

1 **Single nucleotide polymorphism of *MTHFR* rs1801133 associated with elevated Hcy levels**
2 **affects susceptibility to cerebral small vessel disease**

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22 Abstract

23 **Background.** Methylenetetrahydrofolate reductase (MTHFR) is indispensable for the conversion
24 of homocysteine (Hcy) to methionine. The single nucleotide polymorphism (SNP) of MTHFR
25 gene (rs1801133, C667T) is correlated with decreased enzyme activity and eventually results in
26 elevated plasma Hcy levels. Hyperhomocysteinemia has been confirmed to be involved in the
27 pathogenesis of stroke, cerebral small vessel disease (CSVD), various metabolic disorders and so
28 on. However, the relationship among the MTHFR gene polymorphisms, Hcy, and CSVD has not
29 been investigated. In this study, the relationship between SNPs of MTHFR gene and CSVD was
30 investigated after adjusting cardiovascular risk factors, and the potential mechanism was
31 explored.

32 **Methods.** A total of 163 consecutive CSVD patients were collected as the case group. In the
33 corresponding period, 326 healthy people were selected as the control group, who were matched
34 to these cases according to age (± 2 years) and gender at the ratio of 2:1. SNPs of MTHFR
35 rs1801133, rs1801131, rs2274976, rs4846048, rs4846049, rs13306561 and rs3737964 were
36 genotyped with TaqMan Pre-Designed SNP Genotyping Assays. Plasma Hcy levels were detected

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37 using Hcy reagent through enzymatic cycling assay. Multivariate analysis was used to identify the
38 SNPs associated with CSVD susceptibility. Plasma Hcy levels were compared between different
39 genotypes.

40 **Results.** The MTHFR rs1801133 TT and CT genotype had increased risk for CSVD, and the *OR*
41 was higher in the TT genotype than in the CT genotype (2.307 vs 1.473). The plasma Hcy levels
42 of different genotypes showed the tendency of the TT genotype >CT genotype >CC genotype
43 (19.91±8.73 pg/ml vs 17.04±5.68 pg/ml vs 14.96±4.85 pg/ml).

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44 **Conclusions.** The SNP of MTHFR rs1801133 was correlated with CSVD, and the TT and CT
45 genotypes had increased risk for CSVD compared to the CC genotype. The potential mechanism
46 was associated with elevated Hcy levels.

47 **Key words:** Methylenetetrahydrofolate reductase; Single nucleotide polymorphism;
48 Homocysteine; Cerebral small vessel disease

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50 Introduction

51 As a generic term for intracranial vascular disease associated with various neurological and
52 pathological processes, cerebral small vessel disease (CSVD) refers to a syndrome of different
53 clinical features and neuroimaging findings induced by pathological changes in capillaries,
54 perforating cerebral arterioles and venules (Shi and Wardlaw, 2016). CSVD is the cause of 45%

55 of all the cases of dementia, and accounts for 25% of the ischemic stroke cases around the world

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56 (Pantoni, 2010; Petty et al., 2000; Wardlaw et al., 2013). Additionally, CSVD is a **major** cause for
57 depression, cognitive impairment, disability and so on in the aged (Li et al., 2018).

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58 The pathogenesis for CSVD mainly includes impairment of cerebral autoregulation, reduction of
59 cerebral blood flow and increase of blood-brain barrier (BBB) permeability (Joutel and
60 Chabriat, 2017; Li et al., 2018; Li et al., 2019). However, its molecular mechanisms are not
61 completely elucidated. Genetic studies have demonstrated that CSVD is highly heritable,
62 especially in young-onset stroke patients, and that disease processes of some CSVD subtypes
63 may be associated with common variants in monogenic disease genes (Tan et al., 2017).

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64 Methylene tetrahydrofolate reductase (MTHFR) gene is located on chromosome 1p36.3 and is
65 associated with the biosynthesis pathway of amino acid and purine (Goyette et al., 1994; Cui et
66 al., 2011). As an important regulatory enzyme catalyzing the transformation of 5,
67 10-methylene tetrahydrofolate to 5-methyl tetrahydrofolate, MTHFR is indispensable for the
68 conversion of homocysteine (Hcy) to methionine (Qin et al., 2012; Pogliani et al., 2015). The
69 single nucleotide polymorphism (SNP) of MTHFR gene (rs1801133, C667T) is correlated with
70 decreased enzyme activity that eventually results in elevated plasma homocysteine levels

