

A comprehensive analysis for associations between multiple microRNAs and prognosis of osteosarcoma patients

Wen Yang^{1,2}, Yu-bin Qi³, Meng Si¹, Yong Hou¹, Lin Nie^{Corresp. 1}

¹ Department of Orthopaedics, Qilu Hospital of Shandong University, Jinan, Shandong Province, China

² Department of Spinal Surgery, Heze Municipal Hospital, Heze, Shandong Province, China

³ Department of Orthopaedics, Shandong Provincial Qianfoshan Hospital, Jinan, Shandong Province, China

Corresponding Author: Lin Nie

Email address: nielinforest@163.com

Background. Osteosarcoma is the most common malignant primary bone tumor occurring in children and young adults, which occupies the second important cause of tumor-associated deaths among children and young adults. Recent studies have demonstrated that many microRNAs have abnormal expression in osteosarcoma, and can function as prognostic factors of osteosarcoma patients. However, no previous studies have comprehensively analyzed the relationship between multiple miRNAs and prognosis of osteosarcoma patients. **Methods.** A total of 63 osteosarcoma patients were retrospectively enrolled. The clinical characteristics were collected, and the expression levels of microRNA-21, microRNA-30c, microRNA-34a, microRNA-101, microRNA-133a, microRNA-214, microRNA-218, microRNA-433 and microRNA-539 in tumor tissues were measured through quantitative real-time polymerase chain reaction. Kaplan-Meier analysis was used to perform univariate survival analysis, and Cox regression model was used to perform multivariate survival analysis which included the variables with $P < 0.1$ in univariate survival analysis. **Results.** The cumulative survival for 1, 2 and 5 years was 90.48%, 68.25% and 38.10%, respectively, and mean survival time was (45.39 ± 3.60) months (95% confidence interval: 38.34-52.45). Kaplan-Meier analysis demonstrated that TNM stage, metastasis or recurrence, microRNA-21, microRNA-214, microRNA-34a, microRNA-133a and microRNA-539 were correlated with cum survival, but gender, age, tumor diameter, differentiation, microRNA-30c, microRNA-433, microRNA-101 and microRNA-218 were not. Multivariate survival analysis demonstrated that microRNA-21 (hazard ratio: 3.457, 95% confidence interval: 2.165-11.518), microRNA (hazard ratio: 3.138, 95% confidence interval: 2.014-10.259), microRNA-34a (hazard ratio: 0.452, 95% confidence interval: 0.202-0.915), microRNA-133a (hazard ratio: 0.307, 95% confidence interval: 0.113-0.874) and microRNA-539 (hazard ratio: 0.358, 95% confidence interval: 0.155-0.896) were independent prognostic markers of osteosarcoma patients after adjusting for TNM stage

(*hazard ratio: 2.893, 95% confidence interval: 1.496-8.125*), metastasis or recurrence (*hazard ratio: 3.628, 95% confidence interval: 2.217-12.316*) and microRNA-30c (*hazard ratio: 0.689, 95% confidence interval: 0.445-1.828*). **Conclusions.** High expression of microRNA-21 and microRNA-214 and low expression of microRNA-34a, microRNA-133a and microRNA-539 were associated with poor prognosis of osteosarcoma patients after adjusting for TNM stage, metastasis or recurrence and microRNA-30c.

A comprehensive analysis for associations between multiple microRNAs and prognosis of osteosarcoma patients

Wen Yang^{1, 2}, Yu-bin Qi³, Meng Si¹, Yong Hou¹, Lin Nie¹

1. Department of Orthopaedics, Qilu Hospital of Shandong University, Jinan 250012, China

2. Department of Spinal Surgery, Heze Municipal Hospital, Heze 274031, China

3. Department of Orthopaedics, Shandong Provincial Qianfoshan Hospital, Jinan 250014, China

Address correspondence to:

Lin Nie

Department of Orthopaedics, Qilu Hospital of Shandong University

No. 107, Wenhua Xi Road, Jinan 250012, China

Tel: +86-531-82166551

Email: nielinforest@163.com

Abstract

Background. Osteosarcoma is the most common malignant primary bone tumor occurring in children and young adults, which occupies the second important cause of tumor-associated deaths among children and young adults. Recent studies have demonstrated that many microRNAs have abnormal expression in osteosarcoma, and can function as prognostic factors of osteosarcoma patients. However, no previous studies have comprehensively analyzed the relationship between multiple miRNAs and prognosis of osteosarcoma patients.

Methods. A total of 63 osteosarcoma patients were retrospectively enrolled. The clinical characteristics were collected, and the expression levels of microRNA-21, microRNA-30c, microRNA-34a, microRNA-101, microRNA-133a, microRNA-214, microRNA-218, microRNA-433 and microRNA-539 in tumor tissues were measured through quantitative real-time polymerase chain reaction. Kaplan-Meier analysis was used to perform univariate survival analysis, and Cox regression model was used to perform multivariate survival analysis which included the variables with $P < 0.1$ in univariate survival analysis.

Results. The cumulative survival for 1, 2 and 5 years was 90.48%, 68.25% and 38.10%, respectively, and mean survival time was (45.39 ± 3.60) months (95% confidence interval: 38.34-52.45). Kaplan-Meier analysis demonstrated that TNM stage, metastasis or recurrence, microRNA-21, microRNA-214, microRNA-34a, microRNA-133a and microRNA-539 were correlated with cum survival, but gender, age, tumor diameter, differentiation, microRNA-30c, microRNA-433, microRNA-101 and microRNA-218 were not. Multivariate survival analysis

demonstrated that microRNA-21 (*hazard ratio*: 3.457, 95% *confidence interval*: 2.165-11.518), microRNA (*hazard ratio*: 3.138, 95% *confidence interval*: 2.014-10.259), microRNA-34a (*hazard ratio*: 0.452, 95% *confidence interval*: 0.202-0.915), microRNA-133a (*hazard ratio*: 0.307, 95% *confidence interval*: 0.113-0.874) and microRNA-539 (*hazard ratio*: 0.358, 95% *confidence interval*: 0.155-0.896) were independent prognostic markers of osteosarcoma patients after adjusting for TNM stage (*hazard ratio*: 2.893, 95% *confidence interval*: 1.496-8.125), metastasis or recurrence (*hazard ratio*: 3.628, 95% *confidence interval*: 2.217-12.316) and microRNA-30c (*hazard ratio*: 0.689, 95% *confidence interval*: 0.445-1.828).

