

Genetic links between endometriosis and cancers in women

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Abstract

Endometriosis is a chronic disease occurring during the reproductive stage of women. Although there is only limited association between endometriosis and gynecological cancers with regard to clinical features, the molecular basis of the relationship between these diseases is unexplored. We conducted a systematic study by integrating literature-based evidence, gene expression and large-scale cancer genomics data in order to reveal any genetic relationships between endometriosis and cancers in women. We curated 984 endometriosis-related genes from 3270 PubMed articles and then conducted a meta-analysis of the two public gene expression profiles related to endometriosis which identified Differential Expression of Genes (DEGs). Following an overlapping analysis, we identified 39 key endometriosis-related genes common in both literature and DEG analysis. Finally, the functional analysis confirmed that all the 39 genes were associated with the vital processes of tumour formation and cancer progression and that two genes (*PGR* and *ESR1*) were common to four cancers of women. From network analysis, we identified a novel linker gene, *C3AR1*, which had not been implicated previously in endometriosis. The shared genetic mechanisms of endometriosis and cancers in women identified in this study provided possible new avenues of multiple disease management and treatments through early diagnosis.

Introduction

Endometriosis is a chronic disease of women occurring at the reproductive stage of their lives. Worldwide, approximately 176 million (6-10%) women are affected by this disease (Kvaskoff et al., 2017) diagnosed by the formation of endometrial-like tissues and lesions, not only on the walls of the uterus but also in the fallopian tubes and on the pelvic wall. It is a common disease that can cause pelvic inflammation, chronic pain and infertility (Giudice and Kao, 2004). The first report on endometriosis-associated ovarian cancer was published in 1927 (Sampson, 1927) and the histological transformation of endometriosis to endometrioid ovarian cancers (EnOC) was reported in 1996 (de la Cuesta et al., 1996). This process has been confirmed by subsequent epidemiological studies (Heidemann et al., 2014, Kim et al., 2014, Kvaskoff et al., 2015). The occurrence of synchronous endometriosis in ovarian cancer lesions was reported by Jimbo et al. (1997) with 23.1 and 43% of EnOC. A separate epidemiological study showed a 2-3% higher risk in ovarian cancer patients with endometriosis than controls (Wei et al., 2011). A significant positive association was observed with different histological subtypes of ovarian cancers, particularly with endometrioid, epithelial and clear cell types (Saavalainen et al., 2018, Gandini et al., 2019, Pearce et al., 2012). Pearce et al. (2012) reported that endometriosis is associated with an increased risk of clear cell type ovarian cancer. The highest rate of incidence ratio of ovarian cancer was observed among women with ovarian endometriosis, particularly endometrioid and clear cell types (Saavalainen et al., 2018). In addition, endometriosis has been found to show more modest association with seromucinous borderline (Maeda and Shih, 2013, Samartzis et al., 2013) and low-grade serous ovarian carcinomas (Pearce et al., 2012). Sequencing and immune-histochemical studies have demonstrated a clonal relationship

between benign and malignant counterparts suggesting that the cancers may have arisen from the endometriotic lesions (Anglesio et al., 2015, Chene et al., 2015, Stamp et al., 2016, Wiegand et al., 2010).

Endometriosis, along with breast cancer, are estrogen-dependent chronic gynecological disorders and have similar risk factors associated with reproduction and the use of hormone replacement therapy (Munksgaard and Blaakaer, 2011). In addition, both endometriosis and cancer exhibit uncontrolled, estrogen-dependent proliferation, invasion, neo-angiogenesis and metastases (Burney and Giudice, 2012, Thomas and Campbell, 2000). Previous studies have identified the connection between endometriosis and breast cancer (BC) (Kokcu, 2011, Kvaskoff et al., 2015). In Denmark, data collected from 1977 to 2012 on 114,327 women showed that women who were diagnosed as endometriosis positive before age 40 had a 14% reduced risk of breast cancer than those diagnosed between age 40 and 50. Women diagnosed at age 50 or older were more than twice as likely to develop breast cancer as women of the same age who did not have endometriosis (Mogensen et al., 2016). Bulletti et al. (2010) reported that 20-25% of patients might be asymptomatic during the diagnosis of endometriosis. Although it is plausible that endometriosis is associated with increased risks for endometrial and breast cancer, epidemiological studies of the association are inconsistent.

Cervical cancer is the abnormal growth of cells in the lining of the cervix. The most common cervical cancer is squamous cell carcinoma (Marwah et al., 2012). On the other hand, cervical endometriosis is rare and often shows no symptoms. An epidemiological study of the association between the two diseases has reported the decreased risk of cervical cancer (Kvaskoff et al., 2015). A recent study suggested an association between cervical clear cell carcinoma (CCC) and cervical endometriosis (Hashiguchi et al., 2018). However, a decreased risk of cervical cancer of squamous cell histology among women with endometriosis has been suggested (Saavalainen et al., 2018).

Although endometriosis and endometrial cancer are two separate diseases, epidemiological, biological, and molecular data suggest that there could be links between the two disorders (Kvaskoff et al., 2015). Endometriosis is characterized by the formation of endometrial-resembling tissue external to the uterus while endometrial cancer (sometimes called uterine cancer) begins in the layer of cells that form the lining (endometrium) of the uterus. Endometriosis and endometrial cancer are both hormonally regulated diseases, having a common risk factor (higher levels of estrogen) and similar treatment measures (contraceptive pill and hormonal therapies) (Wetendorf and DeMayo, 2012). Both are involved in increased risk of uterine fibroids (Rowlands et al., 2011, Uimari et al., 2011). The predominant symptom of endometrial cancer is pelvic pain, which is also one of the main symptoms of endometriosis. In a recent genetic study, it was identified that these two diseases have common genetic causes (Painter et al., 2018). However, some previous studies showed the opposite or no significant association. Kvaskoff (2015) reported eight studies on the association between endometriosis and endometrial cancer and that there was no association in five studies where numbers of endometrial cancer cases ranging from 12 to 97 (Brinton et al., 1997, Brinton et al., 2005, Melin et al., 2007, Venn et al., 1999). Two studies

showed an association between endometriosis and increased endometrial cancer risk (Melin et al., 2006, Zucchetto et al., 2009) but one study suggested a significant inverse association (Borgfeldt and Andolf, 2004).

Therefore, a detailed investigation on the genetic control of the different cancer types along with endometriosis is necessary in order to explore the relationships between these two diseases. Over the last few decades, a large number of genetic data have been generated using next generation sequencing (NGS) technology and these can be used to decipher the genetic relationships between the two types of diseases. Therefore, we have conducted a meta-analysis of NGS data to explore evidence concerning the genomic and functional relationships of endometriosis and cancers in women.

Materials and Methods

Endometriosis-related gene curation from the literature

To explore the genetics of endometriosis, we conducted a literature search and extracted 3270 records from the GeneRIF (Gene Reference into Function), a database providing lists of genes and their functions (Maglott et al., 2010). We curated the literature data manually to extract the corresponding gene names. In total, we curated 984 genes related to studies of endometriosis and used these as a basis to explore similarities in the genetic mechanisms of cancers.