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71 (Engbersen et al., 1995; Kang et al., 1993). **Hyperhomocysteinemia has been confirmed to be**
72 **involved in the pathogenesis of stroke, CSVD, various metabolic disorders and so on (Inamoto et**
73 **al., 2003; Zee et al., 2007; Chutinet et al., 2012; Qin et al., 2017; Nam et al., 2019; Piao et al.,**
74 **2018; Kloppenborg et al., 2011; Jeon et al., 2014; Pavlovic et al., 2011). In addition, a recent**
75 **study suggested that MTHFR C677T genotype was associated with CSVD subtype**

79 (Rutten-Jacobs et al., 2016). However, no previous studies have investigated the mechanism
80 associated with the effect of MTHFR C677T genotype on CSVD susceptibility. In this study, the
81 relationship between SNPs of MTHFR gene (rs1801133, rs1801131, rs2274976, rs4846048,
82 rs4846049, rs13306561 and rs3737964) and CSVD was investigated after adjusting
83 cardiovascular risk factors, and the potential mechanism was explored. The aim was to provide
84 useful clues for identifying susceptible populations of CSVD.

85

86 **Materials & Methods**

87 **Participants**

88 A total of 163 consecutive patients with CSVD were enrolled in the case group in Heze
89 Municipal Hospital between April 2017 and October 2018. In the corresponding period, 326
90 healthy people were selected as the control group, who were matched to these cases according to
91 age (± 2 years) and gender at the ratio of 2:1. This study received the permission of the ethic
92 committee of Heze Municipal Hospital (20160141022), and written informed consent was
93 provided by each participant.

94 The inclusion criteria of CSVD patients were: (1) in accordance with the diagnostic criteria
95 suggested by Shi Y and Wardlaw JM (Shi and Wardlaw, 2016); (2) to demonstrate typical
96 neuroimaging changes in the subcortical grey matter and white matter, including white matter
97 hyperintensities (WMHs), prominent perivascular spaces (PVS), cerebral microbleeds (CMBs),

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103 atrophy, [lacunas](#) and recent small subcortical infarct (Wardlaw et al., 2013); (3) complete clinical
104 data; (4) written informed consent. The exclusion criteria included (1) large-artery atherosclerosis;
105 (2) a definite history of subarachnoid hemorrhage or cerebral hemorrhage; (3) acute ischemic
106 stroke caused by cardiogenic embolism.

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107 Data Collection

108 Demographic data, vascular risk factors and laboratory indexes were collected in each participant.
109 Demographic data included age, gender, height, weight, annual family income, education level
110 and occupation. Vascular risk factors included hypertension, hyperlipemia, diabetes, smoking,
111 drinking and blood pressure. Laboratory indexes included total cholesterol, triacylglycerol (TG),
112 low density lipoprotein cholesterol (LDL-C), high density lipoprotein cholesterol (HDL-C) and
113 fasting blood glucose (FBG).

114 SNP genotyping for MTHFR gene

115 The DNA used in SNP genotyping was extracted from peripheral blood using the salting out
116 method suggested by Hashemi et al (Hashemi et al., 2010). TaqMan Pre-Designed SNP
117 Genotyping Assays (Applied Biosystems, Carlsbad, USA) was employed to perform SNP
118 genotyping for the MTHFR gene (rs1801133, rs1801131, rs2274976, rs4846048, rs3737966,
119 rs1537515, rs4846049, rs3834044, rs13306561 and rs3737964). The ABI 7500 Fast real-time
120 PCR system (Applied Biosystems, Carlsbad, USA) was employed to perform PCR amplification
121 and allelic discrimination.

124 Plasma Hcy assay

125 A blood sample was collected from the antecubital vein after an overnight fasting in each
126 participant. The blood sample was then separated through centrifugation at 3000g for 5 min. The
127 level of Hcy was detected by enzymatic cycling assay using Hcy reagent (Beijing Strong
128 Biotechnologies, Inc, Beijing, China) and the ROCHE Cobas 8000 automatic biochemical
129 analyzer (Roche Ltd., Switzerland).

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Deleted: and Hcy reagent (Beijing Strong Biotechnologies, Inc, Beijing, China) through enzymatic cycling assay.

130 Statistical analysis

131 The SNPstats software, a web tool for the analysis of association studies:
132 <http://bioinfo.iconcologia.net/SNPStats>, was employed to perform Hardy-Weinberg equilibrium
133 (HWE) test and to calculate allele frequencies and genotype frequencies for all the SNPs (Solé et
134 al., 2006). Univariate analysis was performed with Student's *t*-test or chi-square test for all
135 variables, including demographic data, vascular risk factors, laboratory indexes and SNP
136 genotyping data. The variables with a *P* value less than 0.10 were then included in the
137 multivariate analysis, which was used to identify the SNPs associated with CSVD susceptibility
138 through a backward stepwise logistic regression model. Plasma Hcy levels among different
139 genotypes were compared using ANOVA. All statistical analysis was conducted using the SPSS
140 version 20.0 for Windows (SPSS Inc., USA), and significance was set at $P < 0.05$.