Conclusions. High expression of microRNA-21 and microRNA-214 and low expression of microRNA-34a, microRNA-133a and microRNA-539 were associated with poor prognosis of osteosarcoma patients after adjusting for TNM stage, metastasis or recurrence and microRNA-30c.

Key words: MicroRNAs; Survival; Kaplan-Meier analysis; Multivariate Cox regression analysis

Introduction

Osteosarcoma (OS) is the most common malignant primary bone tumor occurring in children and young adults, which occupies the second important cause of tumor-associated deaths among children and young adults (Mirabello et al., 2009; Mirabello et al., 2009; Biermann et al., 2013; Yu et al., 2017). It is highly aggressive and occurs mainly in the proximal tibia, proximal humerus, and metaphyseal regions of the distal femur, with an incidence of 4.4 per million people around the world (Zhu et al., 2016). OS responds poorly to chemotherapy and the 5-year

survival rate is still very low for OS patients with metastasis or recurrence (Hutanu et al., 2017; Zhou et al., 2016), although its prognosis has been improved gradually over the past 30 years (Rytting et al., 2000; Kunz et al., 2015). Therefore, it is crucial to identify new biomarkers that can exactly evaluate the prognosis of OS.

MicroRNAs (miRNAs) are a group of non-coding RNAs, which consist of 18-25 nucleotides (Ambros, 2004; Chang et al., 2016; Jamieson et al., 2012). They widely exist in animals, plants and even some viruses, and have an important role in post-transcriptional modulation of gene expression and gene silencing (Bartel, 2004; Hayes et al., 2014; Griffiths-Jones et al., 2008; Liu et al., 2017). Approximately 50% of miRNAs are confirmed to be associated with human tumorigenesis through directly targeting tumor suppressor genes or oncogenes (Li & Rana, 2014; Bracken et al., 2016). MiRNAs are able to be circulated in body fluid, suggesting their potential as non-invasive markers (Bahrami et al., 2018). In OS, abnormal expression of miRNAs is involved in its occurrence and development. In addition, the expression of some miRNAs is associated with OS chemoresistance. Therefore, miRNAs have been widely applied in prediction of prognosis, detection of patients at early stages, and monitoring of the patients in response to chemotherapy. Studies have demonstrated that many miRNAs can function as prognostic factors of OS patients (Cheng et al., 2017; Zhang et al., 2015). Among them, miRNA-21, miRNA-30c, miRNA-34a, miRNA-101, miRNA-133a, miRNA-214, miRNA-218, miRNA-433 and miRNA-539 have been studied extensively and confirmed a potential association with the prognosis of OS patients. However, no previous studies have comprehensively analyzed the relationship between multiple miRNAs and prognosis of OS patients. There may be interactions

among them. In this study, the expression levels of these 9 miRNAs in tumor tissues of OS patients were measured through quantitative real-time PCR (qRT-PCR). Kaplan-Meier method was employed to determine the survival rate of OS patients, and long-rank test was employed to compare the survival rates between groups. Multivariate Cox regression analysis was finally performed to identify the independent prognostic factors with adjusting for confounders.

Materials & Methods

Patients

A total of 63 OS patients were retrospectively collected in Heze Municipal Hospital between January 2012 and January 2018. Surgery was performed in all of them, and tumor tissues and adjacent normal bone tissues were sampled. None of them received chemotherapy and radiotherapy before surgery. All tissue samples, obtained during surgery, were frozen immediately in liquid nitrogen and stored at -80°C. The diagnosis and histological grading were determined with histopathological examination. This study received the approval of the ethic committee of Heze Municipal Hospital (20185261), and was performed according to the Declaration of Helsinki. All patients provided written informed consents.

Quantitative real-time PCR

Total RNA was extract from tumor tissues and adjacent normal bone tissues through miRNeasy kit (Qiagen, Germany) in accordance with instructions of the manufacturer. The TaqMan miRNA assey kit (Applied Biosystems, USA) was used to quantitate the expression levels of miRNAs. Rotor Gene 6000 Real-Time PCR (Qiagen, Germany) was used to perform Real-Time

106 PCR with a TaqMan universal PCR master mix and an invitrogen kit. U6 was chosen as the
 107 reference gene, and the $2^{-\Delta\Delta C_t}$ method was used to assess the relative expression levels of
 108 miRNAs. The primers of the included miRNAs and U6 were designed and chemosynthesized by
 109 Shanghai Jima Biotech Ltd (Shanghai, China). The primers used were as follows: miRNA-21-3p:
 110 5'-GCCACCACACCAGCTAATTT-3' (forward) and 5'-CTGAAGTCGCCATGCAGATA-
 111 3' (reverse); miRNA-30c-3p: 5'-GCCCCAAGTGGTTCTGTGTTT-3' (forward) and 5'-
 112 TCCATGGCAGAAGGAGTAAA-3' (reverse); miRNA-34a-5p: 5'-
 113 TATGGCAGTGTCTTAGCTGGTTGT-3' (forward) and 5'-GGCCAACCGCGAGAAGATG-3'
 114 (reverse); miRNA-101-3p: 5'-GCCGAGTACAGTACTGTGA-3' (forward) and 5'-
 115 CTCAACTGGTGTCGTGGA-3' (reverse); miRNA-133a-5p: 5'-
 116 TGCTTTGCTAGAGCTGGTAAAATG-3' (forward) and 5'-AGCTACAGCTGGTTGAAGGG-
 117 3' (reverse); miRNA-214-3p: 5'-TGCAGTAGTGTCTTAGCTGGAATG-3' (forward) and 5'-
 118 GGCTAACCGCGAGAAGTTT-3' (reverse); miRNA-218-5p: 5'-
 119 GCGCTTGTGCTTGATCTAA-3' (forward) and 5'-GTGCAGGGTCCGAGGT-3' (reverse);
 120 miRNA-433-3p: 5'-GCTTTAGTGGTTCTGTGTGA-3' (forward) and 5'-
 121 TCCGCGACAGAAGGAGTTTA-3' (reverse); miRNA-539-3p: 5'-
 122 GCTTGTACACCAGCTAGTGC-3' (forward) and 5'-CTTAGCTCGCCATGCAGAAG-
 123 3' (reverse); and U6: 5'-GATCAAGGATGACAC GCAAATTCG-3' (forward) and 5'-
 124 GGCCAACCGCGAGAAGATG-3' (reverse).