Differential gene expression for endometriosis dataset

To obtain independent evidence at the expression level, we focused on those studies which compared endometriosis patients with healthy individuals. We downloaded two datasets (GSE2339 and GSE5108) related to endometriosis that were collected from the gene expression omnibus (GEO) database, a repository of high-throughput gene expression data of original submitter-supplied records and curated records. Using GEO2R, we compared endometriosis samples with that of healthy individuals and, using the data filtering criteria of adjusted P values < 0.05 and an absolute fold change > 2 , identified genes that are differentially expressed (Barrett et al., 2012). Overlapping analysis was conducted to identify genes common to the two GEO datasets and the curated endometriosis gene lists.

Mutational Analysis

The cancer related mutational analyses were conducted using the cBio portal (Cerami et al., 2012) which enables the exploration, visualization and analysis of multidimensional cancer genomics data. We selected the TCGA datasets (Tomczak et al., 2015) of cancers in women including ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma, breast invasive carcinoma, and endometrial carcinoma in order to analyse the mutational frequency of the genes that are related to endometriosis. The combined studies dataset contained a total of 7462 samples.

Functional Enrichment and Network analysis

To investigate the biological systems in the endometriosis-related genes, we analysed the genes common to endometriosis and the differential expression of endometriosis by using the online tools Toppfun (Chen et al., 2009), REVIGO (Supek et al., 2011) and GeneMania (Warde-Farley et al., 2010). The molecular functions of the key endometriosis-related genes were analysed using ToppFun. ToppFun is a web tool which allows users to explore the molecular functions of gene ontology (GO) including cellular components, biological processes and molecular functions. We extracted gene IDs and their corresponding P values for the visualization process using REVIGO. REVIGO summarized and removed the redundant GO terms from a long list. The GO results served as input data in REVIGO and provided a semantic similarity-based scatterplot of GO terms from Toppfun. To perform the network analysis we used GeneMania to identify the interactions of the selected genes and Cytoscape (Shannon et al., 2003) to characterize and visualise the network results.

Results

Identification of potential endometriosis-related genes

In order to explore the association between endometriosis and four cancers of women, we identified the genes commonly implicated in those diseases (Figure 1). To achieve this, we manually curated endometriosis-related genes from 3273 PubMed abstracts (Table S1). The curated results were mapped based on their official gene names in concordance with data integration for other cancer genomics data sources. In total, we curated 984 endometriosis genes. To further refine the key genetic factors related to the endometriosis, we performed differential expression analyses on two endometriosis-related expression datasets: GSE2339 and GSE5108. GSE2339 is a two disease (endometrioma and eutopic endometrium) state set presenting the molecular mechanisms underlying the pathology of endometriosis (Hawkins et al., 2011). GSE5108 presents the gene expression profile of endometriosis (Eyster et al., 2007).

Using a cut-off as the adjusted P value <0.05 and the absolute fold change >2 , we identified 1037 genes associated with endometriosis (Table S2) from GSE2339 and 800 from the GSE5108 datasets. The overlapping results revealed that 165 (39+65+61) genes (Figure 1A) shared at least one GEO dataset and curated gene list and, of these, 65 and 61 genes are common to GSE2339 and GSE5108 , and 39 are in both GEO datasets. Using these lists, we explored the key molecular processes for the endometriosis. In summary, our computational workflow integrated the literature curated endometriosis genes with two other GEO endometriosis datasets.

Functional analysis of major endometriosis-related genes common in GEO datasets and curated list

Functional enrichment analysis of the 165 genes shared by our curated endometriosis-related gene list and at least one GEO dataset was conducted using Gene Ontology (GO)

terms as functional units (Figure 1C). The results revealed the genes enriched with cell proliferation (GO: 0042127; p-value: 1.03E-41), growth (GO: 0040007; p-value: 2.17E-21), apoptotic process (GO: 0006915; p-value: 5.54E-21), and cell adhesion (GO: 0007155; p-value: 1.47E-17) (Table S3). All these gene functions are associated with cancer progression. In addition, we identified hormone stimulating genes involved in the occurrence of endometriosis. Eighteen genes were found to be involved in the responses to estrogen (P-value = 2.45E-12), five (*PTGER4*, *PTN*, *THBS1*, *CCL2*, *C3*) to progesterone (P-value = 7.45E-05) and 45 to cytokine (P-value = 9.567E-24). Thirty-one genes were associated with cytokine production (P-value = 7.03E-14) and 17 genes involved in the *ERK1* and *ERK2* cascades (P-value = 5.27E-10) (Table S3). In addition, biological adhesion (GO: 0022610; P-value = 2.04E-17) and *MAPK* cascade (GO: 0000165; P-value = 2.00E-17) play important roles in cell development processes related to cancer progression.

We conducted another functional enrichment analysis using ToppFun to explore the functions of the 39 genes common to both GEO data sets and the curated endometriosis gene list (Figure 1B). This showed that the genes are enriched in most of the vital processes in cancer and tumor progression including: apoptotic process (GO: 0006915; P-value = 3.47E-06), the regulation of the cell cycle (GO: 0051726; P-value = 4.48E-06), cell proliferation (GO: 0042127; P-value = 1.26E-11), cell death (GO: 0010941; P-value = 4.95E-07), and cell differentiation (GO: 0045595; P-value = 1.06E-07). Our study also found that five genes are also involved in the enrichment of estrogen (GO: 0043627; P-value = 1.22E-04) and four in the enrichment of progesterone (GO: 0032570; P-value = 3.53E-06). Ten genes were found to be associated with angiogenesis (GO: 0001525; P-value = 2.79E-08), twelve genes in responses to cytokine (GO: 0034097; P-value = 6.83E-08) and seven genes in cytokine production (GO: 0001816; P-value = 5.54E-04). Eleven genes were involved in the *MAPK* pathway (GO: 0000165; P-value = 2.05E-06) and five genes were involved in *ERK1* and *ERK2* cascade (GO: 00703; P-value = 3.00E-04) (Table S4).

As presented in supplementary Table S3 and S4, our study identified that *C3* is common in both estrogen, progesterone, angiogenesis, cytokine production, *MAPK* pathway, *ERK1*, and *ERK2*. Pleiotrophin (*PTN*), a member of a highly conserved human gene family (Rauvala, 1989), is common in the apoptotic process, cell cycle, cell proliferation, cell death, cell differentiation, progesterone, estrogen, and angiogenesis. *LEP* is common in the apoptotic process, cell cycle, cell proliferation, cell death, cell differentiation, estrogen, angiogenesis, cytokine, cytokine production, and *MAPK* pathway. Both *CCL2* and *THBS1* are involved in the apoptotic process, cell proliferation, cell death, progesterone, angiogenesis, cytokine production, and *MAPK* pathway. *AGTR1* is involved in the apoptotic process, cell proliferation, cell death, cell differentiation, estrogen, and cytokine production. *ESR1* is involved in cell cycle, cell proliferation, cell death, estrogen, and the *MAPK ERK1* and *ERK2* pathways. *PLCB1* and *CSF1R* are concerned in the cytokine and *MAPK* pathways. *GATA2*, *SOX17*, *DCN*, *PTGIS*, and *NRP2* are involved in angiogenesis; *AIF1*, *CXCL16*, *IRF6*, *MME*, *FOXA2*, *DCN*, *PTGIS*, *TNFSF13B* in cytokine; and *NGF*, *PLA2G2A*, *LPAR3*, and *NTF3* in the *MAPK* pathway only.

