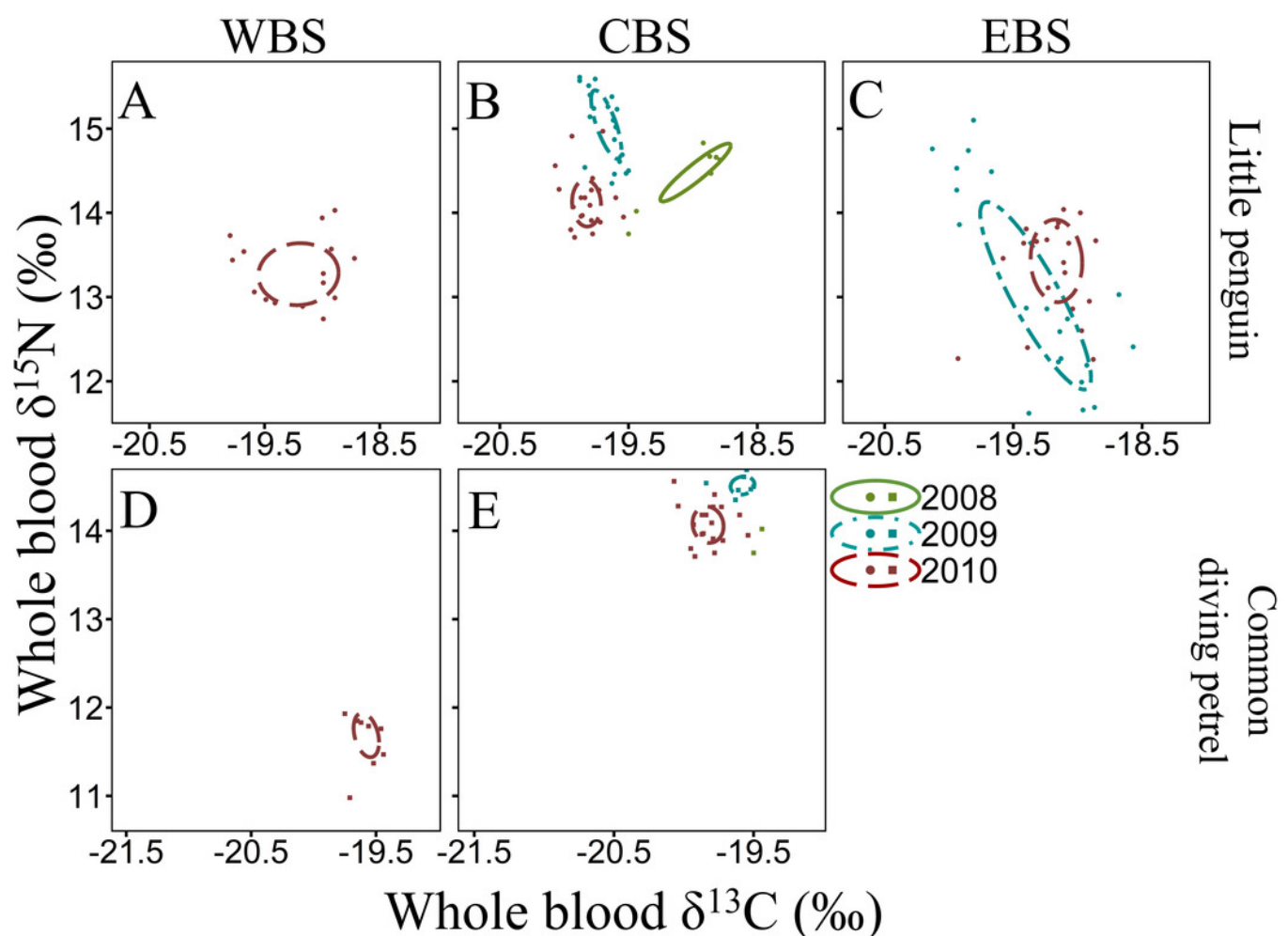


Figure 5

Summary of $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values (‰) in whole blood of the main marine predators in Bass Strait region.

Little penguin (LP, $n = 278$; present study; summer and winter combined), short-tailed shearwater (STSW, $n = 177$; present study; summer and winter combined), fairy prion (FP, $n = 88$; present study; summer and winter combined), common diving petrel (CDP, $n = 38$; present study; summer and winter combined), Australasian gannet (GA, $n = 27$; Angel et al. 2016; summer), white-faced storm petrel (WFST, $n = 17$; Underwood 2012; summer), shy albatross (SA, $n = 8$; Cherel et al. 2013; summer) and Australian fur seal (AFS, $n = 242$; Arnould et al. 2011; winter). The isotopic values of WFST and SA were calculated from data on chick feathers (Underwood 2012) and adult feathers (Cherel et al. 2013), respectively, and corrected using mean correction factors in Cherel et al. 2014.