125 Statistical analysis

126 Statistical analysis was conducted using the SPSS version 20.0 for Windows (SPSS Inc., USA).

Kolmogorov-Smirnov test was used to determine the normality of quantitative data. Normal data were expressed as mean \pm standard deviation (SD), and non-normal data were expressed as median (interquartile range). Qualitative data were expressed as percentages or ratios (%). Kaplan-Meier analysis was used to perform univariate survival analysis, and Cox regression model was used to perform multivariate survival analysis which included the variables with $P < 0.1$ in univariate survival analysis. Significance was set at $P < 0.05$.

Results

General data

These 63 osteosarcoma patients included 36 males and 27 females, and the median age of onset for them was 17 years with an interquartile range of 10 years. The other detailed clinical characteristics were demonstrated in *Table 1*. The follow-up was up to January 2019. The cumulative survival for 1, 2 and 5 years was 90.48%, 68.25% and 38.10%, respectively, and mean survival time was (45.39 \pm 3.60) months (95% *confidence interval*: 38.34-52.45).

Expression levels of microRNAs in tumor tissues and adjacent normal bone tissues

According to the results of quantitative real-time polymerase chain reaction (*Table 2* and *Fig. 1*), the expression levels of microRNA-21, microRNA-214 and microRNA-433 were higher in tumor tissues than in adjacent normal bone tissues, and the expression levels of microRNA-30c, microRNA-34a, microRNA-101, microRNA-133a and microRNA-539 was lower in tumor tissues than in adjacent normal bone tissues, and the expression level of microRNA-218 was not statistically different.

Univariate survival analysis

The osteosarcoma patients were divided into high expression group and low expression group according to the median expression levels of microRNAs. Kaplan-Meier analysis demonstrated that TNM stage (Fig. 2), metastasis or recurrence (Fig. 3), microRNA-21 (Fig. 4A), microRNA-214 (Fig. 4B), microRNA-34a (Fig. 4C), microRNA-133a (Fig. 4D) and microRNA-539 (Fig. 4E) were correlated with cum survival, but gender, age, tumor diameter, differentiation, microRNA-30c (Fig. 4F), microRNA-433, microRNA-101 and microRNA-218 were not. Median time of survival and log rank χ^2 were demonstrated in Table 3.

Multivariate survival analysis

TNM stage, metastasis or recurrence, microRNA-21, microRNA-214, microRNA-30c, microRNA-34a, microRNA-133a and microRNA-539 were included in Cox proportional hazards model. According to the results of multivariate survival analysis (Table 4), microRNA-21 (hazard ratio: 3.457, 95% confidence interval: 2.165-11.518), microRNA-214 (hazard ratio: 3.138, 95% confidence interval: 2.014-10.259), microRNA-34a (hazard ratio: 0.452, 95% confidence interval: 0.202-0.915), microRNA-133a (hazard ratio: 0.307, 95% confidence interval: 0.113-0.874) and microRNA-539 (hazard ratio: 0.358, 95% confidence interval: 0.155-0.896) were independent prognostic markers of osteosarcoma patients after adjusting for TNM stage (hazard ratio: 2.893, 95% confidence interval: 1.496-8.125), metastasis or recurrence (hazard ratio: 3.628, 95% confidence interval: 2.217-12.316) and microRNA-30c (hazard ratio: 0.689, 95% confidence interval: 0.445-1.828). In other words, high expression of microRNA-21 and microRNA-214 and low expression of microRNA-34a, microRNA-133a and microRNA-539

were associated with poor prognosis of osteosarcoma patients.

Discussion

The prognosis of OS patients has been significantly improved with the development of multiple chemotherapy regimens. However, OS patients receiving the same treatment often demonstrate different clinical outcomes, suggesting an urgent need for developing reliable prognostic biomarkers to improve the prognosis of OS patients. MiRNAs modulate protein expression through regulating the degradation and translation of mRNAs at post-transcriptional level (Chang et al., 2016; Jamieson et al., 2012). They play a critical role in various biological processes which are involved in the development and progression of tumors, including proliferation, apoptosis, differentiation and metastasis (Hayes et al., 2014; Ebert & Sharp, 2012; Rogers & Chen, 2013; Liu et al., 2012).

Additionally, they are very stable and easily detected in the blood and tissues (Gilad et al., 2008). Therefore, plenty of miRNAs are employed as new biomarkers for the diagnosis and prognosis of tumors. Regarding to OS, a variety of miRNAs has been reported to be associated with its prognosis. Kim et al. demonstrated that the pooled HR was 1.40 (95%CI: 1.01-1.94) for OS patients with lower expression miRNAs, and proposed that miRNAs with increased expression should also be investigated for their effects on the prognosis of OS patients. Additionally, the expression of some miRNAs is associated with OS chemoresistance (Xie et al., 2018). In our study, the 9 miRNAs, having been studied widely, were chosen as research targets. Our results demonstrated that miRNA-21, miRNA-214, miRNA-34a, miRNA-133a and miRNA-539 were

independently associated with the prognosis of OS patients.

MiRNA-21 has been confirmed to act as tumor oncogene in many types of tumors. For OS, it may regulate the proliferation, invasion and metastasis of OS cells through directly targeting PTEN and RECK (Ziyan et al., 2011; Lv et al., 2016). Li et al. demonstrated that the elevated expression of miRNA-21 might lead to elevated expression of the proteins in the PI3K/AKT signaling pathway and decreased expression of PTEN, which was associated with the increased invasiveness of OS cells (Li et al., 2018). Hu et al. indicated that inhibition of miRNA-21 might reduce the proliferation of OS cells through modulating the TGF- β 1 signaling pathway and targeting PTEN (Hu et al., 2018). Additionally, miRNA-21 might decrease the anti-tumor effect of cisplatin through modulating the expression of Bcl-2 (Ziyan & Yang, 2016). Our results demonstrated that high expression of miRNA-21 was independently associated with poor prognosis of OS patients with a *HR* of 3.457 (95%*CI*: 2.165-11.518). MiRNA-214 may act as either a tumor suppressor gene or an oncogene. For OS, the elevated expression of miRNA-214 is associated with enhanced invasion and proliferation of OS cells through modulating the expression of LZTS1 (Xu & Wang, 2014). However, Rehei et al. found that the expression of miRNA-214 was negatively associated with the expression of TRAF3 in OS tissues, and over-expression of miRNA-214 could inhibit the invasion and metastasis of OS cells through targeting TRAF3 (Rehei et al., 2018). Our results demonstrated that high expression of miRNA-214 was independently associated with poor prognosis of OS patients with a *HR* of 3.138 (95%*CI*: 2.014-10.259).