Interconnectivity of Endometriosis-related genes

In order to identify the interconnectivity of endometriosis-related genes, we conducted a network analysis of the 165 genes which shared at least one GEO dataset (Figure S1) and the curated gene list of endometriosis. We performed network analysis, using GeneMania and Cytoscape, and identified correlations among the genes of interest. To avoid the potential of a large number of non-significant correlations, we used only reliable interactions (Cerami et al., 2010). Additionally, we compared the results from the 165 genes with another network analysis focusing on 39 common genes of curated endometriosis genes and both GEO data sets.

The network analysis of the 165 endometriosis related genes showed that most of the genes has more than 20 connections. We found maximum connections (75) in *SPARC* followed by *FN1* (72), *VCAM1* (61), *DAPK1* (54), *CD14* (53), *HLA-DRA* (53), *TFAP2C* (51), *CCR1* (50), *CDH3* (47), *CXCL12* (46), *TIMP1* (46), *BGN* (45), *CSF1R* (43), *MMP9* (43), *CNN1* (41), *LIPC* (41), *CDH1* (40). From the derived network, we identified novel linker genes showing functional links with the endometriosis related genes. Among the identified linker genes, twenty genes (*CXCL9*, *TNFAIP6*, *FAP*, *A2M*, *TIMP3*, *CCL18*, *FCER1G*, *MMP2*, *CD68*, *C1QB*, *TNC*, *FPRI*, *CCL5*, *C3AR1*, *IGF1*, *ACTA2*, *TYROBP*, *MMP1*, *AOAH* and *COL3A1*) have more than >20 connections. Results suggested that these linker genes can be used in prognostic studies of endometriosis.

A network analysis of 39 genes is presented in (Figure 2A). By focusing on genes with the highest number of interactions, we found eight genes with 10 or more connections. Gene *TAGLN* had highest number of connections (23) followed by *CSF1R* (17), *ESR1* (16), *THBS1* (16), *IRF6* (14), *PLA2G2A* (12), *BDNF* (10), and *PLCB1* (10). We also revealed 20 novel linker genes (*VEGFA*, *NTF4*, *PAX2*, *PAX5*, *CD47*, *CFI*, *CSF1*, *C3AR1*, *IGFBP1*, *PGF*, *FOXF1*, *OVGP1*, *IRF8*, *IRF5*, *MDK*, *PARD6A*, *PARD3*, *TNFRSF13C*, *FEZ1*, and *IGFBP5*) connected with the 39 genes. Among the linker genes, 16 genes had 10 or more connections with highest in *CFI* (25) followed by *C3AR1* (24), *IGFBP1* (21), *VEGFA* (19), *IRF8* (18), *IGFBP5* (18), *CSF1* (15), *PARD3* (15), *FEZ1* (14), *FOXF1* (14), *OVGP1* (13), *PARD6A* (13), *CD47* (12), *PGF* (12), *IRF5* (10), and *PAX2* (10).

The network topological analysis identified 59 gene nodes and 364 gene-gene interactions, which showed that the majority of the gene nodes have multiple connections (Figure 2B). Of the 59 nodes, 39 were identified from our gene list and the remaining 20 were linker genes. The degrees of the nodes in the map fit a power law distribution $y=ax^b$, where $a=2.684$ and b is an exponent with an estimated value of -0.110. The correlation between the given data point and the corresponding point on the fitted curve is 0.195 ($R^2=0.014$). Topological analysis on the shortest path length distribution analysis shows that the average length of the shortest path is 2, which indicates that the number of nodes which instantly connected (Figure 2C).

Mutation frequency of major endometriosis-related genes common in three data sets

We conducted a mutational analysis of the 39 listed genes of endometriosis using TCGA datasets associated with four cancers of women. For mutational analysis, we used 7462 TCGA samples collected from 11 studies, consisting of 3834 breast, 1754 ovarian, 1577 endometrial and 297 cervical cancers. The frequency of alteration in cBio Cancer Genomics Portal is defined by mutation, copy number amplification, and homozygous deletion in tumor samples (Cerami et al., 2012). Alteration frequency of the TCGA samples revealed that all the 39 genes have a high alteration rate in the tumor samples as revealed by gene amplifications. For example, of the 585 cases of TCGA ovarian epithelial tumor, the frequency of genetic alterations was 60.17% of which the highest alteration was observed in amplification (37.44%, 219 cases) followed by deep deletion (10.6%, 62 cases) (Figure 3C). In 1169 cases of ovarian cancer, we observed 54.06% alteration and 41.75% of these were due to amplification. For endometrial carcinoma, the frequency was tested in 529 cases and 59.74% showed genetic alteration with maximum of these being in mutation (34.85%). For invasive breast carcinoma TCGA cohort (of 1086 cases), 40.88% of all were genetically altered and 21.73% of these were due to amplification. The cervical squamous cell carcinoma had 41.83% genetic alteration from 251 cases with highest number due to amplification (17.93%) (Figure 3E). These results showed the importance of endometriosis-related genes in the development of cancers in women as a consequence of their function in promoting a large number of copy number gains (Wee et al., 2018).

In addition to the sample analysis, we explored the genomic alterations in multiple genes across several tumor samples (Figure 3A-D). We used the OncoPrint in cBioportal for a query search of alterations in the 39 endometriosis-related genes in TCGA ovarian serous cystadenocarcinoma, TCGA breast invasive carcinoma, TCGA uterine corpus endometrial carcinoma, and TCGA cervical squamous cell carcinoma tumor samples. An oncoprint is a graphical display of gene mutations in human cancer tumor samples. We further identified the top 10 genes with the highest amplification rate in samples of four cancer types. (Figure 3A-D). For ovarian serous cystadenocarcinoma, there was a total of 12 (*MME*, *AGTR1*, *SOX17*, *PTN*, *NTF3*, *MSR1*, *PGR*, *PLCB1*, *BNC2*, *PTGIS*, *LEP*, and *PAX8*) genes with >4% of genetic alterations frequency. *IRF6* showed 6% and *SOX17*, *MSR1*, *PTGIS* showed 3% in the TCGA breast invasive carcinoma tumor samples. In TCGA uterine corpus endometrial carcinoma samples, there were nine (*SOX17*, *C3*, *PAX8*, *ESR1*, *MME*, *MSR1*, *BNC2*, *IRF6*, and *PGR*) genes with >5% alteration frequency. Three genes (*MME*, *PGR*, and *AGTR1*) showed >7% alteration frequency in the TCGA cervical squamous cell carcinoma. Most of these alterations were related to amplification. *MME*, *SOX17*, *AGTR1*, *PGR*, and *ESR1* had the highest amplifications frequency ranging from 4% to 11% in ovarian serous cystadenocarcinoma. We observed similar features in the uterine corpus endometrial carcinoma dataset: the genes with the highest frequency included *SOX17* and *C3* (8%), *PAX8*, *MME* and *ESR1* (7%). In breast invasive carcinoma, *IRF6* showed the highest rate of alteration (6%) followed by *SOX17* and *MSR1* (3%). In the case of cervical squamous cell carcinoma, alteration frequency was greatest in *MME* (10%), followed by *PGR* (9%) and

AGTR1 (8%). Therefore, all these endometriosis-related genes are associated with gene amplification events across the four cancers of women TCGA samples.