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142 Results

147 **Univariate analysis of demographic data, vascular risk factors and laboratory indexes**

148 The case group included 94 males and 69 females, and the control group included 188 males and
149 138 females. Their average age was (63.28±7.09) years. Univariate analysis demonstrated that
150 age, body mass index (BMI), annual family income, education level, occupation, drinking,
151 hyperlipidaemia, total cholesterol, TG, LDL-C and HDL-C were not statistically different
152 between the case group and the control group, and hypertension, diabetes, smoking, systolic
153 blood pressure (SBP), diastolic blood pressure (DBP) and FBG were statistically different (*Table*
154 *1*).

155 **SNP analysis**

156 All the SNPs were successfully genotyped in both the case group and the control group. As
157 shown in *Table 2*, the genotype frequencies of all the SNPs were not statistically different from
158 those evaluated using Hardy-Weinberg equilibrium. Univariate analysis demonstrated that the
159 genotype frequencies of rs1801133 ($\chi^2=12.852$, $P=0.002$) and rs1801131 ($\chi^2=6.203$, $P=0.045$)
160 were statistically different between the case group and the control group, and rs2274976,
161 rs4846048, rs4846049, rs13306561 and rs3737964 were not statistically different (all $P>0.05$).

162 **Multivariate analysis**

163 Multivariate analysis was conducted to identify the independent association between different
164 genotypes of the MTHFR rs1801133 and rs1801131 and CSVD. The results demonstrated that
165 the polymorphism of rs1801133 was correlated with CSVD after adjusting hypertension, diabetes,

166 smoking, SBP, DBP, FBG, hyperlipidaemia, LDL-C and HDL-C, but rs1801131 was not (*Table*
167 3). The MTHFR rs1801133 TT and CT genotype had increased risk for CSVD, and the *OR* was
168 higher in the TT genotype than in the CT genotype (2.307 vs 1.473).

169 Plasma Hcy levels

170 Plasma Hcy levels were compared using ANOVA among the MTHFR rs1801133 TT, CT and CC
171 genotype. The results demonstrated that plasma Hcy levels were highest in the TT genotype,
172 intermediate in the CT genotype, and lowest in the CC genotype (*Table 4*).

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174 Discussion

175 As a sulfur-containing amino acid, Hcy is an important intermediate product for the metabolism
176 of methionine and has an important role in vascular function (Li et al., 2017). High Hcy levels
177 can predispose vascular smooth muscle cells and endothelial cells to injury, which results in
178 activation of coagulation factors, expression of plasminogen activator inhibitor, endothelial
179 proliferation and so on (Hainsworth et al., 2016). This further inhibits the expression of
180 thrombomodulin and synthesis of tissue-type plasminogen activator and sulfated heparin,
181 eventually leading to atherogenesis and thrombogenesis via secretion of inflammatory
182 cytokines, platelet aggregation, endoplasmic reticulum stress, and oxidative stress.
183 Studies showed that Hyperhomocysteinemia (HHcy) is associated with multiple diseases,
184 including ischemic stroke (IS), CSVD and various metabolic disorders and so on. Pavlovic et al.

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193 showed that elevated total Hcy was correlated with clinical status and severity of white matter
194 changes in symptomatic patients with subcortical small vessel disease (Pavlovic et al., 2011).
195 HHcy has [also](#) been confirmed as an independent risk factor for IS (Wu et al., 2016; Boysen et al.,
196 2003). Wu et al. demonstrated that high Hcy levels were associated with a greater incidence of
197 acute cerebral infarction among patients with carotid artery plaques (Wu et al., 2016). Ji et al.
198 reported that high Hcy levels were associated with a poor functional outcome, severe
199 neurological impairment, and stroke recurrence in large artery atherosclerosis stroke subtype,
200 which confirmed the atherogenic effect of Hcy (Ji et al., 2015). Lu et al. demonstrated that high
201 Hcy levels were correlated with strong plaque enhancement and acute ischemic stroke after
202 adjusting sex, age, serum creatinine levels and other atherosclerotic risk factors (Lu et al., 2018).
203 Several previous cohort studies also showed that high Hcy levels were correlated with increased
204 risk of IS, including the British Regional Heart Study, the Framingham Study and the Northern
205 Manhattan cohort study (Perry et al., 1995; Bostom et al., 1999; Sacco et al., 2004). As a risk
206 factor of atherosclerosis, high Hcy levels are associated with white matter lesions, lacunar
207 infarcts and cognitive impairment. Kioppborg et al. (Kloppenborg et al., 2011) found that a
208 higher Hcy level was associated with presence of lacunar infarcts and a higher volume of white
209 matter lesions among patients with symptomatic atherosclerotic disease. Piao et al. (Piao et al.,
210 2018) evaluated the association between Hcy levels and CSVD with the method of meta-analysis.
211 Their results demonstrated that Hcy levels were higher in CSVD patients than in healthy controls.
212 Nam et al. (Nam et al., 2019) found that serum Hcy levels were associated with the presence of
213 cerebral microbleeds, white matter hyperintensity volume enlarged perivascular spaces in a