MiRNA-34a has various target genes which play important roles in biological function of OS

cells, such as Fag1, Wnt, p53 and Notch (Wu et al., 2013; Yan et al., 2012). Gang et al. demonstrated that miRNA-34a was correlated with the apoptosis, proliferation and adhesion of OS cells, and could function as a new tumor suppressor gene by reducing the expression of DUSP1 (Gang et al., 2017). Zhang et al. proved that miRNA-34a was a crucial regulator in the dedifferentiation of OS cells through modulating PAI-1-Sox2 axis (Zhang et al., 2018). In addition, Wang et al. showed that down-modulated expression of miRNA-34a was a prognostic biomarker for poor prognosis of OS patients through a meta-analysis (Wang et al., 2018). Our results demonstrated that low expression of miRNA-34a was independently associated with poor prognosis of OS patients with a *HR* of 0.452 (95%*CI*: 0.202-0.915). MiRNA-133a has been proved to be a crucial modulator for osteogenesis, and have a key role in osteoblast differentiation (Bao et al., 2010). It can act as an antionco-miRNA or a tumor suppressor gene in the development and progression of tumors (Ji et al., 2013). It has been reported to be associated with many cancers, including esophagus cancer, bladder cancer and prostate cancer. The underlying mechanisms of pro-apoptotic function of miRNA-133a may be associated with the inhibition of Mcl-1 and Bcl-xL expression (Wang et al., 2010). Our results confirmed that low expression of miRNA-133a was independently associated with poor prognosis of OS patients with a *HR* of 0.307 (95%*CI*: 0.113-0.874). Few reports have investigated the biological functions of miRNA-539. Muthusamy et al. found that miRNA-539 could inhibit O-GlcNAcase expression (Muthusamy et al., 2014). Wang et al. demonstrated that miRNA-539 was involved in the regulation of apoptosis and mitochondrial activity by means of targeting PHB2 (Wang et al., 2014). The expression of miRNA-539 is down-regulated in thyroid cancer, and moreover, it has

a suppressor role in the invasion and metastasis of thyroid cancer cells through targeting CARMA1 (Gu & Sun, 2015). Our results demonstrated that low expression of miRNA-539 was independently associated with poor prognosis of OS patients with a *HR* of 0.358 (95%*CI*: 0.155-0.896).

Conclusions

High expression of miRNA-21 and miRNA-214 and low expression of miRNA-34a, miRNA-133a and miRNA-539 were associated with poor prognosis of OS patients after adjusting for TNM stage, metastasis or recurrence and miRNA-30c.

Acknowledgements

None.

References

- Ambros V. 2004. The functions of animal microRNAs. *Nature* 431(7006):350-355.
- Bartel DP. 2004. MicroRNAs: Genomics, biogenesis, mechanism, and function. *Cell* 116(2):281-297.
- Bao B, Rodriguez-Melendez R, Wijeratne SS, Zemleni J. 2010. Biotin regulates the expression of holocarboxylase synthetase in the miR-539 pathway in HEK-293 cells. *J Nutr* 140(9):1546-1551. DOI: 10.3945/jn.110.126359.
- Biermann JS, Adkins DR, Agulnik M, Benjamin RS, Brigman B, Butrynski JE, Cheong D,

253 Chow W, Curry WT, Frassica DA, Frassica FJ, Hande KR, Hornicek FJ, Jones RL, Mayerson J,
254 McGarry SV, McGrath B, Morris CD, O'Donnell RJ, Randall RL, Santana VM, Satcher RL,
255 Siegel HJ, von Mehren M, Bergman MA, Sundar H; National comprehensive cancer network.
256 2013. Bone cancer. *J Natl Compr Canc Netw* 11(6):688-723.

257 Bracken CP, Scott HS, Goodall GJ. 2016. A network-biology perspective of microRNA function
258 and dysfunction in cancer. *Nat Rev Genet* 17(12):719-732. DOI: 10.1038/nrg.2016.134.

259 Bahrami A, Aledavood A, Anvari K, Hassanian SM, Maftouh M, Yaghobzade A, Salarzaee O,
260 ShahidSales S, Avan A. 2018. The prognostic and therapeutic application of microRNAs in
261 breast cancer: Tissue and circulating microRNAs. *J Cell Physiol* 233(2):774-786. DOI:
262 10.1002/jcp.25813.

263 Chang J, Yao M, Li Y, Zhao D, Hu S, Cui X, Liu G, Shi Q, Wang Y, Yang Y. 2016. MicroRNAs
264 for osteosarcoma in the mouse: a meta-analysis. *Oncotarget* 7(51):85650-85674. DOI:
265 10.18632/oncotarget.13333.

266 Cheng D, Qiu X, Zhuang M, Zhu C, Zou H, Liu Z. 2017. MicroRNAs with prognostic
267 significance in osteosarcoma: a systemic review and meta-analysis. *Oncotarget* 8(46):81062-
268 81074. DOI: 10.18632/oncotarget.19009.

269 Ebert MS, Sharp PA. 2012. Roles for microRNAs in conferring robustness to biological
270 processes. *Cell* 149(3):515-524. DOI: 10.1016/j.cell.2012.04.005.

271 Griffiths-Jones S, Saini HK, van Dongen S, Enright AJ. 2008. miRBase: tools for microRNA
272 genomics. *Nucleic Acids Res* 36(Database issue):D154-158.

273 Gilad S, Meiri E, Yogeve Y, Benjamin S, Lebanony D, Yerushalmi N, Benjamin H, Kushnir M,
274 Cholakh H, Melamed N, Bentwich Z, Hod M, Goren Y, Chajut A. 2008. Serum microRNAs are
275 promising novel biomarkers. *PLoS One* 3(9):e3148. DOI: 10.1371/journal.pone.0003148.

276 Gu L, Sun W. 2015. MiR-539 inhibits thyroid cancer cell migration and invasion by directly
277 targeting CARMA1. *Biochem Biophys Res Commun* 464(4):1128-1133. DOI:
278 10.1016/j.bbrc.2015.07.090.

279 Gang L, Qun L, Liu WD, Li YS, Xu YZ, Yuan DT. 2017. MicroRNA-34a promotes cell cycle
280 arrest and apoptosis and suppresses cell adhesion by targeting DUSP1 in osteosarcoma. *Am J*
281 *Transl Res* 9(12):5388-5399.

282 Hayes J, Peruzzi PP, Lawler S. 2014. MicroRNAs in cancer: biomarkers, functions and therapy.
283 *Trends Mol Med* 20(8):460-469. DOI: 10.1016/j.molmed.2014.06.005.