Mutational analysis of linker genes involved in the endometriosis network

We applied the network analysis and constructed a gene network to identify the global connections of the 39 genes identified by overlapping analysis between endometriosis-related genes and two GEO datasets. The derived network comprised 20 linker genes that were shown to connect with core genes. Interestingly, one of those linker genes (*C3AR1*) is within the 165 full gene list, which was identified by overlapping analysis between literature-based endometriosis-related genes and one set of GEO datasets.

The additional mutational analysis on the 20 linker genes, including *C3AR1* (Figure 4A), showed a significant amplification frequency across tumor samples. Eighty percent of the genes showed more than 1% genetic alterations in eleven TCGA datasets from four cancer types: breast invasive carcinoma, cervical squamous cell carcinoma, ovarian serous cystadenocarcinoma, and uterine corpus endometrial carcinoma. The TCGA ovarian serous cystadenocarcinoma patients had >40% genetic alteration/expression (median) (Figure 4B). Overall, most of cancer cohort patients showed >20% genetic alterations. Based on 7462 samples, the greatest rate of alteration was observed in *PARD3* and *C3AR1* (both 3%), followed by *CD47* (2.6%) and *IRF8* (2.4%). The graphical presentation of expression result from cBioportal (Figure 4B) revealed that the maximum expression of *C3AR1* was found in breast cancer samples followed by ovarian then cervical cancers and minimum in uterine or endometrial cancers.

Mutational and functional analysis of common genes involved in endometriosis and cancers in women

Overlapping analysis of 165 endometriosis genes and previously identified 52 genes of four women cancers (Bhyan et al., 2019) revealed that nine genes were common: *SPARC*, *CDH1*, *MET*, *TIMP1*, *BRCA1*, *IGF2*, *PGR*, *MMP9* and *ESR1* (Figure S2). *PGR* and *ESR1* are also found in the list of the common 39 genes from the three endometriosis-related data sets. Further analysis revealed that all the nine genes are frequently mutated and the frequency varied from 1% to 8% (Figure 5A). Gene *CDH1* had the highest rates of alteration (8%) followed by *BRCA1* (4%), *PGR* (4%) and *ESR1* (4%). *IGF2* had the lowest rate of alteration (1%). TCGA data sets collected from the eleven studies provided 7462 samples in which the maximum rate of genetic alteration occurred in endometrial carcinoma (36.86% of 529 cases), followed by ovarian epithelial tumor (27.69% of 585 cases), invasive breast carcinoma (26.7% of 1086 cases), and ovarian cancer (26.26% of 1169 cases) (Figure 5B).

Functional enrichment analysis revealed that the nine genes were associated with a number of key cancer pathways and reproductive system biological processes including: the progesterone receptor signalling pathway (GO: 0050847; P-value=2.90E-03); the hormone-mediated signalling pathway (GO: 0009755; P-value=1.46E-04); and the intracellular receptor signalling pathway (GO: 0030522; P-value=3.20E-04) (Table S3). Among the nine

genes, *PGR* and *ESR1* were involved in all three pathways and *PGR* alone was involved in the progesterone receptor signalling pathway. These results suggested that endometriosis and four women cancers arise from common genetic mutations.

Discussion

Endometriosis is the hormone-dependent abnormal growth of endometrial epithelial and stromal cells outside the uterine cavity which may cause chronic pelvic pain, subfertility and result in an increased risk of ovarian cancer (Vercellini et al., 2014, Saavalainen et al., 2018, Pearce et al., 2012). In a previous study, we identified some key driver genes of four cancers of women which function as hormonal stimulants (Bhyan et al., 2019). But, the high-grade serous ovarian cancers data set from TCGA are not strongly associated with endometriosis and the clear cell and endometrioid OCs that are associated with OC are not represented in TCGA. However, Dawson et al. (2018) suggested that endometriosis and four cancers of women may share common biological mechanisms. Previous studies demonstrated that there was an increased risk of ovarian and breast cancer due to endometriosis (Saavalainen et al., 2018, Pearce et al., 2012, Saavalainen et al., 2019). Our research involved a meta-genomic study on endometriosis-related genes and an assessment of their involvement in four women cancers: breast, cervical, ovarian and endometrial. Integrated analyses, including functional enrichment, network, and mutational analysis provided a list of key genes playing dual role in endometriosis and women cancers.

In this study, functional analysis of endometriosis-related genes revealed shared mechanisms of endometriosis concerned with four cancers of women. This was particularly the case for cell proliferation, growth, apoptotic process, cell adhesion, regulation of cell cycle, regulation of cell death, and regulation of cell differentiation. The control of cell proliferation and apoptosis are the key regulatory mechanisms of cancer progression. The apoptotic process is highly regulated in human cells and essential for maintaining the physiological balance between cell death and cell growth (Koff et al., 2015). Different extracellular and intracellular signals, including hormones, growth factors and cytokines, can stimulate the different pathways (Dhillon et al., 2007) and regulate cell differentiation, growth and apoptosis (Kim and Choi, 2010). We identified common hormone stimulant pathways, including enrichment of estrogen and progesterone, angiogenesis, response to cytokine, and cytokine production, as functional roles of endometriosis-related genes. Estrogen and progesterone are the key hormones involved in reproductive development but are also associated with tumor growth and the spread of some cancers (Subramani et al., 2017). It has been shown that estrogen exposure is directly associated with an increase in the risk of developing breast cancer (Begg et al., 1987, Pike et al., 1979), whereas reducing exposure is thought to be protective against breast cancer (Hulka, 1997). Estrogen also increases the risk of ovarian cancer, particularly after menopause (Ho, 2003). Progesterone and progesterone receptors (PR) are important because of their role as critical regulators of breast and gynecological cancers. Although the uptake of both estrogen and progesterone reduces the risk of ovarian cancer, the mechanisms explaining the role of these two hormones in carcinogenesis is unclear (Ho, 2003). Angiogenesis, which is responsible for metastasis in

ovarian cancer (Gavalas et al., 2013), also has a key functional role in endometriosis. Several studies have indicated that the further outgrowth of ectopic endometrial implants through endometriosis leads to tumor formation (Shubik, 1982). Furthermore, the cancer-modifier cytokines were found to be involved in endometriosis (Brower, 2005) and the cytokinins are both tumor necrosis factors (Esquivel-Velázquez et al., 2015) as well as being associated with a number of gynaecological cancers (Heikkilä et al., 2008, Murooka et al., 2005).