214 healthy population. In addition, Hcy levels may be correlated with the susceptibility for NAFLD
215 (Hu et al., 2016; Polyzos et al., 2015).

216 MTHFR is a key controlling enzyme involved in the metabolism of Hcy and folate. It is
217 indispensable for the conversion of homocysteine to methionine via catalyzing the transformation
218 of 5, 10-methylenetetrahydrofolate to 5-methyltetrahydrofolate (Qin et al., 2012; Pogliani et al.,
219 2015). Additionally, it has a role in chromosomal integrity, DNA methylation and maintaining the
220 stability of single- and double-strand DNA (Robien & Ulrich, 2003). It is encoded by the
221 MTHFR gene, which is located on chromosome 1p36.3 (Goyette et al., 1994). For the MTHFR
222 gene, the cytosine (C) to thymine (T) substitution at position 677 (rs1801133) in the gene
223 encoding region is the most common SNP. This variation leads to the conversion from alanine to
224 valine at amino acid 222 (Jadavji et al., 2015), and is correlated with decrease of thermal stability
225 of MTHFR and subsequent decrease of enzyme activity (Atadzhanov et al., 2013; Ou et al., 2014).

226 Compared to the CC genotype, the enzyme activity of the CT and TT genotypes is less than 35%
227 and 70%, respectively (Frosst et al., 1995). The decreased enzyme activity eventually results in
228 the elevation of Hcy levels (Atadzhanov et al., 2013; Ou et al., 2014). In other words, the CT and
229 TT genotypes are correlated with elevated Hcy levels through reducing the activity of MTHFR.

230 Wang et al. (Wang et al., 2018) found that the SNP of the MTHFR rs1801133 and NAFLD have a
231 potential synergistic effect on elevated Hcy levels. Li et al. (Li et al., 2017) found that the plasma
232 Hcy levels of different genotypes of the MTHFR rs1801133 showed the tendency of the TT
233 genotype >CT genotype >CC genotype. They concluded that a possible synergistic effect of the

234 MTHFR rs1801133 SNP on plasma Hcy levels increased risk of IS. In addition, Rutten-Jacobs et
235 al. demonstrated that MTHFR C677T genotype was associated with CSVD subtype
236 (Rutten-Jacobs et al., 2016).

237 In this study, the SNP of the MTHFR rs1801133 was correlated with CSVD, and the TT and CT
238 genotypes had increased risk for CSVD compared to the CC genotype. Moreover, the [overall](#)
239 [response \(OR\)](#) was higher in the TT genotype than in the CT genotype. At the same time, the
240 plasma Hcy levels of different genotypes showed the tendency of the TT genotype >CT
241 genotype >CC genotype. Therefore, the SNP of rs1801133 was correlated with CSVD through
242 elevating Hcy levels.

243 The inclusion and exclusion criteria of CSVD patients used in this paper included
244 clinical symptoms, signs and typical neuroimaging changes. Additionally, CSVD patients should
245 have complete clinical data. The aim was to evaluate the relationship between SNPs of MTHFR
246 gene and CSVD comprehensively and precisely. The main limitations of this study included a
247 small sample size and not studying the joint effect of MTHFR rs1801133 and rs1801131 on
248 CSVD susceptibility and Hcy levels. We will investigate their joint effects using a larger sample
249 size in the next step.

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251 Conclusions

252 The SNP of the MTHFR rs1801133 was correlated with CSVD, and the TT and CT genotypes

253 had increased risk for CSVD compared to the CC genotype. The potential mechanism was
254 associated with elevated Hcy levels.

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256 Acknowledgments

257 None.

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