284 Hutanu D, Popescu R, Stefanescu H, Pirtea L, Candea A, Sarau C, Boruga O, Mehdi L, Ciuca I,
285 Tanasescu S. 2017. The molecular genetic expression as a novel biomarker in the evaluation and
286 monitoring of patients with osteosarcoma-subtype bone cancer disease. *Biochem Genet*
287 55(4):291-299. DOI: 10.1007/s10528-017-9801-1.

288 Hu X, Li L, Lu Y, Yu X, Chen H, Yin Q, Zhang Y. 2018. miRNA-21 inhibition inhibits
289 osteosarcoma cell proliferation by targeting PTEN and regulating the TGF- β 1 signaling pathway.
290 *Oncol Lett* 16(4):4337-4342. DOI: 10.3892/ol.2018.9177.

291 Jamieson NB, Morran DC, Morton JP, Ali A, Dickson EJ, Carter CR, Sansom OJ, Evans TR,
292 McKay CJ, Oien KA. 2012. MicroRNA molecular profiles associated with diagnosis,
293 clinicopathologic criteria, and overall survival in patients with resectable pancreatic ductal

294 adenocarcinoma. *Clin Cancer Res* 18(2):534-545. DOI: 10.1158/1078-0432.CCR-11-0679.

295 Ji F, Zhang H, Wang Y, Li M, Xu W, Kang Y, Wang Z, Wang Z, Cheng P, Tong D, Li C, Tang
 296 H. 2013. MicroRNA-133a, downregulated in osteosarcoma, suppresses proliferation and
 297 promotes apoptosis by targeting Bcl-xL and Mcl-1. *Bone* 56(1):220-226. DOI:
 298 10.1016/j.bone.2013.05.020.

299 Kunz P, Fellenberg J, Moskovszky L, Sapi Z, Krenacs T, Machado I, Poeschl J, Lehner B,
 300 Szendroi M, Ruef P, Bohlmann M, Bosch AL, Ewerbeck V, Kinscherf R, Fritzsche B. 2015.
 301 Improved survival in osteosarcoma patients with atypical low vascularization. *Ann Surg Oncol*
 302 22(2):489-496. DOI: 10.1245/s10434-014-4001-2.

303 Liu Y, Yan W, Zhang W, Chen L, You G, Bao Z, Wang Y, Wang H, Kang C, Jiang T. 2012.
 304 MiR-218 reverses high invasiveness of glioblastoma cells by targeting the oncogenic
 305 transcription factor LEF1. *Oncol Rep* 28(3):1013-1021. DOI: 10.3892/or.2012.1902.

306 Li Z, Rana TM. 2014. Therapeutic targeting of microRNAs: current status and future challenges.
 307 *Nat Rev Drug Discov* 13(8):622-638. DOI: 10.1038/nrd4359.

308 Lv C, Hao Y, Tu G. 2016. MicroRNA-21 promotes proliferation, invasion and suppresses
 309 apoptosis in human osteosarcoma line MG63 through PTEN/Akt pathway. *Tumour Biol*
 310 37(7):9333-9342. DOI: 10.1007/s13277-016-4807-6.

311 Liu H, Li P, Chen L, Jian C, Li Z, Yu A. 2017. MicroRNAs as a novel class of diagnostic
 312 biomarkers for the detection of osteosarcoma: a meta-analysis. *Onco Targets Ther* 10:5229-5236.
 313 DOI: 10.2147/OTT.S143974.

314 Li C, Xu B, Miu X, Deng Z, Liao H, Hao L. 2018. Inhibition of miRNA-21 attenuates the

315 proliferation and metastasis of human osteosarcoma by upregulating PTEN. *Exp Ther Med*
 316 15(1):1036-1040. DOI: 10.3892/etm.2017.5477.

317 Mirabello L, Troisi RJ, Savage SA. 2009. Osteosarcoma incidence and survival rates from 1973
 318 to 2004: Data from the surveillance, epidemiology, and end results program. *Cancer*
 319 115(7):1531-1543. DOI: 10.1002/cncr.24121.

320 Mirabello L, Troisi RJ, Savage SA. 2009. International osteosarcoma incidence patterns in
 321 children and adolescents, middle ages and elderly persons. *Int J Cancer* 125(1):229-234. DOI:
 322 10.1002/ijc.24320.

323 Muthusamy S, DeMartino AM, Watson LJ, Brittian KR, Zafir A, Dassanayaka S, Hong KU,
 324 Jones SP. 2014. MicroRNA-539 is up-regulated in failing heart, and suppresses O-GlcNAcase
 325 expression. *J Biol Chem* 289(43):29665-29676. DOI: 10.1074/jbc.M114.578682.

326 Rytting M, Pearson P, Raymond AK, Ayala A, Murray J, Yasko AW, Johnson M, Jaffe N. 2000.
 327 Osteosarcoma in preadolescent patients. *Clin Orthop Relat Res* (373):39-50.

328 Rogers K, Chen X. 2013. Biogenesis, turnover, and mode of action of plant microRNAs.
 329 *Plant Cell* 25(7):2383-2399. DOI: 10.1105/tpc.113.113159.

330 Rehei AL, Zhang L, Fu YX, Mu WB, Yang DS, Liu Y, Zhou SJ, Younusi A. 2018. MicroRNA-
 331 214 functions as an oncogene in human osteosarcoma by targeting TRAF3. *Eur Rev Med*
 332 *Pharmacol Sci* 22(16):5156-5164. DOI: 10.26355/eurrev_201808_15711.

333 Wang ZX, Yang JS, Pan X, Wang JR, Li J, Yin YM, De W. 2010. Functional and biological
 334 analysis of Bcl-xL expression in human osteosarcoma. *Bone* 47(2):445-454. DOI:
 335 10.1016/j.bone.2010.05.027.

336 Wu X, Zhong D, Gao Q, Zhai W, Ding Z, Wu J. 2013. MicroRNA-34a inhibits human
337 osteosarcoma proliferation by downregulating ether à go-go 1 expression. *Int J Med Sci*
338 10(6):676-682. DOI: 10.7150/ijms.5528.

339 Wang K, Long B, Zhou LY, Liu F, Zhou QY, Liu CY, Fan YY, Li PF. 2014. CARL lncRNA
340 inhibits anoxia-induced mitochondrial fission and apoptosis in cardiomyocytes by impairing
341 miR-539-dependent PHB2 downregulation. *Nat Commun* 5:3596. DOI: 10.1038/ncomms4596.

342 Wang W, Hu S, Chang J, Ruan H, Zhi W, Wang X, Shi Q, Wang Y, Yang Y. 2018. Down-
343 Regulated microRNA-34a Expression as a Prognostic Marker for Poor Osteosarcoma in Mice: A
344 Systematic Review and Meta-Analysis. *J Cancer* 9(22):4179-4186. DOI: 10.7150/jca.27483.