Our study showed that kinase signalling pathways, such as *MAPK* (mitogen-activated protein kinases), *ERK1* and *ERK2*, are activated in endometriosis. *MAPK* act as integration points for many biochemical signals and are involved in a variety of cellular processes, including cell proliferation, differentiation, transcription regulation and development (Imajo et al., 2006). In association with several environmental stimuli such as, hormones and cytokines play a role in activating *MAPK* pathways (Imajo et al., 2006). The dysregulation of protein kinase stimulated by several oncogenic driver mutations was found to accelerate uncontrolled cellular proliferation in kinase-dependent tumour growth (Burotto et al., 2014, Sawyers, 2003). *ERK1* and *ERK2* are two extracellular regulated kinases and the final effectors of the *MAPK* pathway (Robinson and Cobb, 1997, Liu et al., 2018). In addition, they are both regulators of malignant breast cancer cells (Milde-Langosch et al., 2005). Therefore, these kinases were identified as potential targets for the treatment of cancers and endometriosis. Endometriosis-related genes identified in this study were found to be associated with key biological mechanisms controlling cancers in women. *C3* is a plasma protein which increases cell proliferation once synthesized in malignant ovarian epithelial cells (Cho et al., 2014). *PTN* is a member of a highly conserved human gene family (Rauvala, 1989) and is a key gene in the process of endometriosis and regulates multiple functions, including apoptosis, cell cycle, cell proliferation, cell differentiation, progesterone and estrogen production, and angiogenesis (Table S4). Overexpression of *PTN* in breast cancer cells, such as *MCF-7* gene enhanced angiogenesis in the rabbit corneal assay (Choudhuri et al., 1997). In addition, a truncated form of *PTN* was shown to act as a dominant-negative effector on the proliferation and angiogenesis of breast cancer cells, *in vitro* and *in vivo* (Ducès et al., 2008). Another endometriosis-related gene, *LEP*, functions as a key mediator in obesity-associated cancers including breast, colorectal and prostate (Renahan et al., 2010).

From the network analysis of 165 and 39 genes, we identified only one common linker gene, *C3AR1*, although there is no evidence of its involvement with endometriosis. *C3AR1* is an oncogenic gene that down regulates in tumour cells (Nabizadeh et al., 2016, Yamada et al., 2017). Formerly, *C3AR1* was considered to be involved in the innate immune response but is now regarded as a factor in cancer (Opstal-van Winden et al., 2012). Additionally, *C3AR1* was found to activate the *PI3K-AKT* pathways that result in cell proliferation (Cho et al., 2016, Towner et al., 2016). Further investigation of the involvement of this gene in endometriosis and four cancers of women is important for future disease management.

We identified that the alteration frequency was highest (5%) in *SOX17*, which is involved in oncogenesis through tumour suppression, down-regulating *MAML3* expression,

modulating nuclear β -catenin and antagonizing Wnt signaling (Zhang et al., 2016). Gene *IRF6*, a transcriptional activator which plays critical roles in endometrial gene expression and the growth and differentiation conceptus trophoctoderm (Fleming et al., 2009), also showed 5% alteration frequency in our study. High rate (5%) of mutation frequency was observed in macrophage metalloelastase (*MME*), which is a zinc-dependent endoprotease and also involved in *ECM* re-modulation and conversion of plasminogen to angiostatin (Lavilla-Alonso et al., 2012). Finally, we discovered eight genes (*MME*, *SOX17*, *AGTR1*, *PGR*, *ESR1*, *PAX8*, *C3* and *IRF6*) with high amplification frequency compared to other genes across the cancer types. These genes are all involved in endometriosis suggesting their involvement in the progression of cancers in women.

Conclusion

Using integrated bioinformatic analysis we discovered evidence of genetic link between endometriosis and women cancers. We utilized information from next generation sequence data and compiled a list of a large number of endometriosis related genes. Functional analysis confirmed that 39 genes were associated with the processes of tumour formation and cancer progression of which two (*PGR* and *ESR1*) were common to four cancers of women. Mutational analysis proved that eight endometriosis genes had a higher rate of alterations across the four women cancers. Finally, we explored a novel linker gene, *C3AR1*, which had not been implicated previously in endometriosis. The evidence of shared genetic mechanisms of endometriosis and cancers in women may be an avenue of future disease management and treatment through early diagnosis. This paper provides a catalogue of genetic links between endometriosis and cancer as a guide for further investigation and analysis.

Acknowledgements

The authors thank to two anonymous reviewers and the academic editor (Kate Lawrenson) of PeerJ for reviewing the manuscript and providing suggestions to improve the manuscript. We thank Professor Richard Burns (University of the Sunshine Coast) and Dr Mobashwer Alam (The University of Queensland) for their assistance in language editing and putting the manuscript in order.

References

- ANGLESIO, M. S., BASHASHATI, A., WANG, Y. K., SENZ, J., HA, G., YANG, W., ANIBA, M. R., PRENTICE, L. M., FARAHANI, H. & LI CHANG, H. 2015. Multifocal endometriotic lesions associated with cancer are clonal and carry a high mutation burden. *The Journal of pathology*, 236, 201-209.
- BARRETT, T., WILHITE, S. E., LEDOUX, P., EVANGELISTA, C., KIM, I. F., TOMASHEVSKY, M., MARSHALL, K. A., PHILLIPPY, K. H., SHERMAN, P. M. & HOLKO, M. 2012. NCBI GEO: archive for functional genomics data sets—update. *Nucleic acids research*, 41, D991-D995.
- BEGG, L., KULLER, L. H., GUTAI, J. P., CAGGIULA, A. G., WOLMARK, N., WATSON, C. G. & RAO, D. 1987. Endogenous sex hormone levels and breast cancer risk. *Genetic epidemiology*, 4, 233-247.

465 BHYAN, S. B., WEE, Y. K., LIU, Y., CUMMINS, S. & ZHAO, M. 2019. Integrative analysis of common
 466 genes and driver mutations implicated in hormone stimulation for four cancers in women.
 467 *PeerJ*, 7, e6872.

468 BORGFELDT, C. & ANDOLF, E. 2004. Cancer risk after hospital discharge diagnosis of benign ovarian
 469 cysts and endometriosis. *Acta obstetrica et gynecologica Scandinavica*, 83, 395-400.

470 BRINTON, L. A., GRIDLEY, G., PERSSON, I., BARON, J. & BERGQVIST, A. 1997. Cancer risk after a
 471 hospital discharge diagnosis of endometriosis. *American journal of obstetrics and*
 472 *gynecology*, 176, 572-579.

473 BRINTON, L. A., WESTHOFF, C. L., SCOCCIA, B., LAMB, E. J., ALTHUIS, M. D., MABIE, J. E. & MOGHISSI,
 474 K. S. 2005. Causes of infertility as predictors of subsequent cancer risk. *Epidemiology*, 16,
 475 500-507.

476 BROWER, V. 2005. Researchers Attempting To Define Role of Cytokines in Cancer Risk. *JNCI: Journal*
 477 *of the National Cancer Institute*, 97, 1175-1177.

478 BULLETTI, C., COCCIA, M. E., BATTISTONI, S. & BORINI, A. 2010. Endometriosis and infertility. *Journal*
 479 *of assisted reproduction and genetics*, 27, 441-447.

480 BURNEY, R. O. & GIUDICE, L. C. 2012. Pathogenesis and pathophysiology of endometriosis. *Fertility*
 481 *and sterility*, 98, 511-519.

482 BUROTTO, M., CHIOU, V. L., LEE, J. M. & KOHN, E. C. 2014. The MAPK pathway across different
 483 malignancies: a new perspective. *Cancer*, 120, 3446-3456.

484 CERAMI, E., GAO, J., DOGRUSOZ, U., GROSS, B. E., SUMER, S. O., AKSOY, B. A., JACOBSEN, A., BYRNE,
 485 C. J., HEUER, M. L. & LARSSON, E. 2012. The cBio cancer genomics portal: an open platform
 486 for exploring multidimensional cancer genomics data. *AACR*.

487 CERAMI, E. G., GROSS, B. E., DEMIR, E., RODCHENKOV, I., BABUR, Ö., ANWAR, N., SCHULTZ, N.,
 488 BADER, G. D. & SANDER, C. 2010. Pathway Commons, a web resource for biological pathway
 489 data. *Nucleic acids research*, 39, D685-D690.

490 CHEN, J., BARDES, E. E., ARONOW, B. J. & JEGGA, A. G. 2009. ToppGene Suite for gene list
 491 enrichment analysis and candidate gene prioritization. *Nucleic acids research*, 37, W305-
 492 W311.

493 CHENE, G., OUELLET, V., RAHIMI, K., BARRES, V., PROVENCHER, D. & MES-MASSON, A. M. 2015. The
 494 ARID1A pathway in ovarian clear cell and endometrioid carcinoma, contiguous
 495 endometriosis, and benign endometriosis. *International Journal of Gynecology & Obstetrics*,
 496 130, 27-30.

497 CHO, M. S., RUPAIMOOLE, R., CHOI, H.-J., NOH, K., CHEN, J., HU, Q., SOOD, A. K. & AFSHAR-
 498 KHARGHAN, V. 2016. Complement component 3 is regulated by TWIST1 and mediates
 499 epithelial-mesenchymal transition. *The Journal of Immunology*, 196, 1412-1418.

500 CHO, M. S., VASQUEZ, H. G., RUPAIMOOLE, R., PRADEEP, S., WU, S., ZAND, B., HAN, H.-D.,
 501 RODRIGUEZ-AGUAYO, C., BOTTSFORD-MILLER, J. & HUANG, J. 2014. Autocrine effects of
 502 tumor-derived complement. *Cell reports*, 6, 1085-1095.

503 CHOUDHURI, R., ZHANG, H.-T., DONNINI, S., ZICHE, M. & BICKNELL, R. 1997. An angiogenic role for
 504 the neurokines midkine and pleiotrophin in tumorigenesis. *Cancer research*, 57, 1814-1819.

505 DAWSON, A., FERNANDEZ, M. L., ANGLÉSIO, M., YONG, P. J. & CAREY, M. S. 2018. Endometriosis and
 506 endometriosis-associated cancers: new insights into the molecular mechanisms of ovarian
 507 cancer development. *Ecancermedicalscience*, 12.

508 DE LA CUESTA, R. S., EICHHORN, J. H., RICE, L. W., FULLER JR, A. F., NIKRUI, N. & GOFF, B. A. 1996.
 509 Histologic transformation of benign endometriosis to early epithelial ovarian cancer.
 510 *Gynecologic oncology*, 60, 238-244.

511 DHILLON, A. S., HAGAN, S., RATH, O. & KOLCH, W. 2007. MAP kinase signalling pathways in cancer.
 512 *Oncogene*, 26, 3279-3290.

513 DUCÈS, A., KARAKY, R., MARTEL-RENOIR, D., MIR, L., HAMMA-KOURBALI, Y., BIÉCHE, I., OPOLON, P.,
 514 DELBE, J., COURTY, J. & PERRICAUDET, M. 2008. 16-kDa fragment of pleiotrophin acts on

- endothelial and breast tumor cells and inhibits tumor development. *Molecular cancer therapeutics*, 7, 2817-2827.
- ESQUIVEL-VELÁZQUEZ, M., OSTOA-SALOMA, P., PALACIOS-ARREOLA, M. I., NAVA-CASTRO, K. E., CASTRO, J. I., MORALES-MONTOR, J. & RESEARCH, C. 2015. The role of cytokines in breast cancer development and progression. *Journal of Interferon*, 35, 1-16.
- EYSTER, K. M., KLINKOVA, O., KENNEDY, V. & HANSEN, K. A. 2007. Whole genome deoxyribonucleic acid microarray analysis of gene expression in ectopic versus eutopic endometrium. *Fertility and sterility*, 88, 1505-1533.
- FLEMING, J.-A. G., SONG, G., CHOI, Y., SPENCER, T. E. & BAZER, F. W. 2009. Interferon regulatory factor 6 (IRF6) is expressed in the ovine uterus and functions as a transcriptional activator. *Molecular and cellular endocrinology*, 299, 252-260.
- GANDINI, S., LAZZERONI, M., PECCATORI, F., BENDINELLI, B., SAIEVA, C., PALLI, D., MASALA, G. & CAINI, S. 2019. The risk of extra-ovarian malignancies among women with endometriosis: A systematic literature review and meta-analysis. *Critical reviews in oncology/hematology*, 134, 72-81.
- GAVALAS, N., LIONTOS, M., TRACHANA, S.-P., BAGRATUNI, T., ARAPINIS, C., LIACOS, C., DIMOPOULOS, M. & BAMIAS, A. 2013. Angiogenesis-related pathways in the pathogenesis of ovarian cancer. *International journal of molecular sciences*, 14, 15885-15909.
- GIUDICE, L. C. & KAO, L. C. 2004. Endometriosis. *The Lancet*, 364, 1789-1799.
- HASHIGUCHI, M., KAI, K., NISHIYAMA, S., NAKAO, Y., YOKOYAMA, M. & AISHIMA, S. 2018. Clear Cell Carcinoma of the Uterine Cervix Presented as a Submucosal Tumor Arising in a Background of Cervical Endometriosis. *International Journal of Gynecological Pathology*, 37, 88-92.
- HAWKINS, S. M., CREIGHTON, C. J., HAN, D. Y., ZARIFF, A., ANDERSON, M. L., GUNARATNE, P. H. & MATZUK, M. M. 2011. Functional microRNA involved in endometriosis. *Molecular endocrinology*, 25, 821-832.
- HEIDEMANN, L. N., HARTWELL, D., HEIDEMANN, C. H. & JOCHUMSEN, K. M. 2014. The relation between endometriosis and ovarian cancer—a review. *Acta obstetrica et gynecologica Scandinavica*, 93, 20-31.
- HEIKKILÄ, K., EBRAHIM, S. & LAWLOR, D. A. 2008. Systematic review of the association between circulating interleukin-6 (IL-6) and cancer. *European Journal of Cancer*, 44, 937-945.
- HO, S.-M. 2003. Estrogen, progesterone and epithelial ovarian cancer. *Reproductive Biology and Endocrinology*, 1, 73.
- HULKA, B. S. 1997. Epidemiologic analysis of breast and gynecologic cancers. *Progress in clinical biological research*, 396, 17-29.
- IMAJO, M., TSUCHIYA, Y. & NISHIDA, E. J. I. L. 2006. Regulatory mechanisms and functions of MAP kinase signaling pathways. *IUMB Life*, 58, 312-317.
- JIMBO, H., YOSHIKAWA, H., ONDA, T., YASUGI, T., SAKAMOTO, A. & TAKETANI, Y. 1997. Prevalence of ovarian endometriosis in epithelial ovarian cancer. *International Journal of Gynecology & Obstetrics*, 59, 245-250.
- KIM, E. K. & CHOI, E.-J. 2010. Pathological roles of MAPK signaling pathways in human diseases. *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease*, 1802, 396-405.
- KIM, H., KIM, T., CHUNG, H. & SONG, Y. 2014. Risk and prognosis of ovarian cancer in women with endometriosis: a meta-analysis. *British journal of cancer*, 110, 1878.
- KOFF, J. L., RAMACHANDIRAN, S. & BERNAL-MIZRACHI, L. 2015. A time to kill: targeting apoptosis in cancer. *International journal of molecular sciences*, 16, 2942-2955.
- KOKCU, A. 2011. Relationship between endometriosis and cancer from current perspective. *Archives of gynecology and obstetrics*, 284, 1473-1479.
- KONDI-PAFITI, A., SPANIDOU-CARVOUNI, H., PAPADIAS, K., HATZISTAMOU-KIARI, I., KONTOGIANNI, K., LIAPIS, A. & SMYRNIOTIS, V. 2004. Malignant neoplasms arising in endometriosis: clinicopathological study of 14 cases. *Clinical and experimental obstetrics & gynecology*, 31, 302-304.