345 Xu Z, Wang T. 2014. miR-214 promotes the proliferation and invasion of osteosarcoma cells
346 through direct suppression of LZTS1. *Biochem Biophys Res Commun* 449(2):190-195. DOI:
347 10.1016/j.bbrc.2014.04.140.

348 Xie B, Li Y, Zhao R, Xu Y, Wu Y, Wang J, Xia D, Han W, Chen D. 2018.
349 Identification of Key Genes and miRNAs in Osteosarcoma Patients with Chemoresistance by Bi
350 oinformatics Analysis. *Biomed Res Int* 2018:4761064. DOI: 10.1155/2018/4761064.

351 Yan K, Gao J, Yang T, Ma Q, Qiu X, Fan Q, Ma B. 2012. MicroRNA-34a inhibits the
352 proliferation and metastasis of osteosarcoma cells both in vitro and in vivo. *PLoS One*
353 7(3):e33778. DOI: 10.1371/journal.pone.0033778.

354 Yu W, Zhu J, Wang Y, Wang J, Fang W, Xia K, Shao J, Wu M, Liu B, Liang C, Ye C, Tao H.
355 2017. A review and outlook in the treatment of osteosarcoma and other deep tumors with

356 photodynamic therapy: from basic to deep. *Oncotarget* 8(24):39833-39848. DOI:
 357 10.18632/oncotarget.16243.

358 Ziyan W, Shuhua Y, Xiufang W, Xiaoyun L. 2011. MicroRNA-21 is involved in osteosarcoma
 359 cell invasion and migration. *Med Oncol* 28(4):1469-1474. DOI: 10.1007/s12032-010-9563-7.

360 Zhang J, Yan YG, Wang C, Zhang SJ, Yu XH, Wang WJ. 2015. MicroRNAs in osteosarcoma.
 361 *Clin Chim Acta* 444:9-17. DOI: 10.1016/j.cca.2015.01.025.

362 Zhu K, Liu L, Zhang J, Wang Y, Liang H, Fan G, Jiang Z, Zhang CY, Chen X, Zhou G. 2016.
 363 MiR-29b suppresses the proliferation and migration of osteosarcoma cells by targeting CDK6.
 364 *Protein Cell* 7(6):434-444. DOI: 10.1007/s13238-016-0277-2.

365 Zhou H, Zhang M, Yuan H, Zheng W, Meng C, Zhao D. 2016. MicroRNA-154 functions as a
 366 tumor suppressor in osteosarcoma by targeting Wnt5a. *Oncol Rep* 35(3):1851-1858. DOI:
 367 10.3892/or.2015.4495.

368 Ziyan W, Yang L. 2016. MicroRNA-21 regulates the sensitivity to cisplatin in a human
 369 osteosarcoma cell line. *Ir J Med Sci* 185(1):85-91. DOI: 10.1007/s11845-014-1225-x.

370 Zhang Y, Pan Y, Xie C, Zhang Y. 2018. miR-34a exerts as a key regulator in the
 371 dedifferentiation of osteosarcoma via PAI-1-Sox2 axis. *Cell Death Dis* 9(7):777. DOI:
 372 10.1038/s41419-018-0778-4.

Table 1 (on next page)

Clinical characteristics of OS patients

1 **Table 1** Clinical characteristics of OS patients

Clinical characteristics	No. of patients	Percentages (%)
Gender		
Male	36	57.14%
Female	27	42.86%
Age (years)		
≤25	55	87.30%
>25	8	12.70%
Tumor diameter (cm)		
≤5	37	58.73%
>5	26	41.27%
TNM stage		
I + II	25	39.68%
III+IV	38	60.32%
Metastasis or recurrence		
Yes	37	58.73%
No	26	41.27%
Differentiation		
Well and moderate	31	49.21%
Poor	32	50.79%

2

Table 2(on next page)

Expression levels of microRNAs in tumor tissues and adjacent normal bone tissues

1 **Table 2** Expression levels of microRNAs in tumor tissues and adjacent normal bone tissues

	microR								
	microR	microR	microR	microR	microR	microR	NA-	microR	microR
	NA-21	NA-214	NA-433	NA-30c	NA-34a	NA-101	133a	NA-539	NA-218
Tumor	7.35±2.	6.12±2.	2.26±1.	3.93±1.	3.09±0.	3.16±1.	3.78±2.	2.35±1.	2.16±1.
tissues	96	25	34	77	94	72	17	08	07
Adjace									
nt									
normal									
bone	3.14±1.	3.37±1.	1.17±0.	5.34±1.	5.24±1.	5.19±2.	11.89±	5.23±1.	2.31±1.
tissues	58	49	91	32	35	74	4.16	84	18
<i>t</i>	9.959	8.088	5.341	-5.069	-10.374	-4.981	-13.719	-10.714	-0.747
<i>P</i>	<0.05	<0.05	<0.05	<0.05	<0.05	<0.05	<0.05	<0.05	>0.05

2

3

Table 3(on next page)

Median time of survival and log rank χ^2 for the K-M survival plots

1 **Table 3** Median time of survival and log rank χ^2 for the K-M survival plots

		No. of patients	Median time of survival (months)	Log rank χ^2	<i>P</i>
TNM stage	I + II	25	39.67±4.43	4.199	0.040
	III+IV	38	49.15±5.14		
Metastasis or recurrence	Yes	37	32.72±3.85	28.970	<0.001
	No	26	63.42±6.29		
microRNA-21	Low expression	24	61.75±5.60	11.847	0.001
	High expression	39	35.32±4.25		
microRNA-214	Low expression	26	58.24±6.17	7.338	0.007
	High expression	37	36.36±4.28		
microRNA-34a	Low expression	33	35.58±4.22	5.372	0.020

microRNA-133a	High expression	30	56.18±5.87	16.258	<0.001
	Low expression	42	36.35±4.38		
microRNA-539	High expression	21	63.47±5.89	7.390	0.007
	Low expression	34	35.27±4.13		
microRNA-30c	High expression	29	57.26±6.07	3.378	0.066
	Low expression	32	42.41±4.72		
	High expression	31	48.47±5.06		

2

3

Table 4(on next page)

Results of Cox proportional hadards model

1 **Table 4** Results of Cox proportional hadards model

	Regression coefficient	Standard error	Wald χ^2	<i>Hazard</i> <i>ratio</i>	95% <i>confidence</i> <i>interval</i>	<i>P</i>
microRNA-21	1.107	0.465	5.923	3.457	2.165-11.518	0.013
microRNA-214	1.058	0.446	5.642	3.138	2.014-10.259	0.017
microRNA-34a	-0.835	0.371	5.148	0.452	0.202-0.915	0.021
microRNA-133a	-0.946	0.382	6.137	0.307	0.113-0.874	0.011
microRNA-539	-0.887	0.369	5.474	0.358	0.155-0.896	0.018
TNM stage	0.953	0.392	5.016	2.893	1.496-8.125	0.024
metastasis or recurrence	1.154	0.458	6.529	3.628	2.217-12.316	0.007

microRNA-30c	-0.738	0.426	3.045	0.689	0.445-1.828	0.074
--------------	--------	-------	-------	-------	-------------	-------

Figure 1

Expression levels of microRNAs in tumor tissues and adjacent normal bone tissues.