- KVASKOFF, M., HORNE, A. W. & MISSMER, S. A. 2017. Informing women with endometriosis about ovarian cancer risk. *The Lancet*, 390, 2433-2434.
- KVASKOFF, M., MU, F., TERRY, K. L., HARRIS, H. R., POOLE, E. M., FARLAND, L. & MISSMER, S. A. 2015. Endometriosis: a high-risk population for major chronic diseases? *Human reproduction update*, 21, 500-516.
- LAVILLA-ALONSO, S., BAUER, M., ABO-RAMADAN, U., RISTIMÄKI, A., HALAVAARA, J., DESMOND, R., WANG, D., ESCUTENAIRE, S., AHTIAINEN, L. & SAKSELA, K. 2012. Macrophage metalloelastase (MME) as adjuvant for intra-tumoral injection of oncolytic adenovirus and its influence on metastases development. *Cancer gene therapy*, 19, 126-134.
- LIU, F., YANG, X., GENG, M. & HUANG, M. J. A. P. S. B. 2018. Targeting ERK, an Achilles' Heel of the MAPK pathway, in cancer therapy. 8, 552-562.
- MAEDA, D. & SHIH, I.-M. 2013. Pathogenesis and the role of ARID1A mutation in endometriosis-related ovarian neoplasms. *Advances in anatomic pathology*, 20, 45-52.
- MAGLOTT, D., OSTELL, J., PRUITT, K. D. & TATUSOVA, T. 2010. Entrez Gene: gene-centered information at NCBI. *Nucleic Acids Research*, 39, D52-D57.
- MARWAH, N., GARG, M., SINGH, S., SETHI, D., SEN, R. J. I. J. O. A. & RESEARCH, B. M. 2012. Unusual form of squamous cell carcinoma of the cervix extending in situ into the endometrium: Three case reports and review of literature. *International Journal of Applied and Basic Medical Research*, 2, 139-141.
- MASAND, R. P., EUSCHER, E. D., DEEVERS, M. T. & MALPICA, A. 2013. Endometrioid stromal sarcoma: a clinicopathologic study of 63 cases. *The American journal of surgical pathology*, 37, 1635-1647.
- MELIN, A., SPAREN, P. & BERGQVIST, A. 2007. The risk of cancer and the role of parity among women with endometriosis. *Human reproduction*, 22, 3021-3026.
- MELIN, A., SPAREN, P., PERSSON, I. & BERGQVIST, A. 2006. Endometriosis and the risk of cancer with special emphasis on ovarian cancer. *Human reproduction*, 21, 1237-1242.
- MILDE-LANGOSCH, K., BAMBERGER, A., RIECK, G., GRUND, D., HEMMINGER, G., MÜLLER, V. & LÖNING, T. J. B. J. O. C. 2005. Expression and prognostic relevance of activated extracellular-regulated kinases (ERK1/2) in breast cancer. *British journal of cancer*, 92, 2206-2215.
- MOGENSEN, J. B., KJÆR, S. K., MELLEMKJÆR, L. & JENSEN, A. 2016. Endometriosis and risks for ovarian, endometrial and breast cancers: a nationwide cohort study. *Gynecologic oncology*, 143, 87-92.
- MUNKSGAARD, P. S. & BLAAKAER, J. 2011. The association between endometriosis and gynecological cancers and breast cancer: a review of epidemiological data. *Gynecologic oncology*, 123, 157-163.
- MUROOKA, T. T., WARD, S. E. & FISH, E. N. 2005. Chemokines and cancer. In: L.C., P. (ed.) *Cytokines and Cancer*. Boston, MA: Springer.
- NABIZADEH, J. A., MANTHEY, H. D., STEYN, F. J., CHEN, W., WIDIAPRADJA, A., AKHIR, F. N. M., BOYLE, G. M., TAYLOR, S. M., WOODRUFF, T. M. & ROLFE, B. 2016. The complement C3a receptor contributes to melanoma tumorigenesis by inhibiting neutrophil and CD4+ T cell responses. *The journal of Immunology*, 196, 4783-4792.
- OPSTAL-VAN WINDEN, A. W., VERMEULEN, R. C., PEETERS, P. H., BEIJNEN, J. H., VAN GILS, C. H. J. B. C. R. & TREATMENT 2012. Early diagnostic protein biomarkers for breast cancer: how far have we come? *Breast Cancer Research and Treatment*, 134, 1-12.
- PAINTER, J. N., O'MARA, T. A., MORRIS, A. P., CHENG, T. H., GORMAN, M., MARTIN, L., HODSON, S., JONES, A., MARTIN, N. G. & GORDON, S. 2018. Genetic overlap between endometriosis and endometrial cancer: evidence from cross-disease genetic correlation and GWAS meta-analyses. *Cancer medicine*, 7, 1978-1987.
- PEARCE, C. L., TEMPLEMAN, C., ROSSING, M. A., LEE, A., NEAR, A. M., WEBB, P. M., NAGLE, C. M., DOHERTY, J. A., CUSHING-HAUGEN, K. L. & WICKLUND, K. G. 2012. Association between

endometriosis and risk of histological subtypes of ovarian cancer: a pooled analysis of case-control studies. *The lancet oncology*, 13, 385-394.

PIKE, M. C., GERKINS, V. R., CASAGRANDE, J. T., GRAY, G. E., BROWN, J. & HENDERSON, B. E. J. N. C. I. M. 1979. The hormonal basis of breast cancer. *ational Cancer Institute monograph*, 187-193.

RAUVALA, H. J. T. E. J. 1989. An 18-kd heparin-binding protein of developing brain that is distinct from fibroblast growth factors. *he EMBO Journal* 8, 2933-2941.

RENEHAN, A. G., SOERJOMATARAM, I. & LEITZMANN, M. F. J. E. J. O. C. 2010. Interpreting the epidemiological evidence linking obesity and cancer: a framework for population-attributable risk estimations in Europe. *European Journal of Cancer*, 46, 2581-2592.