*: $P < 0.05$, vs Tumor tissues.

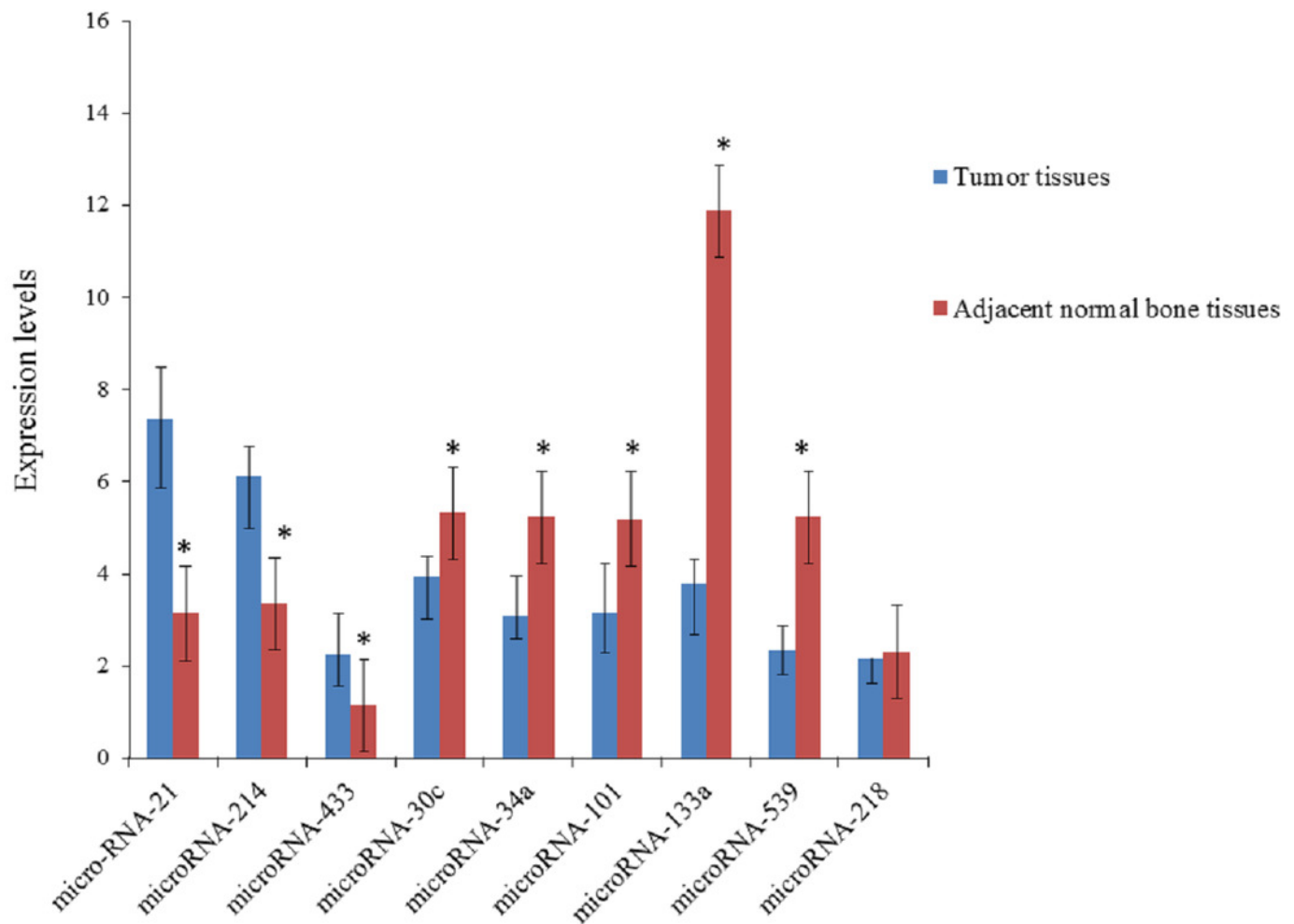


Figure 2

Kaplan-Meier analysis of cumulative survival for TNM stage using Log Rank test.

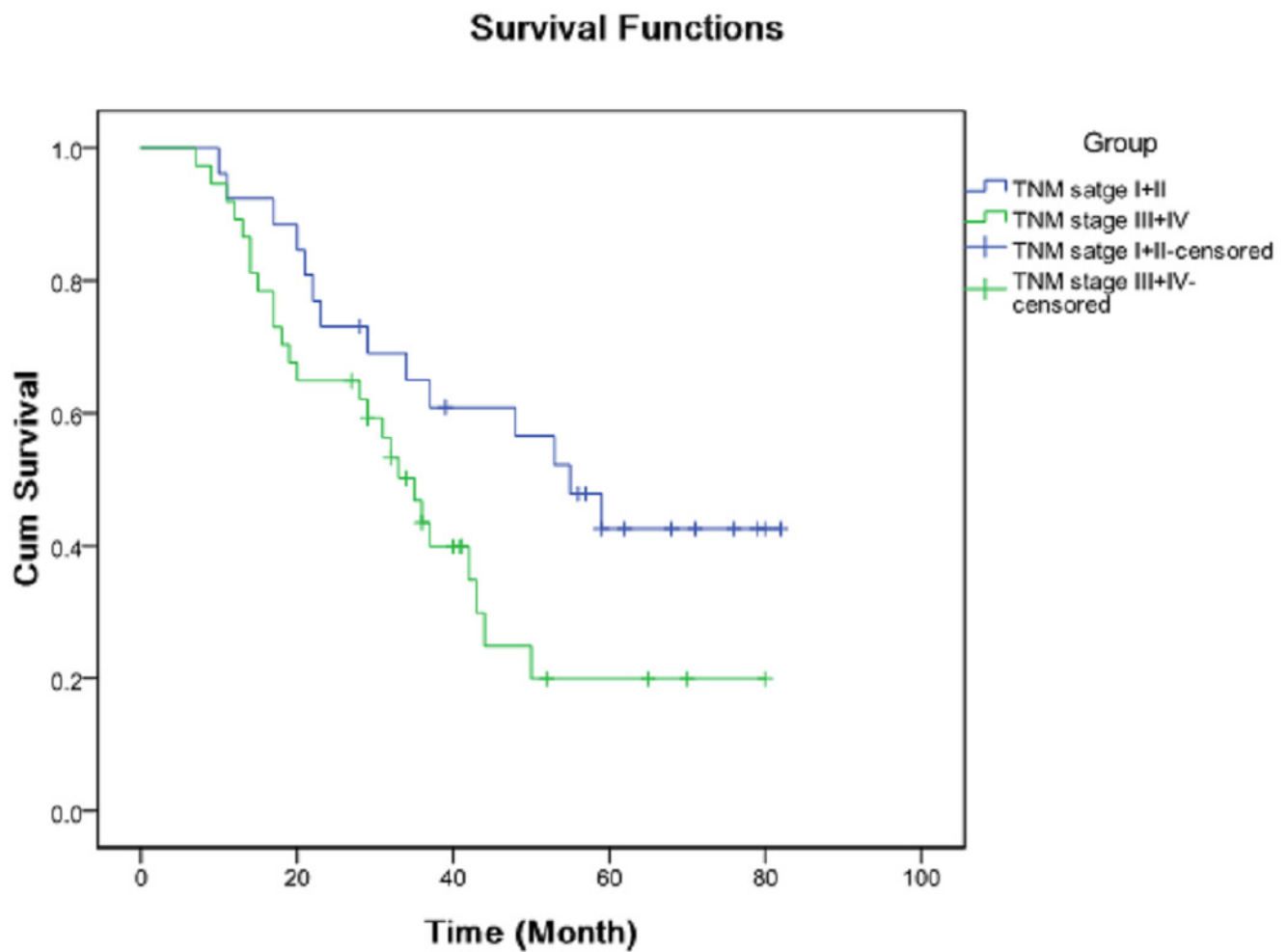


Figure 3

Kaplan-Meier analysis of cumulative survival for metastasis or recurrence using Log Rank test.

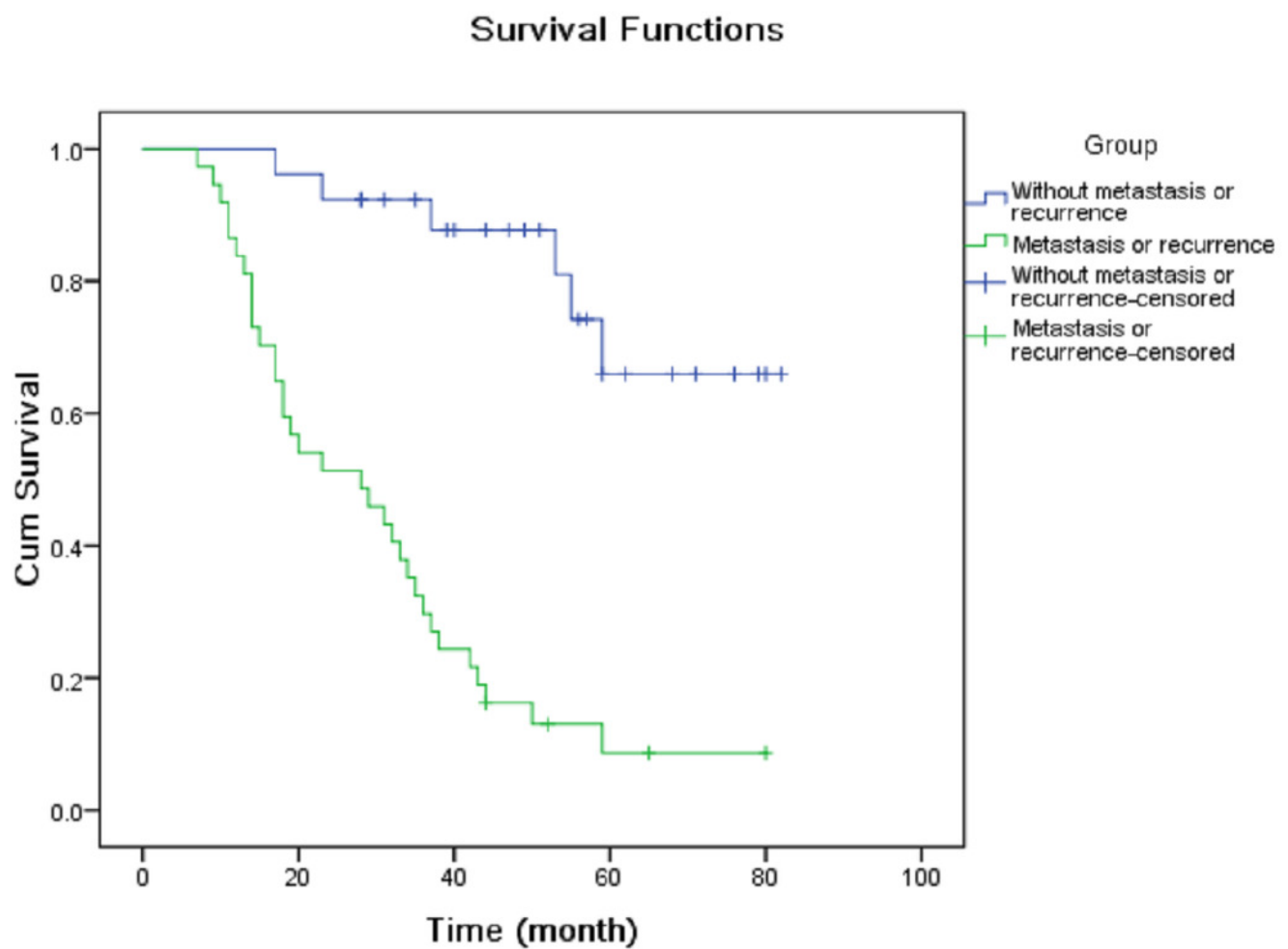


Figure 4

Kaplan-Meier analysis of cumulative survival for microRNAs using Log Rank test.

A: microRNA-21, B: microRNA-214, C: microRNA-34a, D: microRNA-133a, E: microRNA-539, and F: microRNA-30c.