ROBINSON, M. J. & COBB, M. H. 1997. Mitogen-activated protein kinase pathways. *Current opinion in cell biology*, 9, 180-186.

ROWLANDS, I. J., NAGLE, C. M., SPURDLE, A. B., WEBB, P. M., GROUP, A. N. E. C. S. & GROUP, A. O. C. S. 2011. Gynecological conditions and the risk of endometrial cancer. *Gynecologic oncology*, 123, 537-541.

SAAVALAINEN, L., LASSUS, H., BUT, A., TIITINEN, A., HÄRKKI, P., GISSLER, M., PUKKALA, E. & HEIKINHEIMO, O. 2018. Risk of Gynecologic Cancer According to the Type of Endometriosis. *Obstetrics & Gynecology*, 131, 1095-1102.

SAAVALAINEN, L., LASSUS, H., BUT, A., TIITINEN, A., HÄRKKI, P., GISSLER, M., PUKKALA, E. & HEIKINHEIMO, O. 2019. A cohort study of 49 933 women with surgically verified endometriosis: Increased incidence of breast cancer below the age of 40. *Acta obstetricia et gynecologica Scandinavica*, 1-7.

SAMARTZIS, E. P., NOSKE, A., DEDES, K. J., FINK, D. & IMESCH, P. 2013. ARID1A mutations and PI3K/AKT pathway alterations in endometriosis and endometriosis-associated ovarian carcinomas. *International journal of molecular sciences*, 14, 18824-18849.

SAMPSON, J. A. 1927. Metastatic or embolic endometriosis, due to the menstrual dissemination of endometrial tissue into the venous circulation. *The American journal of pathology*, 3, 93-110.

SAWYERS, C. L. 2003. Opportunities and challenges in the development of kinase inhibitor therapy for cancer. *Genes & development*, 17, 2998-3010.

SHANNON, P., MARKIEL, A., OZIER, O., BALIGA, N. S., WANG, J. T., RAMAGE, D., AMIN, N., SCHWIKOWSKI, B. & IDEKER, T. 2003. Cytoscape: a software environment for integrated models of biomolecular interaction networks. *Genome research*, 13, 2498-2504.

SHUBIK, P. 1982. Vascularization of tumors: a review. *Journal of cancer research clinical oncology*, 103, 211-226.

STAMP, J. P., GILKS, C. B., WESSELING, M., ESHRAGH, S., CEBALLOS, K., ANGLESIO, M. S., KWON, J. S., TONE, A., HUNTSMAN, D. G. & CAREY, M. S. 2016. BAF250a Expression in Atypical Endometriosis and Endometriosis-Associated Ovarian Cancer. *International Journal of Gynecological Cancer*, 26, 825-832.

SUBRAMANI, R., NANDY, S. B., PEDROZA, D. A. & LAKSHMANASWAMY, R. 2017. Role of growth hormone in breast cancer. *Endocrinology*, 158, 1543-1555.

SUPEK, F., BOŠNJAK, M., ŠKUNCA, N. & ŠMUC, T. 2011. REVIGO summarizes and visualizes long lists of gene ontology terms. *PloS one*, 6, e21800.

THOMAS, E. J. & CAMPBELL, I. 2000. Evidence that endometriosis behaves in a malignant manner. *Gynecologic and obstetric investigation*, 50, 2-10.

TOMCZAK, K., CZERWIŃSKA, P. & WIZNEROWICZ, M. 2015. The Cancer Genome Atlas (TCGA): an immeasurable source of knowledge. *Contemporary oncology*, 19, A68.

TOWNER, L. D., WHEAT, R. A., HUGHES, T. R. & MORGAN, B. P. 2016. Complement Membrane Attack and Tumorigenesis A SYSTEMS BIOLOGY APPROACH. *Journal of Biological Chemistry*, 291, 14927-14938.

UIMARI, O., JÄRVELÄ, I. & RYYNÄNEN, M. 2011. Do symptomatic endometriosis and uterine fibroids appear together? *Journal of human reproductive sciences*, 4, 34-38.

- VENN, A., WATSON, L., BRUINSMA, F., GILES, G. & HEALY, D. 1999. Risk of cancer after use of fertility drugs with in-vitro fertilisation. *The Lancet*, 354, 1586-1590.
- VERCELLINI, P., VIGANÒ, P., SOMIGLIANA, E. & FEDELE, L. 2014. Endometriosis: pathogenesis and treatment. *Nature Reviews Endocrinology*, 10, 261-265.
- WARDE-FARLEY, D., DONALDSON, S., COMES, O., ZUBERI, K., BADRAWI, R., CHAO, P., FRANZ, M., GROUIOS, C., KAZI, F., TANNUS LOPES, C., MAITLAND, A., MOSTAFAVI, S., MONTOJO, J., SHAO, Q., WRIGHT, G., D BADER, G. & MORRIS, Q. 2010. *The GeneMANIA prediction server: Biological network integration for gene prioritization and predicting gene function*.
- WEE, Y., WANG, T., LIU, Y., LI, X. & ZHAO, M. 2018. A pan-cancer study of copy number gain and up-regulation in human oncogenes. *Life sciences*, 211, 206-214.
- WEI, J.-J., WILLIAM, J. & BULUN, S. 2011. Endometriosis and ovarian cancer: a review of clinical, pathologic, and molecular aspects. *International journal of gynecological pathology: official journal of the International Society of Gynecological Pathologists*, 30, 553.
- WETENDORF, M. & DEMAYO, F. J. 2012. The progesterone receptor regulates implantation, decidualization, and glandular development via a complex paracrine signaling network. *Molecular and cellular endocrinology*, 357, 108-118.
- WIEGAND, K. C., SHAH, S. P., AL-AGHA, O. M., ZHAO, Y., TSE, K., ZENG, T., SENZ, J., MCCONECHY, M. K., ANGLESIO, M. S. & KALLOGER, S. E. 2010. ARID1A mutations in endometriosis-associated ovarian carcinomas. *New England Journal of Medicine*, 363, 1532-1543.
- YAMADA, Y., TAKAYAMA, K. I., FUJIMURA, T., ASHIKARI, D., OBINATA, D., TAKAHASHI, S., IKEDA, K., KAKUTANI, S., URANO, T. & FUKUHARA, H. 2017. A novel prognostic factor TRIM44 promotes cell proliferation and migration, and inhibits apoptosis in testicular germ cell tumor. *Cancer science*, 108, 32-41.
- YANG, C., OH, H. K. & KIM, D. 2014. Müllerian adenosarcoma arising from rectal endometriosis. *Annals of coloproctology*, 30, 232-236.
- ZHANG, Y., BAO, W., WANG, K., LU, W., WANG, H., TONG, H. & WAN, X. 2016. SOX17 is a tumor suppressor in endometrial cancer. *Oncotarget*, 7, 76036.
- ZUCCHETTO, A., SERRAINO, D., POLESEL, J., NEGRI, E., DE PAOLI, A., DAL MASO, L., MONTELLA, M., LA VECCHIA, C., FRANCESCHI, S. & TALAMINI, R. 2009. Hormone-related factors and gynecological conditions in relation to endometrial cancer risk. *European journal of cancer prevention*, 18, 316-321.